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Occupational Medicine Implications of Engineered Nanoscale

Particulate Matter

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Abstract

The imminent nanotechnology revolution promises dramatic advancements in science, technology, medicine and society as a whole. First generation products containing engineered nanoscale materials are already appearing in the marketplace, while more sophisticated products are being developed in laboratories around the world. Researchers and manufacturing employees are potentially exposed to dispersible nanoscale particulate matter via inhalation, ingestion and skin contact. Preliminary research indicates that in some cases nanoparticulate matter may be more toxic than other forms of the same or similar material. Application of the classical tools of occupational medicine and industrial hygiene is hampered by the lack of consensus guidelines for medical monitoring, exposure assessment, and exposure control.

Introduction

A revolution is underway in academic and industrial laboratories and factories around the world, where developments in nanotechnology promise a huge range of benefits for science, technology, and society. More than evolutionary, new nanoscale materials will likely prove revolutionary in many fields. This revolution will have a dramatic impact in engineering, materials sciences, chemistry, computer technology, aerospace, medicine and biological sciences, as well as a wide range of manufacturing. Potential applications of these new materials are wide open to innovation.

“Nanotechnology” is most generally defined as the intentional manipulation of matter to form novel structures with one or more dimension or features less than 100 nm. In the broadest sense nanotechnology includes work at the nanoscale in the fields of inorganic and organic chemistry, biochemistry, engineering, electronics and materials science.

While many of the points in this review may be applicable to the entire field of nanotechnology, I focus on *engineered dispersible inorganic nanoparticulate matter*, hereafter referred to as *nanoparticles*. These nanoparticles are not attached to a substrate, not part of a larger structure and can be inhaled, ingested or contaminate the skin. They are distinct from naturally occurring environmental ultrafine particles and incidentally produced nanoparticles such as diesel soot, although some engineered structures are also found in air pollution¹.

The first generation of “passive” nanoscale materials is now appearing in industrial and consumer products. This includes carbon nanotubes in composite materials used to make sporting equipment, nanoclays in cements and plastics, metals oxides in batteries, paints and sunscreens and fluoropolymers in stain repellent clothing.

Second generation “active” nanomaterials are being developed in laboratories around the world and a few are on the verge of commercialization. Medicine, in particular, is predicted to benefit tremendously from these more advanced materials. Third and fourth generation materials, which include the holy grail of nanotechnology, molecular manufacturing, are still largely beyond the horizon at this time.

Nanoparticle behavior is often strikingly different from the behavior of the chemically similar material of larger particle size. These new properties form the basis for the optimistic claims of nanotechnology pundits. The toxicity of new nanoparticles may also vary qualitatively or quantitatively from that of similar materials at the micro- or macro scale. To date, limited evidence suggests that some materials are uncharacteristically toxic at the nanoscale.

Employees involved in the development, production, distribution and use of these nanoparticles are already potentially exposed to materials of uncertain toxicity. The public is also exposed, through the use of topical sunscreens and cosmetics and ultimately through the breakdown of other nanomaterial-containing consumer products.

The challenge to occupational health professionals is to prevent the development of disease in employees handling these novel nanomaterials despite the lack of toxicological information, consensus exposure standards, air sampling methodologies and medical monitoring protocols. This is particularly difficult in R&D laboratories, where completely novel materials are developed and processes change frequently.

Sources of Concern

Nanotechnology involves a wide range of chemistries and structures, many so dramatically new as to have highly unpredictable properties. The range of chemistries used for nanoparticles is vast, as shown in table 1². Possible structures are almost unlimited, as suggested by figure 1 which reveals the many different nanoparticles possible for just one chemistry (zinc oxide). In effect, nanoscale structures may be thought of as entirely new chemicals with regard to their potential toxicity.

Unfortunately, it is very apparent that discerning the toxicity of engineered nanoparticles will not be a simple task, as details of chemistry, crystalline structure, morphology, contaminants, size and many other factors must be considered³⁻⁵. The emerging field of “nanotoxicology” has experienced growing pains due to methodological problems. For example, some early studies of the toxicity of carbon nanotubes were conducted without due consideration of the residual metal catalysts left over from synthesis. This resulted in some inconsistencies in the literature, with the assignment of toxic properties to the nanotubes that may have been the consequence of the synthetic method and residual catalyst⁶. Another early study that purported to expose rodents to single walled carbon

nanotubes used a material that was only about 50% nanotubes². A very recent inhalation study that seemed to expose animals to 95% pure carbon nanotubes used materials that actually were heavily contaminated with other fibrous carbon nanostructures.

ELEMENTS USED IN ENGINEERED NANOPARTICLES		
Aluminum	Antimony	Barium
Bismuth	Boron	Cadmium
Calcium	Carbon	Cerium
Chromium	Cobalt	Copper
Dysprosium	Erbium	Europium
Gadolinium	Gallium	Germanium
Gold	Hafnium	Holmium
Indium	Iridium	Iron
Lanthanum	Lead	Lithium
Lutetium	Magnesium	Manganese
Molybdenum	Neodymium	Nickel
Niobium	Nitrogen	Osmium
Oxygen	Palladium	Platinum
Potassium	Praseodymium	Promethium
Rhodium	Rhenium	Ruthenium
Samarium	Scandium	Silicon
Silver	Sodium	Strontium
Sulfur	Tantalum	Technetium
Terbium	Thulium	Tin
Titanium	Tungsten	Vanadium
Ytterbium	Yttrium	Zinc
Zirconium		

Table 1: Elements used in engineered nanoparticles⁴

Figure 1: The many forms of nanoscale zinc oxide may pose diverse health hazards⁷

The traditional experimental models used to evaluate the toxicity of nanoparticles may not be up to the task and may lead to false-positive or false-negative conclusions. The authors of several early *in vitro* studies using the colorimetric “MTT” assay to measure toxicity of carbon nanotubes to mitochondria failed to recognize that the carbon nanotubes directly interfered with the test, which resulted in flawed conclusions⁸. Species variability is likely to be substantial, for example the pulmonary toxicity of nanoscale TiO₂ at high doses differs substantially between rats and most other species including humans⁹⁻¹². More generally, simple, widely employed *in vitro* assays may not reliably predict *in vivo* toxicity for many nanoparticles^{13, 14}.

However, for perspective it is important to remember that people have been occupationally exposed for years to incidental nanoparticles from welding, the production of carbon black and combustion smokes, among others. The safe handling of engineered nanoparticles should build on what we know of the toxicity of these materials.

Emerging Dose Metric: Surface Area

A dramatic difference in toxicity as a function of particle size is well established in the case of quartz. Quartz particles much larger than 10 μm in equivalent aerodynamic diameter are readily removed from the upper and middle part of the respiratory tract where they deposit without consequence. However, smaller particles have the opportunity to reach the alveolar spaces in the lungs, where oxygen is transferred across the capillary membranes into the blood. Once lodged in the alveolar space, the quartz is not readily

removed by pulmonary defensive mechanisms, is toxic to pulmonary macrophages and initiates a cascade of events characterized by chronic inflammation and ending in lung fibrosis and cancer^{14, 15}.

Much of the size-specific toxicity of quartz can be ascribed to differential deposition in the respiratory tract, but two other factors, surface area and surface activity, are likely involved as well¹⁶⁻¹⁸. For a given mass of particles, as the diameter of the particles is reduced, the number of particles increases exponentially and the surface-to-volume ratio increases linearly, as shown in figure 2.

Figure 2: Increase in particle number and surface area with decrease in size¹⁹

It is well established in the use of industrial catalysts that atoms in the core of a catalyst particle contribute little; surface area is the key factor in accelerating a chemical reaction. There is accumulating evidence that the toxicity of quartz is to some extent related to a catalytic effect that causes the generation of reactive oxygen species and thus is enhanced by particles of very high surface area²⁰. This oxidant stress effect, a product both of surface area and surface reactivity, when combined with the alveolar deposition of sub-10 μm particles, makes quartz a very serious occupational health concern.

For nanoscale materials, a surprisingly large fraction of the atoms in a particle are on the surface, available for interaction with biological molecules, as shown in figure 3. In the

case of a ZnS quantum nanodot, 4 micrometers in diameter, roughly half of all the molecules in the particle are on the surface. For a single walled carbon nanotube or buckyball, every atom in the particle is on the surface.

Figure 3: Percentage of atoms on the surface of particles as a function of particle diameter^{21, 22}

In numerous studies, poorly soluble low toxicity nanoscale particles have been shown to be more toxic than microscale materials of the same composition and mass^{17, 23-28}. There is significant evidence that in the nanoscale these materials are uncharacteristically toxic due in part to accelerated generation of free radicals, hydrogen peroxide and hydroxyl atoms, driven by high surface area^{20, 22}. While there are several proposed pathways leading to these reactive oxygen species, in the end they all ultimately result in damaged DNA, proteins, lipids and other biomolecules, inflammation and even cell death.

The work of Oberdörster and collaborators in this regard is widely referenced. As shown in figure 4, Oberdörster demonstrated that nanoscale TiO₂ appeared to be much more inflammatory in lung than microscale TiO₂ particles when compared on the basis of *mass* of material introduced into the lung²⁰. However, when the data were plotted on the basis of *surface area* rather than mass, the inflammatory response was identical for both nano- and microscale particles^{17, 23}. Others have reported this surface area effect for various

particles of low toxicity including metal oxides, polymers¹⁷, carbon black²⁹⁻³², and other carbonaceous nanoparticles²²

Figure 4: Relationship between mass (left) or surface area (right) and toxicity for TiO₂ [17, 19]

However, for perspective, figure 6 shows that per unit surface area, quartz is much more toxic than TiO₂^{17, 28}. Similarly, nanoscale nickel and cobalt particles are much more toxic than TiO₂³³. Thus, it is not surface area alone that determines the toxicity of all nanoparticles, but rather the product of surface area, surface reactivity, and elemental toxicity. This variability in surface reactivity even extends to the various crystalline forms of TiO₂^{14, 15}.

Indeed, even in the well-worn case of quartz there is a wide range of bioactivity in samples obtained from different parts of the earth or handled differently¹⁷ and much of the toxicity can be erased by prior treatment with aluminum¹⁹

Figure 5: Relative inflammatory potency of SiO₂ compared to TiO₂ and BaSO₄ particles on an equal surface area basis³⁴

It is important to note that not all researchers have been able to reproduce this surface area-dependent effect for a range of nanoscale particles^{33, 35, 36}, and some of the aforementioned positive studies have been criticized on the way in which they derived and interpreted their data³⁷. However, at this time, for nanomaterials of low solubility and low intrinsic toxicity, surface area as determined empirically is likely the best dose metric.

Case Study: Environmental Ultrafines

There is mounting evidence that exposure to environmental ultrafine (~nano) particles, particularly combustion derived nanoparticles (CDNP), contributes to community respiratory and cardiovascular morbidity and mortality³⁸⁻⁴⁰. Epidemiological and experimental studies have consistently indicated that exposure to these incidental nanoparticles predisposes compromised people to illness one or a few days post-exposure^{37, 41-45}. Originally attributed to larger particles, it is now likely that much of this observed health impact is due to ultrafine CDNP in air pollution¹.

It is likely that the toxic potential of some engineered nanoparticles will parallel this effect of CDNP. For example it appears that pulmonary deposition of carbon nanotubes has some of the same adverse cardiovascular effects as CDNP. Indeed, it is now known that CDNP air pollution includes large numbers of multi-walled carbon nanotubes^{46, 47}.

Case Study: Carbon Nanotubes

The special case of carbon nanotubes is illustrative of many of the difficulties in assessing the toxicity of novel nanostructures.

Carbon nanotubes come in two primary forms—single walled nanotubes (SWCNT) and nested multiwalled nanotubes (MWCNT). They are being produced by the ton and incorporated into many commercial products including baseball bats, bicycles and other sporting equipment. Nanotubes range in diameter from about one nanometer (SWCNT) to dozens of nanometers (MWCNT) and can have lengths into the micrometer range.

Figure 6: Single and multi-walled carbon nanotubes

A large number of in vitro toxicity studies have been reported for carbon nanotubes, with most demonstrating unusual cytotoxicity to a range of target cells, as shown in table 2.

Year	Author	Cell Line	Main findings
2003	Shvedova	Human skin fibroblasts	Cell death, oxidative stress
2005	Ding	Human skin/lung fibroblasts	MWCNT induce dose-dependent cytotoxicity, induce genes indicative of a strong immune, stress and inflammatory response
2005	Jia	Human lung macrophages	SWCNT more toxic than MWCNT10, both more toxic than quartz
2005	Murr	Mouse lung macrophages	S/MWCNT “ropes” showed dose related cytotoxicity, more toxic than asbestos
2005	Fiorito	Mouse & Human macrophages	CNT were not well taken up by macrophages and caused little toxicity, metals cause CNT toxicity
2006	Kagan	Human lung macrophages	Oxidative stress is related to iron contamination, macrophages do not effectively engulf CNTs
2006	Tian	Human fibroblasts	Surface area predicts cytotoxicity, SWCNT more toxic than MWCNT
2006	Pluscamp	Lung macrophage and epithelial cells	Little acute cytotoxicity of MWCNT, toxicity related to metal contamination
2006	Tian	Human fibroblast	Surface area predicts cytotoxicity, SWCNT more toxic than MWCNT or other carbon. Refined SWCNT more toxic than unrefined SWCNT
2007	Wick	Human mesothelioma	Nanoropes more toxic than asbestos, dispersed CNTs less toxic

Table 2: Selected *in vitro* studies of carbon nanotube cytotoxicity

In some of these *in vitro* studies, nanotubes appeared to be more toxic than quartz or asbestos, both of which induce lung inflammation, fibrosis and ultimately cancer. Several of the authors ascribed the observed toxicity to the metals contaminating impure carbon nanotubes^{6, 48-51}.

If carbon nanotubes are instilled or aspirated into the lungs of rodents they induce signs of oxidative stress, much like the metal oxide nanoparticles discussed previously, and in

most cases cause fibrosis and granuloma formation⁵². Depletion of vitamin E, a potent antioxidant, exacerbates the oxidative stress and profibrotic activity⁵³. All of the published studies of this type are summarized in table 3.

However, these dosing methods are clearly non-physiological, it is quite possible that some of these pathologies, particularly the granulomas, are artifacts of the assay and will not occur in occupationally exposed individuals. A very recent publication from NIOSH supports this hypothesis, where they found no granulomas when they used extremely finely ground single walled carbon nanotubes rather than larger particles of agglomerated tubes⁵⁴.

Year	Author	Species	Granuloma	Inflammation	Fibrosis
2001	Huczko	G. pig	NA	–	NA
2004	Warheit	Rat	+	+/-	+
2004	Lam	Rat	+	+	NA
2005	Muller	Rat	+	+	+
2005	Grubek - Jaworska	G. Pig	+	+	+
2005	Shvedova	Mouse	+	+	+
2006	Mangum	Rats	+	–	+
2006	Carrero - Sanchez	Mice	+	+	NA
2007	Shvedova	Mice	+	+	+
2008	Mercer	Mice	–	+	+

Table 3: Summary of findings from all published carbon nanotube instillation/aspiration pulmonary toxicology studies

Mitchell recently published an inhalation study where mice were exposed to multi-walled carbon nanotubes⁵⁵. No evidence of lung inflammation, fibrosis or granuloma formation was detected, but the authors did find evidence of impairment of the animal's immune systems, a new and unique finding. The Mitchell study has been challenged due to a number of significant methodological problems, thus its validity is uncertain^{56 32}.

In contrast, preliminary reports from NIOSH indicate that inhaled single and multiwalled carbon nanotubes cause rapid but transient inflammation and consistent diffuse lung fibrosis. In the SWCNT inhalation study, the fibrosis is reported to be four times as severe than was seen for the same doses via aspiration^{57, 58}. No mention of granulomas was made in this context.

Carbon nanotubes may also cause cancer, based on their morphology and biodurability. Carbon nanotubes can be viewed as rolled-up layers of graphite that forms a single tube about a nanometer in diameter (SWCNT) or a series of concentric nanotubes that can be a few or dozens of nanometers in diameter (MWCNTs). Carbon nanotubes can be thousands of nanometers in length, they have high tensile strength⁴⁹ and relatively low solubility in biological systems^{1, 59}. They tend to cling together to make larger structures called nanoropes that are many nanometers or even micrometers in diameter.

These characteristics are all remarkably similar to a naturally occurring magnesium silicate nanotube, chrysotile asbestos, shown in figure 7⁵⁹.

Figure 7: Cross section of fibril of chrysotile asbestos showing lamellar structure and size very similar to MWCNTs^{60, 61}

Inhaled chrysotile asbestos causes macrophage death, respiratory inflammation, fibrosis, lung cancer and probably mesothelioma, a cancer of the lining of the lungs and other organs⁶². However, these effects are not unique to the chemistry of chrysotile. Indeed, the amphibole forms of “asbestos”, which are chemically unrelated to chrysotile and do not share the lamellar structure, also induce fibrosis and cancer. The occurrence of fibrous erionite (a form of zeolite) in the Cappadokia region of Turkey and elsewhere is associated with a highly elevated risk of mesothelioma⁶³. Even man made fibers, such as some mineral fibers and refractory ceramic fibers, have the potential to induce these diseases⁶⁴.

Originally proposed by Stanton and Wrench in 1972⁶³, it is now generally accepted that inhaled fibrous particles have the potential to cause fibrosis and cancer if they meet certain criteria of size, shape and biodurability:

- Particles must be small enough to be deposited in the alveoli
- Particles must have the right shape, including a high aspect ratio, a length of over 5 μm or more and a sub-micrometer diameter.
- Particles must resist dissolution and clearance in the lungs

As shown in table 4, widely divergent fiber chemistries cause the same toxic endpoints if they have the right size, shape and biodurability⁴⁹.

It is evident that CNTs have the requisite size, strength and morphology to be suspect in this model. Very limited data are available on their biodurability. Muller⁶³ showed that 80% of unground and 36% of ground MWCNTs were retained in lung tissue after 60 days, suggesting that MWCNTs may be adequately persistent to cause fibrosis and cancer.

Fiber	Type	Lung Half Life (Days)	Fibrosis	Tumors
MMVF34	Stone Wool	6	–	–
MMVF11	Glass Wool	9	–	–
MMVF10	Glass Wool	37	–	–
MMVF33	475 Glass	49	+	+/- ^a
RCF1a	Refractory	55	+	+
MMVF32	E Glass	79	+	+
Amosite	Asbestos	418	+	+
Crocidolite	Asbestos	817	+	+

^a. Positive in hamsters but not rats

Table 4: Correlation between lung biopersistence of long fibers and lung pathology³²

Limited *in vitro* testing indicates that carbon nanotubes, like asbestos, can interact with DNA⁶⁵ and cause large scale chromosomal damage, but show no activity in the Ames point mutation assay^{66, 67}. Carbon nanotubes and asbestos interact with tissue to create reactive oxygen species^{32, 68}. However, no one has studied whole animals past 90 days to truly assess the carcinogenicity of inhaled carbon nanotubes.

Surprisingly, carbon nanotubes deposited in the respiratory tract can induce toxic effects in other organ systems. A recent NIOSH report, described in more detail below, shows a range of cardiovascular toxicity due to inspired single walled carbon nanotubes⁵⁸.

While some material safety data sheets have suggested that the exposure limit for carbon nanotubes should be based on the graphite standard (TLV = 5 mg/m³ for respirable dust), the instillation/aspiration studies indicate that this exposure level may be unsafe⁶⁹. Some authors have suggested the use of the PEL for quartz, 0.1µg/m³, as a better starting point⁷⁰.

The hazard posed by any workplace chemical is a product of both the chemicals intrinsic toxicity and the potential for exposure. Few studies have assessed the exposure potential during the handling of carbon nanotubes. The most notable investigation found levels of up to 53 µg/m³ in an occupational setting where nanotubes were made and harvested⁷¹. Free fibers were rare; almost all of the carbon nanotubes measured were large aggregates. In this regard carbon nanotubes are very different from chrysotile asbestos, where handling of processed mineral fiber does release a large number of free nanofibers.

Case Study: Quantum Dots

Quantum nanodots are single-digit sized particles made up of semiconductor metals that demonstrate the amazing feature of changing fluorescence wavelength based on their size. Quantum nanodots present an interesting case in that many of these are intrinsically cytotoxic due to their metal content (e.g. Cd, Pb, Se)⁷². Uncoated nanodots are quite

cytotoxic, and it is possible that their toxicity exceeds the sum of the toxicity of the constituent metals. For example, Cho⁷³ showed that cytotoxicity of a variety of coated nanodots in a breast cancer cell line did not fully correlate with the generation of Cd²⁺ ions. Instead, the quantum nanodots were consistently *more toxic* than predicted by their release of Cd²⁺ ion (figure 8). In this study quantum dot net toxicity appears to be a result of both intrinsic metal ion toxicity and induction of oxidative stress by the surface of the intact nanoparticle, the latter effect the same as seen for TiO₂, carbon nanotubes and other nanoparticles.

Fortunately, quantum dots can be coated with various polymers and biologically compatible molecules that shield the semiconducting core from dissolving or interacting with tissue, which greatly reduces their toxicity⁷².

Figure 8: Plot showing reduction in cell viability as a function the propensity of Cd²⁺ ions to plate off of CdSe quantum nanodots with different protective coatings. The dotted line represents the dose/toxicity relationship for pure Cd²⁺ ions. The nanodots are uniformly more toxic than would be predicted by their release of Cd²⁺ ions^{74, 75}.

Distribution Across Anatomical Barriers and Systemic Effects

Exposure to nanoscale particles can occur via any of the usual routes of exposure, that is, inhalation, ingestion and skin contact. As with other chemical occupational stressors,

each of these routes of exposure must be evaluated to determine the extent of deposition, absorption, distribution, excretion and toxicity.

Insoluble nanoparticles may be more mobile across anatomical barriers than microscale or larger insoluble particles of the same chemistry. Also, nanoparticles may exert systemic toxic effects that may not depend on translocation of the particles.

Digestive and Respiratory Tracts: It has been known for many years that some intact nanoparticles cross the digestive tract and respiratory tract and appear in the body⁷⁶.

Although subject to ongoing controversy related to methodological limitations^{76, 77}, inhaled nanoparticles have some ability to cross through or around the cells in the lungs, enter the interstitial space and are distributed systemically⁷⁶. This clearly happens, but the extent of this process and significance remains in question. Early studies that suggested very rapid translocation of nanoparticles out of lung^{53, 78-80} were likely flawed, more recent work has suggested a slow migration of a small percentage of particles out of the lung that is exacerbated by lung inflammation⁸¹.

Skin: Many sunscreens contain micro or nanoscale zinc or titanium oxide particles. In general, studies suggest that intact skin is a pretty good barrier to these particles⁸². Under some circumstances, sub micrometer particles can penetrate the skin, at least as far as the living tissue underlying the stratum corneum^{82, 83}. It is not clear that these particles travel as far as the systemic circulation or if they are toxicologically significant. This route of

exposure is likely to be of greater importance for damaged skin or concomitant exposure to solvents and nanoparticles⁸³.

Figure 9: Image showing 500 nm fluorescent beads penetrating to the living layers of the skin, but 4 μ m beads stopped on the surface of the skin⁸⁴.

Brain: Although not completely unprecedented⁸⁵, it was certainly remarkable when it was demonstrated that carbon nanoparticles⁸⁶ and manganese oxide nanoparticles^{87, 88} deposited in the olfactory mucosa in rodents translocated up the olfactory neuron axons into the brain and in some studies triggered inflammation in neural tissue. This is intriguing in light of the lung deposition model proposed by the International Council on Radiation Protection that shows nanoparticles less than 10 nm in aerodynamic diameter will preferentially deposit in the head airways region rather than the alveolar space⁸⁹⁻⁹¹. There is also evidence that some, but not all, nanoparticles can penetrate the blood-brain barrier and gain access to the brain via the bloodstream^{91, 92}.

The importance of these brain translocation mechanisms in humans is unknown. Humans have far less olfactory mucosa than rodents. For perspective, welders have been inhaling adventitious metal oxide nanoparticles for decades with relatively little obvious adverse neurological effect for metals other than manganese.

Placenta: The placenta seems to present a more formidable barrier to nanoparticle translocation⁹³, although at least one report suggested translocation into the fetus^{91, 94}.

Systemic Toxicity

In general, the consequence of translocated nanoparticles has not been established. As might be expected, nanoparticles in the blood stream are sometimes accumulated in the liver and lymph nodes, as shown for quantum dots in a mouse in figure 10⁹⁴.

*Figure 10: Accumulation of injected quantum nanodots in the lymph nodes, bone marrow and liver of a mouse*⁶⁸.

Li of NIOSH studied the cardiovascular toxicity of carbon nanotubes aspirated into the lungs of rats genetically modified to rapidly develop atherosclerosis⁴². This pulmonary exposure resulted in cardiovascular toxicity, including accelerated atherosclerosis, oxidative stress in aorta and heart tissue and damage to aortic mitochondrial DNA. The exposure level used in this study was intended to approximate the potential human exposure at the PEL for respirable graphite.

It is not known if this cardiovascular toxicity was due to translocated nanotubes interacting directly with aortic and heart tissue or some type of secondary response, due to the observed inflammation of lung tissue. Recall that inhalation of environmental ultrafine combustion particulate matter also induces cardiovascular toxicity³².

As yet unpublished work by Castranova at NIOSH finds that TiO₂ particles instilled into the lungs of rodents causes dysfunction of the microvascular system³². Treated animals showed blunting and even reversal of the response to dilators that was dose dependent, more severe for nanoparticles than for microparticles and appeared within 1 day of dosing. Confocal microscopy demonstrated rapid accumulation of polymorphonuclear leukocytes all along the microvascular walls. This is likely a systemic effect rather than due to translocated particles based on the rapidity of response.

NIOSH has also presented preliminary data which suggest that pulmonary exposure to MWCNTs and TiO₂ nanowires may degrade the integrity of the blood brain barrier and cause brain damage, primarily in the olfactory bulb, hippocampus and frontal cortex⁹⁵. This was not due to translocation up the olfactory nerve. In the case of the MWCNT this effect was seen with doses in the range of what a person would be exposed to at the PEL for graphite.

Very little work has been done to assess the potential reproductive toxicity of engineered nanoparticles. One brief report indicates that gold nanoparticles may have a negative impact on sperm function in vitro⁹⁶. A recent study found that pulmonary deposition of carbon black had negative impact on the reproductive system of male mice^{93, 97}. Older references suggests that C60 may have fetotoxic potential⁹⁸. These studies are two few and incomplete to allow any conclusions regarding the reproductive toxicity of nanoparticles.

Goals of an Occupational Medicine Program

Broadly construed, occupational medicine programs attempt to limit the health effects of chemical and physical stressors in the workplace. With respect to chemically induced disease, the goals of an occupational medicine program are, in order of preference:

- 1. Prevent occupational diseases from occurring**
- 2. Quickly detect occupational diseases that do occur**
- 3. Intervene to cure occupational diseases**

Goal 1: Prevent Occupational Disease

This goal, preventing occupational illness entirely, is the loftiest ambition of an occupational medicine program. Traditionally, the paradigm for achieving this goal relies on four key program elements:

Workplace Exposure Monitoring—Exposure to chemical agents is assessed either by environmental monitoring (e.g., air monitoring, dermal exposure assessment) or biological monitoring (e.g., blood analysis, urinalysis, lung counting). The results of these assays are compared to established limits as an index of the risk.

Establish Workplace Controls—Controls are established to reduce employee exposure to occupational stressors. Controls may include engineered controls

(e.g., ventilation, filtration, enclosure), administrative controls (e.g., safe work practices, training) and personal protective equipment (e.g., gloves, respirators, goggles).

Medical Pre-Screening for People at Elevated Risk—Prior to exposure to an occupational stressor, the working population is screened for conditions that may put them at elevated risk of occupational disease. At-risk employees may be offered alternative assignments or enhanced protection to reduce their chances of becoming ill.

Medical Surveillance— In this context, medical surveillance is narrowly defined to describe a process of looking for health trends in the worker community that might warrant further action. This is distinct from “medical monitoring”, which is the key element of goal two and has a clinical focus⁹⁸. Note that in practice many people use the terms medical monitoring and medical surveillance interchangeably, but in fact they are conceptually distinct. Some OSHA standards have “medical surveillance” provisions, but these are better described as “medical monitoring”.¹⁰

This paradigm requires several elements to be in place:

- Exposure monitoring methods are available and affordable
- A standard exists to which measured exposures can be compared
- Control methods applicable to larger particles are effective for nanoparticles

- The target organ(s) and health effects are known and can be screened
- The impact of pre-existing conditions on risk are known
- Alternatives are available for employees found to be at increased risk of disease due to preexisting conditions

For most nanoscale particulate matter, there are no accepted exposure monitoring methods, no exposure standards, the effectiveness of traditional control methods is only now being elucidated, the target organs are not always obvious and the impact of pre-existing conditions on risk is not clear. This makes it very difficult to establish an evidence-based program to prevent the manifestation of occupational disease related to nanoparticles.

Exposure Monitoring

As of December 2007, there are almost no published consensus methods to measure exposure to nanoscale materials or exposure standards to compare to the results. NIOSH has proposed a draft an exposure limit of 0.1 mg/m³ for nanoscale TiO₂, which stands alone as a widely recognized exposure standard specific for engineered nanostructured materials in the U.S.⁹⁹.

It is a simple matter to buy handheld condensation nuclei counters that can enumerate airborne nanoparticles down to 10 nm in diameter, but it is difficult even to obtain relative measurements with these instruments due to the extremely high and variable background level of natural and anthropogenic ultrafine particles. In laboratory settings,

the signal of interest is likely to be a small fraction of the background noise. Also these instruments are not size or chemistry specific. Even with these limitations these instruments have been used successfully in factory settings as part of research efforts¹⁰⁰.

Size-selective real time aerosol monitors for measuring nanoscale particulate matter, such as mobility particle spectrometers, are available but this equipment is very expensive, large, and requires special training to operate. A number of researchers have reported successful deployment of this type of instrument in labs and factories¹⁰¹⁻¹⁰³. Like the simpler condensation nuclei counters, these instruments are not able to distinguish engineered nanoparticles from background ultrafines.

Particles can be collected on filters or other media with subsequent analysis by electron microscopy. This allows for speciation and sizing of nanoparticles, but at huge cost in terms of time and expense and requires expertise that is of very limited availability right now.

NIOSH has reported the results of an investigation where they built a clean room around a carbon nanotube source, thus removing the confounding background particles. They also avoided the problem with nanoparticle measurement techniques by measuring the level of airborne residual catalyst metals and back-calculating the exposure to carbon nanotubes. Interestingly, the authors found only relatively low levels of carbon nanotubes in their air samples and attributed this result to the extensive agglomeration of the newly synthesized tubes into micro- and macro clumps that did not readily become airborne.

This level of effort is feasible for a funded research study, but not for routine exposure monitoring, especially in research labs where the work changes all the time.

The vast majority of industrial hygiene exposure limits for particulate matter are specified on a mass basis. Thus OSHA limits exposure to lead to 50 $\mu\text{g}/\text{m}^3$ of air averaged over an 8-hour day. The exposure limit for some fibrous materials is based on particle count. However, there is no generally accepted sampling method to evaluate particle surface area, likely the most relevant exposure metric for many nanoscale particles. Some research in this area has been done¹⁰⁴ and at least one vendor offers an instrument purported to measure surface area directly, but this suffers from the same interference from ambient ultrafines and lack of chemical specificity as the simple particle counting instruments. This instrument is primarily a research tool at this time.

Where existing exposure limits based on chemistry are available, great caution must be exercised before applying them to nanoparticles. For example, the toxicity of uncoated CdSe quantum nanodots may be a function both of the intrinsic metal ion toxicity and the catalytic promotion of oxidative stress that is a property of the intact nanoparticle. Existing Cd exposure limits do not account for these dual pathways to toxicity. Similarly, use of the graphite exposure limit for structurally related carbon nanotubes may lead to disease, as the morphology of the nanotube induces additional toxicological mechanisms that are not accounted for in the exposure standard.

Medical Pre-Screening

There is currently no technical basis for recommending medical prescreening criteria for most nanoparticle exposed workers.

Exposure Control

Until quite recently, it was not clear that the usual triumvirate of engineered, administrative and personal protective controls was adequate to control exposure to nanoparticles.

In 1991 it was proposed that nanoparticles smaller than about 10 nm might not be effectively captured by mechanical air filters due to a “thermal rebound” effect¹⁰⁵. The authors calculated that at some size the nanoparticles would rebound from the filter matrix due to their propensity to diffuse and thus not be captured, much in the way that individual vapor or gas molecules pass unchecked through a filter. In 2004 a study was published that purported to demonstrate this effect for very small nanoparticles¹⁰⁶. Other authors reported similar findings¹⁰⁷. About this time it was also reported that N95 respirator filters, especially those that rely on electrostatic charge effects for particle deposition, might not perform quite at their stated efficiency for nanoparticles¹⁰⁸⁻¹¹³.

Overwhelming data are now available from numerous investigators showing filters work as expected for particles as small as 2 nanometers^{109, 110, 114, 115}. The earlier negative reports suffered from methodological problems that resulted in erroneous conclusions¹¹⁰.

Of course, at some size, filtration efficiency must drop off, as air and vapor molecules are not captured in a particulate filter. Data from Kim suggest that thermal rebound does appear at about 2 nm, just about the diameter of a buckyball¹⁰⁸.

Discussion continues regarding the permeability of gloves and other elastomeric materials to nanoparticles. However, limited studies to date indicate that latex and nitrile rubber gloves form a reliable barrier to nanoparticles under test conditions¹¹⁰.

Figure 11: Penetration of nanoparticles through a low efficiency filter as a function of size, showing that particles below 2 nm may not be captured as predicted by filtration theory^{100, 116}

Despite some suggestions to the contrary, local exhaust ventilation will function for engineered nanoscale particles exactly as it has all along for incidentally produced nanoparticles¹⁰⁰.

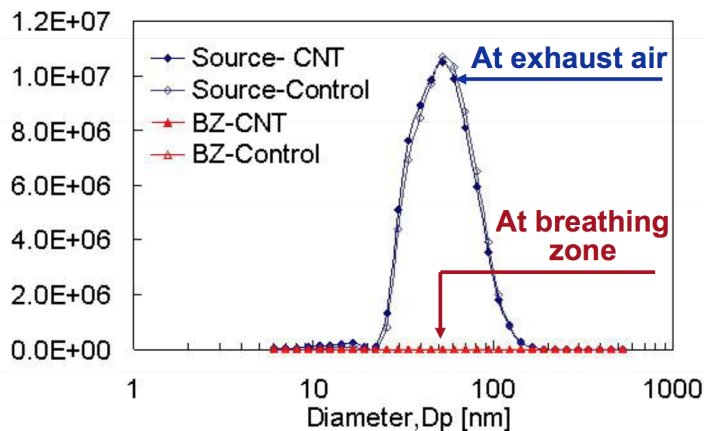


Figure 12: Demonstration of the effectiveness of a fume hood at preventing exposure to carbon nanotubes¹¹⁷⁻¹¹⁹.

In summary, the management of exposure to nanoparticles can in most cases be achieved using familiar engineered, administrative and personal protective control measures.

Medical Surveillance

NIOSH, the NNI and Nasterlick from BASF have recommended the establishment of worker medical surveillance programs to monitor for the emergence of sentinel cases of new disease^{98, 117}. At this time it is unlikely that most employers have the capability or will to establish a meaningful sentinel event medical surveillance program, so as a practical matter, medical surveillance is not likely to figure prominently in occupational medicine programs for nanoparticles for the near term.

Goal 2: Detect Occupational Disease Quickly

The second goal of occupational health surveillance is to detect subclinical signs of illness in a worker population, with an eye toward quick intervention to prevent development of overt disease. This process is most commonly called “medical monitoring” in the United States¹¹⁹ and is mandated by the Occupational Safety and Health Administration for some chemical agents such as asbestos, lead and benzene (although most laboratory work is exempted from these requirements).

The criteria for establishing a medical monitoring program include^{117, 119}:

- Understanding who is being exposed
- Knowledge of the target organ(s) and specific health effects
- Availability of reliable and safe medical tests
- Action criteria to compare to test results
- Availability of interventions to arrest or reverse disease

Medical monitoring often includes diagnostic studies to identify perturbations that disclose the preliminary stages of occupational disease. Thus for asbestos, which causes pulmonary fibrosis, lung cancer and mesothelioma, OSHA mandates that the medical evaluation include a chest x-ray and pulmonary function testing. Whereas preclinical detection of mesothelioma is largely irrelevant (it is essentially 100% fatal), the discovery of subclinical disease may result in limiting further asbestos exposure that can reduce the progression of fibrosis. Early-stage lung cancer may be treatable via surgery and other interventions.

For most nanomaterials, it is unclear what diagnostic studies should be included in a medical monitoring program¹¹⁹. While many suggestions have been, including measurement of heart rate variability, proinflammatory cytokines, lung CT studies, liver enzyme tests, etc., none of these rise to the level of validation normally required for inclusion in a targeted medical monitoring program¹¹⁹. The sensitivity, specificity and risk/benefit ratio of such testing is unknown with respect to most nanoparticles.

It is also not always clear who should be included in a monitoring program, as the usual inclusion criteria incorporate the results of exposure monitoring that are not likely to be available for nanoparticles.

Nasterlack and colleagues at BASF in Germany published an opinion paper that argues that routine medical monitoring of workers exposed to nanoparticles is not warranted or feasible at this time, and effort should instead be expended on control measures to prevent exposure¹¹⁷.

NIOSH has recently published a draft guideline that proposes “Insufficient scientific and medical evidence now exists to recommend the specific medical screening of workers potentially exposed to engineered nanoparticles”.

The feasibility of medical monitoring is likely to evolve as the findings of more whole animal toxicological studies become available. For example, if new inhalation studies confirms the relationship between carbon nanotubes inhalation and lung fibrosis and granuloma formation, it will be reasonable to formulate specific guidelines for medical surveillance of workers exposed to these materials.

Goal 3: Treatment of Disease

If all else fails and occupational disease is manifested, the third goal of occupational medicine is to heal those injured by their experience at work. This might be affected by

removing the injured individual from further exposure via transfer, or via some form of treatment.

Medical removal is not always effective at limiting the progression of disease and raises real concerns for both the employer and employee. This inevitably leads to ethical and practical quandaries. Specifically, is it justifiable to remove a worker from his or her job based on the results of uncertain tests, without knowing if this intervention will make any difference in disease outcome?

Conclusion

As the discovery and commercialization of nanoscale materials expands, occupational health professionals such as physicians and industrial hygienists will be forced to develop hazard assessment, exposure control and health monitoring strategies without the usual panoply of tools.

This quandary is not that unusual in a research setting such as a university or pharmaceutical company, where the creation of novel materials is the stock-in-trade. However the widespread use of materials of uncertain hazard in diverse industries, big and small, is unusual and may pose an unacceptable risk that will not be recognized until cases of disease start appearing in number.

The emerging nanotechnology revolution is another grand step in the industrial revolution that has been underway for over 200 years. As with prior steps in the revolution there will likely be anticipated and unanticipated consequences, both good and bad, of new technologies. The goal should be to anticipate and mitigate adverse consequences before people are injured or the environment is contaminated. If history is any indication, this will be a very difficult task.

Strategies to manage the poorly defined risk of nanoparticulate matter are beginning to appear from various government and consensus standard setting organizations in the United States and Europe. The companion manuscript to this paper presents the hazard assessment and control recommendations for research laboratories developed by the five Department of Energy Nanoscale Science Research Centers

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References:

1. L. E. Murr; K. F. Soto, *Journal of Materials Science* **2004**, 39, (15), 4941-4947.
2. ETCGroup, *Occasional Paper Series* **2003**, 7, (1), 20.
3. P. Wick; P. Manser; L. G. Limbach; U. Dettlaff-Weglikowski; F. Krumeich; S. Roth; W. J. Stark; A. Bruinink, *Toxicology Letters* **2007**, 168, (2), 121-131.
4. L. Z. Wang, *Materials Today* **2004**, 7, 7.
5. V. E. Kagan; Y. Y. Tyurina; V. A. Tyurin; N. V. Kondura; A. I. Potapovich; A. N. Osipov; E. R. Kisin; R. Schwegler-Berry; V. Mercer; V. Castranova; A. A. Shvdova, *Toxicology Letters* **2005**, 165, (1), 13.
6. D. B. Warheit; B. R. Laurence; K. L. Reed; D. H. Roach; G. A. M. Reynolds; T. R. Webb, *Toxicological Sciences* **2004**, 77, (1), 117-125.
7. J. M. Wörle-Knirsch; K. Pulskamp; H. F. Krug, *Nano Letters* **2006**, 6, (6), 1261-1268.
8. L. S. Levy, *Indoor and Built Environment* **1995**, 4, (5), 9.
9. ILSI, *Inhalation toxicology* **2000**, 12, (1-2), 17.
10. NIOSH, in: November 22, 2005 ed.; N. I. f. O. S. a. Health, (Ed.) 2005.
11. E. Bermudex; J. Mangum; B. Wong; A. Bahman; P. Hext; Warheit, *Toxicological sciences* **2004**, 77, 11.
12. C. M. Sayes; K. L. Reed; D. B. Warheit, *Toxicol Sci* **2007**, 97, (1), 18.
13. C. M. Sayes; A. Marchione; K. Reed; D. Warheit, *Nano Letters* **2007**, 7, (8), 8.
14. K. Donaldson; P. Borm, *Annals of Occupational Hygiene* **1998**, 42, (5), 8.
15. C. Albrecht; A. M. Knaapen; A. Becker; D. Höhr; P. Haberzettl; F. J. van Schooten; P. J. A. Borm; R. Schins, *Respiratory Research* **2005**, 6, (129).
16. D. B. Warheit, *Toxicological sciences* **2007**, 95, (1), 11.
17. R. Duffin; L. Tran; D. Brown; V. Stone; K. Donaldson, *Inhalation Toxicology* **2007**, 19, (10), 849-856.

18. B. Railsback, in: teaching materials, Basic Geology, U. o. G. Department of Geology, 2002, "Increase in particle number and surface area with decrease in particle size", <http://www.gly.uga.edu/railsback/1121WeatheringArea.jpeg>
19. A. D. Maynard, *Annals of Occupational Hygiene* **2007**, 51, (1), 12.
20. G. Oberdörster; E. Oberdörster; J. Oberdörster, *Environmental Health Perspectives* **2005**, 113, (7), 823.
21. Q. Zhang; Y. Kusaka; X. Zhu; K. Sato; Y. Mo; T. Kluz; K. Donaldson, *Journal of Occupational Health* **2003**, 45, (1), 8.
22. G. Oberdörster, *Philosophical transactions: Mathematical, Physical and Engineering Sciences* **2000**, 358, (1175), 22.
23. D. M. Brown; M. R. Wilson; W. MacNee; V. Stone; K. Donaldson, *Toxicology and applied Pharmacology* **2001**, 175, 9.
24. V. Stone; M. Tuinman; J. Vamvakopoulos; J. Shaw; D. Brown; S. Petterson; Faux; S. P.; W. Borm; W. Macnee; F. Michaelangeli; K. Donaldson, *Eruopean Respiratory Journal* **2000**, 15, 7.
25. X. Y. Li; P. Gilmour; K. Donaldson; W. MACNee, *Environmental health prespectives* **1997**, 105, (S5), 3.
26. K. Donaldson; V. Stone; D. M. Gilmour; M. Brown; W. MacNee, *Philosophical Transactions: Mathematical, physical and engineering sciences* **2000**, 358, (1175), 9.
27. C. M. Sayes; A. M. Gobin; K. D. Ausman; J. Mendez; J. L. Wst; V. L. Colvin, *Biomaterials* **2005**, 26, (36), 9.
28. Q. Zhang; Y. Kusaka; K. Sato; K. Makakuku; N. Kohyama; K. Donaldson, *53* **1998**, 6, 16.
29. T. Stoeger; C. Reinhard; S. Takenaka; A. Schroepel; E. Karg; B. Ritter; J. Heyder; H. Schulz, *Environmental health perspectives* **2006**, 114, (3), 6.
30. C. Monteiller; L. Tran; W. MacNee; S. Faux; A. Jones; B. Miller; K. Donaldson, *Occup Environ Med* **2007**, 64, (9), 7.
31. A. Maynard; E. Kuempel, *Journal of Nanoparticle reseasrch* **2005**, 7, 29.
32. V. Castranova, in: *Nanotechnology Update for IHs: Toxicology and exposure assessment*, A. I. H. Association, (Ed.) American Industrial Hygiene Association: 2008.

33. D. Warheit; T. Webb; S. Frerichs; C. Sayes, **2007**.
34. K. Soto; K. M. Garza; L. E. Murr, *Acta Biomaterialia* **2007**, 3, (3), 351-358.
35. D. Warheit; T. Webb; C. M. Sayes; V. L. Colvin; K. L. Reed, *Toxicological sciences* **2006**, 91, (1), 10.
36. K. Wittmaak, *Environmental health perspectives* **2007**, 115, (2), 8.
37. K. Donaldson; L. Tran; L. A. Jimenez; R. Duffin; D. E. Newby; N. Mills; W. MacNee; V. Stone, *Particle and Fibre Toxicology* **2005**, 2, (10), 1743-8977.
38. D. W. Dockery; C. A. Pope; X. Xu; J. Spengler; J. Ware; M. E. Fay; B. G. Ferris; F. E. Speizer, *New England Journal of Medicine* **1993**, 329, 7.
39. S. v. Klot; A. Peters; P. Aalto; T. Bellander; N. B. D. D'Ippoliti; R. Elosua; A. Hörmann; M. Kulmala; T. Lanki; H. Löwel; J. Pekkanen; S. Picciotto; J. Sunyer; F. Forastiere, *Circulation* **2005**, 112, 7.
40. R. Rückerl; R. Phillips; A. Schneider; M. Frampton; J. Cyrus; G. Oberdörster; H. E. Wichmann; A. Peters, *Particle and Fibre Toxicology* **2007**, 4, (1), 14.
41. G. Oberdörster; J. ZFinkelstein; J. Ferin; R. Gelein; C. Johnson; J. Godleski; L.-Y. Change; J. D. Crapo, *Chest* **1996**, 109, 2.
42. A. Nemmar; M. F. Hoylaerts; P. Hoet; B. Nemery, *Toxicology letters* **2004**, 149, 11.
43. P. Penttinen; K. L. Timonen; P. Tiittanen; A. Mirme; J. Ruuskanen; J. Pekkanen, *European REspiratory Journal* **2001**, 17, 8.
44. A. Peters; H. E. Wichmann; T. Tuch; J. Heinrich; J. Heyder, *American Journal of Respiratory Critical Care Medicine* **1997**, 155, 8.
45. R. Duffin; N. Mills; K. Donaldson, *Yonsei Medical Journal* **2007**, 48, (4), 12.
46. K. Pulskamp; S. Diabaté; H. F. Krug, *Toxicology Letters* **2007**, 168, (1), 58-74.
47. V. E. Kagan; Y. Y. Tyurina; V. A. Tyurin; N. V. Konduru; A. I. Potapovich; A. N. Osipov; E. R. Kisin; D. Schwegler-Berry; R. Mercer; V. Castranova; A. A. Shvedova, *Toxicology Letters* **2006**, 165, (1), 88-100.
48. J. B. Mangum; E. A. Turpin; A. Antao-Menezes; M. F. Cesta; E. Bermudez; J. C. Bonner, *Particle & Fibre Toxicology* **2006**, 3, (15).

49. J. Muller; F. Huaux; N. Moreau; P. Misson; J.-F. Heilier; M. Delos; M. Arras; A. Fonseca; J. B. Nagy; D. Lison, *Toxicology and Applied Pharmacology* **2005**, 207, (3), 221-231.
50. A. A. Shvedova; E. R. Kisin; R. Mercer; A. R. Murray; V. J. Johnson; A. I. Potapovich; Y. Y. Tyurina; O. Gorelik; S. Arepalli; D. Schwegler-Berry; A. F. Hubbs; J. Antonini; D. E. Evans; B.-K. Ku; D. Ramsey; A. Maynard; V. E. Kagan; V. Castranova; P. Baron, *American Journal of Physiology - Lung Cellular and Molecular Physiology* **2005**, 289, 698-708.
51. K. Donaldson; R. Aitken; L. Tran; V. Stone; R. Duffin; G. Forrest; A. Alexander, *Toxicological Sciences* **2006**, 92, (1), 5-22.
52. A. A. Shvedova; E. R. Kisin; A. R. Murray; O. Gorelik; S. Arepalli; V. Castranova; S.-H. Young; F. Gao; Y. Y. Tyurina; T. D. Oury; V. E. Kagan, *Toxicology and Applied Pharmacology* **2007**, 221, (3), 339-348.
53. R. Mercer; J. Scabilloni; L. Wang; E. Kisin; A. Murray; S.-B. D; A. Shvedova; V. Castranova, *American Journal of LungCell Molecular Physiology* **2008**, 294, (1), 11.
54. L. A. Mitchell; J. Gao; R. Vander Wall; A. Gigliotti; S. W. Burchiel; J. D. McDonald, *Toxicological sciences* **2007**, 100, (1), 12.
55. D. Lison; J. Muller, *Toxicological Sciences* **2008**, 101, (1), 2.
56. J. McDonald; L. Mitchell, *Toxicological Sciences* **2007**, 101, (1), 2.
57. C. Schöenberger; L. Forró, *Physicsworld* **2000**, 13, (June), 37.
58. O. L. Min-Feng Yu, Mark Dyer, Katerina Moloni, Thomas Kelly, Rodney Ruoff, *Science* **2000**, 287, 4.
59. K. Yada, *Acta Crystallographica Section A* **1971**, 27, (6), 5.
60. J. A. Merchant, *Environmental Health Perspectives* **1990**, 88, 7.
61. I. J. Selikoff, *Environmental Health Perspectives* **1990**, 88, 8.
62. S. H. Constantopoulos, *Regulatory Toxicology and Pharmacology* **2007**.
63. T. W. Hesterberg; G. Chase; C. Axten; W. C. Miller; R. P. Musselman; O. Kamstrup; J. Hadley; C. Morscheidt; D. B. Bernstein; P. Thevanaz, *Toxicology and Applied Pharamcology* **1998**, 151, 14.
64. M. F. Stanton; C. Wrench, *Journal of the National Cancer Institute* **1972**, 48, 24.

65. E. R. Kisin; A. R. Murray; M. J. Keane; X. C. Shi; D. Schwegler-Berry; O. Gorelik; S. Arepalli; V. Castranova; W. E. Wallace; V. E. Kagan; A. Shvedova, A., *Toxicology and Environmental Health* **2007**, 70, (24), 9.
66. H. H. Nelson; K. T. Kelsey, *Oncogene* **2002**, 21, (48), 7284-7288.
67. T. Ollikainen; K. Linnainmaa; V. L. Kinnula, *Environ. Mol. Mutagen.* **1999**, 33, 153-160.
68. Z. Li; T. Hulderman; R. Salmen; R. Chapman; S. S. Leonard; S.-H. Young; A. Shvedova; M. I. Luster; P. P. Simeonova, *Environmental Health Perspectives* **2007**, 115, (3), 377.
69. C.-w. Lam; J. T. James; R. McCluskey; S. Arepalli; R. L. Hunter, *Critical Reviews in Toxicology* **2006**, 36, (3), 189-217.
70. A. D. Maynard; P. A. Baron; M. Foley; A. A. Shvedova; E. R. Kisin; V. Castranova, *Journal of Toxicology and Environmental Health, A* **2004**, 67, (1), 87-107.
71. A. M. Derfus; W. C. W. Chan; S. Bhatia, *Nano Letters* **2004**, 4, (1), 8.
72. S. J. Cho; D. Maysinger; M. Jain; B. Röder; S. Hackbarth; F. M. Winnik, *Langmuir* **2007**, 23, (4), 1974-1980.
73. R. Hardman, *Environmental Health Perspectives* **2006**, 114, (2), 165-172.
74. J. P. Berry; B. Arnoux; G. Stanislas; P. Galle; J. Chretien, *Biomedicine* **1977**, 29, (9-10), 4.
75. P. Jani; G. Halbert; J. Langridge; A. T. Florence, *Journal of Pharm Pharmacol* **1990**, 42, (12), 6.
76. A. Nemmar; P. H. M. Hoet; D. Vanquickenborne; M. Dinsdale; M. F. Thomeer; H. Hoylaerts; L. Vanbilloen; L. Mortelmans; B. Nemery, *Circulation* **2002**, 105, 411-414.
77. S. Taenaka; E. Karg; C. Roth; H. Shultz; A. Ziesenis; U. Heinzmann; P. Schramel, *Environmental health perspectives* **2001**, 109, (4), 5.
78. W. G. Kreyling; M. Semmler; F. Erbe; P. Mayer; S. Takenaka, *Journal of Toxicology and Environmental Health, A* **2002**, 65, (20), 1513-1530.
79. M. Semmler-Behnke; S. Takenaka; S. Fertsch; A. Wenk; J. Seitz; P. Mayer, **2007**, 115, (5), 6.
80. J. Chen; M. Tan; A. Nemmar; W. Song; M. Dong; G. Zhang; Y. Li, *Toxicology* **2006**, 222, (3), 195-201.
81. J. Ladermann; H.-J. Weigmann; C. Rickmeyer; H. Barthelmes; H. Schaefer; G. Muller; W. Sterry, *Skin, Pharmacology and Applied Skin Physiology* **1999**, 12, (5), 10.

82. J. P. Ryman-Rasmussen; J. E. Riviere; N. A. Monteiro-Reviere, *Toxicological Sciences* **2006**, 91, (1), 7.
83. S. S. Tinkle; J. M. Antonini; B. A. Rich; J. R. Roberts; R. Salmen; K. DePree; E. J. Adkins, *Environmental Health Perspectives* **2003**, 111, (9), 1202-1208.
84. H. A. Howe; D. Bodian, *Proc Soc Exp Biol med* **1940**, 43, 4.
85. G. Oberdörster; Z. Sharp; W. Kreyling; C. Cox; V. Atudorei; A. Elder; R. Gelein, *Inhalation Toxicology* **2004**, 16, (7), 437-445.
86. A. Elder; R. Gelein; V. Silva; T. Feikert; L. Opanashuk; J. Carter; R. Potter; A. Maynard; Y. Ito; J. Finkelstein; G. Oberdörster, *Environmental Health Perspectives* **2006**, 114, (8), 1172-1178.
87. I. C. o. R. Protection, *Annals of the ICRP* **1995**, 24, (1-3).
88. B. Asgharian; O. Price, *Inhalation toxicology* **2007**, 19, (13), 10.
89. K. K. Jain, *Neurodegenerative Diseases* **2007**, 4, (4), 5.
90. P. Lockman; R. Mumper; M. Khan; D. Allen, *Drug Dev Ind Pharm* **2002**, 28, (1), 13.
91. E. Sadauskas; H. Wallin; M. Stoltenberg; U. Vogel; P. Doering; A. Larsen; G. Danscher, *Particle and fiber toxicology* **2007**, 4.
92. J. Challier; M. Panigel; E. Meyer, *Int J Nucl Med Biol* **1973**, 1, (2), 4.
93. T. Tsuchiya; I. Oguri; Y. Yamakoshi; N. Miyata, *FEBS Letters* **1996**, 393, (1), 7.
94. B. Ballou; B. C. Lagerholm; L. A. Ernst; M. P. Bruchez; A. S. Waggoner, *Bioconjugate Chemistry* **2003**, 15, (1), 79-86.
95. V. Wiwanitkit; A. Sereemasun; R. Rojanathanes, in: *Fertil Steril*, 2007.
96. S. Yoshida; K. Hiyoshi; T. Ichinose; H. Takano; S. Oshio; I. Sugawara; K. Takeda; T. Shibamoto, in: *Int J Androl*, January 22: 2008.
97. T. Tsuchiya; Y. Yamakoshi; N. Miyata, *Biochem Biophys Res Comm* **1995**, 206, (3), 10.
98. OSHA Medical Screening and Surveillance.
<http://www.osha.gov/SLTC/medicalsurance/index.html> (1/29/08),

99. D. Evans; W. Heitbrink; T. Slavin; T. Peters, *Annals of Occupational Hygiene* **2008**, 52, (1), 13.
100. C. Tsai; E. Ada; M. Ellenbacker; M. Hallock; K. Ahn, in: *CampusHealth Safety and environmental Management*, Boston, 2007.
101. W. Heitbrink; D. Evans; T. Peters; T. Slavin, *Journal of Occupational and Environmental Hygiene* **2007**, 4, (5), 11.
102. D. Brouwer; J. Gijbers; M. Lurvink, *Annals of Occupational Hygiene* **2004**, 48, (5), 15.
103. H. Fissan; S. Neumann; A. Trampe; D. Pui; W. SHin, *Journal of Nanoparticle Research* **2007**, 9, 7.
104. H. C. Wang; G. Kasper, *Journal of Aerosol Science* **1991**, 22, 11.
105. A. Balazy; A. Podgorsky; L. Gradon in: *Filtration of nanosized aerosol particles in fibrous filters. I-Experimental results*, European Aerosol Conference, 2004, 2004; 2004; p 2.
106. E. A. Bal; C. v. Guliji in: *Measurement of aerosol filtration for ultrafine particles*, European Aerosol Conference, 2004; Thermap rebound measurement theory: 2004; pp 979-980.
107. A. Balazy; M. Toivola; T. Reponen; A. Podgorsky; A. Zimmer; S. Grinshpun, *Annals of Occupational Hygiene* **2005**, Baazy 2005 REspirators 12.
108. B. Halford, *Chemical & Engineering News* **2006**, 84, (18), 1.
109. M. Heim; B. Mullins; M. Wild; J. Meyer; G. Kasper, *Aerosol Science and Technology* **2005**, 39, (8), 8.
110. C. S. Kim; L. Bao; K. Okuyama; M. Shimada; H. Niinuma, *Journal of Nanoparticle research* **2006**, 8, (2), 7.
111. D. Japuntich; L. Franklin; D. Pui; T. Kuehen; S. C. Kim; A. Viner, *Journal of Nanoparticle Research* **2007**, 9, 15.
112. J. Wang; D. Chen; D. Pui, *Journal of nanoparticle research* **2007**, 9, 7.
113. S. C. Kim; M. Harrington; D. Pui, *Journal of Nanoparticle Research* **2007**, 9, 9.
114. 3M, *3M Technical Data Bulletin* **2006**, 171, 6.
115. S.-H. Huang; C.-W. Chen; C.-P. Chang; C.-Y. Lai; C.-C. Chen, *Journal of Aerosol Sciences* **2007**, 38, (7), 9.

116. M.-H. Lee; W. McClellan; J. Candela; D. Andrews; P. Biswas, *Journal of Nanoparticle REsearch* **2007**, 9, 10.
117. NIOSH, in: N. I. f. O. S. a. Health, (Ed.) Department of Health and Human Services: 2007.
118. R. Russel; R. Cresanti, in: N. S. a. T. COuncil, (Ed.) 2006.
119. M. Nasterlack; A. Zobor; C. Oberlinner, *Int Arch Occup Environ Health* **2007**, Published online September 12, 2007, 6.