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Genome sequence of *Thermofilum pendens* reveals an exceptional loss of biosynthetic pathways without genome reduction

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Running title: *Thermofilum pendens* genome

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Abstract

We report the complete genome of *Thermofilum pendens*, a deep-branching, hyperthermophilic member of the order Thermoproteales within the archaeal kingdom Crenarchaeota. *T. pendens* is a sulfur-dependent, anaerobic heterotroph isolated from a solfatara in Iceland. It is an extracellular commensal, requiring an extract of *Thermoproteus tenax* for growth, and the genome sequence reveals that biosynthetic pathways for purines, most amino acids, and most cofactors are absent. In fact *T. pendens* has fewer biosynthetic enzymes than obligate intracellular parasites. *T. pendens* does not display other features common among obligate parasites and thus does not appear to be in the process of becoming a parasite. It appears that *T. pendens* has adapted to life in an environment rich in nutrients. *T. pendens* was known to utilize peptides as an energy source, but the genome reveals substantial ability to grow on carbohydrates. *T. pendens* is the first crenarchaeote and only the second archaeon found to have a transporter of the phosphotransferase system. In addition to fermentation, *T. pendens* may gain energy from sulfur reduction with hydrogen and formate as electron donors. It may also be capable of sulfur-independent growth on formate with formate hydrogenlyase. Additional novel features are the presence of a monomethylamine:corrinoid methyltransferase, the first time this enzyme has been found outside of Methanosarcinales, and a presenilin-related protein. Predicted highly expressed proteins do not include housekeeping genes, and instead include ABC transporters for carbohydrates and peptides, and CRISPR-associated proteins.

Introduction

Crenarchaeota is one of the two major divisions of the Archaea, and it is the least well-represented in terms of genome sequences. Only six crenarchaeal complete genomes have been published so far, and three of these are from the genus *Sulfolobus*. Within the order Thermoproteales, only one organism has been completely sequenced so far, *Pyrobaculum aerophilum*, although several more species of *Pyrobaculum*, *Caldivirga maquilingsis*, and *Thermoproteus tenax* are currently being sequenced (23).

Thermofilum pendens represents a deep branch within the order Thermoproteales, and the organism grows only in the presence of a fraction of the polar lipids of *T. tenax* (55), a property that has not been seen before in archaea. Therefore, it was an attractive sequencing target. We report here the genome sequence and analysis of the type strain *Thermofilum pendens* Hrk5.

T. pendens is an anaerobic, sulfur-dependent hyperthermophile isolated from a solfatara in Iceland. It forms long thin filaments and may have an unusual mode of reproduction in which spherical bulges form at one end of the cell. It requires complex media and a lipid extract from the related organism *T. tenax* for growth (55). The unknown lipid may be a cellular component or may make sulfur more available to the cells. Complex media such as tryptone or yeast extract are required for growth, and CO₂ and H₂S are produced, similar to other anaerobic members of the Crenarchaeota and the euryarchaeal family Thermococcaceae. The genome reveals an organism that appears to have lost many biosynthetic capabilities yet does not have a reduced genome size compared to other Crenarchaeota.

Materials and Methods

Frozen *T. pendens* Hrk5 cells were obtained from Dr. Karl Stetter. Cells were resuspended in 0.25 M sucrose in TE buffer. Sodium dodecyl sulfate was added to 1% concentration, and cells were lysed by three cycles of freezing and thawing. Proteinase K was added to 50 µg/ml, and the lysate was incubated at 60°C for 30 minutes. Undigested proteins were precipitated by addition of NaCl to 0.5 M concentration and removed by centrifugation. Nucleic acids in the supernatant were precipitated by the addition of an equal volume of cold isopropanol and collected by centrifugation. After digestion with RNase A, the DNA was purified by successive extractions with phenol and phenol:chloroform and recovered by ethanol precipitation. DNA was resuspended in TE buffer and sent to the Joint Genome Institute.

The genome of *T. pendens* was sequenced at the Joint Genome Institute (JGI) using a combination of 3kb, 6kb and 40kb (fosmid) DNA libraries. All general aspects of library construction and sequencing performed at the JGI can be found at <http://www.jgi.doe.gov/>. Draft assemblies were based on 21478 total reads. All three libraries provided 11x coverage of the genome. The Phred/Phrap/Consed software package (www.phrap.com) was used for sequence assembly and quality assessment (5-7). After the shotgun stage, reads were assembled with parallel phrap (High Performance Software, LLC). Possible mis-assemblies were corrected with Dupfinisher (11) or transposon bombing of bridging clones (Epicentre Biotechnologies, Madison, WI). Gaps between contigs were closed by editing in Consed, custom primer walk or PCR amplification (Roche Applied Science, Indianapolis, IN). A total of 465 additional reactions were necessary to close gaps and to raise the quality of the finished sequence.

The sequences of *T. pendens*, consisting of one chromosome and one plasmid, can be accessed using the GenBank accession numbers [NC_008698](#) and [NC_008696](#) and the Genomes On Line Database accession number Gc00473. Genes were identified using a combination of Critica (2) and Glimmer (4) followed by a round of manual curation.

Analysis of the *T. pendens* genome was carried out with the Integrated Microbial Genomes (IMG) system (27). Protein families unique to *T. pendens* or missing from *T. pendens* but present in other Crenarchaeota were identified with the phylogenetic profiler in IMG. Analysis of signal transduction was carried out using the MiST database (51). A cumulative GC-skew plot was generated with a 35 kb sliding window using GraphDNA (48).

Predicted highly expressed (PHX) genes were determined with the EMBOSS (38) programs *cus* and *cai*. The training set of PHX genes was compiled from Karlin et al. (16). The genes with a codon adaptation index (CAI) in the top 5% were taken to be PHX genes.

Results

General features

The genome of *T. pendens* Hrk5 consists of a circular chromosome of 1.78 Mbp and a plasmid of 31,504 bp (Table 1). The G+C percentage is 58%, higher than that of other Crenarchaeota. 1,923 genes were identified, of which 1,883 encode proteins. The percentage of the genome devoted to encoding genes is 91%, slightly higher than for other sequenced Crenarchaeota. About 59% of protein-coding genes begin with an AUG codon, 32% with a GUG, and 10% with UUG. About 66% of protein-coding genes have

COG domains, and about 63% have Pfam domains, similar to other archaeal genomes. There is one copy of each ribosomal RNA. *T. pendens* has the highest percentage of fusion genes among the Crenarchaeota. Several proteins in *T. pendens* have not been found in Crenarchaeotes or in archaea before (Table 2) and several proteins found in all other crenarchaeota are missing from the *T. pendens* genome (Table 3).

The plasmid is predicted to encode 52 proteins, of which only two have similarity to proteins in the GenBank nonredundant protein database. Tpen_1849 is similar to a *T. pendens* chromosomal protein of unknown function (Tpen_0735), and Tpen_1875 is a predicted helicase. In addition, Tpen_1891 is predicted to be a site-specific recombinase (COG4974). The function of the plasmid and whether or not it is beneficial to the host is currently unknown.

Cumulative GC-skew analysis of the *T. pendens* genome was used to identify the potential origin(s) of replication (9). A global minimum was located at position 488,884 which is near a 478 bp intergenic region between positions 487,890 and 488,368. The intergenic spacer contains several repetitive sequences similar to conserved crenarchaeal origin recognition boxes (39).

No repetitive elements were found when the ISfinder database (46) was searched with the *T. pendens* coding sequences. However, within a 100,000 bp section of the genome there are twelve stretches of clustered regularly interspaced short palindromic repeat (CRISPR) elements interspersed with protein-coding genes. Among the 97 predicted highly expressed (PHX) genes (supplementary table 1) there are 6 CRISPR-associated proteins (Tpen_1263, Tpen_1287, Tpen_1288, Tpen_1316, Tpen_1342, Tpen_1356). Interestingly, a group of seven consecutive genes (Tpen_1287-Tpen_1293) including two

CRISPR-associated proteins is PHX. While other thermophilic archaea have similar numbers of CRISPR-associated proteins within their genomes, most do not have CRISPR-associated proteins as PHX genes, although *Staphylothermus marinus* and *P. aerophilum* do (4 and 2 genes respectively). However *T. pendens* has the highest number of CRISPR-associated genes as PHX genes, and the highest percentage. Thus, protection against viral infection appears to be a major priority for *T. pendens*. Crenarchaeota from hot spring environments are known to host a wide variety of viruses with distinctive morphologies (reviewed in 35).

Central metabolism

T. pendens contains complete glycolysis and gluconeogenesis pathways.

Glyceraldehyde 3-phosphate:ferredoxin oxidoreductase, found in some archaeal hyperthermophiles as an alternative step in glycolysis (30), is not present in *T. pendens*. Phosphoenolpyruvate (PEP) synthase, used as the last step in glycolysis in *Thermococcus kodakaraensis* (14), is present in *T. pendens* (Tpen_0588). PEP synthase could be involved in glycolysis and/or in gluconeogenesis. Starch synthesis and utilization pathways are also present.

Pentoses are synthesized through the ribulose monophosphate pathway common in archaea (reviewed in 17). *T. pendens* encodes two ribose 5-phosphate isomerases, one RpiA-type (Tpen_0327) and one RpiB-type (Tpen_1241). This is the first time an RpiB has been found in archaea. The RpiB is adjacent to uridine phosphorylase (Tpen_1240) suggesting a function in nucleoside utilization. Under conditions in which ribonucleosides are present in excess, RpiB may be involved in conversion of ribose

phosphate to hexoses through the ribulose monophosphate pathway, a reversal of the pathway from its predicted function in archaea.

ATP can be generated from pyruvate through the consecutive action of pyruvate:ferredoxin oxidoreductase (PFOR) and ADP-forming acetyl-CoA synthase similar to Thermococcales (26). The *T. pendens* PFOR (Tpen_0571-0574) is similar to the characterized *Thermotoga maritima* enzyme (18). In Thermococcales and Crenarchaeota, ADP-forming acetyl-CoA synthase is split into two subunits, alpha and beta. *T. pendens* contains one alpha subunit (Tpen_0336), one beta subunit (Tpen_0109), and one protein with both alpha and beta subunits fused together (Tpen_0602). Two AMP-forming acyl-CoA synthases are also present (Tpen_0893, Tpen_1611). *T. pendens* has four other enzymes similar to pyruvate:ferredoxin oxidoreductase (Tpen_0540-0543, Tpen_0781-0782, Tpen_0856-0857, Tpen_1455-1456), and these are likely to be involved in amino acid degradation pathways in which the amino acid is first converted to the 2-ketoacid, then to the acyl-CoA, and finally to an acid, with ATP generated by acyl-CoA synthases (26). Four aldehyde:ferredoxin oxidoreductases are also present (Tpen_0094, Tpen_0176, Tpen_1413, Tpen_1817), and these could be involved in peptide fermentation (1). The 2-oxoacid oxidoreductases produce aldehydes which are then converted to acids. Reduced ferredoxin is produced but there is no ATP production by this pathway.

T. pendens appears to assimilate glycerol. There is a glycerol kinase (Tpen_1128) adjacent to subunit A of glycerol 3-phosphate dehydrogenase (Tpen_1127). Next to these are three genes with similarity to subunits B, C, and D of succinate dehydrogenases (Tpen_1124-1126). A gene encoding subunit A of succinate dehydrogenase is not found

within the genome. It appears that the three succinate dehydrogenase-related subunits along with the glycerol 3-phosphate dehydrogenase subunit A may form a novel glycerol 3-phosphate dehydrogenase that may transfer electrons to a quinone or other acceptor.

Unlike most of the sequenced Crenarchaeota, *T. pendens* has ribulose 1,5-bisphosphate carboxylase (Rubisco, Tpen_1227). It also has the recently discovered enzymes involved in conversion of the ribose phosphate group of AMP to ribulose 1,5-bisphosphate (43): AMP phosphorylase (Tpen_0093) and ribose-1,5-bisphosphate isomerase (Tpen_0384). Under conditions in which acetate is incorporated into the gluconeogenesis pathway, the AMP-forming acetyl-CoA synthetase and phosphoenolpyruvate synthase could produce substantial AMP. The *T. pendens* AMP-forming acetyl-CoA synthetase (Tpen_0893) has very high similarity to the *P. aerophilum* characterized enzyme (3). A large amount of AMP may also be generated by ribose-phosphate pyrophosphokinase, required for pyrimidine synthesis, and phosphoribosyltransferases.

Biosynthesis

T. pendens is known to require an extract of *T. tenax* for growth. While the specific compound required from *T. tenax* could not be identified, the genome reveals a vast reduction in its ability to synthesize basic metabolites. *T. pendens* appears to be dependent on its environment for purines, most cofactors, and most amino acids. A list of 125 COGs involved in synthesis of nucleobases, amino acids, and cofactors was compiled (supplementary table 2). COGs encoding archaeal biosynthetic enzymes were included where they are known. The presence of these COGs in all complete bacterial and archaeal genomes was determined using the function profile feature in IMG. *T.*

pendens possesses only 11 of these COGs. The only organisms with fewer of this COG set were obligate parasites or commensals. In fact some obligate parasites, such as *Rickettsia* species, have greater biosynthetic capabilities than *T. pendens*.

While it is possible that *T. pendens* has different pathways for metabolite synthesis or has many enzymes replaced through nonorthologous gene displacement, this is unlikely to account for the lack of biosynthetic enzymes because other Crenarchaeota have recognized pathways for basic metabolites. For example, all Crenarchaeota except for *T. pendens* have homologs of the pyridoxine biosynthesis genes PDX1 and PDX2 (*yaaD* and *yaaE* in *Bacillus subtilis*) and the bifunctional coenzyme A biosynthetic enzyme phosphopantothenoylcysteine synthetase/decarboxylase. Table 3 lists the COGs missing from *T. pendens* that are found in all other sequenced Crenarchaeota. The top nine of these COGs are involved in pyridoxine, coenzyme A, riboflavin, ubiquinone, and thiamine biosynthesis. In addition, most Crenarchaeota have homologs of several heme biosynthetic enzymes, but *T. pendens* lacks these. They are not found in Table 3 because they are also missing from the *Staphylothermus marinus* genome. Also COG1731, archaeal riboflavin synthase, is not found in Table 3 because it is missing from both *T. pendens* and *Cenarchaeum symbiosum*; however, *C. symbiosum* has the bacterial-type riboflavin synthase (COG0307) but *T. pendens* lacks both the bacterial and archaeal enzymes.

In accordance with the predicted lack of biosynthetic capacity, *T. pendens* is the only Crenarchaeote to have a bioY family biotin transporter and a riboflavin transporter (see Table 2). In addition *T. pendens* has an expansion of ABC transporters related to those involved in cobalt uptake. While most Crenarchaeota have 0-2 representatives from this

family, *T. pendens* has 7. One of these transporters has an additional membrane protein related to *B. subtilis* YkoE, and such transporters are predicted to transport the thiamine precursor hydroxymethylpyrimidine (40).

T. pendens has genes for limited amino acid synthesis. There is a putative cysteine synthase (Tpen_1605) related to a characterized *A. pernix* enzyme (34), but no serine acetyltransferase. Probably, like *A. pernix*, *T. pendens* uses O-phosphoserine rather than O-acetylserine as the intermediate in cysteine synthesis. Cysteine synthesis may have been preserved in *T. pendens* so that cysteine can help to protect the cell against oxidative stress, as is thought to occur in some parasitic protists (reviewed in 31).

Glutamine can be synthesized from glutamate on its tRNA (Tpen_0360-0361), and also by a cytosolic glutamine synthase (Tpen_1089). Cytosolic glutamine synthesis has probably been preserved for its role as a nitrogen donor. *T. pendens* has six proteins with glutamine amidotransferase domains including CTP synthase (Tpen_1163) and glucosamine 6-phosphate synthetase (Tpen_0085, Tpen_1094). Asparagine can be synthesized by a tRNA synthetase-related, archaeal asparagine synthetase (Tpen_1140; 41).

T. pendens has a methionine synthase (Tpen_1819) but no homoserine biosynthesis genes, thus it can probably not make methionine *de novo*, but it can recycle homoserine resulting from S-adenosylmethionine-dependent methylation reactions. Interestingly *T. pendens* has genes related to monomethylamine and trimethylamine methyltransferases from Methanosarcinales (Tpen_1211, Tpen_1467). The *T. pendens* monomethylamine methyltransferase is related to the Methanosarcina enzymes, and this is the first time this protein family has been found outside the Methanosarcinales. Both putative

methyltransferases are adjacent to corrinoid proteins (Tpen_1212, Tpen_1468), supporting their function as methyltransferases. Where the *Methanosarcina* proteins have pyrrolysine residues, *T. pendens* has leucine in both proteins. The methyl groups transferred from methylamines could be used to recycle methionine after methylation reactions.

T. pendens can synthesize pyrimidines *de novo* but not purines. Carbamoyl phosphate for pyrimidine synthesis is generated by carbamate kinase (Tpen_0172), not by carbamoyl phosphate synthase, similar to *P. furiosus* (52). There are a variety of phosphorylases and phosphoribosyltransferases that could be used for salvage of bases. In addition *T. pendens* has an ORF (Tpen_1649) with 66% similarity to *A. pernix* APE0012, which is a broad-range nucleoside kinase as well as a phosphofructokinase (12), thus nucleosides may also be salvaged. No transporters belonging to known families of nucleobase or nucleoside transporters could be identified in the genome.

T. pendens appears to be able to synthesize phospholipids *de novo*. It may have a modified mevalonate pathway as predicted for *Methanocaldococcus jannaschii* (Grochowski et al., 2006) as it has a homolog of the MJ0044 protein which was shown to be an isopentyl phosphate kinase (Tpen_0607). It has the enzymes for synthesis of sn-glycerol 1-phosphate (Tpen_1231) and geranylgeranyl diphosphate (Tpen_0606) and for attaching the geranylgeranyl groups to glycerol 1-phosphate (Tpen0633, Tpen_0636, Tpen_1449). Like many archaea it has only one identifiable CDP-alcohol phosphatidyltransferase (Tpen_0218), and this is most closely related to archaetidylinositol synthases. Myo-inositol-1-phosphate synthase is present (Tpen_1660). It is unknown whether *T. pendens* makes additional phospholipids.

Carbohydrate metabolism and transport

T. pendens requires a complex growth medium such as yeast extract, tryptone, or gelatin, and sucrose stimulates growth (55). It was concluded that *T. pendens* grows mainly by peptide fermentation. While *T. pendens* does have enzymes for amino acid degradation, the genome reveals that sugars and sugar polymers may also be important growth substrates for this organism.

One source of evidence that carbohydrates are important growth substrates is the set of transporters encoded in the genome. *T. pendens* encodes 8 ABC transporters of family 1, which are involved in sugar uptake (Tpen_1055-1057, Tpen_1149-1152, Tpen_1174-1177, Tpen_1255-1257, Tpen_1451-1453, Tpen_1547-1550, Tpen_1588-1590, Tpen_1617-1619). Within the archaea, only *Haloarcula marismortui* possesses as many family 1 ABC transporters. *T. pendens* also has one ABC transporter from family 2, likely to be involved in sugar uptake (Tpen_1208-1210). The only other family 2 ABC transporter in the archaea is in *Sulfolobus acidocaldarius*. One of the four family 5 ABC transporters in *T. pendens* (Tpen_1676-1680) is similar to a *P. furiosus* cellobiose transporter (19) and a *T. maritima* transporter for mannobiose (TM1223; 32). Two members of the glycoside-pentoside-hexuronide (GPH): cation symporter family are also present (Tpen_1599, Tpen_1831).

T. pendens is the only sequenced crenarchaeote to have the phosphotransferase system (PTS) for carbohydrate uptake (see Table 2). The only other sequenced archaeon to have a PTS transporter is *H. marismortui*. *Haloquadratum walsbyi* has Enzyme I and HPr proteins from the PTS, but it does not have identifiable PTS transporters. A phylogenetic tree of Enzyme I shows that the *T. pendens* and halophile proteins are not closely related,

suggesting they were independently acquired through separate lateral transfer events (not shown). *T. pendens* Enzyme I is adjacent to a predicted N-acetylglucosamine 6-phosphate deacetylase, suggesting that N-acetylglucosamine may be the substrate for this transporter.

T. pendens has a set of 15 glycosyl hydrolases, about the same number as *Sulfolobus* species, and greater than other Crenarchaeota. There are several genes involved in starch utilization. One cluster of genes encoding two glycosyl hydrolases and an ABC transporter (Tpen_1451-1454, 1458) is similar to a cluster from *Thermococcus* sp. B1001 involved in extracellular formation of cyclomaltodextrins, transport of cyclomaltodextrins into the cell, and intracellular degradation of the cyclomaltodextrins (13). In addition, there is an alpha-glucosidase (Tpen_1511) similar to the characterized NAD⁺-dependent *T. maritima* enzyme (37).

Cellulose may also be utilized by *T. pendens*. There is a secreted family 12 glycosyl hydrolase (Tpen_1681) with weak similarity to cellulases as well as an ABC transporter with high similarity to a characterized cellobiose transporter from *P. furiosus* (Tpen_1676-1680; 19). Cellobiose and larger oligosaccharides may be broken down by an intracellular beta-glucosidase (Tpen_1494).

Sucrose stimulates growth of *T. pendens* but does not serve as the sole energy source (55). The enzymes involved in sucrose metabolism can not be identified from the genome sequence. No beta-fructofuranosidase (invertase) or sucrose phosphorylase can be identified, and there is no homolog of PF0132, which encodes the invertase purified from *P. furiosus* (22).

Three glycosidases (Tpen_1511, Tpen_1269, Tpen_1458) and three ABC transporter-associated sugar-binding proteins (Tpen_1055, Tpen_1208, Tpen_1257) are among the PHX genes of *T. pendens*, providing further evidence of the importance of carbohydrate metabolism. Subunits of two peptide ABC transporters are also PHX genes (Tpen_1635-1636, Tpen_1638, Tpen_1245, Tpen_1247-1249). This reflects the need of *T. pendens* to obtain many amino acids from external sources and the utilization of peptides for energy.

Electron transport

T. pendens requires sulfur for growth and produces H₂S, and some of the potential catalysts for this metabolism can be identified in the genome sequence. *T. pendens* does not have a hydrogenase related to sulfhydrogenase and hydrogenase II of *P. furiosus*, which reduce sulfur as well as protons (25). It also does not possess a sulfide dehydrogenase (24). There is a homolog (Tpen_0143, 48% identity, 66% similarity) of the recently identified CoA-dependent NADPH:sulfur oxidoreductase from *P. furiosus* (45). However *T. pendens* does not have the *mbx* protein complex that is predicted to transfer electrons from ferredoxin to NADPH. Also *T. pendens* does not have a homolog of bacterial ferredoxin-NADP⁺ reductases (COG1018), so the pathway for recycling ferredoxin is unknown. *T. pendens* has a large set of adjacent genes (Tpen_1070-Tpen_1088) with similarity to NADH dehydrogenases and membrane bound hydrogenases. This cluster may encode one or more multisubunit enzymes that oxidize ferredoxin and transfer the electrons to NADP, a quinone, or another electron carrier.

T. pendens has an operon (Tpen_1121-Tpen_1123) similar to the *psrABC* genes of the polysulfide reductase from *Wolinella succinogenes* (20). The protein similarity is weak, but the three proteins in the *T. pendens* operon belong to the same protein families as the

polysulfide reductase subunits. The A subunit is predicted by ProSite to have a twin-arginine signal peptide, so the enzyme probably reduces its substrate extracellularly.

A substantial amount of formate may be produced by fermentative organisms in the environments in which *T. pendens* lives, and *T. pendens* appears to have two pathways for utilizing formate. Like *P. aerophilum* and *Hyperthermus butylicus*, *T. pendens* has a three-subunit, membrane-bound, molybdopterin-dependent formate dehydrogenase. The alpha subunit has a predicted twin arginine signal peptide, so the topology of the enzyme is likely to be similar to the solved structure of *E. coli* formate dehydrogenase N with formate oxidation occurring outside the cell (reviewed in 15). This enzyme likely channels electrons from formate to a quinone or other carrier and then to sulfur as the final electron acceptor.

T. pendens is the only Crenarchaeote to have a formate transporter (Tpen_0191). The transporter is found adjacent to a putative operon (Tpen_0190-Tpen_0178) with high similarity to *E. coli* hydrogenase 4. In *E. coli*, hydrogenase 4 forms part of the formate hydrogenlyase complex which oxidizes formate and produces hydrogen under conditions in which no electron acceptors other than protons are present. The *T. pendens* operon contains a formate dehydrogenase alpha subunit, providing strong evidence that this operon encodes formate hydrogenlyase. The formate dehydrogenase protein does not have a signal peptide, suggesting that formate oxidation occurs in the cytoplasm as for the *E. coli* complex (reviewed in 44). Formate hydrogenlyase contributes to the generation of a proton gradient in two ways: by using protons from inside the cell to make H₂, which then diffuses out of the cell, and under some conditions by pumping protons out of the cell (Hakobyan et al., 2005). This enzyme complex is expressed in *E.*

coli only when no electron acceptors are present, suggesting that *T. pendens* may use this pathway when sulfur is scarce.

T. pendens may also use hydrogen as an electron donor as it contains genes (Tpen_0591-0594) similar to the four subunits of a membrane-bound uptake hydrogenase from *Acidianus ambivalens* (21). The *A. ambivalens* hydrogenase is predicted to use a quinone to transfer electrons from hydrogen to sulfur. This type of pathway is common among archaeal autotrophs, and in *T. pendens* it may supplement the energy derived from peptides and sugars.

Signal transduction

Archaea have significantly fewer signal transduction systems than bacteria. On average, 2.63% of archaeal proteomes and 5.4% of bacterial proteomes consist of signal transduction proteins (51). Moreover, it has been shown previously that archaeal signal transduction utilizes a substantially reduced repertoire of sensory (input) and regulatory (output) domains (50). The median level of archaeal, one-component systems per genome is roughly 50 times greater than that of two-component systems, and the majority of these systems regulate gene expression at the transcriptional level (51). Two-component systems have only been found in Euryarchaeota and appear to have been laterally transferred from bacteria. In general, crenarchaeal species have fewer signal transduction systems (only 0.7% of the proteome) than Euryarchaeotes.

The *T. pendens* genome contains 45 one-component systems – regulatory proteins that contain one or more sensory domains (50) – that comprise 2.4% of its proteome. This percentage is over three times as high as the average for Crenarchaeota. Thirty-one (69%) of these are located in operons containing predominantly enzymatic genes and are

predicted to regulate their transcription in response to environmental and intracellular signals. Interestingly, *T. pendens* possesses three families of transcription regulators that have not previously been found in Crenarchaeota (Table 2). *T. pendens* contains more PadR domains than any other Crenarchaeal species, possibly indicating a high level of phenolic acid metabolism. Unlike all other Crenarchaeotes, *T. pendens* does not have a member of the *fur* family, which is responsible for metal-ion uptake, although it does possess an iron-dependent repressor (Tpen_0973), which is positioned beside an iron transporter.

Presenilin

A protein belonging to the presenilin family is present in the *T. pendens* genome (Tpen_0870). In eukaryotes presenilin is an integral membrane protease and part of the gamma-secretase complex (54). Mutations in presenilin cause it to cut amyloid precursor peptide (APP) in a different place and generate APP forms that are more likely to aggregate and form plaques. A family of proteins weakly related to presenilins, known as presenilin homologs, has been identified in eukaryotes and archaea (8, 36). The *T. pendens* protein and a related protein from *P. aerophilum* are not closely related to these presenilin homologs; they represent a new subfamily of presenilins. These crenarchaeal proteins are about 150 amino acids shorter than the mammalian presenilins, lacking hydrophilic regions at the N-terminus and in an internal loop (Figure 1). They contain 7-9 predicted transmembrane helices and the conserved YD, LGXGD, and PALP motifs. The gamma-secretase complex includes three other proteins, but none of these are present in *T. pendens* or *P. aerophilum*. Characterization of this new subfamily of presenilins may shed light on the structure and function of the eukaryotic proteins.

Discussion

The genome sequence of *T. pendens* shows a loss of biosynthetic pathways to a degree that it is no longer a free-living organism, and has become a commensal, dependent on another archaeon. Along with this lack of biosynthesis, several nutrient transporters that are not found in any other crenarchaeote are present in *T. pendens*. A lack of biosynthetic capability and an increase in nutrient transport capability are features commonly found in obligate parasites (42). However *T. pendens* lacks other features of obligate intracellular parasites, such as genome size reduction (33, 53), loss of signal transduction and DNA repair proteins (29), increased percentage of A-T base pairs (28, 53), and decreased number of fusion proteins (Mavromatis, K. and Kyrpides, N. C., unpublished results). It is hypothesized that one reason genome size reduction in intracellular bacteria occurs because there is no possibility for lateral gene transfer from other bacteria (47), however *T. pendens* does have the opportunity to be exposed to DNA of other bacteria and archaea, and this may help to explain its maintenance of a normal genome size. Also since *T. pendens* is an extracellular rather than intracellular symbiont, it may require a larger genome to deal with environmental perturbations. *T. pendens* does not appear to be parasitic, as it is not known to cause harm to another organism. However, it is limited to growth in nutrient-rich environments, to the point of depending on a specific organism for an essential nutrient. This type of dependence may be one reason why many microbes are not able to be cultivated.

Predicted highly expressed genes in archaea are generally found to be housekeeping genes (16), however this is not the case in *T. pendens*. Surprisingly *T. pendens* PHX genes contain many CRISPR associated genes and ABC transporters for carbohydrates

and peptides. These findings suggest that *T. pendens* is constantly under attack from viruses in its environment. The large number of CRISPR elements also supports this conclusion. The presence of peptide ABC transporters as PHX genes suggests that *T. pendens* places a higher priority on nutrient acquisition than on maximization of cell growth and division, which is in agreement with its lack of biosynthetic pathways for most amino acids and cofactors.

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Table 1. General statistics

Chromosome size (bp)	1781889
Chromosome G+C (bp)	1027538 (57.6%)
Plasmid size (bp)	31504
Plasmid G+C (bp)	17813 (56.5%)
Total genome size (bp)	1813393
Total genome G+C (bp)	1045351 (57.6%)
Total genes	1923
RNA genes	40 (2.1%)
Protein-coding genes	1883 (97.9%)
Genes with function prediction	1170 (60.8%)
Genes in ortholog clusters	1541 (80.1%)
Genes in paralog clusters	805 (41.9%)
Genes assigned to COGs	1264 (65.7%)
Genes assigned Pfam domains	1209 (62.9%)
Genes with signal peptides	134 (7.0%)
Genes with transmembrane helices	437 (22.7%)
Fusion genes	79 (4.11%)

Table 2. Unique genes in *T. pendens* with COG hits

CDS	COG	Function
COGs not found in any other sequenced Archaea		
Tpen_1241	0698	Ribose 5-phosphate isomerase rpiB
Tpen_1297	3525	Glycosyl hydrolase family 20
Tpen_1097	3444	Phosphotransferase system IIB subunit
Tpen_1100	3715	Phosphotransferase system IIC subunit
Tpen_1100	3716	Phosphotransferase system IID subunit
Tpen_1090	4821	Phosphosugar binding protein, SIS domain
COGs not found in any other sequenced Crenarchaeota		
Tpen_1155	1554	Glycoside hydrolase family 65
Tpen_1624	3836	2-Dehydro-3-deoxyglucarate aldolase
Tpen_0948	0207	Thymidylate synthase
Tpen_0017	3613	Nucleoside 2-deoxyribosyltransferase
Tpen_1467	5598	Trimethylamine:corrinoid methyltransferase
Tpen_1211		Monomethylamine:corrinoid methyltransferase
Tpen_1092	1080	Phosphoenolpyruvate-protein kinase (Enzyme I of PTS)
Tpen_1091	1925	Phosphotransferase system HPr protein
Tpen_1098	2893	Phosphotransferase system IIA component
Tpen_1491	1268	Biotin transporter bioY
Tpen_0929	3601	Riboflavin transporter
Tpen_0191	2116	Formate transporter
Tpen_1479	2060	Potassium-transporting ATPase, A chain
Tpen_1480	2216	Potassium-transporting ATPase, B chain
Tpen_1481	2156	Potassium-transporting ATPase, c chain
Tpen_0197	0474	Cation transport ATPase (P-type ATPase)
Tpen_1427		
Tpen_1010	1327	Predicted transcriptional regulator, Zn ribbon and ATP-cone
Tpen_1048	1510	Predicted transcriptional regulator
Tpen_0270	4190	Predicted transcriptional regulator
Tpen_0889	2150	Predicted regulator of amino acid metabolism, ACT domain
Tpen_0253	2229	Predicted GTPase
Tpen_1457	1773	Rubredoxin
Tpen_1536	1811	Uncharacterized membrane protein, DUF554
Tpen_0198	2047	Uncharacterized protein, ATP-grasp superfamily
Tpen_0838	2164	Uncharacterized conserved protein, DUF369
Tpen_1835		
Tpen_1118	2908	Uncharacterized conserved protein
Tpen_0381	3863	Uncharacterized relative of cell wall-associated hydrolases
Tpen_1090	4821	Uncharacterized protein with phosphosugar binding domain

Table 3. Genes present in all Crenarchaeota except *T. pendens* with COG hits

COG	Function
0214	Pyridoxine biosynthesis enzyme (yaaD)
0311	Glutamine amidotransferase involved in pyridoxine synthesis (yaaE)
0413	Ketopantoate hydroxymethyltransferase
0452	Phosphopantothenoylcysteine synthetase/decarboxylase
0108	3,4-dihydroxy-2-butanone 4-phosphate synthase
1985	Pyrimidine reductase, riboflavin biosynthesis
0054	Riboflavin synthase beta chain
0163	3-polyprenyl-4-hydroxybenzoate decarboxylase
1635	Flavoprotein involved in thiazole biosynthesis
0112	Glycine/serine hydroxymethyltransferase
0189	Glutathione synthase/Ribosomal protein S6P modification enzyme/L-2-aminoadipate N-acetyltransferase
0105	Nucleoside diphosphate kinase
2046	Sulfate adenylyltransferase
1650	Uncharacterized protein conserved in archaea
1701	Uncharacterized protein conserved in archaea

Figure 1. Alignment of presenilins from human and Thermoproteales. Alignment was carried out with Clustal W (49) and shaded with GeneDoc (<http://www.nrbsc.org/gfx/genedoc/index.html>).