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Journal

Microbiology Resource Announcements, 5(41)

ISSN

2576-098X

Authors

Parales, Rebecca E
Navara, Rachita
Gettys, Rebecca
et al.

Publication Date

2017-10-12

DOI

10.1128/genomea.01169-17

Peer reviewed



Genome Sequence of *Pseudomonas putida* Strain ASAD, an Acetylsalicylic Acid-Degrading Bacterium

Rebecca E. Parales,^a Rachita Navara,^b Rebecca Gettys,^b Jean J. Huang^b

Department of Microbiology and Molecular Genetics, University of California, Davis, California, USA^a; Franklin W. Olin College of Engineering, Needham, Massachusetts, USA^b

ABSTRACT *Pseudomonas putida* strain ASAD was isolated from compost because of its ability to utilize aspirin (acetylsalicylic acid) as a carbon and energy source. We report the draft genome sequence of strain ASAD, with an estimated length of 6.9 Mb. Study of this isolate will provide insight into the aspirin biodegradation pathway.

Pharmaceutical chemicals are frequently detected in municipal sewage, and only some are efficiently degraded during wastewater treatment (1, 2). In one study, of 18 pharmaceuticals measured, aspirin (acetylsalicylic acid) was most abundant in the influent, and it was efficiently degraded during standard primary and secondary treatment (2). To date, however, only one pure culture has been reported to degrade aspirin (3, 4). We isolated *Pseudomonas putida* strain ASAD from compost soil using minimal medium (5) containing acetylsalicylic acid as the sole carbon and energy source. *P. putida* ASAD grew on acetylsalicylic acid with a doubling time of 1.8 h, and preliminary analyses demonstrated that cells are chemotactic to acetylsalicylic acid. Study of this strain can provide insight into how this degradation process occurs and enable exploration of the relationship between degradation and chemotaxis in bioremediation processes.

Genomic DNA was extracted from *P. putida* ASAD using the PowerLyzer PowerSoil DNA isolation kit (Mo Bio) and sequenced to 40× coverage by an Illumina MiSeq instrument at Molecular Research DNA (Shallowater, TX). The genome sequence of *P. putida* ASAD consisted of 6,963,567 bp, with an average GC content of 60.19%. Of the 6,346 predicted genes in the genome, 97% encode proteins of which 79% have a predicted function (6).

Acinetobacter lwoffii NCIB 10553 was reported to degrade acetylsalicylic acid using an intracellular esterase that generates acetate and salicylic acid (3, 4), but the gene encoding this enzyme has not been identified. The genome of *P. putida* ASAD contains five annotated acetyl esterases (pput 00474, 00825, 01806, 01976, and 06025), which may be involved in aspirin degradation. In contrast, the genome of *P. putida* F1 (a strain that cannot grow on acetylsalicylic acid) does not carry any acetyl esterase genes. Within the *P. putida* ASAD genome, one salicylate hydroxylase gene (EC 1.14.13.1, GenBank locus OMQ39226, pput 01717) is present. Salicylate hydroxylase catalyzes the formation of catechol from salicylic acid; catechol is then cleaved by a dioxygenase (7). In the same gene cluster encoding the salicylate hydroxylase are genes predicted to encode an aromatic acid transporter, a catechol 1,2-dioxygenase (*catA*), an acyl-CoA reductase, and aryl alcohol dehydrogenase. This putative operon is conserved in the genomes of 3 out of 64 *P. putida* strains examined. At an additional locus, the *P. putida* ASAD genome also carries genes that comprise the β -keto adipate pathway (8), including a second copy of *catA* that is associated with *catB* and *catC*, as well as *pcaD*, *pcaJ*,

Received 15 September 2017 Accepted 18 September 2017 Published 12 October 2017

Citation Parales RE, Navara R, Gettys R, Huang JJ. 2017. Genome sequence of *Pseudomonas putida* strain ASAD, an acetylsalicylic acid-degrading bacterium. Genome Announc 5:e01169-17. <https://doi.org/10.1128/genomeA.01169-17>.

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Address correspondence to Jean J. Huang, jean.huang@olin.edu.

and *pcaF*, which together encode the conversion of catechol to TCA cycle intermediates.

Study of *P. putida* ASAD can determine how it senses acetylsalicylic acid, the enzymes involved in degradation of this compound, and whether these processes are regulated. Strain ASAD has 26 predicted methyl-accepting chemotaxis protein (MCP)-encoding genes. Several studies of pollutant-degrading bacteria such as *P. putida* strains F1 and G7 have shown that genes for chemotaxis to pollutant compounds that are degraded by the bacteria are coordinately regulated with genes for degradation (9). Further study of this strain will provide insight into the sensing and degradation capabilities of *P. putida* ASAD.

Accession number(s). This whole-genome shotgun project has been deposited in GenBank under the accession no. [MNAH00000000](https://ncbi.nlm.nih.gov/nucl/MNAH00000000). The version described in this paper is the first version, MNAH01000000. The genome has also been deposited and annotated in the Joint Genome Institute Integrated Microbial Genomes portal (<https://img.jgi.doe.gov/cgi-bin/m/main.cgi>) with genome ID 2576861821.

ACKNOWLEDGMENTS

This research was supported by Olin College. We thank the staff at the Wellesley College greenhouse for allowing R. Navara to obtain a compost sample and the spring 2010 microbial diversity course at Olin College for feedback in development of this work.

REFERENCES

1. Nakada N, Tanishima T, Shinohara H, Kiri K, Takada H. 2006. Pharmaceutical chemicals and endocrine disruptors in municipal wastewater in Tokyo and their removal during activated sludge treatment. *Water Res* 40:3297–3303. <https://doi.org/10.1016/j.watres.2006.06.039>.
2. Deblonde T, Cossu-Leguille C, Hartemann P. 2011. Emerging pollutants in wastewater: a review of the literature. *Int J Hyg Environ Health* 214:442–448. <https://doi.org/10.1016/j.ijheh.2011.08.002>.
3. Grant DJW, de Szöcs JD, Wilson JV. 1970. Utilization of acetylsalicylic acid as sole carbon source and the induction of its enzymatic hydrolysis by an isolated strain of *Acinetobacter lwoffii*. *J Pharm Pharmacol* 22:461–464. <https://doi.org/10.1111/j.2042-7158.1970.tb08564.x>.
4. Grant DJ. 1971. Degradation of acetylsalicylic acid by a strain of *Acinetobacter lwoffii*. *J Appl Bacteriol* 34:689–698. <https://doi.org/10.1111/j.1365-2672.1971.tb01006.x>.
5. Leadbetter JR, Greenberg EP. 2000. Metabolism of acyl-homoserine lactone quorum-sensing signals by *Variovorax paradoxus*. *J Bacteriol* 182:6921–6926. <https://doi.org/10.1128/JB.182.24.6921-6926.2000>.
6. Markowitz VM, Mavromatis K, Ivanova NN, Chen IMA, Chu K, Kyrpides NC. 2009. IMG ER: a system for microbial genome annotation expert review and curation. *Bioinformatics* 25:2271–2278. <https://doi.org/10.1093/bioinformatics/btp393>.
7. Caspi R, Foerster H, Fulcher CA, Kaipa P, Krummenacker M, Latendresse M, Paley S, Rhee SY, Shearer AG, Tissier C, Walk TC, Zhang P, Karp PD. 2007. The MetaCyc Database of metabolic pathways and enzymes and the BioCyc collection of Pathway/Genome Databases. *Nucleic Acids Res* 36:D623–D631. <https://doi.org/10.1093/nar/gkm900>.
8. Harwood CS, Parales RE. 1996. The β -keto adipate pathway and the biology of self-identity. *Annu Rev Microbiol* 50:553–590. <https://doi.org/10.1146/annurev.micro.50.1.553>.
9. Parales RE, Harwood CS. 2002. Bacterial chemotaxis to pollutants and plant-derived aromatic molecules. *Curr Opin Microbiol* 5:266–273. [https://doi.org/10.1016/S1369-5274\(02\)00320-X](https://doi.org/10.1016/S1369-5274(02)00320-X).