

UCSF

UC San Francisco Previously Published Works

Title

Elevated miR-499 Levels Blunt the Cardiac Stress Response

Permalink

<https://escholarship.org/uc/item/082069xg>

Journal

PLOS ONE, 6(5)

ISSN

1932-6203

Authors

Shieh, Joseph TC
Huang, Yu
Gilmore, Jacqueline
[et al.](#)

Publication Date

2011

DOI

10.1371/journal.pone.0019481

Peer reviewed

Elevated miR-499 Levels Blunt the Cardiac Stress Response

Joseph T. C. Shieh^{1,2}, Yu Huang¹, Jacqueline Gilmore³, Deepak Srivastava^{1,4,5*}

1 Gladstone Institute of Cardiovascular Disease, San Francisco, California, United States of America, **2** Division of Medical Genetics, Department of Pediatrics, University of California San Francisco, San Francisco, California, United States of America, **3** School of Medicine, University of California San Francisco, San Francisco, California, United States of America, **4** Division of Pediatric Cardiology, Department of Pediatrics, University of California San Francisco, San Francisco, California, United States of America, **5** Department of Biochemistry and Biophysics, University of California San Francisco, San Francisco, California, United States of America

Abstract

Background: The heart responds to myriad stresses by well-described transcriptional responses that involve long-term changes in gene expression as well as more immediate, transient adaptations. MicroRNAs quantitatively regulate mRNAs and thus may affect the cardiac transcriptional output and cardiac function. Here we investigate miR-499, a microRNA embedded within a ventricular-specific myosin heavy chain gene, which is expressed in heart and skeletal muscle.

Methodology/Principal Findings: We assessed miR-499 expression in human tissue to confirm its potential relevance to human cardiac gene regulation. Using a transgenic mouse model, we found that elevated miR-499 levels caused cellular hypertrophy and cardiac dysfunction in a dose-dependent manner. Global gene expression profiling revealed altered levels of the immediate early stress response genes (*Egr1*, *Egr2* and *Fos*), β -myosin heavy chain (*Myh7*), and skeletal muscle actin (*Acta1*). We verified the effect of miR-499 on the immediate early response genes by miR-499 gain- and loss-of-function in vitro. Consistent with a role for miR-499 in blunting the response to cardiac stress, asymptomatic miR-499-expressing mice had an impaired response to pressure overload and accentuated cardiac dysfunction.

Conclusions: Elevated miR-499 levels affect cardiac gene expression and predispose to cardiac stress-induced dysfunction. miR-499 may titrate the cardiac response to stress in part by regulating the immediate early gene response.

Citation: Shieh JTC, Huang Y, Gilmore J, Srivastava D (2011) Elevated miR-499 Levels Blunt the Cardiac Stress Response. PLoS ONE 6(5): e19481. doi:10.1371/journal.pone.0019481

Editor: Masa Tsuchiya, Keio University, Japan

Received: August 11, 2010; **Accepted:** April 7, 2011; **Published:** May 9, 2011

Copyright: © 2011 Shieh et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by the National Institutes of Health Grant HL092970 (JS) and grants from the National Institutes of Health, the California Institute for Regenerative Medicine, and the Younger Family Foundation (DS). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: dsrivastava@gladstone.ucsf.edu

Introduction

Heart failure affects over five million individuals in the United States and is characterized by progressive cardiac dysfunction. Such cardiomyopathies are characterized by significant changes in gene expression [1–7], many of which represent adaptive or maladaptive responses to stress. The immediate early gene response is activated rapidly upon cardiac stress and in a number of models of cardiac failure [8]. These genes are rapidly upregulated in response to environmental stimuli [9–11] and include the early growth response gene *Egr1* and the proto-oncogene *Fos*, both of which are involved in the cardiac stress response [12–14]. The induction of these genes is dependent upon serum response factor (SRF) and mediates further changes in gene expression, including regulation of sarcomeric and hypertrophic genes [15,16].

MicroRNAs are small non-coding RNAs that play an important role in gene regulation. Each of the over 650 known human microRNAs likely regulates hundreds of mRNAs and thus has the potential to affect many biologic processes, although the effect on rapid transcriptional responses has not been extensively studied. Some microRNAs are expressed ubiquitously while others display

tissue-specific expression patterns, suggesting unique functions within tissues. Several microRNAs, including miR-1, miR-133, miR-206 and miR-208 [17–29], are found in cardiac and/or skeletal muscle, and each has a potentially distinct regulatory function. Furthermore, many microRNAs reside within introns of coding genes, suggesting that their function may have co-evolved with their host gene [30,31].

In screening for microRNAs enriched in the human heart, we identified an abundant microRNA, miR-499, which has been the subject of several recent studies. Global microRNA expression profiling studies have identified miR-499 in the heart [32–35], however its function is just beginning to be elucidated. miR-499 is an evolutionarily conserved muscle-specific microRNA that is encoded within the intron of myosin heavy chain 7B (*Myh7B*) and is highly enriched in the cardiac ventricles. miR-499 plays a role in myosin gene regulation [36–38], however the functional effects of altered microRNA dosage may depend on the tissue's physiologic state. Furthermore, microRNA levels vary in many cardiac disease states [30,39–41], and in cardiac samples from individuals with aortic stenosis leading to pressure-overload and heart failure, miR-499 levels are altered [40]. In this study we show that elevated levels of miR-499 in hearts of transgenic mice result in

cardiomyocyte hypertrophy and stress-dependent cardiac dysfunction. Furthermore, we found miR-499 alters the immediate early gene response to cardiac stress, which may partially contribute to the effects of elevated miR-499.

Results

miR-499 is expressed in human heart and skeletal muscle

To identify microRNAs that are enriched in the human heart, we compared the expression of microRNAs in human fetal heart and fetal liver using microRNA microarrays. miR-499 was among the top cardiac-enriched microRNAs (**Fig. 1A**, **Table S1**), along with the well-studied microRNAs, miR-1 and miR-133. miR-499 is distinct from miR-1 and miR-133 in that it is encoded in only one genomic locus. By genomic sequence alignment, miR-499 is completely conserved throughout evolution with the exception of a single nucleotide change in chicken (**Fig. 1B**). We verified that the predicted human miR-499 genomic region encoded the microRNA by cloning the pre-miR-499 region, expressing it in cell culture, and performing Northern blot analysis (**Fig. 1C**). To determine the tissue-specificity of miR-499, we surveyed various human tissue types by Northern blot; miR-499 was detected only in the heart and in skeletal muscle (**Fig. 1D**) in agreement with

what has been reported in mice [36,37]. *In silico* analysis revealed that miR-499 was located in an intron of the human myosin heavy chain gene, *MYH7B* (**Fig. 1E**), analogous to the arrangement in *Myh7B* in mice.

Elevated miR-499 levels are associated with cardiac hypertrophy *in vivo*

We hypothesized that miR-499 expression may be playing an important role in cardiac gene regulation given its expression pattern, its evolutionary conservation, and its decrement in pressure-overload conditions such as human aortic stenosis [40]. To determine the consequence of persistent miR-499 expression in pressure-overload stress, we first generated transgenic mice with increased levels of miR-499 in the heart by expressing miR-499 under control of the cardiac, alpha-myosin heavy chain (*Myh6*) promoter, as it confers increasing postnatal ventricular cardiomyocyte expression [42–44]. We established two transgenic lines, which varied in degree of miR-499 expression compared to littermate controls (**Fig. 2A**). Mice expressing lower levels of miR-499 (line #17 or TG-17) appeared normal under basal conditions ($n = 20$). Transgenic mice expressing higher levels of miR-499 (line #9 or TG-9) developed enlarged hearts ($n = 16$) as demonstrated

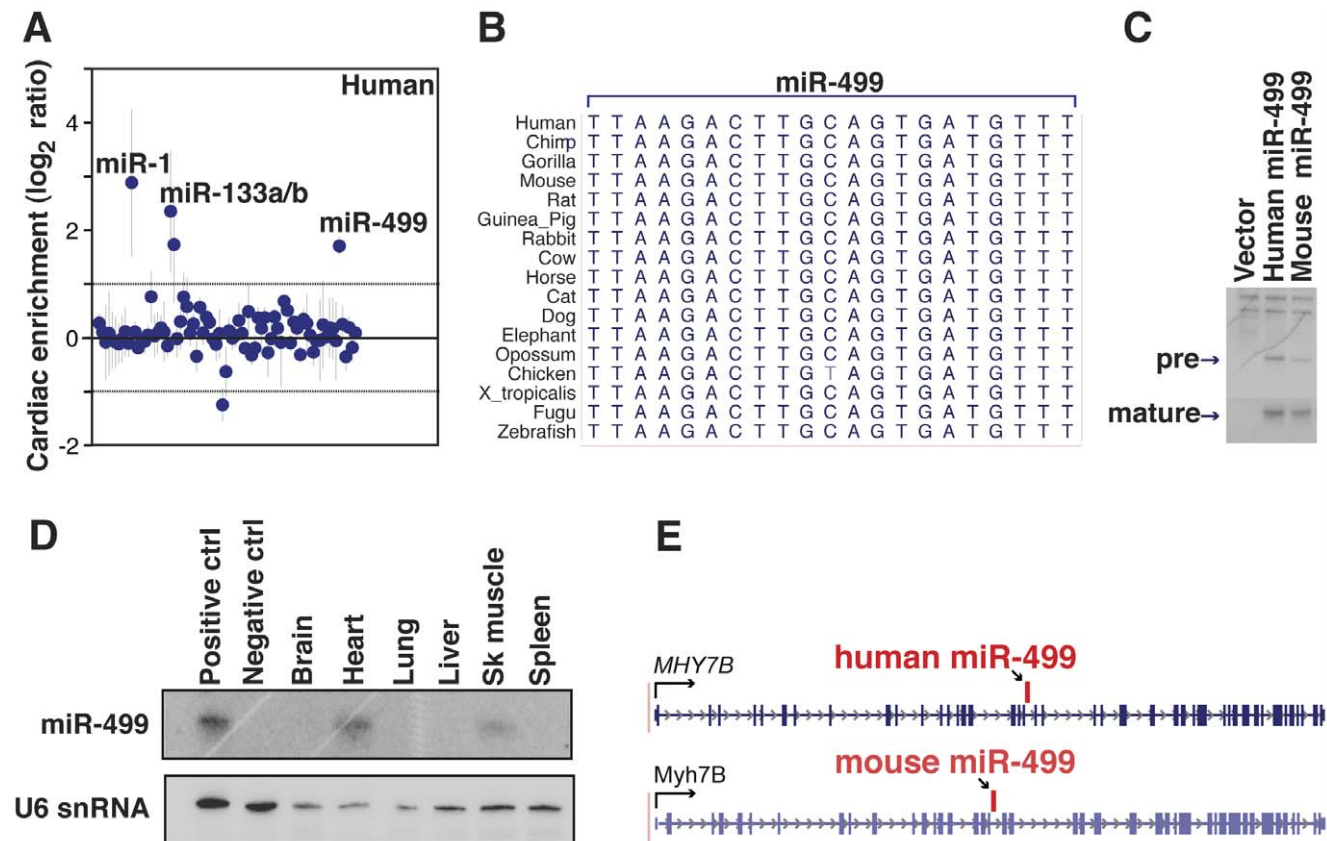


Figure 1. Human miR-499 is a conserved muscle-specific microRNA. (A) microRNA expression in human fetal heart compared to liver. Positive \log_2 ratio values indicate cardiac enrichment. $n = 3$ independent RNA samples per group, dots with vertical line indicate mean \pm standard deviation for each microRNA. (B) Alignment of genomic sequences corresponding to mature miR-499 in multiple species. (C) Northern blot with a miR-499 specific probe with RNA from 293T cells transfected with the genomic sequence surrounding the human or mouse miR-499 locus. Pre-miR and mature microRNA bands are visible. (D) Mature miR-499 was expressed in human heart and skeletal muscle as detected by Northern blot; U6 small nuclear RNA was probed as a loading control. RNA from miR-499 transfected 293T cells or control vector-transfected cells was used as positive or negative controls (ctrl). (E) Genomic organization of miR-499 in the myosin heavy chain gene of human *MYH7B* or mouse *Myh7b* adapted from the UCSC genome browser. The conserved intronic location of miR-499 is indicated between exons 20 and 21 in humans and exons 19 and 20 in mouse; arrows indicate the direction of transcription. doi:10.1371/journal.pone.0019481.g001

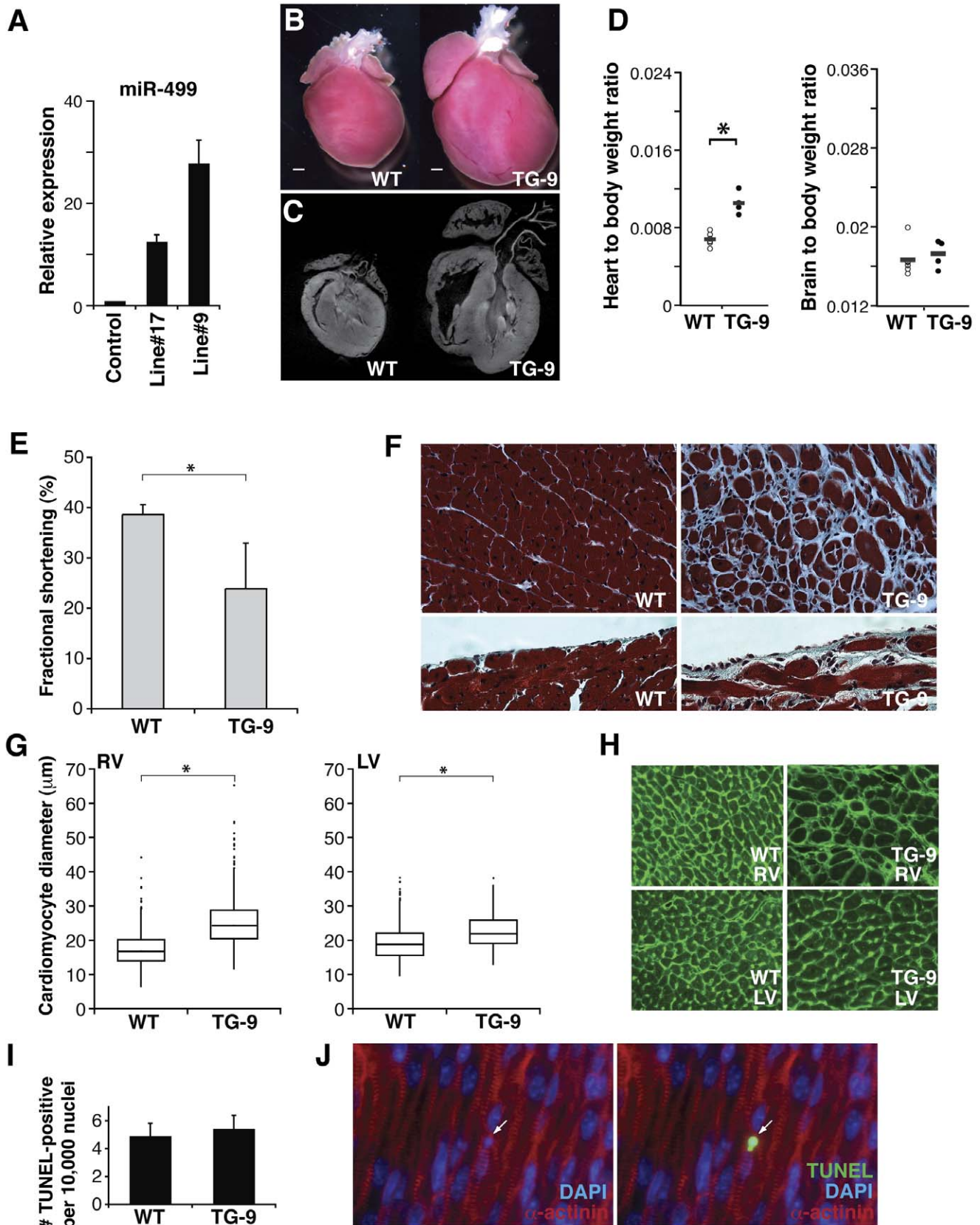


Figure 2. Characterization of hypertrophy phenotype in miR-499 transgenic mice. (A) Cardiac miR-499 expression levels (mean \pm standard deviation) in transgenic mouse lines compared to littermate controls. (B) Hearts from miR-499 transgenic mice (line #9, TG-9) were larger than those from littermate controls (6 weeks of age), $n = 16$ mice per group. (C) Coronal optical tomography section of miR-499 TG-9 heart demonstrated

chamber enlargement compared to control. (D) Heart-to-body weight ratios of miR-499 transgenic (TG-9) or littermate control (WT) mice. Brain-to-body weight ratios are also shown. Data from individual mice are plotted as dots with the mean indicated by a horizontal bar. * $P=0.0002$. (E) Fractional shortening measured by echocardiography of WT or TG-9 mice. * $P=0.00047$, $n=8$ mice for WT, 4 for TG. (F) Histological section of heart revealing area of fibrosis in TG-9 hearts compared to WT as shown by Masson trichrome staining in myocardium (upper panels) or beneath the endocardial surface (lower panels); magnification 400 \times . (G) Quantification of cardiomyocyte size from TG-9 mice compared to WT mice (RV, right ventricle, and LV, left ventricle, both * $P<0.05$; box plots depicts size distribution of over 300 cells per heart in each of two mice per group). (H) Wheat germ agglutinin (WGA) staining; magnification 200 \times . (I) TG-9 and WT heart demonstrated similar levels of apoptosis, $n=3$ mice per group; representative stained histologic section (J) demonstrated the alpha-actinin staining (red) and DAPI-stained nuclei (blue) overlaid with TUNEL-positive nuclei (green, right panel).
doi:10.1371/journal.pone.0019481.g002

by gross pathology at 5 weeks of life (Fig. 2B). The higher expressing miR-499 transgenic mice (TG-9) had larger hearts (Fig. 2C) and increased heart-to-body weight ratios compared to littermate controls (Fig. 2D), while the brain-to-body weight ratios were similar. Echocardiography demonstrated decreased fractional shortening in TG-9 mice indicating contractile dysfunction (Fig. 2E, Table 1).

On histopathological analysis, we found extensive cellular hypertrophy and patchy interstitial fibrosis in TG-9 hearts (Fig. 2F) ($n=4$). To assess the degree of hypertrophy, we quantified cardiomyocyte cross-sectional area after staining cardiac sections using wheat germ agglutinin (WGA). TG-9 hearts displayed a marked increase in cardiomyocyte size indicating cellular hypertrophy (Fig. 2G and 2H). There was no increase in TUNEL-positive/alpha-actinin-positive cells in TG-9 mice at 1 week (Fig. 2I and J) or 7 weeks of age, suggesting that cardiomyocyte apoptosis was not a primary contributor to the phenotype.

A second line of miR-499 transgenic mice, expressing relatively lower levels of miR-499 (line #17, TG-17), had hearts that appeared unremarkable under basal conditions (Fig. 3A). The ratio of the heart-to-body weights was similar in transgenic and control mice (Fig. 3B), and echocardiographic evaluation of cardiac function in TG-17 mice trended lower, although the results were not statistically significant, at least under basal conditions (Fig. 3C, Table 1). However, WGA-staining of TG-17 hearts revealed a mild but significant increase in cardiomyocyte size (Fig. 3D and E), indicating mild cellular hypertrophy.

miR-499 targeting of Sox6

Given the baseline hypertrophy in miR-499-transgenic mice, we hypothesized that miR-499 expression may alter genes important in this cellular program, either directly or indirectly. Initially, we examined genes predicted to be targets of miR-499 using a targeted approach. From a survey of microRNA target prediction programs including MiRanda, TargetScan, and

Mirtarget, Sox6 emerged as an attractive candidate, given three potential miR-499 binding sites and potential roles for Sox6 in muscle and heart [36–38,45,46]. We verified that Sox6 was an *in vitro* target of miR-499 by cloning the Sox6 3'-UTR and then tested for microRNA repression by luciferase assays. We transfected increasing amounts of miR-499 into 293T cells in culture and found dose-dependent inhibition of the Sox6 UTR-luciferase construct, however another cardiac microRNA, miR-133, had no effect regardless of the dose (Fig. 4A). Surprisingly, Sox6 protein levels were not diminished in transgenic mice, however the myosin heavy chain classically involved in hypertrophy, β -myosin heavy chain (β -MyHC), was markedly increased (Fig. 4B). This was accompanied by an increase in the corresponding *Myh7* (β -MyHC) transcript, while Sox6 mRNA was unaltered, similar to its protein levels (Fig. 4C).

A distinctive pattern of cardiac gene regulation mediated by miR-499

We investigated miR-499-dependent changes in cardiac gene expression by comparing global gene expression in miR-499-transgenic mice and littermate controls. To avoid changes secondary to heart failure, we used the miR-499 transgenic line (TG-17) that did not display overt cardiac dysfunction. RNA was prepared from the ventricles of postnatal day 17 miR-499 transgenic mice or littermate controls, and gene expression was compared by microarray analysis (Fig. 5A). When we analyzed for differentially expressed genes, 0.03% of the transcripts (nine transcripts of the 28,853 interrogated) were dysregulated when we used a stringent cut-off (P value <0.05 , Benjamini-Hochberg correction for false-discovery rate), whereas a less stringent cut-off demonstrated that 0.5% of the transcripts (172 transcripts of the 28,853 interrogated) were significantly altered (Bayes log odds of differential expression $B>0$). The average fold change was 1.43 for upregulated genes and -1.37 for downregulated genes with a range of 2.51 to -2.53 fold. miR-499 levels were increased 12.6-fold, in agreement with our microRNA qPCR results.

Table 1. Echo Assessment of miR-499 Transgenic Mice.

	WT	TG-9	P value	WT	TG-17	P value	WT [†]	TG-17 [†]	P value	WT TAB [‡]	TG-17 TAB [‡]	P value
%FS	38.63 \pm 1.95	23.88 \pm 9.06	0.00047*	40.67 \pm 0.88	37.42 \pm 4.82	0.31	39.7 \pm 4.16	36.3 \pm 4.13	0.34	30.9 \pm 2.29	19.9 \pm 4.10	0.00079*
LVEDD	3.85 \pm 0.18	4.18 \pm 1.06	0.37	3.71 \pm 0.23	3.83 \pm 0.09	0.38	3.79 \pm 0.41	3.81 \pm 0.11	0.95	3.74 \pm 0.08	4.18 \pm 0.27	0.0095*
LVESD	2.37 \pm 0.17	3.24 \pm 1.21	0.047*	2.20 \pm 0.14	2.40 \pm 0.24	0.26	2.29 \pm 0.37	2.43 \pm 0.21	0.57	2.58 \pm 0.09	3.35 \pm 0.38	0.0023*
LVPWd	0.72 \pm 0.05	0.76 \pm 0.11	0.43	0.72 \pm 0.08	0.68 \pm 0.03	0.42	0.79 \pm 0.07	0.81 \pm 0.04	0.66	1.08 \pm 0.03	1.05 \pm 0.04	0.26
LVPWs	1.20 \pm 0.07	1.00 \pm 0.15	0.0054*	1.19 \pm 0.06	1.12 \pm 0.08	0.25	1.31 \pm 0.06	1.26 \pm 0.09	0.44	1.46 \pm 0.09	1.23 \pm 0.11	0.0078*

Measurements of left ventricle (LV) in mm \pm standard deviation, LVEDD, end-diastolic dimension; LVESD, end-systolic dimension; LVPWd, posterior wall diastole; LVPWs, posterior wall systole. $n=8$ for WT, 4 for TG-9, 3–5 for TG-17. P values calculated using student's T-test.

[†]Second assessment 5 weeks later.

[‡]TAB 3.5 mo after thoracic aortic banding.

* $P<0.05$.

doi:10.1371/journal.pone.0019481.t001

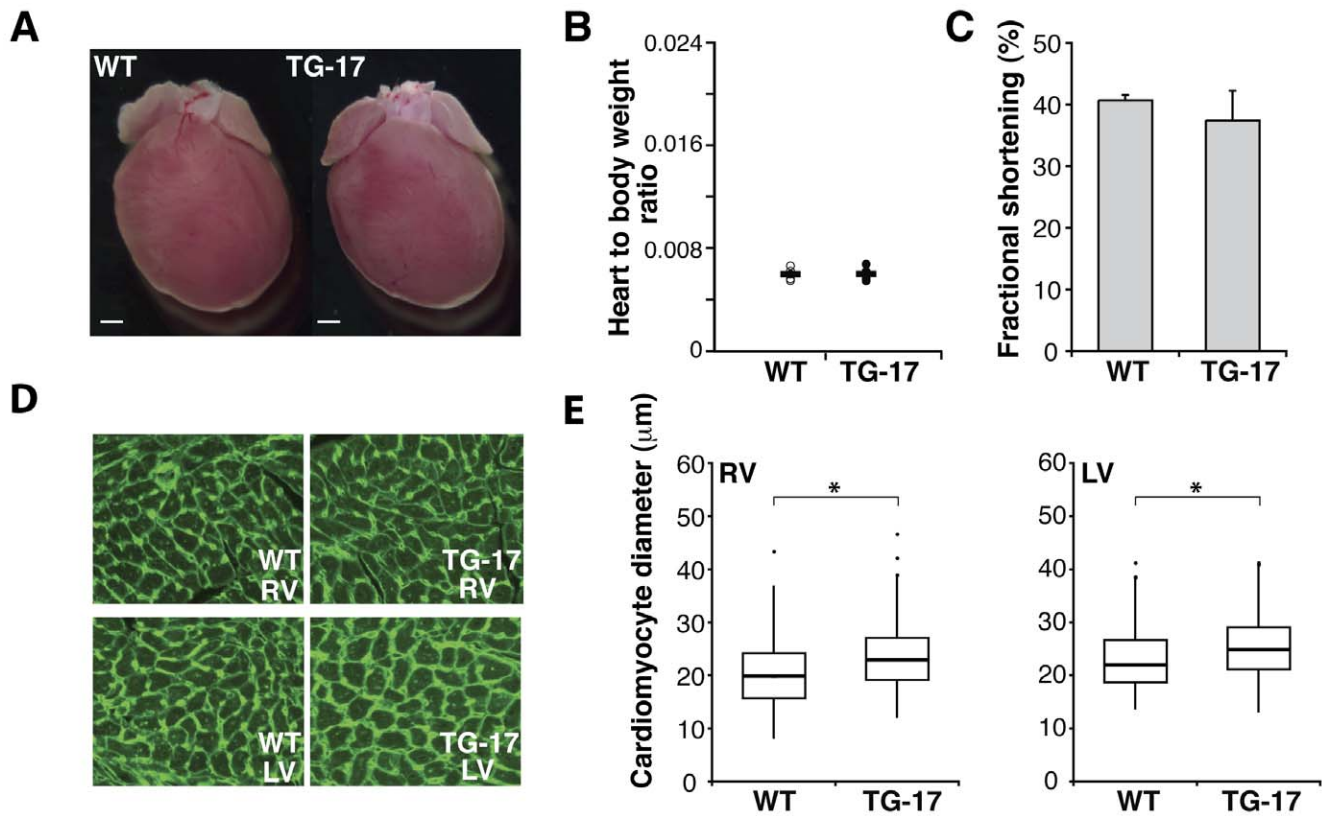


Figure 3. Modest levels of miR-499 expression results in mild hypertrophy. (A) Gross appearance of hearts from miR-499 transgenic mice (line #17, TG-17) and littermate controls (WT) was similar. (B) Heart-to-body weight ratios in TG-17 and WT, $n=4-6$ mice per group. (C) Fractional shortening of WT or TG-17 mice by echocardiography. (D) WGA staining of histological sections of hearts from TG-17 and WT mice; magnification 200 \times . (E) Quantification of cardiomyocyte size revealed an increase in TG-17 (RV and LV both $*P<0.05$). doi:10.1371/journal.pone.0019481.g003

Transcripts such as *natriuretic peptide precursor type B (Nppb)*, β -myosin heavy chain (*Myh7*), and *alpha 1 skeletal muscle actin (Acta1)*, genes that are known to be upregulated in hypertrophy, were increased 2.0, 1.9, and 1.6 fold, respectively (Fig. 5B). *Sox6*, which was a target for miR-499 in luciferase assays, was not altered in transgenic mice (1.0 fold, not significant). We noted that the most downregulated transcripts were the immediate early response genes, *early growth response 1 (Egr1)* (-2.5 fold, the most downregulated transcript in the entire array) and *FBj osteosarcoma oncogene (Fos)* (-1.9 fold, the second most downregulated transcript) (Fig. 5C). *Egr2*, an immediate early gene with an unknown role in the heart, was diminished -1.6 fold, and was also among the ten most downregulated transcripts. As the immediate early genes may be involved in the response to stress, we validated the reduced *Egr1*, *Egr2*, and *Fos* levels using qPCR, which further suggested dysregulation (Fig. 5D).

The Immediate Early Gene Response is Altered by miR-499 Levels

We reasoned that the immediate early response genes may be playing a role in the hypertrophic phenotype for several reasons: 1) the immediate early response genes are rapidly altered in response to cardiac stress [8,47,48]; 2) perturbation of the immediate early response alters cardiac hypertrophy [49,50]; and 3) immediate early genes are typically SRF-dependent, and the cardiac abnormalities in the miR-499 transgenic partially resembled that of mice where SRF was temporally deleted [51]. First, to examine the miR-499:immediate early gene relationship independent of the

potential complexity of the *in vivo* transgenic system, we tested whether the immediate early gene response was altered using cultured cells where miR-499 levels were manipulated. We used an antisense morpholino to inhibit miR-499 generation in the ventricular cardiomyocyte line, H9c2. Knockdown of miR-499 levels led to an increase in *Egr1* and *Fos* levels 30 min following immediate early gene response activation by serum (Fig. 6A), however the effect was temporally limited, as it resolved by 60 min. Conversely, introduction of miR-499 mimic lowered *Egr1* levels in cell culture after serum stimulation (Fig. 6B). There was no significant effect on *Fos* in this setting. These data are consistent with a role for miR-499 in regulating the immediate early gene response.

We next investigated whether elevated miR-499 levels affected the epidermal growth factor (EGF)-induced immediate early gene response *in vivo* [52]. miR-499-transgenic mice (TG-17) displayed a blunted activation of *Egr1* in response to systemic administration of EGF (Fig. 6C). Activation of *Fos* was not different in the miR-499 transgenic mice, similar to what we observed in cultured cells. These data suggested that miR-499 levels negatively regulate the stress-induced activation of *Egr1* and this may be important in the cardiac response to stress and in hypertrophy.

We assessed whether serum response factor (SRF), which is upstream of the immediate early genes, was altered in the miR-499 transgenics. When we compared SRF protein levels in WT and TG mice, there was no difference (Fig. 6D), suggesting that the immediate early genes are regulated by miR-499 independent of SRF levels. To test whether miR-499 directly or indirectly

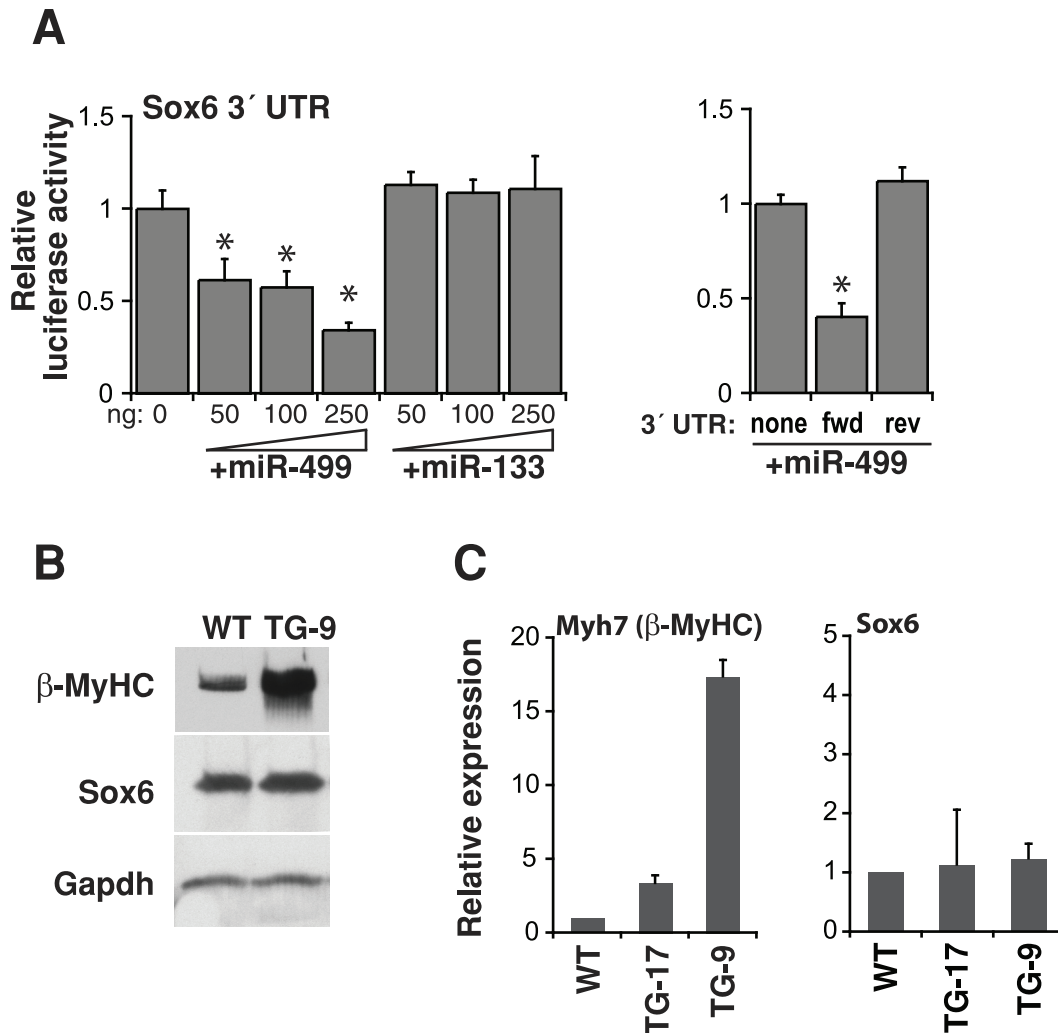


Figure 4. miR-499 targeting of Sox6. (A) The 3'UTR of Sox6 was placed downstream of a luciferase reporter construct and tested for repression by miR-499 in 293T cells. Sox6 3'UTR-mediated repression increased as amounts of miR-499 was increased; this was not observed with miR-133 or when the UTR orientation was reversed, $n=3-4$ transfections per condition, $*P<0.05$. (B) Western blots indicate protein levels in TG-9 or WT hearts using antibodies against Sox6, β -myosin heavy chain (β -MyHC), or Gapdh. Results are representative of three hearts each. (C) Cardiac expression of β -myosin heavy chain (β -MyHC) or Sox6 mRNA transcripts in TG lines and WT mice by qPCR, $n=3$. doi:10.1371/journal.pone.0019481.g004

regulates the immediate early genes, we cloned the 3' UTRs of the immediate early response genes and placed each downstream of luciferase and tested these in H9c2 and 293T cells. miR-499 did not repress luciferase constructs with the Egr1, Egr2, or Fos UTRs (Fig. 6E), whereas miR-499 repressed constructs containing the Sox6 3'UTR. Since the 3' UTRs of Egr1, Egr2, and Fos were insufficient to mediate miR-499 repression, we suspect they are affected by miR-499 in a yet unknown manner that is not dependent on altered SRF levels.

miR-499 Expressing Mice Are Predisposed to Stress-Induced Cardiac Dysfunction

Given the effect of miR-499 levels on the immediate early gene response, which is known to be involved in cardiac stress and hypertrophy [12–14], we hypothesized that increased miR-499 levels may predispose to stress-induced cardiac dysfunction. We therefore tested whether the TG-17 miR-499-transgenic mice, which had normal function at baseline, were predisposed to increased cardiac dysfunction upon cardiac pressure overload. We

used the well-characterized model of thoracic aortic banding (TAB) to induce pressure overload in miR-499 transgenic (TG-17) or control mice and monitored cardiac function over 12 weeks. The banded transgenic hearts demonstrated accentuated cardiac enlargement (Fig. 7A) and a marked increase in heart weight to body weight ratio (Fig. 7B) compared to banded WT controls. After TAB, the miR-499 transgenic mice displayed more severe contractile dysfunction as indicated by the decline in fractional shortening compared to control mice (Fig. 7C, Table 1) that underwent the same degree of TAB (peak aortic pressure gradient: WT 49 ± 10 mmHg, TG 43 ± 9 mmHg, $p=0.251$). Interestingly, the cardiomyocyte size was not higher in the LV, but was slightly increased in the RV of transgenic mice (Fig. 7D and 7E), similar to TG-9 mice. Overall, these results suggest elevated miR-499 levels predispose the heart to cardiac dysfunction.

To determine the dose-responsiveness of miR-499 sensitive genes, we analyzed gene expression in the higher miR-499 expressing TG-9 line at 17 days of age. Thirty-two common transcripts were altered in both TG-17 and TG-9, and thirty of

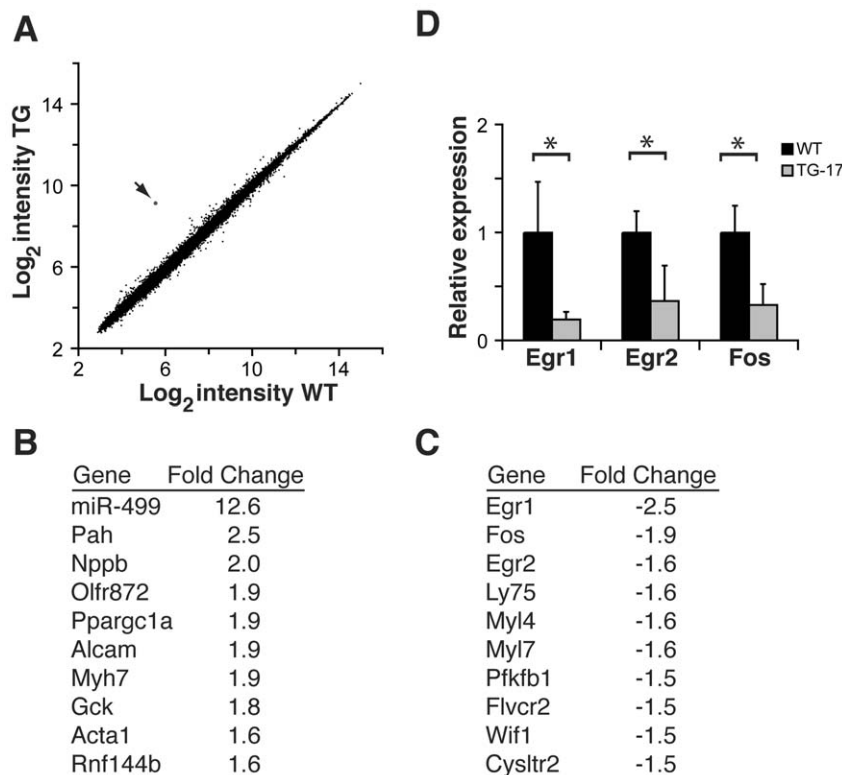


Figure 5. Altered cardiac transcripts from global analysis of miR-499 TG hearts. (A) Expression microarray results depicting average log₂ intensity of gene expression from post-natal day 17 WT hearts (x-axis) versus TG-17 hearts (y-axis). Each dot represents a single transcript from three miR-499 transgenic mice and three littermate controls. The dot indicated in red (arrow) represents miR-499 expression. (B) and (C) Gene expression alterations in miR-499 mice (top ten up- and downregulated genes). Egr1 and Fos were the most downregulated transcripts. Full expression data have been deposited in NCBI GEO. (D) qPCR validation of microarray results regarding the immediate early response gene transcripts Egr1, Egr2, and Fos. *P<0.05, n=3 hearts per group with samples assessed in triplicate. Bars indicate mean with 95% confidence interval. doi:10.1371/journal.pone.0019481.g005

these were greater in magnitude in TG-9 than TG-17 (**Table S2**). None of the genes were predicted by Targetscan. Importantly, pathway analysis revealed enrichment for the GO terms sarcomere (P=0.0074), contractile fiber part (P=0.0085), myofibril (P=0.0095), and contractile fiber (P=0.010), further highlighting the potential role of miR-499 in regulating sarcomeric function in the heart.

Discussion

In this study, we found that elevated levels of miR-499 in the heart can lead to cardiomyocyte hypertrophy and cardiomyopathy in a dose-dependent manner. High levels of miR-499 led to spontaneous cardiac contractile dysfunction, while more modest levels of transgenic expression conferred susceptibility to pressure-induced dysfunction.

Our studies support an association between elevated cardiac miR-499 levels and cardiac dysfunction, particularly in the setting of pressure overload. Interestingly, in a study that analyzed microRNAs in human aortic valve stenosis, miR-499 levels were decreased [40], and it is possible that this decrement may regulate cardiac gene expression changes that are important in the physiologic response to cardiac pressure load. We suspect the failure of normal miR-499 down-regulation in our transgenic mice disrupted the normal response to cardiac pressure stress; similar theories have recently been put forward for cardiac transgenic mice expressing miR-133a [53]. Interestingly, a recent study [39] reported that miR-499 levels increase in human cardiac failure,

and the findings in our transgenic mouse model may support a detrimental role for elevated miR-499 levels.

We identified a discrete set of altered transcripts in unmanipulated, normally functioning, miR-499 transgenic hearts, including natriuretic peptide precursor type B (*Nppb*), β -MyHC (*Myh7*) and alpha 1 skeletal muscle actin (*Acta1*). This suggests gene expression changes due to miR-499 that favor hypertrophy. These changes were confirmed in the second transgenic line. The limited magnitude of transcript changes by miR-499 suggests that the hypertrophic phenotype observed was not due to gross gene expression disruption. Although the Sox6 3'UTR could be targeted by miR-499, as we and others have demonstrated [36–38,46], Sox6 levels were not altered in the transgenic mice, suggesting that Sox6 was not targeted by increased miR-499 levels *in vivo*, or that additional compensatory mechanisms may be involved *in vivo*. It is possible that targets seen in luciferase assays may not necessarily translate into targeting *in vivo*.

We present several lines of evidence that demonstrate that miR-499 levels fundamentally alter the immediate early gene response, which is known to be important in the cardiac stress response. First, Egr1 and Fos were the most diminished transcripts in TG-17 miR-499-altered hearts, suggesting a robust alteration in the immediate early gene expression program, while in TG-9 Egr1 was also altered (although Fos was not significantly changed). Second, inhibition of miR-499 in culture led to increases in induced Egr1 and Fos levels, which demonstrates that this regulation is present even outside of the context of the cardiac transgene. Pressure-induced stress, which is known to invoke the

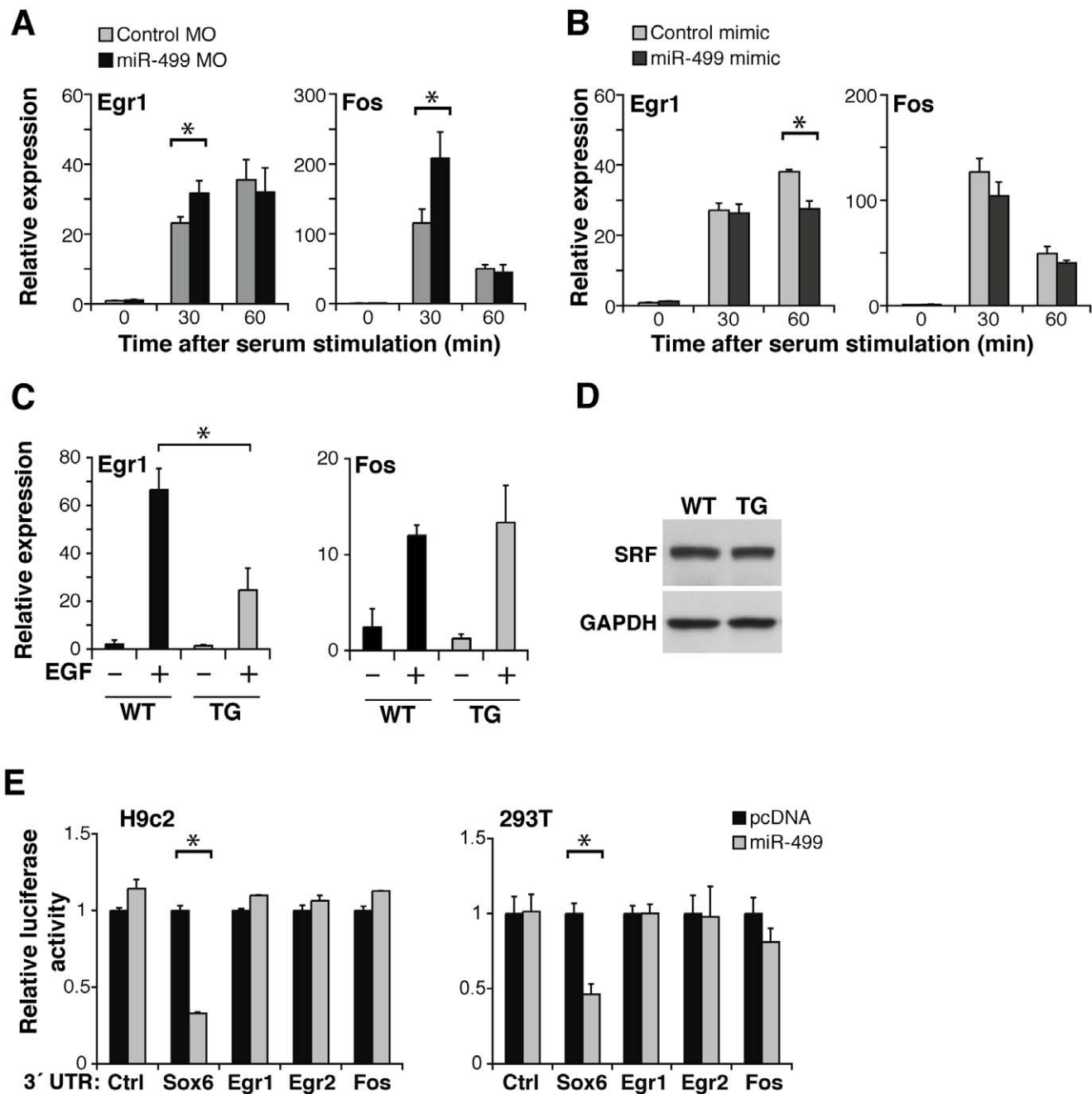


Figure 6. miR-499 blunts the induction of the immediate early response genes. (A) Egr1 and Fos mRNA levels by qPCR relative to Gapdh in the ventricular cell line H9c2, upon introduction of a morpholino (MO) that blocks miR-499 generation or a control MO. (B) Egr1 and Fos mRNA levels by qPCR in H9c2 cells after the introduction of miR-499 or control mimic. The x-axis in (A) and (B) indicates the time after serum stimulation to activate the immediate early gene response; $n=3$ per group. $*P<0.05$. (C) Egr1 or Fos mRNA levels in miR-499 TG-17 or WT hearts as assessed by qPCR 1.5 hours after *in vivo* stimulation with epidermal growth factor (EGF). Bars indicate mean \pm standard deviation; $n>3$ per group. $*P<0.05$. (D) Western blot indicates protein levels in TG-9 or WT hearts using SRF antibodies with Gapdh as loading control. Results are representative of three hearts each. (E) The 3' UTR of Sox6, Egr1, Egr2, or Fos was placed downstream of a luciferase reporter construct and tested for repression by miR-499 in culture. Sox6 3' UTR-mediated repression by miR-499 was evident, however Egr1, Egr2, and Fos 3' UTRs were not repressed by miR-499 ($n=3$ per condition, $*P<0.05$).

doi:10.1371/journal.pone.0019481.g006

immediate early gene response, dramatically altered cardiac function in transgenic mouse hearts. We therefore hypothesize that miR-499 alters the cardiac response to stress in part by modulating the immediate early gene response.

The connection between miR-499 and the immediate early gene response is important since activation of this pathway is

thought to precede further transcriptional responses to stress. Similar to TG-17 mice expressing miR-499, mice deficient in Egr1 are normal at baseline, but have an impaired response to cardiac stress [49], as do transgenic mice expressing the Egr1-repressor, Nab1 [50]. Interestingly, mice lacking the cardiac-specific microRNA miR-208a have decreased amounts of miR-499

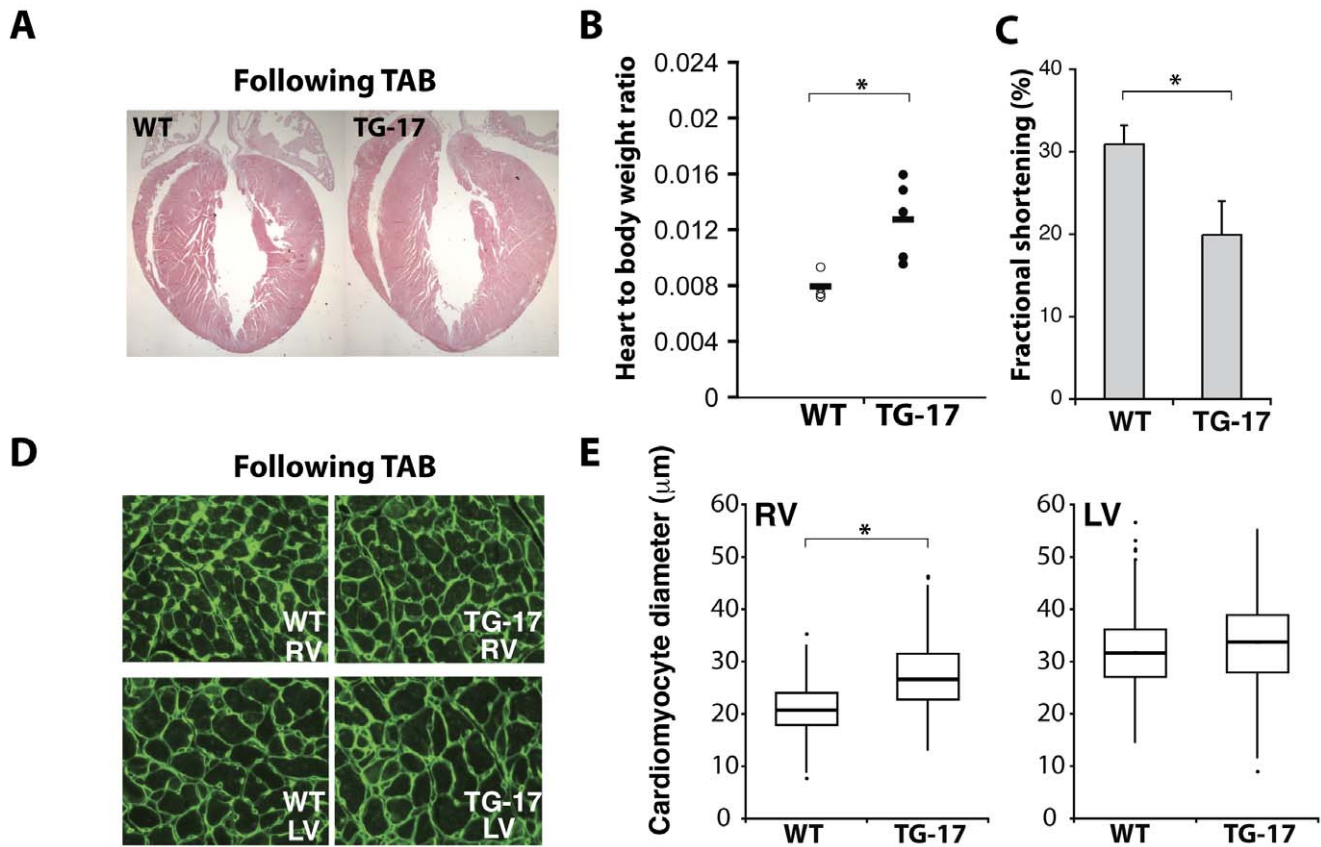


Figure 7. miR-499 elevation predisposes to stress-induced cardiac dysfunction in vivo. (A) Following thoracic aortic banding (TAB), TG-17 hearts were slightly larger compared to banded WT hearts, and (B) the heart-to-body weight ratio was increased to a greater degree in TG-17 compared to similarly banded WT controls ($n=4-5$ per group, $*P<0.05$). (C) Fractional shortening of WT and TG-17 hearts by echocardiography, $*P=0.00079$. (D) WGA staining and quantification revealed increased size of cardiomyocytes following TAB. $*P<0.05$. doi:10.1371/journal.pone.0019481.g007

[25,36]. Evaluation of genes reported to be dysregulated in miR-208a mutant mice revealed elevated levels of *Egr1*, *Egr2*, and *Fos* in the heart, suggesting a potential relationship between miR-499 and miR-208 in regulation of the immediate early gene response to cardiac stress. The observation that miR-499 transgenic mice develop or are predisposed to cardiac dysfunction may reflect the fact that the immediate early response genes hold a key position in the hierarchy of the cardiac transcriptional response to stress. It will be interesting to determine if levels of this non-coding RNA affect the progression of disease in the setting of pressure overload, such as in hypertension or aortic valve disease.

Materials and Methods

Ethics Statement

Care and handling of all experimental animals used in this study were in accordance with the University of California San Francisco's Institutional Animal Care and Use Committee policies and approved under protocol #AN080001.

MicroRNA detection by Northern blotting

Total RNA (12 mcg) was separated on 12% polyacrylamide/urea gels and transferred to nitrocellulose membranes. Membranes were probed with ^{32}P -labelled locked nucleic acid probe (Exiqon) or DNA oligonucleotide probe complementary to mature miR-499 or to U6. We transfected 293T cells (ATCC) with miR-

499-expression construct or vector alone for RNA for positive and negative controls. Signals were detected by phosphorimaging.

MicroRNA and messenger RNA detection by PCR

RNA samples were reverse transcribed for microRNA detection with microRNA-specific primers (Applied Biosystems) or for mRNAs with oligo-dT (Invitrogen). miR-499 levels and mRNAs were quantified using RNA from transgenic mouse hearts and littermate controls. Quantitative real-time PCR (qPCR) was performed in an ABI HT7900 with Taqman primers (Applied Biosystems). Taqman primers used: miR-499 (001352), miR-16 (000391), β -myosin heavy chain (*Myh7*, Mm00600555_m1), *Sox6* (Mm01274768_m1), *Egr1* (Mm00656724_m1), *Egr2* (Mm00456650_m1), *Fos* (Mm01302932_g1).

MicroRNA genomic cloning

We amplified fragments containing miR-499 from genomic DNA using high-fidelity PCR (PlatinumTaq, Invitrogen) and directional cloning into pcDNA3.1-TOPO (Invitrogen) for expression studies in cell culture. The UCSC genome browser and Sanger miRBase were used to analyze cloned products, to assess microRNA conservation, and for primer design.

MicroRNA target assays

We surveyed multiple bioinformatics prediction programs that predict potential microRNA targets. MiRanda, TargetScan, and

Mirtarget [54–56] included Sox6 as a potential target. The 3' untranslated regions (UTRs) of Sox6, Egr1, Egr2, or Fos were cloned into pCR2.1-TOPO (Invitrogen) and subcloned downstream of the luciferase coding sequence in pMIR-REPORT (Ambion). For microRNA target assays, we transfected cells with microRNA expression vector (50, 100, and 250 ng) and luciferase construct containing the target sequence of interest. Briefly, we expressed microRNAs in H9c2 or 293T cells with expression plasmids, equalized the total amount of DNA transfected, and expressed the luciferase construct with the 3' UTR of interest (Lipofectamine 2000, Invitrogen). We also tested whether repression was lost when we transfected the reversed 3'UTR sequence construct with our miR-499 vector (250 ng). Renilla luciferase-expressing plasmid (pRL-CMV, Promega) was used to normalize transfection efficiency.

Transgenic mice

To express miR-499 *in vivo*, we amplified DNA surrounding the mouse pre-miR-499 region and cloned the 574-bp fragment into the alpha-myosin heavy chain promoter vector using the following primers and SalI linkers: 5'-CGT GTC GAC CAA GTC TGG GGT GAA AGA GAA G-3' (forward), 5'-TGT GTC GAC GGT CAT GAG CTT GTT GAG GTT C-3' (reverse) and injected into one-cell FVB/NCrl embryos before implantation in pseudopregnant females. Phenotyping was performed by echocardiography (Vevo 770, VisualSonics) with standard measurement techniques (reviewed in [57]). Echocardiography was performed using anesthesia with isoflurane and with continuous monitoring of mice to ensure the heart rate was kept constant. Thoracic aortic banding was performed as described [58] with slight modifications. Briefly, the aorta was exposed through a mediastinal approach, and suture was tied around a 27-gauge needle placed adjacent to the aorta. Echocardiography confirmed aortic constriction by peak velocity measurement. For analysis of transgenic hearts, hearts were harvested in PBS containing 25 mM KCl and optical projection tomography was performed for virtual sectioning [59]. Histologic analysis was performed using Masson Trichrome staining, DAPI, alpha-actinin staining (clone EA-53, Sigma), and wheat germ agglutinin staining on corresponding coronal sections followed by quantification of cardiomyocyte cross-sectional area by measuring the cell diameter with visible nuclei in over 300 cells. Cell death was assessed by quantification using alpha actinin-staining combined with tunel analysis (Roche) at different ages. For analysis of the immediate early gene response *in vivo*, recombinant epidermal growth factor (30 mcg, Invitrogen) or vehicle (normal saline) was injected intraperitoneally into control or transgenic mice, and hearts were collected after 1.5 hours for analysis of immediate early gene expression.

Western blotting

Protein lysates were run on SDS-PAGE gels, transferred to PVDF membranes, and probed with antibodies to Sox6 (ab12054, Abcam), slow myosin (clone NOQ7.5.4D, Sigma), SRF (sc-335, Santa Cruz), or Gapdh (Santa Cruz).

Gene expression analysis

Data have been deposited in The National Center for Biotechnology Information (NCBI) Gene Expression Omnibus (GEO, <http://www.ncbi.nlm.nih.gov/geo/>) series accession number GSE21104 (<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?token=nrqpfuaaqeacso&acc=GSE21104>). Total RNA was isolated from ventricular tissue from three miR-499-transgenic

mice (line #17, TG-17) and three littermate controls at postnatal day 17. Gene expression was assessed using Mouse Gene 1.0 ST Array as recommended by the manufacturer (Affymetrix). Differentially expressed genes were identified using the limma package. Adjustment for multiple testing was performed as described in the text. Further validation was performed using qPCR as described above. Human heart and liver RNA samples (Virogen), gestational age 6–8 weeks, were labeled and run for microRNA expression as described by the manufacturer (Exiqon). Gene ontology analysis was performed using DAVID Bioinformatics Resources 6.7 [60,61].

MicroRNA modulation in cell culture

To modulate microRNA levels, miR-499 mimic or control mimic (Dharmacon) or a morpholino directed against the miR-499 precursor or a scrambled control morpholino (GeneTools) was electroporated (Amaxa) into H9c2 and allowed to recover overnight. After serum starvation in 1% fetal bovine serum, 10% serum was applied for designated times and RNA collected for RT-PCR analysis. Sequence of morpholino directed against the miR-499 precursor 5'-ATG CAG AGG AGC TAA ACA TCA CTG C-3' or a scrambled control morpholino 5'-GAT TGC GAA GAG TCT CAC AAG ACC A-3'.

RNA isolation

Total RNA was isolated using guanidine thiocyanate and phenol (Molecular Research Center, Inc.) and following manufacturer's instructions for chloroform extraction, isopropanol precipitation, and 75% ethanol washing. For tissue RNA extraction, tissues were processed using a bead homogenizer (WisBiomed).

Primer sequences

Genomic amplification primers for human miR-499: 5'-CAC CCA AGT CTG GGG TGA AAG AGA AG-3' (forward) and 5'-GGT CAT CAG CTT GTT GAG GTT C-3' (reverse). 3'UTR sequence primers: Egr1 5'-GGC AGG AAA GAC ATA AAA GCA C-3' (forward) and 5'-ACA TAT CCC ATG GGC AAT AGA G-3' (reverse), Egr2 5'-AAC ACT ACC ACC CTT CCC TGT T-3' (forward) and 5'-AGC CAT CCA TTA TCT GAA CTC C-3' (reverse), Fos 5'-TGG TGC ATT ACA GAG AGG AGA A-3' (forward) and 5'-TGG AAC AAT AAG CAA ACA ATG C-3' (reverse), Sox6 5'-CTC ACT AGT TGG CTC CAC AAT ACA TCA GC-3' (forward) and 5'-CTC AAG CTT CCA AGT GAC AAA ATG GCT CA-3' (reverse).

Supporting Information

Table S1 Enrichment of microRNAs in human heart versus liver. Log ratio for each microRNA and standard deviation are shown. Positive ratios indicate enrichment in heart compared to the liver. (DOC)

Table S2 Common genes altered in miR-499 transgenic lines, TG-17 and TG-9. For each gene, the gene symbol, genomic location, log ratios and fold change of TG versus WT are shown. (XLS)

Author Contributions

Conceived and designed the experiments: JS. Performed the experiments: JS YH JG. Analyzed the data: JS DS. Wrote the paper: JS DS.

References

- Liew CC, Dzau VJ (2004) Molecular genetics and genomics of heart failure. *Nat Rev Genet* 5: 811–825.
- Nabel EG (2003) Cardiovascular disease. *N Engl J Med* 349: 60–72.
- Lowes BD, Gilbert EM, Abraham WT, Minobe WA, Larrabee P, et al. (2002) Myocardial gene expression in dilated cardiomyopathy treated with beta-blocking agents. *N Engl J Med* 346: 1357–1365.
- Kaab S, Barth AS, Margerie D, Dugas M, Gebauer M, et al. (2004) Global gene expression in human myocardium-oligonucleotide microarray analysis of regional diversity and transcriptional regulation in heart failure. *J Mol Med* 82: 308–316.
- Hwang JJ, Allen PD, Tseng GC, Lam CW, Fananapazir L, et al. (2002) Microarray gene expression profiles in dilated and hypertrophic cardiomyopathic end-stage heart failure. *Physiol Genomics* 10: 31–44.
- Barth AS, Kuner R, Bunes A, Ruschhaupt M, Merk S, et al. (2006) Identification of a common gene expression signature in dilated cardiomyopathy across independent microarray studies. *J Am Coll Cardiol* 48: 1610–1617.
- Movassagh M, Choy MK, Goddard M, Bennett MR, Down TA, et al. (2010) Differential DNA methylation correlates with differential expression of angiogenic factors in human heart failure. *PLoS One* 5: e8564.
- Izumo S, Nadal-Ginard B, Mahdavi V (1988) Protooncogene induction and reprogramming of cardiac gene expression produced by pressure overload. *Proc Natl Acad Sci U S A* 85: 339–343.
- Sassone-Corsi P, Verma IM (1987) Modulation of c-fos gene transcription by negative and positive cellular factors. *Nature* 326: 507–510.
- Sukhatme VP, Cao XM, Chang LC, Tsai-Morris CH, Stamenkovich D, et al. (1988) A zinc finger-encoding gene coregulated with c-fos during growth and differentiation, and after cellular depolarization. *Cell* 53: 37–43.
- Chavrier P, Zerial M, Lemaire P, Almendral J, Bravo R, et al. (1988) A gene encoding a protein with zinc fingers is activated during G0/G1 transition in cultured cells. *Embo J* 7: 29–35.
- Komuro I, Kaida T, Shibazaki Y, Kurabayashi M, Katoh Y, et al. (1990) Stretching cardiac myocytes stimulates protooncogene expression. *J Biol Chem* 265: 3595–3598.
- Iwaki K, Sukhatme VP, Shubeita HE, Chien KR (1990) Alpha- and beta-adrenergic stimulation induces distinct patterns of immediate early gene expression in neonatal rat myocardial cells. fos/jun expression is associated with sarcomere assembly; Egr-1 induction is primarily an alpha 1-mediated response. *J Biol Chem* 265: 13809–13817.
- Shubeita HE, McDonough PM, Harris AN, Knowlton KU, Glembocki CC, et al. (1990) Endothelin induction of inositol phospholipid hydrolysis, sarcomere assembly, and cardiac gene expression in ventricular myocytes. A paracrine mechanism for myocardial cell hypertrophy. *J Biol Chem* 265: 20555–20562.
- Ebisuya M, Yamamoto T, Nakajima M, Nishida E (2008) Ripples from neighbouring transcription. *Nat Cell Biol* 10: 1023–1024.
- Gupta MP, Gupta M, Zak R, Sukhatme VP (1991) Egr-1, a serum-inducible zinc finger protein, regulates transcription of the rat cardiac alpha-myosin heavy chain gene. *J Biol Chem* 266: 12813–12816.
- Zhao Y, Samal E, Srivastava D (2005) Serum response factor regulates a muscle-specific microRNA that targets Hand2 during cardiogenesis. *Nature* 436: 214–220.
- Kwon C, Han Z, Olson EN, Srivastava D (2005) MicroRNA1 influences cardiac differentiation in *Drosophila* and regulates Notch signaling. *Proc Natl Acad Sci USA* 102: 18986–18991.
- Zhao Y, Ransom JF, Li A, Vedantham V, von Drehle M, et al. (2007) Dysregulation of cardiogenesis, cardiac conduction, and cell cycle in mice lacking miRNA-1-2. *Cell* 129: 303–317.
- Ivey KN, Muth A, Arnold J, King FW, Yeh R-F, et al. (2008) MicroRNA regulation of cell lineages in mouse and human embryonic stem cells. *Cell Stem Cell* 2: 219–229.
- Chen JF, Mandel EM, Thomson JM, Wu Q, Callis TE, et al. (2006) The role of microRNA-1 and microRNA-133 in skeletal muscle proliferation and differentiation. *Nat Genet* 38: 228–233.
- Rao PK, Kumar RM, Farkhondeh M, Baskerville S, Lodish HF (2006) Myogenic factors that regulate expression of muscle-specific microRNAs. *Proc Natl Acad Sci U S A* 103: 8721–8726.
- Care A, Catalucci D, Felicetti F, Bonci D, Addario A, et al. (2007) MicroRNA-133 controls cardiac hypertrophy. *Nat Med* 13: 613–618.
- Liu N, Bezprozvannaya S, Williams AH, Qi X, Richardson JA, et al. (2008) microRNA-133a regulates cardiomyocyte proliferation and suppresses smooth muscle gene expression in the heart. *Genes Dev* 22: 3242–3254.
- van Rooij E, Sutherland LB, Qi X, Richardson JA, Hill J, et al. (2007) Control of stress-dependent cardiac growth and gene expression by a microRNA. *Science* 316: 575–579.
- Duisters RF, Tijssen AJ, Schroen B, Leenders JJ, Lentink V, et al. (2009) miR-133 and miR-30 regulate connective tissue growth factor: implications for a role of microRNAs in myocardial matrix remodeling. *Circ Res* 104: 170–178.
- Callis TE, Pandya K, Seok HY, Tang RH, Tatsuguchi M, et al. (2009) MicroRNA-208a is a regulator of cardiac hypertrophy and conduction in mice. *J Clin Invest* 119: 2772–2786.
- Yan D, Dong XD, Chen X, Wang L, Lu C, et al. (2009) MicroRNA-1/206 targets c-Met and inhibits rhabdomyosarcoma development. *J Biol Chem* 284: 29596–29604.
- Huang TH, Zhu MJ, Li XY, Zhao SH (2008) Discovery of porcine microRNAs and profiling from skeletal muscle tissues during development. *PLoS One* 3: e3225.
- van Rooij E, Sutherland LB, Liu N, Williams AH, McAnally J, et al. (2006) A signature pattern of stress-responsive microRNAs that can evoke cardiac hypertrophy and heart failure. *Proc Natl Acad Sci U S A* 103: 18255–18260.
- Wang D, Lu M, Miao J, Li T, Wang E, et al. (2009) Cepred: predicting the co-expression patterns of the human intronic microRNAs with their host genes. *PLoS One* 4: e4421.
- Kloosterman WP, Steiner FA, Berezikov E, de Bruijn E, van de Belt J, et al. (2006) Cloning and expression of new microRNAs from zebrafish. *Nucleic Acids Res* 34: 2558–2569.
- Landgraf P, Rusu M, Sheridan R, Sewer A, Iovino N, et al. (2007) A mammalian microRNA expression atlas based on small RNA library sequencing. *Cell* 129: 1401–1414.
- Liang Y, Ridzon D, Wong L, Chen C (2007) Characterization of microRNA expression profiles in normal human tissues. *BMC Genomics* 8: 166.
- Reddy AM, Zheng Y, Jagadeeswaran G, Macmill SL, Graham WB, et al. (2009) Cloning, characterization and expression analysis of porcine microRNAs. *BMC Genomics* 10: 65.
- van Rooij E, Quiat D, Johnson BA, Sutherland LB, Qi X, et al. (2009) A family of microRNAs encoded by myosin genes governs myosin expression and muscle performance. *Dev Cell* 17: 662–673.
- Bell ML, Buvoli M, Leinwand LA (2010) Uncoupling of Expression of an Intronic microRNA and its Myosin Host Gene by Exon Skipping. *Mol Cell Biol* 30: 1937–1945.
- Sluijter JP, van Mil A, van Vliet P, Metz CH, Liu J, et al. (2010) MicroRNA-1 and -499 Regulate Differentiation and Proliferation in Human-Derived Cardiomyocyte Progenitor Cells. *Arterioscler Thromb Vasc Biol* 30: 859–868.
- Matkovich SJ, Van Booven DJ, Youker KA, Torre-Amione G, Diwan A, et al. (2009) Reciprocal regulation of myocardial microRNAs and messenger RNA in human cardiomyopathy and reversal of the microRNA signature by biomechanical support. *Circulation* 119: 1263–1271.
- Ikedo S, Kong SW, Lu J, Bisping E, Zhang H, et al. (2007) Altered microRNA expression in human heart disease. *Physiol Genomics* 31: 367–373.
- Naga Prasad SV, Duan ZH, Gupta MK, Surampudi VS, Volinia S, et al. (2009) Unique microRNA profile in end-stage heart failure indicates alterations in specific cardiovascular signaling networks. *J Biol Chem* 284: 27487–27499.
- Lyons GE, Schiaffino S, Sassoon D, Barton P, Buckingham M (1990) Developmental regulation of myosin gene expression in mouse cardiac muscle. *J Cell Biol* 111: 2427–2436.
- Subramaniam A, Jones WK, Gulick J, Wert S, Neumann J, et al. (1991) Tissue-specific regulation of the alpha-myosin heavy chain gene promoter in transgenic mice. *J Biol Chem* 266: 24613–24620.
- Molkentin JD, Robbins J (2009) With great power comes great responsibility: using mouse genetics to study cardiac hypertrophy and failure. *J Mol Cell Cardiol* 46: 130–136.
- Hagiwara N, Yeh M, Liu A (2007) Sox6 is required for normal fiber type differentiation of fetal skeletal muscle in mice. *Dev Dyn* 236: 2062–2076.
- McCarthy JJ, Esser KA, Peterson CA, Dupont-Versteegden EE (2009) Evidence of MyomiR network regulation of beta-myosin heavy chain gene expression during skeletal muscle atrophy. *Physiol Genomics* 39: 219–226.
- Komuro I, Kurabayashi M, Takaku F, Yazaki Y (1988) Expression of cellular oncogenes in the myocardium during the developmental stage and pressure-overloaded hypertrophy of the rat heart. *Circ Res* 62: 1075–1079.
- Rozich JD, Barnes MA, Schmid PG, Zile MR, McDermott PJ, et al. (1995) Load effects on gene expression during cardiac hypertrophy. *J Mol Cell Cardiol* 27: 485–499.
- Saadane N, Alpert L, Chalifour LE (2000) Altered molecular response to adrenergic-induced cardiac hypertrophy in Egr-1-deficient mice. *Am J Physiol Heart Circ Physiol* 278: H796–805.
- Buitrago M, Lorenz K, Maass AH, Oberdorf-Maass S, Keller U, et al. (2005) The transcriptional repressor Nabl1 is a specific regulator of pathological cardiac hypertrophy. *Nat Med* 11: 837–844.
- Parlakian A, Charvet C, Escoubet B, Mericskay M, Molkentin JD, et al. (2005) Temporally controlled onset of dilated cardiomyopathy through disruption of the SRF gene in adult heart. *Circulation* 112: 2930–2939.
- Liu L, Tsai JC, Aird WC (2000) Egr-1 gene is induced by the systemic administration of the vascular endothelial growth factor and the epidermal growth factor. *Blood* 96: 1772–1781.
- Matkovich SJ, Wang W, Tu Y, Eschenbacher WH, Dorn LE, et al. (2009) MicroRNA-133a protects against myocardial fibrosis and modulates electrical repolarization without affecting hypertrophy in pressure-overloaded adult hearts. *Circ Res* 106: 166–175.
- John B, Enright AJ, Aravin A, Tuschl T, Sander C, et al. (2004) Human MicroRNA targets. *PLoS Biol* 2: e363.
- Lewis BP, Burge CB, Bartel DP (2005) Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 120: 15–20.
- Wang X, El Naqa IM (2008) Prediction of both conserved and nonconserved microRNA targets in animals. *Bioinformatics* 24: 325–332.

57. Rottman JN, Ni G, Brown M (2007) Echocardiographic evaluation of ventricular function in mice. *Echocardiography* 24: 83–89.
58. Tarnavski O, McMullen JR, Schinke M, Nie Q, Kong S, et al. (2004) Mouse cardiac surgery: comprehensive techniques for the generation of mouse models of human diseases and their application for genomic studies. *Physiol Genomics* 16: 349–360.
59. Sharpe J, Ahlgren U, Perry P, Hill B, Ross A, et al. (2002) Optical projection tomography as a tool for 3D microscopy and gene expression studies. *Science* 296: 541–545.
60. Huang da W, Sherman BT, Lempicki RA (2009) Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc* 4: 44–57.
61. Huang da W, Sherman BT, Lempicki RA (2009) Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. *Nucleic Acids Res* 37: 1–13.