

## **UC Davis**

### **UC Davis Previously Published Works**

#### **Title**

Geography of speciation affects rate of trait divergence in haemulid fishes

#### **Permalink**

<https://escholarship.org/uc/item/08t7h9ft>

#### **Journal**

Proceedings of the Royal Society B, 286(1896)

#### **ISSN**

0962-8452

#### **Authors**

Tavera, José J  
Wainwright, Peter C

#### **Publication Date**

2019-02-13

#### **DOI**

10.1098/rspb.2018.2852

Peer reviewed

## Research



**Cite this article:** Tavera JJ, Wainwright PC. 2019 Geography of speciation affects rate of trait divergence in haemulid fishes. *Proc. R. Soc. B* **286**: 20182852. <http://dx.doi.org/10.1098/rspb.2018.2852>

Received: 14 December 2018

Accepted: 23 January 2019

**Subject Category:**

Evolution

**Subject Areas:**

evolution

**Keywords:**

sympatry, allopatry, haemulidae, character displacement, trait evolution

**Author for correspondence:**

José J. Tavera

e-mail: [jose.tavera@correounivalle.edu.co](mailto:jose.tavera@correounivalle.edu.co)

Electronic supplementary material is available online at <https://dx.doi.org/10.6084/m9.figshare.c.4397888>.

# Geography of speciation affects rate of trait divergence in haemulid fishes

José J. Tavera<sup>1</sup> and Peter C. Wainwright<sup>2</sup>

<sup>1</sup>Departamento de Biología, Universidad del Valle, Cali, Colombia

<sup>2</sup>Department of Evolution and Ecology, University of California, Davis, CA 95616, USA

JJT, 0000-0003-4517-9238

Speciation and the interactions between recently diverged species are thought to be major causes of ecological and morphological divergence in evolutionary radiations. Here, we explore the extent to which geographical overlap and time since speciation may promote divergence in marine species, which represent a small fraction of currently published studies about the patterns and processes of speciation. A time-calibrated molecular phylogeny of New World haemulid fishes, a major radiation of reef and shore fishes in the tropical West Atlantic and East Pacific, reveals 21 sister species pairs, of which eight are fully sympatric and 13 are allopatric. Sister species comparisons show a non-significant relation between most of the phenotypic traits and time since divergence in allopatric taxa. Additionally, we find no difference between sympatric and allopatric pairs in the rate of divergence in colour pattern, overall body shape, or functional morphological traits associated with locomotion or feeding. However, sympatric pairs show a significant decrease in the rate of divergence in all of these traits with increasing time since their divergence, suggesting an elevated rate of divergence at the time of speciation, the effect of which attenuates as divergence time increases. Our results are consistent with an important role for geographical overlap driving phenotypic divergence early in the speciation process, but the lack of difference in rates between sympatric and allopatric pairs indicates that the interactions between closely related species are not dominant drivers of this divergence.

## 1. Background

The relation between present-day patterns of biodiversity and evolutionary processes (i.e genetic divergence, speciation, extinction, range-size variation) has generated major discussion points among evolutionary biologists and ecologists [1–5]. Many studies [4–10] have documented the connection of geographical range with speciation and extinction in marine taxa, and yet marine groups are under-represented, comprising only a small fraction of all currently published studies about the patterns and processes of speciation [11].

It has been recognized that speciation works on ecological and evolutionary timescales not easily captured by any single approach [12–15]. Advancements in genetic techniques and evolutionary models have allowed biological studies to understand mechanisms and processes involved in the isolation of populations and whether these groups can become sufficiently isolated to become species or regain genetic connectivity in the marine realm [16–20]. Additionally, genetic sequence data are now used to generate robust phylogenies of marine lineages leading to an increasing interest in using these data to investigate the role of geography in marine speciation [21–25].

The relative strength of biological and geographical isolating mechanisms can vary over the course of speciation and between different modes of speciation [15]. Owing to the expected importance of character displacement and reinforcement, models of speciation predict a strong effect of geography on the evolution of traits associated with reproductive isolation. Sister species and very closely related species that occur in sympatry may experience

diversifying selection on mate choice traits, patterns of resource use, and traits that underlie the ability to make use of resources [26,27]. Indeed, models of sympatric speciation rely on a linkage between assortative mating and disruptive selection on niche traits [28–30].

More generally, there are a number of reasons why speciation may be a period in lineage history with elevated rates of trait evolution [26,31,32]. Whether adapting to a new region following a dispersal event, or in the absence of gene flow following vicariance, isolation promotes adaptive adjustment to the current geographical distribution. But while palaeontologists have explored patterns of change in rates of morphological evolution through much deeper time than seen in contemporary ecology, it has proven difficult to compare the impact of speciation versus other factors in contributing to the rates of phenotypic change in recent evolutionary radiations (e.g. [33]).

In this study, we ask how the range overlap affects rates of trait evolution in sibling species of New World haemulid fishes, a prominent group of marine shorefishes that has undergone a substantial radiation on coral reefs of the Caribbean and Eastern Pacific regions. Our recent development of a complete phylogeny of New World haemulids reveals 21 sister species pairs, with estimates of the divergence time for each pair. For each sister pair, we estimate trophic overlap and the rate of divergence of trophic morphology, locomotor morphology, overall body shape, and colouration. Range overlap is strongly bimodal with eight pairs of fully sympatric sister species and 13 allopatric pairs. Given our expectation that rates of trait evolution are high during speciation, we predict and test for a negative relationship between the rate of trait divergence and time since divergence between sister species. We also ask whether rates of divergence are affected by range overlap, predicting that sympatric pairs will experience higher rates of divergence.

## 2. Methods

The phylogeny used in this study was pruned from a larger phylogeny of 107 haemulid taxa and 44 out-group species [34]. Thus, the present study focused on the phylogeny of the monophyletic radiation of all 61 species of haemulids that occur in the Western Atlantic and East Pacific (figure 1). This phylogeny was constructed from a dataset of three mitochondrial and four nuclear genes, and extends results from previous studies [35,36]. For methodological details on phylogenetic reconstruction, see [34]. Specimens were collected throughout their range including regions of sympatry. Preserved specimens were stored and measured at the collections of Centro Interdisciplinario de Ciencias Marinas, Centro de Investigaciones Biológicas del Noroeste, and California Academy of Sciences.

### (a) Species pairs

Our approach was to measure trait divergence between sister species, and using the estimated divergence time, we calculated the rate of trait evolution. The phylogenetic analysis revealed 21 sister species pairs. For each of these 42 species, we measured geographical range, habitat preference, diet, body shape, colouration, and a set of functional morphological traits associated with feeding and locomotion.

### (b) Geographical overlap

Species ranges and distribution records were obtained from published sources [13–24]. Ranges were drawn onto equal area maps using the free geographic information system (GIS) software

GRASS (Geographic Resources Analysis Support System). These maps were not intended to be highly exhaustive reconstructions of species distributions; rather they are approximations, which we used to calculate species geographical overlay. Note that distributions in this context may be seen as large-scale overlaps; however, we confirmed that all sympatric species pairs identified in this study co-occur at the microhabitat scale and have been seen during our dives within a few metres of each other. Range overlap was defined as the percentage of the more restricted species range overlapped by its more widespread sister [37]. Species were coded allopatric if ranges did not overlap or if overlap was less than 20%, whereas if species ranges overlapped more than 90% then they were categorized as sympatric.

### (c) Geography of speciation

We explored the ‘age–range correlation’ for the New World haemulids, using our time-calibrated phylogeny and the values of range overlap. We plotted range overlap against time since divergence for the 21 sister pairs. Range overlap for the deeper nodes combined areas of species distributions and were calculated following the range merging procedure as in Barraclough & Vogler [38]. If a significant age–range relationship is present, then younger pairs are expected to be informative on the geography of speciation. Intercept values of 0.5 or higher and a negative slope are indicative of a predominantly sympatric model. Whereas an intercept of less than 0.5 and a positive slope suggests that the most frequent mode of speciation is allopatric [39,40].

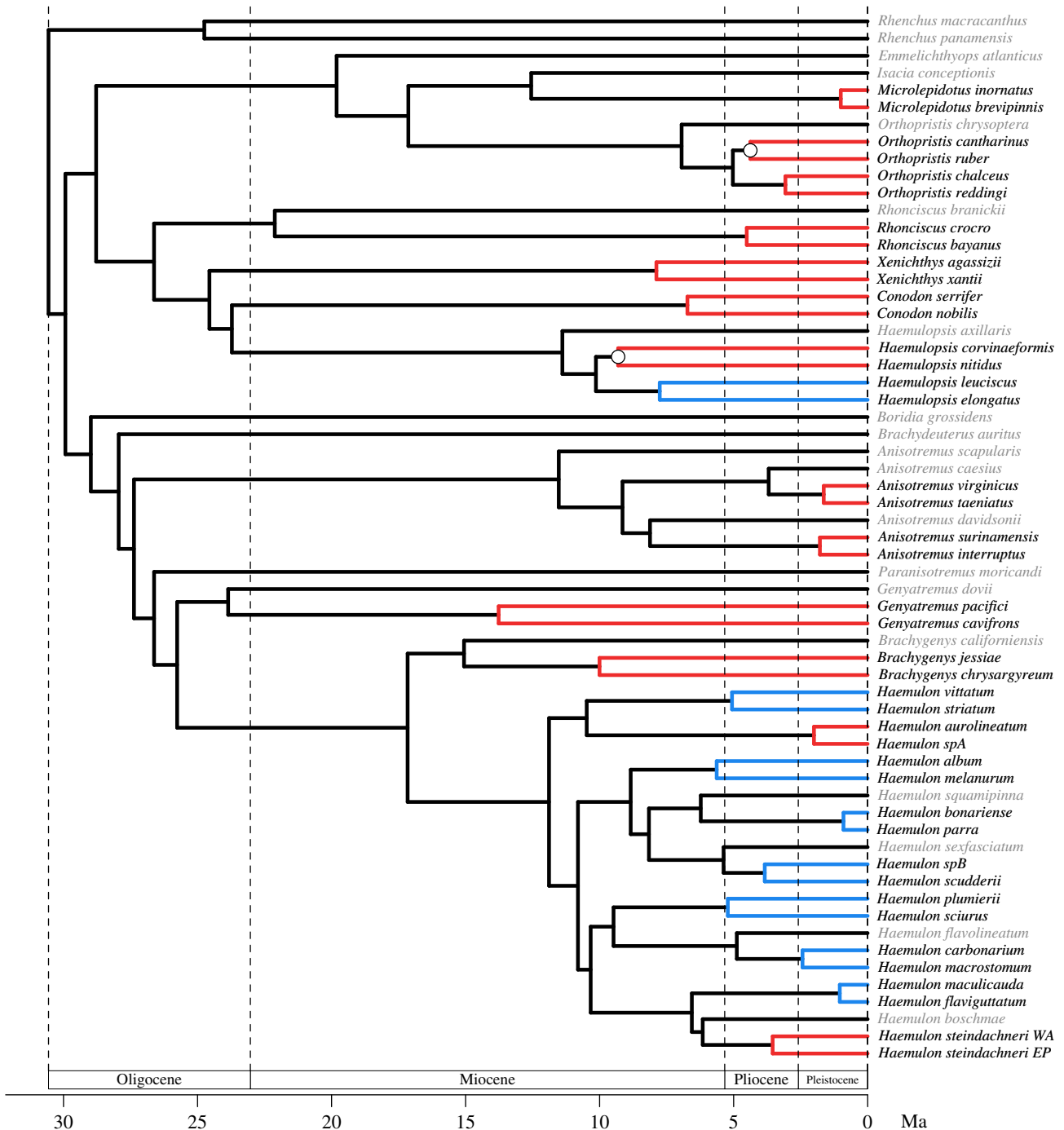
### (d) Niche overlap

To estimate niche overlap between sister species, we compiled a dataset of ecological traits using published information [41–50]. We focused on two main axes of niche differentiation: habitat and diet. Habitat included nine categories of habitat substrate type. Dietary overlap was quantified from the use of 13 discrete categories for prey items. Both diet and habitat overlap were calculated as the number of items shared by the two species divided by the sum of total unique items included.

### (e) Morphological and functional traits

We quantified body shape using geometric morphometric methods. We selected 23 anatomical landmarks and 13 semilandmarks, the latter describing head profile and lateral line contour. The  $x,y$  coordinates of these landmarks were collected from digital lateral-view photographs of three to five adult individuals per species. A total of 183 pictures of 61 haemulid species were used. In geometric morphometrics, landmarks are analysed assuming biological homology [51,52]. Semilandmarks were used to represent homologous curves by sets of points, establishing a geometric homology. Landmark configurations for each specimen were aligned using a generalized Procrustes superimposition and average species shape was calculated using the Geomorph [53] package. TPSrelw v. 1.5.1 was used to perform a relative warp analysis. This corresponds to a principal component analysis (PCA) of variation in shape. Finally, a matrix of scores of all species in each of the relative warps was generated.

Additionally, we analysed a set of functional traits related to feeding (11 traits) and locomotion (13 traits) [54]. All linear measurements were log transformed, while masses were cube root transformed prior to analysis. We performed a phylogenetic size correction and phylogenetic PCA [55] to reduce dimensionality of each dataset. The broken stick method was used to determine how many principal components were retained for further analysis. All dataset manipulations and statistics were performed in R (R Development Core Team 2012) using the *ape* [56], *geiger* [57], and *phytools* [53] libraries.



**Figure 1.** Time-calibrated maximum clade credibility tree of New World haemulids [34]. Unlabelled nodes correspond to 1 in posterior probability. White circles indicate 0.75–0.95 posterior probability. Red and blue branches refer to allopatric and sympatric pairs, respectively.

### (f) Body colouration

We selected one photograph per species from which to measure colouration and digitized 73 anatomical landmarks and semilandmarks using TPSDig v. 1.2.1. Haemulids are not known to show sexual dichromatism. The landmarks captured the overall body shape of the species. We superimposed all of these images, fitted all images to the consensus configuration, and then averaged the unwarped images. This procedure resulted in identical (shape and size) images for each species. These images were modified in Photoshop by averaging pixel colours using a pre-determined palette, based on dominant haemulid colours. Each pixel on the body had a numerical value corresponding to a combination of three independently stored colour channels: red, green, and blue. These three matrices were extracted and concatenated. Colour extraction and manipulation of edited images was done in R with the pixmap library.

### (g) Standardized contrasts

We used phylogenetic standardized contrasts of trait values between sister species to estimate the rate of trait evolution [58,59]. Contrasts were calculated for body colouration and the first phylogenetic principal component (PC) of body shape, trophic traits, and locomotor traits using 1000 trees sampled randomly from the posterior distribution of time-calibrated phylogenies. Contrasts were calculated as the Euclidean distance between species in the trait space and standardized by dividing by the square root of the sum of the branch lengths (time) separating the species [60]. The variance of standardized contrasts is an estimate of the Brownian rate parameter for that trait on the phylogeny when the branch lengths are in units of time [59] and thus individual standardized contrasts estimate the rate of trait evolution across the section to the tree they are calculated on. We used a linear regression to test for a significant

relationship between standardized contrasts and the divergence time between sister species among sympatric and allopatric pairs separately. We used ANOVA to test for overall differences between allopatric and sympatric pairs in the rate of trait evolution. To account for phylogenetic uncertainty and uncertainty in divergence times, all analyses were performed on a set of 1000 trees sampled from the posterior distribution of trees (electronic supplementary material). However, results and statistics reported here are based on the maximum clade credibility tree.

### 3. Results

Overall, the combined molecular data yielded a robust phylogenetic hypothesis with strong support (Bayesian posterior = 1.0) for more than two-thirds of all nodes. Divergence times between sister species exhibit a range of ages from a maximum of 17.5–10.3 Ma between *Genyatremus cavifrons* and *G. pacifici* to 1.6–0.3 Ma in *Haemulon bonariense* and *H. parra*.

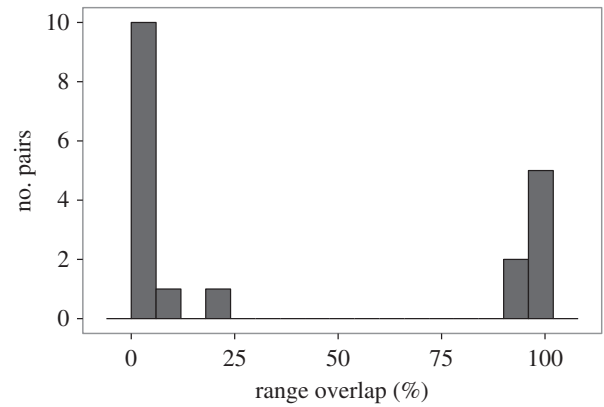
#### (a) Geography of speciation

Twenty-one pairs of sister species were identified with only three of them being supported by less than 0.95 posterior probability. Two distinct clusters of pairs were found in degree of range overlap (figure 2). Among the 13 allopatric pairs, seven were geminate pairs across the Panamanian isthmus and two previously unidentified pairs each include one member from the Galapagos Islands and one from the Caribbean.

A regression of arcsine-transformed range overlap against time since divergence performed on the entire phylogeny was significant; however, the adjusted  $R^2$  was extremely low (adjusted  $R^2 = 0.085$ ;  $F$ -statistic: 6.385 on 1 and 57 d.f.,  $p$ -value = 0.01). The intercept of  $0.59 \pm 0.025$  s.e. was not significantly different from 0.5, with most of the recent pairs showing an even distribution between 0 and 1 in range overlap. A significant positive slope of 0.026 was found. Under an allopatric model, sympatry tends to increase with node age, suggesting that range changes have occurred with increasing times since divergence in this group. Since the intercept is not different from 0.5 (indicative of sympatry) but the slope is slightly positive (indicative of allopatry), we consider this result inconclusive on the issue of geography during species formation. However, a plot of range overlap against time of divergence resembled closely the simulated data under a null model of random ranges and 50% sympatry [9].

Among the 13 allopatric pairs, 11 showed zero range overlap, with the Panamanian isthmus being the most common geographical barrier, separating six geminate pairs. These species pairs ranged in age from 2.95 to 14.12 Ma. An interesting pattern can be seen in two Galapagos endemics each of which has a sister species in the Western Atlantic and not in the adjacent eastern Pacific as might be expected. Further discussion can be found in Tavera *et al.* [34].

Seven of the eight sympatric pairs are found within *Haemulon*. From these seven, five are broadly distributed throughout the Western Atlantic from eastern USA south to Brazil; including the Gulf of Mexico and the Caribbean and the remaining two pairs are distributed through the eastern Pacific, from Mexico to Peru. Time since divergence for these seven pairs range from 0.946 to 5.68 Ma (95% highest posterior density (HPD): 0.377–7.9682 Ma) (table 1).



**Figure 2.** Proportion of range overlap in haemulid sister species.

The average age of the allopatric pairs (5.53 Ma; s.d. 3.94) was not significantly different from that of the sympatric pairs (4.04 Ma; s.d. 2.44) as indicated by the Welch two-sample  $t$ -test ( $t = 1.066$ , d.f. = 18.98,  $p$ -value = 0.3) in spite of a greater overall range of allopatric ages: allopatric pairs ranging from 1 to 14.2 Ma (95% HPD: 0.48–17.56 Ma) versus 1.8 to 7.8 Ma (95% HPD: 0.37–10.37 Ma) for the sympatric pairs. Results were consistent after running the test independently on 1000 trees sampled from the posterior distribution and after removing two pairs that did not have full posterior probability support (see electronic supplementary material).

#### (b) Geography and ecology

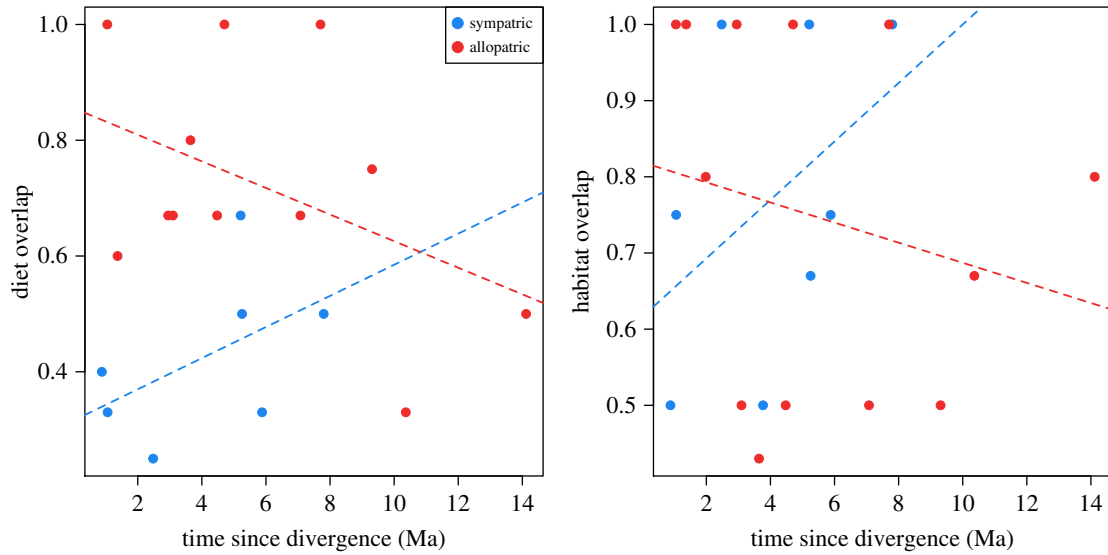
Diet overlap was significantly lower in sympatric pairs (mean = 0.43) than allopatric pairs (mean = 0.72) (Welch two-sample  $t$ -test;  $t = 3.3$ , d.f. = 17,  $p$ -value = 0.0038) (figure 3). However, there was no relationship between diet overlap and age of divergence within either sympatric or allopatric pairs or when the groups were combined (maximum clade credibility tree: allopatric pairs:  $p$ -value: 0.151; sympatric pairs:  $p$ -value: 0.2533). The same pattern was found for habitat overlap; however, the latter did not show significant differences between the two groups (Welch two-sample  $t$ -test;  $t = -0.24$ , d.f. = 17,  $p$ -value = 0.80) or relationship with age of divergence (maximum clade credibility tree: allopatric  $p$ -value = 0.44; sympatric  $p$ -value = 0.29) (figure 3). These results were confirmed when analyses were repeated on the 1000 trees from the posterior distribution (see electronic supplementary material, figure S3).

#### (c) Geography and phenotypic evolution

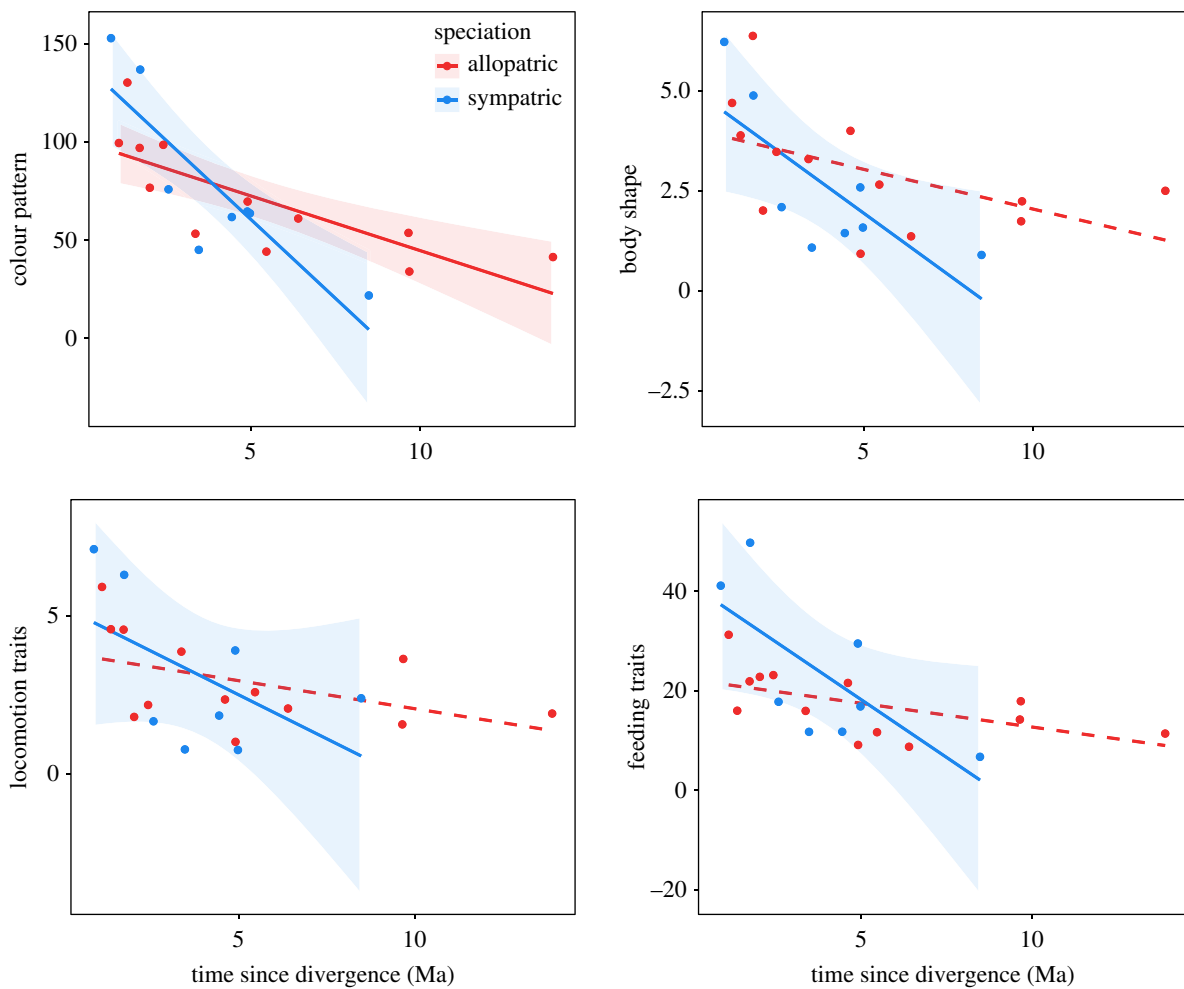
Contrasts for body colouration PC1, body shape PC1, locomotion PC1, and feeding PC1 all decreased significantly with divergence time in sympatric pairs ( $p$ -values: 0.00027; 0.0028; 0.039; 0.019, respectively) (figure 4). Only body colouration contrasts showed a significant relationship with divergence time in the allopatric pairs ( $p = 0.0032$ ), although the trend was towards decreasing contrasts with age across allopatric pairs in all traits. Declining rates of trait evolution with time since divergence in the sympatric pairs are consistent with speciation involving a period of enhanced rates of trait evolution. The ANOVA indicated no overall difference in rates of trait evolution between sympatric and allopatric pairs for any of the trait categories.

**Table 1.** Sister pairs included in this study, including mean time since divergence (MTSD), 95% of credible interval, habitat overlap, diet overlap, and standardized values of contrast for the different axis of differentiation. Distribution codes are as follows: EP, Eastern Pacific; GC, Gulf of California; GI, Galapagos Islands; GM, Gulf of Mexico; WA, Western Atlantic.

pairs	speciation	distribution	95% CI age	MTSD	habitat overlap	diet overlap	standardized phylogenetic contrasts			
							colour	shape	locomotion	feeding
<i>Anisotremus surinamensis/A. interruptus</i>	allopatric	WA/EP	1.123–2.677	1.824	1	0.6	130.120	15.997	3.882	4.566
<i>A. virginicus/A. taeniatus</i>	allopatric	WA/EP	0.969–2.408	1.669	1	0.67	96.855	21.904	6.370	4.553
<i>Brachygenys chrysargyreum/B. jessiae</i>	allopatric	WA/GI	7.104–13.427	10.057	0.67	0.33	33.921	17.904	2.234	3.625
<i>Conodon nobilis/C. serrifer</i>	allopatric	WA/EP	3.986–10.397	6.899	0.5	0.67	60.908	8.743	1.358	2.057
<i>Genyatremus cavifrons/G. pacifici</i>	allopatric	WA/EP	10.328–17.561	13.876	0.8	0.5	41.294	11.398	2.497	1.898
<i>Haemulon aurolineatum/H. spA</i>	allopatric	WA/GM	1.197–3.043	2.056	0.8	NA	98.413	23.161	3.471	2.169
<i>H. steindachneri/H. steindachneri</i>	allopatric	WA/EP	2.136–5.139	3.605	0.43	0.8	69.549	9.111	0.923	1.003
<i>Haemulopsis corvinaeformis/Hae. Nitidus</i>	allopatric	WA/EP	6.863–12.125	9.350	0.5	0.75	53.608	14.223	1.735	1.554
<i>Microlepidotus inornatus/M. brevipinnis</i>	allopatric	EP/GC	0.475–1.707	1.049	1	1	99.344	31.279	4.693	5.904
<i>Orthopristis chalcus/O. reddingi</i>	allopatric	EP/GC	1.797–4.409	3.107	0.5	0.67	76.576	22.812	2.004	1.792
<i>O. ruber/O. cantharinus</i>	allopatric	WA/GI	2.879–6	4.426	0.5	0.67	NA	21.586	3.996	2.342
<i>Rhombiscus croca/ R. bayanus</i>	allopatric	WA/EP	2.629–7.296	4.678	1	1	53.180	15.961	3.291	3.857
<i>Xenichthys xantii/X. agassizii</i>	allopatric	EP/GI	4.816–11.269	8.022	1	1	44.064	11.663	2.652	2.573
<i>Haemulon album/H. melanurum</i>	sympatric	WA	3.697–7.968	5.684	0.75	0.33	63.568	16.852	1.578	0.744
<i>H. bonariense/H. parra</i>	sympatric	WA	0.377–1.597	0.946	0.5	0.4	152.711	41.163	6.219	7.098
<i>H. carbonarium/H. macrostomum</i>	sympatric	WA	1.368–3.627	2.484	1	0.25	75.765	17.794	2.088	1.652
<i>H. flaviguttatum/H. maculicauda</i>	sympatric	EP	0.464–1.888	1.099	0.75	0.33	136.724	49.800	4.879	6.286
<i>H. plumieri/H. sciurus</i>	sympatric	WA	3.49–7.237	5.264	1	0.67	61.652	11.765	1.440	1.833
<i>H. scudderii/H. spB</i>	sympatric	EP	2.345–5.661	3.876	0.5	NA	44.958	11.745	1.077	0.763
<i>H. striatum/H. vittatum</i>	sympatric	WA	3.247–7.233	5.142	0.67	0.5	64.489	29.516	2.582	3.894
<i>Haemulopsis elongatus/Hae. Leudiscus</i>	sympatric	EP	5.541–10.37	7.815	1	0.5	21.745	6.716	0.893	2.381



**Figure 3.** Standardized contrast (calculated on the maximum clade credibility tree) of the diet and habitat overlap in haemulid sister species pairs, plotted against time since divergence. Dashed lines indicate a non-significant linear relationship with time since divergence. Diet significance values are as follows: allopatric pairs:  $p$ -value: 0.151; sympatric pairs:  $p$ -value: 0.2533. Habitat significance values: allopatric pairs  $p$ -value = 0.44; sympatric pairs  $p$ -value = 0.29. (Online version in colour.)



**Figure 4.** Standardized contrast (calculated on the maximum clade credibility tree) of the different axes of differentiation plotted against age of sister species pairs. The Y-axis corresponds to contrast values between sister species and the X-axis represents time since divergence measured in million years. Solid lines mean significant linear relationship (error ranges correspond to 95% confidence intervals). Dashed lines without error ranges indicate that the relationship was not significant. Allopatric pairs  $p$ -values for body colouration, body shape, locomotion traits, and feeding traits: (0.0033; 0.14; 0.17; 0.16), sympatric pairs  $p$ -values: (0.00027; 0.0028; 0.039; 0.019). (Online version in colour.)

## 4. Discussion

A number of major macroevolutionary issues revolve around the temporal pattern of evolutionary change through the history of lineages. For many reasons, it is expected that the rate of ecological and phenotypic evolution will not be constant through time as both the biotic and abiotic factors that cause niche and trait evolution likely change in response to a wide range of time-dependent factors. Phylogenies and comparative methods allow one to evaluate variation in the rate of trait change across speciation events in the phylogeny, but it has been difficult to evaluate changes in the pace of trait change during the time between speciation events [61]. By focusing on species pairs of varying time since divergence, we find evidence that the time period near speciation exhibits elevated rates of trait evolution in sympatric sister species of New World haemulids. The inferred rate of evolution declined with increasing time since speciation, consistent with an interpretation in which sympatric species diverge most rapidly during or immediately following speciation, and thus have lower net rates of divergence the longer they have been separated.

Although the present-day geography of sister species is not conclusive evidence of their geography during speciation, we found no overall age difference between sympatric and allopatric sister species, suggesting that our results are not an artefact of an increasing probability of range overlap among older sister species pairs [27]. The observed level of sympatry (38%) among New World grunt sister taxa was within the range reported from other animals (33–45% mammalian; 35% flycatchers; 50% *Drosophila*; 50% marine invertebrates) but low if compared to some other coral reef fishes (64%) and with plants (80%) [62–64]. This result is consistent with the view that allopatric speciation is frequent in the sea, even in organisms with wide dispersal capabilities [15,65]. The importance of biogeographic barriers in separating sister species in coral reef fishes has been extensively studied [63,66–73] demonstrating that allopatric speciation is an important engine in marine fishes diversification, although it appears not to be the only mode of speciation in the sea [63,74–76] as sometimes suggested [77]. Sympatric speciation, on the other hand, has always been contentious. Nevertheless, it has been demonstrated as one of the most probable modes of speciation for many marine fishes [74,76,78] and marine invertebrates [62]. We found two sympatric species pairs to be younger than or close to 1 Ma, which disagrees with previous suggestions that 2 Ma is the minimum time required for sister species to develop sympatric distributions following allopatric divergence in hermit crabs [79] and 4 Ma in other coral reef fishes [80]. Indeed, Quenouille *et al.* [80] found that after speciation, some species expanded their range and became sympatric, while others appear to have experienced limited or no range expansion, with the second of these dynamics being the most frequent. However, hybridization and introgression cannot be easily ruled out in these young pairs which is expected to confound attempts to reconstruct the history of divergence [81].

Shallow marine environments, such as the coral reefs and near shore habitats inhabited by haemulids, move with a frequency that is much greater than the rate of new species formation. As others have pointed out [11], global warming and cooling cycles have changed sea levels by as much as 150 m several times in the Pleistocene, meaning that the

species ranges will have had to move with the moving habitats and, thus, present distribution may be a relatively poor predictor of the geography of populations at the time of species formation.

The rapid phenotypic divergence of young sympatric sister species is consistent with a role for disruptive selection and character displacement acting during time periods close to divergence. Further, the very young age of some of the sympatric sister species pairs in our study is suggestive of the possibility that these species were formed in sympatry, in which case, disruptive selection on ecological traits is clearly expected [82]. Finding this pattern in functional morphological traits involved in feeding and locomotion, as well as overall body shape and body colouration further suggests that ecological differentiation in sympatric species is most intense during and shortly after speciation. Our results differ from observations in some other groups, for example, there was a dominance of similarity in ecological and reproductive traits found in closely related but sympatric plant species within the California Floristic Province [64,83].

Interestingly, although sympatric sister species appear to undergo elevated rates of phenotypic divergence at the time of speciation, the overall rate of divergence between pairs was not generally greater in sympatric pairs than allopatric pairs. Perhaps, allopatric ranges encompass a greater diversity of environments, and so some diversity of selective pressures, which might increase adaptive responses and rates of divergence [84]. While sympatric species pairs showed less diet overlap than allopatric pairs, our study does not find strong evidence that sympatry of sister species results in elevated rates of phenotypic divergence. Allopatric pairs showed a trend towards declining rates of divergence since speciation, similar to that seen in the sympatric pairs, but this relationship was only significant in colour pattern. The combination of decreasing rates of divergence with time in sympatry and no difference in rate of divergence between allopatric and sympatric pairs suggests that ecological interactions between sister species may not be a dominant cause of rates of trait evolution in young sister pairs. Nevertheless, this conclusion is tentative because of the relatively low number of young allopatric and sympatric pairs. Indeed, the highest rates of divergence in colour pattern, the overall body shape, and feeding traits were seen in the youngest sympatric pairs, suggesting that a larger sample of these young pairs may confirm an enhanced rate of divergence in sympatry.

Our results partially agree with those of Pitteloud *et al.* [85] in *Pyrgus* butterflies in which they found higher rates of niche evolution along climatic dimensions for sister lineages that diverge in sympatry. Haemulids are an ecologically diverse group of reef and shore fishes in the tropical Western Atlantic and the Eastern Pacific. The feeding ecology of species ranges from benthic sediment sifting to mollusc crushing, piscivory, and mid-water zooplankton feeding. Trophic ecology is reflected in functional morphological features of the feeding apparatus and the locomotor system [34,54]. We find the lowest diet overlap in the youngest sympatric sister species, indicating that these lineages diverged rapidly in patterns of prey use (figure 4), in association with the highest inferred rates of divergence in feeding traits and among the highest rates in locomotion traits (figure 4). These results are consistent with an important role for ecological interactions in shaping early patterns of



divergence in resource use and morphology of recently diverged sympatric sister species of New World haemulid fishes.

## 5. Conclusion

Although our study does not support differences in the rate of divergence between allopatric and sympatric pairs, we found that recently diverged sympatric pairs exhibit higher levels of diet divergence and higher rates of trait evolution, diverging most rapidly during or immediately following speciation. This pattern is consistent with a role for disruptive selection and character displacement during these speciation events. Whether these cases represent *in situ* sympatric speciation or secondary contact remains unsettled, but the results suggest that in these species pairs that are presently

sympatric, particularly rapid trait evolution occurred early in their divergence.

**Data accessibility.** The dataset supporting this article is available in the Dryad Digital Repository: <https://doi.org/10.5061/dryad.5qd0gd0> [86] and the R code has been included as part of the electronic supplementary material.

**Authors' contributions.** J.J.T. and P.C.W. conceived and designed the study, made fish measurements, and drafted the manuscript. J.J.T. collected specimens, carried out the molecular laboratory work, and analyses.

**Competing interests.** The authors have no competing interests.

**Funding.** This research was supported by NSF grant no. DEB-1061981 and IOS-0924489 to P.C.W. and Universidad del Valle project CI 71028 to J.J.T.

**Acknowledgements.** We thank Arturo Acero and many other people for assistance during fieldwork.

## References

- Darwin C. 1859 *On the origin of species by means of natural selection*. London, UK: John Murray.
- Wallace AR. 1855 On the law which has regulated the introduction of new species. *Ann. Mag. Nat. Hist.* **16**, 184–196. (doi:10.1080/037454809495509)
- Mayr E. 1963 *Animal species and evolution*. London, UK: Harvard University.
- Jablonski D. 1994 Extinctions in the fossil record. *Phil. Trans. R. Soc. Lond. B* **344**, 11–17. (doi:10.1098/rstb.1994.0045)
- Jablonski D, Roy K, Valentine JW. 2003 *Evolutionary macroecology and the fossil record*. Cambridge, UK: Cambridge University Press.
- Reynolds JD, Dulvy NK, Goodwin NB, Hutchings JA. 2005 Biology of extinction risk in marine fishes. *Proc. R. Soc. B* **272**, 2337–2344. (doi:10.1098/rspb.2005.3281)
- Gaston K, Blackburn T. 2008 *Pattern and process in macroecology*. Oxford, UK: Blackwell Science.
- Giam X, Ting HN, Lok AFSL, Hoek HN. 2011 Local geographic range predicts freshwater fish extinctions in Singapore. *J. Appl. Ecol.* **48**, 356–363. (doi:10.1111/j.1365-2664.2010.01953.x)
- Friedman M, Sallan LC. 2012 Five hundred million years of extinction and recovery: a Phanerozoic survey of large-scale diversity patterns in fishes. *Paleontology* **55**, 707–742. (doi:10.1111/j.1475-4983.2012.01165.x)
- Burgess MG, Costello C, Fredston-Hermann A, Pinsky ML. 2017 Range contraction enables harvesting to extinction. *Proc. Natl Acad. Sci. USA* **114**, 3945–3950. (doi:10.1073/pnas.1607551114)
- Miglietta MP, Faucci A, Santini F. 2011 Speciation in the sea: overview of the symposium and discussion of future directions. *Integr. Comp. Biol.* **51**, 449–455. (doi:10.1093/icb/ict024)
- Simpson GG. 1944 *Tempo and mode in evolution*. New York, NY: Columbia University Press.
- Jablonski D. 2008 Biotic interactions and macroevolution: extensions and mismatches across scales and levels. *Evolution* **62**, 715–739. (doi:10.1111/j.1558-5646.2008.00317.x)
- Reznick DN, Ricklefs RE. 2009 Darwin's bridge between microevolution and macroevolution. *Nature* **457**, 837–842. (doi:10.1038/nature07894)
- Norris RD, Hull PM. 2012 The temporal dimension of marine speciation. *Evol. Ecol.* **26**, 393–415. (doi:10.1007/s10682-011-9488-4)
- Williams ST, Benzie JAH. 1997 Indo-West Pacific patterns of genetic differentiation in the high-dispersal starfish *Linckia laevigata*. *Mol. Ecol.* **6**, 559–573. (doi:10.1046/j.1365-294X.1997.00221.x)
- Williams ST. 2000 Species boundaries in the starfish genus *Linckia*. *Mar. Biol.* **136**, 137–148. (doi:10.1007/s002270050016)
- Lessios HA, Kessing BD, Pearse JS. 2001 Population structure and speciation in tropical seas: global phylogeography of the sea urchin *Diadema*. *Evolution* **55**, 955–975. (doi:10.1554/0014-3820(2001)055[0955:PSASIT]2.0.CO;2)
- Bernardi G, Glacomo B. 2005 Phylogeography and demography of sympatric sister surfperch species, *Embiotoca jacksoni* and *E. lateralis* along the California coast: historical versus ecological factors. *Evolution* **59**, 386–394. (doi:10.1554/04-367)
- Kochzius M, Nuryanto A. 2008 Strong genetic population structure in the boring giant clam, *Tridacna crocea*, across the Indo-Malay Archipelago: implications related to evolutionary processes and connectivity. *Mol. Ecol.* **17**, 3775–3787. (doi:10.1111/j.1365-294X.2008.03803.x)
- Bowen BW, Bass AL, Rocha LA, Grant WS, Robertson DR. 2001 Phylogeography of the trumpetfishes (*Aulostomus*): ring species complex on a global scale. *Evolution* **55**, 1029–1039. (doi:10.1554/0014-3820(2001)055[1029:POTTAR]2.0.CO;2)
- McCafferty S, Bermingham E, Quenouille B, Planes S, Hoelzer G, Asoh K. 2002 Historical biogeography and molecular systematics of the Indo-Pacific genus *Dascyllus* (Teleostei: Pomacentridae). *Mol. Ecol.* **11**, 1377–1392. (doi:10.1046/j.1365-294X.2002.01533.x)
- Collin R. 2003 Phylogenetic relationships among calyptraeid gastropods and their implications for the biogeography of marine speciation. *Syst. Biol.* **52**, 618–640. (doi:10.1080/10635150390235430)
- Krug PJ. 2011 Patterns of speciation in marine gastropods: a review of the phylogenetic evidence for localized radiations in the sea. *Am. Malacol. Bull.* **29**, 169–186. (doi:10.4003/006.029.0210)
- Ingram T. 2011 Speciation along a depth gradient in a marine adaptive radiation. *Proc. R. Soc. B* **287**, 20101127. (doi:10.1098/rspb.2010.1127)
- Schluter D. 2000 *The ecology of adaptive radiation*. Oxford: Oxford Series in Ecology and Evolution, 300. (doi:10.2307/3558417)
- Grossenbacher DL, Whittall JB. 2011 Increased floral divergence in sympatric monkeyflowers. *Evolution* **65**, 2712–2718. (doi:10.1111/j.1558-5646.2011.01306.x)
- Brooks DR. 1994 Animal species and their evolution. *Trends Ecol. Evol.* **9**, 153–154. (doi:10.1016/0169-5347(94)90187-2)
- Nagel L, Schluter D. 1998 Body size, natural selection, and speciation in sticklebacks. *Evolution* **52**, 209–218. (doi:10.2307/2410936)
- Dieckmann U, Doebeli M. 1999 On the origin of species by sympatric speciation. *Nature* **400**, 354–357. (doi:10.1038/22521)
- Pfennig DW, Pfennig KS. 2012 *Evolution's wedge: competition and the origins of diversity*. Berkeley, CA: University of California Press.
- Rabosky DL, Santini F, Eastman J, Smith SA, Sidlauskas B, Chang J, Alfaro ME. 2013 Rates of speciation and morphological evolution are correlated across the largest vertebrate radiation. *Nat. Commun.* **4**, 1–38. (doi:10.1038/ncomms2958)
- Ricklefs R. 2006 Time, species, and the generation of trait variance in clades. *Syst. Biol.* **55**, 151–159. (doi:10.1080/10635150500431205)
- Tavera J, Acero PA, Wainwright PC. 2018 Multilocus phylogeny, divergence times, and a major role for the benthic-to-pelagic axis in the diversification of

- grunts (Haemulidae). *Mol. Phylogenet. Evol.* **121**, 212–223. (doi:10.1016/j.ympev.2017.12.032)
35. Tavera JJ, Acero PA, Balart EF, Bernardi G, Acero AP, Balart EF, Bernardi G. 2012 Molecular phylogeny of grunts (Teleostei, Haemulidae), with an emphasis on the ecology, evolution, and speciation history of new world species. *BMC Evol. Biol.* **12**, 57. (doi:10.1186/1471-2148-12-57)
36. Sanciangco MD, Rocha LA, Carpenter KE. 2011 A molecular phylogeny of the Grunts (Perciformes: Haemulidae) inferred using mitochondrial and nuclear genes. *Zootaxa* **50**, 37–50.
37. Chesser RT, Zink RM. 1994 Modes of speciation in birds: a test of Lynch's method. *Source Evol.* **48**, 490–497. (doi:10.2307/2410107)
38. Barraclough T, Vogler A. 2000 Detecting the geographical pattern of speciation from species-level phylogenies. *Am. Nat.* **155**, 419–434. (doi:10.1086/303332)
39. Barraclough TG, Vogler AP, Harvey PH. 1998 Revealing the factors that promote speciation. *Phil. Trans. R. Soc. Lond. B* **353**, 241–249. (doi:10.1098/rstb.1998.0206)
40. Fitzpatrick BM, Turelli M. 2006 The geography of mammalian speciation: mixed signals from phylogenies and range maps. *Evolution* **60**, 601. (doi:10.1554/05-453.1)
41. Böhlke JE, Chaplin CCG. 1968 *Fishes of the Bahamas and adjacent tropical waters*. Wynnewood, PA: Livingston Publishing Co.
42. Randall J. 1968 *Caribbean reef fishes*, 1st edn. New York, NY: T.F.H. Publications, Inc.
43. Uyeno T, Matsuura K, Fujii E. 1983 *Fishes trawled off Suriname and French Guiana*. Tokyo, Japan: Marine Fishery Resource Research Center.
44. Cervigón F. 1993 *Los peces marinos de Venezuela*. Caracas, Venezuela: Fundación Científica Los Roques.
45. Allen GR, Robertson DR. 1994 *Fishes of the tropical eastern Pacific*. Honolulu, HI: University of Hawaii Press.
46. McKay RJ, Schneider M. 1995 Haemulidae. In *Guía FAO para la identificación de especies para los fines de la pesca: pacífico centro-oriental Vol. 3 vertebrados—parte 2* (eds W Fischer, F Krupp, W Schneider, C Sommer, KE Carpenter, VH Niem), pp. 1136–1173. Rome, Italy: FAO.
47. Chirichigno NF, Cornejo MU. 2001 *Catálogo comentado de los peces marinos del Perú*. Callao, Perú: Instituto del Mar del Perú.
48. Humann P, DeLoach N. 2002 *Reef fish identification: Florida Caribbean Bahamas*, 3rd edn. Jacksonville, FL: New World Publications Inc.
49. Lindeman KC, Toxey CS. 2002 Haemulidae. In *FAO species identification guide for fishery purposes. The living marine resources of the Western Central Atlantic* (ed KE Carpenter), pp. 1522–1550. Rome, Italy: Food and Agriculture Organization of the United Nations.
50. McEachran JD, Feckhelm JD. 2005 *Fishes of the Gulf of Mexico: scorpaeniformes to tetraodontiformes*. Austin, TX: University of Texas Press.
51. Rohlf FJ. 2002 Geometric morphometrics and phylogeny. *Syst. Assoc. Spec.* **64**, 175–193. (doi:10.1016/j.tree.2004.08.005)
52. Klingenberg CP, Gidaszewski NA. 2010 Testing and quantifying phylogenetic signals and homoplasy in morphometric data. *Syst. Biol.* **59**, 245–261. (doi:10.1093/sysbio/syp106)
53. Revell LJ. 2012 phytools: an R package for phylogenetic comparative biology (and other things). *Methods Ecol. Evol.* **3**, 217–223. (doi:10.1111/j.2041-210X.2011.00169.x)
54. Price SA, Tavera JJ, Near TJ, Wainwright PC. 2012 Elevated rates of morphological and functional diversification in reef-dwelling haemulid fishes. *Evolution* **67**, 1–12. (doi:10.5061/dryad.s049 s)
55. Revell LJ. 2009 Size-correction and principal components for interspecific comparative studies. *Evolution* **63**, 3258–3268. (doi:10.1111/j.1558-5646.2009.00804.x)
56. Paradis E, Claude J, Strimmer K. 2004 APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)
57. Harmon LJ, Weir JT, Brock CD, Glor RE, Challenger W. 2008 GEGE: investigating evolutionary radiations. *Bioinformatics* **24**, 129–131. (doi:10.1093/bioinformatics/btm538)
58. Freckleton RP, Harvey PH. 2006 Detecting non-Brownian trait evolution in adaptive radiations. *PLoS Biol.* **4**, e373. (doi:10.1371/journal.pbio.0040373)
59. Mahler DL, Revell LJ, Glor RE, Losos JB. 2010 Ecological opportunity and the rate of morphological evolution in the diversification of Greater Antillean anoles. *Evolution* **64**, 2731–2745. (doi:10.1111/j.1558-5646.2010.01026.x)
60. Felsenstein J. 1985 Phylogenies and the comparative method. *Am. Nat.* **125**, 1–15. (doi:10.1086/284325)
61. McEntee JP, Tobias JA, Sheard C, Burleigh JG. 2018 Tempo and timing of ecological trait divergence in bird speciation. *Nat. Ecol. Evol.* **2**, 1120–1127. (doi:10.1038/s41559-018-0570-y)
62. Knowlton N. 1993 Sibling species in the Sea. *Annu. Rev. Ecol. Syst.* **24**, 189–216. (doi:10.1146/annurev.es.24.110193.001201)
63. Hodge JR, Bellwood DR. 2016 The geography of speciation in coral reef fishes: the relative importance of biogeographical barriers in separating sister-species. *J. Biogeogr.* **43**, 1324–1335. (doi:10.1111/jbi.12729)
64. Anacker BL, Strauss SY. 2014 The geography and ecology of plant speciation: range overlap and niche divergence in sister species. *Proc. R. Soc. B* **281**, 20132980. (doi:10.1098/rspb.2013.2980)
65. Williams ST, Reid DG. 2004 Speciation and diversity on tropical rocky shores: a global phylogeny of snails of the genus *Echinolittorina*. *Evolution* **58**, 2227–2251. (doi:10.1111/j.0014-3820.2004.tb01600.x)
66. Lessios H. 1998 The first stage of speciation as seen in organism separated by the Isthmus of Panama. In *Endless forms: species and speciation* (eds DJ Howard, SH Berlocher), pp. 185–201. New York, NY: Oxford University Press.
67. Bernardi G, Findley L, Rocha-Olivares A. 2003 Vicariance and dispersal across Baja California in disjunct marine fish populations. *Evolution* **57**, 1599–1609. (doi:10.1554/02-669)
68. Rocha LA. 2003 Patterns of distribution and process of speciation in Brazilian reef fishes. *J. Biogeogr.* **30**, 1161–1171. (doi:10.1046/j.1365-2699.2003.00900.x)
69. Lessios HA. 2008 The Great American Schism: divergence of marine organisms after the rise of the Central American Isthmus. *Annu. Rev. Ecol. Syst.* **39**, 63–91. (doi:10.1146/annurev.ecolsys.38.091206.095815)
70. Bernardi G, Alva-Campbell YR, Gasparini JL, Floeter SR. 2008 Molecular ecology, speciation, and evolution of the reef fish genus *Anisotremus*. *Mol. Phylogenet. Evol.* **48**, 929–935. (doi:10.1016/j.ympev.2008.05.011)
71. Renema W *et al.* 2008 Hopping hotspots: global shifts in marine biodiversity. *Science* **321**, 654–657. (doi:10.1126/science.1155674)
72. Cowman PF, Bellwood DR. 2013 The historical biogeography of coral reef fishes: global patterns of origination and dispersal. *J. Biogeogr.* **40**, 209–224. (doi:10.1111/jbi.12003)
73. Cowman PF, Bellwood DR. 2013 Vicariance across major marine biogeographic barriers: temporal concordance and the relative intensity of hard versus soft barriers. *Proc. R. Soc. B* **280**, 20131541. (doi:10.1098/rspb.2013.1541)
74. Rocha LA, Bowen BW. 2008 Speciation in coral-reef fishes. *J. Fish Biol.* **72**, 1101–1121. (doi:10.1111/j.1095-8649.2007.01770.x)
75. Hodge JR, Read CI, van Herwerden L, Bellwood DR. 2012 The role of peripheral endemism in species diversification: evidence from the coral reef fish genus *Anampses* (Family: Labridae). *Mol. Phylogenet. Evol.* **62**, 653–663. (doi:10.1016/j.ympev.2011.11.007)
76. Hodge JR, Read CI, Bellwood DR, van Herwerden L. 2013 Evolution of sympatric species: a case study of the coral reef fish genus *Pomacanthus* (Pomacanthidae). *J. Biogeogr.* **40**, 1676–1687. (doi:10.1111/jbi.12124)
77. Nelson G, Platnick N. 1981 *Systematics and biogeography: cladistics and vicariance*. New York, NY: Columbia University Press.
78. Crow KD, Munehara H, Bernardi G. 2010 Sympatric speciation in a genus of marine reef fishes. *Mol. Ecol.* **19**, 2089–2105. (doi:10.1111/j.1365-294X.2010.04611.x)
79. Malay MCM, Paulay G. 2010 Peripatric speciation drives diversification and distributional pattern of reef hermit crabs (Decapoda: Diogenidae: *Calcinus*). *Evolution* **64**, 634–662. (doi:10.1111/j.1558-5646.2009.00848.x)
80. Quenouille B, Hubert N, Bermingham E, Planes S. 2011 Speciation in tropical seas: allopatry followed by range change. *Mol. Phylogenet. Evol.* **58**, 546–552. (doi:10.1016/j.ympev.2010.12.009)

81. Bernal MA, Gaither MR, Simison WB, Rocha LA. 2017 Introgression and selection shaped the evolutionary history of sympatric sister-species of coral reef fishes (genus: *Haemulon*). *Mol. Ecol.* **26**, 639–652. (doi:10.1111/mec.13937)
82. Schluter D, Rambaut A. 1996 Ecological speciation in postglacial fishes [and Discussion]. *Phil. Trans. R. Soc. B* **351**, 807–814. (doi:10.1098/rstb.1996.0075)
83. Burns JH, Strauss SY. 2011 More closely related species are more ecologically similar in an experimental test. *Proc. Natl Acad. Sci. USA* **108**, 5302–5307. (doi:10.1073/pnas.1116085108)
84. Jablonski D, Roy K. 2003 Geographical range and speciation in fossil and living molluscs. *Proc. R. Soc. B* **270**, 401–406. (doi:10.1098/rspb.2002.2243)
85. Pitteloud C *et al.* 2017 Climatic niche evolution is faster in sympatric than allopatric lineages of the butterfly genus *Pyrgus*. *Proc. R. Soc. B* **284**, 20170208. (doi:10.1098/rspb.2017.0208)
86. Tavera JJ, Wainwright PC. 2019 Data from: Geography of speciation affects rate of trait divergence in haemulid fishes. Dryad Digital Repository. (doi:10.5061/dryad.5qd0gd0)