UC Irvine

UC Irvine Previously Published Works

Title

Draft Genome Sequence of Salmonella enterica subsp. enterica Serovar Orion Strain CRJJGF_00093 (Phylum Gammaproteobacteria)

Permalink

https://escholarship.org/uc/item/08w0d7pk

Journal

Microbiology Resource Announcements, 4(5)

ISSN

2576-098X

Authors

Gupta, Sushim K McMillan, Elizabeth A Jackson, Charlene R et al.

Publication Date

2016-10-27

DOI

10.1128/genomea.01063-16

Peer reviewed







Draft Genome Sequence of Salmonella enterica subsp. enterica Serovar Orion Strain CRJJGF_00093 (Phylum Gammaproteobacteria)

Sushim K. Gupta,^a Elizabeth A. McMillan,^{a,b} Charlene R. Jackson,^a Prerak T. Desai,^c Steffen Porwollik,^c Michael McClelland,^c Lari M. Hiott,^a Shaheen B. Humayoun,^a Jonathan G. Frye^a

Bacterial Epidemiology and Antimicrobial Resistance Research Unit, USDA-ARS, Athens, Georgia, USA^a; Department of Microbiology, University of Georgia, Athens, Georgia, USA^b; Department of Microbiology and Molecular Genetics, University of California Irvine, Irvine, California, USA^c

Here, we report a 4.70-Mbp draft genome sequence of *Salmonella enterica* subsp. *enterica* serovar Orion strain CRJJGF_00093, isolated from a dog in 2005.

Received 5 August 2016 Accepted 8 August 2016 Published 29 September 2016

Citation Gupta SK, McMillan EA, Jackson CR, Desai PT, Porwollik S, McClelland M, Hiott LM, Humayoun SB, Frye JG. 2016. Draft genome sequence of Salmonella enterica subsp. enterica serovar Orion strain CRJJGF_00093 (phylum Gammaproteobacteria). Genome Announc 4(5):e01063-16. doi:10.1128/genomeA.01063-16.

Copyright © 2016 Gupta et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license. Address correspondence to Jonathan G. Frye, Jonathan.Frye@ars.usda.gov.

Salmonella enterica subsp. enterica serovar Orion belongs to lowinvasive groups of Salmonella pathogens (1), though S. Orion has been isolated from a variety of hosts, including humans (2, 3), cattle (4), ducks (5), and birds (6), across the globe. Here, we announce the draft genome sequence of S. Orion, isolated from a dog in 2005.

The Salmonella strain was isolated from a sick dog by NVSL (National Veterinary Services Laboratory) using standard microbiology techniques. The isolates were serotyped using Salmonella multiplex assay for rapid typing PCR (7), and a serotype antigenic formula was predicted using reads with SeqSero (8), which predicted the antigenic formula of 3,10:y:1,5, designated Orion. Using pulsed-field gel electrophoresis (PFGE), as described by PulseNet, the isolate displayed the PFGE pattern XLAX01.0009.ARS (9). Susceptibility testing for the strain was performed using broth microdilution plates for the Sensititre semiautomated antimicrobial susceptibility system (TREK Diagnostic Systems, Inc, Westlake, OH, USA), and guidelines of the Clinical and Laboratory Standards Institute (CLSI) were followed to interpret the results (10).

Genomic DNA was isolated from an overnight culture using the GenElute bacterial genomic DNA kit (Sigma-Aldrich, St. Louis, MO, USA). DNA libraries were constructed using the Nextera-XT DNA preparation kit, and paired-end sequencing was performed on the Illumina HiSeq 2500 platform (Illumina Inc., San Diego, CA, USA) using a 500-cycle MiSeq reagent kit. A total of 4,183,624 reads were generated. Reads were de novo assembled using Velvet (11), which assembled to 178 contigs ≥200 bp with an 83-fold average coverage. The combined length of the contigs was 4,701,421 bases with a G+C content of 52.19% and an N_{50} value of 58.9 kb. The contigs were ordered with Mauve (12) using the Salmonella LT2 genome sequences as reference; coding sequences and tRNAs were predicted with Prodigal (13) and ARAGORN (14), respectively. A total of 4,445 coding sequences (≥50 amino acids) and 43 tRNAs were predicted within the genome. Signal-peptide, CRISPR regions and prophages were predicted using Signalp (15), CRISPRFinder (16), and PHAST (17), respectively. We identified signal peptides in 414 coding sequences, two CRISPR loci, and three intact phages (Salmon_SP_004 [NC021774], Stx2_converting_1717 [NC011357], and Mannhe_

YB_MnM_3927AP2 [NC028766]) in the analyzed contigs. The strain was susceptible to all the tested antibiotics, though we detected a cryptic aminoglycoside resistance gene, *aac6*-Iy, with ARG-ANNOT (18). The genome data generated from *S*. Orion can be helpful to understand the variations in genomic features that make *S*. Orion a member of a low-invasive group of *Salmonella* pathogens.

Accession number(s). The genome sequence of Salmonella enterica subsp. enterica serovar Orion strain CRJJGF_00093 has been deposited in the GenBank database (NCBI) under the accession number JQVX00000000. This paper describes the first version of the genome, JQVX01000000.

ACKNOWLEDGMENTS

J.G.F. and C.R.J. were supported by USDA Project Numbers 6040-32000-006-00 and 6040-32000-009-00, and a grant from the Foundation for Meat and Poultry Research and Education. M.M. was supported in part by NIH grants R01AI052237, AI039557, AI052237, AI073971, AI075093, AI077645, and AI083646; USDA grants 2009-03579 and 2011-67017-30127; the Binational Agricultural Research and Development Fund; and a grant from the Center for Produce Safety. We also thank Calvin Williams for all the technical support.

FUNDING INFORMATION

This work, including the efforts of Charlene R. Jackson and Jonathan Gray Frye, was funded by Foundation for Meat and Poultry Research and Education. This work, including the efforts of Michael McClelland, was funded by HHS | National Institutes of Health (NIH) (R01AI052237, AI039557, AI052237, AI073971, AI075093, AI077645, and AI083646). This work, including the efforts of Charlene R. Jackson and Jonathan Gray Frye, was funded by U.S. Department of Agriculture (USDA) (6040-32000-006-00 and 6040-32000-009-00). This work, including the efforts of Michael McClelland, was funded by U.S. Department of Agriculture (USDA) (2009-03579 and 2011-67017-30127). This work, including the efforts of Michael McClelland, was funded by United States - Israel Binational Agricultural Research and Development Fund (BARD). This work, including the efforts of Michael McClelland, was funded by Center for Produce Safety (CPS).

The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

REFERENCES

- McWhorter AR, Davos D, Chousalkar KK. 2015. Pathogenicity of Salmonella strains isolated from egg shells and the layer farm environment in Australia. Appl Environ Microbiol 81:405–414. http://dx.doi.org/10.1128/AEM.02931-14.
- Cabrera R, Ruiz J, Ramírez M, Bravo L, Fernández A, Aladueña A, Echeíta A, Gascón J, Alonso PL, Vila J. 2006. Dissemination of Salmonella enterica serotype Agona and multidrug-resistant Salmonella enterica serotype Typhimurium in Cuba. Am J Trop Med Hyg 74:1049–1053.
- 3. Trafny EA, Kozłowska K, Szpakowska M. 2006. A novel multiplex PCR assay for the detection of *Salmonella enterica* serovar Enteritidis in human faeces. Lett Appl Microbiol 43:673–679. http://dx.doi.org/10.1111/j.1472 -765X.2006.02007.x.
- Alam MJ, Renter DG, Ives SE, Thomson DU, Sanderson MW, Hollis LC, Nagaraja TG. 2009. Potential associations between fecal shedding of Salmonella in feedlot cattle treated for apparent respiratory disease and subsequent adverse health outcomes. Vet Res 40:2. http://dx.doi.org/ 10.1051/vetres:2008040.
- Rampersad J, Johnson J, Brown G, Samlal M, Ammons D. 2008. Comparison of polymerase chain reaction and bacterial culture for *Salmonella* detection in the Muscovy duck in Trinidad and Tobago. Rev Panam Salud Publica 23:264–267. http://dx.doi.org/10.1590/S1020-49892008000400006.
- Münch S, Braun P, Wernery U, Kinne J, Pees M, Flieger A, Tietze E, Rabsch W. 2012. Prevalence, serovars, phage types, and antibiotic susceptibilities of *Salmonella* strains isolated from animals in the United Arab Emirates from 1996 to 2009. Trop Anim Health Prod 44:1725–1738. http://dx.doi.org/10.1007/s11250-012-0130-4.
- Leader BT, Frye JG, Hu J, Fedorka-Cray PJ, Boyle DS. 2009. Highthroughput molecular determination of *Salmonella enterica* serovars by use of multiplex PCR and capillary electrophoresis analysis. J Clin Microbiol 47:1290–1299. http://dx.doi.org/10.1128/JCM.02095-08.
- 8. Zhang S, Yin Y, Jones MB, Zhang Z, Deatherage Kaiser BL, Dinsmore BA, Fitzgerald C, Fields PI, Deng X. 2015. *Salmonella* serotype determi-

- nation utilizing high-throughput genome sequencing data. J Clin Microbiol 53:1685–1692. http://dx.doi.org/10.1128/JCM.00323-15.
- 9. Gerner-Smidt P, Hise K, Kincaid J, Hunter S, Rolando S, Hyytiä-Trees E, Ribot EM, Swaminathan B, Pulsenet Taskforce. 2006. PulseNet USA: a five-year update. Foodborne Pathog Dis 3:9–19. http://dx.doi.org/10.1128/JCM.00323-15.
- CLSI. 2015. Performance standards for antimicrobial susceptibility testing: 25th informational supplement (m100-S25). Clinical and Laboratory Standards Institute, Wayne, PA.
- 11. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res 18:821–829. http://dx.doi.org/10.1101/gr.074492.107.
- Rissman AI, Mau B, Biehl BS, Darling AE, Glasner JD, Perna NT. 2009. Reordering contigs of draft genomes using the mauve aligner. Bioinformatics 25:2071–2073. http://dx.doi.org/10.1093/bioinformatics/btp356.
- 13. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. http://dx.doi.org/10.1186/1471-2105-11-119.
- 14. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. http://dx.doi.org/10.1093/nar/gkh152.
- Petersen TN, Brunak S, von Heijne G, Nielsen H. 2011. SignalP 4.0: discriminating signal peptides from transmembrane regions. Nat Methods 8:785–786. http://dx.doi.org/10.1038/nmeth.1701.
- 16. **Grissa I, Vergnaud G, Pourcel C**. 2007. CRISPRFinder: a web tool to identify clustered regularly interspaced short palindromic repeats. Nucleic Acids Res 35:W52–W57. http://dx.doi.org/10.1093/nar/gkm360.
- 17. Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. Nucleic Acids Res 39:W347–W352. http://dx.doi.org/10.1093/nar/gkr485.
- Gupta SK, Padmanabhan BR, Diene SM, Lopez-Rojas R, Kempf M, Landraud L, Rolain JM. 2014. Arg-Annot, a new bioinformatic tool to discover antibiotic resistance genes in bacterial genomes. Antimicrob Agents Chemother 58:212–220. http://dx.doi.org/10.1128/ AAC.01310-13.