UC Davis

UC Davis Previously Published Works

Title

Complete Genome Sequence of a Putative Densovirus of the Asian Citrus Psyllid, Diaphorina citri

Permalink

https://escholarship.org/uc/item/0bz2k4z0

Journal

Microbiology Resource Announcements, 4(4)

ISSN

2576-098X

Authors

Nigg, Jared C Nouri, Shahideh Falk, Bryce W

Publication Date

2016-08-25

DOI

10.1128/genomea.00589-16

Peer reviewed







Complete Genome Sequence of a Putative Densovirus of the Asian Citrus Psyllid, *Diaphorina citri*

Jared C. Nigg, Shahideh Nouri, Bryce W. Falk

Department of Plant Pathology, University of California Davis, Davis, California, USA

Here, we report the complete genome sequence of a putative densovirus of the Asian citrus psyllid, *Diaphorina citri*. *Diaphorina citri* densovirus (DcDNV) was originally identified through metagenomics, and here, we obtained the complete nucleotide sequence using PCR-based approaches. Phylogenetic analysis places DcDNV between viruses of the *Ambidensovirus* and *Iteradensovirus* genera.

Received 5 May 2016 Accepted 10 June 2016 Published 28 July 2016

Citation Nigg JC, Nouri S, Falk BW. 2016. Complete genome sequence of a putative densovirus of the Asian citrus psyllid, *Diaphorina citri*. Genome Announc 4(4):e00589-16. doi:10.1128/genomeA.00589-16.

Copyright © 2016 Nigg et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Bryce W. Falk, bwfalk@ucdavis.edu.

metagenomic survey of viruses associated with *Diaphorina citri* revealed contigs displaying similarity to densovirus (DNV) structural (VP) and nonstructural (NS) genomic regions (1). These contigs were reported as genomic fragments of the tentatively named *Diaphorina citri* densovirus (DcDNV). To expand on the genomic sequence of DcDNV, DNA from *D. citri* collected in Taiwan was used for PCR, with primers designed to amplify the unknown sequence between the NS and VP coding regions, followed by Sanger sequencing. Sequence analysis revealed that DcDNV has an ambisense genome organization, with 25 nucleotides (nt) separating the VP and NS cassettes. Additional PCR-based strategies and Sanger sequencing were used to obtain the sequences at the extremities of the genome, which contain 210-nt inverted terminal repeats (ITRs) predicted to form simple 210-nt hairpins characteristic of subgroup B ambisense DNVs (2–4).

The complete genome sequence of DcDNV is 5,071 nt and contains four predicted open reading frames (ORFs). ORF1 (nt 300 to 1598) has a coding capacity of 432 amino acids (aa). BLASTp analysis of the full-length putative protein encoded by ORF1 indicates similarity with uncharacterized insect proteins (accession numbers XP_011214328.1 and XP_003248352.1). ORF2 (nt 338 to 2377) begins with a TTG codon at position 338 and is present in a -1 reading frame relative to ORF1. BLASTp analysis of the putative 679-aa protein encoded by ORF2 indicated the highest identity with Cherax quadricarinatus densovirus NS1 (query coverage, 67%; identity, 34%) (accession no. YP_ 009134732.1). Additionally, the ORF2-encoded protein possesses the rolling-circle replication initiator and helicase superfamily 3 motifs characteristic of DNV NS1 proteins (5-7). We did not identify DNV NS2 or NS3 ORFs. Among ambisense DNVs, lack of an NS3 ORF has been reported only for *Myzus persicae* densovirus (8). Although the phylogenetic position of DcDNV is unclear, phylogenetic analysis based on the NS1 amino acid sequence places DcDNV in an intermediate position between the subgroup B ambisense DNVs and the iteradensoviruses. Indeed, the organization of the NS cassette of DcDNV resembles that of iteradensoviruses more than other ambisense DNVs.

The VP cassettes of ambisense DNVs are on the complementary strand of that containing the NS cassettes and encode four or five structural proteins from one or two ORFs, respectively (4). ORF3 (nt 2402 to 4168) encodes a putative 588-aa protein that displays the highest identity to the VP1 protein of densovirus SC1065 based on BLASTp analysis (query coverage, 37%; identity, 31%). ORF4 (nt 4149 to 4766) has a coding capacity of 205 aa and encodes a putative protein containing the HDXXY and YXGXG phospholipase A2 motifs characteristic of DNV VP1 proteins (9). The full-length putative protein encoded by ORF4 shows the highest similarity with Periplaneta fuliginosa densovirus VP1 based on BLASTp analysis (query coverage, 43%; identity, 42%) (accession no. BAA82965.1). The structural proteins of ambisense DNVs are generated by leaky scanning and/or alternative splicing mechanisms. Splice site prediction using NNSPLICE (version 0.9) (10) indicates seven potential splicing donor sites and four potential splicing acceptor sites within the VP cassette.

Nucleotide sequence accession number. The GenBank accession number of the complete nucleotide sequence of DcDNV is KX165268.

ACKNOWLEDGMENTS

We thank Hsin-Hung Yeh for providing *D. citri* samples. We also thank Martha Wohfeil for lab assistance.

This material is based upon work supported by the National Science Foundation Graduate Research Program under Grant No. 1148897.

FUNDING INFORMATION

This work, including the efforts of Jared C. Nigg, was funded by National Science Foundation (NSF) (1148897). This work, including the efforts of Bryce W. Falk, was funded by U.S. Department of Agriculture (USDA) (13-002NU-781 and 2015-70016-23011).

REFERENCES

1. Nouri S, Salem N, Nigg JC, Falk BW. 2016. A diverse array of new viral sequences identified in worldwide populations of the Asian citrus psyllid (*Diaphorina citri*) using viral metagenomics. J Virol 90:2434–2445. http://dx.doi.org/10.1128/JVI.02793-15.

- Guo H, Zhang J, Hu Y. 2000. Complete sequence and organization of Periplaneta fuliginosa densovirus genome. Acta Virol 44:315–322.
- 3. Bochow S, Condon K, Elliman J, Owens L. 2015. First complete genome of an *Ambidensovirus*; *Cherax quadricarinatus densovirus*, from freshwater crayfish *Cherax quadricarinatus*. Mar Genomics 24:305–312. http://dx.doi.org/10.1016/j.margen.2015.07.009.
- Tijssen P, Bando H, Li Y, Jousset F, Zadori Z, Fediere G, El-Far M, Szelei J, Bergoin M. 2005. Evolution of densoviruses. Parvoviruses 5:55–60.
- 5. Ilyina TV, Koonin EV. 1992. Conserved sequence motifs in the initiator proteins for rolling circle DNA replication encoded by diverse replicons from eubacteria, eucaryotes and archaebacteria. Nucleic Acids Res 20: 3279–3285. http://dx.doi.org/10.1093/nar/20.13.3279.
- Koonin EV. 1993. A common set of conserved motifs in a vast variety of putative nucleic acid-dependent ATPases including MCM proteins involved in the initiation of eukaryotic DNA replication. Nucleic Acids Res 21:2541–2547. http://dx.doi.org/10.1093/nar/21.11.2541.
- 7. Gorbalenya AE, Koonin EV, Wolf YI. 1990. A new superfamily of putative NTP-binding domains encoded by genomes of small DNA and RNA viruses. FEBS Lett 262:145–148. http://dx.doi.org/10.1016/0014-5793(90)80175-I.
- Van Munster M, Dullemans AM, Verbeek M, van den Heuvel JF, Reinbold C, Brault V, Clérivet A, Van der Wilk F. 2003. A new virus infecting *Myzus persicae* has a genome organization similar to the species of the genus *Densovirus*. J Gen Virol 84:165–172. http://dx.doi.org/ 10.1099/vir.0.18650-0.
- Zádori Z, Szelei J, Lacoste M-C, Li Y, Gariépy S, Raymond P, Allaire M, Nabi IR, Tijssen P. 2001. A viral phospholipase A 2 is required for parvovirus infectivity. Dev Cell 1:291–302. http://dx.doi.org/10.1016/S1534 -5807(01)00031-4.
- Reese MG, Eeckman FH, Kulp D, Haussler D. 1997. Improved splice site detection in Genie. J Comput Biol 4:311–323. http://dx.doi.org/10.1089/ cmb.1997.4.311.