

UC Irvine

UC Irvine Previously Published Works

Title

Extended imaging depth of en-face optical coherence tomography based on fast measurement of a reflection matrix by wide-field heterodyne detection.

Permalink

<https://escholarship.org/uc/item/0dx693ms>

Journal

Optics Letters, 45(4)

ISSN

0146-9592

Authors

Yang, Qiang
Cao, Jing
Miao, Yusi
[et al.](#)

Publication Date

2020-02-15

DOI

10.1364/ol.382898

Peer reviewed



HHS Public Access

Author manuscript

Opt Lett. Author manuscript; available in PMC 2021 February 15.

Published in final edited form as:

Opt Lett. 2020 February 15; 45(4): 828–831. doi:10.1364/OL.382898.

Extended imaging depth of *en-face* optical coherence tomography based on fast measurement of a reflection matrix by wide-field heterodyne detection

Qiang Yang¹, Jing Cao¹, Yusi Miao^{1,2}, Jiang Zhu¹, Zhongping Chen^{1,2,*}

¹Beckman Laser Institute, University of California, Irvine, Irvine, California 92612, USA

²Department of Biomedical Engineering, University of California, Irvine, Irvine, California 92697, USA

Abstract

Multiple light scattering in biomedical tissue limits the penetration depth of optical imaging systems such as optical coherence tomography. To increase the imaging depth in scattering media, a computational method based on coherent reflection matrix measurement has been developed using low coherence interferometry. The complex reflection matrix is obtained via point-by-point scanning followed by a phase-shifting method; then singular value decomposition is used to retrieve the singly back-scattered light. However, the in vivo application of the current reported method is limited due to the slow acquisition speed of the matrix. In this Letter, a wide-field heterodyne-detection method is adopted to speed up the complex matrix measurement at a deep tissue layer. Compared to the phase-shifting method, the heterodyne-detection scheme retrieves depth-resolved complex amplitudes faster and is more stable without mechanical movement of the reference mirror. As a result, the matrix measurement speed is increased by more than one order of magnitude.

Optical imaging inside a scattering media at a depth beyond what is currently accessible poses a great challenge to biomedical communities. Great efforts have been made in various modalities to increase the depth of imaging while maintaining the microscopic resolution [1]. Among all the modalities to wrestle with the light scattering, optical coherence tomography (OCT) is able to provide depth-resolved images with high spatial resolution by adopting low coherence interferometry with a broadband light source [2,3]. Nevertheless, the penetration depth of conventional OCT is limited to the range of 1–2 mm inside human skin tissue due to the increased ratio of multiple-scattered photons to the ballistic photons along the penetration depth. The multiple photons bounced from different layers will dominate and hinder the signal retrieval of ballistic signal light when they have the same optical path difference with the reference light.

Different methods have been proposed to highlight and distinguish the ballistic signal light from the background multiple scattered light from unrelated multi-layers. One solution can

*Corresponding author: z2chen@uci.edu.

Disclosures. Dr. Zhongping Chen has a financial interest in OCT Medical Imaging, Inc., which, however, did not support this Letter.

The target particle is placed in the focal plane of the incident beam underneath a scattering layer. A half-wave plate (HWP) and linear polarizer (LP) pair in the incident arm, reference arm, and sample arm work as adjustable attenuators. Two acoustic optical modulators (AOMs) [405AF3, IntraAction Corp.] are cascade connected to shift the optical frequency in the reference arm by 40 kHz. To minimize the return loss of back-scattered light from the sample, the combination of a rotatable QWP with PBS1 is adopted in the sample arm.

To test the performance of the wide-field heterodyne detection, microbead targets are placed ~1 mm underneath a strong scattering sample layer, as in Fig. 2. The spatial light modulator (SLM) works as a two-axis galvo scanner by successively displaying hundreds of phase-only grating patterns that have different periods and orientations to deflect the incident beam into different diffractive angles [Fig. 2(a)]. The sample beam is focused on the top surface of the cover glass for raster scanning after passing through the objective. Like a stone being thrown into the water to trigger a wide-field wavefront distribution nearby, an extended 2D optical field will be formed around the focal spot due to the multiple refractions and inner reflections of light inside the scatterer target, if the target is just positioned on the trajectory of the focused scan beam [Fig. 2(b)]. The fast Fourier transform (FFT) is applied to the recorded complex field to reconstruct the original speckle complex field in the target plane. By using an adjustable digital pinhole in the post-processing right after FFT, only the complex field in the peripheral area of the scanning point is chosen to be reconstructed [Fig. 2(c)]. The focused beam is then scanned point-by-point along the transverse direction in the target plane; thus, the whole reflection matrix of the scanned area can be obtained by using the lock-in complementary metal–oxide–semiconductor (CMOS) camera (300x300 pixels, 16 bits, Helicam C3, Heliotis, Switzerland).

The principle of wide-field heterodyne-detection of interference field in the camera sensor plane is illustrated in Fig. 3. The wave function of the light takes the general form of $Ae^{i(\omega t + \varphi)}$. The superposition of two light waves (the sample beam and the reference beam in the interferometer) with different frequencies and amplitudes at a fixed point (i.e., one pixel of the lockin camera) is shown below, using the complex exponential expressions:

$$\begin{aligned} E &= A_1 e^{i(\omega_1 t + \varphi_1)} + A_2 e^{i(\omega_2 t + \varphi_2)} \\ &= e^{i(\omega_2 t)} (A_1 e^{i[\Delta\omega t + \varphi_1]} + A_2 e^{i\varphi_2}), \end{aligned} \quad (1)$$

where $e^{i(\omega_2 t)}$ is the rapidly changing component which is undetectable by the detector, and $A_1 e^{i(\omega_1 t + \varphi_1)} + A_2 e^{i\varphi_2}$ is the slowly changing component, $\omega = \omega_1 - \omega_2$. The corresponding detectable intensity of the interference light field is

$$\begin{aligned} I &= |E|^2 = |A_1 e^{i[\Delta\omega t + \varphi_1]} + A_2 e^{i\varphi_2}|^2 = |A_1|^2 + |A_2|^2 \\ &\quad + 2A_1 A_2 \cos(\Delta\omega t + \Delta\varphi). \end{aligned} \quad (2)$$

If the DC item $|A_1|^2 + |A_2|^2$ is removed as background, the remaining signal is $2A_1 A_2 \cos(\omega t + \varphi) = I_1 \cos(\omega t + \varphi)$. Here, A_2 and φ_2 are the reference beam's amplitude and phase which are regarded as the constants, since the reference beam is approximately a plane wave. The lock-in camera outputs two local oscillations (LOs) of $\cos(\omega t)$ and $\sin(\omega t)$ in

two individual channels of the camera's driving circuit. By multiplying the two LOs with the detected signal $I_1 \cos(\omega t + \varphi)$, the corresponding output signals become $1/2 I_1 \cos(\varphi)$ and $1/2 I_1 \sin(\varphi)$, since the high-frequency components are removed by a low-pass filter inside the camera. These two signals are called in-phase (I) and quadrature (Q) signals. The phase of the sample beam can be calculated by $\varphi_I = \varphi = \arctan(Q/I)$, and the amplitude can be calculated by $A_1 \propto I_1 = \sqrt{I^2 + Q^2}$. Only a constant coefficient difference exists between A_1 and I_1 .

The I and Q signals are retrieved from the interference signal through the camera's output function, so as to calculate the phase and amplitude profile of the back-scattered sample beam. The two profiles acquired in Fig. 4 by the proposed heterodyne-detection method have undergone only one acquisition without a frame averaging process which is time-consuming and typically adopted by the four-step phase-shifting method to improve the signal-to-noise ratio (SNR).

One focal spot in the target plane corresponds to one frame of 2D complex field in the sensor plane. Each frame of the 2D complex amplitude is reshaped into a vector for the matrix rearrangement. The rearrangement order complies with the scanning trajectory of the incident sample beam. Thus, hundreds of complex amplitudes are recombined into one reflection matrix for post-analysis. The digital pinhole in the target plane makes the original reflection matrix become a filtered matrix with multiple diagonals, as in Fig. 5. This step of wide-field detection with an enlarged pinhole filter is a key factor for the matrix analysis.

After obtaining the filtered reflection matrix $RM(r_{in}, r_{out})$, a singular value decomposition (SVD) is applied to this matrix which can be expressed by the form of $RM(r_{in}, r_{out}) = U \Sigma V^\dagger$, as in Fig. 6.

SVD is an effective tool for finding lower-rank approximations to a given matrix U and V are the unitary matrices whose columns correspond to the input and output singular vectors U_i and V_i , respectively. The subscript i represents the column number for the matrix U or the row number for the matrix V^\dagger . Σ is the diagonal matrix containing the real and positive singular values σ_i in a decreased order of $\sigma_1 > \sigma_2 \dots > \sigma_N > \sigma_{N+1}$. \dagger is the conjugate transpose. The colors in the matrix above denote different complex values. The singular vectors U_i and V_i having the biggest singular values are chosen and reshaped from the original one-dimensional vectors into two 2D wavefronts \bar{U}_i and \bar{V}_i respectively. The reshaping order is matched with the scanning trajectory of the focused beam. The image can be reconstructed by $I(x, y) = |\bar{U}_1 \cdot \bar{V}_1| \cdot \sigma_1 + |\bar{U}_2 \cdot \bar{V}_2| \cdot \sigma_2 + \dots$, where σ_i weighting factor. Compared with the counterpart of ultrasonic imaging [8], the optical pulse and echo in our system are in the same focal plane, so there is no need to back-propagate the wavefront of \bar{U}_i and \bar{V}_i . Thus, the 2D target image is reconstructed according to the complex modulus of the Hadamard product between the two reshaped wavefronts.

In the experiment, the optical beating frequency is set at 40 kHz by tuning the two AOM drivers. To satisfy the Nyquist criterion, the matrix is measured by the lock-in camera operating at a demodulation rate of 80 kHz. The microbead images are successfully

reconstructed by applying SVD onto the filtered reflection matrix followed by the reconstruction algorithm, as in Fig. 7. To demonstrate the extended imaging depth of our system compared to conventional OCT, a standard *en-face* OCT imaging process by conventional OCT with the same targets and scattering layer was also conducted. However, the commercial OCT (OCS1310V1 1300 nm MEMS-VCSEL swept source OCT, 100 kHz, Thorlabs) in the lab has a lateral resolution larger than 13 μm , due to low numerical aperture (NA) scan lens (OCT-LSM03, NA 0.06, Thorlabs) used. Thus, the *en-face* image of the microbeads with a 10 μm diameter in Fig. 7 cannot be successfully acquired by the OCS1310V1 system. To make the image results comparable, microbeads with a larger diameter (20 μm) are chosen as targets in Fig. 8, while a ~ 0.8 mm thickness lab-made phantom tissue (TiO₂ powder dissolved in a curing agent and silicone with a scattering coefficient more than two times that of human skin) is used as the scattering layer. These microbeads made of polydimethylsiloxane (PDMS) are phase objects and evenly distributed via interference nanolithography on the same PDMS material base which is soft and flexible.

The images of microbeads are recovered successfully by the proposed method, whether or not the strong scattering tissue is placed above [Figs. 8(e) and 8(f)], while the commercial system fails to recover the image with the scattering tissue put above [Figs. 8(b) and 8(d)], even though its source has a longer center wavelength at 1310 nm. In previous systems using four-step phase shifting, the microbead image with the same tissue cannot be recovered either because of the decorrelation noises generated during the long-time measurement. The axial resolution is decided by the bandwidth of the light source, as in conventional OCT. Due to the large dispersion introduced by the AOMs and the objective, the measured axial resolution is ~ 20 μm , which is larger than the theoretical value and can be improved by further dispersion compensation in both arms. According to the Nyquist sampling criterion, the lateral resolution is two times that of the distance between each scanning point, that is, two times larger than conventional OCT. The lateral resolution in the current system is ~ 4 μm . One drawback of the heterodyne detection is that the pixel pitch of the lock-in camera is as large as 40 μm , which decreases the SNR of the recovered image.

In summary, deep tissue imaging at a depth beyond conventional OCT is demonstrated by combining the reflection matrix measurement with the wide-field heterodyne detection. The number of scattering mean free paths (MFPs) of the tissue is measured as 16.4 [7], approximately two times deeper than conventional OCT (which can image at a depth with ~ 6 -9 scattering MFPs according to the literature) [5,10]. Here the number of scattering MFPs is measured according to Refs. [5,7]. The photodetector (DCC1645C, Thorlabs) used to measure the MFP is placed ~ 1 m away from the scattering layer. By using a detector with higher sensitivity and lower noise level, the ratio of ballistic light to scattering light can be further decreased, and the calculated number of scattering MFPs regarding the tissue will increase. Importantly, the speed of matrix measurement is improved by one order above by using a wide-field heterodyne-detection method with the lock-in camera. The consuming time of matrix measurement in the proposed system is ~ 6 min for 961 scanning points, compared with ~ 20 -30 min for only 289 scanning points in previous works [5,7]. That is, the speed is ~ 0.37 sec/pt versus ~ 4.15 sec/pt, a significant improvement of more than one order of magnitude. The current matrix measurement speed is mainly limited by the liquid crystal SLM. Using the full speed of the lock-in camera (operated at 3800 fps) enables the entire

scattering matrix to be acquired in just seconds. This experiment paves the way for deep tissue imaging in the next phase by using living tissues.

Acknowledgment.

The authors thank Kaustubh Asawa and Prof. Chang-Hwan Choi at the Stevens Institute of Technology for providing the PDMS samples.

Funding. National Institutes of Health (R01EY-026091, R01EY-028662, R01HL-125084, R01HL-127271).

REFERENCES

1. Gigan S, Nat. Photonics 11, 14 (2017).
2. Fujimoto J and Swanson E, Invest. Ophthalmol. Visual Sci 57, 101 (2016).
3. Borycki D, Hamkalo M, Nowakowski M, Szkulmowski M, and Wojtkowski M, Biomed. Opt. Express 10, 2032 (2019). [PubMed: 31086716]
4. Liu G, Lin A, Tromberg B, and Chen Z, Biomed. Opt. Express 3, 2669 (2012). [PubMed: 23082305]
5. Badon A, Li D, Lerosey G, Boccara A, Fink M, and Aubry A, Sci. Adv 2, e1600370 (2016).
6. Kang S, Jeong S, Choi W, Ko H, Yang T, Joo J, Lee J, Lim Y, Park Q, and Choi W, Nat. Photonics 9, 253 (2015).
7. Yang Q, Miao Y, Huo T, Li Y, Heidari E, Zhu J, and Chen Z, Appl. Phys. Lett 113, 011106 (2018). [PubMed: 30034015]
8. Popoff S, Lerosey G, Carminati R, Fink M, Boccara A, and Gigan S, Phys. Rev. Lett 104, 100601 (2010). [PubMed: 20366410]
9. Liu Y, Ma C, Shen Y, and Wang LV, Opt. Lett 41, 1321 (2016). [PubMed: 27192226]
10. Pan Y, Birngruber R, and Engelhardt R, Appl. Opt 36, 2979 (1997). [PubMed: 18253302]

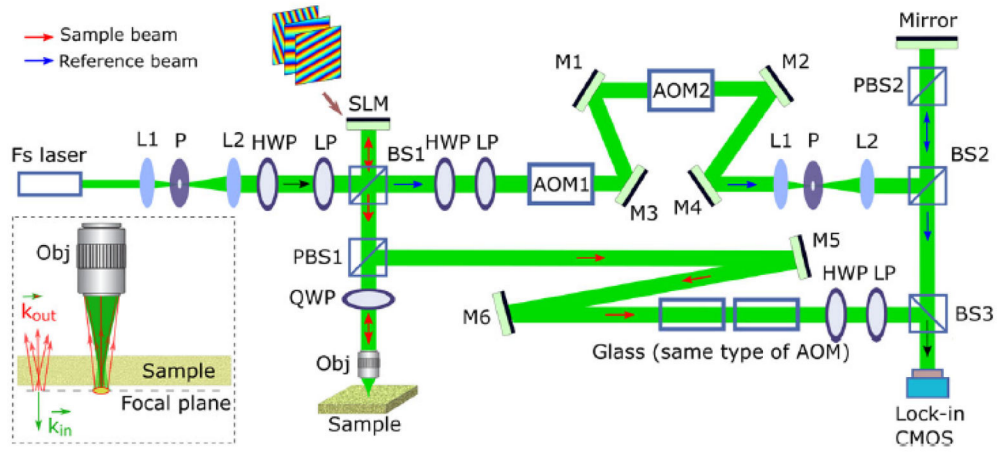


Fig. 1. Schematic of the imaging setup. Fs laser, femtosecond pulse laser; L, lens; BS, beam splitter; PBS, polarized beam splitter; HWP, half-wave plate; LP, linear polarizer; QWP, quarter-wave plate; Obj, objective (10XOlympus Plan Achromat Objective, 0.25 NA, 10.6mm WD); SLM, phase-only spatial light modulator (Pluto-NIR-015, Holoeye, Germany); AOM, acoustic optical modulator.

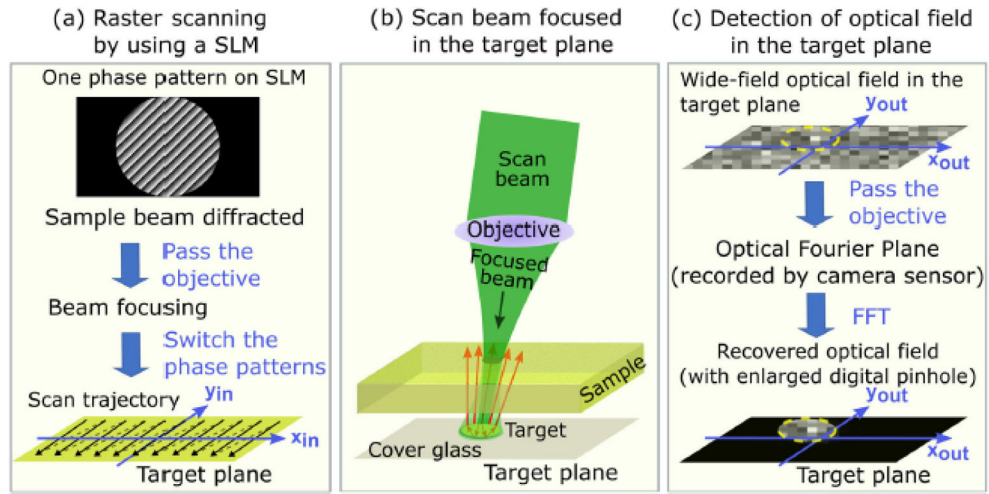


Fig. 2. Illustration of (a) the raster scanning process of the focused beam, which is formed by phase-only grating patterns successively displayed on the SLM. Here, one grating pattern corresponds to one single focal spot in the target plane; (b) the beam is focused on the target, resulting in a wide-field distorted optical distribution around; (c) detection of the wide-field distribution in the target plane by applying FFT to the Fourier plane distribution followed by digital pinhole filtering.

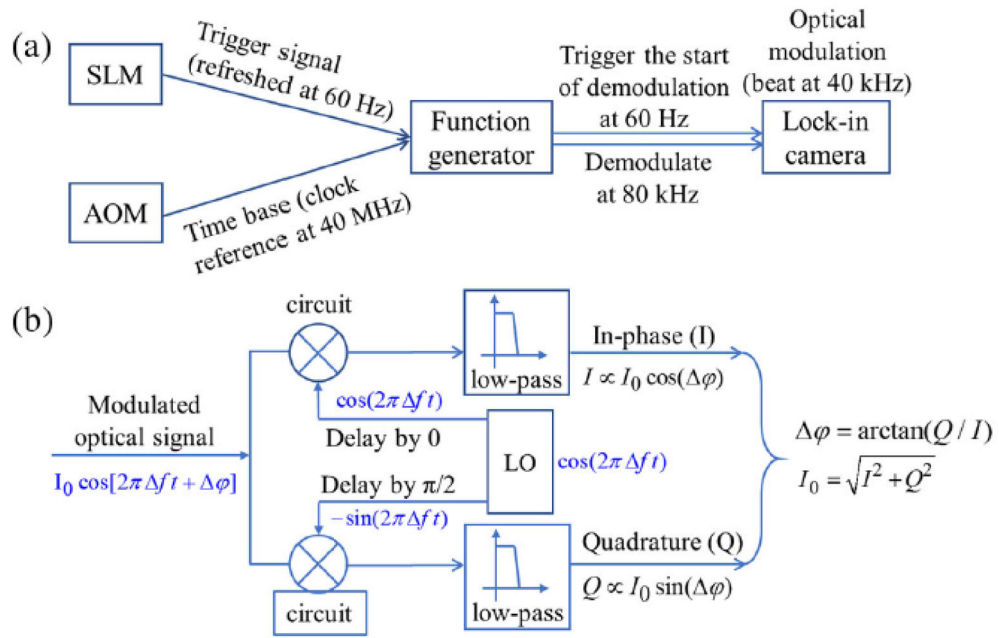


Fig. 3. Flow chart of the control signal and illustration of signal processing. (a) Hardware connection of the proposed system; (b) signal processing flow inside the driver of the lock-in camera.

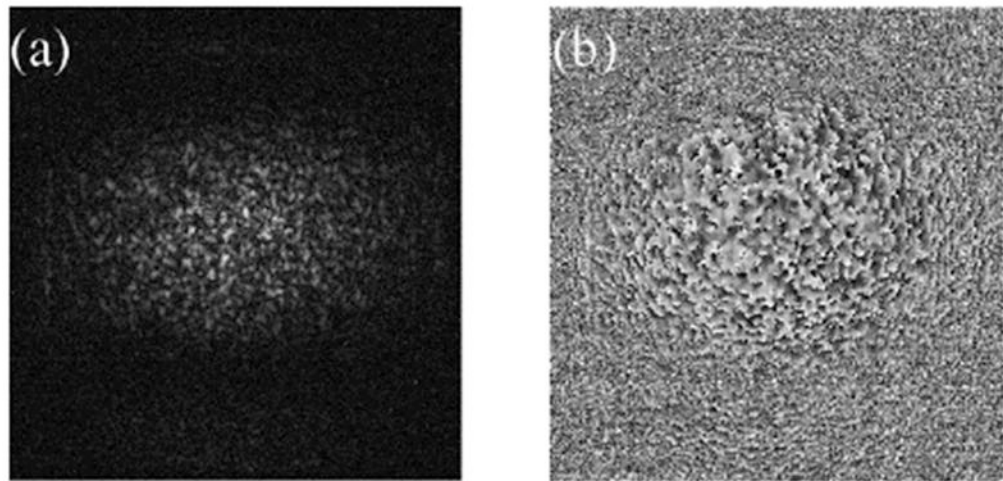


Fig. 4. Back-scattered complex amplitude retrieval by the heterodyne-detection scheme. (a) Amplitude and (b) the phase profile in the camera plane corresponding to one scanning point, as shown in Fig. 1.

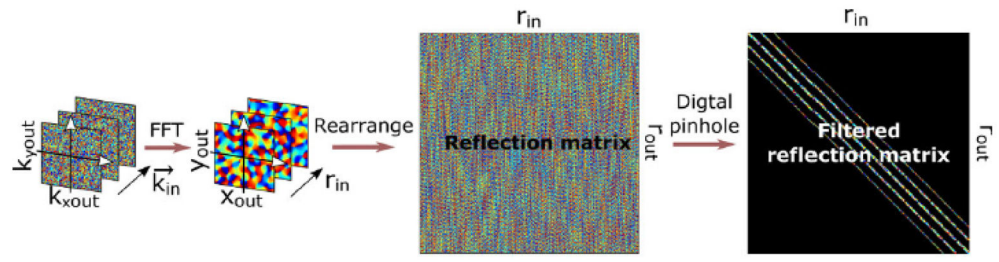


Fig. 5.
Combination of multiple complex frames into one filtered matrix.

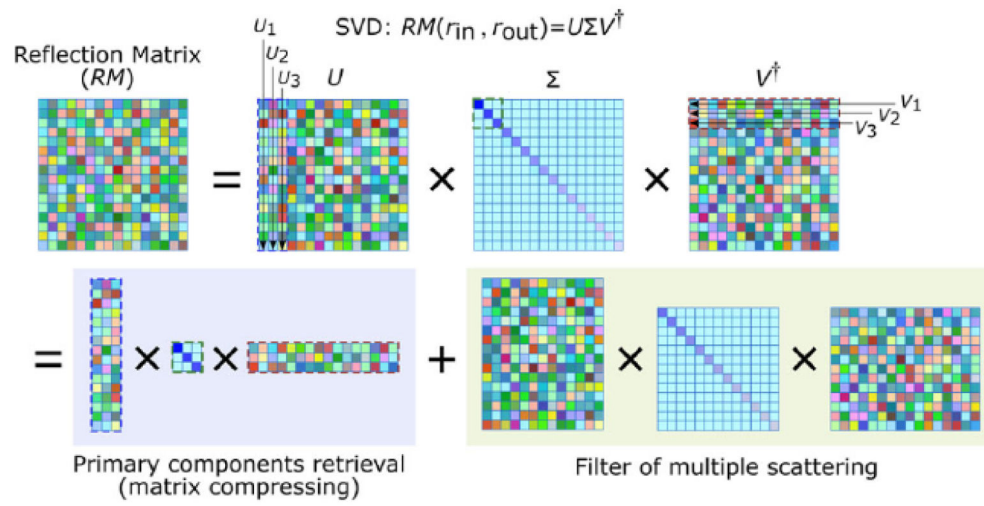


Fig. 6. Primary component analysis for removing multiple scattering noises in the SVD process.

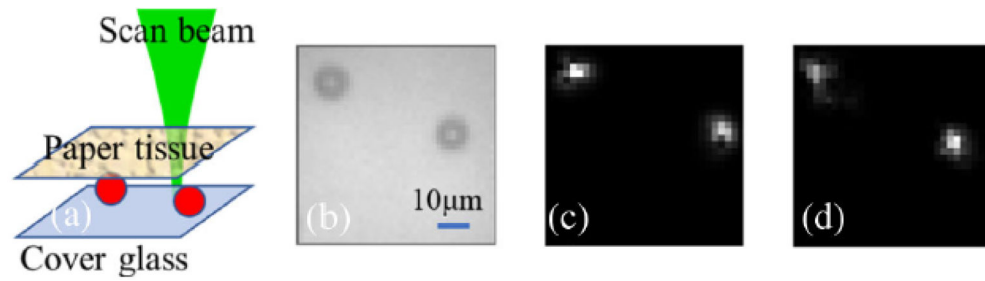


Fig. 7.

Image reconstruction results of the proposed method. (a) Two microbead targets are placed on a cover glass, while a white paper tissue (Amazon Basics 92 Bright Multipurpose Copy Paper) is put ~ 1 mm above; (b) standard microscopic image of the two TiO_2 beads when the scattering paper tissue is removed; the reconstructed image of two microbeads by analyzing the filtered reflection matrix which is measured (c) without and (d) with the scattering tissue placed above.

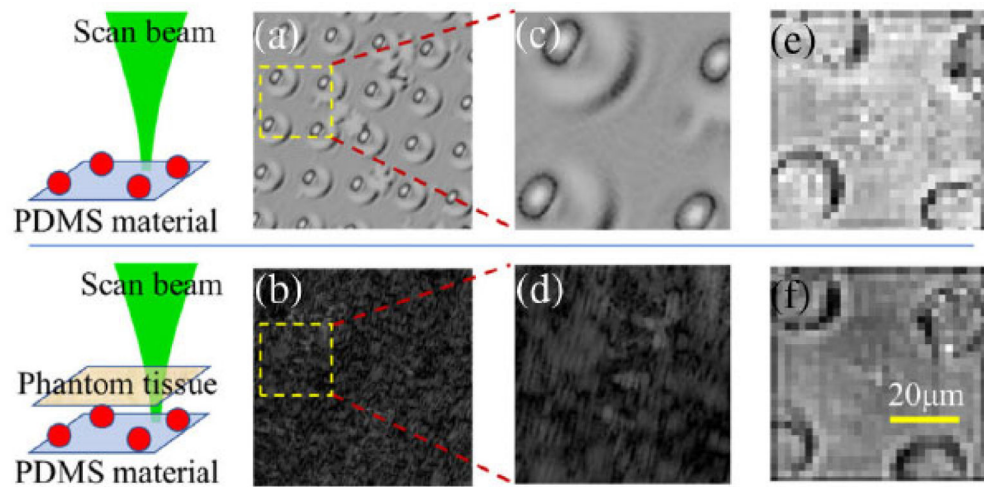


Fig. 8.

(a) Standard *en-face* OCT image by Thorlabs OCS1310V1 OCT where microbeads are formed on the flexible PDMS; (b) enface OCT image of the same beads by the OCS1310V1 when the phantom tissue is put above the beads; (c) and (d) the zoomed images corresponding to the field of view inside the yellow square, as shown in (a) and (b), respectively; (e) and (f) reconstructed images of the microbeads in the same field of view, (e) without and (f) with the phantom tissue put above by the proposed matrix measurement method.