UC Irvine UC Irvine Previously Published Works

Title

Methane Oxidation, Production, and Emission at Contrasting Sites in a Boreal Bog

Permalink https://escholarship.org/uc/item/0hz6m483

Journal Geomicrobiology Journal, 17(3)

ISSN 0149-0451

Author Whalen, WS Reeburgh SC

Publication Date 2000-07-01

DOI

10.1080/01490450050121198

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at https://creativecommons.org/licenses/by/4.0/

Peer reviewed



Methane Oxidation, Production, and Emission at Contrasting Sites in a Boreal Bog

S. C. WHALEN

Department of Environmental Sciences and Engineering University of North Carolina Chapel Hill, North Carolina, USA

W. S. REEBURGH

Department of Earth System Science University of California—Irvine Irvine, California, USA

Boreal peatlands, a major source of atmospheric CH_4 , are characterized by a rapidly fluctuating water table position and meter-scale variations in relief. Regional and ecosystem-based studies show that water table position generally controls CH_4 emission from boreal peatlands by influencing the relative extent of the zones of CH_4 oxidation and production within the peat profile. We used a combined field and laboratory study to assess the influence of local hydrology on the short-term dynamics of CH₄ production, oxidation, and emission from sites in an Alaskan boreal peatland that were characterized by temporarily (site LB1A) and permanently (LB2) water-saturated subsurface peat during the thaw season. The two sites contrasted sharply with respect to the dynamics of CH_4 cycling. Site LB1A, which showed low CH_4 concentrations in pore water (<2 μ M) and unsaturated peat (<2.6 nM), consumed both atmospheric CH_4 and CH_4 diffusing upward from the saturated zone for a net flux of $-0.9 \text{ mg } CH_4 \text{ m}^{-2} d^{-1}$. In contrast, LB2 had pore water CH₄ concentrations, 300 μ M and emitted CH₄ at 69 mg m⁻² d⁻¹. Roughly 55% of the CH_4 diffusing upward from the saturated zone at LB2 was oxidized in transit to the peat surface. Methane oxidation potentials (V_{ox}) were maximum in the 10-cm zone immediately above the local water table at both sites but were greater on a dry mass (dw) basis at LB2 (498-650 ng CH₄ $g_{dw}^{-1} h^{-1}$) than at LB1A (220-233 ng CH₄ $g_{dw}^{-1} h^{-1}$). Methane production potentials (V_p) were low (<2 ng CH₄ $g_{dw}^{-1} h^{-1}$) at LB1A, but the maximum at LB2 (139 ng CH₄ $g_{dw}^{-1} h^{-1}$) was spatially coupled with the maximum V_{ox} . Methanogens exposed to O_2 produced no CH_4 in a subsequent 48 h anoxic incubation, whereas methanotrophs incubated anoxically oxidized CH_4 vigorously within 20 h of return to an oxic environment, indicating that the former are more sensitive than the latter to adverse O_2 conditions. Experiments with ¹⁴CH₄ showed that ~71% of assimilated ¹⁴CH₄ was respired as ${}^{14}CO_2$. Respiration by methanotrophs contributes at most ~ 1.1–1.7% (molar basis) of gross ecosystem respiration (15.6–17.9 mg $CO_2 m^{-2} d^{-1}$) at these sites.

Keywords boreal bog, methane oxidation, methanogenesis, peatlands

A 1% annual increase in the atmospheric concentration of the radiatively important trace gas CH_4 over the last 200 years is well documented (Houghton et al. 1996), and CH_4 currently

Received 29 February 2000; accepted 1 May 2000.

This research was supported by the U.S. Environmental Protection Agency and the National Institute for Global Environmental Change.

Address correspondence to S. C. Whalen, Department of Environmental Sciences and Engineering, CB #7400, University of North Carolina, Chapel Hill, NC 27599-7400, USA. E-mail: steve_whalen@unc.edu

contributes ~20% to global warming (Bouwman 1990). The relative importance of CH_4 as a greenhouse gas is expected to increase, given that, on a mass basis, CH_4 has a global warming potential 56 times that of CO_2 in a 20-year time horizon (Houghton et al. 1996).

Wetlands are the largest natural source of atmospheric CH₄, contributing 110 Tg annually, or 20% of the total emission from all sources (Fung et al. 1991). High-latitude (>45 °N) peatlands emit 38 Tg of CH₄ annually (Bartlett and Harriss 1993) and are therefore particularly important in the atmospheric CH₄ budget. Methane emission from peatlands depends in part on the balance between methanogenesis in an anaerobic, subsurface zone and CH₄ oxidation by methanotrophs in an overlying, aerobic layer (Whalen and Reeburgh 1992; Whalen et al. 1996). Dissolved O₂ profiles in peatlands indicate that the oxic zone extends from the surface of the depth of the water saturation or a few centimeters below (Benstead and Lloyd 1994; Nedwell and Watson 1995; Whalen et al. 1996). Water table position therefore serves as an indicator of the relative extent of the zones of CH₄ production and oxidation. Accordingly, CH₄ emission from peatlands has been positively correlated with water table position through repeated measurements in individual wetlands (e.g., Crill et al. 1988; Moore et al. 1990; Shannon and White 1994) or by regional comparisons of mean CH₄ fluxes and water table positions (reviewed by Bubier and Moore 1994).

Peatlands are characterized by surface heterogeneity and fluctuating water table position (Clymo and Pearce 1995). In particular, differences in relief and microtopography within a specific high-latitude peatland underlain by permafrost may result in local water table fluctuations ranging from near zero on a seasonal scale to 30 cm over hours to days (Whalen and Reeburgh 1992; Moosavi et al. 1996). This results in differences in the thermal regime, nutrient cycling, plant community composition, and organic matter production on a scale of several meters (Waddington and Roulet 1996). Consequently, studies documenting the generalized relationship between wetland CH_4 emission and water table position at the ecosystem to regional scale lack the temporal and spatial resolution necessary to assess the short-term (days) impact of this environmental variable on the dynamics of methanogenesis and CH_4 oxidation. Nonetheless, there is a growing awareness of the need to augment studies of source strengths with process-level investigations at finer spatiotemporal scales, both to improve our understanding of present climates and to forecast more accurately future trends in the concentration of atmospheric CH_4 (Prinn 1994).

The overall aim of this investigation was to examine the short-term dynamics of CH₄ production and oxidation in a high-latitude peat bog in response to water table position. We selected for study two sites that seasonally showed zones of permanently or transiently water-saturated peat. We combined field observations of CH₄ and CO₂ emission and peat CH₄ profiles with a laboratory study of the extant potentials for CH₄ production and oxidation, the short-term (\leq 2-day) response of these potentials to water table fluctuation, and the distribution of end products (biomass and CO₂) of ¹⁴CH₄ oxidation. We expected higher rates and potentials for CH₄ production at the permanently inundated site, but higher rates and potentials for CH₄ oxidation at the transiently saturated site.

Methods

Field Sites

The study site, Lemeta Bog, is a high boreal wetland complex located near Fairbanks, Alaska (64°53'N, 147°30'W). The plant community changes along a moisture gradient from paper birch (*Betula papyrifa*) and grasses in dry areas of the bog margin to a floating mat of *Carex* spp. and *Sphagnum* in the wettest areas. Moosavi et al. (1996) have given a complete description. We selected for study two sites that best represent the plant community and

hydrologic conditions of the wetland complex. The dry bog site (LB1A) was characterized by an overstory of black spruce (*Picea mariana*) and tamarak (*Larix laricinia*) with an undergrowth of *Sphagnum* invaded by Laborador tea (*Ledum palustre*), cloudberry (*Rubus chamaemorus*), lowbush cranberry (*Vaccinium vitis-ideae*), and blueberry (*Vaccinium uliginosum*). The wet site (LB2) was dominated by *Sphagnum* interspersed with cloudberry, blueberry, Laborador tea cotton grass tussocks (*Eriophorum vaginatum*), and *Carex* spp. Peat extended to permafrost at both sites, and our experiments were conducted in late summer (August) of 1994, when thaw depths were maximum (43 and 63 cm at LB1A and LB2, respectively). Site LB1A is drained to permafrost except during rainfall when the bottom several centimeters of the thaw layer are water-saturated for a few days. In contrast, at LB2 a water-saturated zone persists immediately above the thaw layer throughout the summer and may extend to within 10 cm of the peat surface for several days after rainfall.

Field Sampling

Following from our experimental aims, we made single determinations of CH_4 and CO_2 flux as well as depth distributions of temperature and CH_4 in peat each site. Thereafter, we destructively sampled the peat at the exact location where these measurements were made to assess rates, potentials, and depth distributions of CH_4 oxidation and production in a controlled laboratory environment.

Methane and CO_2 flux determinations were made by using the static chamber technique (Whalen and Reeburgh 1992). Each chamber consisted of a skirted aluminum base seated in the peat and a removable aluminum lid that utilized a water-filled channel for a seal. Lids were equipped with an O-seal fitting to allow syringe sampling of the headspace gas and fitted with a capillary bleed to equilibrate with atmospheric pressure. Following flux measurements, the depth distributions of temperature and CH_4 concentration were determined at 5-cm intervals to 40 (LB1A) or 45 cm (LB2) in the peat isolated by each aluminum chamber base. Temperature was measured with a portable thermistor probe. Methane samples were collected through the unsaturated zone (<32 cm at LB1A, <23 cm at LB2) with 30-mL polyethylene syringes fitted with a two-way stopcock and a removable steel tube (1 mm [i.d.] \times 1 m long). Similarly, 30-mL pore water samples were collected into 60-mL polyethylene syringes in the saturated zone. The following day, peat within each aluminum chamber base was sampled to 40 cm with a 15-cm-diameter stainless steel coring apparatus. Each 40-cm core was cut horizontally into eight 5-cm sections that were then divided into triplicate subcores (6.7 cm in diameter $\times 5 \text{ cm}$ long) by using a plastic tube. A single subcore from each depth interval in the saturated zone was immediately placed in a Mason jar (capacity ~ 1 L), amended with 180 mL of pore water from that depth, and sealed under ultrapure N_2 . Additionally, one subcore from each 5-cm depth interval in the lower unsaturated zone (15-30 cm at LB1A, 10-20 cm at LB2) was amended with 180 ml of composite pore water and sealed under N₂. All other core sections from both the saturated and unsaturated zones were sealed in Mason jars under ambient atmosphere for transport to the laboratory.

Laboratory Studies

All laboratory experiments were conducted on the 6.7-cm-diameter \times 5-cm-long core sections in 1-L Mason jars capped with lids fitted with a septum for syringe sampling of the headspace gas. Core sections used for CH₄ oxidation experiments were disjoined and spread at the bottom of the jars before experimentation to avoid any limitation of diffusion and to prevent formation of anoxic zones. During experimentation, all jars were maintained in a controlled temperature environment within $\pm 1^{\circ}$ C of the temperature at the depth of sample collection. Headspace CH₄ concentrations were adjusted to a concentration that had been shown previously to give zero-order kinetics (600 μ 1 L⁻¹; Whalen and Reeburgh 1996), and the resulting reduction in concentration was monitored by hourly sampling over an 8-h time course. Immediately after determination of the CH_4 oxidation potential, a single subcore from each sample depth was randomly selected for assay for the end products of CH_4 oxidation in a single endpoint experiment. Each sample was equilibrated in an ambient air atmosphere and amended with microliter quantities of biogenically produced ${}^{14}CH_4$ (Daniels and Ziekus 1983) tracer (3.5 kBq; specific activity 2005 kBq mol⁻¹). Experiments were terminated after a 6-h incubation in the dark by adding 2 cm³ of acetylene (Bedard and Knowles 1989). Jar headspaces were flushed with He into a stripping/oxidation line where 14 CO₂ was trapped directly and 14 CH₄ was trapped as 14 CO₂ after combustion. Soils were freeze-dried and assayed by dry combustion for ¹⁴C incorporated into microbial biomass. The radioactivity of all samples was determined by liquid scintillation spectrometry. The remaining core section from each depth interval was analyzed for pH, organic content, bulk density, particle density, and percent water-filled pore space. Methodological details of radiocarbon experiments and physicochemical assays are given in Whalen and Reeburgh (1992) and Whalen et al. (1992).

Methane production was determined for the core sections amended with pore water and sealed with N_2 in the field. On return to the lab, the pore water was stripped of CH_4 by bubbling with N_2 for 1 h, the jars were resealed under ultrapure N_2 , and the time course for CH_4 accumulation was measured over 54 h. Cores were allowed to drain for 20 h in an ambient air atmosphere and resealed in Mason jars. Headspaces were adjusted to CH_4 contents of $\sim 20 \,\mu l \, L^{-1}$, and the time course for change in CH_4 concentration was determined over the next 45 h to assess the ability of methanotrophs to resume activity after temporary anoxia. Finally, cores were rewetted with pore water and made anoxic as described above, after which the time course for CH_4 accumulation was reassessed for 48 h to evaluate the ability of methanogens to resume CH_4 production after transient exposure to an ambient atmosphere. This switch between oxic and anoxic conditions was intended to simulate a rapid drop and rise in the water table position, respectively.

Gas Analysis and Calculations

Methane was stripped from pore water samples by equilibration with an equal volume of ultrapure N₂ (McAuliffe 1971). Methane and CO₂ determinations were made by flame ionization and thermal conductivity gas chromatography, respectively, with a precision of < 1% in both cases (Whalen et al. 1992). Calibration gases were relatable to standards from the National Institute for Technology and Standards. Soil CH₄ in the unsaturated zone is expressed as the equilibrium aqueous-phase concentration calcuated with Bunsen solubility coefficients (Yamamoto et al. 1976) so that data from both the unsaturated and waterlogged zones can be presented in comparable units. Details of all calculations, including integration of dry mass–normalized (g_{dw}^{-1}) rates of microbial activity in individual core sections to yield area-based (m⁻²) rates, are given in Whalen et al. (1992) and Whalen and Reeburgh (1992).

Results

The two sampling sites were similar with respect to pH, organic content, and percent airfilled pore space in the unsaturated zone. The pH of core sections varied from 3.7 to 4.7, and the organic content varied from 88% to 96% (average 93%). No depth-dependent



FIGURE 1 Peat temperature distributions at sites LB1A and LB2 in Lemeta Bog.

trends were noted for either variable. Bulk density increased with depth at both sites, from $\sim 0.02 \text{ g cm}^{-3}$ in the 0- to 5-cm zone to 0.25 g cm⁻³ in the 35- to 40-cm zone. The percentage of air-filled pore space decreased from $\sim 81\%$ in the 0- to 5-cm zone at both sites to 43% (LB1A) and 25% (LB2) just above the saturated zone. Peat temperatures decreased with increasing depth at both sites (Figure 1). Relatively lower temperatures at LB1A, especially in the surface peat, were due to shading by the larch and black spruce overstory, which was absent at LB2.

The two sites showed distinctly different depth distributions for CH_4 (Figure 2). The CH_4 concentration in the unsaturated zone of LB1A was lowest (1.8 nM) at 20 cm, and increased slightly with increasing distance both above and below this depth. Methane concentrations in the unsaturated zone were also in the nanomolar range, but decreased steadily to the peat surface. Methane concentrations in the saturated zone were three and five orders of magnitude greater than concentrations in the unsaturated zone at LB1A and LB2,



FIGURE 2 Peat CH₄ distributions at sites LB1A and LB2 in Lemeta Bog. Each value for the unsaturated zone is the calculated aqueous-phase CH₄ concentrations in equilibrium with the measured CH₄ concentration in the air-filled pore space at the temperature of the depth of sample collection. Note the change in scale for CH₄ concentrations between the unsaturated and saturated zones.

respectively. Thus, CH_4 concentrations in the saturated zone at LB2 were ~ 100-fold higher than at LB1.

Methane fluxes in static chamber experiments differed in both sign and magnitude between sites. The flux at LB1A (-0.9 mg of CH₄ m⁻² d⁻¹) indicated consumption of atmospheric CH₄, whereas that at LB2 (69 mg of CH₄ m⁻² d⁻¹) indicated net CH₄ emission from the peat surface to the atmosphere. In contrast to CH₄, fluxes of CO₂ were reasonably similar between sites. Carbon dioxide emission in static chamber experiments was 15.6 and 17.9 g m⁻² d⁻¹ at LB1A and LB2, respectively.

Depth distributions of CH₄ oxidation potential (V_{ox}) were similar between sites; that is, the greatest rates were observed in a 10-cm horizon of the unsaturated zone immediately above the local water table (Figure 3). V_{ox} values in this horizon were about threefold greater at LB2 than at LB1A, ranging from 498 to 650 ng of CH₄ g_{dw}^{-1} h⁻¹ at the former and from 220 to 233 ng of CH₄ g_{dw}^{-1} h⁻¹ at the latter. Methane oxidation potentials in deep (30-40 cm), water-saturated peat were similar at both sites (21 to 33 ng of CH₄ g_{dw}^{-1} h⁻¹) and an order of magnitude or more lower than the V_{ox} immediately above the saturated zone. Methane oxidation potentials in surface peat (0-10 cm) were roughly comparable at both sites (32 to 75 ng of CH₄ g_{dw}^{-1} h⁻¹) and slightly higher than values deep in the saturated zone.

Depth profiles for oxidation of 14 CH₄ (Figure 4) were similar to those for V_{ox} (Figure 3), in that a subsurface maximum was observed in the unsaturated zone just above the water table at both sites. The fraction of assimilated radiocarbon that was respired ranged from 45% to 98% (mean ± SD : 71% ± 4%) for the 16 core sections exposed to 14 CH₄. Fractional respiration showed no pattern with respect to depth at either site and was not significantly different between sites. Mass balances for each core section showed full recovery (100% ± 7%; mean ± SD) of added label as 14 C-biomass, 14 CO₂, and unassimilated 14 CH₄.



FIGURE 3 Depth distribution of CH_4 oxidation potential (V_{ox}) in cores collected from two sites in Lemeta Bog. The mean V_{ox} of duplicate subcores is plotted at the midpoint of each 5-cm sampling interval. Error bars are eliminated for clarity, but the standard deviation averaged 21%. The water table position is indicated by solid (LB1A) and open (LB2) arrows.

Experiments assessing the depth distribution of potential CH₄ production (V_p) at LB1A showed extremely limited V_p in 5-cm core sections from 25 to 40 cm (0.1 to 0.2 ng of CH₄ $g_{dw}^{-1} h^{-1}$) and no V_p at the higher peat horizons tested (15 to 25 cm). In contrast, V_p for 5-cm core sections from 20 to 40 cm deep at LB2 ranged from 32 to 139 ng of CH₄ $g_{dw}^{-1} h^{-1}$, and modest V_p values (4 to 10 ng of CH₄ $g_{dw}^{-1} h^{-1}$) were still evident in higher (10 to 20 cm) peat horizons. Methane began accumulating without lag in all core sections showing V_p at both sites (e.g., Figure 5).



FIGURE 4 Depth distributions of ¹⁴CH₄ consumption (V) in cores from two sites in Lemeta Bog, showing the partitioning of assimilated radiocarbon into respiration (¹⁴CO₂) and microbial biomass. Values of V are plotted at the midpoint of each 5-cm depth interval. Horizontal dashed lines indicate the water table position.



FIGURE 5 Time courses for CH₄ production (left) and oxidation (right) in selected 5-cm core sections (from site LB2 in Lemeta Bog) that were alternately made anoxic and oxic. Methane production by anoxic core sections plus pore water was measured for 54 h. Core sections were drained for 20 h in an oxic environment and then amended with ~20 μ l L⁻¹ CH₄, after which the time course for CH₄ oxidation was determined for 45 h. Not shown: no CH₄ was produced over 48 h by continued incubation of core sections rewetted and made anoxic at 120 h.



FIGURE 6 Depth distribution of CH_4 oxidation and production potentials in cores collected from two sites in Lemeta Bog. Methane oxidation and production potentials for an entire 6.7-cm-diameter × 5-cm-long core section are plotted at the midpoints of each depth interval. Horizontal dashed lines indicate the water table position.

Core sections from the saturated zone of LB2 that were incubated anaerobically in pore water in experiments assessing V_p showed vigorous CH₄ oxidation after draining for 20 h in an ambient air atmosphere and subsequent exposure to a CH₄ headspace of ~20 µl L⁻¹ (Figure 5). Headspace CH₄ concentrations decreased to <1 µl L⁻¹ (subatmospheric) within 45 h for all core sections. Core sections from the saturated zone of LB1A (30-40 cm) behaved similarly when drained and exposed to CH₄ in an oxic environment (data not shown). Core sections from both sites that were subsequently rewetted with pore water and again rendered anaerobic showed no CH₄ production over the next 48 h (data not shown).

When the total dry peat mass in each 5-cm core section was considered, LB1A showed low V_p throughout the peat profile but a high V_{ox} in the unsaturated zone immediately above the water table (Figure 6). In contrast, LB2 showed both high V_p and high V_{ox} at the interface of the saturated and unsaturated peat. The area-based CH₄ oxidation potential (Σ V_{ox}) of 153 mg m⁻² d⁻¹ at LB2 was ~1.5 times the value of 91 mg CH₄ m⁻² d⁻¹ calculated for LB1A. Site-wise differences were even more pronounced for area-based CH₄ production potentials (Σ V_p), where Σ V_p was 39 and <1 mg of CH₄ m⁻² d⁻¹ for LB2 and LB1A, respectively. Thus, Σ V_{ox} exceeded Σ V_p at both sites.

Discussion

The two experimental sites were reasonably similar at comparable depths with respect to pH, percent organic matter, temperature, bulk density, and percent air-filled porosity in the unsaturated zone. However, differences in local hydrology on a scale of meters (continuous [LB2] versus transient [LB1A] presence of a saturated zone during the thaw season) yielded distinct site-wise differences in CH_4 oxidation and production potentials, CH_4 fluxes, and peat CH_4 profiles.

Peat CH₄ concentrations in the unsaturated zone at LB1A (Figure 2) were consistently at or below the aqueous-phase concentration (~ 2.5 nM) in equilibrium with atmospheric

CH₄ (1.8 μ l L⁻¹), whereas CH₄ concentrations in the saturated zone clearly exceeded this value. Thus, peat in the unsaturated zone of LB1A consumed not only atmospheric CH₄, but also all of the CH₄ diffusing upward from the saturated zone. This corroborates well the measured CH₄ flux of $-0.9 \text{ mg m}^{-2} \text{ d}^{-1}$ and also agrees with data from similar sites within this bog ("dry sites"; Moosavi et al. 1996) that oscillate between net CH₄ emission and consumption at low rates throughout the thaw season.

In contrast to LB1A, LB2 showed net CH4 emission to the atmosphere. The measured CH_4 flux of 69 mg m⁻² d⁻¹ is consistent with the 1992 and 1993 seasonal averages of 72 and 43 mg m⁻² d⁻¹ reported by Moosavi et al. (1996) for similar sites ("wet sites") within this bog. Methane concentrations as great as 306 μ M in the saturated zone and nanomolar concentrations immediately above in the unsaturated zone suggest intense CH₄ oxidation at the interface of these two peat horizons (Figure 2). Moreover, the concave upward CH_4 profile in the unsaturated zone (Figure 2) suggests continued oxidation of the CH₄ diffusing upward to the peat surface. Using a stagnant film model for gas transfer and assuming a film thickness of 200 μ m at the water table surface (Whalen et al. 1996), we estimate a CH₄ flux of 152 mg m⁻² d⁻¹ from the water table surface to the unsaturated zone. Comparing this estimated flux with the measured emission from the peat surface implies consumption of 83 mg of CH₄ m⁻² d⁻¹ in the unsaturated zone. Thus, we estimate that 55% of the CH₄ emitted from the water table surface is oxidized while diffusing upward through the unsaturated zone. Our estimate for the fractional oxidation of CH₄ at LB2 is somewhat less than that for other wet peats (King et al. 1990; Yavitt et al. 1990; Fenchner and Hemond 1992), which show $\sim 70\%$ to 90% oxidation of available CH₄. However, our estimate is based on a single observation, and fractional oxidation clearly depends on the position of the water table. Harriss et al. (1982) observed that temperate swamp soils may function seasonally as a source or a sink for atmospheric CH_4 , depending on the water table position.

Methane oxidation potential is generally considered to provide a relative measure of the methanotrophic biomass because measurements are made at uptake-saturating CH₄ concentrations (Sundh et al. 1994). Our observation of the greatest V_{ox} immediately above the saturated zone (Figures 3 and 6) compares favorably with other studies in peatlands, which show the most activity in a similar zone or in anaerobic peat within several centimeters below the local water table (Bubier et al. 1993; Lien et al. 1993; Sundh et al. 1993, 1994, 1995; Nedwell and Watson 1995; Moore and Dalva 1997; Saarnio et al. 1997; Watson et al. 1997; Kettunen et al. 1999). Methanotrophs remain viable after an extended period of anoxia (King 1996). Thus, variability between studies in the location of the greatest V_{ox} probably best reflects the average seasonal condition where CH₄ and O₂ supplies are adequate rather than the water table position at the time of sampling.

Our ranges for V_{ox} (15–650 ng of $CH_4 g_{dw}^{-1} h^{-1}$) and ΣV_{ox} (91–153 mg of $CH_4 m^{-2} d^{-1}$) are generally lower than data for other peatlands. Reported values of V_{ox} range from <1 to 10,390 ng of $CH_4 g_{dw}^{-1} h^{-1}$ for various Canadian wetlands (Moore and Knowles 1990; Bubier et al. 1993; Moore and Dalva 1997) and from 1280 to 5120 ng of $CH_4 g_{dw}^{-1} h^{-1}$ for a Finnish pine fen (Saarnio et al. 1997), whereas we calculate values as great as 23,000 ng of $CH_4 g_{dw}^{-1} h^{-1}$ for an English blanket bog from data given by McDonald et al. (1996). Reported values of ΣV_{ox} range from 40 to 22,100 mg of $CH_4 m^{-2} d^{-1}$ for the Finnish pine fen already mentioned (Saarnio et al. 1997). However, unlike those studies, which were conducted at room temperature, our values were obtained at the in situ peat temperatures. We previously reported an optimum temperature of 23 °C for substrate-saturated CH_4 oxidation in these peats (Whalen and Reeburgh 1996). However, temperatures at the depths of the greatest V_{ox} and ΣV_{ox} values were $\sim 5-7^{\circ}C$ (cf. Figures 1, 3, and 6), indicating that much greater rates are possible at room temperature.

Our data for LB2 indicating that the locations for the Vox and Vp maxima are spatially similar (Figure 6) agree with other reports for peat environments that show proximity or overlap of these potentials (Bubier et al. 1993; Sundh et al. 1993, 1994; Nedwell and Watson 1995; Moore and Dalva 1997; Saarnio 1997; Edwards et al. 1998; Kettunen et al. 1999). Our values of <1-139 ng of $CH_4 g_{dw}^{-1} h^{-1}$ for V_p are similar to the ranges of 0-270, 8-121, and 1-58 ng of $CH_4 g_{dw}^{-1} h^{-1}$ given by Magnusson (1993), Moore and Knowles (1990), and Bubier et al. (1993), respectively, for Swedish Sphagnum peats, a subarctic Canadian fen, and 12 Canadian peatland sites. However, our data are generally lower than those from other peatland studies that used similar methodology to measure V_p . Methane production potentials averaged 129 and 3640 ng of CH₄ g_{dw}⁻¹ h⁻¹ in a comprehensive study of Canadian wetlands (Moore and Dalva 1997) and a Minnesota fen (Williams and Crawford 1984), respectively. Values for V_p ranged from ~100 to 2288 ng of CH₄ g_{dw}^{-1} h⁻¹ for samples collected below the water table in a fen (Saarnio et al. 1997) and in two mires (Kettunnen et al. 1999) in Finland and from 70 to 10,400 ng of $CH_4 g_{dw}^{-1} h^{-1}$ in a temperate bog (Yavitt et al. 1987). As with V_{ox} , our values for V_p probably would have been higher if they had been determined at room temperature because rates of methanogenesis are notably temperature-dependent (e.g., Dunfield et al. 1993).

Here, and in most of the studies cited above, $V_{ox} > V_p$. However, the two values are not directly comparable (Sundh et al. 1994; Moore and Dalva 1997). The latter involves no addition of methanogenic substrate and therefore gives CH₄ production from endogenous substrate only for a methanogenic community that is probably substrate-limited (Valentine et al. 1994; Yavitt et al. 1997; Bergman et al. 1998).

The presence of a viable methanotrophic community in anoxic Lemeta Bog peat is clearly illustrated by the resumption of CH_4 oxidation on a time scale of < 1 day after exposure to ambient air atmosphere (Figure 5). We cannot discount an even shorter response time, because CH₄ consumption measurements were not initiated until 20 h after O₂ reintroduction. Rapid recovery of CH₄ oxidizing activity after anoxia has previously been attributed to reactivation of the indigenous population by O_2 (King 1996; Edwards et al. 1998); the response time is too rapid to involve the germination of cysts and spores, which may take days (Whittenbury et al. 1970). Vegetative cells were probably responsible here also. King et al. (1990) attributed a reduction in post- to preanoxia relative CH_4 oxidizing potential in a Danish wetland sediment to the co-occurrence of populations that recover slowly (days) and those that recover rapidly after reexposure to O_2 . The relative importance of these two functional groups in Lemeta Bog is unclear. However, the highly dynamic water table position favors the development of a methanotrophic population capable of rapid recovery from anoxia, and the consumption of 20 μ l L⁻¹ CH₄ to subatmospheric concentrations (1 μ l L⁻¹) within 48 h by reoxygenated samples (Figure 5) clearly points to a vigorous population of anoxia-tolerant methanotrophs. A rapid response capability to O2 reintroduction may be widespread for at least part of the methanotrophic community in peat. Yavitt et al. (1990) reported immediate CH₄ consumption for surface peats in a temperate bog, but a lag of 12 h for peats 30 to 40 cm deep. Edwards et al. (1998) and McDonald et al. (1996) detected CH₄-oxidizing activity throughout 30-cm-deep cores from a UK blanket bog after ≤ 2 h exposure to air, whereas King et al. (1990) reported immediate CH₄ oxidation by anoxic subtropical peats incubated with O2, and Roslev and King (1996) observed CH4 oxidation by anoxic temperate marshland peat 1 to 7 h after O₂ addition.

We are unaware of other studies analyzing the immediate impact of a change in the O_2 environment on V_p and V_{ox} simultaneously. Although the time course for recovery of methanogenic activity after exposure to O_2 is uncertain, the absence of CH₄ production for 48 h after samples were again made anaerobic clearly points to a greater short-term sensitivity by methanogens than menthanotrophs in Lemeta Bog to adverse conditions

with respect to O_2 . Studies assessing the influence of O_2 exposure on methanogenesis in environmental samples are few and give conflicting results. In general agreement with our data, Ratering and Conrad (1998) observed that CH_4 production in slurries of an Italian rice soil was completely inhibited by a 48-h exposure to air and that this inhibition persisted for 30 days. Similarly, Öquist and Sundh (1998) found that a 40-day exposure of boreal bog peat to air resulted in a 1-week delay in CH_4 production in surface samples and complete inhibition of methanogenesis in deep peat (50 to 55 cm) during the subsequent 27-day anoxic incubation. In contrast, Schutz et al. (1989) observed that incubation of paddy soil in air for 6 h had no effect on methanogenesis, whereas Mayer and Conrad (1990) demonstrated CH_4 production in previously air-dried paddy soil within 2 days after being placed in an anaerobic atmosphere. Clearly, efforts to model peatland CH_4 emission will benefit from an improved understanding of the apparent hysteresis and recovery of activity by methanogens in the response to a shift from oxic to anoxic conditions that will accompany a rising water table.

The low CH₄ concentration in pore water at the infrequently flooded LB1A site (Figure 2) is consistent with demonstrated O₂ sensitivity of Lemeta Bog methanogens (Figure 5). However, relatively high V_{ox} in the 20- to 30-cm-deep horizon (Figure 3) suggests that the residence time of pore water during periodic flooding is sometimes sufficient to allow for methanogenesis. Insofar as aerobic decomposition in normally unsaturated peat reduces the availability of methanogenic precursors during periods of anoxia (Sundh et al. 1994; Kettunnen et al. 1999), it is likely that rates of CH₄ production at LB1A are always less than at LB2 and that the CH₄ oxidizers efficiently intercept upwardly diffusing CH₄ such that the net flux to the atmosphere is near zero. The observation that similar sites in Lemeta Bog alternate between weak CH₄ emitters and consumers throughout the thaw season (Moosavi et al. 1996) supports this contention. Low V_p in core sections from 25 to 40 cm deep at LB1A (0.1 to 0.2 ng of CH₄ g_{dw}⁻¹ h⁻¹) may reflect not only poor substrate quality but also the recent history of the site, in that the time between saturation and sampling may have been insufficient to activate or develop the methanogenic community fully.

Both methanogens and methanotrophs probably are attached to peat particles and remain in the same layer of the peat matrix despite water table fluctuations (Kettunen et al. 1999). Given the high O₂ sensitivity of methanogens in this peat, the fact that the greatest values of V_p and V_{ox} at LB2 overlap in space is enigmatic (Figure 6). However, anaerobic microzones supporting methanogenesis can persist in seemingly dry peat (Öquist and Sundh 1998), although peat higher in the profile is generally more readily decomposed (e.g., Hogg 1993). Consequently, the locus of V_p probably reflects a balance between O₂ tolerence and substrate supply. Methanotrophs not only are able to withstand extended periods of anoxia but also are capable of functioning at dissolved O₂ concentrations as low as 0.1 mg L⁻¹ (Rudd et al. 1976). Hence, the spatial coupling of V_p and V_{ox} is not surprising. Spatial overlap of aerobic respiration and methanogenesis is a well-established component of the CH₄ production process in ecosystems that are characterized by changing O₂ conditions (Wagner et al. 1999).

Recovery of $71\% \pm 4\%$ of assimilated ¹⁴CH₄ as ¹⁴CO₂ (Figure 4) indicates that most of the CH₄ carbon consumed by the peat microbial community was used for energy rather than biomass. Studies assessing the relative allocation of assimilated CH₄ give highly variable results. In agreement with our data, Yavitt et al. (1988, 1990) reported a respiratory loss of 78–85% of assimilated CH₄ for Appalachian peatlands, and Whalen et al. (1992) showed a 60% loss for boreal forest soils. However, other terrestrial environments show less loss to respiration. Roughly 30% of assimilated CH₄ was respired by landfill cover soils (Whalen et al. 1990; Jones and Nedwell 1993) and by a cultivated humisol (Megraw and Knowles 1987). Freshwater and marine ecosystems also show a wide range for respiratory loss of CH₄, 39–98% (summarized in Whalen et al. 1990). Differences among studies may reflect differences among CH₄-oxidizing communities or among their physiological states (Megraw and Knowles 1987). Assuming that 71% of ΣV_{ox} is respired, we calculate (molar basis) as an upper limit that 1.1–1.7% of the observed respiratory CO₂ flux in static chamber measurements is the result of CH₄ oxidation. Thus, although CH₄ oxidation serves as an important modulator of CH₄ emission from Lemeta Bog to the atmosphere, it contributes little to the gross ecosystem respiration.

In conclusion, this study demonstrates that meter-scale differences in local hydrology in a boreal bog may give sharp contrasts in CH_4 production and oxidation, which result in highly variable atmospheric CH_4 fluxes on a similar scale. The methanotrophic and methanogenic communities show spatial overlap, but respond on different time scales to the changes in peat O_2 status that may result from rapid changes in water table position. Process-oriented studies such as these will allow mechanisms underlying CH_4 fluxes to be better incorporated into models to improve our predictive capabilities regarding peatland response to future climates. Future study should be directed toward better understanding of the O_2 sensitivity of methanogenesis in these ecosystems.

References

- Bartlett KB, Harriss RC. 1993. Review and assessment of methane emissions from wetlands. Chemosphere 26:261–320.
- Bedard C, Knowles R. 1989. Physiology, biochemistry and specific inhibitors of CH₄, NH₄⁺ and CO oxidation by methylotrophs and nitrifiers. Microbiol Rev 53:68–84.
- Benstead J, Lloyd D. 1994. Direct mass spectrometric measurements of gases in peat cores. FEMS Microbiol Ecol 13:233–240.
- Bergman I, Svensson BH, Nilsson M. 1998. Regulation of methane production in a Swedish acid mire by pH, temperature and substrate. Soil Biol Biochem 30:729–741.
- Bouwman AF. 1990. Exchange of greenhouse gases between terrestrial ecosystems and the atmosphere. In: Bouwman AF, editor. Soils and the greenhouse effect. New York: Wiley, p 61–127.
- Bubier J, Moore TR. 1994. An ecological perspective on methane emissions from northern wetlands. Trends Ecol Evol 9:460–464.
- Bubier J, Costello A, Moore TR, Roulet NT, Savage K. 1993. Microtopography and methane flux in boreal peatlands, northern Ontario, Canada. Can J Bot 71:1056–1063.
- Clymo RS, Pearce DME. 1995. Methane and carbon dioxide production in, transport through, and efflux from a peatland. Philos Trans R Soc Lond A350:249–259.
- Crill PM, Bartlett KB, Harriss RC, Gorham E, Verry ES, Sebacher DI, Madzar I, Sanner W. 1988. Methane flux from Minnesota peatlands. Global Biogeochem Cycles 2:371–384.
- Daniels L, Zeikus JG. 1983. Convenient biological preparation of pure high specific activity CH₄labeled methane. J Labelled Compd Radiopharm 20:17–24.
- Dunfield P, Knowles R, Dumont R, Moore TR. 1993. Methane production and consumption in temperate and subarctic peat soils: response to temperature and pH. Soil Biol Biochem 25:321–326.
- Edwards C, Hales BA, Hall GH, McDonald IR, Murrell JC, Pickup R, Ritchie DA, Saunders JR, Simon BM, Upton M. 1998. Microbiological processes in the terrestrial carbon cycle: methane cycling in peat. Atmos Environ 32:3247–3255.
- Fechner E, Hemond HF. 1992. Methane transport and oxidation in the unsaturated zone of a *Sphagnum* peatland. Global Biogeochem Cycles 6:33–44.
- Fung I, John J, Lerner J, Matthews E, Prather M, Steele LP, Fraser PJ. 1991. Three-dimensional model synthesis of the global methane cycle. J Geophys Res 96:13033–13065.
- Harriss RC, Sebacher DI, Day FP Jr. 1982. Methane flux in the Great Dismal Swamp. Nature 292:673-674.
- Hogg EH. 1993. Decay potential of hummock and hollow Sphagnum peats at different depths in a Swedish raised bog. Oikos 66:269–278.

- Houghton JT, Filho LG, Bruce J, Lee H, Callander BA, Harris N, Kattenberg A, Maskell K, editors. 1996. Climate change 1995. The science of climate change. Contribution of Working Group I to the Second Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge: Cambridge University Press, 572p.
- Jones HA, Nedwell DB. 1993. Methane emission and methane oxidation in a land-fill cover soil. FEMS Microbiol Ecol 102:185–195.
- Kettunen A, Kaitala V, Lehtinen A, Lohila A, Alm J, Silvola J, Martikainen PJ. 1999. Methane production and oxidation potentials in relation to water table fluctuations in two boreal mires. Soil Biol Biochem 31:1741–1749.
- King GM. 1996. Physiological limitations of methanotrophic activity in situ. In: Murrell JC, Kelley DP, editors. Microbiology of atmospheric trace gases. Berlin: Springer-Verlag, p 17–32.
- King GM, Roslev P, Skovgaard H. 1990. Distribution and rate of methane oxidation in sediments of the Florida everglades. Appl Environ Microbiiol 56:2902–2911.
- Lien T, Martikainen P, Nykänen H, Bakken L. 1993. Methane oxidation and methane fluxes in two drained peat soils. Suo 43:231–236.
- Magnusson T. 1993. Carbon dioxide and methane formation in forest mineral and peat soils during aerobic and anaerobic incubations. Soil Biol Biochem 25:877-883.
- Mayer HP, Conrad R. 1990. Factors influencing the population of methanogenic bacteria and the initiation of methane production upon flooding of paddy soil. FEMS Microbiol Ecol 73:103–112.
- McAuliffe CC. 1971. Gas chromatographic determination of solutes by multiple phase equilibration. Chem Technol 1:46–51.
- McDonald IR, Hall GH, Pickup RW, Murrell JC. 1996. Methane oxidation potential and preliminary analysis of metanotrophs in blanket bog peat using molecular ecology techniques. FEMS Microbiol Ecol 21:197–211.
- Megraw SR, Knowles R. 1987. Methane production and consumption in a cultivated humisol. Biol Fertil Soils 5:56–60.
- Moore TR, Dalva M. 1997. Methane and carbon dioxide exchange potentials of peat soils in aerobic and anaerobic laboratory incubations. Soil Biol Biochem 29:1157–1164.
- Moore TR, Knowles R. 1990. Methane emission from fen, bog and swamp peatlands in Quebec. Biogeochemistry 11:45-61.
- Moore TR, Roulet N, Knowles R. 1990. Spatial and temporal variations of methane flux from subarctic/northern boreal fens. Global Biogeochem Cycles 4:29-46.
- Moosavi SC, Crill PM, Pullman ER, Funk DW, Peterson KM. 1996. Controls on CH₄ flux from an Alaskan boreal wetland. Global Biogeochem Cycles 10:287–296.
- Nedwell DB, Watson A. 1995. CH₄ production, oxidation and emission in a U.K. ombotrophic peat bog: influence of SO_4^{2-} from acid rain. Soil Biol Biochem 27:893–903.
- Öquist M, Sundh I. 1998. Effects of a transient oxic period on mineralization of organic matter to CH₄ and CO₂ in anoxic peat incubations. Geomicrobiol J 15:325–333.
- Prinn RG. 1994. The interactive atmosphere: global atmospheric-biospheric chemistry. Ambio 23:50-61.
- Ratering S, Conrad R. 1998. Effects of short-term drainage and aeration on the production of methane in submerged rice soil. Global Change Biol 4:397–407.
- Roslev P, King GM. 1996. Regulation of methane oxidation in a freshwater wetland by water table changes and anoxia. FEMS Microbiol Ecol 19:105–115.
- Rudd JWM, Furutani A, Flett RJ, Hamilton RD. 1976. Factors controlling methane oxidation in Shield Lakes: the role of nitrogen fixation and oxygen concentration. Limnol Oceanogr 21:357–364.
- Saarnio S, Alm J, Silvola J, Lohila A, Nykanen H, Martikainen PJ. 1997. Seasonal variation in CH₄ emissions and production and oxidation potentials at microsites on an oligotrophic pine fen. Oecologia 110:414-422.
- Schutz H, Seiler W, Conrad R. 1989. Processes involved in the formation and emission of methane in rice paddies. Biogeochemistry 7:33–53.
- Shannon RD, White JR. 1994. A three-year study of controls on methane emissions from two Michigan peatlands. Biogeochemistry 27:35–60.

- Sundh I, Mikkelä C, Nilsson M, Svensson BH. 1995. Potential aerobic methane oxidation in a Sphagnum-dominated peatland: controlling factors in relation to methane emission. Soil Biol Biochem 27:829–837.
- Sundh I, Nilsson M, Granberg G, Svensson BH. 1994. Depth distribution of microbial production and oxidation of methane in northern peatlands. Microbial Ecol 27:253–265.
- Sundh I, Nilsson M, Svensson BH. 1993. Depth distribution of methane production and oxidation in a sphagnum peat bog. Suo 43:267–269.
- Valentine DW, Holland EA, Schimel DS. 1994. Ecosystem and physiological controls over methane production in a northern wetland. J Geophys Res 99:1563–1571.
- Waddington JM, Roulet NT. 1996. Atmosphere-wetland carbon exchanges: scale dependency of CO_2 and CH_4 exchange on the developmental topography of a peatland. Global Biogeochem Cycles 10:233–245.
- Wagner D, Pfeiffer E-M, Bock E. 1999. Methane production in aerated marshland and model soils: effects of microflora and soil texture. Soil Biol Biochem 31:999–1006.
- Watson A, Stephen KD, Nedwell DB, Arah JRM. 1997. Oxidation of methane in peat: kinetics of CH₄ and O2 removal and the role of plant roots. Soil Biol Biochem 29:1257–1267.
- Whalen SC, Reeburgh WS. 1992. Interannual variations in tundra methane emission: a 4-year time series at fixed sites. Global Biogeochem Cycles 6:139–159.
- Whalen SC, Reeburgh WS. 1996. Moisture and temperature sensitivity of CH₄ oxidation in boreal soils. Soil Biol Biochem 28:1271–1281.
- Whalen SC, Reeburgh WS, Barber VA. 1992. Oxidation of methane in boreal forest soils: a comparison of seven measures. Biogeochemistry 16:181–211.
- Whalen SC, Reeburgh WS, Reimers CE. 1996. Controls of tundra methane emission by microbial oxidation. In: Reynolds JF, Tenhunen JD, editors. Landscape function and disturbance in arctic tundra. Berlin: Springer-Verlag, p 257–274.
- Whalen SC, Reeburgh WS, Sandbeck KA. 1990. Rapid methane oxidation in a landfill cover soil. Appl Environ Microbiol 56:3405–3411.
- Whittenbury R, Davies SL, Davey JF. 1970. Exospores and cysts formed by methane utilizing bacteria. J Gen Microbiol 61:219–226.
- Williams RT, Crawford RL. 1984. Methane production in Minnesota peatlands. Appl Environ Microbiol 47:1266–1271.
- Yamamoto S, Alcauskas JB, Crozier TE. 1976. Solubility of methane in distilled water and seawater. J Chem Eng Data 21:78–80.
- Yavitt JB, Downey DM, Lancaster E, Lang GE. 1990. Methane consumption in decomposing Sphagnum-derived peat. Soil Biol Biochem 22:441–447.
- Yavitt JB, Lang GE, Downey DM. 1988. Potential methane production and methane oxidation rates in peatland ecosystems of the Appalachian Mountains, United States. Global Biogeochem Cycles 2:253–268.
- Yavitt JB, Lang GE, Wider RK. 1987. Control of carbon mineralization to CH₄ and CO₂ in anaerobic, *Sphagnum*-derived peat from Big Run Bog, West Virginia. Biogeochemistry 4:141–157.
- Yavitt JB, Williams CJ, Wieder RK. 1997. Production of methane and carbon dioxide in peatland ecosystems across North America: effects of temperature, aeration and organic chemistry of peat. Geomicrobiol J 14:299–316.