UC Irvine UC Irvine Previously Published Works

Title

Imaging of in vivo pseudoxanthoma elasticum via multiphoton microscopy and optical coherence tomography

Permalink https://escholarship.org/uc/item/0rj2p43z

Journal JAAD Case Reports, 6(8)

ISSN 2352-5126

Authors

Mehrabi, Joseph N Doong, Judy Lentsch, Griffin <u>et al.</u>

Publication Date 2020-08-01

DOI

10.1016/j.jdcr.2020.05.029

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <u>https://creativecommons.org/licenses/by/4.0/</u>

Peer reviewed

Imaging of in vivo pseudoxanthoma elasticum via multiphoton microscopy and optical coherence tomography



Joseph N. Mehrabi, MS,^a Judy Doong, MD,^a Griffin Lentsch, MS,^b and Natasha Mesinkovska, MD, PhD^a Irvine, California

Key words: coherence tomography; collagen; elastic fibers; multiphoton microscopy; optical reflectance confocal microscopy; pseudoxanthoma elasticum.

INTRODUCTION

Pseudoxanthoma elasticum (PXE) is an autosomal recessive disorder of abnormal elastic tissue deposition and calcification.¹ PXE primarily affects the elastic fibers in the dermis of the skin, Bruch membrane of the eye, and the media and intima of midsized arteries. The skin findings may present as small yellow, orange, or cream-colored papules, resembling plucked-chicken skin or cobblestone. The skin on the neck and skin flexures may be lax and redundant because of the impaired elastic features, causing a degree of psychosocial distress. Histologic changes of the skin consist of fragmented and disorganized elastin fibers in the mid and deep reticular dermis with overlying calcium deposits.

The diagnosis of PXE is primarily clinical with histologic confirmation. Multiphoton microscopy (MPM) is a noninvasive imaging technique that allows visualization of skin components. MPM is based on optical signals generated through nonlinear light-matter interactions. MPM can easily contrast keratinocytes, collagen, and elastin fibers to a depth of 200 μ m. Optical coherence tomography (OCT) is an imaging technology that uses nearinfrared light and the reflective properties of tissue to generate real-time, noninvasive cross-sectional images of biological tissues to a depth of 1 to 2 mm. OCT makes it possible to visualize structures in the skin such as the epidermis, dermoepidermal junction (DEJ), dermis, hair follicles, sweat glands, and blood vessels. Here we investigate the ability of MPM and OCT to provide a noninvasive diagnosis of skin in a patient with PXE.

Funding sources: None.

Conflicts of interest: None disclosed.

Abbreviations used:

DEJ: dermoepidermal junction MPM: multiphoton microscopy

OCT: optical coherence tomography

PXE: pseudoxanthoma elasticum

CASE REPORT

We present the case of a 65-year-old woman with a long-standing, biopsy-proven diagnosis of PXE. She had subtle yellowish skin papules coalescing into plaques on the lateral neck with skin laxity of the axilla (Fig 1), face, neck, and groin. Noncutaneous history of PXE was significant for angioid streaks, atrophic retinal break of the right eye, angina pectoris, myocardial infarction, peripheral artery disease, coronary artery disease, vitreous degeneration, and bilateral posterior vitreous detachment.

Noninvasive imaging with MPM and OCT was performed to visualize any irregularities in dermal elastin and/or calcium deposits. The patient consented to the imaging procedure, which was approved by the Institutional Review Board. We imaged an area of lax skin on the right axilla with dynamic OCT (VivoSight Dx, Michelson Diagnostics Ltd, London, UK) and MPM tomography (MPTflex, JenLab GmbH, Jena, Germany) and processed images using the freely available ImageJ software (National Institutes of Heath, Bethesda, MD).

Multiphoton microscopy

Specifications and settings of the MPM tomography system and methods are outlined in prior work.²⁻⁴

2352-5126

https://doi.org/10.1016/j.jdcr.2020.05.029

From the Department of Dermatology^a and the Beckman Laser Institute, Laser Microbeam and Medical Program,^b University of California Irvine.

Correspondence to: Joseph N. Mehrabi, MS, Department of Dermatology, University of California, Irvine, 118 Med Surge 1, Irvine, CA 92617. E-mail: jmehrabi@uci.edu.

JAAD Case Reports 2020;6:702-4.

^{© 2020} by the American Academy of Dermatology, Inc. Published by Elsevier, Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/ 4.0/).



Fig 1. PXE in the patient's scanned, right axilla. Note the lax and redundant skin.

The patient's images showed an abundance of chopped, fragmented, and irregular elastin bundles in the lower part of the visualized papillary dermis about 30 μ m below the DEJ with clearance of elastin bundles in the reticular dermis (Fig 2). Calcium deposits were undetectable via visual inspection of MPM images.

Optical coherence tomography

OCT imaging of the lax axilla skin displayed large areas of hyporeflective, homogeneous areas of attenuation, or signal loss, in the mid dermis (Fig 3). In contrast to normal skin, which attenuates the image at about 0.7 to 0.9 mm, the depth of the PXE image exhibits relative superficial attenuation of 50% at 0.3 mm and near complete attenuation at a depth of about 0.53 mm. The observed changes may be associated with the noted abundance of elastin at and above this level on histology.

DISCUSSION

PXE is a systemic condition with multiple phenotypes, making it difficult to diagnose clinically. Noninvasive imaging with MPM was able to capture a dermatologic feature of PXE consistent with fragmented and irregular dermal elastin. Although these findings are not specific for PXE, we find that MPM can be a useful diagnostic tool. The contrast mechanisms of MPM are based on second harmonic generation from collagen fibers and 2-photon—excited fluorescence from nicotinamide adenine dinucleotide plus hydrogen, flavin adenine dinucleotide, keratin, melanin, and elastin fibers. MPM images of age-controlled normal skin from a non—sun-exposed area (Fig 3) leverage the contrast between the autofluorescence from elastin and the second harmonic generation from collagen in the dermis, revealing the harmony of normal dermal collagen and elastin and making such MPM images of PXE clearly characteristic of the condition.

Based on our literature search, no prior reports of in vivo MPM of PXE have been performed. Ex vivo MPM analysis of PXE has demonstrated similar findings showing higher densities and larger amounts of elastin in the dermis with direct comparison to histopathologic images.^{5,6}

OCT imaging of PXE has shown dermal clusters of hyporeflective aggregates with peripheral clefts, which our images do not show.⁷ Another imaging modality known as *reflectance confocal microscopy* (RCM) has been used to image perforating PXE in a prior study, showing a disorganized arrangement at the DEJ and hyperreflective, amorphous material in the papillary dermis which resembled eggs in a basket.⁸ Reflectance confocal microscopy images of our patient's axilla, however, demonstrated no such findings.

Multiple skin diseases require biopsy for diagnosis, and PXE is one such diagnosis that would benefit from histologic confirmation. MPM offers an alternative noninvasive diagnostic method that spares the patient from an invasive biopsy. The main limitation of this modality is that MPM has yet to be widely adopted. It is expensive, imaging time is lengthy, imaging depth maximizes at 200 μ m, its field of view is minimal, and scanning curved skin surfaces is challenging due to the large, 1.5-cm lens. OCT offers another method of dermal imaging at a maximum depth of 1 to 2 mm for some devices, covering the deeper parts of the dermis in a shorter amount of time but at a much lower resolution. Therefore, it cannot yet be used as a reliable means of diagnosis.

The differential diagnosis of PXE includes cutis laxa, solar elastosis, mid-dermal elastolysis, PXE-like papillary dermal elastolysis, and fibroelastolytic papulosis, which are all consequences of altered elastin fibers. PXE-like histologic findings have also been reported in diseases like lipodermatosclerosis, morphea profunda, erythema nodosum, granuloma annulare, lichen sclerosis, tumefactive lipedema, and basal cell carcinoma. Optical imaging in dermatology is in early developmental stages, and we aim to advance the process and help establish guidelines in diagnostic optical imaging of these diseases.

This report describes the findings of MPM and OCT imaging of PXE in vivo, offering what is known as an *optical biopsy*. We show PXE can adequately be imaged in vivo with MPM and OCT to aid in diagnosis, setting the stage to avoid unnecessary



Fig 2. MPM images of normal skin (left) vs PXE (right), 30 µm below the DEJ. PXE is characterized by dense bundles of fragmented elastin (bright green) surrounded by organized collagen (blue). Normal, non-sun-exposed skin of a different patient controlled for age shows organized collagen and harmonized elastin.



Approximate depth of MPM image captured in Figure 2

Fig 3. OCT cross-section of PXE shows homogenous areas of premature signal loss (yellow, dotted circle), with 50% of total attenuation at 0.3 mm below the skin surface and nearcomplete attenuation at about 0.53 mm. The approximate depth of the MPM image at 30 μ m below the DEJ is marked by the red line, and an approximation of the maximum depth MPM

can penetrate is marked by the blue line.

procedures when a diagnosis is uncertain as new versions of these improved technologies are being developed.

REFERENCES

- 1. Sherer DW, Sapadin AN, Lebwohl MG. Pseudoxanthoma elasticum: an update. Dermatology. 1999;199(1):3-7.
- 2. Balu M, Kelly KM, Zachary CB, et al. Distinguishing between benign and malignant melanocytic nevi by in vivo multiphoton microscopy. Cancer Res. 2014;74(10):2688-2697.
- 3. Balu M, Zachary CB, Harris RM, et al. In vivo multiphoton microscopy of basal cell carcinoma. JAMA Dermatol. 2015; 151(10):1068-1074.
- 4. Balu M, Lentsch G, Korta DZ, et al. In vivo multiphoton-microscopy of picosecond-laser-induced optical breakdown in human skin. Lasers Surg Med. 2017;49(6):555-562.

- 5. Murata T, Honda T, Miyachi Y, Kabashima K. Morphological character of pseudoxanthoma elasticum observed by multiphoton microscopy. J Dermatol Sci. 2013;72(2): 199-201.
- 6. Tong PL, Qin J, Cooper CL, et al. A quantitative approach to histopathological dissection of elastin-related disorders using multiphoton microscopy. Br J Dermatol. 2013;169(4): 869-879.
- 7. Persechino F, Giordano D, Marini CD, Franceschini C, Ardigo M, Persechino S. Dermoscopy, optical coherence tomography, and histological correlation of pseudoxanthoma elasticum. Dermatol Pract Concept. 2019;9(3):209-210.
- 8. Nasca MR, Lacarrubba F, Caltabiano R, Verzi AE, Micali G. Perforating pseudoxanthoma elasticum with secondary elastosis perforans serpiginosa-like changes: dermoscopy, confocal microscopy and histopathological correlation. J Cutan Pathol. 2016;43(11):1021-1024.