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Mitochondrial alarmins released by degenerating motor axon terminals activate perisynaptic Schwann cells

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An acute and highly reproducible motor axon terminal degeneration followed by complete regeneration is induced by some animal presynaptic neurotoxins, representing an appropriate and controlled system to dissect the molecular mechanisms underlying degeneration and regeneration of peripheral nerve terminals. We have previously shown that nerve terminals exposed to spider or snake presynaptic neurotoxins degenerate as a result of calcium overload and mitochondrial failure. Here we show that toxin-treated primary neurons release signaling molecules derived from mitochondria: hydrogen peroxide, mitochondrial DNA, and cytochrome c. These molecules activate isolated primary Schwann cells, Schwann cells cocultured with neurons and at neuromuscular junction in vivo through the MAPK pathway. We propose that this inter- and intracellular signaling is involved in triggering the regeneration of peripheral nerve terminals affected by other forms of neurodegenerative diseases.

Significance

The neuromuscular junction is the site of transmission of the nerve impulse to the muscle. This finely tuned synapse relies on at least three components: the motor neuron, the muscle fiber, and the Schwann cells, which assist nerve recovery after injury. Using animal neurotoxins to induce an acute and reversible nerve degeneration, we have identified several mitochondrial molecules through which the damaged nerve terminal communicates with nearby cells, activating signaling pathways in Schwann cells involved in nerve regeneration. Among these messengers, hydrogen peroxide appears to be crucial at the initial stages of regeneration, because its inactivation delays the functional recovery of the damaged neuromuscular junction in vivo. These findings provide important indications about the pharmacological treatment of traumatized patients.

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The authors declare no conflict of interest.

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degeneration and regeneration, which is likely to provide information useful to the understanding of the pathogenesis not only of envenomation but also, more in general, of other human pathological syndromes.

Cell death and injury often lead to the release or exposure of intracellular molecules called damage-associated molecular patterns (DAMPs) or alarmins. Recently, mitochondria have emerged as major sources of DAMPs (25). Mitochondria are abundant subcellular components of the NMJ that have been recently shown to release mitochondrial DNA (mtDNA) and cytochrome c (Cyt c) after trauma or snake myotoxin-induced muscle damage, thus contributing to the systemic or local inflammatory responses associated with such conditions (26, 27). In this study, we tested whether α-Ltx and SPANs induce the release of mitochondrial signaling molecules from primary neuronal cultures and found that, in addition to mtDNA and Cyt c, hydrogen peroxide (H$_2$O$_2$) is released. First candidate targets of these mitochondrial mediators released by damaged neurons are nonmyelinating PSCs, which are intimately associated with the end plate. They play an active role in the formation, function, maintenance, and repair of the NMJ (28–33). PSC activation parallels nerve degeneration and contributes to neuronal regeneration by phagocytosis of cellular debris and by extension of processes that guide reinnervation (34, 35). We therefore investigated whether mitochondrial DAMPs released by injured neurons were able to activate SCs, and through which downstream pathway. Using isolated primary cells, neuron-Schwann cell cocultures, and the NMJ in vivo, we found that PSCs are activated by mitochondrial alarmins and that the MAPK signaling pathway is involved in this process.

**Results**

**Hydrogen Peroxide Is Produced by Neurons Exposed to Spider or Snake Presynaptic Neurotoxins.** Given that mitochondria of stressed cells produce superoxide anion, which is rapidly converted into H$_2$O$_2$, and that in neurons exposed to the neurotoxins, mitochondria functionality is impaired, we asked whether intoxication of neurons by α-Ltx or SPANs leads to H$_2$O$_2$ production, an ideal candidate as intercellular signaling molecule (36–38). We therefore loaded rat cerebellar granular neurons (CGNs) with specific H$_2$O$_2$ probes with different cellular localization and monitored the samples for up to an hour. MitoPY1 is a bifunctional molecule that combines a chemoselective boronate-based switch and a mitochondrial-targeting phosphonium moiety for the detection of mitochondrial reactive oxygen species (ROS) in live cells. Using this probe, we observed a significant increase in fluorescent signal in neurons exposed to α-Ltx or SPANs, indicating that these neurotoxins induce the production of H$_2$O$_2$. This finding is consistent with previous reports demonstrating that α-Ltx and SPANs cause mitochondrial dysfunction, leading to increased ROS production.

**Fig. 1.** Live-imaging of neuronal hydrogen peroxide production. Rat CGNs were loaded with the H$_2$O$_2$-specific probes PF6-AM (A) or MitoPY1 (B), washed, and then exposed to Tpx 6 nM or α-Ltx 0.1 nM for 50 min. Changes in fluorescence resulting from H$_2$O$_2$ production were monitored with time and expressed as a percentage of the fluorescence value at $t = 0$ (Right). ***P < 0.001. Arrows in bright-field images and in the green channel point to neuronal bulges. (Scale bars: 10 μm.)
of H$_2$O$_2$, localized to mitochondria (39). PF6-AM takes advantage of multiple masked carboxylates to increase cellular retention, and hence sensitivity to low levels of peroxide. In its ester-protected form, PF6-AM can readily enter cells. Once inside cells, the protecting groups are rapidly cleaved by intracellular esterases to produce their anionic carbamate forms, which are effectively trapped within cells (40).

After exposure to α-Ltx or Tpx, H$_2$O$_2$ levels increased with time, markedly at the level of neurite enlargements (so-called bulges), which are a hallmark of intoxication (16, 41), as shown in Fig. 1. Bulges are sites of calcium overload and accumulation of depolarized mitochondria (18), and the MitofPy1 signal indicates that these mitochondria produce H$_2$O$_2$. Quantification of the signals indicates a more pronounced effect of α-Ltx with respect to Tpx, in agreement with the fact that the pore formed by the former neurotoxin mediates a larger Ca$^{2+}$ entry than Tpx (21). Similar results were obtained following intoxication of rat spinal cord motor neurons (MNs; Fig. S1). That mitochondria are the major source of H$_2$O$_2$ is reinforced by the finding that toxins failed to induce membrane translocation of cytoplasmic p47phox, a regulatory component of the NADPH oxidase complex, which excludes a role of the NADPH oxidase system (Fig. S2).

**Hydrogen Peroxide Released by Degenerating Nerve Terminals Activates Schwann Cells and Stimulates Regeneration.** Growing evidence indicates that H$_2$O$_2$ is a largely used intercellular signaling molecule regulating kinase-driven pathways (37, 38, 42); it triggers ERK phosphorylation in different cell types (43), with consequent activation of downstream gene transcription, and ERK signaling was recently shown to play a central role in the orchestration of axon repair by SCs (44, 45).

In preliminary experiments, we checked whether primary SCs isolated from rat sciatic nerves were responsive to H$_2$O$_2$ by analyzing ERK phosphorylation by Western blotting and immunofluorescence. Exposure of primary SCs to H$_2$O$_2$ led to ERK phosphorylation and translocation of p-ERK into the nucleus (Fig. S3 A and B). Cocultures of primary spinal cord motor neurons and sciatic nerve-derived SCs were then exposed to α-Ltx or Tpx; bulges appeared within few minutes along neuronal processes, and p-ERK was detected in the cytoplasm and nucleus of SCs (Fig. 2A). Phospho-ERK-positive cells were also positive for S-100, a specific SC marker (Fig. S3C). In cocultures, the score of S-100-positive cells that become p-ERK positive is 59% on intoxication with β-Btx (n = 81) and 78% in the case of α-Ltx (n = 69). These percentages were obtained by counting many S-100-positive cells randomly distributed in different fields, but the value is actually much higher if one considers only clustered SCs in close proximity of intoxicated neurites; this observation further supports the conclusion that molecules released by injured neurons reach nearby SCs, thus activating them.

ERK phosphorylation was reduced in cocultures preincubated with catalase, which converts H$_2$O$_2$ into water and O$_2$, indicating that H$_2$O$_2$ produced inside neurons diffuses to reach nearby SCs, contributing to their ERK activation (Fig. 2B). Residual p-ERK signal might be a result of mediators other than H$_2$O$_2$ released on neuronal injury. Toxins failed to induce a direct ERK phosphorylation either in isolated SCs (Fig. S3D) or in isolated primary neurons (Fig. 2B).

Next we tested whether the ERK pathway is activated also within PSCs at the NMJs of intoxicated mice. Sublethal doses of the neurotoxins were s.c. injected in transgenic mice expressing a cytoplasmic GFP specifically in SCs under the plp promoter (46, 47), in proximity to the levator auris longus (LAL) (48), a thin muscle ideal for imaging. Twenty-four hours later, muscles were collected and processed for indirect immunohistochemistry. A clear p-ERK signal was detected at the level of PSCs in treated NMJs, thus extending in vivo the results obtained in cocultures.
The present toxin-based model of acute nerve degeneration is long known to follow nerve terminal damage (35), and therefore, fibers by the first day of injection (Fig. 2C). The importance of ERK pathway for SCs activation and regeneration was addressed by a pharmacologic approach: SCs-MNs cocultures exposed to the neurotoxins show a decreased ERK phosphorylation in the presence of the MEK 1 inhibitor PD98059 (Fig. S4, A and B); moreover, soleus muscles of mice pretreated with PD98059 and then locally injected with α-Ltx show a delayed recovery from paralysis with respect to mice injected with toxin only (Fig. S4C).

PSCs respond to neurotoxin-induced nerve degeneration by forming long sproutings and bridges between junctions of different fibers by the first day of injection (Fig. S5). This response has been long known to follow nerve terminal damage (35), and therefore, the present toxin-based model of acute nerve degeneration reproduces the known crucial aspects of regeneration.

To test whether H$_2$O$_2$ production by injured nerve terminals is important for functional regeneration, we performed electrophysiological recordings at soleus NMJs 16, 24, 48, and 72 h after i.m. injections of α-Ltx alone or α-Ltx plus catalase. Three days after treatment, fibers injected with α-Ltx plus catalase showed evoked junction potentials (EJPs) with significantly smaller amplitudes than those injected only with the toxin, indicating a slowdown of the regeneration process; muscles treated with catalase alone showed EJPs indistinguishable from the control (Fig. 2D).

Immunohistochemistry on LAL muscles treated as described earlier confirmed the electrophysiological results, showing a delay in the recovery of synaptosomal-associated protein 25 (SNAP-25) staining, a presynaptic marker, in samples exposed to α-Ltx plus catalase compared with muscles injected with α-Ltx only (Fig. S6). At 24 h, SNAP-25 staining is recovered in 80% of the NMJs treated with α-Ltx (90% at 48 h) compared with 17% of the NMJs treated with α-Ltx plus catalase (33% at 48 h; n = 40). The disappearance of SNAP-25 during the degeneration steps takes place with a closely similar kinetic under the two conditions (Fig. S6). Four hours after intoxication, SNAP-25 displays a spotty distribution in nearly all NMJs analyzed (indicative of nerve terminal degeneration), both in the presence and absence of catalase; at 16 h, 68% of α-Ltx-treated NMJs have no more SNAP-25 versus 60% of catalase and α-Ltx-treated NMJs (n = 30).

**Fig. 3.** Mitochondrial DNA and cytochrome c are released by degenerating neurons and activate the ERK pathway, together with hydrogen peroxide. (A) Real-time qPCR performed on CGNs supernatants from control and toxin-treated samples (Tpx 6 nM or α-Ltx 0.1 nM for 50 min), using primers specific for rat mitochondrial genes Cyt b and NADH dhl. DNA copy numbers of control and treated samples have been quantified. *P < 0.05; n = 11. (B) Supernatants from control and neurons treated as described earlier were precipitated with TCA and probed for Cyt c immunoreactivity in Western blot. (C) Time-course of ERK-phosphorylation induced in primary SCs by H$_2$O$_2$ (10 μM), mtDNA (10 μg/mL), and Cyt c (1 μg/mL) added alone or in a mixture and the relative quantification. Phospho-ERK signal was normalized to the Hsc70 band. *P < 0.05; **P < 0.01; n = 3.

mitDNA and Cyt c Are Released by Degenerating Neurons and Activate the ERK Pathway in Schwann Cells. We next tested whether mtDNA and Cyt c could act together with H$_2$O$_2$ as neuronal mediators of PSCs activation. For mtDNA detection, primary neurons were intoxicated, the supernatants collected, and DNA purified. The eluates were subjected to real-time PCR, using primers specific for the rat mitochondrial genes Cyt b and NADH dhl. Fig. 3A shows that mtDNA is indeed released in the neuronal supernatant after treatment with Tpx or α-Ltx. In another set of experiments, TCA-precipitated cell supernatants (sham or toxin-treated) were loaded in SDS/PAGE, followed by Western blotting. Samples were probed with an antibody against Cyt c: only toxin-treated samples showed a clear band corresponding to the intact, monomorphic form of the protein (Fig. 3B). Control experiments showed no amplification when primers for the nuclear gene GAPDH were used (Fig. S7A), and the LDH assay on neuronal supernatant excluded a loss of membrane integrity (Fig. S7B). Thioredoxin 2, a mitochondrial protein with a molecular weight similar to Cyt c, was undetectable by Western blot of toxin-treated supernatants precipitated with TCA, thus supporting the conclusion that neuronal alarmins are released from intact membranes (Fig. S7C). Moreover, CGNs loaded with calcine-AM did not lose dye during 50 min incubation with both the toxins, indicating conservation of plasma membrane integrity (Fig. S7D).

Exposure of isolated SCs to mtDNA or Cyt c led to a sustained ERK phosphorylation, whereas a peak of p-ERK followed by progressive decline was observed upon H$_2$O$_2$ stimulation. When the three mitochondrial alarmins were added together, an additive effect on ERK phosphorylation was observed (Fig. 3C).

**Mitochondrial Alarmins Exit from Neurons.** H$_2$O$_2$ is permeable to biological membranes (49), whereas mtDNA and Cyt c must be released from mitochondrial and plasma membranes to reach the extracellular medium. Pretreatment of neurons with cyclosporin A, a drug that desensitizes the mitochondrial permeability transition pore (PTP) via its binding to cyclophilin D (50), reduces both mtDNA and Cyt c release triggered by the toxins (Fig. 4A and B), suggesting these molecules can exit mitochondria and
reach the cytoplasm through the PTP, whose opening is indeed induced by snake neurotoxins (51).

Because neuronal plasma membrane integrity is preserved, how do these alarmins reach the extracellular medium? We posited that exosomes might be involved and have purified them from control and treated neuronal supernatants. Purified exosomes were found enriched in Hsp90, Hsc70, flotillin, and CD63; no contamination with Golgi, mitochondrial, or plasma membranes was detected (Fig. 4C and Figs. S8A and B). Electron microscopy and immunogold labeling of purified exosomes confirmed their correct morphology, size, and positivity for Hsp90 (Fig. S8C). Next, we purified total DNA from exosomes and performed real-time qPCR to check for their mtDNA content. Fig. 4D shows that exosomes released by α-Ltx- and β-Btx-intoxicated neurons do contain mtDNA. Similar mtDNA copy numbers were found before and after DNase treatment of exosomal fractions, indicating that mtDNA is indeed inside exosomes (Fig. S8D). In contrast, no Cyt c was detected in exosomes by Western blotting; this is likely to be a result of the much lower sensitivity of Western blotting with respect to RT-PCR, but the possibility that Cyt c is released from damaged nerve terminals via other mechanisms cannot be discarded.

**Phagocytosis Is Induced in PSCs During Nerve Terminal Injury.** During toxin-induced neurodegeneration, PSCs at poisoned NMJs undergo evident morphological changes, showing a number of intracellular structures appearing dense by light microscopy (Fig. 5A, Lower). These structures are particularly evident at 4 h after α-Ltx injection, with a reduction in number and size with time (Fig. 5A).

The appearance and life span of these processes parallel nerve terminal degeneration, suggesting they might be phagosomes involved in the clearance of nerve debris. Accordingly, immunostaining of sham or poisoned LAL muscles for the scavenger macrophage receptor CD68 was performed. After α-Ltx injection, perineurial SCs of LAL NMJs do express CD68 on these intracellular structures, supporting their phagocytic role (Fig. 5B). CD68-positive structures also appear after β-Btx treatment, but at a later time (16 h), as expected on the basis of the different time course of pathogenesis of the two neurotoxins (Fig. 5B). Lysotracker-positive staining confirmed the acidic nature of such compartments (Fig. 5C). CD68-positive macrophages were also recruited in the proximity of neurotoxin-treated NMJs, with a typical migrating phenotype (Fig. 5B); this is consistent with the chemoattractant role of H$_2$O$_2$ (52–54). In contrast, polymorphonuclear leukocytes, which are recruited by axonal degradation (54), were rarely seen in the many samples we have inspected.

Four hours after α-Ltx injection, the distribution of the presynaptic markers neurofilaments (NF) and SNAP-25 is altered, with clear fragmentation in many junctions, as a result of the specific and localized nerve terminal degeneration induced by the neurotoxins (Fig. 6A and B). SNAP-25-positive spots localize within PSCs phagosomes (the same holds true for NF), as shown by orthogonal projections (Fig. 6C), confirming that phagocytosis by PSCs and macrophages is taking place during nerve terminal degeneration.

**Discussion**

The present article describes an original approach to study motor axon terminals degeneration and regeneration. This model system is based on the use of animal presynaptic neurotoxins highly specific for nerve terminals with a well-defined biochemical mechanism of action (10, 12, 16, 18). Here, these neurotoxins are used as tools to induce localized and reversible nerve degeneration, followed by complete regeneration. This system is more controllable than the classical cut and crush approaches, which are invasive and inevitably damage several cell types, triggering a pronounced inflammatory response (55). Moreover, this model avoids some adverse effects of techniques such as laser ablation (high temperatures, photooxidation, etc.). The model proposed here is therefore better suited to...
study the inter- and intracellular signaling and transcriptomic events involved in the regeneration process.

Spider and snake presynaptic neurotoxins induce, by different biochemical mechanisms, a large entry of calcium in axon terminals, which in turn leads to mitochondrial failure and nerve terminal degeneration. At the same time, PSCs perceive the damage occurring to the motor axons and respond by dedifferentiating to a progenitor-like state, proliferating and assisting nerve regeneration. They acquire macrophagic-like activities that contribute to the removal of nerve cell debris and facilitate reinnervation, similar to what was found previously after nerve crush (23). Moreover, upon extensive cytoskeletal reorganization, PSCs send out long projections, along which the regenerating nerve terminals extend sprouts, which originate from the nonmyelinated axon terminal to innervate adjacent denervated junctions (34, 35).

It was recently shown that the MAPK signaling pathway has a central role in controlling SC plasticity and peripheral nerve regeneration via the activation of ERK1/2 and JNK, which activate the transcription complex activator protein 1, of which c-Jun is a key component (44, 45, 56). The major result obtained here, using p-ERK as a read-out, is that alarmins released by mitochondria of degenerating axon terminals activate SCs. Mitochondria are abundant components of the motor axons terminals, and here we define them as a source of mediators that are released under cytosolic calcium overload. The rapid accumulation of Ca$^{2+}$ inside mitochondria causes the opening of the PTP and the exit of alarmins (50). Mitochondrial alterations are hallmarks of nerve terminal damage (19, 20), and therefore the present findings can be extrapolated to several other nerve terminal pathological conditions.

Mitochondria of stressed cells produce reactive oxygen species (ROS), among which H$_2$O$_2$ is the most stable species (37, 38, 57). It is a very reactive molecule that can permeate biological membranes. As PSCs and axon terminals are in close contact within the NMJ, significant amounts of H$_2$O$_2$ released by axon terminals can reach PSCs before it becomes inactivated by cellular antioxidant defense systems. Once within the target cell, H$_2$O$_2$ can act as a second messenger via chemoselective oxidation of cysteine residues in signaling proteins and via ERK phosphorylation. Collectively, these properties make H$_2$O$_2$ an ideal mediator of signal transduction processes (36–38, 42, 58). Recent experimental
evidence in different animal models demonstrated that a rapid concentration gradient of \(\text{H}_2\text{O}_2\) is generated during injury and that \(\text{H}_2\text{O}_2\) is a powerful chemoattractant of leukocytes (53, 54). Moreover, lowering ROS levels by pharmacologic or genetic approaches reduces cell proliferation and impairs regeneration (59). We therefore have imaged \(\text{H}_2\text{O}_2\) in living neurons exposed to neurotoxins with novel specific fluorescent probes (39, 40) and found that the degenerating nerve terminals release \(\text{H}_2\text{O}_2\) of mitochondrial origin. This \(\text{H}_2\text{O}_2\) activates PSCs in vitro and in vivo. We also found that macrophages are recruited around the neurotoxin-treated NMJs. It is therefore likely that these macrophages are attracted by \(\text{H}_2\text{O}_2\), as well as by molecules released by activated PSCs, as previously found (60, 61). The prominent role of \(\text{H}_2\text{O}_2\) in neurotoxin-induced nerve degeneration and repair is proved by the impaired regeneration we observed in the presence of \(\text{H}_2\text{O}_2\) inhibitors.

In addition to \(\text{H}_2\text{O}_2\), we found that mtDNA and Cyt c can act as mediators of neuronal damage and activate SCs via ERK pathway. When added in a mixture with \(\text{H}_2\text{O}_2\), an additive effect on ERK phosphorylation is observed. As neuronal membrane integrity is preserved, the question arises of how mtDNA and Cyt c, coming from the mitochondrial matrix or the intermembrane space, respectively, can exit the cell. Several pieces of evidence indicate that mitochondria are central sensors for axonal degenerative stimuli (62), and the release of mtDNA fragments from PTP in isolated mitochondria has been documented (63). Here, the mitochondrial PTP was found to be involved in the exit of both mtDNA and Cyt c from mitochondria, with a significant reduction in the presence of the PTP desensitizing molecule cyclosporin A. Once in the cytosol, mtDNA and Cyt c could be released via the noncanonical or unconventional secretory route, including secreto-lysosomes, membrane blebbing, multivesicular body-derived exosomes, or autophagy (64). Here, we found that exosomes purified from intoxicated neuronal supernatants contain mtDNA, whereas Cyt c was not detected, possibly because of the insufficient sensitivity of Western blot. It is also possible that Cyt c is released directly via contact sites between mitochondria and the presynaptic membrane, similar to those observed by electron microscopy in a closely similar pathological condition caused by autoimmune anti-ganglioside antibodies (65).

The present work has identified three mitochondrial alarmins involved in PSCs activation after an acute nerve injury and proposes \(\text{H}_2\text{O}_2\) as the strongest inducer of PSCs response. Inactivation of \(\text{H}_2\text{O}_2\) by catalase reduces ERK phosphorylation in vivo and delays NMJ recovery in vivo after toxin-induced neuroaplasia and degeneration, supporting a crucial role of this molecule in the regeneration process.

Nerve damage triggers important morphologic and functional changes in PSCs aimed at promoting NMJ regeneration, confirming their endowed high plasticity and their crucial role in the clearance of nerve debris. Indeed, during nerve terminal degeneration, PSCs become CD68-positive, indicating an acquired phagocytic activity. Together with macrophages, but not neutrophils, activated PSCs were found here to remove nerve debris, thus permitting a functional nerve regeneration. This is at variance from what was found during axonal degeneration, where a pronounced neutrophil infiltration was detected (54).

The phagocytic features of PSCs described here represent an additional early read-out of PSCs activation at the injured NMJ. PSCs respond to axonal damage caused by neurotoxin poisoning by engulfing degenerating terminals, by extending long processes, and by activating intracellular signaling pathways crucial for regeneration. On the basis of these perspectives, we plan to study more in detail the intracellular signaling and transcriptomic events taking place inside activated PSCs. More in general, it appears that the present experimental approach can be extended to the investigation of other motor neuron diseases, including the non-cell-autonomous and dying-back axonopathy of ALS and autoimmune neuropathies including Guillain-Barré and Miller-Fisher syndromes (57–66). Such studies are likely to provide relevant insights for future therapeutic endeavors.

### Materials and Methods

**Animal Strains.** C57BL/6 mice expressing cytosolic GFP under the pp2 promoter were kindly provided by B. Macklin (Aurora) in collaboration of T. Misgeld (Munchen, Germany). All experiments were performed in accordance with the European Communities Council Directive n° 2010/63/UE and approved by the Italian Ministry of Health.

**Hydrogen Peroxide Detection.** Hydrogen peroxide generation in primary neurons was measured using Mitochondria Peroxy Yellow 1 (MitoyoY) (39) or Peroxyfluor 6-acetoxymethyl ester (PF6-AM) (40), synthesized in the C.J.C. laboratory (Berkeley, CA), specific probes of \(\text{H}_2\text{O}_2\) production in mitochondria and cytoplasm, respectively. Both probes were loaded at 5 \(\mu\)M for 30 min at 37 °C in Krebs–ringer buffer (KRB: Hepes 25 mM at pH 7.4, NaCl 124 mM, KCl 5 mM, MgSO\(_4\) 1.25 mM, CaCl\(_2\) 1.25 mM, KH\(_2\text{PO}_4\) 1.25 mM, glucose 8 mM). Images were acquired at different points after toxin exposure with a DM6000 inverted epifluorescence microscope (Leica) equipped with a 63× HXL PL APO oil immersion objective NA 1.4. Filter cubes (Chroma Technology) have an excitation range of 470/40 nm, a dichroic mirror 495LPXR, and an emission of 525/50 nm. Images were acquired with an Orca-Flash4 digital camera (Hamamatsu). Illumination was kept at a minimum to avoid ROS generation because of phototoxicity. To detect neuronal bulges, we took advantage of differential interference contrast microscopy. Fluorescence intensity quantification was carried out with ImageJ, and the statistical analysis with Prism (GraphPad).

**Cell Treatments.** CGNs (6 d in culture) plated onto 35-mm dishes (1.2 million cells per well) were exposed for 50–60 min to SPANs (6 nM) or to aLtx (0.1 nM) at 37 °C. In some experiments, neurons were preincubated for 30 min with cyclosporin A 5 \(\mu\)M before toxin addition. Supernatants or cell lysates were collected and then processed for real-time quantitative PCR (qPCR) or Western blot.

Primary SCs were exposed to different mitochondrial alarmins (\(\text{H}_2\text{O}_2\), 10–100 \(\mu\)M, Cyt c (R&D) 1 \(\mu\)g/mL, mtDNA 16 \(\mu\)g/mL) or to the toxins for different times and lysed in Lysis Buffer (Hepes 10 mM, NaCl 150 mM, SDS 1%, EDTA 4 mM, protease inhibitors mixture (Roche), and phosphatase inhibitor mixture).

Cocultures were treated with the toxins and then lysed after different periods; in a set of experiments, 1,000 U per well catalase was added 5 min before intoxication and kept throughout the experiment; in another set, cultures were incubated with the MEK1 inhibitor PD98059 (Cell Signaling; 4 mM, protease inhibitors mixture) 1 h before toxins addition. Samples were then probed for p-ERK.

**Immunofluorescence.** After treatments, isolated SCs or cocultures were fixed for 15 min in 4% (wt/vol) paraformaldehyde (PFA) in PBS, quenched (0.3% glycine, 0.24% NH\(_4\)Cl in PBS), and permeabilized with 0.3% Triton X-100 in PBS for 5 min at room temperature (RT). After saturation with 3% (vol/vol) BSA in PBS for 30 min at RT, SCs were incubated with the following primary antibodies: anti-Phospho-p44/42 MAPK (Cell Signaling), 1:1,000; anti-NF200 (Sigma), 1:200; anti-SNAP-25 (abcam), 1:500; anti-p47phox (AbD Serotec, 1:300); and anti-CD68 (mouse monoclonal, 1:200; Covance). Images were acquired at different points after toxin exposure with a DMI6000 B (Leica) equipped with a 100× oil immersion objective NA 1.4. Filter cubes (Chroma Technology) have an excitation range of 470/40 nm, a dichroic mirror 495LPXR, and an emission of 525/50 nm. Images were acquired with an Orca-Flash4 digital camera (Hamamatsu). Illumination was kept at a minimum to avoid ROS generation because of phototoxicity. To detect neuronal bulges, we took advantage of differential interference contrast microscopy. Fluorescence intensity quantification was carried out with ImageJ, and the statistical analysis with Prism (GraphPad).
primary antibody (anti-Phospho-p44/42 MAPK, 1:1,000; Cell Signaling) was carried out for 72 h and the tyramide signal amplification kit (Perkin-Elmer) was used (45).

To stain acidic compartments, LAL muscles collected after 4 h of intoxication were loaded ex vivo with LysoTracker Red DND-99 (1:5,000; Life Technologies). Samples were then fixed and processed for indirect immunohistochemistry, as described earlier. Images were collected with a Leica SP5 confocal microscope equipped with a 63× HCX PL APO NA 1.4×. Laser excitation line, power intensity, and emission range were chosen according to each fluorophore in different samples to minimize bleed-through.

Electrophysiological Recordings. Electrophysiological recordings were performed with glass microelectrodes filled with 3 M KCl and 3 M NaCl for 2 min. (68) while being continuously perfused with oxygenated Neurobasal A medium (Life Technologies). Samples were then fixed and processed for indirect immunohistochemistry, as described earlier. Images were collected with a Leica SP5 confocal microscope equipped with a 63× HCX PL APO NA 1.4×. Laser excitation line, power intensity, and emission range were chosen according to each fluorophore in different samples to minimize bleed-through.

Evoked neurotransmitter release was measured in current-clamp mode, and resting membrane potential was adjusted with current injection to −70 mV. EJPs were elicited by supramaximal nerve stimulation at 0.5 Hz, using a suction microelectrode connected to a 588 stimulator (Grass). To prevent muscle contraction after dissection, samples were incubated for 10 min with 1 μM Conotoxin GilB (Alomone).

Signals were amplified with intracellular bridge mode amplifier (BA-01X, NPI), sampled using a digital interface (NI PCI-6221, National Instruments) and recorded by means of electrophysiological software (WinEDR, Strathclyde University). EJPs measurements were carried out with Clampfit software (Molecular Devices).

Statistical Analysis. The sample size (N) of each experimental group is described in each corresponding figure legend, and at least three biological replicates were performed. Prism (GraphPad Software) was used for all statistical analyses. Quantitative data displayed as histograms are expressed as means ± SEM (represented as error bars). Results from each group were averaged and used to calculate descriptive statistics. Significance was calculated by Student’s t test (unpaired, two-side). P values less than 0.05 were considered significant.

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