### UCSF UC San Francisco Previously Published Works

### Title

The Crossroads of Iron with Hypoxia and Cellular Metabolism. Implications in the Pathobiology of Pulmonary Hypertension

**Permalink** https://escholarship.org/uc/item/0tv716jz

**Journal** American Journal of Respiratory Cell and Molecular Biology, 51(6)

**ISSN** 1044-1549

### Authors

Robinson, Jeffrey C Graham, Brian B Rouault, Tracey C <u>et al.</u>

**Publication Date** 

2014-12-01

### DOI

10.1165/rcmb.2014-0021tr

Peer reviewed

### The Crossroads of Iron with Hypoxia and Cellular Metabolism Implications in the Pathobiology of Pulmonary Hypertension

Jeffrey C. Robinson<sup>1</sup>, Brian B. Graham<sup>1</sup>, Tracey C. Rouault<sup>2</sup>, and Rubin M. Tuder<sup>1</sup>

<sup>1</sup>Program in Translational Lung Research, Division of Pulmonary Sciences and Critical Care Medicine, Department of Medicine, University of Colorado School of Medicine, Aurora, Colorado; and <sup>2</sup>Molecular Medicine Program, Eunice Kennedy Shriver National Institute of Child Health and Human Development, Bethesda, Maryland

### Abstract

The pathologic hallmark of pulmonary arterial hypertension (PAH) is pulmonary vascular remodeling, characterized by endothelial cell proliferation, smooth muscle hypertrophy, and perivascular inflammation, ultimately contributing to increased pulmonary arterial pressures. Several recent studies have observed that iron deficiency in patients with various forms of PAH is associated with worsened clinical outcome. Iron plays a key role in many cellular processes regulating the response to hypoxia, oxidative stress, cellular proliferation, and cell metabolism. Given the potential importance of iron supplementation in patients with the disease and the broad cellular functions of iron, we review its role in processes that pertain to PAH.

### **Clinical Relevance**

This manuscript details the complexities of how disordered iron metabolism, with iron deficiency being one of the most common nutritional deficits in the world, may act to alter the pathobiology of pulmonary vascular disease and other hypoxia-related illnesses.

Pulmonary arterial hypertension (PAH), strictly defined by a pulmonary arterial pressure  $\geq 25$  mm Hg on right heart catheterization (1), is a heterogeneous disorder. A host of etiologies, including heritable, idiopathic (i.e., idiopathic PAH [IPAH]), infectious (HIV, schistosomiasis), connective tissue disease, and hypoxic conditions (pulmonary obstructive diseases, interstitial lung diseases) have been identified as causing the characteristic pathologic vascular changes (2). These contribute to a progressive increase in pulmonary vascular resistance due to precapillary pulmonary microangiopathy (3) and eventual right ventricular failure (4). Although primary PAH is classically associated with a grave prognosis (5, 6), modern therapies have resulted in improved exercise performance, quality of life, and, in some instances, reduced mortality (7).

However, despite with the use of these new pharmacologic agents, the pulmonary vascular lesions and considerable morbidity and mortality persist (8).

Recently, it has been clinically observed that iron deficiency has an increased prevalence in a population of patients with IPAH as measured by reduced serum iron, elevated circulating transferrin levels, and decreased transferrin saturation. It has been found that iron deficiency is associated with increased morbidity and mortality in IPAH and scleroderma-related PAH (9-12). Furthermore, it has been suggested that iron infusions may attenuate hypoxic vasoconstriction (13, 14), and trials are underway to evaluate the effect of intravenous iron infusions in patients with primary PAH (15). These initiatives are being performed despite a paucity of mechanistic evidence concerning the

pathobiologic relationship between pulmonary hypertension (PH) and iron deficiency. Many of the core pathobiologic features of PH, such as plexogenic arteriopathy, smooth muscle vasoconstriction, and inflammation (16), involve pathways that may depend on iron, and it is clear that iron homeostasis is entwined with cellular responses to hypoxia, cellular proliferation, and mitochondrial function. In contrast to iron deficiency, several hemoglobinopathies have been associated with increased prevalence of PH, which has been identified as a prognostic indicator in these disorders, highlighting the complex and multifactorial nature by which iron influences pulmonary vascular disease. The increased understanding of the role of iron in modifying these key disease pathways may lead to novel insights into the pathobiology of PH. This effort may provide

(Received in original form January 17, 2014; accepted in final form June 27, 2014)

Correspondence and requests for reprints should be addressed to Jeffrey C. Robinson, M.D., Program in Translational Lung Research, Division of Pulmonary Sciences and Critical Care Medicine, Department of Medicine, University of Colorado School of Medicine, Aurora, CO 80045. E-mail: jeffrey.robinson@ucdenver.edu

Am J Respir Cell Mol Biol Vol 51, Iss 6, pp 721-729, Dec 2014

Copyright © 2014 by the American Thoracic Society Originally Published in Press as DOI: 10.1165/rcmb.2014-0021TR on July 2, 2014 Internet address: www.atsjournals.org new therapeutic insights because current therapies are focused on pulmonary vasodilation rather than on reversing pulmonary vascular or right ventricular pathologic remodeling.

Here, we review iron metabolism and homeostasis and then focus on several pathobiologic links to PH, notably, how dysregulated iron metabolism may alter the hypoxic response of the pulmonary vasculature, oxidative stress, and cellular metabolism.

### PH and Iron Deficiency: Prevalence and Prognosis

It has been demonstrated that iron deficiency is highly prevalent in IPAH and hereditary PAH, with rates ranging from 30.1 to 63%, which is much higher than the general population (10). Another recent study reported that 41% of patients with systemic sclerosis-related PH are iron deficient and that patients with systemic sclerosis without PH demonstrated only a 16% prevalence of iron deficiency (11). In these observational studies, patients did not demonstrate overt anemia compared with control patients. When patients with chronic thromboembolic disease were analyzed, the rates of iron deficiency were only 4.9%, suggesting that iron deficiency is unique to group 1 PAH (9).

Retrospective analysis has demonstrated that overt anemia correlates with increased risk of death in PH. In one study, after adjustment for known predictors of death and PH etiology, anemic patients were 3.3 times more likely to die than nonanemic patients (17). In a separate observational study, it was found that of 70 patients with IPAH, 43% were iron deficient and had significantly lower 6-minute walk distance (6MWD) than iron-replete patients, but hemodynamic indices were not significantly different. Furthermore, when the irondeficient individuals were split into anemic and nonanemic groups, there was no significant difference in 6MWD (12). This suggests that poor functional status and outcomes may be related more to defects triggered by low systemic iron than to simple anemia. Supporting this is the fact that red blood cell distribution width, which is elevated in iron deficiency, has greater utility than established biomarkers-including N-terminal fragment of the prohormone B-type natriuretic peptide, growth

differentiation factor-15, IL-6, and serum creatinine-in predicting mortality among patients with PH (18, 19). Offering further evidence that iron status modulates the disease process of PH, zinc-protoporphyrin, a highly sensitive marker of iron metabolic abnormalities and absolute deficiency, has been demonstrated to be significantly increased in individuals with IPAH and in sleep apnea-associated PH groups relative to control subjects, patients with asthma, and those with secondary PAH. Moreover, elevations of zinc-protoporphyrin strongly correlated with disease severity as determined by echocardiographic pulmonary arterial systolic pressures (PASP) and 6MWD measurements (20).

These observations highlight the preponderance of iron deficiency anemia in several forms of group 1 PAH, the mechanisms of which remain unclear. There are multiple studies demonstrating that the presence of iron deficiency correlates with worsened functional status and mortality. These data suggest an important link between iron deficiency and the pathobiology of PH and highlight the need for retrospective and mechanistic studies of this relationship, which may lead to novel insights and therapeutic strategies.

# Effects of Iron Supplementation on PH

It has been observed that iron replacement may be hampered in patients with PH. In one study, patients with IPAH and iron deficiency were given oral iron preparations. At the end of 4 weeks, only 11% of the patients who had completed the intervention demonstrated significant increases in their serum iron or ferritin levels, suggesting underlying absorptive or release dysfunction in PH (12). Nevertheless, ironor chelation in converse (13)-appears to modulate the degree of PH. As outlined below, iron participates in the control of hypoxic transcriptional controls, including stabilization of hypoxia-inducible factors, thereby affecting pulmonary vascular responses.

Alveolar hypoxia triggers demonstrable elevation in pulmonary pressures that is attributable to hypoxic vasoconstriction and pulmonary vascular remodeling. The role of iron supplementation in hypoxic vasoconstriction has been studied in a model of acute mountain sickness (14). Briefly, acute hypoxic PH was induced in human subjects by moving them from sea level (estimated O<sub>2</sub> partial pressures: atmospheric, 21%; lung, 15%) to an altitude of 4,340 m (estimated O<sub>2</sub> partial pressures: atmospheric, 13%; lung, 7%), with a predictable and significant increase in their PASP as measured by echocardiography. After 3 days, subjects were randomized to receive intravenous infusions of iron sucrose or placebo. There was a significant (and rapid) decrease in PASP in the iron-supplemented group. In a separate trial, individuals who chronically lived at an altitude of greater than 2,500 m above sea level (O<sub>2</sub> partial pressures: atmospheric, 16%; lung, 10%) were identified as having chronic mountain sickness (defined by excessive erythrocytosis, hypoxemia, and the absence of chronic pulmonary disease), with baseline echocardiography demonstrating elevated PASP. Patients were then randomized to staged venesection with subsequent intravenous iron or placebo infusion. After 24 days, each group underwent crossover to receive the alternative treatment. With initial venesection, progressive development of iron deficiency correlated with worsening of pulmonary arterial pressures determined by echocardiography. This study suggests that there is a causal relationship between iron deficiency and acute hypoxic PH.

Despite the potential protective role of infused iron in pulmonary vasoconstriction in the acute setting, several clinical and pathologic characteristics underscore the differences between hypoxic and class I PAH, with hypoxia-related PH demonstrating (generally) less severe elevations in right ventricular systolic pressure, a lack of response to PAH-specific therapies, and vascular lesions characterized by medial hypertrophy of small muscular arterials and more distal arteriolar neomuscularization rather than the plexiform lesions that are characteristic in PAH (21, 22). However, recent mechanistic studies have revealed elements of hypoxia-like signaling in PAH (23, 24), warranting investigation into the effects of iron in hypoxic and nonhypoxic signaling and hypertensive pulmonary vascular remodeling.

#### Mechanisms of Iron Homeostasis and Links to PH

Iron is involved in many integral cellular functions in aerobic organisms, including

growth, differentiation, and metabolism. Approximately 60 to 70% of iron is used by red blood cell hemoglobin, 10% is localized in myocytes in the form of myoglobin, and the remainder is an indispensable constituent of over 1,000 different irondependent proteins in eukaryotic organisms (25, 26). Broadly, iron is intimately involved in oxygen transport, electron transport, and DNA synthesis, exemplified by its function as an essential cofactor of ribonucleotide reductase, whereby ribonucleotides are catalytically converted to deoxyribonucleotides (27). It is essential for functioning cytochromes, lipoxygenases, fatty acid desaturases, superoxide dismutase, mitochondrial NADH dehydrogenase (complex 1), and a number of other enzymes. Conversely, when in excess, free iron catalyzes the formation of reactive species through the Fenton reaction, damaging macrostructures within the cell (28, 29). Given its fundamental and dual nature in health and disease, a rich homeostatic system has evolved to regulate iron transport and metabolism.

The average adult male body contains roughly 4 g of iron, which is efficiently recycled within the body, with < 2 mg dailyreplenishment required via dietary intake. No physiologic control of iron excretion exists, leaving regulation of absorption from dietary sources the primary modulator of total body iron. Dietary iron is primarily in the form of insoluble oxidized Fe<sup>3+</sup> (ferric) and must be reduced to the  $Fe^{2+}$  (ferrous) form by the apical duodenal cytochrome B before being shuttled across the intestinal epithelium by the divalent metal transporter 1 (DMT1) (30). DMT1 is an evolutionarily highly conserved protein of the natural resistance-associated macrophage protein class, consisting of transmembrane transporters that are broadly involved in divalent cation uptake (31). Once through the apical duodenal epithelium, iron is exported into the circulation by ferroportin, which works with the ferroxidase hephaestin to generate ferric iron that binds to serum transferrin, an abundant plasma glycoprotein with high affinity for ferric iron that binds iron, attenuating its reactivity and transporting it to cells. After localization at the cell wall, the transferrin/ iron complex binds to transferrin receptors localized on the cell surface, with subsequent endocytosis via clathrin-coated pits. Within the endosome, acidification

via proton pumps on the endosomal surface leads to protein conformational changes that result in the release of iron from transferrin and transport to the cytosol by DMT1 (32). At this point, iron functions within a wide array of cellular functions as part of the labile iron pool (33).

#### Hepcidin Regulation of Iron Absorption

Maintenance of systemic iron homeostasis is partially regulated by hepcidin, a defensin family member that is induced in the setting of dietary and parenteral iron loading largely in liver cells. Its central role in iron physiology is highlighted by inherited disorders of hepcidin deficiency, which result in severe forms of hemochromatosis (34). Mechanistically, hepcidin decreases absorption of iron by binding ferroportin in hepatocytes, enterocytes, and macrophages, triggering its ubiquitination and subsequent lysosomal degradation (35). Hepcidin expression is markedly up-regulated by iron overload and IL-6 driven signaling, whereas hypoxia and iron depletion lead to a decline in hepcidin levels (36, 37).

The precise mechanisms that regulate hepcidin expression remain unclear, but it is apparent that IL-6 and bone morphogenic protein receptor (BMPR) signaling are critical. IL-6, a proinflammatory cytokine that is up-regulated in PAH and is associated with higher mortality (38, 39), is sufficient to induce hepcidin expression and leads to hypoferremia in animal models of inflammation (40). However, in a study of patients with IPAH, although IL-6 levels were increased, this failed to correlate with increased hepcidin activity, arguing that IL-6 is unlikely to be the sole cause of the effect (9). BMP receptors are ubiquitously expressed receptors in the TGF- $\beta$  family, which bind in a heteromeric fashion and lead to phosphorylation (thus activation) and nuclear translocation of receptor-activated SMADs, which signal through a multitude of downstream pathways, including pSmad, p38, pERK, JNK, and Akt/PI3K. Heterozygous germline mutations in BMPR type 2 (BMPR2) have been described in both sporadic IPAH and hereditary PAH (41). Interestingly, when BMPR2 is disrupted by *loxP* knockout or siRNA knockdown in pulmonary arterial smooth muscle cells, there is paradoxical increased activation of SMAD and p38 by BMP6, whereas BMP2 and BMP4 resulted

in reduced signaling (42). BMP6 has been demonstrated to be a major positive regulator of hepcidin expression, with BMP6 knockout mice developing severe iron overload in the setting of reduced hepcidin levels (43). It is unclear whether patients with hereditary PAH with heterozygous mutations in BMPR2 have altered BMP control of hepcidin in the liver. However, some patients with PAH have increased levels of circulating hepcidin (10), and although it remains unclear if the mechanisms above are causal, increased hepcidin may explain the difficulty in supplementing these individuals with oral iron preparations (12).

# Iron Regulatory Proteins and Iron Responsive Elements

The iron regulatory proteins (IRPs) tightly regulate the flow of iron by modulating expression of target proteins posttranscriptionally by binding to cis-regulatory iron responsive elements (IREs), which are found in the untranslated regions (UTRs) of mRNAencoding proteins involved in iron metabolism (Figure 1). In iron-replete conditions, an iron-sulfur cluster (ISC) is bound to IRP-1, which relegates it to the cytoplasm to function as an aconitase. In iron-deplete conditions, without an ISC occupying its IRE binding site, IRPs are free to bind to transcripts that contain IREs (44). In contrast, IRP-2 does not contain an ISC but is regulated through F-box and leucine-rich repeat protein 5-regulated ubiquitination and proteasomal degradation (45, 46). Although IRP-1 and IRP-2 differ in their molecular nature and regulation of expression, both bind to IREs located in the 3' UTR of transcripts, such as transferrin receptor and likely to an alternative splice form of DMT1, resulting in transcript stabilization and facilitation of translation, leading to in increased iron uptake. In contrast, when the IRP binds to transcripts containing a 5' UTR IRE, translation is blocked, as with transcripts of H- and L-ferritin, ferroportin, and hypoxiainducible factor  $2\alpha$  (HIF $2\alpha$ ) (33, 44, 47). From a teleological perspective, IRP repression of HIF2 $\alpha$  via a phylogenetically conserved 5' UTR IRE (48, 49) allows for adjustment of erythrocyte production based on iron availability because HIF2a controls erythropoietin production in the kidney.

The IRP/IRE regulatory system is physiologically essential, as indicated by



**Figure 1.** Iron-deplete conditions translationally repress hypoxia-inducible factor  $2\alpha$  (HIF2 $\alpha$ ) via iron regulatory protein (IRP) and iron responsive element (IRE) interaction. IRP1 functions as a cytosolic aconitase in iron-replete conditions when the iron sulfur cluster (ISC) is bound to the catalytic apoprotein. In iron-deplete conditions, ISCs are unavailable, which enables IRP1 to bind to either the 5' or 3' IREs. In the 5' position, this results in translational repression, as is the case with HIF2 $\alpha$ . In contrast, when IRP1 binds to a 3' IREs (which are located on the transcripts of transferrin receptor 1 [TfR1] and divalent metal transporter 1 [DMT1]), the transcript is stabilized, resulting in increased iron uptake.

early embryonic lethality in transgenic mice that have both IRP-1 and IRP-2 knocked out. However, mice in which IRP-1 alone was knocked out did not have an immediate apparent phenotype, suggesting that IRP-1 and IRP-2 have redundant physiologic functions (i.e., IRP-2 rescues the deficiency of IRP-1) (50). However, detailed inspection found that mice with targeted deletion of IRP-1 developed polycythemia (51, 52) and spontaneous PH (53). This response was found to be due to translational derepression of HIF2 $\alpha$ , with a subsequent increase in erythropoietin and endothelin-1 (a potent pulmonary vascular bed constrictor involved in the development of PH [54]). The PH persisted from 3 months to 1 year but without apparent hemodynamic progression. There was also the absence of muscular hypertrophy, suggesting that derepression of HIF2a may induce PH primarily via increased pulmonary vascular vasoconstriction. A known target of HIF, endothelin-1 could mediate vasoconstriction and remodeling.

IRPs act as a master transcriptional regulator modulated by iron. In addition to their well-documented role in the maintenance of iron homeostasis, IRPs are increasingly recognized to play a role in the hypoxic response. The finding that homozygous deletion of IRP-1 in mice results in spontaneous PH has led further credence to the possibility that this homeostatic axis is pertinent to the pathobiology of pulmonary vascular disease, although the exact mechanism by which IRP-1 contributes to the disease remains to be defined. Dysregulated iron metabolism, mediated by defects in the hepcidin and/or IRP regulatory axis, alters cellular availability of iron and has multiple potential effects on the pathobiology of PH.

### Cellular Destinations and Functions of Iron in Relation to PH

With the vast number of enzymatically driven cellular functions that rely on iron as a cofactor, numerous iron-dependent, cellular-specific, and organ-specific processes may have a pathobiologic role in pulmonary vascular disease. Namely, there is potential interplay between iron with hypoxia and hypoxia signaling within the pulmonary vasculature, the function of iron in congestive heart failure, and iron's role in carcinogenesis.

## Iron and the Hypoxic Response of the Pulmonary Vasculature

As recently reviewed, stabilization of HIF1 $\alpha$ in hypoxic murine models results in pulmonary arterial smooth muscle cell (PASMC) proliferation, migration, and hypertrophy, ultimately contributing to the pathogenesis of PH (55). Highlighting the importance of HIFs in the development of PH, their role is not limited to World Health Organization group 3 (hypoxic) disease; increased expression of HIFs was observed in tissues of patients with non-hypoxia-related PAH (23, 56). Although similar phenotypic attenuation of PH is seen in HIF1 $\alpha(+/-)$  and HIF2 $\alpha(+/-)$  mice, it remains to be shown if this occurs through similar mechanisms in PASMCs (57, 58).

Targeted knock-out of IRP-1 resulted in PH via derepression of HIF2 $\alpha$ . This suggests that, in iron-deficient conditions, there may be increased IRP-1 binding of IREs (due to decreased ISC binding to the cytosolic aconitase), leading to suppressed HIF2 $\alpha$  translation and protection from the maladaptive PASMC remodeling. On the other hand, iron deficiency may worsen PH via HIF-dependent mechanisms. The ubiquitin ligase von Hippel-Lindau protein (VHL) mediates normoxic degradation of HIF1 $\alpha$  and HIF2 $\alpha$  to very low levels after prolyl hydroxylation (mediated by prolyl hydroxylase [PHD]) and subsequent proteasome-mediated degradation. PHDs require not only oxygen but also 2-oxoglutarate ( $\alpha$ -ketoglutarate), ascorbate, and iron (Figure 2) as essential cofactors (59, 60). It has been observed that iron chelation acutely elevates pulmonary vascular resistance in normal individuals subjected to acute hypoxia, which has been attributed to increased HIF stabilization (61).

Chuvash polycythemia (CP) is a rare autosomal recessive disorder in which individuals carry a dysfunctional VHL with the missense mutation R200W, resulting in increased HIF1 $\alpha$  and HIF2 $\alpha$  stabilization and increased expression of downstream target genes, including erythropoietin, glucose transporter member 1, transferrin, transferrin receptor, and vascular endothelial growth factor (62). Human physiologic studies have demonstrated that patients with CP have elevated baseline



**Figure 2.** Iron-deplete conditions stabilize HIF via a prolyl hydroxylase (PHD)-dependent mechanism. Iron is an essential cofactor for PHD, which allows for rapid ubiquitination and degradation by Von-Hippel Lindau (VHL) protein E3 ubiquitin ligase. HRE, hormone response element.

PASP when measured by echocardiography without evidence of clinical PH. When exposed to hypoxic conditions, patients with CP exhibit extraordinary pulmonary vascular sensitivity relative to control subjects (63). It is conceivable that this is due to unchecked HIF-mediated pulmonary vasoconstriction. Furthermore, many patients with CP require phlebotomy, which results in iron deficiency (as reflected by low serum ferritin); this iron deficiency, even when corrected for reduced blood volume (as represented by left ventricular internal diastolic diameter, hemoglobin concentration, and left atrial diameter), was associated with significantly increased tricuspid regurgitation velocity, a surrogate for the severity of pulmonary vascular pressures (64).

In a murine model of CP, mice carrying the R200W mutation in the VHL gene in a homozygous fashion develop PH independent of polycythemia. HIF2 $\alpha$ , but not HIF1 $\alpha$ , was the driver of PH and polycythemia in the animals with CP (65). The hypertensive process is accompanied by increased perivascular inflammation and vascular remodeling. The interesting parallels and differences of the pulmonary hypertensive phenotype between IRP-1 null (as described above) and VHL R200W mutant suggest that these proximal genetic events affect the pulmonary vascular phenotype, possibly by affecting downstream effector genes. In addition to the shared up-regulation of HIF2 $\alpha$  in both models, IRP-1 and VHL differentially targeted transcripts and proteins, respectively, may account for the specific findings in both models.

An interesting correlate is found in humans who carry HIF2a gain-offunction mutations that induce a more vasoconstrictive phenotype of PH along with an exaggerated vasoconstrictive response to hypoxia (66, 67). Two separate genome studies have demonstrated a significant increase in SNPs in EPAS1the gene coding for HIF2 $\alpha$ —in ethnic Tibetans living at elevations exceeding 4,000 m (68, 69). There is an associated decrease in hemoglobin concentration, ameliorating one of the key findings of chronic mountain sickness. It is posited that, either through this or through one of the many other transcriptional effects of HIF2 $\alpha$ , a naturally selected survival advantage is offered by decreased HIF2a function at altitude (69).

The experimental data highlighted here demonstrate that HIF plays a role in the pathogenesis of hypoxic PH and may play a role in primary PAH. Low iron could therefore act to stabilize HIF via inhibition of 2-oxoglutarate-dependent PHDs. However, low iron, via stabilization of IRP-1, can selectively suppress HIF2 $\alpha$ translation, leaving HIF1 $\alpha$  to exert its prohypertensive effects. The mechanisms by which HIF levels are elevated in PAH remain unclear, but the role of iron, either through IRP-regulated repression or VHL/PHD-mediated stabilization, represents an enticing therapeutic target once mechanisms can be delineated.

#### Iron and Oxidative Stress

Iron's essential role in cellular biology is largely dependent on its ability to undergo cyclic oxidation and reduction. The most basic form of the Fenton reaction involves an electron transfer from hydrogen peroxide to ferrous iron, resulting in the creation of potentially damaging hydroxyl radicals (70). Organisms have highly conserved mechanisms of preventing this reaction in a harmful context, primarily through the binding of ferric iron to ferritin and transferrin receptor, allowing for safe storage and distribution of iron. However, although this reaction is of concern in the setting of elevated iron levels, there is extensive evidence demonstrating the role of iron in antioxidant contexts.

Erythroid cells require the ironcontaining prosthetic group heme for  $\alpha$ - and  $\beta$ -globin chain synthesis during reticulocyte maturation. Although free heme has cytotoxic effects, it can also act as a signaling molecule, triggering the canonical cytoprotective hemeoxygenase-1 (HO-1) transcriptional program. Heme interacts directly with Bach1, a transcriptional repressor, to deactivate it and allow for transcription of a multitude of genes in the metabolism of heme, including HO-1 (71). An additional mechanism includes heme-mediated stabilization of the transcription factor NF-E2-related factor 2 (Nrf2), a potent regulator of antioxidant proteins for which there is experimental evidence suggesting a protective effect in pulmonary vascular disease (72). This tightly regulated program likely serves as a protective mechanism in periods of high oxidant stress. Oxidative stress is ubiquitous and is increased in many disease states. Iron is a central mediator of this, as illustrated by the hemoglobinopathies.

#### Hemoglobinopathies and PH

It has been suggested that hemoglobinopathies may be one of the most common causes of PH worldwide owing to the high prevalence of this heterogeneous group of diseases (73). Nearly every form of hemolytic anemia, including sickle cell disease, thalassemia, hereditary spherocytosis, paroxysmal nocturnal hemoglobinuria, and microangiopathic hemolytic anemias, has been associated with PH (74). Homozygous sickle cell anemia is frequently associated with PH, with several studies demonstrating prevalence by echocardiographic criteria at 30% (75).

Although the pathobiology of these hemolytic disorders is heterogeneous, there may be a shared pathophysiology that gives rise to PH. Several experimental and clinical studies have demonstrated that chronic hemolysis results in nitric oxide depletion, increased endothelin-1, and augmented platelet activation, all of which have pathogenic links with PH (76). Iron loading occurs in B-thalassemia major primarily from blood transfusions; however, in non-transfusion-dependent thalassemia, iron overload is still a frequent occurrence, thought to result from ineffective erythropoiesis and hypoxia, culminating in suppression of hepcidin activity, which leads to increased iron uptake (77). In addition to nitric oxide depletion, there may be increased oxidant formation and myocardial iron deposition contributing to associated cardiovascular disease (74, 78). Emphasizing the biologic importance of iron overload, human studies have demonstrated that chelation therapy in thalassemia intermedia results in a significant reduction in pulmonary arterial pressures by echocardiogram (79).

Given the significant global health burden of hemoglobinopathy-associated PH and the lack of effective therapies (75), further research is necessary to delineate the mechanisms by which these disorders lead to pulmonary vascular disease, which could inform unique iron-targeted therapeutics.

## Iron, Metabolic Dysregulation, and Mitochondrial Dysfunction

Many pathologic and molecular features of PH, including apoptotic resistance, increased cellular proliferation, and a preferential aerobic glycolysis, are analogous with characteristics seen in cancers (80, 81). These are supported by complementary human and experimental studies. Further contributing to the quasimalignancy paradigm of PH is evidence of clonal endothelial cell expansion in patients with IPAH (82), somatic chromosomal abnormalities in IPAH lungs (83), increased uptake of 18-fluorodeoxy-glucose in the lung fields of patients with IPAH (84), and the presence of DNA microsatellite instability within the IPAH vascular proliferative lesions (85). The metabolic switch toward aerobic glycolysis and mitochondrial dysfunction in PH is increasingly recognized as a key pathophysiologic feature shared with cancers.

Experimentally, rodents with PH exhibit aerobic glycolysis- and HIF-1  $\alpha$ -dependent vascular cell growth and PH (86); this is paralleled by increased uptake of 18-fluorodeoxy-glucose uptake, as observed in humans with the disease (56). HIF activity may be intimately tied to this pathologic shift because hormone response element promoter transcription results in target gene expression, resulting in a shift from oxidative phosphorylation to glycolysis (87). Highlighting the wide reach of HIF-regulated cellular control, microRNA-210 has been identified as a transcriptional target of HIF (88) and has been implicated in suppression of the ISC assembly proteins, effectively downregulating mitochondrial respiratory complexes via decreasing ISC biogenesis (89). As detailed above, deregulated iron metabolism has the ability to either upregulate or attenuate HIF, providing a potential pathobiologic link in PH.

Mitochondria are the synthetic site of ISCs and heme, which function as prosthetic groups that are essential for numerous cellular functions. Heme in addition to its critical role in erythroid cells, has antioxidant effects. ISCs are integrated into proteins within the electron transport chain and into enzymes within the Krebs cycle, such as aconitase and succinate dehvdrogenase (90). Although it remains unclear whether altered iron availability contributes to the pathobiology of PH through disruption of this metabolic machinery, there is evidence that mutations in NFU1-a protein that participates in iron sulfur complex assembly-results in a fatal mitochondrial disease, with PH being a feature in 70% of the individuals identified (91).

Given the fundamental role of iron in metabolism and mitochondrial function, there are several potential mechanisms by which altered iron availability may contribute to the pathobiology of PH. Future work will need to determine the importance of ISC availability on mitochondrial function and pulmonary vascular cell metabolism as well as the role of HIF in this relationship.

### Conclusion

Iron deficiency has a striking prevalence and association with increased morbidity and mortality in various forms of PH. It is





a compelling therapeutic target given the relative ease with which it can be manipulated via supplementation and chelators. However, iron has such critical and wide-ranging cellular roles (including HIF regulation, cellular metabolism, and oxidative stress) that specific mechanisms must first be delineated given the potential pathogenic role of either increased or decreased iron availability (summarized in Figure 3). It remains to be shown whether systemic iron deficiency results in tissue- or cellspecific iron depletion within the pulmonary vasculature. Parsing the mechanisms of iron on cellular function is of critical importance to understanding the role of iron in the pathobiology of PH. Furthermore, with iron deficiency affecting an estimated 2 billion people worldwide (92), a deeper understanding of how iron availability alters such fundamental cellular processes has broad implications for human disease.

<u>Author disclosures</u> are available with the text of this article at www.atsjournals.org.

#### References

- Galiè N, Hoeper MM, Humbert M, Torbicki A, Vachiery JL, Barbera JA, Beghetti M, Corris P, Gaine S, Gibbs JS, et al.; ESC Committee for Practice Guidelines (CPG). Guidelines for the diagnosis and treatment of pulmonary hypertension: the Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS), endorsed by the International Society of Heart and Lung Transplantation (ISHLT). *Eur Heart J* 2009;30: 2493–2537.
- Simonneau G, Gatzoulis MA, Adatia I, Celermajer D, Denton C, Ghofrani A, Gomez Sanchez MA, Krishna Kumar R, Landzberg M, Machado RF, *et al.* Updated clinical classification of pulmonary hypertension. *J Am Coll Cardiol* 2013;62(25 Suppl):D34–D41.
- Tuder RM. Pathology of pulmonary arterial hypertension. Semin Respir Crit Care Med 2009;30:376–385.
- Voelkel NF, Gomez-Arroyo J, Abbate A, Bogaard HJ, Nicolls MR. Pathobiology of pulmonary arterial hypertension and right ventricular failure. *Eur Respir J* 2012;40:1555–1565.
- Rich S, Dantzker DR, Ayres SM, Bergofsky EH, Brundage BH, Detre KM, Fishman AP, Goldring RM, Groves BM, Koerner SK, *et al*. Primary pulmonary hypertension: a national prospective study. *Ann Intern Med* 1987;107:216–223.
- D'Alonzo GE, Barst RJ, Ayres SM, Bergofsky EH, Brundage BH, Detre KM, Fishman AP, Goldring RM, Groves BM, Kernis JT, et al. Survival in patients with primary pulmonary hypertension: results from a national prospective registry. Ann Intern Med 1991;115:343–349.
- 7. Humbert M, Sitbon O, Simonneau G. Treatment of pulmonary arterial hypertension. *N Engl J Med* 2004;351:1425–1436.
- Stacher E, Graham BB, Hunt JM, Gandjeva A, Groshong SD, McLaughlin VV, Jessup M, Grizzle WE, Aldred MA, Cool CD, et al. Modern age pathology of pulmonary arterial hypertension. Am J Respir Crit Care Med 2012;186:261–272.
- Soon E, Treacy CM, Toshner MR, MacKenzie-Ross R, Manglam V, Busbridge M, Sinclair-McGarvie M, Arnold J, Sheares KK, Morrell NW, *et al.* Unexplained iron deficiency in idiopathic and heritable pulmonary arterial hypertension. *Thorax* 2011;66:326–332.
- Rhodes CJ, Howard LS, Busbridge M, Ashby D, Kondili E, Gibbs JSR, Wharton J, Wilkins MR. Iron deficiency and raised hepcidin in idiopathic pulmonary arterial hypertension: clinical prevalence, outcomes, and mechanistic insights. *J Am Coll Cardiol* 2011;58:300–309.
- Ruiter G, Lanser IJ, de Man FS, van der Laarse WJ, Wharton J, Wilkins MR, Howard LS, Vonk-Noordegraaf A, Voskuyl AE. Iron deficiency in systemic sclerosis patients with and without pulmonary hypertension. *Rheumatology (Oxford)* 2014;53:285–292.
- Ruiter G, Lankhorst S, Boonstra A, Postmus PE, Zweegman S, Westerhof N, van der Laarse WJ, Vonk-Noordegraaf A. Iron deficiency is common in idiopathic pulmonary arterial hypertension. *Eur Respir J* 2011;37:1386–1391.
- Smith TG, Balanos GM, Croft QPP, Talbot NP, Dorrington KL, Ratcliffe PJ, Robbins PA. The increase in pulmonary arterial pressure caused by hypoxia depends on iron status. *J Physiol* 2008;586:5999–6005.
- Smith TG, Talbot NP, Privat C, Rivera-Ch M, Nickol AH, Ratcliffe PJ, Dorrington KL, León-Velarde F, Robbins PA. Effects of iron supplementation and depletion on hypoxic pulmonary hypertension: two randomized controlled trials. *JAMA* 2009;302:1444–1450.

- Howard LSGE, Watson GMJ, Wharton J, Rhodes CJ, Chan K, Khengar R, Robbins PA, Kiely DG, Condliffe R, Elliott CA, *et al.* Supplementation of iron in pulmonary hypertension: rationale and design of a phase II clinical trial in idiopathic pulmonary arterial hypertension. *Pulm Circ* 2013;3:100–107.
- Tuder RM, Groves B, Badesch DB, Voelkel NF. Exuberant endothelial cell growth and elements of inflammation are present in plexiform lesions of pulmonary hypertension. *Am J Pathol* 1994;144:275–285.
- Krasuski RA, Hart SA, Smith B, Wang A, Harrison JK, Bashore TM. Association of anemia and long-term survival in patients with pulmonary hypertension. *Int J Cardiol* 2011;150:291–295.
- Rhodes CJ, Wharton J, Howard LS, Gibbs JSR, Wilkins MR. Red cell distribution width outperforms other potential circulating biomarkers in predicting survival in idiopathic pulmonary arterial hypertension. *Heart* 2011;97:1054–1060.
- Hampole CV, Mehrotra AK, Thenappan T, Gomberg-Maitland M, Shah SJ. Usefulness of red cell distribution width as a prognostic marker in pulmonary hypertension. *Am J Cardiol* 2009;104:868–872.
- Decker I, Ghosh S, Comhair SA, Farha S, Tang WHW, Park M, Wang S, Lichtin AE, Erzurum SC. High levels of zinc-protoporphyrin identify iron metabolic abnormalities in pulmonary arterial hypertension. *Clin Transl Sci* 2011;4:253–258.
- Barberà JA, Blanco I. Pulmonary hypertension in patients with chronic obstructive pulmonary disease: advances in pathophysiology and management. *Drugs* 2009;69:1153–1171.
- Nathan SD, Hassoun PM. Pulmonary hypertension due to lung disease and/or hypoxia. Clin Chest Med 2013;34:695–705.
- Tuder RM, Chacon M, Alger L, Wang J, Taraseviciene-Stewart L, Kasahara Y, Cool CD, Bishop AE, Geraci M, Semenza GL, et al. Expression of angiogenesis-related molecules in plexiform lesions in severe pulmonary hypertension: evidence for a process of disordered angiogenesis. J Pathol 2001;195:367–374.
- 24. Farha S, Asosingh K, Xu W, Sharp J, George D, Comhair S, Park M, Tang WHW, Loyd JE, Theil K, *et al.* Hypoxia-inducible factors in human pulmonary arterial hypertension: a link to the intrinsic myeloid abnormalities. *Blood* 2011;117:3485–3493.
- Andreini C, Bertini I, Rosato A. Metalloproteomes: a bioinformatic approach. Acc Chem Res 2009;42:1471–1479.
- Andreini C, Banci L, Bertini I, Elmi S. Non-heme iron through the three domains of life. *Proteins* 2007;67:317–324.
- Zhang A-S, Enns CA. Iron homeostasis: recently identified proteins provide insight into novel control mechanisms. *J Biol Chem* 2009; 284:711–715.
- Jomova K, Valko M. Advances in metal-induced oxidative stress and human disease. *Toxicology* 2011;283:65–87.
- Koskenkorva-Frank TS, Weiss G, Koppenol WH, Burckhardt S. The complex interplay of iron metabolism, reactive oxygen species, and reactive nitrogen species: insights into the potential of various iron therapies to induce oxidative and nitrosative stress. *Free Radic Biol Med* 2013;65:1174–1194.
- Mims MP, Prchal JT. Divalent metal transporter 1. *Hematology* 2005; 10:339–345.
- Cellier MFM. Nramp: from sequence to structure and mechanism of divalent metal import. *Curr Top Membr* 2012;69:249–293.
- Shawki A, Knight PB, Maliken BD, Niespodzany EJ, Mackenzie B. H(+)-coupled divalent metal-ion transporter-1: functional properties, physiological roles and therapeutics. *Curr Top Membr* 2012;70: 169–214.

- Hentze MW, Muckenthaler MU, Andrews NC. Balancing acts: molecular control of mammalian iron metabolism. *Cell* 2004;117: 285–297.
- 34. Pietrangelo A. Hereditary hemochromatosis: pathogenesis, diagnosis, and treatment. *Gastroenterology* 2010;139:393–408, 408.e1–2.
- Nemeth E, Tuttle MS, Powelson J, Vaughn MB, Donovan A, Ward DM, Ganz T, Kaplan J. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* 2004;306: 2090–2093.
- Nicolas G, Viatte L, Bennoun M, Beaumont C, Kahn A, Vaulont S. Hepcidin, a new iron regulatory peptide. *Blood Cells Mol Dis* 2002; 29:327–335.
- Merle U, Fein E, Gehrke SG, Stremmel W, Kulaksiz H. The iron regulatory peptide hepcidin is expressed in the heart and regulated by hypoxia and inflammation. *Endocrinology* 2007;148:2663–2668.
- Humbert M, Monti G, Brenot F, Sitbon O, Portier A, Grangeot-Keros L, Duroux P, Galanaud P, Simonneau G, Emilie D. Increased interleukin-1 and interleukin-6 serum concentrations in severe primary pulmonary hypertension. *Am J Respir Crit Care Med* 1995; 151:1628–1631.
- Selimovic N, Bergh C-H, Andersson B, Sakiniene E, Carlsten H, Rundqvist B. Growth factors and interleukin-6 across the lung circulation in pulmonary hypertension. *Eur Respir J* 2009;34:662–668.
- Nemeth E, Rivera S, Gabayan V, Keller C, Taudorf S, Pedersen BK, Ganz T. IL-6 mediates hypoferremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. *J Clin Invest* 2004; 113:1271–1276.
- 41. Rabinovitch M. Molecular pathogenesis of pulmonary arterial hypertension. *J Clin Invest* 2012;122:4306–4313.
- Yu PB, Beppu H, Kawai N, Li E, Bloch KD. Bone morphogenetic protein (BMP) type II receptor deletion reveals BMP ligand-specific gain of signaling in pulmonary artery smooth muscle cells. *J Biol Chem* 2005;280:24443–24450.
- 43. Andriopoulos B Jr, Corradini E, Xia Y, Faasse SA, Chen S, Grgurevic L, Knutson MD, Pietrangelo A, Vukicevic S, Lin HY, *et al.* BMP6 is a key endogenous regulator of hepcidin expression and iron metabolism. *Nat Genet* 2009;41:482–487.
- 44. Rouault TA. The role of iron regulatory proteins in mammalian iron homeostasis and disease. *Nat Chem Biol* 2006;2:406–414.
- 45. Vashisht AA, Zumbrennen KB, Huang X, Powers DN, Durazo A, Sun D, Bhaskaran N, Persson A, Uhlen M, Sangfelt O, *et al.* Control of iron homeostasis by an iron-regulated ubiquitin ligase. *Science* 2009; 326:718–721.
- 46. Salahudeen AA, Thompson JW, Ruiz JC, Ma H-W, Kinch LN, Li Q, Grishin NV, Bruick RK. An E3 ligase possessing an iron-responsive hemerythrin domain is a regulator of iron homeostasis. *Science* 2009;326:722–726.
- 47. Evstatiev R, Gasche C. Iron sensing and signalling. *Gut* 2012;61: 933–952.
- 48. Sanchez M, Galy B, Dandekar T, Bengert P, Vainshtein Y, Stolte J, Muckenthaler MU, Hentze MW. Iron regulation and the cell cycle: identification of an iron-responsive element in the 3'-untranslated region of human cell division cycle 14A mRNA by a refined microarray-based screening strategy. *J Biol Chem* 2006;281: 22865–22874.
- Sanchez M, Galy B, Muckenthaler MU, Hentze MW. Iron-regulatory proteins limit hypoxia-inducible factor-2alpha expression in iron deficiency. *Nat Struct Mol Biol* 2007;14:420–426.
- 50. Smith SR, Ghosh MC, Ollivierre-Wilson H, Hang Tong W, Rouault TA. Complete loss of iron regulatory proteins 1 and 2 prevents viability of murine zygotes beyond the blastocyst stage of embryonic development. *Blood Cells Mol Dis* 2006;36:283–287.
- 51. Anderson SA, Nizzi CP, Chang Y-I, Deck KM, Schmidt PJ, Galy B, Damnernsawad A, Broman AT, Kendziorski C, Hentze MW, *et al.* The IRP1-HIF- $2\alpha$  axis coordinates iron and oxygen sensing with erythropoiesis and iron absorption. *Cell Metab* 2013;17:282–290.
- 52. Wilkinson N, Pantopoulos K. IRP1 regulates erythropoiesis and systemic iron homeostasis by controlling HIF2 $\alpha$  mRNA translation. Blood 2013;122:1658–1668.
- 53. Ghosh MC, Zhang D-L, Jeong SY, Kovtunovych G, Ollivierre-Wilson H, Noguchi A, Tu T, Senecal T, Robinson G, Crooks DR, *et al*. Deletion of iron regulatory protein 1 causes polycythemia and pulmonary

hypertension in mice through translational derepression of HIF2 $\alpha$ . Cell Metab 2013;17:271–281.

- 54. Kawanabe Y, Nauli SM. Endothelin. Cell Mol Life Sci 2011;68:195–203.
- Shimoda LA, Semenza GL. HIF and the lung: role of hypoxia-inducible factors in pulmonary development and disease. *Am J Respir Crit Care Med* 2011;183:152–156.
- 56. Fijalkowska I, Xu W, Comhair SAA, Janocha AJ, Mavrakis LA, Krishnamachary B, Zhen L, Mao T, Richter A, Erzurum SC, et al. Hypoxia inducible-factor1alpha regulates the metabolic shift of pulmonary hypertensive endothelial cells. *Am J Pathol* 2010;176: 1130–1138.
- 57. Yu AY, Shimoda LA, Iyer NV, Huso DL, Sun X, McWilliams R, Beaty T, Sham JS, Wiener CM, Sylvester JT, *et al.* Impaired physiological responses to chronic hypoxia in mice partially deficient for hypoxiainducible factor 1alpha. *J Clin Invest* 1999;103:691–696.
- 58. Brusselmans K, Compernolle V, Tjwa M, Wiesener MS, Maxwell PH, Collen D, Carmeliet P. Heterozygous deficiency of hypoxia-inducible factor-2α protects mice against pulmonary hypertension and right ventricular dysfunction during prolonged hypoxia. *J Clin Invest* 2003; 111:1519–1527.
- Ivan M, Kondo K, Yang H, Kim W, Valiando J, Ohh M, Salic A, Asara JM, Lane WS, Kaelin WG Jr. HIFalpha targeted for VHL-mediated destruction by proline hydroxylation: implications for O2 sensing. *Science* 2001;292:464–468.
- 60. Jaakkola P, Mole DR, Tian Y-M, Wilson MI, Gielbert J, Gaskell SJ, von Kriegsheim A, Hebestreit HF, Mukherji M, Schofield CJ, *et al.* Targeting of HIF-α to the von Hippel-Lindau ubiquitylation complex by O2-regulated prolyl hydroxylation. *Science* 2001;292:468–472.
- Balanos GM, Dorrington KL, Robbins PA. Desferrioxamine elevates pulmonary vascular resistance in humans: potential for involvement of HIF-1. 2002;92:2501–2507.
- 62. Ang SO, Chen H, Hirota K, Gordeuk VR, Jelinek J, Guan Y, Liu E, Sergueeva AI, Miasnikova GY, Mole D, *et al.* Disruption of oxygen homeostasis underlies congenital Chuvash polycythemia. *Nat Genet* 2002;32:614–621.
- 63. Smith TG, Brooks JT, Balanos GM, Lappin TR, Layton DM, Leedham DL, Liu C, Maxwell PH, McMullin MF, McNamara CJ, *et al.* Mutation of von Hippel-Lindau tumour suppressor and human cardiopulmonary physiology. *PLoS Med* 2006;3:e290.
- 64. Sable CA, Aliyu ZY, Dham N, Nouraie M, Sachdev V, Sidenko S, Miasnikova GY, Polyakova LA, Sergueeva AI, Okhotin DJ, et al. Pulmonary artery pressure and iron deficiency in patients with upregulation of hypoxia sensing due to homozygous VHL(R200W) mutation (Chuvash polycythemia). *Haematologica* 2012;97: 193–200.
- 65. Hickey MM, Richardson T, Wang T, Mosqueira M, Arguiri E, Yu H, Yu Q-C, Solomides CC, Morrisey EE, Khurana TS, et al. The von Hippel-Lindau Chuvash mutation promotes pulmonary hypertension and fibrosis in mice. J Clin Invest 2010;120:827–839.
- 66. Gale DP, Harten SK, Reid CDL, Tuddenham EGD, Maxwell PH. Autosomal dominant erythrocytosis and pulmonary arterial hypertension associated with an activating HIF2 α mutation. Blood 2008;112:919–921.
- 67. Formenti F, Beer PA, Croft QPP, Dorrington KL, Gale DP, Lappin TRJ, Lucas GS, Maher ER, Maxwell PH, McMullin MF, *et al.* Cardiopulmonary function in two human disorders of the hypoxiainducible factor (HIF) pathway: von Hippel-Lindau disease and HIF-2α gain-of-function mutation. *FASEB J* 2011;25:2001–2011.
- 68. Yi X, Liang Y, Huerta-Sanchez E, Jin X, Cuo ZXP, Pool JE, Xu X, Jiang H, Vinckenbosch N, Korneliussen TS, et al. Sequencing of 50 human exomes reveals adaptation to high altitude. Science 2010;329: 75–78.
- 69. Beall CM, Cavalleri GL, Deng L, Elston RC, Gao Y, Knight J, Li C, Li JC, Liang Y, McCormack M, *et al*. Natural selection on EPAS1 (HIF2α) associated with low hemoglobin concentration in Tibetan highlanders. *Proc Natl Acad Sci USA* 2010;107:11459–11464.
- Winterbourn CC. Toxicity of iron and hydrogen peroxide: the Fenton reaction. *Toxicol Lett* 1995;82-83:969–974.
- 71. Kitamuro T, Takahashi K, Ogawa K, Udono-Fujimori R, Takeda K, Furuyama K, Nakayama M, Sun J, Fujita H, Hida W, et al. Bach1 functions as a hypoxia-inducible repressor for the heme oxygenase-1 gene in human cells. J Biol Chem 2003;278:9125–9133.

- 72. Eba S, Hoshikawa Y, Moriguchi T, Mitsuishi Y, Satoh H, Ishida K, Watanabe T, Shimizu T, Shimokawa H, Okada Y, *et al.* The nuclear factor erythroid 2-related factor 2 activator oltipraz attenuates chronic hypoxia-induced cardiopulmonary alterations in mice. *Am J Respir Cell Mol Biol* 2013;49:324–333.
- 73. Barnett CF, Hsue PY, Machado RF. Pulmonary hypertension: an increasingly recognized complication of hereditary hemolytic anemias and HIV infection. *JAMA* 2008;299:324–331.
- Gladwin MT, Kato GJ. Cardiopulmonary complications of sickle cell disease: role of nitric oxide and hemolytic anemia. *Hematology* 2005; 2005:51–57.
- 75. Klings ES, Machado RF, Barst RJ, Morris CR, Mubarak KK, Gordeuk VR, Kato GJ, Ataga KI, Gibbs JS, Castro O, et al.; American Thoracic Society Ad Hoc Committee on Pulmonary Hypertension of Sickle Cell Disease. An official American Thoracic Society clinical practice guideline: diagnosis, risk stratification, and management of pulmonary hypertension of sickle cell disease. Am J Respir Crit Care Med 2014;189:727–740.
- Farmakis D, Aessopos A. Pulmonary hypertension associated with hemoglobinopathies: prevalent but overlooked. *Circulation* 2011; 123:1227–1232.
- Musallam KM, Cappellini MD, Wood JC, Taher AT. Iron overload in non-transfusion-dependent thalassemia: a clinical perspective. *Blood Rev* 2012;26:S16–S19.
- Gladwin MT, Sachdev V. Cardiovascular abnormalities in sickle cell disease. J Am Coll Cardiol 2012;59:1123–1133.
- Akrawinthawong K, Chaowalit N, Chatuparisuth T, Siritanaratkul N. Effectiveness of deferiprone in transfusion-independent betathalassemia/HbE patients. *Hematology* 2011;16:113–122.
- Rai PR, Cool CD, King JAC, Stevens T, Burns N, Winn RA, Kasper M, Voelkel NF. The cancer paradigm of severe pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2008;178:558–564.
- Guignabert C, Tu L, Le Hiress M, Ricard N, Sattler C, Seferian A, Huertas A, Humbert M, Montani D. Pathogenesis of pulmonary arterial hypertension: lessons from cancer. *Eur Respir Rev* 2013;22:543–551.
- Lee SD, Shroyer KR, Markham NE, Cool CD, Voelkel NF, Tuder RM. Monoclonal endothelial cell proliferation is present in primary but not secondary pulmonary hypertension. *J Clin Invest* 1998;101: 927–934.

- Aldred MA, Comhair SA, Varella-Garcia M, Asosingh K, Xu W, Noon GP, Thistlethwaite PA, Tuder RM, Erzurum SC, Geraci MW, et al. Somatic chromosome abnormalities in the lungs of patients with pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2010; 182:1153–1160.
- 84. Zhao L, Ashek A, Wang L, Fang W, Dabral S, Dubois O, Cupitt J, Pullamsetti SS, Cotroneo E, Jones H, *et al*. Heterogeneity in lung (18) FDG uptake in pulmonary arterial hypertension: potential of dynamic (18)FDG positron emission tomography with kinetic analysis as a bridging biomarker for pulmonary vascular remodeling targeted treatments. *Circulation* 2013;128:1214–1224.
- 85. Yeager ME, Halley GR, Golpon HA, Voelkel NF, Tuder RM. Microsatellite instability of endothelial cell growth and apoptosis genes within plexiform lesions in primary pulmonary hypertension. *Circ Res* 2001;88:E2–E11.
- Marsboom G, Wietholt C, Haney CR, Toth PT, Ryan JJ, Morrow E, Thenappan T, Bache-Wiig P, Piao L, Paul J, et al. Lung <sup>18</sup>Ffluorodeoxyglucose positron emission tomography for diagnosis and monitoring of pulmonary arterial hypertension. Am J Respir Crit Care Med 2012;185:670–679.
- Seagroves TN, Ryan HE, Lu H, Wouters BG, Knapp M, Thibault P, Laderoute K, Johnson RS. Transcription factor HIF-1 is a necessary mediator of the pasteur effect in mammalian cells. *Mol Cell Biol* 2001;21:3436–3444.
- Kulshreshtha R, Ferracin M, Wojcik SE, Garzon R, Alder H, Agosto-Perez FJ, Davuluri R, Liu CG, Croce CM, Negrini M, *et al*. A microRNA signature of hypoxia. *Mol Cell Biol* 2007;27:1859–1867.
- Chan SY, Loscalzo J. MicroRNA-210: a unique and pleiotropic hypoxamir. *Cell Cycle* 2010;9:1072–1083.
- Rouault TA, Tong W-H. Iron-sulphur cluster biogenesis and mitochondrial iron homeostasis. *Nat Rev Mol Cell Biol* 2005;6: 345–351.
- Navarro-Sastre A, Tort F, Stehling O, Uzarska MA, Arranz JA, Del Toro M, Labayru MT, Landa J, Font A, Garcia-Villoria J, *et al*. A fatal mitochondrial disease is associated with defective NFU1 function in the maturation of a subset of mitochondrial Fe-S proteins. *Am J Hum Genet* 2011;89:656–667.
- 92. Zimmermann MB, Hurrell RF. Nutritional iron deficiency. *Lancet* 2007; 370:511–520.