

UC San Diego

UC San Diego Previously Published Works

Title

Methamphetamine and Cannabis: A Tale of Two Drugs and their Effects on HIV, Brain, and Behavior

Permalink

<https://escholarship.org/uc/item/0zw5r5sk>

Journal

Journal of Neuroimmune Pharmacology, 15(4)

ISSN

1557-1890

Authors

Saloner, Rowan
Fields, Jerel Adam
Marcondes, Maria Cecilia Garibaldi
[et al.](#)

Publication Date

2020-12-01

DOI

10.1007/s11481-020-09957-0

Peer reviewed



Published in final edited form as:

J Neuroimmune Pharmacol. 2020 December ; 15(4): 743–764. doi:10.1007/s11481-020-09957-0.

Methamphetamine and Cannabis: A Tale of Two Drugs and their Effects on HIV, Brain, and Behavior

Rowan Saloner^{1,2}, Jerel Adam Fields¹, Maria Cecilia Garibaldi Marcondes³, Jennifer E. Iudicello¹, Sofie von Känel¹, Mariana Cherner¹, Scott L. Letendre¹, Marcus Kaul^{1,4}, Igor Grant¹ Translational Methamphetamine AIDS Research Center (TMARC) Group¹

¹Department of Psychiatry, University of California, San Diego, HIV Neurobehavioral Research Program, San Diego, California

²San Diego State University/University of California, San Diego Joint Doctoral Program in Clinical Psychology, San Diego, California

³San Diego Biomedical Research Institute, San Diego, California

⁴Division of Biomedical Sciences, University of California, Riverside, Riverside, California

Abstract

HIV infection and drug use intersect epidemiologically, and their combination can result in complex effects on brain and behavior. The extent to which drugs affect the health of persons with HIV (PWH) depends on many factors including drug characteristics, use patterns, stage of HIV disease and its treatment, comorbid factors, and age. To consider the range of drug effects, we have selected two that are in common use by PWH: methamphetamine and cannabis. We compare the effects of methamphetamine with those of cannabis, to illustrate how substances may potentiate, worsen, or even buffer the effects of HIV on the CNS. Data from human, animal, and ex vivo studies provide insights into how these drugs have differing effects on the persistent inflammatory state that characterizes HIV infection, including effects on viral replication, immune activation, mitochondrial function, gut permeability, blood brain barrier integrity, glia and neuronal signaling. Moving forward, we consider how these mechanistic insights may inform interventions to improve brain outcomes in PWH.

Graphical Abstract

This review summarizes literature from clinical and preclinical studies demonstrating the adverse effects of METH, as well as the potentially beneficial effects of cannabis, on the interacting systemic (e.g., gut barrier leakage/microbial translocation, immune activation, inflammation) and

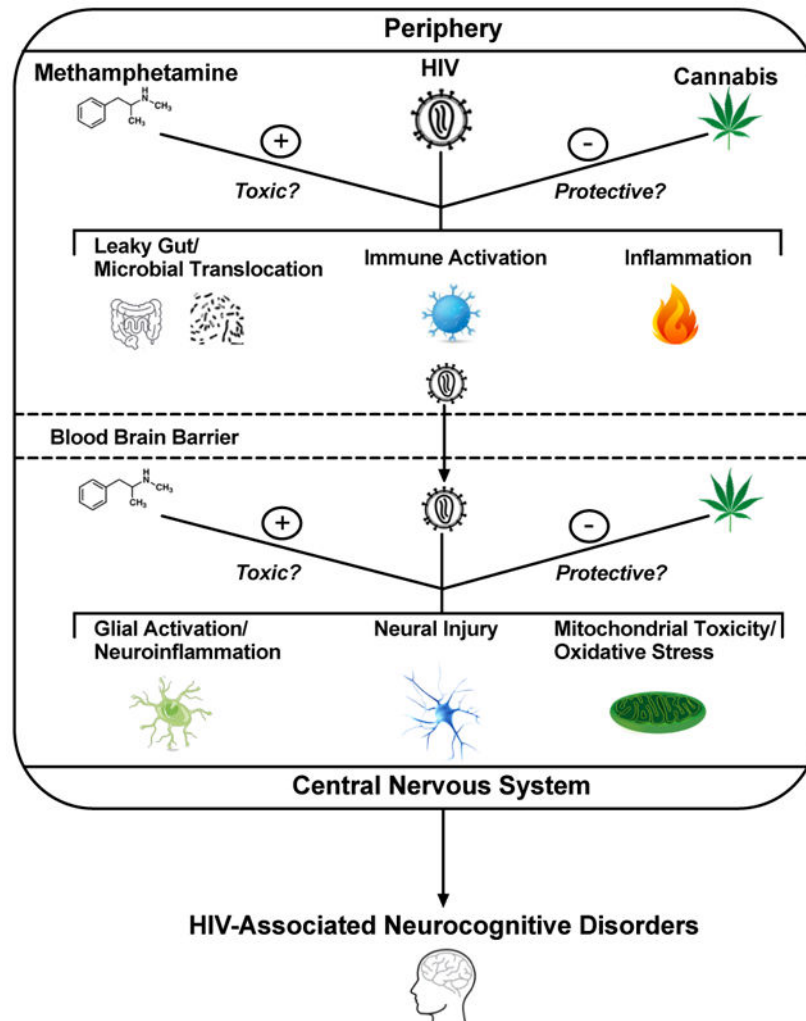
Terms of use and reuse: academic research for non-commercial purposes, see here for full terms. <http://www.springer.com/gb/open-access/authors-rights/aam-terms-v1>

Correspondence and Request for Reprints: Rowan Saloner, M.S., Predoctoral Research Fellow, SDSU/UC San Diego Joint Doctoral Program in Clinical Psychology, University of California, San Diego, HIV Neurobehavioral Research Program, 220 Dickinson Street, Suite B, Mail Code 8231, San Diego, CA 92103-8231, Phone: 619-543-5085, Fax: 619-543-1235, rsaloner@ucsd.edu.

Conflict of Interest: The authors declare no conflicts of interest.

Publisher's Disclaimer: This Author Accepted Manuscript is a PDF file of a an unedited peer-reviewed manuscript that has been accepted for publication but has not been copyedited or corrected. The official version of record that is published in the journal is kept up to date and so may therefore differ from this version.

CNS-specific (e.g., glial activation/neuroinflammation, neural injury, mitochondrial toxicity/oxidative stress) mechanisms underlying HIV-associated neurocognitive disorders.



Keywords

HIV-associated neurocognitive disorders; methamphetamine; cannabis; inflammation; blood-brain-barrier; gut-brain-axis

Introduction

Despite the success of combined antiretroviral therapy (cART) in prolonging the lifespan of persons with HIV (PWH), HIV-associated neurocognitive disorders (HAND) remain prevalent. HIV-associated dementia, a more severe complication from the persistence of HIV in the brain, has become relatively rare since the introduction of cART. However, milder forms of HAND, specifically asymptomatic neurocognitive impairment and mild neurocognitive disorder, have been reported in 30-50% of PWH, even despite suppressive ART, and can affect quality of life as well as everyday function (Heaton et al., 2010; Heaton

et al., 2011; Saloner and Cysique, 2017). The profile of HAND is heterogeneous; fronto-striatal dysfunction, manifested by disruptions in planning and reasoning (executive function) is most commonly observed, but problems in learning and memory, reduction in speed and efficiency of information processing, and attentional difficulties can also occur. The domain-specific pattern of deficits and the trajectory of these deficits over time varies between and within individuals (Woods et al., 2009; Morgan et al., 2011; Heaton et al., 2015; Dastgheyb et al., 2019; Arce Rentería et al., 2020). Importantly, HIV-associated neurocognitive deficits confer risk for impairments in real-world functioning, including unemployment, poor medication adherence, and impaired driving (Heaton et al., 2004; Marcotte et al., 2004; Moore et al., 2018a).

The heterogeneous presentation of HAND is not fully explained by inter-individual differences in disease management, but is also attributable to interactions of HIV disease with comorbidities that alter CNS function. Substance use disorders (SUDs) have long been considered a risk factor for the acquisition and transmission of HIV as well as a barrier to optimal HIV disease management (Moore et al., 2012; Elkbuli et al., 2019). Methamphetamine (METH) use has resurged nationally as indicated by the exponential increase in overdose deaths involving METH over the past decade (National Institute on Drug Abuse, 2020). METH users are at high risk of being exposed to HIV through unsafe sex practices and needle sharing (Hoenigl et al., 2016) and PWH are more likely to report METH use than persons without HIV. National rates of cannabis use have also increased due to the growing legalization of cannabis for medical and recreational use (Substance Abuse and Mental Health Services Administration, 2019). PWH frequently use cannabis recreationally and to treat HIV-related medical symptoms including neuropathic pain; compared to persons without HIV, PWH report significantly higher rates of cannabis use in their lifetime (77% vs. 45%) and within the past year (34% vs. 11%) and month (25% vs. 7%; (Montgomery et al., 2019).

The aim of this review is to provide an overview of the preclinical and clinical studies examining the pathological mechanisms, neuroimmune response, and general CNS consequences of METH and cannabis in HIV. We will not exhaustively review the literature regarding all possible pathological mechanisms. Rather, we will focus on select mechanisms involved in the neuroimmune response to METH and their neurobehavioral implications in the context of HIV infection. We will also contrast how cannabis use may exert an opposing influence on the CNS in HIV disease. Toward this end, we will in part draw upon our own experience conducting translational research studies on the CNS effects of METH and cannabis at the Translational Methamphetamine AIDS Research Center (TMARC; (Soontornniyomkij et al., 2016a).

Early studies on neuropsychological implications of HIV have highlighted the need to consider comorbid SUDs in the evaluation of HAND (Antinori et al., 2007). One influential study from Rippeth et al. (2004) reported that neurocognitive impairment was most prevalent among PWH who had METH dependence (HIV+/METH+: 58%) compared to single-risk groups (HIV+/METH-: 38%; HIV-/METH+: 40%) and HIV-/METH- controls (18%; (Rippeth et al., 2004). Carey et al. (2006) further demonstrated additive effects of immunosuppression and METH-dependence on neurocognitive impairment within PWH

(Carey et al., 2006). METH+ individuals with CD4⁺ counts lower than 200 exhibited a 77% rate of global impairment compared to 55% among METH+ individuals with CD4⁺ ≥ 200, 44% among METH- individuals with CD4⁺ < 200, and 33% among METH- individuals with CD4⁺ ≥ 200. In the absence of HIV, Gonzalez et al. (2004) also observed higher rates of impairment in METH+ individuals (50%) compared to METH- individuals (22%); (Gonzalez et al., 2004). Interestingly, it was also observed that impairment rates were attenuated in METH+ individuals with a comorbid history of cannabis use disorder (33%). Collectively, these findings have stimulated work aimed at elucidating possible neuroimmunological mechanisms underpinning the increased risk of HAND in some persons with SUDs.

HIV-mediated activation of cells of the immune system triggers inflammatory processes, as well as the production of neurotoxic viral proteins, which contribute to neuronal injury. In addition to the release of pro-inflammatory cytokines, the presence and persistence of HIV in the brain causes alterations in glutamatergic and dopaminergic systems, mitochondrial damage and oxidative stress, with important degradation of blood-brain-barrier (BBB) integrity. *In vivo* neuroimaging studies have shown compromised integrity of gray and white matter, as well as neurometabolic alterations during early infection, and many of these neuroimaging abnormalities are still observed in chronically-infected PWH on cART (Ances et al., 2009; Valcour et al., 2012; Masters and Ances, 2014; Ragin et al., 2015). Neuropathological data has demonstrated reduced concentrations of dopamine in frontostriatal regions, cerebral atrophy, synaptodendritic and myelin loss, astrogliosis, and microglial activation (Everall et al., 2009; Kumar et al., 2009; Desplats et al., 2013). Thus, HIV infection of the CNS affects neuronal and BBB integrity, through the development of an inflammatory environment, which is largely influenced by substantial changes in neurotransmitter systems in the context of drug use (Nolan and Gaskill, 2019). Neurotransmitters such as dopamine, in turn, further enhance infection and inflammation (Gaskill et al., 2014; Basova et al., 2018).

Addiction is also recognized as a condition with a neurobiological disease state characterized by the “hi-jacking” of endogenous neurotransmitter systems (Volkow et al., 2016). For example, similar to HIV, chronic stimulant use is associated with reduced dopaminergic tone, gray and white matter abnormalities, microglial activation, and frontostriatal hypometabolism (London et al., 2015; Moszczynska and Callan, 2017). From a neuroimmunological perspective, the intersection between neuroHIV and drug use results in dynamic CNS alterations that are commensurate with the complexity of the clinical manifestations of HAND. NeuroHIV as a CNS condition can be affected by drug use at different molecular and cellular levels, due to the numerous modes of action of different drugs and the diversity of molecular pathways activated during signaling, not only leading to addictive behaviors, but also interfering with cells that are targets of HIV infection in the brain, such as macrophages and microglia (Bortell et al., 2015).

Neural Mechanisms and Neuroinflammation

Although cART limits viral replication and improves CD4⁺ T-cell counts, chronic inflammation due to sustained immune activation and possibly cART itself are implicated as

a driving force behind the progression of HIV disease in the CNS in the cART era, including the persistence of HAND (Heaton et al., 2010; Heaton et al., 2011; Xu et al., 2017; Zulu et al., 2018; Fields et al., 2019). Microglia and perivascular macrophages (both cell types of myeloid lineage) initiate and regulate immune responses in the brain, signaling to astrocytes and other brain cells (Minagar et al., 2002; Liddelow et al., 2017). Microglia and perivascular macrophages also regulate clearance of extracellular aggregates, such as beta amyloid, from the brain, a process that may be impaired in reactive brain macrophages in PWH (Green et al., 2005; Achim et al., 2009; Fields et al., 2018; Mackiewicz et al., 2019; Fields et al., 2020). Astrocytes react rapidly and robustly with gene expression, metabolic, and morphological changes, that when unchecked, perpetuate chronic inflammatory signaling (Khakh and Sofroniew, 2015; Bortell et al., 2017a; Liu et al., 2018). The expression of chemokine receptors CCR5 and CXCR4 are thought to mediate the viral infection of perivascular macrophages and microglia (Kaul et al., 2007), whereas astrocytes may be capable of harboring the virus within CD81-lined vesicles (Gray et al., 2014). Even with systemic viral suppression, these CNS viral reservoirs may generate low-level HIV replication that triggers the release of viral proteins and proinflammatory cytokines and chemokines (Hellmuth et al., 2015; Marban et al., 2016). Though the extent to which astrocytes may harbor virus remains a matter of debate (Li et al., 2016; Ko et al., 2019), there is little question that they contribute to increased chemotaxis and monocyte activation following activation via HIV-infected macrophages (Muratori et al., 2010; Bortell et al., 2017a).

The notion of a chronic neuroinflammatory state in the context of suppressive cART is supported by elevated CSF and MRS markers of monocyte activation and neuroinflammation among well-treated PWH, particularly those with HAND (Harezlak et al., 2011; Masters and Ances, 2014; Anderson et al., 2015b; Schrier et al., 2015). Neuropathological studies have revealed increased presence of reactive microglia and astrocytes concomitant with increased expression of inflammatory cytokines in brain tissues from PWH and HAND (Fields et al., 2018; Swinton et al., 2019). Recently, positron emission tomography (PET) imaging studies have focused on the expression of the translocator protein (TSPO) 18kDa, a marker for microglial activation and thus neuroinflammation. These studies confirm increased microglial activation in PWH on suppressive cART and report associations between higher TSPO binding in the medial temporal lobe and thalamus with worse memory (Coughlin et al., 2014; Garvey et al., 2014; Vera et al., 2016).

Methamphetamine (METH)

Non-neuronal cells, especially innate immune HIV targets in the brain, microglia and infiltrating macrophages, are phenotypically affected by drugs of abuse, such as METH (Marcondes et al., 2010; Bortell et al., 2015; Mediouni et al., 2015; Najera et al., 2016; Basova et al., 2018). These cells may directly experience interactions with drug chemical structures, but also express receptors for the neurotransmitters that are enhanced locally by neuronal responses to drugs, including dopamine (Gaskill et al., 2012). Innate immune HIV target cells express all the DA receptors, making them responsive to the hyperdopaminergic environment of the brain in METH users (Gaskill et al., 2012; Gaskill et al., 2014; Basova et

al., 2018). One of the innate immune phenotypic characteristics that are relevant in HIV infection is the expression of the chemokine receptor and co-receptor for viral entry CCR5, which is upregulated in the brain upon METH chronic administration in macaques and in human cells (Marcondes et al., 2010; Bortell et al., 2015; Najera et al., 2016). Our group has shown that DA signaling that is increased in the context of METH can increase the expression of CCR5 and influence viral entry and spread (Basova et al., 2018). Others have shown that DC-SIGN, a molecule that also facilitates HIV dissemination, is upregulated via DA receptors (Nair et al., 2006).

Microglia and astrocytes also exhibit inflammatory responses in the presence of drugs of abuse, which in part contribute to the maladaptive alterations to neural circuitry that underlie addictive behaviors (Hauser and Knapp, 2014; Bortell et al., 2017b; Kohno et al., 2019). Our group recently reported that compared to non-transgenic control mice, Tat-transgenic mice display increased recruitment of midbrain dopamine neurons and locomotor sensitization in response to METH (Kesby et al., 2017). These reward-system adaptations in Tat mice following METH exposure were also accompanied by the elevated expression of ionized calcium binding adaptor molecule (IBA-1), a marker for microglial activation (Fig. 1). Importantly, this finding corroborated our group's prior observation that IBA-1 levels are higher in post-mortem temporo-parietal tissue of PWH with a lifetime history of METH dependence relative to those without a history of METH dependence, suggesting that METH use contributes to focal cerebral microgliosis among PWH (Soontornniyomkij et al., 2016b). An early MRS study from Chang et al. (2005) similarly found elevated glial activation and neuroinflammation, indicated by higher *myo*-inositol and choline levels, in the frontal white matter of PWH with METH use (Chang et al., 2005). Taylor et al. (2007) did not detect main effects of METH-dependence on MRS metabolites in PWH; however, higher plasma HIV RNA levels correlated with higher *myo*-inositol levels in frontal white and gray matter only among the METH-dependent group (Taylor et al., 2007).

Astrocytes regulate many aspects of brain homeostasis including BBB permeability, glucose transport, neurotransmission, synapse formation, and other processes that are disturbed during chronic inflammation (Sofroniew, 2000; Hamby and Sofroniew, 2010; Sofroniew and Vinters, 2010; Khakh and Sofroniew, 2015). These critical astrocytic functions are compromised in preclinical models of HIV-induced neuroinflammation, and the addition of drugs of abuse, including opiates and stimulants, exacerbates astrocytic dysfunction (Buch et al., 2012; Hauser and Knapp, 2014; Borgmann and Ghorpade, 2015). Astrocytic gliosis and decreased expression of excitatory amino acid transporter-2 (EAAT-2), suggestive of glutamatergic excitotoxicity, is evident in cortical tissue from PWH (Xing et al., 2009). METH has also been shown to downregulate astrocytic EAAT-2 expression via the trace amine associated receptor-1 (TAAR-1; (Borgmann and Ghorpade, 2015). Recent clinical data from our center also implicates disruption of the Wnt/ β -catenin signaling pathway, which promotes astrocytic-mediated reuptake of glutamate via EAAT-2, in the pathogenesis of HAND (Yu et al., 2017a). Nevertheless, the influence of METH on biomarkers of astrocyte homeostasis in PWH remains poorly understood.

Cannabis

There is emerging evidence that cannabis use may buffer the deleterious neuroimmune effects of high inflammation (O'Sullivan and Kendall, 2010; Bilkei-Gorzo et al., 2017; Rizzo et al., 2019; Ellis et al., 2020; Henriquez et al., 2020; Watson et al., 2020). This may occur through cannabinoid (CB)₁ receptor-mediated dampening of glutamatergic excitotoxicity and CB₂ receptor-mediated initiation of anti-inflammatory cascades (Rom and Persidsky, 2013). Some recent human studies suggest that active cannabis use may limit HIV viral replication and attenuate HIV-related immunosuppression and inflammation (Thames et al., 2016; Rizzo et al., 2018; Chaillon et al., 2019). Recent data from our group suggest that a lifetime history of cannabis use disorders lowers the odds of neurocognitive impairment in PWH (Fig. 2; (Watson et al., 2020) and may even promote “youthful” and resilient neurocognitive abilities among adults aging with HIV (Saloner et al., 2019b). Despite these promising findings, reports are inconsistent on the effects of cannabis on the brain in PWH. In a study of the combined and independent effects of chronic cannabis use and HIV on brain metabolites, Chang et al. (2006) found that cannabis use was associated with a decrease in neuronal and glial metabolites, yet a normalization of glutamate levels in PWH (Chang et al., 2006). Chronic cannabis use has also been associated with reduced gray matter volumes and memory deficits in cohorts comprising both PWH and seronegative controls (Cristiani et al., 2004; Chang et al., 2006; Battistella et al., 2014; Thames et al., 2017).

The effects of cannabis use on neuroimmune function and neurocognition are highly complex and may be dependent upon patterns of consumption, among other cannabis use characteristics. Moderate use may mitigate HIV-induced neuroinflammation and microglial activation, while heavy exposure may promote toxicity that eclipses any anti-inflammatory benefits (Childs et al., 2017; Calabrese and Rubio-Casillas, 2018). Δ^9 -tetrahydrocannabinol (THC) effects on neurogenesis and memory are not linear, with low to moderate concentrations stimulating neurogenesis and high doses inhibiting neurogenesis and memory in multiple model systems. For example, THC provides protection from neurodegenerative processes by reducing inflammation in aged mouse models for neurodegenerative diseases but induces memory impairment in healthy mice, young or aged mice (Fishbein-Kaminietsky et al., 2014; Bilkei-Gorzo et al., 2017). Similarly, endocannabinoids exert neuroprotective effects through CB₁ in models for HIV Tat-induced neurotoxicity (Xu et al., 2017).

THC may be neuroprotective by disrupting macrophage and T-cell to astrocyte inflammatory signaling, resulting in reduced inflammatory gene expression (Rizzo et al., 2019; Henriquez et al., 2020). It has been suggested that cannabinoids (e.g., THC), and possibly other agonists particularly of the CB₂ receptor, promote the transformation of microglia from their proinflammatory, cytotoxic (M1) state to neuroprotective, healing (M2) state (Tang and Le, 2016). The modulatory effects of THC on microglial/macrophage and astrocyte activation may be mediated by activation of peroxisome proliferator-activated receptors (PPARs), which has been shown to block glial inflammatory responses (Janabi, 2002; Drew et al., 2006; Xu et al., 2006; Pautz et al., 2010; Kozela et al., 2017). This may happen through direct effects of THC on PPAR transcriptional activity (O'Sullivan et al., 2005; O'Sullivan et

al., 2009; O'Sullivan and Kendall, 2010; Takeda et al., 2014), as well as indirectly through a protein kinase-dependent mechanism of PPAR α and γ phosphorylation (Rueda et al., 2000). Activation of PPAR α / γ and CB₁ and CB₂ in microglia and astrocytes by inhaled cannabis may explain the neuroprotective properties in PWH, but these interactions *in vivo* in the context of HIV and cART have not been studied.

Blood Brain Barrier Mechanisms

The BBB is a complex structure that divides the CNS from the periphery and is critical in preventing CNS infiltration of peripheral pathogens (Ballabh et al., 2004). The BBB endothelium, composed of brain microvascular endothelial cells (BMVECs) and tight junction proteins (TJPs), communicates with other components of the neurovascular unit (i.e., astrocytes, microglia, perivascular macrophages, pericytes, neurons) to coordinate biomolecular traffic in and out of the CNS (Ballabh et al., 2004). In the setting of HIV infection, proinflammatory cytokines and viral proteins induce the expression of matrix metalloproteinases and adhesion molecules (including E-selectin, vascular cell adhesion molecule 1 [VCAM-1], and intercellular cell adhesion molecule 1 [ICAM-1]) on BMVECs, which in turn facilitates the neuroinvasion of HIV-infected monocytes and CD4⁺ cells (Eugenin et al., 2006; Atluri et al., 2015). Over time, the neuroimmune response to CNS infection can produce a vicious cycle of neuroinflammation and endothelial damage that accelerates deterioration of the BBB and further promotes transmigration of HIV and peripheral toxins into the CNS (Kaul et al., 2005; Persidsky et al., 2006).

METH

Using *in vitro* cell cultures of primary BMVECs and an *in vitro* BBB model, Mahajan et al. (2008) demonstrated that HIV protein gp120 in combination with METH increased BBB permeability and altered the expression of TJPs, including ZO-1, JAM-2, Occludin, Claudin-3, and Claudin-5 (Mahajan et al., 2008). Furthermore, gp120 and Tat mice treated with METH show reductions in TJPs that can be restored with antioxidant treatment, highlighting a role of oxidative stress in stimulant-induced BBB compromise (Ramirez et al., 2009; Banerjee et al., 2010). Yao et al. demonstrated that cocaine induces the expression of monocyte chemoattractant protein-1 (MCP-1) in rodent microglia as well as activated leukocyte cell adhesion molecule in human BMVECs, which results in the accelerated adhesion and transmigration of HIV-infected monocytes across the BBB (Yao et al., 2010; Yao et al., 2011).

These cellular studies are consistent with clinical studies indicating increased neuroinvasion of peripheral viruses, including HIV and hepatitis C virus (HCV), in drug users (Kousik et al., 2012). Increased BBB permeability has also been proposed as an explanation for the unexpected observation that METH use is associated with *increased* cortical area and volume in PWH (Jernigan et al., 2005; MacDuffie et al., 2018). Under this hypothesis, HIV- and METH-related neuroinflammation alters aquaporin 4 function, a water channel protein expressed in astrocytes at cortical BBB junctions, resulting in cerebral edema that is detected as an increased volumetric signal on MRI (St Hillaire et al., 2005; Benga and Huber, 2012; MacDuffie et al., 2018). However, there is a dearth of clinical literature on the interactions

between HIV and drugs on CSF and neuroimaging biomarkers of BBB integrity. Elevated CSF/serum albumin ratio (CSAR), indicative of BBB leakage, is observed even in PWH on effective cART and has been linked to CSF biomarkers of neuronal injury and diffusion tensor imaging (DTI) markers of white matter integrity (Wright et al., 2015; Calcagno et al., 2016; Rahimy et al., 2017; Farhadian et al., 2019). A recent study also reported elevated capillary permeability in the basal ganglia and anterior frontal white matter, estimated using dynamic contrast enhanced perfusion MRI, in virally-suppressed HAND patients compared to controls (Chaganti et al., 2019). These *in vivo* studies excluded participants with drug abuse histories, did not report drug use histories, or included PWH with comorbid SUDs but did not examine SUDs as a predictor of BBB integrity. In preliminary analysis of the TMARC cohort, HIV and METH-dependence additively contributed to higher plasma levels of VCAM-1 (Fig. 3; Iudicello et al., unpublished data), a marker of endothelial cell activation that is linked to inflammation and vascular conditions such as atherosclerosis and small vessel disease (Arba et al., 2018; Kong et al., 2018). Higher levels of VCAM-1 in plasma were in turn associated with worse neurocognitive functioning, providing insight into the potential role of vascular/BBB mechanisms in HIV and METH associated neurocognitive impairment. Nevertheless, more clinical studies are needed to determine whether substance-related compromise of BBB integrity observed *in vitro* translates to fluid-based and neuroimaging biomarkers of the BBB in PWH.

Cannabis

In contrast to stimulants, cannabinoids may stabilize the BBB under conditions of high neuroinflammation. CB₂ receptor expression is increased in post-mortem BMVECs of patients with HIV encephalitis (Persidsky et al., 2011), yet several studies demonstrate that CB₂ receptor agonists reduce lipopolysaccharide (LPS)-induced inflammatory responses at the BBB, thereby increasing TJP expression and trans-endothelial resistance while reducing the surface expression of ICAM-1 and VCAM-1 (Calapai et al., 2020). Cannabinoid agonists inhibit HIV gp120-induced release of Ca²⁺ and significantly decrease the down-regulation of TJPs and permeability of BMVECs *in vitro* (Lu et al., 2008). The same study also reported that cannabinoid agonists inhibited the transmigration of human monocytes across the BBB using an *in vivo* model of mannitol-induced BBB permeability in mice. Very recent work from our group has extended these findings into the clinical realm. Using CSAR and soluble urokinase plasminogen activator receptor values to derive a composite BBB index, Ellis et al. (2020) reported a significant HIV by cannabis interaction on BBB integrity such that more frequent use of cannabis in the past month related to lower BBB index values (less “leakage”) in PWH but not HIV– individuals (Ellis et al., 2020). Notably, lower BBB index values associated with less CSF neurofilament light, a marker of axonal injury.

Mitochondrial Function and Oxidative Stress Mechanisms

Since the beginning of the epidemic, PWH have presented with disease markers associated with mitochondrial damage and oxidative stress. HIV proteins and cART induce changes in mitochondria that likely lead to increases in oxidative stress (reviewed by (Fields and Ellis, 2019). In postmortem brain specimens of PWH, evidence has been found for disruptions in mitochondrial biogenesis, mitochondrial dynamics fission and fusion, mitochondrial

transport, and recycling of damaged mitochondrial via mitophagy (Fields et al., 2015a; Fields et al., 2015b; Avdoshina et al., 2016; Swinton et al., 2019). Metabolomic studies have also found that PWH with depression exhibit reduced plasma levels of acylcarnitine, indicative of mitochondrial dysfunction (Cassol et al., 2015). Another study found reduced levels of N-acetylaspartate (NAA), a putative marker of neuronal integrity and mitochondrial injury (Bates et al., 1996), in frontal white matter of PWH taking didanosine or stavudine (Schweinsburg et al., 2005). Moreover, neuroimaging studies from the CNS HIV Anti-Retroviral Therapy Effects Research (CHARTER) cohort indicate negative correlations between frontal cortex levels of NAA and markers of monocyte activation (IP-10 and MCP-1) in PWH (Anderson et al., 2015a), as well as lower basal ganglia and frontal white matter levels of NAA on MRS in PWH with severe comorbidity burden (Saloner et al., 2019a).

HIV proteins, namely gp120, Tat, Nef, and Vpr are all associated with mitochondrial damage and oxidative stress in *in vitro* and *in vivo* models (Fields and Ellis, 2019). Moreover, antiretroviral drugs have also been associated with mitochondrial damage. The original reverse transcriptase inhibitors damaged mitochondrial DNA in the periphery, likely through inhibiting mitochondrial polymerase γ , which is responsible for mtDNA replication (Fields and Ellis, 2019). More recently, the new generation of cART drugs has been shown to alter mitochondrial function *in vitro* and *in vivo* (Fields et al., 2019). Despite these findings, surprisingly little is known about how various drugs of abuse affect mitochondrial function and oxidative stress in PWH.

METH

In the aforementioned MRS studies, Chang et al. (2005) observed additive effects of HIV and METH on lower NAA in the basal ganglia and frontal gray and white matter (Chang et al., 2005), while Taylor et. al (2007) observed a correlation between higher plasma HIV RNA and lower NAA in frontal white matter in HIV+/METH+ (Taylor et al., 2007). In a post-mortem analysis of frontal brain tissues from controls without a history of METH use and PWH with and without a history of METH use, Var et al. (2016) quantified mitochondrial injury as the proportion of mitochondrial DNA carrying the “common deletion,” a genetic mutation reflective of oxidative stress (Var et al., 2016). HIV+/METH– demonstrated the highest levels of mitochondrial injury, which in turn correlated with worse global cognition within the HIV+/METH– group. An opposite effect was detected in tissues from HIV+/METH+ donors such that this group had the lowest proportion of mtDNA carrying the common deletion, yet this apparent reduction in mitochondrial injury also conferred risk for poorer neurocognition. These findings are consistent with prior observations from our group that increased cortical volumes and higher fractional anisotropy values relate to better neurocognition in PWH but worse neurocognition in METH+ individuals (Jernigan et al., 2005; Soontornniyomkij et al., 2016a), which may reflect a compensatory consequence of METH-related neuropathology as opposed to a neuroprotective effect of METH.

In a rodent model of self-administration of METH under long access conditions, which elicits compulsive METH intake similar to patterns of consumption in METH-dependent

humans, HIV-transgenic rats exhibited greater evidence of impaired aerobic glucose metabolism, neural injury, and inflammation compared to wild-type rats (de Guglielmo et al., 2020). Langford et al. (2003; 2004) demonstrated that the combination of HIV and METH reduces calbindin-immunoreactivity in nonpyramidal neurons and these neurotoxic effects were accompanied by mitochondrial damage and oxidative stress, possibly through a mechanism involving the mitochondrial calcium potential (Langford et al., 2003; Langford et al., 2004). Importantly, a follow-up study of PWH with METH use reported that memory deficits 6-months prior to death correlated with loss of frontal calbindin interneurons at autopsy (Chana et al., 2006). Similarly, a DTI study in mice demonstrated that METH-induced increases in hippocampal mean diffusivity also correlated with a loss of hippocampal calbindin expression (McKenna et al., 2016). *In vitro* studies have shown that METH may induce mitochondrial damage and oxidative stress in astrocytes exposed to HIV (Borgmann and Ghorpade, 2018). Banjeree et al found that a combination of gp120, Tat and METH induced robust increases in markers for oxidative stress, GSH and MDA. Importantly, N-acetylcysteine amide (NACA) protected the BBB from oxidative stress-induced damage, though the mechanisms of this protection were not determined. However, a study by Zeng et al. (2018) using *in vitro* and *in vivo* models showed that Tat- and METH-induced oxidative cellular injury is mitigated by NACA via modulation of mTOR signaling (Zeng et al., 2018).

Cannabis

Cannabinoids, namely THC and cannabidiol (CBD), and some synthetic cannabinoid receptor agonists affect mitochondrial function in different types of brain cells. The effects of cannabis on HIV-induced mitochondrial function and oxidative stress have been investigated using *in vitro* models for brain cells. Some studies have investigated the effects of cannabinoids on HIV-induced neurotoxicity using *in vivo* models, but data on mitochondrial alterations are largely lacking. We recently found that a cannabinoid receptor agonist blocks mitochondrial dysfunction in astrocytes after exposure to pro-inflammatory cytokines that are relevant to HAND (Swinton et al., 2019). Moreover, conditioned media from the reactive astrocytes in culture reduced mitochondrial biogenesis markers in neurons and this was blocked by treating the astrocytes with the CB receptor agonist WIN55,212-2 (Fig. 4). Another study found that THC blocks monocyte mediated activation of astrocytes and subsequent expression of MCP-1, possibly explaining how cannabis users may be protected from HIV and cART-induced neurotoxicity (Rizzo et al., 2019).

Gut Microbiome Mechanisms

Alterations to the gut microbiome (i.e., changes in enteric bacterial diversity) coupled with increased gut permeability can lead to greater microbial translocation, which can in turn disrupt neuroendocrine and neuroimmune homeostasis (Pellegrini et al., 2018). For example, regulatory T-cells survey gut-associated lymphoid tissue (GALT) and changes in gut microbiome composition can promote T-cell brain infiltration. Circulating bacterial factors, such as bacterial LPS which acts on endothelial toll-like receptors, can alter BBB integrity and promote neuroinflammation. Taken together, gut dysbiosis is now implicated in a host of neurological and psychiatric disorders (Kim and Shin, 2018; Ma et al., 2019). An emerging

topic in HIV and substance use research is how these conditions alter the composition of the gut microbiome and how gut changes influence systemic organ and brain functioning. HIV replication in GALT results in immune-mediated damage to intestinal epithelial cells, thereby degrading the integrity of the gut-blood barrier and facilitating the leakage of intestinal bacteria into systemic circulation (Dillon et al., 2016). Similarly, gut dysbiosis has been observed in response to alcohol, stimulants, and opiates, although no specific dysbiotic signature has emerged across or within drug classes (Meckel and Kiraly, 2019). In addition to substance-induced alterations to gut integrity, lifestyle and dietary factors are likely to influence the composition of the gut microbiome in individuals with SUDs (Volpe et al., 2014; Singh et al., 2017).

METH

Recent studies have investigated the impact of METH use on the gut microbiome in HIV-seropositive men who have sex with men. In these studies, METH use moderated the effects of HIV on microbial composition and was also independently associated with microbial alterations favoring the expression of pro-inflammatory bacteria (Fulcher et al., 2018b; Cook et al., 2019). Furthermore, recent METH use related to increased levels of IL-6 and TNF-alpha in rectal sponge samples, regardless of HIV serostatus (Fulcher et al., 2018a). Despite the putative relevance of microbial translocation in the neuropathogenesis of HIV and METH, the combined effects of HIV and METH on gut-brain relationships remain vastly understudied. Some studies suggest that treatments that alter the gut microbiome and reduce microbial translocation may mitigate the effects of HIV disease and drug use on HAND (Gori et al., 2011; Ceccarelli et al., 2017), however future studies must clarify the role of confounds such as poor diet and HCV, integrate comprehensive gut microbiome, CNS biomarker, and neurobehavioral phenotypes, and examine the impact of gut health interventions (e.g., prebiotic supplementation) on neurocognition to allow for such a conclusion.

Cannabis

In addition to the brain, CB₁ and CB₂ receptors are expressed on enteroendocrine L cells that are innervated by enteric glial cells and afferent neurons (Yoo and Mazmanian, 2017). CB receptors in the large intestine are activated by the bioactive lipids anandamide and 2-arachidonoylglycerol and the synthesis and degradation of these cannabinoids is modulated by structurally similar bioactive lipids (Cani et al., 2016a; Acharya et al., 2017; Chiurchiù et al., 2018). Some endocannabinoids and related bioactive lipids function as epithelial barrier “gate-keepers” by reinforcing TJP integrity and reducing inflammation while others function as “gate-openers” by decreasing the thermogenesis of brown adipose tissue and increasing inflammation (Cani et al., 2016b). In addition to CB receptor-dependent mechanisms, the endocannabinoid system may facilitate gut barrier homeostasis via PPAR-dependent pathways (Muccioli et al., 2010). In a recent simian immunodeficiency virus (SIV) model, gut epithelial disruption was accompanied by reduced PPAR α signaling and mitochondrial dysfunction, yet restoration of the intestinal barrier was accomplished via probiotic-induced enhancement of PPAR α signaling and restoration of mitochondrial function and fatty acid β -oxidation (Crakes et al., 2019). In another SIV study, chronic THC administration inhibited activation of immune and pro-inflammatory pathways in lamina propria leukocytes and gut

epithelium (Kumar et al., 2016). Macrophage shift to the proinflammatory M1 state also contributes to gut cytotoxic cascades. Interestingly CB₂ agonists reverse some of the inflammatory processes in models of inflammatory bowel disease, possibly by facilitating a shift to the M2 phenotype. Further, in LPS models of neural injury, CB₂ agonists may protect against neurotoxicity, possibly via a similar M1 to M2 phenotype shift (Reiner et al., 2014; Presley et al., 2015). While these preclinical findings suggest that cannabis may ameliorate HIV-induced immune activation, inflammation, and oxidative stress in the gut, either independently or in synergy with probiotics, these mechanisms have not been investigated in humans. Translational paradigms that examine the interface between the endocannabinoid system and the gut-brain axis in both PWH and HIV transgenic animals would importantly bridge this critical gap in knowledge (Fig. 5).

Animal Models of NeuroHIV

Overview

Animal models have been critical for research on HIV-associated brain injury as well as for exploring the mechanistic underpinnings of SUDs. These models span a range of species, including chimpanzees and other non-human primates, cats, rats and mice (Gardner and Luciw, 1989; Klotman and Notkins, 1996; Toggas and Mucke, 1996; Nath et al., 2000; Reid et al., 2001; Keppler et al., 2002; Ambrose et al., 2007; Van Duyne et al., 2009). Of those, non-human primates infected with SIV and rodents (rats and mice) have been the primary models used for neuroHIV and SUD research, including METH and cannabinoids (Olmsted et al., 1989; Clements et al., 1994; Madden et al., 2005; Ambrose et al., 2007; Clements et al., 2008; Williams et al., 2008; Liu et al., 2009; Winsauer et al., 2011; Molina et al., 2015; Najera et al., 2016; Simon et al., 2016; Thaney et al., 2018). One study treated SIV-infected rhesus macaques with THC for several months and found that the animals became tolerant to the behavioral effects of THC while lacking any increase of viral titers in plasma, CSF and brain tissue and displaying reduced CNS pathology in comparison vehicle controls (Winsauer et al., 2011). In contrast, METH exposure increased SIV levels and immune activation in the brain of SIV-infected rhesus macaques (Marcondes et al., 2010).

Rodents are not permissive to productive infection with wild-type HIV-1. However, certain immuno-compromised mouse strains can be 'humanized' by reconstitution with human immune or hematopoietic stem cells. The resulting human peripheral blood cell populations are permissive to HIV infection and thus provide small animal models for HIV/AIDS and neuroAIDS research (Van Duyne et al., 2009; Dash et al., 2011; Thaney et al., 2018). Another important advantage of rodents, both mice and rats, is that they can be genetically modified (Klotman and Notkins, 1996; Toggas and Mucke, 1996; Reid et al., 2001; Van Duyne et al., 2009). Several transgenic mouse lines and a rat have been generated that express an entire HIV genome or a truncated version with certain viral components, such as gp120, Tat or Vpr (Leonard et al., 1988; Iwakura et al., 1992; Hanna et al., 1998b; Hanna et al., 1998a; Reid et al., 2001). A number of studies investigating the intersection of HIV and stimulants have been performed in the HIV gp120-transgenic mouse model, which expresses a soluble viral envelope gp120 of HIV-1 in the brain (Toggas et al., 1994; Roberts et al., 2010; Bandaru et al., 2011; Soontornniyomkij et al., 2016a). This gp120 model shares

Author Manuscript

hallmarks of neuropathology with human neuroHIV patients, including loss of neuronal dendrites and synapses, activated microglia, astrocytosis, and compromised neurogenesis (Toggas et al., 1994; Okamoto et al., 2007; Crews et al., 2011; Lee et al., 2011; Lee et al., 2013; Avraham et al., 2014; Fields et al., 2014; Maung et al., 2014; Avraham et al., 2015; Steiner et al., 2015; Thaney et al., 2017). In comparison to non-transgenic littermate controls, gp120 mice also display memory impairments, perturbations in electrophysiological function, and share patterns of differential gene expression with human HIV brains (Krucker et al., 1998; D'hooge et al., 1999; Maung et al., 2014; Hofer et al., 2015; Thaney et al., 2017).

Author Manuscript

In comparison to non-transgenic controls, the gp120 mice display an altered acute response to METH that is discernable in stereotypic behavior (Roberts et al., 2010). Another study detected in gp120 mice an increased preference for both METH and a highly palatable non-drug reinforcer (saccharin) as well as increased sensitivity to METH-induced conditioned reward, providing a potentially explanation for a frequent abuse by HIV-infected individuals (Kesby et al., 2014). The HIV-transgenic rat has also been employed to assess alterations of behavior and the dopaminergic system associated with HIV infection and the effects of METH on sensorimotor gating and locomotor activity (Liu et al., 2009; Moran et al., 2012; Moran et al., 2013). HIV-expressing rats displayed greater behavioral sensitization due to METH than non-transgenic controls.

Translational Behavioral Findings

Author Manuscript

A more direct example of a translational, cross-species paradigm is the behavioral pattern monitor (BPM), which constitutes a modification for humans of the traditional open field test for rodents (Perry et al., 2009; Young et al., 2016). The mouse BPM assesses potential inhibition deficits in male and female mice, reflected by increased motor activity, inappropriate perseverative behavior, and elevated exploration of novel stimuli (Henry et al., 2013). The reported observations indicated that both gp120 and chronic METH exposure affected behavioral inhibition in a sex-dependent fashion. While robust gender differences have not been reported in human studies using the BPM, known gender differences in HIV disease warrant further investigation of potential behavioral correlates (Wilson et al., 2006; Addo and Altfeld, 2014).

Author Manuscript

Prepulse inhibition (PPI) is a measure of sensorimotor gating that is regulated by several neural networks including the dopaminergic circuitry implicated in inhibition and can also be investigated across species. Compromised sensorimotor inhibition, assessed by PPI of the eyeblink startle response, has been reported in PWH with neurocognitive impairment when compared to neurocognitively intact PWH (Minassian et al., 2013). This observation suggested that early inhibition deficits accompany or possibly precede downstream neurocognitive impairment in PWH. In rodent models, PPI is quantified using the whole-body startle response, and PPI in gp120 and METH-exposed mice was investigated (Henry et al., 2014). Prior to METH exposure, female gp120 mice exhibited decreased PPI while male gp120 mice displayed increased acoustic startle response compared to their respective non-transgenic controls. The observations in gp120 and METH treated mice and the results in humans indicating PPI deficits in PWH with neurocognitive impairment (Minassian et al.,

2013) indicate that inhibition deficits are affected by HIV- and METH-induced alterations of dopaminergic neurotransmission, which manifest not as a global phenomenon but rather in association with higher-order cognitive deficits or biological variations, such as sexual dimorphism.

Using an attentional-set-shifting task (Young et al., 2010) to assess discrimination learning in gp120 mice exposed to an escalating-dose, multiple-binge METH regimen, Kesby et al. (2015) demonstrated impaired learning in gp120 mice, regardless of METH exposure, which was concordant with the pattern of learning deficits observed on standard neuropsychological testing in humans stratified by HIV serostatus and METH-dependence (Kesby et al., 2015). In a separate study employing the same METH regimen in gp120 mice, Hoefer et al. (2015) showed that similar to gp120, METH triggered a significant loss of pre-synaptic terminals and neuronal dendrites in the hippocampus and cerebral cortex of non-transgenic animals (Hoefer et al., 2015). Electrophysiology analysis of hippocampal slices demonstrated that METH-treated gp120 mice exhibit significantly reduced post-tetanic potentiation, while both METH and gp120 expression resulted in impaired long-term potentiation. Notably, these pre- and post-synaptic alterations also occurred in conjunction with impaired learning and memory in the METH-exposed gp120 mice. More recently, Kesby et al. (2018) demonstrated additive effects of Tat and METH exposure in mice on perseverative errors during reversal learning, which may reflect Tat- and METH-induced alterations in dopaminergic tone in the orbitofrontal cortex and caudate putamen (Kesby et al., 2018). This executive dysfunction in METH-exposed Tat-transgenic mice is also concordant with perseverative responding observed in METH-using PWH on the Wisconsin Card Sorting Task (Fig. 6), a gold standard neuropsychological measure of executive function. Overall, the similar pattern of outcomes suggests that gp120 and Tat independently and in combination with METH can contribute to behavioral deficits across species, with some indication of a protein-specific dissociation in the constructs involved such that gp120 may preferentially compromise learning and memory whereas Tat may modulate dopaminergic circuitry involved in perseverative responding.

Additional Considerations

Aging

As the proportion of PWH over the age of 50 steadily rises (Centers for Disease Control and Prevention, 2018), attention is shifting toward the interactions of HIV and drug use on neurocognitive and brain aging. In a cross-sectional design, Iudicello et al. (2014) observed a negative impact of a remote history of METH dependence on neurocognition (including memory, attention, and executive function) and everyday functioning (including employment) in older, but not younger PWH (Iudicello et al., 2014), suggesting that older age may enhance vulnerability to the detrimental neurocognitive effects of prior METH use even in currently abstinent PWH. Using longitudinal data from the CHARTER cohort, Heaton et al. (2015) reported that a lifetime history of METH use disorder at baseline resulted in a 70% increase in risk of experiencing clinically-significant neurocognitive decline over an average study period of three-years (Heaton et al., 2015). This indication of an adverse “legacy” effect of historical METH use on age-related neurobehavior in PWH

was supported by a recent study (Paolillo et al., 2019). Frailty, a clinical proxy of biological age indexed by the proportion of accumulated age-related multi-system health deficits, was higher among PWH with a lifetime METH use disorder compared to PWH and HIV– individuals without a history of METH use (Fig. 7). Notably, higher frailty exhibited the strongest association with worse executive function and working memory in the dual-risk METH+ PWH group, suggesting that prefrontal dysfunction linked to past METH use among PWH may in part be explained by the presence of a geriatric phenotype. This is consistent with studies suggesting that HIV-induced immune dysfunction may accelerate cellular aging (Deeks, 2011), which may progress even more rapidly in the setting of METH abuse (Cohen and Torres, 2017; Papageorgiou et al., 2019). HIV and METH can induce astrocyte senescence *in vitro* and across multiple animal models, which contributes to neuronal toxicity via downregulation of β -catenin signaling (Yu et al., 2017b). Other cellular and molecular mechanisms of neuroimmunosenescence that overlap in HIV and METH pathogenesis may involve autophagic dysregulation (Cao et al., 2017), particularly mitophagy (Borgmann and Ghorpade, 2018; Teodorof-Diedrich and Spector, 2018), and brain DNA methylome alterations in genes enriched in neurodegenerative disease (Desplats et al., 2014).

Older adults represent the fastest growing segment of cannabis users nationally, as indicated by a 71.4% increase in the past-year prevalence of cannabis use between 2006 and 2013 among adults aged 50 and older (Han et al., 2017; Lloyd and Striley, 2018). However, findings on the effects of cannabis use on neurocognition and brain integrity in older adults without HIV are scattered and inconsistent (Yoo et al., 2020). A recent systematic review of older adults (ages 50 and older) with and without clinical disorders (e.g., Parkinson’s disease) provided some evidence of a modest adverse effect of heavy cannabis exposure on neurocognition, predominantly verbal memory, yet many studies yielded null results and a few studies reported better subjective and global neurocognition in older cannabis users compared to non-users (Scott et al., 2019). The influence of cannabis on neurocognitive and brain aging in HIV is even more limited, and therefore represents a major gap in our current understanding of mechanisms of neurobehavioral resilience and vulnerability in the growing population of older PWH.

Host Genetics

Clinical studies frequently observe heterogeneous patterns of neurocognitive and biomarker profiles in substance-dependent PWH, necessitating an increased focus on moderating co-factors that may help explain inter-individual differences. Self-reported parameters of METH use (e.g., duration of use, frequency of use, length of abstinence) do not strongly correlate with neurocognitive performance (Johanson et al., 2006; Cherner et al., 2010b), suggesting that other factors such as host genetics may better explain susceptibility to METH-related neural injury. To this end, candidate gene studies have examined associations of single-nucleotide polymorphisms (SNPs) involved in putative pathways of HAND and drug use with neurocognitive phenotypes. For example, METH-dependent individuals who carry SNPs that confer sustained dopaminergic signaling through increased dopamine binding affinity or decreased dopamine metabolism do not exhibit the same neurocognitive benefits of high dopaminergic tone as non-users and may be at enhanced risk for

neurocognitive impairment due to dopamine-mediated increases in viral replication (Gaskill et al., 2009; Bousman et al., 2010; Gupta et al., 2011; Cherner et al., 2019). Dopaminergic SNPs similarly moderate the neurocognitive effects of other drugs in PWH and in general are implicated in the heritability of impulsivity and sensation-seeking traits that subserve addictive behaviors (Derringer et al., 2010; Khadka et al., 2014; Holmes et al., 2016). A composite approach that aggregates dopaminergic SNPs into a continuous polygenic risk score has shown promise in identifying individual risk for neurobehavioral impairments in other neuropsychiatric conditions such as schizophrenia and affective disorders (Pearson-Fuhrhop et al., 2014; Wang et al., 2018), but has not been comprehensively explored in the context of HIV and SUDs. Genetic variation in the metabolism of drugs and cART may also alter risk for HAND as they directly influence expression of enzymatic factors (e.g., COMT, cytochrome enzymes) that modulate CNS exposure (levels and duration of exposure) to the exogenous substance and its metabolites, which may also be neurotoxic (Cherner et al., 2010a; Saloner et al., 2020). The crosstalk between drugs in the CNS and neuroimmune factors is likely influenced by genetic variation in other putative pathogenic pathways of HAND, including inflammatory, mitochondrial, immune, iron-regulation, and BBB pathways (Jia et al., 2017; Olivier et al., 2018; Sundermann et al., 2019), yet this is also underexplored.

Summary

It has been long understood that abusive drugs can worsen HIV-related health outcomes and have neurotoxic potential. However, in this review we demonstrate how two commonly used substances in PWH, METH and cannabis, can have opposing actions on the CNS in HIV disease, which in turn contributes to the clinical complexity of HAND. With respect to METH, the extant literature provides converging lines of evidence that HIV and METH conspire to overburden the neuroimmunological system. Pro-inflammatory signaling in the periphery and microbial translocation due to increased gut barrier permeability facilitate the transmigration of activated monocytes across a BBB that is already compromised due to local immunomodulatory effects of HIV and METH. Subsequent enhancement of HIV RNA replication and pro-inflammatory signaling in the CNS contributes to microglial activation and disruption of astrocytic homeostasis. In addition to glial-mediated mechanisms of neuronal injury, such as mitochondrial damage, oxidative stress, and excitotoxicity, viral proteins and METH promote neuroadaptations and neurotoxicity in frontostriatal dopaminergic pathways that impair higher-order neurocognitive functions and perpetuate addictive behaviors. Together, these mechanisms modify HIV infection in the brain, conferring novel characteristics and causing the pathogenesis to become fundamentally different in the context of METH, when compared to neuroHIV in non-using individuals.

Behavioral interventions that promote abstinence from METH and adherence to cART in METH-dependent PWH are considered the first-step in mitigating the CNS burden of METH in HIV disease (Moore et al., 2012; Moore et al., 2018b). However, many individuals struggle to maintain abstinence and METH-related neurobehavioral deficits can persist even among PWH with protracted periods of abstinence (Iudicello et al., 2014; Paolillo et al., 2019). Given that there are no established adjuvant therapies for treating HAND (Bougea et al., 2019), identification of molecular targets within these pathogenic pathways is of critical

importance for neurotherapeutic development. HIV and METH may interfere with trophic factor gene expression and signaling cascades, thereby limiting the production of neurotrophic and neuroprotective factors that promote synaptodendritic plasticity and reduce glial activation and neuroinflammation (Ellis et al., 2007). For example, fibroblast growth factor (FGF)-1 inactivates the pro-apoptotic kinase glycogen synthase kinase (GSK) 3 β signaling pathway (Crews et al., 2009), which may confer protection against dendritic spine loss due to HIV gp120 and promote the survival of glutamatergic neurons and calbindin-immunoreactive interneurons (Everall et al., 2001; Everall et al., 2002; Ellis et al., 2007). Our group reported that HIV, METH-dependence, and accompanying neurocognitive deficits are associated with lower CSF levels of fibroblast growth factor FGF-1 (Bharti et al., 2016). Pharmacological approaches that upregulate neurotrophic factors like FGF-1, or modulate downstream signaling factors (e.g., GSK 3 β), as is the case with oral lithium, may ameliorate neuronal injury due to HIV and METH. The second-generation NMDAR receptor antagonist nitromemantine is a nascent therapeutic that may also protect against glutamatergic toxicity but has yet to be tested in PWH (Takahashi et al., 2015; Nakanishi et al., 2016; Bougea et al., 2019).

Importantly, not all substance use worsens neuroHIV and in the case of cannabis, there may in fact be therapeutic levels that mitigate the aforementioned pro-inflammatory mechanisms of HIV-related neuroimmune injury. Moreover, emerging preclinical data also suggests that CBD may attenuate addictive behaviors across multiple drug classes (Prud'homme et al., 2015; Gonzalez-Cuevas et al., 2018), with recent data showing that CBD administration reduced self-administration of METH and METH-seeking behavior in rats (Hay et al., 2018). Activation of CB₂ receptors may shift the expression of macrophages to an anti-inflammatory phenotype and these anti-inflammatory effects may limit mitochondrial toxicity and help restore gut and BBB integrity. Reduced BBB permeability would importantly limit the entry of HIV and neurotoxins into the CNS, however it would also limit the distribution of cART into the CNS, which may lead to poorer viral suppression but also less potential for cART-induced neurotoxicity. Nevertheless, there is a paucity of research with respect to cannabis exposure in gp120 and Tat mouse models and studies of cannabis effects in PWH are similarly lacking rigorous scientific investigation. Translational paradigms, similar to those that have yielded valuable insights into the effects of METH on neuroHIV, may help inform evidence-based approaches to cannabis in HIV, including the possibility of new neuroprotective strategies. The ecological validity of these paradigms may also be advanced with careful consideration for real-world patterns of drug use, including polysubstance use and differences in route of administration (e.g., METH injection vs. inhalation, cannabis ingestion vs. inhalation). Toward this end, emerging technologies such as ecological momentary assessment offer a promising avenue for real-time measurement of polysubstance use and its neurobehavioral antecedents and consequences in humans (Paolillo et al., 2018), which may in turn inform preclinical experimental manipulations to multiple agents. This may be particularly relevant for METH-dependent PWH who concomitantly use cannabis, as the extent to which cannabis may mitigate the adverse effects of METH is likely dependent upon the relative timing, dosage, and context in which these two drugs are consumed. Given that cannabis may function via a hormetic effect on the brain, with low doses showing beneficial effects while high doses show detrimental effects

(Fig. 8), it is critical to understand both the potential utility and risks of cannabis, and specific cannabinoids, in the treatment and control of chronic HIV disease and its neurologic complications.

Acknowledgments

This work was supported by grants from the National Institutes of Health. National Institute of Drug Abuse: P50 DA026306 (Translational Methamphetamine AIDS Research Center [TMARC]) to IG, R01 DA036164 and R01 DA047822 to MCGM, K23 DA037793 and R01 DA047879 to JEI; National Institute of Mental Health: K01 MH115819 to JAF, P30 MH062512 to JEI, R01 MH087332, R01 MH104131, and R01 MH105330 to MK. Stipend support to RS is funded by National Institute of Aging award F31 AG064989.

We thank the HIV Neurobehavioral Research Program and TMARC investigators that graciously provided material for producing manuscript figures. Specifically, we thank Ronald J. Ellis, James Kesby, David J. Moore, Emily W. Paolillo, Mary K. Swinton, and Caitlin Wei-Ming Watson.

The Translational Methamphetamine AIDS Research Center (TMARC) is supported by Center award P50DA026306 from the National Institute on Drug Abuse (NIDA) and is affiliated with the University of California, San Diego (UCSD), the Sanford-Burnham Medical Discovery Institute (SBMDI), and the University of California, Riverside (UCR). The TMARC comprises: Administrative Coordinating Core (ACC) – Executive Unit: Director – Igor Grant, M.D.; Co-Directors – Ronald J. Ellis, M.D., Ph.D., Scott L. Letendre, M.D., and Cristian L. Achim, M.D., Ph.D.; Center Manager – Mariana Cherner, Ph.D.; Associate Center Managers – Erin E. Morgan, Ph.D. and Jared Young, Ph.D.; Data Management and Information Systems (DMIS) Unit: Ian S. Abramson, Ph.D. (Unit Chief), Clint Cushman, B.A. (Unit Manager); ACC – Statistics Unit: Florin Vaida, Ph.D. (Unit Chief), Ian S. Abramson, Ph.D., Reena Deutsch, Ph.D., Anya Umlauf, M.S.; ACC – Participant Unit: J. Hampton Atkinson, M.D. (Unit Chief), Jennifer Marquie-Beck, M.P.H. (Unit Manager); Behavioral Assessment and Medical (BAM) Core – Neuromedical and Laboratory Unit (NLU): Scott L. Letendre, M.D. (Core Co-Director/NLU Chief), Ronald J. Ellis, M.D., Ph.D.; BAM Core – Neuropsychiatric Unit (NPU): Robert K. Heaton, Ph.D. (Core Co-Director/NPU Chief), J. Hampton Atkinson, M.D., Thomas D. Marcotte, Ph.D., Erin E. Morgan, Ph.D., Matthew Dawson (NPU Manager); Neuroimaging (NI) Core: Gregory G. Brown, Ph.D. (Core Director), Thomas T. Liu, Ph.D., Miriam Scadeng, Ph.D., Christine Fennema-Notestine, Ph.D., Sarah L. Archibald, M.A., John R. Hesselink, M.D., Mary Jane Meloy, Ph.D., Craig E.L. Stark, Ph.D.; Neuroscience and Animal Models (NAM) Core: Cristian L. Achim, M.D., Ph.D. (Core Director), Marcus Kaul, Ph.D., Virawudh Soontornniyomkij, M.D.; Pilot and Developmental (PAD) Core: Mariana Cherner, Ph.D. (Core Director), Stuart A. Lipton, M.D., Ph.D.; Project 1: Arpi Minassian, Ph.D. (Project Director), William Perry, Ph.D., Mark A. Geyer, Ph.D., Jared W. Young, Ph.D.; Project 2: Amanda B. Grethe, Ph.D. (Project Director), Susan F. Tapert, Ph.D., Assawin Gongvatana, Ph.D.; Project 3: Erin E. Morgan, Ph.D. (Project Director), Igor Grant, M.D.; Project 4: Samuel Barnes, Ph.D. (Project Director); Project 5: Marcus Kaul, Ph.D. (Project Director), Ana Sanchez, Ph.D.

The views expressed in this article are those of the authors and do not reflect the official policy or position of the United States Government.

References

- Acharya N, Penukonda S, Shcheglova T, Hagymasi AT, Basu S, Srivastava PK (2017) Endocannabinoid system acts as a regulator of immune homeostasis in the gut. *Proc Natl Acad Sci U S A* 114:5005–5010. [PubMed: 28439004]
- Achim CL, Adame A, Dumaop W, Everall IP, Masliah E, Neurobehavioral Research C (2009) Increased accumulation of intraneuronal amyloid beta in HIV-infected patients. *Journal of neuroimmune pharmacology : the official journal of the Society on NeuroImmune Pharmacology* 4:190–199. [PubMed: 19288297]
- Addo MM, Altfeld M (2014) Sex-based differences in HIV type 1 pathogenesis. *J Infect Dis* 209 Suppl 3:S86–92. [PubMed: 24966195]
- Ambrose Z, Kewalramani VN, Bieniasz PD, Hatzioannou T (2007) HIV/AIDS: in search of an animal model. *Trends Biotechnol* 25:333–337. [PubMed: 17574286]
- Ances BM, Sisti D, Vaida F, Liang CL, Leontiev O, Perthen JE, Buxton RB, Benson D, Smith DM, Little SJ, Richman DD, Moore DJ, Ellis RJ (2009) Resting cerebral blood flow: a potential biomarker of the effects of HIV in the brain. *Neurology* 73:702–708. [PubMed: 19720977]

- Anderson AM, Fennema-Notestine C, Umlauf A, Taylor MJ, Clifford DB, Marra CM, Collier AC, Gelman BB, McArthur JC, McCutchan JA, Simpson DM, Morgello S, Grant I, Letendre SL (2015a) CSF biomarkers of monocyte activation and chemotaxis correlate with magnetic resonance spectroscopy metabolites during chronic HIV disease. *Journal of neurovirology* 21:559–567. [PubMed: 26069183]
- Anderson AM, Fennema-Notestine C, Umlauf A, Taylor MJ, Clifford DB, Marra CM, Collier AC, Gelman BB, McArthur JC, McCutchan JA, Simpson DM, Morgello S, Grant I, Letendre SL, Group C (2015b) CSF biomarkers of monocyte activation and chemotaxis correlate with magnetic resonance spectroscopy metabolites during chronic HIV disease. *Journal of neurovirology* 21:559–567. [PubMed: 26069183]
- Antinori A et al. (2007) Updated research nosology for HIV-associated neurocognitive disorders. *Neurology* 69:1789–1799. [PubMed: 17914061]
- Arba F et al. (2018) Small vessel disease and biomarkers of endothelial dysfunction after ischaemic stroke. *European Stroke Journal* 4:119–126. [PubMed: 31259260]
- Arce Renteria M, Byrd D, Coulehan K, Miranda C, Fuentes A, Rosario AK, Morris EP, Rivera Mindt M (2020) Neurocognitive intra-individual variability within HIV+ adults with and without current substance use. *Neuropsychology* 34:321–330. [PubMed: 31886690]
- Atluri VSR, Hidalgo M, Samikkannu T, Kurapati KRV, Jayant RD, Sagar V, Nair MPN (2015) Effect of human immunodeficiency virus on blood-brain barrier integrity and function: an update. *Front Cell Neurosci* 9:212–212. [PubMed: 26113810]
- Avdoshina V, Fields JA, Castellano P, Dedoni S, Palchik G, Trejo M, Adame A, Rockenstein E, Eugenin E, Masliah E, Mochetti I (2016) The HIV Protein gp120 Alters Mitochondrial Dynamics in Neurons. *Neurotox Res* 29:583–593. [PubMed: 26936603]
- Avraham HK, Jiang S, Fu Y, Rockenstein E, Makriyannis A, Zvonok A, Masliah E, Avraham S (2014) The cannabinoid CB(2) receptor agonist AM1241 enhances neurogenesis in GFAP/Gp120 transgenic mice displaying deficits in neurogenesis. *Br J Pharmacol* 171:468–479. [PubMed: 24148086]
- Avraham HK, Jiang S, Fu Y, Rockenstein E, Makriyannis A, Wood J, Wang L, Masliah E, Avraham S (2015) Impaired neurogenesis by HIV-1-Gp120 is rescued by genetic deletion of fatty acid amide hydrolase enzyme. *Br J Pharmacol* 172:4603–4614. [PubMed: 24571443]
- Bandaru VV, Patel N, Ewaleifoh O, Haughey NJ (2011) A failure to normalize biochemical and metabolic insults during morphine withdrawal disrupts synaptic repair in mice transgenic for HIV-gp120. *J Neuroimmune Pharmacol* 6:640–649. [PubMed: 21748284]
- Banerjee A, Zhang X, Manda KR, Banks WA, Ercal N (2010) HIV proteins (gp120 and Tat) and methamphetamine in oxidative stress-induced damage in the brain: Potential role of the thiol antioxidant N-acetylcysteine amide. *Free Radical Biology and Medicine* 48:1388–1398. [PubMed: 20188164]
- Basova L, Najera JA, Bortell N, Wang D, Moya R, Lindsey A, Semenova S, Ellis RJ, Marcondes MCG (2018) Dopamine and its receptors play a role in the modulation of CCR5 expression in innate immune cells following exposure to Methamphetamine: Implications to HIV infection. *PLoS One* 13:e0199861. [PubMed: 29944719]
- Bates TE, Strangward M, Keelan J, Davey GP, Munro PM, Clark JB (1996) Inhibition of N-acetylaspartate production: implications for 1H MRS studies in vivo. *Neuroreport* 7:1397–1400. [PubMed: 8856684]
- Battistella G, Fornari E, Annoni JM, Chtioui H, Dao K, Fabritius M, Favrat B, Mall JF, Maeder P, Giroud C (2014) Long-Term Effects of Cannabis on Brain Structure. *Neuropsychopharmacology* 39:2041–2048. [PubMed: 24633558]
- Benga O, Huber VJ (2012) Brain water channel proteins in health and disease. *Molecular aspects of medicine* 33:562–578. [PubMed: 22504060]
- Bharti AR, Woods SP, Ellis RJ, Cherner M, Rosario D, Potter M, Heaton RK, Everall IP, Masliah E, Grant I, Letendre SL (2016) Fibroblast growth factors 1 and 2 in cerebrospinal fluid are associated with HIV disease, methamphetamine use, and neurocognitive functioning. *HIV AIDS (Auckl)* 8:93–99. [PubMed: 27199571]

- Bilkei-Gorzo A, Albayram O, Draffehn A, Michel K, Piyanova A, Oppenheimer H, Dvir-Ginzberg M, Racz I, Ulas T, Imbeault S, Bab I, Schultze JL, Zimmer A (2017) A chronic low dose of Delta(9)-tetrahydrocannabinol (THC) restores cognitive function in old mice. *Nat Med* 23:782–787. [PubMed: 28481360]
- Borgmann K, Ghorpade A (2015) HIV-1, methamphetamine and astrocytes at neuroinflammatory Crossroads. *Frontiers in Microbiology* 6:1143. [PubMed: 26579077]
- Borgmann K, Ghorpade A (2018) Methamphetamine Augments Concurrent Astrocyte Mitochondrial Stress, Oxidative Burden, and Antioxidant Capacity: Tipping the Balance in HIV-Associated Neurodegeneration. *Neurotox Res* 33:433–447. [PubMed: 28993979]
- Bortell N, Morsey B, Basova L, Fox HS, Marcondes MC (2015) Phenotypic changes in the brain of SIV-infected macaques exposed to methamphetamine parallel macrophage activation patterns induced by the common gamma-chain cytokine system. *Front Microbiol* 6:900. [PubMed: 26441851]
- Bortell N, Basova L, Semenova S, Fox HS, Ravasi T, Marcondes MCG (2017a) Astrocyte-specific overexpressed gene signatures in response to methamphetamine exposure in vitro. *Journal of neuroinflammation* 14:49–49. [PubMed: 28279172]
- Bortell N, Basova L, Semenova S, Fox HS, Ravasi T, Marcondes MC (2017b) Astrocyte-specific overexpressed gene signatures in response to methamphetamine exposure in vitro. *J Neuroinflammation* 14:49. [PubMed: 28279172]
- Bougea A, Spantideas N, Galanis P, Gkekas G, Thomaidis T (2019) Optimal treatment of HIV-associated neurocognitive disorders: myths and reality. A critical review. *Ther Adv Infect Dis* 6:2049936119838228–2049936119838228. [PubMed: 31001421]
- Bousman CA, Cherner M, Glatt SJ, Atkinson JH, Grant I, Tsuang MT, Everall IP (2010) Impact of COMT Val158Met on executive functioning in the context of HIV and methamphetamine. *Neurobehavioral HIV medicine* 2010:1–11. [PubMed: 24078782]
- Buch S, Yao H, Guo M, Mori T, Mathias-Costa B, Singh V, Seth P, Wang J, Su T-P (2012) Cocaine and HIV-1 interplay in CNS: cellular and molecular mechanisms. *Curr HIV Res* 10:425–428. [PubMed: 22591366]
- Calabrese EJ, Rubio-Casillas A (2018) Biphasic effects of THC in memory and cognition. *Eur J Clin Invest* 48:e12920. [PubMed: 29574698]
- Calapai F, Cardia L, Sorbara EE, Navarra M, Gangemi S, Calapai G, Mannucci C (2020) Cannabinoids, Blood-Brain Barrier, and Brain Disposition. *Pharmaceutics* 12.
- Calcagno A, Atzori C, Romito A, Vai D, Audagnotto S, Stella ML, Montrucchio C, Imperiale D, Di Perri G, Bonora S (2016) Blood brain barrier impairment is associated with cerebrospinal fluid markers of neuronal damage in HIV-positive patients. *Journal of neurovirology* 22:88–92. [PubMed: 26246357]
- Cani PD, Plovier H, Van Hul M, Geurts L, Delzenne NM, Druart C, Everard A (2016a) Endocannabinoids—at the crossroads between the gut microbiota and host metabolism. *Nat Rev Endocrinol* 12:133–143. [PubMed: 26678807]
- Cani PD, Plovier H, Van Hul M, Geurts L, Delzenne NM, Druart C, Everard A (2016b) Endocannabinoids — at the crossroads between the gut microbiota and host metabolism. *Nature Reviews Endocrinology* 12:133–143.
- Cao L, Glazyrin A, Kumar S, Kumar A (2017) Role of Autophagy in HIV Pathogenesis and Drug Abuse. *Molecular neurobiology* 54:5855–5867. [PubMed: 27660273]
- Carey CL, Woods SP, Rippeth JD, Gonzalez R, Heaton RK, Grant I, The HIVNRCG (2006) Additive Deleterious Effects of Methamphetamine Dependence and Immunosuppression on Neuropsychological Functioning in HIV Infection. *AIDS and behavior* 10:185. [PubMed: 16477511]
- Cassol E, Misra V, Morgello S, Kirk GD, Mehta SH, Gabuzda D (2015) Altered Monoamine and Acylcarnitine Metabolites in HIV-Positive and HIV-Negative Subjects With Depression. *J Acquir Immune Defic Syndr* 69:18–28. [PubMed: 25942456]
- Ceccarelli G, Fratino M, Selvaggi C, Giustini N, Serafino S, Schietroma I, Corano Scheri G, Pavone P, Passavanti G, Alunni Fegatelli D, Mezzaroma I, Antonelli G, Vullo V, Scagnolari C, d'Ettore G (2017) A pilot study on the effects of probiotic supplementation on neuropsychological

- performance and microRNA-29a-c levels in antiretroviral-treated HIV-1-infected patients. *Brain Behav* 7:e00756–e00756. [PubMed: 28828217]
- Centers for Disease Control and Prevention (2018) HIV Among People Aged 50 and Over. In: Centers for Disease Control and Prevention.
- Chaganti J, Marripudi K, Staub LP, Rae CD, Gates TM, Moffat KJ, Brew BJ (2019) Imaging correlates of the blood-brain barrier disruption in HIV-associated neurocognitive disorder and therapeutic implications. *AIDS* 33.
- Chaillon A, Nakazawa M, Anderson C, Christensen-Quick A, Ellis RJ, Franklin D, Morris SR, Gianella S (2019) Effect of Cannabis Use on Human Immunodeficiency Virus DNA During Suppressive Antiretroviral Therapy. *Clinical Infectious Diseases* 70:140–143.
- Chana G, Everall IP, Crews L, Langford D, Adame A, Grant I, Cherner M, Lazzaretto D, Heaton R, Ellis R, Masliah E (2006) Cognitive deficits and degeneration of interneurons in HIV+ methamphetamine users. *Neurology* 67:1486–1489. [PubMed: 17060582]
- Chang L, Ernst T, Speck O, Grob CS (2005) Additive effects of HIV and chronic methamphetamine use on brain metabolite abnormalities. *The American journal of psychiatry* 162:361–369. [PubMed: 15677602]
- Chang L, Cloak C, Yakupov R, Ernst T (2006) Combined and Independent Effects of Chronic Marijuana Use and HIV on Brain Metabolites. *Journal of neuroimmune pharmacology : the official journal of the Society on Neuroimmune Pharmacology* 1:65–76. [PubMed: 18040792]
- Cherner M, Bousman C, Everall I, Barron D, Letendre S, Vaida F, Atkinson JH, Heaton R, Grant I (2010a) Cytochrome P450-2D6 extensive metabolizers are more vulnerable to methamphetamine-associated neurocognitive impairment: preliminary findings. *Journal of the International Neuropsychological Society : JINS* 16:890–901. [PubMed: 20727252]
- Cherner M, Watson CW, Saloner R, Halpin LE, Minassian A, Murray SS, Vaida F, Bousman C, Everall I (2019) Adverse effect of catechol-O-methyltransferase (COMT) Val158Met met/met genotype in methamphetamine-related executive dysfunction. *Addictive behaviors* 98:106023. [PubMed: 31301644]
- Cherner M, Suarez P, Casey C, Deiss R, Letendre S, Marcotte T, Vaida F, Atkinson JH, Grant I, Heaton RK, Group H (2010b) Methamphetamine use parameters do not predict neuropsychological impairment in currently abstinent dependent adults. *Drug Alcohol Depend* 106:154–163. [PubMed: 19815352]
- Childs E, Lutz JA, de Wit H (2017) Dose-related effects of delta-9-THC on emotional responses to acute psychosocial stress. *Drug Alcohol Depend* 177:136–144. [PubMed: 28599212]
- Chiurchiù V, Leuti A, Maccarrone M (2018) Bioactive Lipids and Chronic Inflammation: Managing the Fire Within. *Frontiers in immunology* 9:38–38. [PubMed: 29434586]
- Clements JE, Mankowski JL, Gama L, Zink MC (2008) The accelerated simian immunodeficiency virus macaque model of human immunodeficiency virus-associated neurological disease: from mechanism to treatment. *J Neurovirol* 14:309–317. [PubMed: 18780232]
- Clements JE, Anderson MG, Zink MC, Joag SV, Narayan O (1994) The SIV model of AIDS encephalopathy. Role of neurotropic viruses in diseases. *Res Publ Assoc Res Nerv Ment Dis* 72:147–157. [PubMed: 8115711]
- Cohen J, Torres C (2017) HIV-associated cellular senescence: A contributor to accelerated aging. *Ageing Res Rev* 36:117–124. [PubMed: 28017881]
- Cook RR, Fulcher JA, Tobin NH, Li F, Lee DJ, Woodward C, Javanbakht M, Brookmeyer R, Shoptaw S, Bolan R, Aldrovandi GM, Gorbach PM (2019) Alterations to the Gastrointestinal Microbiome Associated with Methamphetamine Use among Young Men who have Sex with Men. *Scientific Reports* 9:14840. [PubMed: 31619731]
- Coughlin JM et al. (2014) Regional brain distribution of translocator protein using [(11)C]DPA-713 PET in individuals infected with HIV. *Journal of neurovirology* 20:219–232. [PubMed: 24567030]
- Crakes KR, Santos Rocha C, Grishina I, Hirao LA, Napoli E, Gaulke CA, Fenton A, Datta S, Arredondo J, Marco ML, Sankaran-Walters S, Cortopassi G, Giulivi C, Dandekar S (2019) PPAR α -targeted mitochondrial bioenergetics mediate repair of intestinal barriers at the host-microbe intersection during SIV infection. *Proceedings of the National Academy of Sciences* 116:24819.

- Crews L, Patrick C, Achim CL, Everall IP, Masliah E (2009) Molecular pathology of neuroAIDS (CNS-HIV). *Int J Mol Sci* 10:1045–1063. [PubMed: 19399237]
- Crews L, Ruf R, Patrick C, Dumaop W, Trejo-Morales M, Achim CL, Rockenstein E, Masliah E (2011) Phosphorylation of collapsin response mediator protein-2 disrupts neuronal maturation in a model of adult neurogenesis: Implications for neurodegenerative disorders. *Mol Neurodegener* 6:67. [PubMed: 21943307]
- Cristiani SA, Pukay-Martin ND, Bornstein RA (2004) Marijuana use and cognitive function in HIV-infected people. *The Journal of neuropsychiatry and clinical neurosciences* 16:330–335. [PubMed: 15377740]
- D'hooge R, Franck F, Mucke L, De Deyn PP (1999) Age-related behavioural deficits in transgenic mice expressing the HIV-1 coat protein gp120. *Eur J Neurosci* 11:4398–4402. [PubMed: 10594667]
- Dash PK, Gorantla S, Gendelman HE, Knibbe J, Casale GP, Makarov E, Epstein AA, Gelbard HA, Boska MD, Poluektova LY (2011) Loss of neuronal integrity during progressive HIV-1 infection of humanized mice. *J Neurosci* 31:3148–3157. [PubMed: 21368026]
- Dastgheyb RM, Sacktor N, Franklin D, Letendre S, Marcotte T, Heaton R, Grant I, McArthur JC, Rubin LH, Haughey NJ (2019) Cognitive Trajectory Phenotypes in Human Immunodeficiency Virus–Infected Patients. *JAIDS Journal of Acquired Immune Deficiency Syndromes* 82.
- de Guglielmo G, Fu Y, Chen J, Larrosa E, Hoang I, Kawamura T, Lorrain I, Zorman B, Bryant J, George O, Sumazin P, Lefebvre C, Repunte-Canonigo V, Sanna PP (2020) Increases in compulsivity, inflammation, and neural injury in HIV transgenic rats with escalated methamphetamine self-administration under extended-access conditions. *Brain research* 1726:146502. [PubMed: 31605699]
- Deeks SG (2011) HIV infection, inflammation, immunosenescence, and aging. *Annu Rev Med* 62:141–155. [PubMed: 21090961]
- Derringer J, Krueger RF, Dick DM, Saccone S, Grucza RA, Agrawal A, Lin P, Almasy L, Edenberg HJ, Foroud T, Nurnberger JI Jr., Hesselbrock VM, Kramer JR, Kuperman S, Porjesz B, Schuckit MA, Bierut LJ, Gene Environment Association Studies C (2010) Predicting sensation seeking from dopamine genes. A candidate-system approach. *Psychol Sci* 21:1282–1290. [PubMed: 20732903]
- Desplats P, Dumaop W, Cronin P, Gianella S, Woods S, Letendre S, Smith D, Masliah E, Grant I (2014) Epigenetic alterations in the brain associated with HIV-1 infection and methamphetamine dependence. *PLoS One* 9:e102555–e102555. [PubMed: 25054922]
- Desplats P, Dumaop W, Smith D, Adame A, Everall I, Letendre S, Ellis R, Cherner M, Grant I, Masliah E (2013) Molecular and pathologic insights from latent HIV-1 infection in the human brain. *Neurology* 80:1415–1423. [PubMed: 23486877]
- Dillon SM, Frank DN, Wilson CC (2016) The gut microbiome and HIV-1 pathogenesis: a two-way street. *AIDS* 30:2737–2751. [PubMed: 27755100]
- Drew PD, Xu J, Storer PD, Chavis JA, Racke MK (2006) Peroxisome proliferator-activated receptor agonist regulation of glial activation: relevance to CNS inflammatory disorders. *Neurochem Int* 49:183–189. [PubMed: 16753239]
- Elkbuli A, Polcz V, Dowd B, McKenney M, Prado G (2019) HIV prevention intervention for substance users: a review of the literature. *Substance Abuse Treatment, Prevention, and Policy* 14:1.
- Ellis R, Langford D, Masliah E (2007) HIV and antiretroviral therapy in the brain: neuronal injury and repair. *Nature Reviews Neuroscience* 8:33–44. [PubMed: 17180161]
- Ellis RJ, Peterson S, Cherner M, Morgan E, Schrier R, Tang B, Hoenigl M, Letendre S, Iudicello J (2020) Beneficial Effects of Cannabis on Blood Brain Barrier Function in HIV. *Clinical Infectious Diseases*.
- Eugenin EA, Osiecki K, Lopez L, Goldstein H, Calderon TM, Berman JW (2006) CCL2/monocyte chemoattractant protein-1 mediates enhanced transmigration of human immunodeficiency virus (HIV)-infected leukocytes across the blood-brain barrier: a potential mechanism of HIV-CNS invasion and NeuroAIDS. *J Neurosci* 26:1098–1106. [PubMed: 16436595]
- Everall I, Vaida F, Khanlou N, Lazzaretto D, Achim C, Letendre S, Moore D, Ellis R, Cherner M, Gelman B, Morgello S, Singer E, Grant I, Masliah E, National Neuro ATC (2009)

- Cliniconeuropathologic correlates of human immunodeficiency virus in the era of antiretroviral therapy. *Journal of neurovirology* 15:360–370. [PubMed: 20175693]
- Everall IP, Trillo-Pazos G, Bell C, Mallory M, Sanders V, Masliah E (2001) Amelioration of Neurotoxic Effects of HIV Envelope Protein gp120 by Fibroblast Growth Factor: A Strategy for Neuroprotection. *Journal of Neuropathology & Experimental Neurology* 60:293–301. [PubMed: 11245213]
- Everall IP, Bell C, Mallory M, Langford D, Adame A, Rockenstein E, Masliah E (2002) Lithium ameliorates HIV-gp120-mediated neurotoxicity. *Molecular and cellular neurosciences* 21:493–501. [PubMed: 12498789]
- Farhadian SF, Mistry H, Kirchwey T, Chiarella J, Calvi R, Chintanaphol M, Patel P, Landry ML, Robertson K, Spudich SS (2019) Markers of CNS Injury in Adults Living With HIV With CSF HIV Not Detected vs Detected <20 Copies/mL. *Open Forum Infectious Diseases* 6.
- Fields J, Dumaop W, Langford TD, Rockenstein E, Masliah E (2014) Role of neurotrophic factor alterations in the neurodegenerative process in HIV associated neurocognitive disorders. *J Neuroimmune Pharmacol* 9:102–116. [PubMed: 24510686]
- Fields J, Dumaop W, Elueteri S, Campos S, Serger E, Trejo M, Kosberg K, Adame A, Spencer B, Rockenstein E, He JJ, Masliah E (2015a) HIV-1 Tat alters neuronal autophagy by modulating autophagosome fusion to the lysosome: implications for HIV-associated neurocognitive disorders. *J Neurosci* 35:1921–1938. [PubMed: 25653352]
- Fields JA, Ellis RJ (2019) HIV in the cART era and the mitochondrial: immune interface in the CNS. *Int Rev Neurobiol* 145:29–65. [PubMed: 31208526]
- Fields JA, Swinton MK, Soontornniyomkij B, Carson A, Achim CL (2020) Beta amyloid levels in cerebrospinal fluid of HIV-infected people vary by exposure to antiretroviral therapy. *AIDS* 34.
- Fields JA, Spencer B, Swinton M, Qvale EM, Marquine MJ, Alexeeva A, Gough S, Soontornniyomkij B, Valera E, Masliah E, Achim CL, Desplats P (2018) Alterations in brain TREM2 and Amyloid- β levels are associated with neurocognitive impairment in HIV-infected persons on antiretroviral therapy. *J Neurochem* 147:784–802. [PubMed: 30152135]
- Fields JA, Swinton MK, Carson A, Soontornniyomkij B, Lindsay C, Han MM, Frizzi K, Sambhwani S, Murphy A, Achim CL, Ellis RJ, Calcutt NA (2019) Tenofovir disoproxil fumarate induces peripheral neuropathy and alters inflammation and mitochondrial biogenesis in the brains of mice. *Sci Rep* 9:17158. [PubMed: 31748578]
- Fields JA, Serger E, Campos S, Divakaruni AS, Kim C, Smith K, Trejo M, Adame A, Spencer B, Rockenstein E, Murphy AN, Ellis RJ, Letendre S, Grant I, Masliah E (2015b) HIV alters neuronal mitochondrial fission/fusion in the brain during HIV-associated neurocognitive disorders. *Neurobiol Dis.*
- Fishbein-Kaminietsky M, Gafni M, Sarne Y (2014) Ultralow doses of cannabinoid drugs protect the mouse brain from inflammation-induced cognitive damage. *J Neurosci Res* 92:1669–1677. [PubMed: 25042014]
- Fulcher JA, Shoptaw S, Makgoeng SB, Elliott J, Ibarondo FJ, Ragsdale A, Brookmeyer R, Anton PA, Gorbach PM (2018a) Brief Report: Recent Methamphetamine Use Is Associated With Increased Rectal Mucosal Inflammatory Cytokines, Regardless of HIV-1 Serostatus. *Journal of acquired immune deficiency syndromes (1999)* 78:119–123. [PubMed: 29419567]
- Fulcher JA, Hussain SK, Cook R, Li F, Tobin NH, Ragsdale A, Shoptaw S, Gorbach PM, Aldrovandi GM (2018b) Effects of Substance Use and Sex Practices on the Intestinal Microbiome During HIV-1 Infection. *The Journal of infectious diseases* 218:1560–1570. [PubMed: 29982500]
- Gardner MB, Luciw PA (1989) Animal models of AIDS. *FASEB J* 3:2593–2606. [PubMed: 2556312]
- Garvey LJ, Pavese N, Politis M, Ramlackhansingh A, Brooks DJ, Taylor-Robinson SD, Winston A (2014) Increased microglia activation in neurologically asymptomatic HIV-infected patients receiving effective ART. *AIDS* 28:67–72. [PubMed: 23887068]
- Gaskill PJ, Carvalho L, Eugenin EA, Berman JW (2012) Characterization and function of the human macrophage dopaminergic system: implications for CNS disease and drug abuse. *J Neuroinflammation* 9:203. [PubMed: 22901451]

- Gaskill PJ, Yano HH, Kalpana GV, Javitch JA, Berman JW (2014) Dopamine receptor activation increases HIV entry into primary human macrophages. *PLoS One* 9:e108232–e108232. [PubMed: 25268786]
- Gaskill PJ, Calderon TM, Luers AJ, Eugenin EA, Javitch JA, Berman JW (2009) Human immunodeficiency virus (HIV) infection of human macrophages is increased by dopamine: a bridge between HIV-associated neurologic disorders and drug abuse. *Am J Pathol* 175:1148–1159. [PubMed: 19661443]
- Gonzalez R, Rippeth JD, Carey CL, Heaton RK, Moore DJ, Schweinsburg BC, Cherner M, Grant I (2004) Neurocognitive performance of methamphetamine users discordant for history of marijuana exposure. *Drug and Alcohol Dependence* 76:181–190. [PubMed: 15488342]
- Gonzalez-Cuevas G, Martin-Fardon R, Kerr TM, Stouffer DG, Parsons LH, Hammell DC, Banks SL, Stinchcomb AL, Weiss F (2018) Unique treatment potential of cannabidiol for the prevention of relapse to drug use: preclinical proof of principle. *Neuropsychopharmacology* 43:2036–2045. [PubMed: 29686308]
- Gori A, Rizzardini G, Van't Land B, Amor KB, van Schaik J, Torti C, Quirino T, Tincati C, Bandera A, Knol J, Benlhassan-Chahour K, Trabattoni D, Bray D, Vriesema A, Welling G, Garssen J, Clerici M (2011) Specific prebiotics modulate gut microbiota and immune activation in HAART-naive HIV-infected adults: results of the "COPA" pilot randomized trial. *Mucosal immunology* 4:554–563. [PubMed: 21525866]
- Gray LR, Turville SG, Hitchen TL, Cheng W-J, Ellett AM, Salimi H, Roche MJ, Wesselingh SL, Gorry PR, Churchill MJ (2014) HIV-1 entry and trans-infection of astrocytes involves CD81 vesicles. *PLoS One* 9.
- Green DA, Masliah E, Vinters HV, Beizai P, Moore DJ, Achim CL (2005) Brain deposition of beta-amyloid is a common pathologic feature in HIV positive patients. *AIDS* 19.
- Gupta S, Bousman CA, Chana G, Cherner M, Heaton RK, Deutsch R, Ellis RJ, Grant I, Everall IP (2011) Dopamine receptor D3 genetic polymorphism (rs6280TC) is associated with rates of cognitive impairment in methamphetamine-dependent men with HIV: preliminary findings. *Journal of neurovirology* 17:239–247. [PubMed: 21491142]
- Hamby ME, Sofroniew MV (2010) Reactive astrocytes as therapeutic targets for CNS disorders. *Neurotherapeutics* 7:494–506. [PubMed: 20880511]
- Han BH, Sherman S, Mauro PM, Martins SS, Rotenberg J, Palamar JJ (2017) Demographic trends among older cannabis users in the United States, 2006–13. *Addiction* 112:516–525. [PubMed: 27767235]
- Hanna Z, Kay DG, Rebai N, Guimond A, Jothy S, Jolicoeur P (1998a) Nef harbors a major determinant of pathogenicity for an AIDS-like disease induced by HIV-1 in transgenic mice. *Cell* 95:163–175. [PubMed: 9790524]
- Hanna Z, Kay DG, Cool M, Jothy S, Rebai N, Jolicoeur P (1998b) Transgenic mice expressing human immunodeficiency virus type 1 in immune cells develop a severe AIDS-like disease. *J Virol* 72:121–132. [PubMed: 9420207]
- Harezlak J, Buchthal S, Taylor M, Schifitto G, Zhong J, Daar E, Alger J, Singer E, Campbell T, Yiannoutsos C, Cohen R, Navia B, Consortium HIVN (2011) Persistence of HIV-associated cognitive impairment, inflammation, and neuronal injury in era of highly active antiretroviral treatment. *AIDS* 25:625–633. [PubMed: 21297425]
- Hauser KF, Knapp PE (2014) Interactions of HIV and drugs of abuse: the importance of glia, neural progenitors, and host genetic factors. *Int Rev Neurobiol* 118:231–313. [PubMed: 25175867]
- Hay GL, Baracz SJ, Everett NA, Roberts J, Costa PA, Arnold JC, McGregor IS, Cornish JL (2018) Cannabidiol treatment reduces the motivation to self-administer methamphetamine and methamphetamine-primed relapse in rats. *Journal of Psychopharmacology* 32:1369–1378. [PubMed: 30260267]
- Heaton RK, Marcotte TD, Mindt MR, Sadek J, Moore DJ, Bentley H, McCutchan JA, Reicks C, Grant I (2004) The impact of HIV-associated neuropsychological impairment on everyday functioning. *Journal of the International Neuropsychological Society* : JINS 10:317–331. [PubMed: 15147590]

- Heaton RK et al. (2015) Neurocognitive change in the era of HIV combination antiretroviral therapy: the longitudinal CHARTER study. *Clin Infect Dis* 60:473–480. [PubMed: 25362201]
- Heaton RK et al. (2010) HIV-associated neurocognitive disorders persist in the era of potent antiretroviral therapy: CHARTER Study. *Neurology* 75:2087–2096. [PubMed: 21135382]
- Heaton RK et al. (2011) HIV-associated neurocognitive disorders before and during the era of combination antiretroviral therapy: differences in rates, nature, and predictors. *Journal of neurovirology* 17:3–16. [PubMed: 21174240]
- Hellmuth J, Valcour V, Spudich S (2015) CNS reservoirs for HIV: implications for eradication. *Journal of virus eradication* 1:67–71. [PubMed: 26430703]
- Henriquez JE, Bach AP, Matos-Fernandez KM, Crawford RB, Kaminski NE (2020) 9-Tetrahydrocannabinol (THC) Impairs CD8+ T Cell-Mediated Activation of Astrocytes. *Journal of Neuroimmune Pharmacology*.
- Henry BL, Geyer MA, Buell M, Perry W, Young JW, Minassian A, Translational Methamphetamine ARCG (2013) Behavioral effects of chronic methamphetamine treatment in HIV-1 gp120 transgenic mice. *Behav Brain Res* 236:210–220. [PubMed: 22960458]
- Henry BL, Geyer MA, Buell MR, Perry W, Young JW, Minassian A, Translational Methamphetamine ARCG (2014) Prepulse inhibition in HIV-1 gp120 transgenic mice after withdrawal from chronic methamphetamine. *Behav Pharmacol* 25:12–22. [PubMed: 24281153]
- Hoefler MM, Sanchez AB, Maung R, de Rozieres CM, Catalan IC, Dowling CC, Thaney VE, Piña-Crespo J, Zhang D, Roberts AJ, Kaul M (2015) Combination of methamphetamine and HIV-1 gp120 causes distinct long-term alterations of behavior, gene expression, and injury in the central nervous system. *Experimental Neurology* 263:221–234. [PubMed: 25246228]
- Hoenigl M, Chaillon A, Moore DJ, Morris SR, Smith DM, Little SJ (2016) Clear Links Between Starting Methamphetamine and Increasing Sexual Risk Behavior: A Cohort Study Among Men Who Have Sex With Men. *Journal of acquired immune deficiency syndromes (1999)* 71:551–557. [PubMed: 26536321]
- Holmes AJ, Hollinshead MO, Roffman JL, Smoller JW, Buckner RL (2016) Individual Differences in Cognitive Control Circuit Anatomy Link Sensation Seeking, Impulsivity, and Substance Use. *The Journal of Neuroscience* 36:4038. [PubMed: 27053210]
- Iudicello JE, Morgan EE, Gongvatana A, Letendre SL, Grant I, Woods SP (2014) Detrimental impact of remote methamphetamine dependence on neurocognitive and everyday functioning in older but not younger HIV+ adults: evidence for a legacy effect? *Journal of neurovirology* 20:85–98. [PubMed: 24470237]
- Iwakura Y, Shioda T, Tosu M, Yoshida E, Hayashi M, Nagata T, Shibuta H (1992) The induction of cataracts by HIV-1 in transgenic mice. *AIDS* 6:1069–1075. [PubMed: 1466838]
- Janabi N (2002) Selective inhibition of cyclooxygenase-2 expression by 15-deoxy-Delta(12,14) (12,14)-prostaglandin J(2) in activated human astrocytes, but not in human brain macrophages. *J Immunol* 168:4747–4755. [PubMed: 11971025]
- Jernigan TL, Gamst AC, Archibald SL, Fennema-Notestine C, Mindt MR, Marcotte TD, Heaton RK, Ellis RJ, Grant I (2005) Effects of methamphetamine dependence and HIV infection on cerebral morphology. *The American journal of psychiatry* 162:1461–1472. [PubMed: 16055767]
- Jia P et al. (2017) Genome-wide association study of HIV-associated neurocognitive disorder (HAND): A CHARTER group study. *Am J Med Genet B Neuropsychiatr Genet* 174:413–426. [PubMed: 28447399]
- Johanson C-E, Frey KA, Lundahl LH, Keenan P, Lockhart N, Roll J, Galloway GP, Koeppe RA, Kilbourn MR, Robbins T (2006) Cognitive function and nigrostriatal markers in abstinent methamphetamine abusers. *Psychopharmacology* 185:327–338. [PubMed: 16518646]
- Kaul M, Zheng J, Okamoto S, Gendelman HE, Lipton SA (2005) HIV-1 infection and AIDS: consequences for the central nervous system. *Cell Death & Differentiation* 12:878–892. [PubMed: 15832177]
- Kaul M, Ma Q, Medders KE, Desai MK, Lipton SA (2007) HIV-1 coreceptors CCR5 and CXCR4 both mediate neuronal cell death but CCR5 paradoxically can also contribute to protection. *Cell Death & Differentiation* 14:296–305. [PubMed: 16841089]

- Keppeler OT, Welte FJ, Ngo TA, Chin PS, Patton KS, Tsou CL, Abbey NW, Sharkey ME, Grant RM, You Y, Scarborough JD, Ellmeier W, Littman DR, Stevenson M, Charo IF, Herndier BG, Speck RF, Goldsmith MA (2002) Progress toward a human CD4/CCR5 transgenic rat model for de novo infection by human immunodeficiency virus type 1. *J Exp Med* 195:719–736. [PubMed: 11901198]
- Kesby JP, Hubbard DT, Markou A, Semenova S (2014) Expression of HIV gp120 protein increases sensitivity to the rewarding properties of methamphetamine in mice. *Addict Biol* 19:593–605. [PubMed: 23252824]
- Kesby JP, Fields JA, Chang A, Coban H, Achim CL, Semenova S (2018) Effects of HIV-1 TAT protein and methamphetamine exposure on visual discrimination and executive function in mice. *Behavioural Brain Research* 349:73–79. [PubMed: 29709610]
- Kesby JP, Heaton RK, Young JW, Umlauf A, Woods SP, Letendre SL, Markou A, Grant I, Semenova S (2015) Methamphetamine Exposure Combined with HIV-1 Disease or gp120 Expression: Comparison of Learning and Executive Functions in Humans and Mice. *Neuropsychopharmacology* 40:1899–1909. [PubMed: 25652249]
- Kesby JP, Najera JA, Romoli B, Fang Y, Basova L, Birmingham A, Marcondes MCG, Dulcis D, Semenova S (2017) HIV-1 TAT protein enhances sensitization to methamphetamine by affecting dopaminergic function. *Brain Behav Immun* 65:210–221. [PubMed: 28495611]
- Khadka S, Narayanan B, Meda SA, Gelernter J, Han S, Sawyer B, Aslanzadeh F, Stevens MC, Hawkins KA, Anticevic A, Potenza MN, Pearson GD (2014) Genetic association of impulsivity in young adults: a multivariate study. *Translational Psychiatry* 4:e451–e451. [PubMed: 25268255]
- Khakh BS, Sofroniew MV (2015) Diversity of astrocyte functions and phenotypes in neural circuits. *Nat Neurosci* 18:942–952. [PubMed: 26108722]
- Kim Y-K, Shin C (2018) The Microbiota-Gut-Brain Axis in Neuropsychiatric Disorders: Pathophysiological Mechanisms and Novel Treatments. *Curr Neuroparmacol* 16:559–573. [PubMed: 28925886]
- Klotman PE, Notkins AL (1996) Transgenic models of human immunodeficiency virus type-1. *Curr Top Microbiol Immunol* 206:197–222. [PubMed: 8608718]
- Ko A, Kang G, Hattler JB, Galadima HI, Zhang J, Li Q, Kim W-K (2019) Macrophages but not Astrocytes Harbor HIV DNA in the Brains of HIV-1-Infected Aviremic Individuals on Suppressive Antiretroviral Therapy. *Journal of neuroimmune pharmacology : the official journal of the Society on NeuroImmune Pharmacology* 14:110–119. [PubMed: 30194646]
- Kohn M, Link J, Dennis LE, McCreedy H, Huckans M, Hoffman WF, Loftis JM (2019) Neuroinflammation in addiction: A review of neuroimaging studies and potential immunotherapies. *Pharmacol Biochem Behav* 179:34–42. [PubMed: 30695700]
- Kong D-H, Kim YK, Kim MR, Jang JH, Lee S (2018) Emerging Roles of Vascular Cell Adhesion Molecule-1 (VCAM-1) in Immunological Disorders and Cancer. *Int J Mol Sci* 19:1057.
- Kousik S, Napier TC, Carvey P (2012) The Effects of Psychostimulant Drugs on Blood Brain Barrier Function and Neuroinflammation. *Frontiers in Pharmacology* 3.
- Kozela E, Juknat A, Vogel Z (2017) Modulation of Astrocyte Activity by Cannabidiol, a Nonpsychoactive Cannabinoid. *Int J Mol Sci* 18.
- Krucker T, Toggas SM, Mucke L, Siggins GR (1998) Transgenic mice with cerebral expression of human immunodeficiency virus type-1 coat protein gp120 show divergent changes in short- and long-term potentiation in CA1 hippocampus. *Neuroscience* 83:691–700. [PubMed: 9483553]
- Kumar AM, Fernandez JB, Singer EJ, Commins D, Waldrop-Valverde D, Ownby RL, Kumar M (2009) Human immunodeficiency virus type 1 in the central nervous system leads to decreased dopamine in different regions of postmortem human brains. *Journal of neurovirology* 15:257–274. [PubMed: 19499455]
- Kumar V, Torben W, Kenway CS, Schiro FR, Mohan M (2016) Longitudinal Examination of the Intestinal Lamina Propria Cellular Compartment of Simian Immunodeficiency Virus-Infected Rhesus Macaques Provides Broader and Deeper Insights into the Link between Aberrant MicroRNA Expression and Persistent Immune Activation. *J Virol* 90:5003–5019. [PubMed: 26937033]

- Langford D, Grigorian A, Hurford R, Adame A, Crews L, Masliah E (2004) The role of mitochondrial alterations in the combined toxic effects of human immunodeficiency virus Tat protein and methamphetamine on calbindin positive-neurons. *J Neurovirol* 10:327–337. [PubMed: 15765804]
- Langford D, Adame A, Grigorian A, Grant I, McCutchan JA, Ellis RJ, Marcotte TD, Masliah E (2003) Patterns of selective neuronal damage in methamphetamine-user AIDS patients. *Journal of acquired immune deficiency syndromes (1999)* 34:467–474. [PubMed: 14657756]
- Lee MH, Amin ND, Venkatesan A, Wang T, Tyagi R, Pant HC, Nath A (2013) Impaired neurogenesis and neurite outgrowth in an HIV-gp120 transgenic model is reversed by exercise via BDNF production and Cdk5 regulation. *J Neurovirol* 19:418–431. [PubMed: 23982957]
- Lee MH, Wang T, Jang MH, Steiner J, Haughey N, Ming GL, Song H, Nath A, Venkatesan A (2011) Rescue of adult hippocampal neurogenesis in a mouse model of HIV neurologic disease. *Neurobiol Dis* 41:678–687. [PubMed: 21146610]
- Leonard JM, Abramczuk JW, Pezen DS, Rutledge R, Belcher JH, Hakim F, Shearer G, Lamperth L, Travis W, Fredrickson T, . (1988) Development of disease and virus recovery in transgenic mice containing HIV proviral DNA. *Science* 242:1665–1670. [PubMed: 3201255]
- Li G-H, Henderson L, Nath A (2016) Astrocytes as an HIV Reservoir: Mechanism of HIV Infection. *Curr HIV Res* 14:373–381. [PubMed: 27719663]
- Liddelov SA et al. (2017) Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 541:481–487. [PubMed: 28099414]
- Liu CY, Yang Y, Ju WN, Wang X, Zhang HL (2018) Emerging Roles of Astrocytes in Neuro-Vascular Unit and the Tripartite Synapse With Emphasis on Reactive Gliosis in the Context of Alzheimer's Disease. *Frontiers in cellular neuroscience* 12:193. [PubMed: 30042661]
- Liu X, Chang L, Vigorito M, Kass M, Li H, Chang SL (2009) Methamphetamine-induced behavioral sensitization is enhanced in the HIV-1 transgenic rat. *J Neuroimmune Pharmacol* 4:309–316. [PubMed: 19444617]
- Lloyd SL, Striley CW (2018) Marijuana Use Among Adults 50 Years or Older in the 21st Century. *Gerontology and Geriatric Medicine* 4:2333721418781668. [PubMed: 29977980]
- London ED, Kohno M, Morales AM, Ballard ME (2015) Chronic methamphetamine abuse and corticostriatal deficits revealed by neuroimaging. *Brain research* 1628:174–185. [PubMed: 25451127]
- Lu T-S, Avraham HK, Seng S, Tachado SD, Koziel H, Makriyannis A, Avraham S (2008) Cannabinoids inhibit HIV-1 Gp120-mediated insults in brain microvascular endothelial cells. *J Immunol* 181:6406–6416. [PubMed: 18941231]
- Ma Q, Xing C, Long W, Wang HY, Liu Q, Wang R-F (2019) Impact of microbiota on central nervous system and neurological diseases: the gut-brain axis. *Journal of Neuroinflammation* 16:53. [PubMed: 30823925]
- MacDuffie KE, Brown GG, McKenna BS, Liu TT, Meloy MJ, Tawa B, Archibald S, Fennema-Notestine C, Atkinson JH Jr., Ellis RJ, Letendre SL, Hesselink JR, Cherner M, Grant I (2018) Effects of HIV Infection, methamphetamine dependence and age on cortical thickness, area and volume. *NeuroImage Clinical* 20:1044–1052. [PubMed: 30342393]
- Mackiewicz M, Overk C, Achim CL, Masliah E (2019) Pathogenesis of age-related HIV neurodegeneration. *Journal of neurovirology* 25:622–633. [PubMed: 30790184]
- Madden LJ, Flynn CT, Zandonatti MA, May M, Parsons LH, Katner SN, Henriksen SJ, Fox HS (2005) Modeling human methamphetamine exposure in nonhuman primates: chronic dosing in the rhesus macaque leads to behavioral and physiological abnormalities. *Neuropsychopharmacol ogy* 30:350–359.
- Mahajan SD, Aalinkeel R, Sykes DE, Reynolds JL, Bindukumar B, Adal A, Qi M, Toh J, Xu G, Prasad PN, Schwartz SA (2008) Methamphetamine alters blood brain barrier permeability via the modulation of tight junction expression: Implication for HIV-1 neuropathogenesis in the context of drug abuse. *Brain research* 1203:133–148. [PubMed: 18329007]
- Marban C, Forouzanfar F, Ait-Ammar A, Fahmi F, El Mekdad H, Daouad F, Rohr O, Schwartz C (2016) Targeting the Brain Reservoirs: Toward an HIV Cure. *Frontiers in Immunology* 7:397. [PubMed: 27746784]

- Marcondes MC, Flynn C, Watry DD, Zandonatti M, Fox HS (2010) Methamphetamine increases brain viral load and activates natural killer cells in simian immunodeficiency virus-infected monkeys. *Am J Pathol* 177:355–361. [PubMed: 20489154]
- Marcotte TD, Wolfson T, Rosenthal TJ, Heaton RK, Gonzalez R, Ellis RJ, Grant I (2004) A multimodal assessment of driving performance in HIV infection. *Neurology* 63:1417–1422. [PubMed: 15505158]
- Masters MC, Ances BM (2014) Role of neuroimaging in HIV-associated neurocognitive disorders. *Semin Neurol* 34:89–102. [PubMed: 24715492]
- Maung R, Hoefler MM, Sanchez AB, Sejbuk NE, Medders KE, Desai MK, Catalan IC, Dowling CC, de Rozieres CM, Garden GA, Russo R, Roberts AJ, Williams R, Kaul M (2014) CCR5 knockout prevents neuronal injury and behavioral impairment induced in a transgenic mouse model by a CXCR4-using HIV-1 glycoprotein 120. *J Immunol* 193:1895–1910. [PubMed: 25031461]
- McKenna BS, Brown GG, Archibald S, Scadeng M, Bussell R, Kesby JP, Markou A, Soontornniyomkij V, Achim C, Semenova S, Translational Methamphetamine Aids Research Center Tmarc G (2016) Microstructural changes to the brain of mice after methamphetamine exposure as identified with diffusion tensor imaging. *Psychiatry Res Neuroimaging* 249:27–37. [PubMed: 27000304]
- Meckel KR, Kiraly DD (2019) A potential role for the gut microbiome in substance use disorders. *Psychopharmacology* 236:1513–1530. [PubMed: 30982128]
- Mediouni S, Marcondes MC, Miller C, McLaughlin JP, Valente ST (2015) The cross-talk of HIV-1 Tat and methamphetamine in HIV-associated neurocognitive disorders. *Front Microbiol* 6:1164. [PubMed: 26557111]
- Minagar A, Shapshak P, Fujimura R, Ownby R, Heyes M, Eisdorfer C (2002) The role of macrophage/microglia and astrocytes in the pathogenesis of three neurologic disorders: HIV-associated dementia, Alzheimer disease, and multiple sclerosis. *J Neurol Sci* 202:13–23. [PubMed: 12220687]
- Minassian A, Henry BL, Woods SP, Vaida F, Grant I, Geyer MA, Perry W, Translational Methamphetamine ARCG (2013) Prepulse inhibition in HIV-associated neurocognitive disorders. *Journal of the International Neuropsychological Society : JINS* 19:709–717. [PubMed: 23552464]
- Molina PE, Amedee AM, Winsauer P, Nelson S, Bagby G, Simon L (2015) Behavioral, Metabolic, and Immune Consequences of Chronic Alcohol or Cannabinoids on HIV/AIDS: Studies in the Non-Human Primate SIV Model. *J Neuroimmune Pharmacol* 10:217–232. [PubMed: 25795088]
- Montgomery L, Bagot K, Brown JL, Haeny AM (2019) The Association Between Marijuana Use and HIV Continuum of Care Outcomes: a Systematic Review. *Curr HIV/AIDS Rep* 16:17–28. [PubMed: 30671919]
- Moore DJ, Montoya JL, Casaletto KB, Hampton Atkinson J (2018a) Medication Adherence and HIV-Associated Neurocognitive Disorders (HAND) In: *Encyclopedia of AIDS* (Hope TJ, Richman DD, Stevenson M, eds), pp 1312–1318. New York, NY: Springer New York.
- Moore DJ, Pasipanodya EC, Umlauf A, Rooney AS, Gouaux B, Depp CA, Atkinson JH, Montoya JL (2018b) Individualized texting for adherence building (iTAB) for methamphetamine users living with HIV: A pilot randomized clinical trial. *Drug and alcohol dependence* 189:154–160. [PubMed: 29958127]
- Moore DJ, Blackstone K, Woods SP, Ellis RJ, Atkinson JH, Heaton RK, Grant I, the HG, the TG (2012) Methamphetamine use and neuropsychiatric factors are associated with antiretroviral non-adherence. *AIDS Care* 24:1504–1513. [PubMed: 22530794]
- Moran LM, Booze RM, Webb KM, Mactutus CF (2013) Neurobehavioral alterations in HIV-1 transgenic rats: evidence for dopaminergic dysfunction. *Exp Neurol* 239:139–147. [PubMed: 23063600]
- Moran LM, Aksenov MY, Booze RM, Webb KM, Mactutus CF (2012) Adolescent HIV-1 transgenic rats: evidence for dopaminergic alterations in behavior and neurochemistry revealed by methamphetamine challenge. *Curr HIV Res* 10:415–424. [PubMed: 22591365]

- Morgan EE, Woods SP, Delano-Wood L, Bondi MW, Grant I, Group HIVNRP (2011) Intraindividual variability in HIV infection: evidence for greater neurocognitive dispersion in older HIV seropositive adults. *Neuropsychology* 25:645–654. [PubMed: 21574712]
- Moszczynska A, Callan SP (2017) Molecular, Behavioral, and Physiological Consequences of Methamphetamine Neurotoxicity: Implications for Treatment. *The Journal of pharmacology and experimental therapeutics* 362:474–488. [PubMed: 28630283]
- Muccioli GG, Naslain D, Bäckhed F, Reigstad CS, Lambert DM, Delzenne NM, Cani PD (2010) The endocannabinoid system links gut microbiota to adipogenesis. *Mol Syst Biol* 6:392–392. [PubMed: 20664638]
- Muratori C, Mangino G, Affabris E, Federico M (2010) Astrocytes contacting HIV-1-infected macrophages increase the release of CCL2 in response to the HIV-1-dependent enhancement of membrane-associated TNF α in macrophages. *Glia* 58:1893–1904. [PubMed: 20737475]
- Nair MPN, Mahajan S, Sykes D, Bapardekar MV, Reynolds JL (2006) Methamphetamine modulates DC-SIGN expression by mature dendritic cells. *Journal of Neuroimmune Pharmacology* 1:296–304. [PubMed: 18040806]
- Najera JA, Bustamante EA, Bortell N, Morsey B, Fox HS, Ravasi T, Marcondes MC (2016) Methamphetamine abuse affects gene expression in brain-derived microglia of SIV-infected macaques to enhance inflammation and promote virus targets. *BMC Immunol* 17:7. [PubMed: 27107567]
- Nakanishi N, Kang Y-J, Tu S, McKercher SR, Masliah E, Lipton SA (2016) Differential Effects of Pharmacologic and Genetic Modulation of NMDA Receptor Activity on HIV/gp120-Induced Neuronal Damage in an In Vivo Mouse Model. *J Mol Neurosci* 58:59–65. [PubMed: 26374431]
- Nath BM, Schumann KE, Boyer JD (2000) The chimpanzee and other non-human-primate models in HIV-1 vaccine research. *Trends Microbiol* 8:426–431. [PubMed: 10989311]
- National Institute on Drug Abuse (2020) Overdose Death Rates. In: National Institute on Drug Abuse website.
- Nolan R, Gaskill PJ (2019) The role of catecholamines in HIV neuropathogenesis. *Brain research* 1702:54–73. [PubMed: 29705605]
- O'Sullivan SE, Kendall DA (2010) Cannabinoid activation of peroxisome proliferator-activated receptors: potential for modulation of inflammatory disease. *Immunobiology* 215:611–616. [PubMed: 19833407]
- O'Sullivan SE, Kendall DA, Randall MD (2009) Time-dependent vascular effects of Endocannabinoids mediated by peroxisome proliferator-activated receptor gamma (PPARgamma). *PPAR Res* 2009:425289. [PubMed: 19421417]
- O'Sullivan SE, Tarling EJ, Bennett AJ, Kendall DA, Randall MD (2005) Novel time-dependent vascular actions of Delta9-tetrahydrocannabinol mediated by peroxisome proliferator-activated receptor gamma. *Biochem Biophys Res Commun* 337:824–831. [PubMed: 16213464]
- Okamoto S, Kang YJ, Brechtel CW, Siviglia E, Russo R, Clemente A, Harrop A, McKercher S, Kaul M, Lipton SA (2007) HIV/gp120 decreases adult neural progenitor cell proliferation via checkpoint kinase-mediated cell-cycle withdrawal and G1 arrest. *Cell Stem Cell* 1:230–236. [PubMed: 18371353]
- Olivier IS, Cacabelos R, Naidoo V (2018) Risk Factors and Pathogenesis of HIV-Associated Neurocognitive Disorder: The Role of Host Genetics. *Int J Mol Sci* 19:3594.
- Olmsted RA, Barnes AK, Yamamoto JK, Hirsch VM, Purcell RH, Johnson PR (1989) Molecular cloning of feline immunodeficiency virus. *Proc Natl Acad Sci U S A* 86:2448–2452. [PubMed: 2928341]
- Paolillo EW, Obermeit LC, Tang B, Depp CA, Vaida F, Moore DJ, Moore RC (2018) Smartphone-based ecological momentary assessment (EMA) of alcohol and cannabis use in older adults with and without HIV infection. *Addict Behav* 83:102–108. [PubMed: 29126667]
- Paolillo EW, Saloner R, Montoya JL, Campbell LM, Pasipanodya EC, Iudicello JE, Moore RC, Letendre SL, Jeste DV, Moore DJ (2019) Frailty in Comorbid HIV and Lifetime Methamphetamine Use Disorder: Associations with Neurocognitive and Everyday Functioning. *AIDS research and human retroviruses* 35:1044–1053. [PubMed: 31303012]

- Papageorgiou M, Raza A, Fraser S, Nurgali K, Apostolopoulos V (2019) Methamphetamine and its immune-modulating effects. *Maturitas* 121:13–21. [PubMed: 30704560]
- Pautz A, Art J, Hahn S, Nowag S, Voss C, Kleinert H (2010) Regulation of the expression of inducible nitric oxide synthase. *Nitric Oxide* 23:75–93. [PubMed: 20438856]
- Pearson-Fuhrhop KM, Dunn EC, Mortero S, Devan WJ, Falcone GJ, Lee P, Holmes AJ, Hollinshead MO, Roffman JL, Smoller JW, Rosand J, Cramer SC (2014) Dopamine genetic risk score predicts depressive symptoms in healthy adults and adults with depression. *PLoS One* 9:e93772–e93772. [PubMed: 24834916]
- Pellegrini C, Antonioli L, Colucci R, Blandizzi C, Fornai M (2018) Interplay among gut microbiota, intestinal mucosal barrier and enteric neuro-immune system: a common path to neurodegenerative diseases? *Acta Neuropathologica* 136:345–361. [PubMed: 29797112]
- Perry W, Minassian A, Paulus MP, Young JW, Kincaid MJ, Ferguson EJ, Henry BL, Zhuang X, Masten VL, Sharp RF, Geyer MA (2009) A reverse-translational study of dysfunctional exploration in psychiatric disorders: from mice to men. *Archives of general psychiatry* 66:1072–1080. [PubMed: 19805697]
- Persidsky Y, Ramirez SH, Haorah J, Kanmogne GD (2006) Blood–brain Barrier: Structural Components and Function Under Physiologic and Pathologic Conditions. *Journal of Neuroimmune Pharmacology* 1:223–236. [PubMed: 18040800]
- Persidsky Y, Ho W, Ramirez SH, Potula R, Abood ME, Unterwald E, Tuma R (2011) HIV-1 infection and alcohol abuse: neurocognitive impairment, mechanisms of neurodegeneration and therapeutic interventions. *Brain Behav Immun* 25 Suppl 1:S61–S70. [PubMed: 21397004]
- Presley C, Abidi A, Suryawanshi S, Mustafa S, Meibohm B, Moore Ii BM (2015) Preclinical evaluation of SMM-189, a cannabinoid receptor 2-specific inverse agonist. *Pharmacology Research & Perspectives* 3:e00159. [PubMed: 26196013]
- Prud'homme M, Cata R, Jutras-Aswad D (2015) Cannabidiol as an Intervention for Addictive Behaviors: A Systematic Review of the Evidence. *Subst Abuse* 9:33–38.
- Ragin AB, Wu Y, Gao Y, Keating S, Du H, Sammet C, Kettering CS, Epstein LG (2015) Brain alterations within the first 100 days of HIV infection. *Annals of Clinical and Translational Neurology* 2:12–21. [PubMed: 25642430]
- Rahimy E, Li F-Y, Hagberg L, Fuchs D, Robertson K, Meyerhoff DJ, Zetterberg H, Price RW, Gisslen M, Spudich S (2017) Blood-Brain Barrier Disruption Is Initiated During Primary HIV Infection and Not Rapidly Altered by Antiretroviral Therapy. *The Journal of Infectious Diseases* 215:1132–1140. [PubMed: 28368497]
- Ramirez SH, Potula R, Fan S, Eidem T, Papugani A, Reichenbach N, Dykstra H, Weksler BB, Romero IA, Couraud PO, Persidsky Y (2009) Methamphetamine disrupts blood-brain barrier function by induction of oxidative stress in brain endothelial cells. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* 29:1933–1945.
- Reid W et al. (2001) An HIV-1 transgenic rat that develops HIV-related pathology and immunologic dysfunction. *Proc Natl Acad Sci U S A* 98:9271–9276. [PubMed: 11481487]
- Reiner A, Heldt SA, Presley CS, Guley NH, Elberger AJ, Deng Y, D'Surney L, Rogers JT, Ferrell J, Bu W, Del Mar N, Honig MG, Gurley SN, Moore BM 2nd (2014) Motor, visual and emotional deficits in mice after closed-head mild traumatic brain injury are alleviated by the novel CB2 inverse agonist SMM-189. *Int J Mol Sci* 16:758–787. [PubMed: 25561230]
- Rippeth JD, Heaton RK, Carey CL, Marcotte TD, Moore DJ, Gonzalez R, Wolfson T, Grant I, Group TH (2004) Methamphetamine dependence increases risk of neuropsychological impairment in HIV infected persons. *Journal of the International Neuropsychological Society* 10:1–14. [PubMed: 14751002]
- Rizzo MD, Crawford RB, Bach A, Sermet S, Amalfitano A, Kaminski NE (2019) Delta(9)-Tetrahydrocannabinol Suppresses Monocyte-Mediated Astrocyte Production of Monocyte Chemoattractant Protein 1 and Interleukin-6 in a Toll-Like Receptor 7-Stimulated Human Coculture. *J Pharmacol Exp Ther* 371:191–201. [PubMed: 31383729]

- Rizzo MD, Crawford RB, Henriquez JE, Aldhamen YA, Gulick P, Amalfitano A, Kaminski NE (2018) HIV-infected cannabis users have lower circulating CD16+ monocytes and IFN- γ -inducible protein 10 levels compared with nonusing HIV patients. *AIDS* 32:419–429. [PubMed: 29194121]
- Roberts AJ, Maung R, Sejbuk NE, Ake C, Kaul M (2010) Alteration of Methamphetamine-induced stereotypic behaviour in transgenic mice expressing HIV-1 envelope protein gp120. *J Neurosci Methods* 186:222–225. [PubMed: 19917310]
- Rom S, Persidsky Y (2013) Cannabinoid receptor 2: potential role in immunomodulation and neuroinflammation. *Journal of neuroimmune pharmacology : the official journal of the Society on Neuroimmune Pharmacology* 8:608–620. [PubMed: 23471521]
- Rueda D, Galve-Roperh I, Haro A, Guzman M (2000) The CB(1) cannabinoid receptor is coupled to the activation of c-Jun N-terminal kinase. *Mol Pharmacol* 58:814–820. [PubMed: 10999952]
- Saloner R, Cysique LA (2017) HIV-Associated Neurocognitive Disorders: A Global Perspective. *Journal of the International Neuropsychological Society : JINS* 23:860–869. [PubMed: 29198283]
- Saloner R, Paolillo EW, Kohli M, Murray SS, Moore DJ, Grant I, Cherner M (2020) Genetic variation in alcohol dehydrogenase is associated with neurocognition in men with HIV and history of alcohol use disorder: preliminary findings. *Journal of neurovirology*.
- Saloner R, Heaton RK, Campbell LM, Chen A, Franklin D Jr., Ellis RJ, Collier AC, Marra C, Clifford DB, Gelman B, Sacktor N, Morgello S, McCutchan JA, Letendre S, Grant I, Fennema-Notestine C (2019a) Effects of comorbidity burden and age on brain integrity in HIV. *AIDS* 33:1175–1185. [PubMed: 30870195]
- Saloner R et al. (2019b) Neurocognitive Super Aging in Older Adults Living With HIV: Demographic, Neuromedical and Everyday Functioning Correlates. *Journal of the International Neuropsychological Society : JINS* 25:507–519. [PubMed: 30890191]
- Schrier RD, Hong S, Crescini M, Ellis R, Perez-Santiago J, Spina C, Letendre S (2015) Cerebrospinal fluid (CSF) CD8+ T-cells that express interferon-gamma contribute to HIV associated neurocognitive disorders (HAND). *PLoS One* 10:e0116526. [PubMed: 25719800]
- Schweinsburg BC, Taylor MJ, Alhassoon OM, Gonzalez R, Brown GG, Ellis RJ, Letendre S, Videen JS, McCutchan JA, Patterson TL, Grant I, Group H (2005) Brain mitochondrial injury in human immunodeficiency virus-seropositive (HIV+) individuals taking nucleoside reverse transcriptase inhibitors. *J Neurovirol* 11:356–364. [PubMed: 16206458]
- Scott EP, Brennan E, Benitez A (2019) A Systematic Review of the Neurocognitive Effects of Cannabis Use in Older Adults. *Curr Addict Rep* 6:443–455. [PubMed: 32477850]
- Simon L, Song K, Vande Stouwe C, Hollenbach A, Amedee A, Mohan M, Winsauer P, Molina P (2016) Delta9-Tetrahydrocannabinol (Delta9-THC) Promotes Neuroimmune-Modulatory MicroRNA Profile in Striatum of Simian Immunodeficiency Virus (SIV)-Infected Macaques. *J Neuroimmune Pharmacol* 11:192–213. [PubMed: 26607731]
- Singh RK, Chang H-W, Yan D, Lee KM, Ucmak D, Wong K, Abrouk M, Farahnik B, Nakamura M, Zhu TH, Bhutani T, Liao W (2017) Influence of diet on the gut microbiome and implications for human health. *Journal of Translational Medicine* 15:73. [PubMed: 28388917]
- Sofroniew MV (2000) Astrocyte failure as a cause of CNS dysfunction. *Mol Psychiatry* 5:230–232. [PubMed: 10889520]
- Sofroniew MV, Vinters HV (2010) Astrocytes: biology and pathology. *Acta Neuropathol* 119:7–35. [PubMed: 20012068]
- Soontornniyomkij V, Kesby JP, Morgan EE, Bischoff-Grethe A, Minassian A, Brown GG, Grant I (2016a) Effects of HIV and Methamphetamine on Brain and Behavior: Evidence from Human Studies and Animal Models. *Journal of neuroimmune pharmacology : the official journal of the Society on Neuroimmune Pharmacology* 11:495–510. [PubMed: 27484318]
- Soontornniyomkij V, Umlauf A, Soontornniyomkij B, Batki IB, Moore DJ, Masliah E, Achim CL (2016b) Lifetime methamphetamine dependence is associated with cerebral microgliosis in HIV-1-infected adults. *Journal of neurovirology* 22:650–660. [PubMed: 27098516]
- St Hillaire C, Vargas D, Pardo CA, Gincel D, Mann J, Rothstein JD, McArthur JC, Conant K (2005) Aquaporin 4 is increased in association with human immunodeficiency virus dementia:

implications for disease pathogenesis. *Journal of neurovirology* 11:535–543. [PubMed: 16338747]

Steiner JP, Bachani M, Wolfson-Stofko B, Lee MH, Wang T, Li G, Li W, Strayer D, Haughey NJ, Nath A (2015) Interaction of paroxetine with mitochondrial proteins mediates neuroprotection. *Neurotherapeutics* 12:200–216. [PubMed: 25404050]

Substance Abuse and Mental Health Services Administration (2019) Key substance use and mental health indicators in the United States: Results from the 2018 National Survey on Drug Use and Health. In. Rockville, MD: Center for Behavioral Health Statistics and Quality, Substance Abuse and Mental Health Services Administration.

Sundermann EE, Hussain MA, Moore DJ, Horvath S, Lin DTS, Kobor MS, Levine A (2019) Inflammation-related genes are associated with epigenetic aging in HIV. *Journal of neurovirology* 25:853–865. [PubMed: 31286441]

Swinton MK, Carson A, Telese F, Sanchez AB, Soontornniyomkij B, Rad L, Batki I, Quintanilla B, Perez-Santiago J, Achim CL, Letendre S, Ellis RJ, Grant I, Murphy AN, Fields JA (2019) Mitochondrial biogenesis is altered in HIV+ brains exposed to ART: Implications for therapeutic targeting of astroglia. *Neurobiol Dis* 130:104502. [PubMed: 31238091]

Takahashi H, Xia P, Cui J, Talantova M, Bodhinathan K, Li W, Saleem S, Holland EA, Tong G, Pina-Crespo J, Zhang D, Nakanishi N, Larrick JW, McKercher SR, Nakamura T, Wang Y, Lipton SA (2015) Pharmacologically targeted NMDA receptor antagonism by NitroMemantine for cerebrovascular disease. *Scientific Reports* 5:14781. [PubMed: 26477507]

Takeda S, Ikeda E, Su S, Harada M, Okazaki H, Yoshioka Y, Nishimura H, Ishii H, Kakizoe K, Taniguchi A, Tokuyasu M, Himeno T, Watanabe K, Omiecinski CJ, Aramaki H (2014) Delta(9)-THC modulation of fatty acid 2-hydroxylase (FA2H) gene expression: possible involvement of induced levels of PPARalpha in MDA-MB-231 breast cancer cells. *Toxicology* 326:18–24. [PubMed: 25291031]

Tang Y, Le W (2016) Differential Roles of M1 and M2 Microglia in Neurodegenerative Diseases. *Molecular neurobiology* 53:1181–1194. [PubMed: 25598354]

Taylor MJ, Schweinsburg BC, Alhassoon OM, Gongvatana A, Brown GG, Young-Casey C, Letendre SL, Grant I (2007) Effects of human immunodeficiency vims and methamphetamine on cerebral metabolites measured with magnetic resonance spectroscopy. *Journal of neurovirology* 13:150–159. [PubMed: 17505983]

Teodorof-Diedrich C, Spector SA (2018) Human Immunodeficiency Vims Type 1 gp120 and Tat Induce Mitochondrial Fragmentation and Incomplete Mitophagy in Human Neurons. *J Virol* 92:e00993–00918. [PubMed: 30158296]

Thames AD, Mahmood Z, Burggren AC, Karimian A, Kuhn T (2016) Combined Effects of HIV and Marijuana Use on Neurocognitive Functioning and Immune Status. *AIDS care* 28:628–632. [PubMed: 26694807]

Thames AD, Kuhn TP, Williamson TJ, Jones JD, Mahmood Z, Hammond A (2017) Marijuana effects on changes in brain structure and cognitive function among HIV+ and HIV–adults. *Drug Alcohol Depend* 170:120–127. [PubMed: 27889592]

Thaney VE, O'Neill AM, Hoefler MM, Maung R, Sanchez AB, Kaul M (2017) IFNβ Protects Neurons from Damage in a Murine Model of HIV-1 Associated Brain Injury. *Scientific Reports* 7:46514. [PubMed: 28425451]

Thaney VE, Sanchez AB, Fields JA, Minassian A, Young JW, Maung R, Kaul M (2018) Transgenic mice expressing HIV-1 envelope protein gp120 in the brain as an animal model in neuroAIDS research. *JNeurovirol* 24:156–167. [PubMed: 29075998]

Toggas SM, Mucke L (1996) Transgenic models in the study of AIDS dementia complex. *Curr Top Microbiol Immunol* 206:223–241. [PubMed: 8608719]

Toggas SM, Masliah E, Rockenstein EM, Rall GF, Abraham CR, Mucke L (1994) Central nervous system damage produced by expression of the HIV-1 coat protein gp120 in transgenic mice. *Nature* 367:188–193. [PubMed: 8114918]

Valcour V, Chalermchai T, Sailasuta N, Marovich M, Lerdlum S, Suttichom D, Suwanwela NC, Jagodzinski L, Michael N, Spudich S, van Griensven F, de Souza M, Kim J, Ananworanich J,

- Group RSS (2012) Central nervous system viral invasion and inflammation during acute HIV infection. *The Journal of infectious diseases* 206:275–282. [PubMed: 22551810]
- Van Duyne R, Pedati C, Guendel I, Carpio L, KeHN-Hall K, Saifuddin M, Kashanchi F (2009) The utilization of humanized mouse models for the study of human retroviral infections. *Retrovirology* 6:76. [PubMed: 19674458]
- Var SR, Day TRC, Vitomirov A, Smith DM, Soontornniyomkij V, Moore DJ, Achim CL, Mehta SR, Perez-Santiago J (2016) Mitochondrial injury and cognitive function in HIV infection and methamphetamine use. *AIDS* 30:839–848. [PubMed: 26807965]
- Vera JH, Guo Q, Cole JH, Boasso A, Greathead L, Kelleher P, Rabiner EA, Kalk N, Bishop C, Gunn RN, Matthews PM, Winston A (2016) Neuroinflammation in treated HIV-positive individuals: A TSPO PET study. *Neurology* 86:1425–1432. [PubMed: 26911637]
- Volkow ND, Koob GF, McLellan AT (2016) Neurobiologic Advances from the Brain Disease Model of Addiction. *New England Journal of Medicine* 374:363–371. [PubMed: 26816013]
- Volpe GE, Ward H, Mwamburi M, Dinh D, Bhalchandra S, Wanke C, Kane AV (2014) Associations of cocaine use and HIV infection with the intestinal microbiota, microbial translocation, and inflammation. *J Stud Alcohol Drugs* 75:347–357. [PubMed: 24650829]
- Wang C, Liu B, Zhang X, Cui Y, Yu C, Jiang T (2018) Multilocus genetic profile in dopaminergic pathway modulates the striatum and working memory. *Scientific Reports* 8:5372. [PubMed: 29599495]
- Watson CW, Paolillo EW, Morgan EE, Umlauf A, Sundermann EE, Ellis RJ, Letendre S, Marcotte TD, Heaton RK, Grant I (2020) Cannabis Exposure is Associated With a Lower Likelihood of Neurocognitive Impairment in People Living With HIV. *J Acquir Immune Defic Syndr* 83:56–64. [PubMed: 31809361]
- Williams R, Bokhari S, Silverstein P, Pinson D, Kumar A, Buch S (2008) Nonhuman primate models of NeuroAIDS. *J Neurovirol* 14:292–300. [PubMed: 18780230]
- Wilson ME, Dimayuga FO, Reed JL, Curry TE, Anderson CF, Nath A, Bruce-Keller AJ (2006) Immune modulation by estrogens: role in CNS HIV-1 infection. *Endocrine* 29:289–297. [PubMed: 16785604]
- Winsauer PJ, Molina PE, Amedee AM, Filipeanu CM, McGoey RR, Troclair DA, Walker EM, Birke LL, Stouwe CV, Howard JM, Leonard ST, Moerschbaecher JM, Lewis PB (2011) Tolerance to chronic delta-9-tetrahydrocannabinol (Delta-THC) in rhesus macaques infected with simian immunodeficiency virus. *Exp Clin Psychopharmacol* 19:154–172. [PubMed: 21463073]
- Woods SP, Moore DJ, Weber E, Grant I (2009) Cognitive Neuropsychology of HIV-Associated Neurocognitive Disorders. *Neuropsychology Review* 19:152–168. [PubMed: 19462243]
- Wright PW, Vaida FF, Fernández RJ, Rutlin J, Price RW, Lee E, Peterson J, Fuchs D, Shimony JS, Robertson KR, Walter R, Meyerhoff DJ, Spudich S, Ances BM (2015) Cerebral white matter integrity during primary HIV infection. *AIDS* 29:433–442. [PubMed: 25513818]
- Xing HQ, Hayakawa H, Gelpi E, Kubota R, Budka H, Izumo S (2009) Reduced expression of excitatory amino acid transporter 2 and diffuse microglial activation in the cerebral cortex in AIDS cases with or without HIV encephalitis. *Journal of Neuropathology & Experimental Neurology* 68:199–209. [PubMed: 19151621]
- Xu J, Chavis JA, Racke MK, Drew PD (2006) Peroxisome proliferator-activated receptor-alpha and retinoid X receptor agonists inhibit inflammatory responses of astrocytes. *J Neuroimmunol* 176:95–105. [PubMed: 16764943]
- Xu P, Wang Y, Qin Z, Qiu L, Zhang M, Huang Y, Zheng JC (2017) Combined Medication of Antiretroviral Drugs Tenofovir Disoproxil Fumarate, Emtricitabine, and Raltegravir Reduces Neural Progenitor Cell Proliferation In Vivo and In Vitro. *J Neuroimmune Pharmacol* 12:682–692. [PubMed: 28735382]
- Yao H, Yang Y, Kim KJ, Bethel-Brown C, Gong N, Funa K, Gendelman HE, Su T-P, Wang JQ, Buch S (2010) Molecular mechanisms involving sigma receptor-mediated induction of MCP-1: implication for increased monocyte transmigration. *Blood* 115:4951–4962. [PubMed: 20354174]
- Yao H, Kim K, Duan M, Hayashi T, Guo M, Morgello S, Prat A, Wang J, Su T-P, Buch S (2011) Cocaine hijacks σ 1 receptor to initiate induction of activated leukocyte cell adhesion molecule:

implication for increased monocyte adhesion and migration in the CNS. *J Neurosci* 31:5942–5955. [PubMed: 21508219]

- Yoo BB, Mazmanian SK (2017) The Enteric Network: Interactions between the Immune and Nervous Systems of the Gut. *Immunity* 46:910–926. [PubMed: 28636959]
- Yoo HB, DiMuzio J, Filbey FM (2020) Interaction of Cannabis Use and Aging: From Molecule to Mind. *Journal of dual diagnosis* 16:140–176. [PubMed: 31570066]
- Young JW, Minassian A, Geyer MA (2016) Locomotor Profiling from Rodents to the Clinic and Back Again. *Curr Top Behav Neurosci* 28:287–303. [PubMed: 27418071]
- Young JW, Powell SB, Geyer MA, Jeste DV, Risbrough VB (2010) The mouse attentional-set-shifting task: a method for assaying successful cognitive aging? *Cognitive, affective & behavioral neuroscience* 10:243–251.
- Yu C, Seaton M, Letendre S, Heaton R, Al-Harhi L (2017a) Plasma dickkopf-related protein 1, an antagonist of the Wnt pathway, is associated with HIV-associated neurocognitive impairment. *AIDS* 31:1379–1385. [PubMed: 28358733]
- Yu C, Narasipura SD, Richards MH, Hu X-T, Yamamoto B, Al-Harhi L (2017b) HIV and drug abuse mediate astrocyte senescence in a β -catenin-dependent manner leading to neuronal toxicity. *Aging Cell* 16:956–965. [PubMed: 28612507]
- Zeng X-F, Li Q, Li J, Wong N, Li Z, Huang J, Yang G, Sham PC, Li S-B, Lu G (2018) HIV-1 Tat and methamphetamine co-induced oxidative cellular injury is mitigated by N-acetylcysteine amide (NACA) through rectifying mTOR signaling. *Toxicol Lett* 299:159–171. [PubMed: 30261225]
- Zulu SS, Simola N, Mabandla MV, Daniels WMU (2018) Effect of long-term administration of antiretroviral drugs (Tenofovir and Nevirapine) on neuroinflammation and neuroplasticity in mouse hippocampi. *J Chem Neuroanat* 94:86–92. [PubMed: 30336207]

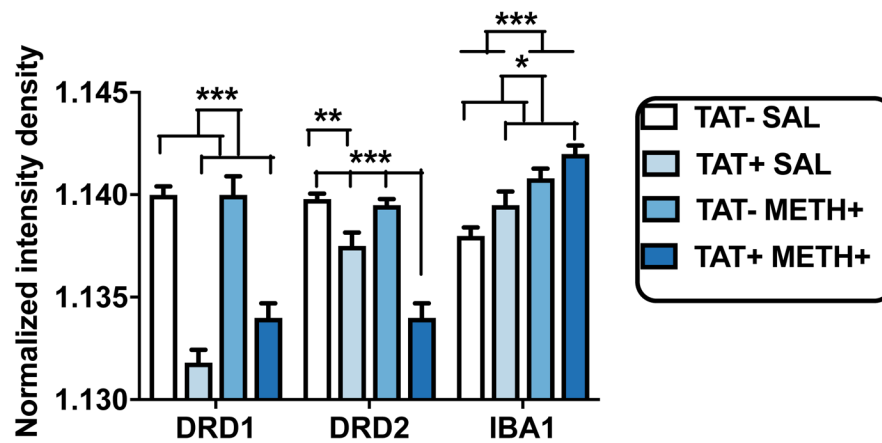


Fig. 1. Caudate putamen dopamine receptors expression and IBA-1 expression in METH-exposed Tat-transgenic mice. Immunohistochemistry on paraffin embedded sections was utilized to examine the protein distribution and levels of dopamine receptor D1, dopamine receptor D2, as well as of IBA-1 in TAT⁻ and TAT⁺ mice treated with either saline (SAL) or methamphetamine (METH⁺). Normalized intensity density data are expressed as mean \pm standard error of the mean (n=5). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. Adapted with permission from Kesby et al. (2017, *Brain Behavior and Immunity*)

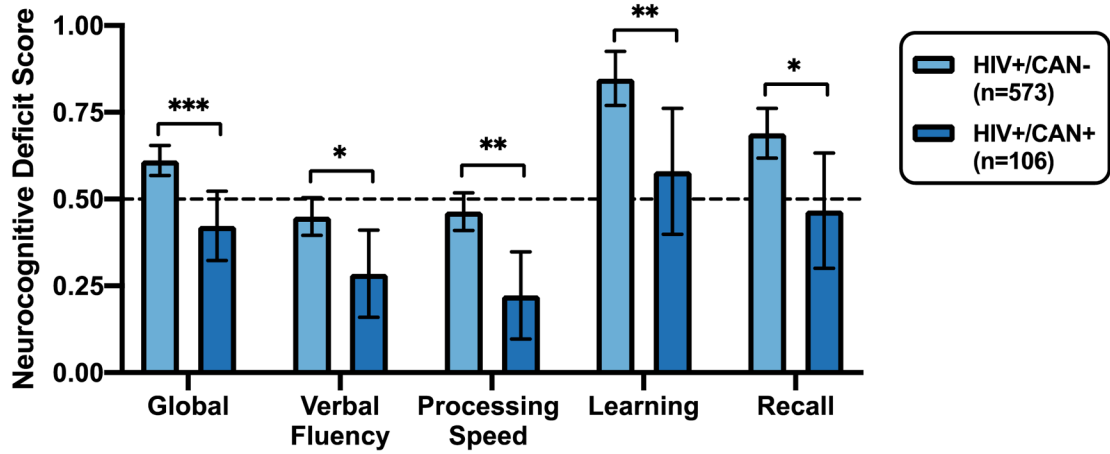


Fig. 2. Cannabis use is associated with less neurocognitive dysfunction in HIV. Substantial past use with recent exposure cannabis exposure (CAN+) was defined as meeting criteria for a lifetime history of cannabis use disorder with self-reported use in the past year. The CAN- group did not have a history of substantial use (i.e., no lifetime cannabis use disorder and estimated lifetime average grams per day of use < 1 gram) nor did they report use in the past year. Deficit scores (higher = worse) for global and domain-specific neurocognition were significantly higher in the HIV+/CAN- group compared to HIV+/CAN+ individuals (*p* range: .001 to .016). Clinically-relevant impairment is defined at a 0.5 deficit score cut-point. Data are expressed as mean ± standard error of the mean. **p* < 0.05, ***p* < 0.01, ****p* < 0.001. Data reanalyzed with permission from Watson et al. (2020)

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

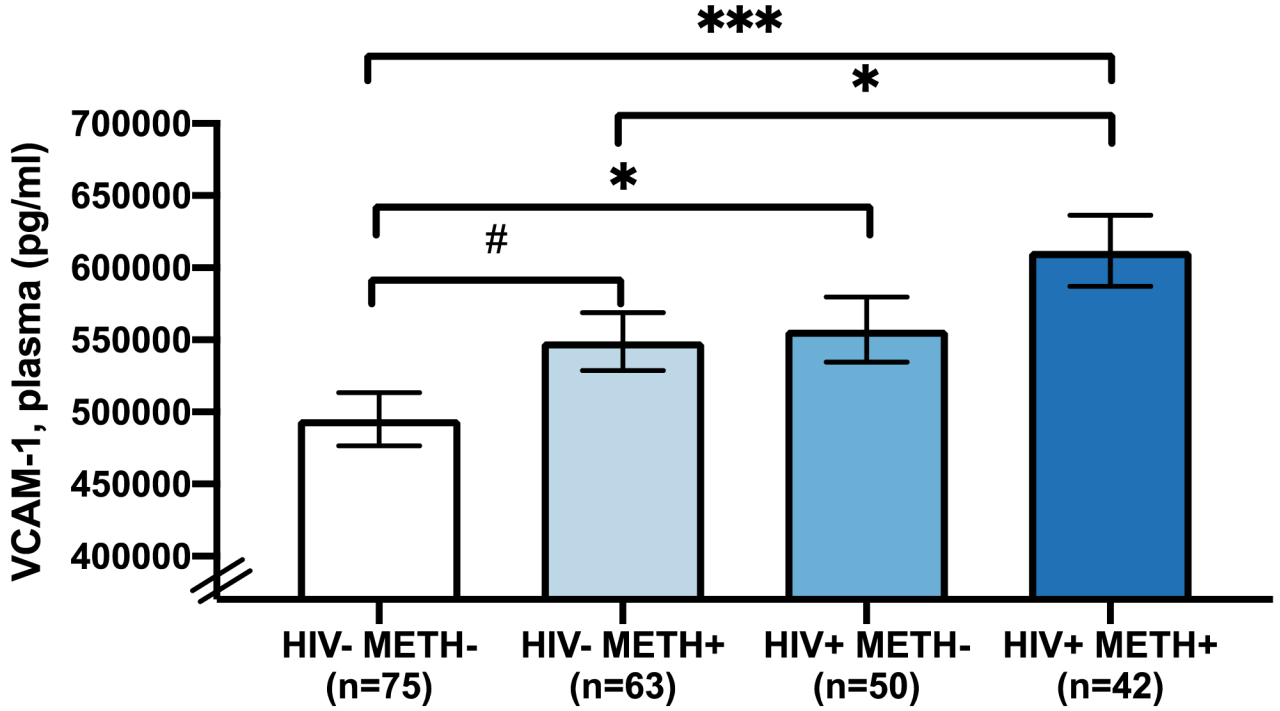


Fig. 3. Preliminary data from the Translational Methamphetamine AIDS Research Cohort (Iudicello et al., unpublished data). Regression analyses examining main and interactive effects of HIV and METH as predictors of plasma vascular cellular adhesion molecule-1 (VCAM-1) levels revealed significant independent main effects of HIV ($p=0.006$) and METH ($p=0.026$). Plasma VCAM-1 levels were highest in the dual-risk group (HIV+ METH+), followed by single-risk groups (HIV- METH+ and HIV+ METH+), and lowest in the control group (HIV- METH-). Data are expressed as mean \pm standard error of the mean. # $p < 0.10$ * $p < 0.05$, *** $p < 0.001$

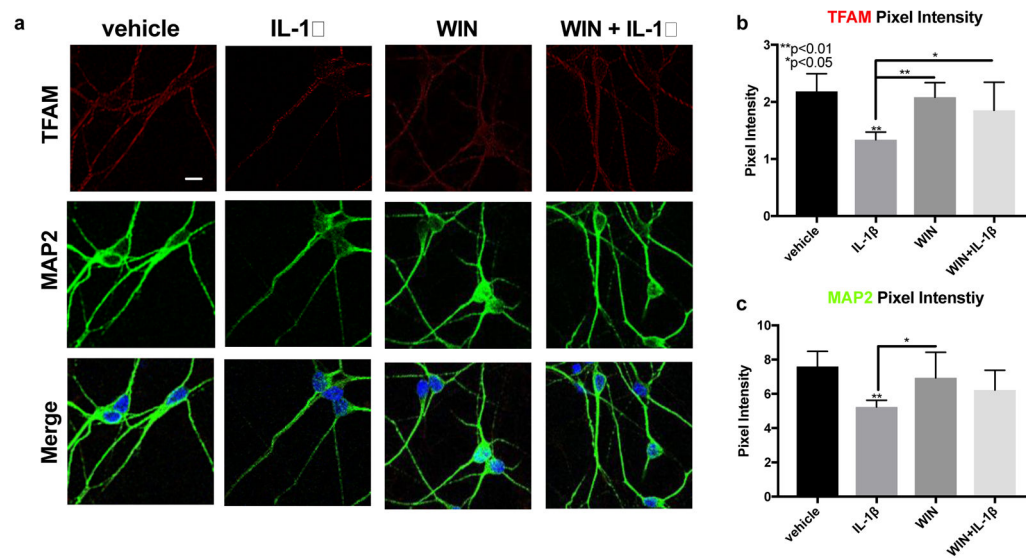


Fig. 4.

Cannabinoid (CB) receptor agonism blocks inflammation-induced toxicity in neuronal mitochondria. Similar to the effects of METH, conditioned media from immune-activated (IL-1 β) astrocytes is toxic and reduces mitochondria biogenesis in neurons. In the left panels (a), the top row shows in red the mitochondrial transcription factor, TFAM (red). The middle row shows MAP2 (green) and the bottom row combines TFAM and MAP2. The panels on the right show TFAM (b) and MAP2 (c) are both decreased in cells treated with conditioned media from reactive astrocytes. However, a CB agonist blocks toxicity as TFAM and MAP2 levels are normalized in neurons that were exposed to conditioned media from astrocytes that were treated with a CB agonist. Hence, CB agonists may protect neurons from mitochondrial damage caused by HIV and METH induced inflammation in the brain. Adapted with permission from Swinton et al. (2019), *Neurobiology of Disease*

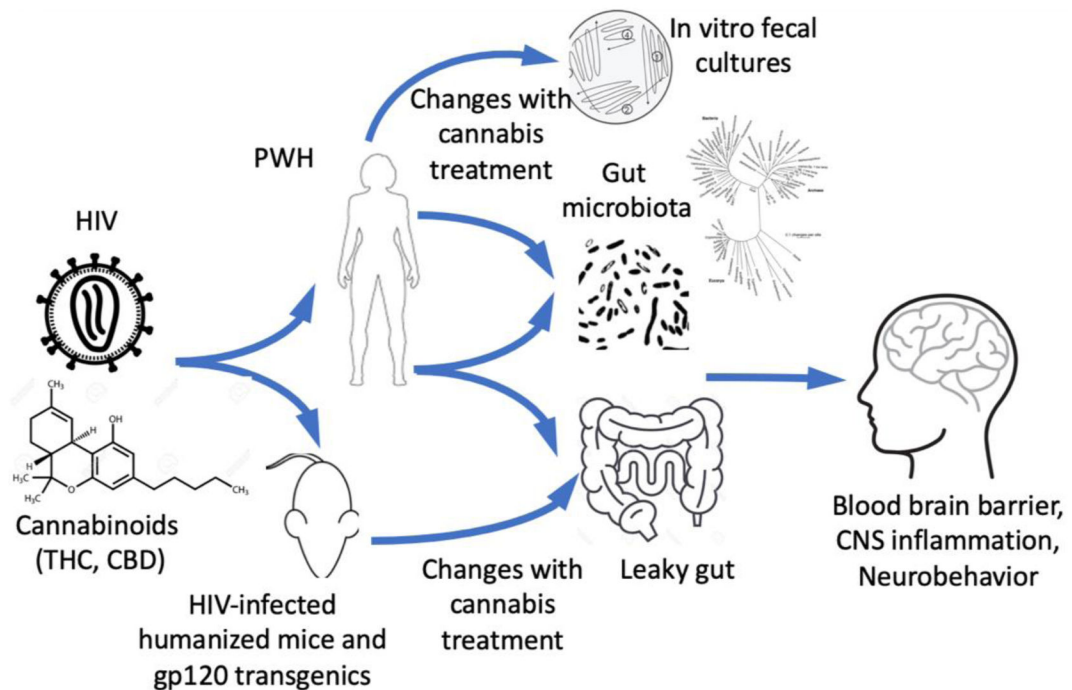


Fig. 5.

This diagram represents a translational approach to evaluating the effects of cannabis and HIV on the gut-brain-axis in five model systems: people with HIV (PWH) and HIV–humans, HIV-infected humanized mice, gp120 transgenic mice and in vitro fecal cultures. Cannabinoid treatment may normalize HIV-related gut dysbiosis and gut barrier permeability, which in turn may reduce systemic and CNS inflammation, restore blood-brain-barrier integrity, and improve neurocognition. Diagram provided courtesy of Ronald J. Ellis, M.D., Ph.D.

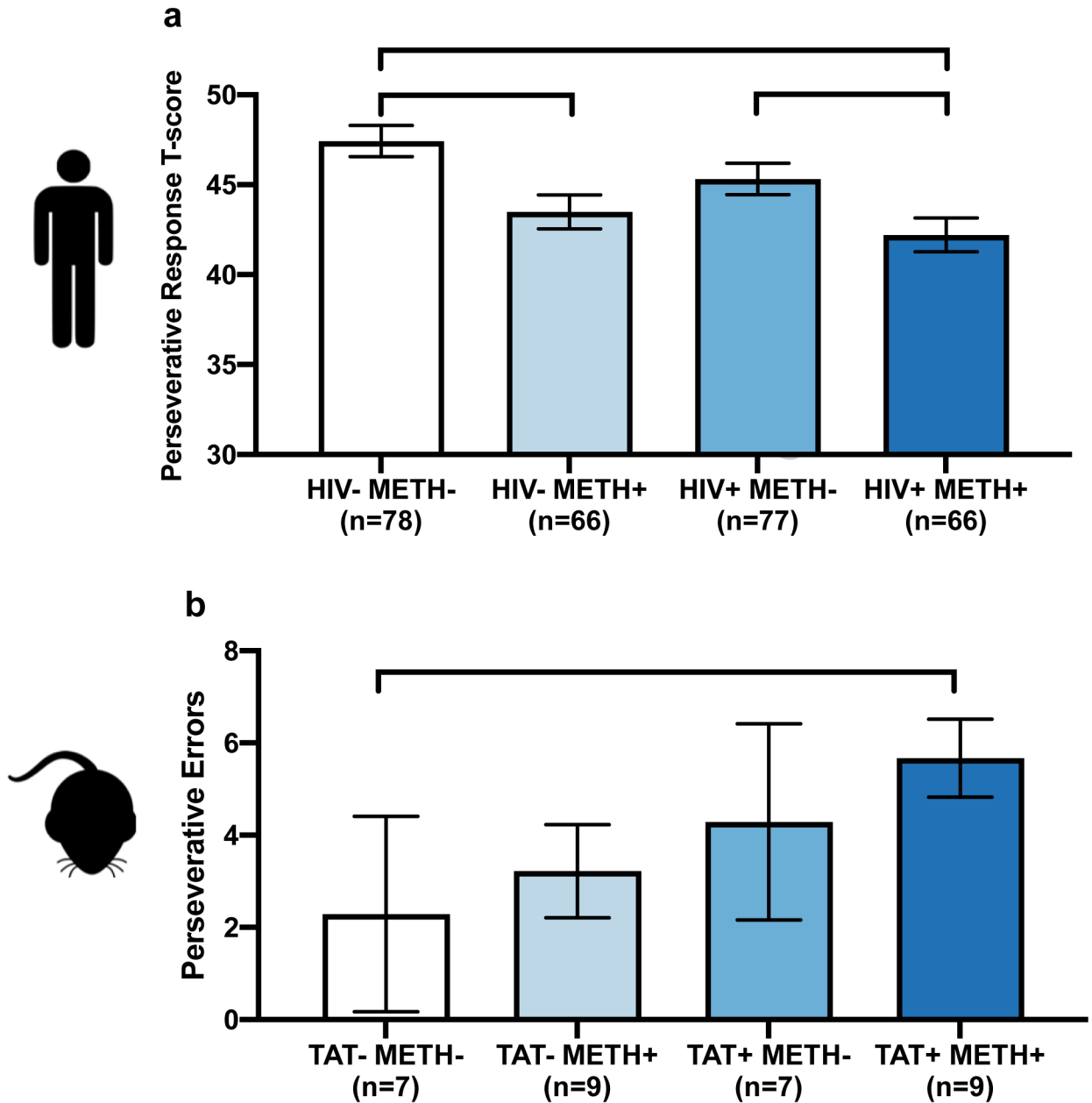


Fig. 6.

Translational evidence of executive dysfunction in HIV and METH. Perseveration was assessed using the Wisconsin Card Sorting Task (WCST) in humans and using a visual discrimination protocol with reversal learning in mice. (a) In the Translational Methamphetamine AIDS Research Center human cohort, demographically adjusted T-scores for perseverative responses on the WCST were significantly lower (signifying more perseveration) in METH-dependent participants within both HIV-serostatus groups. METH+ participants living with HIV (HIV+ METH+) also differed significantly from controls (HIV – METH–). (b) In mice, perseverative errors at the initial reversal of reward contingencies was significantly higher in TAT-transgenic mice exposed to METH (TAT+ METH+)

compared to the control group (TAT– METH–). Data are expressed as mean \pm standard error of the mean. * $p < 0.05$, ** $p < .005$, *** $p < .001$. Mice data (b) adapted with permission from Kesby et al. (2018, *Behavioural Brain Research*)

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

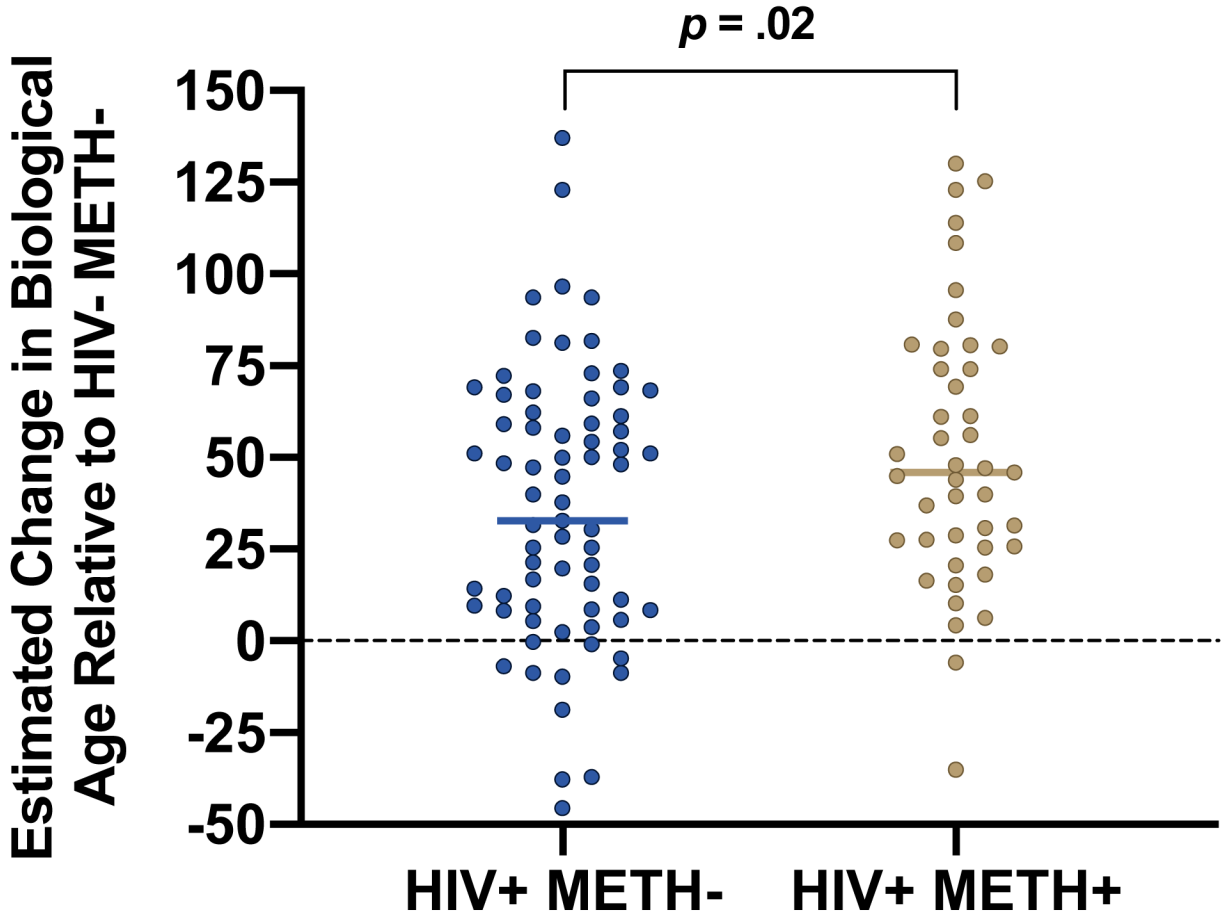


Fig. 7. HIV and a lifetime history of METH use disorder relate to older biological age. The influence of HIV and lifetime METH use disorder on estimated biological age, based on frailty index scores, was calculated relative to the reference group (HIV–METH–). First, linear regression was employed to estimate the effect of age on frailty index values in the HIV–METH– reference group. Next, biological age was estimated by inserting frailty index scores from the HIV+/METH– and HIV+/METH+ groups into the regression equation for the reference group (HIV–METH–) and solving the equation for age. The resulting estimated biological age was subtracted from chronological age to yield the data in Figure 6. Despite comparable chronological age across groups (mean age: HIV–/METH– = 51.2 years, HIV+/METH– = 50.8 years, HIV+/METH+ = 50.0 years; $p = .74$), HIV and METH produced incremental increases in estimated biological age, with a median increase in biological age of 45.9 years in the HIV+METH+ group. Data reanalyzed with permission from Paolillo et al. (2019)

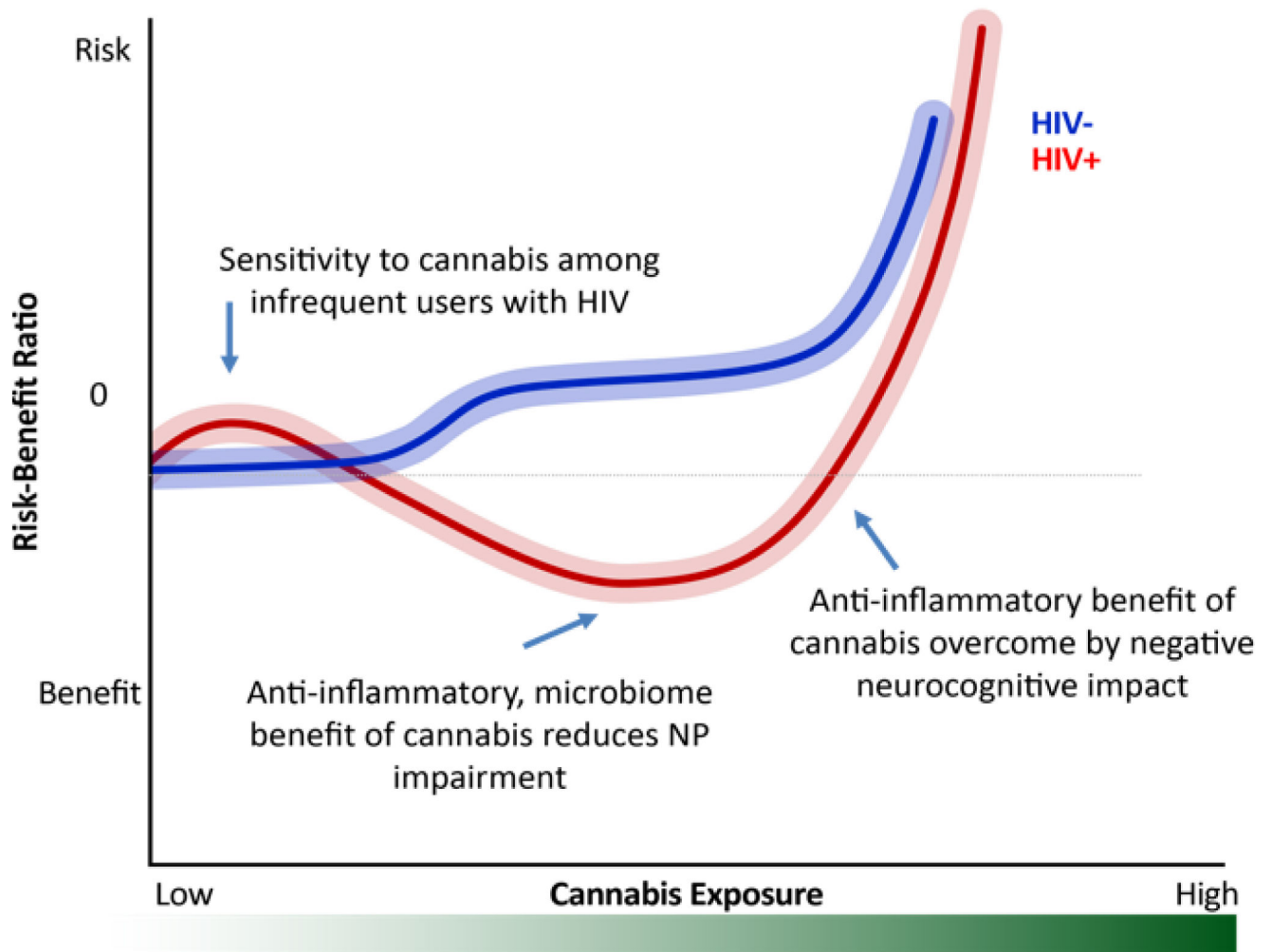


Fig. 8.

Hypothesized HIV group differences in the risk and benefit of cannabis exposure. The risk-benefit ratio of cannabis may differ between PWH (HIV+) and healthy adults (HIV-). In each instance, the risk-benefit ratio is conditioned on cannabis exposure, and in the case of PWH, severity of HIV disease. For HIV- adults, low cannabis exposure may confer little risk and little benefit (since there is no underlying disease process). As use in HIV- increases, risk increases (e.g., neurocognitive impairment [NCI]), which then flattens as tolerance develops. At high exposure, the tolerance is eclipsed by mounting toxicity. For PWH, cannabis's anti-inflammatory effects may dominate with moderate "steady state" exposure [mid-curve] and this may reduce the risk of NCI. This "benefit" would be most pronounced in those who acquire tolerance and maintain moderate use. Infrequent users, in whom the anti-inflammatory effects are intermittent, would experience more risk, because of greater vulnerability to the repeated, acute impairing effects of cannabis. At progressively higher cannabis exposure, mounting toxicity dominates the putative beneficial effects