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Predicting the Next Superspreader

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ABSTRACT The spread of multidrug-resistant zoonotic pathogens, such as *Salmonella*, within livestock is of concern for food safety. The spread of *Salmonella* on the farm is escalated by superspreaders, which shed the pathogen at high numbers with their feces. However, there are currently no biomarkers to identify potential superspreaders. Kempf and coworkers determined that a potent early inflammatory response to *Salmonella* infection and changes in the microbiota composition are associated with the superspreader phenotype in pigs (F. Kempf, G. Cordoní, A.M. Chaussé, R. Drumo, et al., *mSystems*, in press, <https://doi.org/10.1128/msystems.00852-22>). Since these biomarkers only develop during *Salmonella* infection, additional work is needed to predict animals that have the potential to become superspreaders.

KEYWORDS *Salmonella*, antibiotic resistance, gut microbiome, swine

An outbreak caused by a multidrug-resistant *Salmonella enterica* subsp. *enterica* serovar Typhimurium clone, known as monophasic *S. Typhimurium*, started in 2006 in the United Kingdom and other European countries and is ongoing (1). Monophasic *S. Typhimurium* carries a deletion of the *fljAB* operon and is particularly associated with intestinal carriage in pig herds, from where it was introduced into the human food supply (2–4). The emergence of monophasic *S. Typhimurium* coincided with the beginning of a European Union-wide ban of using antibiotics as growth promoters in pig feed, which made inclusion of copper salts a popular alternative to improve growth performance of pig herds (5). Whole-genome analysis of monophasic *S. Typhimurium* isolates from the United Kingdom showed that they form a single clade derived from an ancestral organism carrying a large novel genomic island (designated SGI-4), suggesting that SGI-4 was acquired shortly before clonal expansion of the monophasic *S. Typhimurium* clade (6). SGI-4 encodes a heavy metal RND-family efflux pump conferring enhanced resistance to copper sulfate, thus correlating with the common use of dietary copper supplementation in the porcine reservoir from where this clade originates (5).

The phylogenetic tree of *S. Typhimurium* branches into two major subdivisions, one containing the monophasic *S. Typhimurium* clade and the other including commonly used *S. Typhimurium* laboratory strains (e.g., ATCC 14028, SL1344) as well as multidrug-resistant *S. Typhimurium* clones associated with previous outbreaks in cattle and humans (7). Knowledge about *S. Typhimurium* pathogenesis is largely derived from studies on isolates belonging to the latter subdivision, whereas the monophasic *S. Typhimurium* clade remains poorly studied. Here, Kempf and coworkers investigated which properties are associated with high fecal shedding of monophasic *S. Typhimurium* in pigs (8).

In mice, a fraction of animals, termed superspreaders, exhibit a high luminal abundance of *S. Typhimurium* within the colon and are responsible for pathogen transmission (9). The luminal abundance of *S. Typhimurium* within the colon is controlled initially by the microbiota (10), but it increases once virulence factors trigger intestinal inflammation (11). Increased pathogen growth occurs as the host response escalates the availability of respiratory electron acceptors in the colonic lumen, from which the pathogen then benefits (12–15). *S. Typhimurium* virulence factors trigger severe acute intestinal inflammation in mice (16, 17) and cattle (18), but the pathogen causes less severe disease in pigs. Nonetheless, virulence

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factors enable *S. Typhimurium* to overcome colonization resistance conferred by the microbiota in this host species (19). The ability to reach superspreader status is thought to be important for transmission within pig herds, which is an important food safety concern.

The new research shows that enhanced shedding of monophasic *S. Typhimurium* with the feces of pigs is associated with higher proinflammatory cytokine levels during, but not prior to, infection (8). Furthermore, the superspreader phenotype of monophasic *S. Typhimurium* in pigs is associated with changes in the microbiota composition during infection that predict a functional enrichment for pathways involved in anaerobic respiration (8). These data are consistent with previous work suggesting that *S. Typhimurium* virulence factors trigger intestinal inflammation to increase the availability of respiratory electron acceptors that boost pathogen growth (20, 21). Nevertheless, this work highlights that ongoing work is needed to define the biomarkers that reliably predict which animals will reach superspreader status after they become infected with monophasic *S. Typhimurium*.

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