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Authors

Jain, P
Aoki, E
Keating, M
[et al.](#)

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ORIGINAL ARTICLE

Characteristics, outcomes, prognostic factors and treatment of patients with T-cell prolymphocytic leukemia (T-PLL)

P. Jain^{1†}, E. Aoki^{1†}, M. Keating¹, W. G. Wierda¹, S. O'Brien², G. N. Gonzalez³, A. Ferrajoli¹, N. Jain¹, P. A. Thompson¹, E. Jabbour¹, R. Kanagal-Shamanna⁴, S. Pierce¹, A. Alousi⁵, C. Hosing⁵, I. Khouri⁵, Z. Estrov¹, J. Cortes¹, H. Kantarjian¹, F. Ravandi¹ & T. M. Kadia^{1*}

¹Department of Leukemia, The MD Anderson Cancer Center, Houston; ²Division of Hematology/Oncology, Chao Family Comprehensive Cancer Center, UC Irvine, Irvine; Departments of ³Biostatistics, ⁴Hematopathology; ⁵Stem Cell Transplantation, The MD Anderson Cancer Center, Houston, USA

*Correspondence to: Dr Tapan M. Kadia, Department of Leukemia, The University of Texas, MD Anderson Cancer Center, 1515 Holcombe Blvd., Box 428, Houston, TX 77030, USA. Tel: +1713-792-7305; E-mail: tkadia@mdanderson.org

[†]Both authors contributed equally as senior authors.

Background: T-cell prolymphocytic leukemia (T-PLL) is a rare and aggressive disease. In this study, we report our experience from 119 patients with T-PLL.

Patients and methods: We reviewed the clinico-pathologic records of 119 consecutive patients with T-PLL, who presented to our institution between 1990 and 2016.

Results: One hundred and nineteen patients with T-PLL were analysed. Complex karyotype and aberrations in chromosome 14 were seen in 65% and 52% patients, respectively. Seventy-five patients (63%) were previously untreated and 43 (37%) were initially treated outside our institution. Sixty-three previously untreated patients (84%) received frontline therapies. Overall, 95 patients (80%) have died. Median overall survival (OS) from diagnosis was 19 months [95% confidence interval (CI) 16–26 months]. Using recursive partitioning (RP), we found that patients with hemoglobin < 9.3 g/dl, lactate dehydrogenase (LDH) \geq 1668 IU/l, white blood cell \geq 208 K/l and β 2M \geq 8 mg/l had significantly inferior OS and patients with hemoglobin < 9.3 g/dl had inferior progression-free survival (PFS). In multivariate analysis, we identified that presence of pleural effusion [hazard ratio (HR) 2.08 (95% CI 1.11–3.9); $P = 0.02$], high LDH (\geq 1668 IU/l) [HR 2.5 (95% CI 1.20–4.24); $P < 0.001$], and low hemoglobin (< 9.3 g/dl) [HR 0.33 (95% CI 0.14–0.75); $P = 0.008$] were associated with shorter OS. Fifty-five previously untreated patients received treatment with an alemtuzumab-based regimen (42 monotherapy and 13 combination with pentostatin). Overall response rate, complete remission rate (CR) for single-agent alemtuzumab and alemtuzumab combined with pentostatin were 83%, 66% and 82%, 73% respectively. In patients who achieved initial CR, stem cell transplantation was not associated with longer PFS and OS.

Conclusion: Outcomes in T-PLL remain poor. Multicenter collaborative effort is required to conduct prospective studies.

Key words: T-PLL, prolymphocytic leukemia, prolymphocytes

Introduction

T-cell prolymphocytic leukemia (T-PLL) is an uncommon, mature lymphoid leukemia that is typically associated with a poor prognosis. T-PLL is diagnosed based on characteristic clinical, morphologic, immunophenotypic [1], cytogenetic and molecular features which may explain its aggressive clinical course [2]. Cytogenetic features [3] include chromosome 14

aberrations involving gene rearrangements of proto-oncogenes *TCL1* (T-cell leukemia 1) or *MTCP1* (mature T-cell proliferation 1; on chromosome X), aberration in chromosomes 8 (especially isochromosome 8 potentially leading to overexpression of *MYC*), deletion chromosome 11q22.3 (ATM gene deletion), or high prevalence of complex karyotype. Approximately 70%–80% of patients with T-PLL show an overexpression of *TCL-1* by

immunohistochemistry (IHC) or by chromosomal analysis using FISH and/or karyotyping. TCL1 belongs to beta barrel family of proteins localized in the cytoplasm. TCL1 and MTC1P1 oncoprotein are structurally similar with 40% identical amino acid residues [4]. TCL1 is shown to promote kinase activity and transphosphorylation of AKT and promotes the nuclear transport of AKT [5, 6]. Furthermore, overexpression of TCL-1 has been shown to modulate and amplify the AKT activation mediated by stimulation of T-cell receptor (TCR) in cells from T-PLL patients and induce hyperproliferation [7]. A recent study [8] has demonstrated gain-of-function mutations of *IL-2RG*, *JAK-1/JAK-3* and *STAT5B* using whole-genome and whole-exome sequencing in T-PLL [9, 10]. Specifically, shorter survival was noted in patients with *JAK-3* p.M11I mutations. Inhibition of *STAT5B* promoted apoptosis of T-PLL cells.

Despite an improved understanding into the molecular biology of T-PLL, the disease continues to present a therapeutic challenge for clinicians. In a recent analysis of SEER database [11], the median survival was 21 months and better outcomes were noted after the introduction of alemtuzumab (after 2000) for the therapy of T-PLL. None of the currently available therapies are very effective in producing durable remissions in patients with T-PLL [12]. Overall response rates (ORR) with alemtuzumab-based regimens vary from 50% to 90% and complete remission rates (CR) vary from 60% to 80% in different studies, with short duration of response [13–16]. Other chemotherapies such as nelarabine, single-agent bendamustine [17], combination of pentostatin with alemtuzumab [18], fludarabine, mitoxantrone cyclophosphamide followed by alemtuzumab consolidation [19] have been used with limited success. Allogenic stem cell transplantation (SCT) may induce longer relapse-free survival in few patients [20, 21]. Patients who received total body irradiation and shorter time from diagnosis to SCT had better relapse-free survival; however, treatment-related mortality and relapses remain common [22–24]. Rarity of this disease precludes the development of large-scale prospective clinical trials. Here, in this analysis, we are reporting the clinical experience of the largest series of patients with T-PLL from a single institution.

Methods

We reviewed the clinico-pathologic records of 119 consecutive patients with T-PLL, at our institution. Survival distributions was estimated by Kaplan–Meier method. Cox proportional hazard model for survival was analysed. We also carried out classification and regression tree analysis of various parameters to identify the optimal cut off points for predicting survival. Details are available in supplementary, available at *Annals of Oncology* online.

Results

Patient characteristics

We analysed the characteristics, prognostic factors, outcomes and treatments of 119 patients with T-PLL. Supplementary Table S1, available at *Annals of Oncology* online, provides the summary of the clinical and laboratory characteristics of the patients according to the treatment status. At the initial presentation, 20 patients (17%) presented with performance status 2 or higher.

Lymphadenopathy, splenomegaly, skin lesions, pleural effusion and hepatomegaly were seen in 58%, 38%, 28%, 12% and 11%, of patients, respectively. Small-cell variant of T-PLL was detected in 21/106 patients (20%) and 57% patients with small-cell variant co-expressed CD4 and CD8. Overall, 108 patients had karyotype data available for analysis. Complex karyotype and aberrations in chromosome 14 were seen in 70 (65%) and 56 (52%) patients, respectively.

Treatments and survival outcomes

Overall median survival from initial diagnosis was 19 months [95% confidence interval (CI), 16–26 months] (Figure 1A). At initial presentation to our center, 75 patients (63%) were previously untreated and 43 patients (36%) were previously treated. Treatment and follow-up information were available in 67/75 (89%) previously untreated patients.

Frontline patients

Among these 67 previously untreated patients, 4 patients remained in observation and 63 received treatment (55 with alemtuzumab-based regimens and 8 with non-alemtuzumab-based therapy). The median time from initial diagnosis to first treatment of these 63 patients was 2.3 months (range, 0.1–52 months). Fourteen patients were considered to have an indolent disease, 10 with treatment requirement after ≥ 19 months after initial diagnosis and 4 patients remained under observation. For the 55 patients treated with alemtuzumab-based therapy, 42 (76%) were treated with single-agent alemtuzumab and 13 (24%) with a combination of alemtuzumab and pentostatin. ORR and CR in patients who received frontline single-agent alemtuzumab ($n=42$) and alemtuzumab combined with pentostatin ($n=13$) were 81% and 61% in single-agent alemtuzumab cohort while 82% and 73% in alemtuzumab combined with pentostatin cohort, respectively. Non-alemtuzumab-based therapies are described in supplementary, available at *Annals of Oncology* online.

Patients with relapsed or refractory disease

We next analysed the response to salvage treatments in previously treated patients (before presentation at our center). Information for salvage treatments was available in 29 patients. Relapses occurred in 25/29 patients with a median time to relapse of 2.5 months (0.3–31 months). The ORR with salvage combination alemtuzumab with pentostatin ($n=5$) was 75% compared with 46% with single-agent alemtuzumab ($n=15$), summarized in Table 1. There was no difference in PFS or OS according to the type of salvage therapy (single-agent alemtuzumab versus alemtuzumab with pentostatin versus single-agent nelarabine) (supplementary Figure S1A and B, available at *Annals of Oncology* online).

The median overall survival (OS) and progression-free survival (PFS) of patients are summarized in Table 1. Patients who received frontline treatment had a significantly longer PFS and OS compared with those who were treated with the first salvage ($P=0.0002$ and 0.01 , respectively; Figure 1B and C). Furthermore, patients who received frontline therapy with alemtuzumab-based regimens (monotherapy or combined with

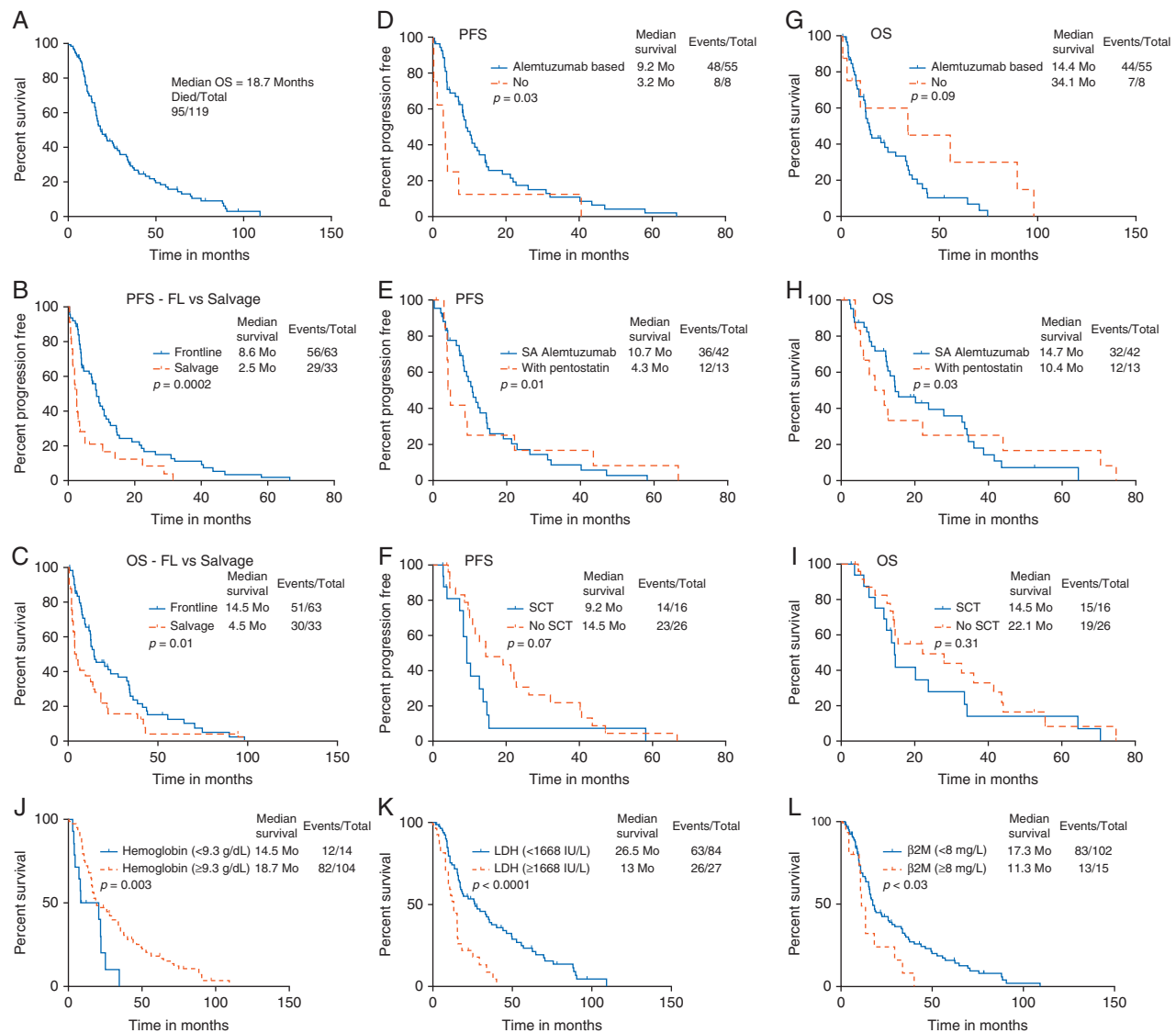


Figure 1. Survival outcomes in patients with T-PLL—overall and according to the treatment status at the time of initial presentation to MDACC—frontline versus salvage treatment. (A) All patients from the time of initial diagnosis, median OS was 19 months. (B) PFS is significantly longer in patients treated in frontline setting ($P = 0.0002$). (C) OS is significantly longer in patients treated in frontline setting ($P = 0.01$). Survival outcomes according to the type of frontline therapy. (D and G) PFS and OS in patients who received therapy with alemtuzumab (single-agent alemtuzumab or in combination with pentostatin) versus other therapies ($P = 0.03$ for PFS and $P = 0.09$ for OS). (E and H) PFS and OS in patients who received alemtuzumab (single agent alemtuzumab versus alemtuzumab with pentostatin ($P = 0.01$ for PFS and $P = 0.03$ for OS). Survival outcomes according to treatment with SCT in patients who achieved complete remission after frontline therapy. (F) PFS was not significantly different in patients with/without SCT ($P = 0.07$). (I) OS was not significantly different in patients with/without SCT ($P = 0.31$). Survival outcomes in patients with T-PLL, according to the prognostic factors identified by *recursive partitioning* (RP) analysis. (J–L) OS was significantly shorter in patients with low hemoglobin (<9.3 g/dl), high serum LDH (≥ 1668 IU/l) and high $\beta 2M$ (≥ 8 mg/l) level.

chemotherapy) had significantly longer PFS when compared with patients treated with non-alemtuzumab-based therapies ($P = 0.03$ for PFS and 0.09 for OS; Figure 1D and E. Moreover, patients who received single-agent alemtuzumab had significantly longer PFS and OS when compared with patients treated with pentostatin and alemtuzumab ($P = 0.01$ and 0.03 , respectively; Figure 1F and G.

Sixteen patients underwent allogeneic SCT in first CR, 73% of 16 patients received an unrelated donor SCT. None of the patients received autologous SCT. We did not observe any significant difference in PFS or OS (Figure 1H and I) among patients

who did or did not undergo SCT after achieving initial CR with any frontline therapies. We have summarized the details of SCT in supplementary Table S2, available at *Annals of Oncology* online. Eight patients (50%) relapsed after SCT.

For the total cohort, 43 patients were treated before initial presentation to our center. Details of treatment responses, follow ups outside our center was not available for review; however, the median number of prior treatments was 2 (range, 1–6); 46% of prior therapies were fludarabine based, 12% with single-agent chlorambucil, 14% with single-agent pentostatin, 16% with single-agent alemtuzumab and 12% with miscellaneous regimen.

Table 1. Summary of the type of treatments, treatment responses and survival outcomes in patients with T-PLL (frontline and salvage setting)

	Alemtuzumab monotherapy		Alemtuzumab + pentostatin		Nelarabine
	First line n = 42	Salvage n = 15	First line n = 13	Salvage n = 5	All n = 5 (one frontline and four salvage)
Overall response (%)	81	46	82	75	20
Complete response (%)	61	46	73	50	0
Median OS (months)*	15	15	10.4	2.6	2
Median PFS (months)*	11	3	4.3	2.6	2
Median number of prior regimen (range)	–	1 (1–4)	–	3 (1–3)	2 (1–3)

*Censored at stem cell transplant, total of 29 patients were treated with salvage therapy and shown above is information on 24 patients, 5 patients were treated with miscellaneous treatments (Fardosine, HyperCVAD, fludarabine-based regimen).

Prognostic factors and survival outcomes

Using recursive partitioning (RP), we have identified an optimal cut off value for certain baseline parameters which significantly correlated with PFS and OS: hemoglobin, serum lactate dehydrogenase (LDH), and β 2M levels. Patients with low hemoglobin (<9.3 g/dl), high serum LDH (\geq 1668 IU/l), and high serum β 2M (\geq 8 mg/l) had significantly inferior PFS (supplementary Figure S2A and C, available at *Annals of Oncology* online) and OS Figure 1J–L. Furthermore, we detected that OS was significantly inferior in patients showing TCL-1 rearrangement by cytogenetics; however, a similar prognostic impact was not observed in patients with positive TCL-1 expression by IHC. PFS was not significantly different in patients with or without TCL-1 expression (supplementary Figure S3A–D, available at *Annals of Oncology* online). Survival was not significantly different in patients with or without small-cell variant of T-PLL.

Finally, we carried out univariate and multivariate analyses to identify the factors predictive for OS. All patients in this analysis were previously untreated and were analysed from the time of initial diagnosis (supplementary Table S3, available at *Annals of Oncology* online). In multivariate analysis, factors independently predictive for significantly increased risk of death were presence of pleural effusion [hazard ratio (HR) 2.08 (95% CI 1.11–3.90); $P=0.02$], low hemoglobin (<9.3 g/dL) 0.33 (0.14–0.75; $P=0.008$), high LDH level (\geq 1668 IU/l) 2.52 (1.2–4.24; $P<0.001$), and high white blood cell count [white blood cell (WBC) \geq 208 K/l] 3.35 (0.98–11.49; $P=0.05$). For PFS, we identified that patients with non-Caucasian ethnicity, absence of small-cell variant of T-PLL, and high β 2M (\geq 8 mg/l) had significantly higher risk of disease progression after initial therapy (supplementary Table S4, available at *Annals of Oncology* online).

Discussion

In this study, we have analysed, to our knowledge, the largest series of patients with T-PLL from a single institution. We described the patient and disease characteristics, responses to treatment, long-term outcomes, and factors that may be useful in determining prognosis.

Patients with T-PLL continue to have poor outcomes [2] and there is only a minority of patients who do not require treatment

after the initial diagnosis. Despite the recent advances in our understanding of the molecular biology of T-PLL, the discovery of newer, more effective therapies in this uncommon and aggressive leukemia remains an unmet need [12]. None of the currently available treatment modalities (including SCT) have significantly improved the outcomes in T-PLL [23]. Our data suggest that single-agent alemtuzumab as an initial therapy is associated with high response rates and is the best available treatment of T-PLL. However, durable remissions are still uncommon and relapsed disease accounts for the majority of failures. Furthermore, in this study, attempts to prolong remission with allogeneic SCT after initial CR did not improve survival. We identified that hemoglobin <9.3 g/dl, serum LDH level \geq 1668 IU/l, high WBC count, and presence of a pleural effusion at initial diagnosis significantly predicted for poor OS. These factors were not previously recognized to have distinct prognostic impact in T-PLL. We also identified that non-Caucasian ethnicity, absence of small-cell variant of T-PLL and high β 2M (\geq 8 mg/l) had significantly higher risk of disease progression after initial therapy. Precise reasons to explain these findings are unclear, as to date, no study has reported on the clinico-pathologic characterization of small-cell variant or impact of race in T-PLL. Additionally, it is possible that similar to other lymphoproliferative disorders, serum LDH and β 2 microglobulin may reflect disease burden in T-PLL. Importantly, the characteristics of the previously treated patients (e.g. elevated β 2 microglobulin, more lymphadenopathy and organomegaly) (Table 1) were suggestive of bulky disease.

Previously published retrospective studies were limited in the number of patients (range, 10–86), and data were obtained from multiple centers and/or different countries. One prior study reported data from 86 patients with T-PLL [7] and identified that overexpression of TCL-1 oncoprotein was associated with aggressive disease, short lymphocyte doubling time, enhanced TCR and AKT signaling, and rapid cell proliferation. Herling et al [7] reported that aged >65 years and higher WBC count (>40 K/ μ l) was associated with an increased risk of death. However, these cut offs were arbitrary and were not derived by statistical analysis. In our study, we based the cut off derived from RP analysis of different baseline variables and identified an optimal, clinically applicable cut-off value, which was not reported in previously published studies. We further applied these variables into the

final cox regression model for survival outcomes and identified their prognostic relevance.

The role of SCT after initial CR in patients with T-PLL remains controversial, largely due to high rate of early relapses, and high transplant related mortality [21]; only a small fraction of patients have 3-year survival of 5%–20% [20, 24]. Two multi-institutional studies have reported that patients with T-PLL who have undergone SCT (auto or allogeneic) have better PFS and OS compared with patients who did not undergo SCT [21, 22]. In contrast, our data show that the median survival in 16 patients who underwent allogeneic SCT in first CR was 14.5 months compared with 22 months (P = not significant) in 26 patients in first CR after different frontline therapies who did not receive SCT. The explanation for this lack of benefit is not clear, but could be related to differences in conditioning regimens, the relative resistance to cytotoxic chemotherapy in T-PLL, and the lack of any robust graft versus leukemia effect in T-PLL [22, 25, 26]. Nonetheless, effective post remission therapy is a critical need for improving outcomes in these patients. Longer term consolidation or maintenance strategies, immunotherapeutic approaches, or targeted therapies should be investigated to maintain remissions achieved after effective frontline therapy.

Currently, data are very sparse on the cytokine milieu of T-PLL. However, the initial clinical presentation with B-symptoms, lymphadenopathy, organomegaly, pleural and pericardial effusions are suggestive of a cytokine-mediated syndrome. Indeed, about 76% patients with T-PLL have activating somatic mutations in *IL2RG/JAK1/JAK3/STAT5B* [8, 10] identified through whole-exome sequencing or by next-generation sequencing [8, 27, 28]. The net effect of these mutations is constitutive activation of JAK-STAT signaling and up regulation of *STAT5B* target genes [8]. It is possible that a pan-JAK inhibitor or *STAT5* inhibitor may have a therapeutic effect in T-PLL.

About 50%–60% of patients exhibited complex karyotype and most abnormalities were observed in chromosome 14, in addition to the previously described del 11q, del 17p and chromosome 8 aberrations. Chromosome 14 abnormalities are a hallmark of T-PLL and involve the *TCL-1* oncogene. Presence of complex karyotype [29, 30] and high incidence of *ATM* mutations [31] is known to be associated with refractory disease and poor prognosis in lymphoid malignancies. Two thirds of patients with T-PLL have *ATM* mutations [32, 33] and patients with ataxia telangiectasia may have a higher risk of transformation to T-PLL [34]. Inactivating mutations in *ATM* or loss of material at chromosome 11q23 (at the *ATM* gene locus) lead to deficient signaling of double-stranded DNA repair and cell cycle arrest after DNA damage. It is possible that in T-PLL [35, 36], these genetic aberrations may explain the molecular complexity and refractoriness to chemotherapy. Exploiting double-stranded DNA repair defects may be another important strategy to selectively target T-PLL cells. At present, this study lacks complete information on the molecular characteristics of these patients; therefore, it is difficult to provide a mechanistic link between cytogenetic finding and clinical course.

We further noted that although the response rate to alemtuzumab with pentostatin is marginally better than single agent alemtuzumab (73 versus 61%), the PFS and OS was significantly longer in patients treated with single-agent alemtuzumab. Since this is a retrospective study, it is possible that selection bias for either regimen may have affected the outcomes i.e. patients with

bulky, more aggressive disease may have received the combination regimen and therefore had inferior outcomes. The PFS was better with alemtuzumab-based regimens compared with non-alemtuzumab-based therapies; for OS, the trend of increased OS with non-alemtuzumab-based therapies could be misleading, since virtually all of these patients received salvage therapy with alemtuzumab. Few patients who were treated with nelarabine and fardosine did not respond well.

There are several limitations to our study. This is a retrospective analysis that allows selection bias and uncontrolled analysis. Treatment and complete follow-up information was not available for all the patients, and third of patients were previously treated before coming to our center. However, when analyzing for outcomes, responses and prognostic factors, care was taken to make sure complete and original source data were available for all analysed patients. There was no uniform pattern of follow-up and management was according to individual physician's choice, but the treatment modalities were examined and presented individually. While cytogenetic, morphology and flow cytometric studies were available for the majority of patients, our study is limited by the lack of complete molecular genetic data on all patients. Collaborations are now underway to retrospectively examine available banked samples and create protocols to prospectively study newly diagnosed patients.

In summary, T-PLL continues to pose a therapeutic challenge and newer modalities are required to treat this leukemia. Rarity of this disease limits the conduct of large-scale clinical trials, and multicenter collaborative effort is required to conduct prospective studies. Studies are underway to further define the genomic imbalances in patients with T-PLL and identify newer therapeutic targets and recognize the signaling pathways which can provide drug resistance in T-PLL.

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Disclosure

The authors have declared no conflicts of interest.

References

1. Chen X, Cherian S. Immunophenotypic characterization of T-cell prolymphocytic leukemia. *Am J Clin Pathol* 2013; 140: 727–735.
2. Dearden C. Management of prolymphocytic leukemia. *Hematol Am Soc Hematol Educ Program* 2015; 2015: 361–367.
3. Urbankova H, Holzerova M, Balcarikova J et al. Array comparative genomic hybridization in the detection of chromosomal abnormalities in T-cell prolymphocytic leukemia. *Cancer Genet Cytogenet* 2010; 202: 58–62.

4. Fu ZQ, Du Bois GC, Song SP et al. Crystal structure of MTCP-1: implications for role of TCL-1 and MTCP-1 in T cell malignancies. *Proc Natl Acad Sci USA* 1998; 95: 3413–3418.
5. Auguin D, Barthe P, Royer C et al. Structural basis for the co-activation of protein kinase B by T-cell leukemia-1 (TCL1) family proto-oncoproteins. *J Biol Chem* 2004; 279: 35890–35902.
6. Pekarsky Y, Koval A, Hallas C et al. Tcl1 enhances Akt kinase activity and mediates its nuclear translocation. *Proc Natl Acad Sci USA* 2000; 97: 3028–3033.
7. Herling M, Patel KA, Teitell MA et al. High TCL1 expression and intact T-cell receptor signaling define a hyperproliferative subset of T-cell prolymphocytic leukemia. *Blood* 2008; 111: 328–337.
8. Kiel MJ, Velusamy T, Rolland D et al. Integrated genomic sequencing reveals mutational landscape of T-cell prolymphocytic leukemia. *Blood* 2014; 124: 1460–1472.
9. Bellanger D, Jacquemin V, Chopin M et al. Recurrent JAK1 and JAK3 somatic mutations in T-cell prolymphocytic leukemia. *Leukemia* 2014; 28: 417–419.
10. Bergmann AK, Schneppenheim S, Seifert M et al. Recurrent mutation of JAK3 in T-cell prolymphocytic leukemia. *Genes Chromosomes Cancer* 2014; 53: 309–316.
11. Chandran R, Gardiner SK, Fenske TS, Spurgeon ES. Survival trends in T cell prolymphocytic leukemia: A SEER database analysis. *Leuk Lymphoma* 2016; 57: 942–944.
12. Dearden C. How I treat prolymphocytic leukemia. *Blood* 2012; 120: 538–551.
13. Dearden CE, Matutes E, Cazin B et al. High remission rate in T-cell prolymphocytic leukemia with CAMPATH-1H. *Blood* 2001; 98: 1721–1726.
14. Dearden CE, Khot A, Else M et al. Alemtuzumab therapy in T-cell prolymphocytic leukemia: comparing efficacy in a series treated intravenously and a study piloting the subcutaneous route. *Blood* 2011; 118: 5799–5802.
15. Keating MJ, Cazin B, Coutre S et al. Campath-1H treatment of T-cell prolymphocytic leukemia in patients for whom at least one prior chemotherapy regimen has failed. *JCO* 2002; 20: 205–213.
16. Ravandi F, O'Brien S, Jones D et al. T-cell prolymphocytic leukemia: a single-institution experience. *Clin Lymphoma Myeloma*. 2005; 6: 234–239.
17. Herbaux C, Genet P, Bouabdallah K et al. Bendamustine is effective in T-cell prolymphocytic leukaemia. *Br J Haematol* 2015; 168: 916–919.
18. Ravandi F, Aribi A, O'Brien S et al. Phase II study of alemtuzumab in combination with pentostatin in patients with T-cell neoplasms. *J Clin Oncol* 2009; 27: 5425–5430.
19. Hopfinger G, Busch R, Pflug N et al. Sequential chemioimmunotherapy of fludarabine, mitoxantrone, and cyclophosphamide induction followed by alemtuzumab consolidation is effective in T-cell prolymphocytic leukemia. *Cancer* 2013; 119: 2258–2267.
20. Guillaume T, Beguin Y, Tabrizi R et al. Allogeneic hematopoietic stem cell transplantation for T-prolymphocytic leukemia: a report from the French society for stem cell transplantation (SFGM-TC). *Eur J Haematol* 2015; 94: 265–269.
21. Krishnan B, Else M, Tjonnfjord GE et al. Stem cell transplantation after alemtuzumab in T-cell prolymphocytic leukaemia results in longer survival than after alemtuzumab alone: a multicentre retrospective study. *Br J Haematol* 2010; 149: 907–910.
22. Wiktor-Jedrzejczak W, Dearden C, de Wreede L et al. Hematopoietic stem cell transplantation in T-prolymphocytic leukemia: a retrospective study from the European Group for Blood and Marrow Transplantation and the Royal Marsden Consortium. *Leukemia* 2012; 26: 972–976.
23. Herling M. Are we improving the outcome for patients with T-cell prolymphocytic leukemia by allogeneic stem cell transplantation? *Eur J Haematol* 2015; 94: 191–192.
24. Matutes E, Polliack A. T-cell prolymphocytic leukemia: survival improves with alemtuzemab, but stem cell transplant eligibility 'counts' even more. *Leuk Lymphoma* 2016; 57: 746–747.
25. de Lavallade H, Faucher C, Furst S et al. Allogeneic stem cell transplantation after reduced-intensity conditioning in a patient with T-cell prolymphocytic leukemia: graft-versus-tumor effect and long-term remission. *Bone Marrow Transplant* 2006; 37: 709–710.
26. Sellner L, Bruggemann M, Schlitt M et al. GvL effects in T-prolymphocytic leukemia: evidence from MRD kinetics and TCR repertoire analyses. *Bone Marrow Transplant* 2017; 52: 544–551.
27. Stengel A, Kern W, Zenger M et al. Genetic characterization of T-PLL reveals two major biologic subgroups and JAK3 mutations as prognostic marker. *Genes Chromosomes Cancer* 2016; 55: 82–94.
28. Lopez C, Bergmann AK, Paul U et al. Genes encoding members of the JAK-STAT pathway or epigenetic regulators are recurrently mutated in T-cell prolymphocytic leukaemia. *Br J Haematol* 2016; 173: 265–273.
29. Herling CD, Klaumunzer M, Rocha CK et al. Complex karyotypes and KRAS and POT1 mutations impact outcome in CLL after chlorambucil-based chemotherapy or chemoimmunotherapy. *Blood* 2016; 128: 395–404.
30. Sarkozy C, Terre C, Jardin F et al. Complex karyotype in mantle cell lymphoma is a strong prognostic factor for the time to treatment and overall survival, independent of the MCL international prognostic index. *Genes Chromosomes Cancer* 2014; 53: 106–116.
31. Cuneo A, Bigoni R, Rigolin GM et al. Acquired chromosome 11q deletion involving the ataxia telangiectasia locus in B-cell non-Hodgkin's lymphoma: correlation with clinicobiologic features. *JCO* 2000; 18: 2607–2614.
32. Stoppa-Lyonnet D, Soulier J, Lauge A et al. Inactivation of the ATM gene in T-cell prolymphocytic leukemias. *Blood* 1998; 91: 3920–3926.
33. Yamaguchi M, Yamamoto K, Miki T et al. T-cell prolymphocytic leukemia with der(11)t(1;11)(q21;q23) and ATM deficiency. *Cancer Genet Cytogenet* 2003; 146: 22–26.
34. Narducci MG, Virgilio L, Isobe M et al. TCL1 oncogene activation in preleukemic T cells from a case of ataxia-telangiectasia. *Blood* 1995; 86: 2358–2364.
35. Soulier J, Pierron G, Vecchione D et al. A complex pattern of recurrent chromosomal losses and gains in T-cell prolymphocytic leukemia. *Genes Chromosomes Cancer* 2001; 31: 248–254.
36. Stilgenbauer S, Schaffner C, Litterst A et al. Biallelic mutations in the ATM gene in T-prolymphocytic leukemia. *Nat Med* 1997; 3: 1155–1159.