

# UCSF

## UC San Francisco Previously Published Works

### Title

Tau pathology in neurodegenerative disease: disease mechanisms and therapeutic avenues

### Permalink

<https://escholarship.org/uc/item/1540k2ff>

### Journal

Journal of Clinical Investigation, 133(12)

### ISSN

0021-9738

### Authors

Samudra, Niyatee

Lane-Donovan, Courtney

VandeVrede, Lawren

et al.

### Publication Date

2023-06-15

### DOI

10.1172/jci168553

### Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

# Tau pathology in neurodegenerative disease: disease mechanisms and therapeutic avenues

Niyatee Samudra, Courtney Lane-Donovan, Lawren VandeVrede, and Adam L. Boxer

Memory and Aging Center, Department of Neurology, and Weill Institute for Neurosciences, UCSF, San Francisco, California, USA.

**Tauopathies are disorders associated with tau protein dysfunction and insoluble tau accumulation in the brain at autopsy. Multiple lines of evidence from human disease, as well as nonclinical translational models, suggest that tau has a central pathologic role in these disorders, historically thought to be primarily related to tau gain of toxic function. However, a number of tau-targeting therapies with various mechanisms of action have shown little promise in clinical trials in different tauopathies. We review what is known about tau biology, genetics, and therapeutic mechanisms that have been tested in clinical trials to date. We discuss possible reasons for failures of these therapies, such as use of imperfect nonclinical models that do not predict human effects for drug development; heterogeneity of human tau pathologies which may lead to variable responses to therapy; and ineffective therapeutic mechanisms, such as targeting of the wrong tau species or protein epitope. Innovative approaches to human clinical trials can help address some of the difficulties that have plagued our field's development of tau-targeting therapies thus far. Despite limited clinical success to date, as we continue to refine our understanding of tau's pathogenic mechanism(s) in different neurodegenerative diseases, we remain optimistic that tau-targeting therapies will eventually play a central role in the treatment of tauopathies.**

## Introduction

Tauopathies are neurodegenerative diseases defined by the accumulation of misfolded, insoluble tau protein aggregates in neuronal and/or glial inclusions detectable in the brain at autopsy (1). These disorders are associated with diverse cognitive, motor, and neuropsychiatric abnormalities (2). Since tau pathologic burden strongly correlates with the severity of neurodegeneration as well as clinical phenomenology, tau has been the focus of therapeutic development, with multiple tau-directed therapeutics evaluated in clinical trials of Alzheimer's disease and other tauopathies (3, 4).

Tau protein has a physiologic role as a soluble cytoplasmic protein interacting with microtubules, primarily through a microtubule-binding region (MTBR), to stabilize cytoskeleton and regulate axonal transport. It also affects a diverse array of other cellular processes, including synaptic function, gene expression, and energy metabolism (5). It is ubiquitously expressed in neurons (6–9). Tau's neurodegenerative role rests on multiple lines of evidence. These include genetic and autopsy data from human tauopathies, as well as nonclinical models of disease, such as induced pluripotent stem cell models and transgenic rodents that express mutant forms of tau associated with autosomal dominant frontotemporal dementia (FTD) (10, 11). Given

that tau loss of function in animal models does not replicate human clinical phenotypes, and genome-wide screens (GWAS) for disease-associated mutations only identify gain-of-function mutations in the tau-encoding gene, *MAPT*, toxic gain of function has been historically suggested as the cause of tauopathies (12). Abnormal tau protein folding has been thought to lead to cytotoxic tau aggregation, accumulation of insoluble tau deposits, and subsequent neuronal loss that correlates with the clinical features of tauopathies during life in clinical-autopsy and clinical-tau PET studies (13, 14).

Here, we review normal and abnormal tau biology, tau genetics, nonclinical models and their relationship to human disease, and hypotheses regarding proposed roles of tau in neurodegeneration. Despite strong evidence for a central pathologic role of tau in neurodegenerative tauopathies, recent human clinical trials of experimental tau-targeting therapies have failed to demonstrate clinical benefit, including drugs purported to interfere with pathologic aggregation, processing, and accumulation of tau (15, 16).

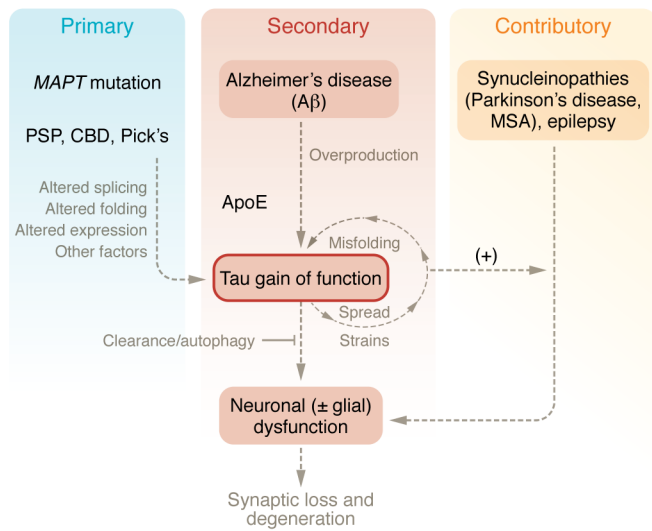
Despite a variety of potential explanations for the negative clinical trials, the lack of efficacy of tau therapies tested to date has raised important questions regarding what is truly understood about tau and its suitability as a drug target in human neurodegenerative disease. Though tau therapeutics have improved pathology in nonclinical models of tauopathy, do the recent negative human clinical trial results reflect flawed nonclinical models that inadequately model human disease? Are the models potentially predictive of therapeutic effects in human disease, but not the disease(s) in which the therapies were tested? Or is the lack of human efficacy explained by problems related to drug development, such as the wrong therapeutic target, inadequate dose, or lack of target engagement? Given these challenges, is it time to rebalance the approach to development of tau therapeutics using nonclinical models and early-stage human clinical trials?

**Conflict of interest:** LV has provided consultation for Retrope. ALB has an ownership interest (stock) in Alector. He has received income as a consultant for AGTC, Alzprotect, Amylyx, Arkuda, Arrowhead, Arvinas, Aviado, Boehringer Ingelheim, Denali, Eli Lilly, GSK, Humana, Life Edit, Merck, Modalis, Oligomerix, Oscotec, Roche, Transposon, and Wave. He has received research support from Biogen and Eisai for serving as a site investigator for clinical trials as well as from Regeneron.

**Copyright:** © 2023, Samudra et al. This is an open access article published under the terms of the Creative Commons Attribution 4.0 International License.

**Reference information:** *J Clin Invest.* 2023;133(12):e168553.

<https://doi.org/10.1172/JCI168553>.



**Figure 1. The hypothesized role of tau in degeneration in various tauopathies (primary, secondary, and contributory).** In particular, gain of function may lead to misfolding and spread in secondary tauopathies. A similar process may contribute in other disorders (synucleinopathies, epilepsy) that are not classically considered tauopathies.

### Normal and abnormal tau biology

Six tau protein isoforms are encoded from the *MAPT* gene by alternative splicing of exons 2, 3, and 10 (17). In particular, exon 10 alternative splicing can generate isoforms with three or four MTBR repeats (3R or 4R tau) (18). In the normal human adult brain, there are approximately equal concentrations of 3R and 4R tau, and changes in this ratio are associated with several neurodegenerative tauopathies, most commonly a relative overexpression of 4R tau (19).

Tau is one of several neuronal proteins responsible for promoting cytoskeletal microtubule assembly and stability. It may also play other cellular roles through its ability to bind to nucleic acids, and its localization to the synapse and mitochondrial compartments (5). Normally, tau is soluble and natively unfolded, whereas it becomes insoluble when hyperphosphorylated, shifting toward polymerization owing to an increase in  $\beta$ -sheet structures, which is also seen in other protein deposition disorders (20). Abnormal hyperphosphorylation and lack of tau clearance in the disease state is associated with diverse intraneuronal and glial inclusions (21). Decreased reversibility of hyperphosphorylation may contribute to pathogenesis in some tauopathies (22). There are many other posttranslational modifications of tau, including O-GlcNAcylation, acetylation, and glycosylation, that may influence the function and pathology of tau (9, 23).

### Classification of tauopathies and conceptualization of tau dysfunction

Tauopathies are often classified based on the primary tau protein isoform deposited in the brain, including 3R tauopathies, exemplified by Pick's disease (PiD); 4R tauopathies, such as progressive supranuclear palsy (PSP), corticobasal degeneration (CBD), argyrophilic grain disease (AGD), and globular glial tauopathy (GGT); and combined 3R/4R tauopathies, such as

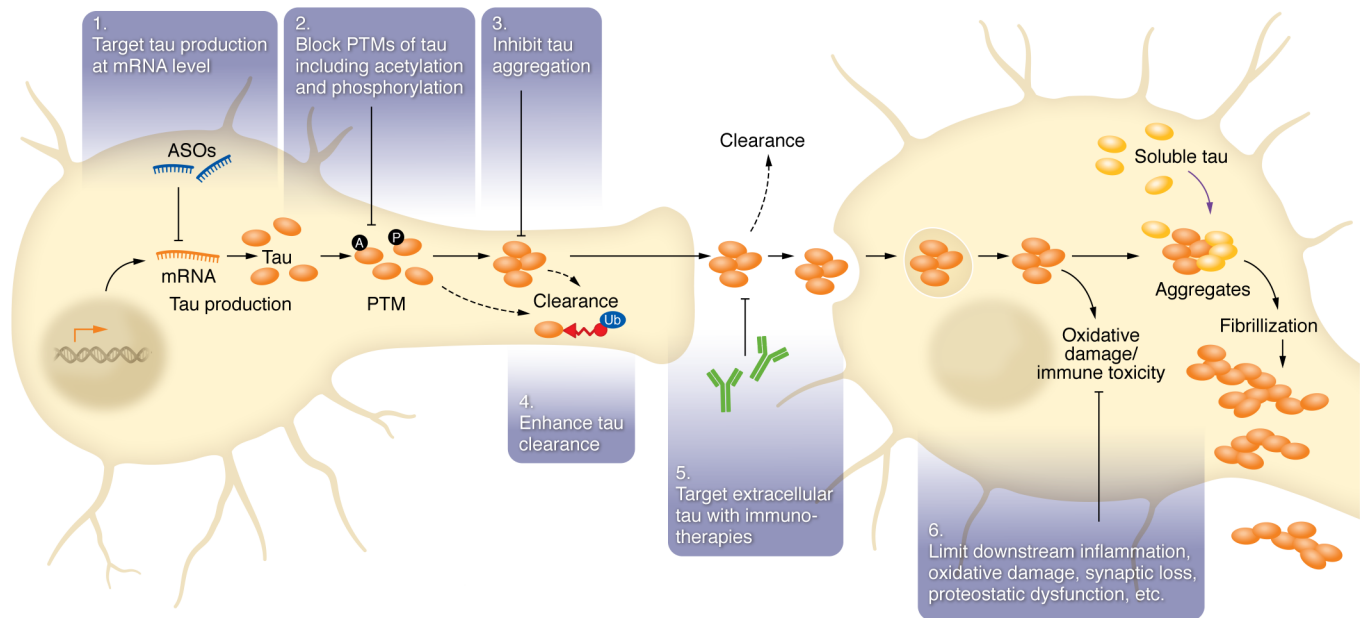
Alzheimer's disease (AD), chronic traumatic encephalopathy (CTE), and primary age-related tauopathy (PART).

Tauopathies can also be classified based on whether tau is the only aggregated protein found in the brain at autopsy or whether other proteins or pathogenic events are believed to initiate tau pathology. See Figure 1 for a summary. More than 20 different tauopathies have been identified, considered "primary," in which tau is the only pathogenic protein found at autopsy, or "secondary," in which tau pathology may accumulate due to the presence of another pathology (24).

Many data from cell culture and animal models, as well as human neuropathologic correlations, suggest that tau is likely to be a key pathogenic driver in most tauopathies. An alternate, less likely hypothesis is that tau pathology is a permissive factor or an epiphenomenon that correlates with disease pathophysiology (25). Tauopathies where tau abnormalities definitely cause disease are autosomal dominant *MAPT* mutations, which lead to hereditary forms of frontotemporal lobar degeneration. Different *MAPT* mutations are associated with specific clinical phenotypes and biomarker profiles (26). Intron 10 (IVS10) and other *MAPT* mutations that increase 4R tau production often lead to movement disorder phenotypes similar to sporadic PSP or CBD (27). The strongest genetic risk factors for these sporadic primary tauopathies are in and around the *MAPT* gene, including the H1c subhaplotype, which is believed to increase *MAPT* mRNA expression (28). These human genetic data strongly support a central role for tau protein pathogenesis in CBD and PSP. Separately, GWAS evidence suggests that tau may play a role in the pathogenesis of synucleinopathies, such as Parkinson's disease and multisystem atrophy, as well as certain forms of epilepsy, such as Dravet's syndrome (29).

The trans-synaptic spread ("prion") hypothesis of tau spread has garnered recent interest. This hypothesis is supported by the predictable progression intracerebrally of tau protein in various diseases, including AD, correlating with clinical symptoms (30). In AD, the spread of tau neurofibrillary tangles (NFTs) from entorhinal cortex to hippocampus to cortical regions prior to and in tandem with the development of clinical symptoms suggests tau's causal role (31). In animal and cell culture models, tau spreads in a prion-like manner, potentially explaining the stereotypical pattern of progression of tau accumulation in neurodegenerative diseases like AD (31–33). Seeding-based mouse models expressing human *MAPT* gene (wild type or mutant) have demonstrated conversion of tau monomers to oligomers, and then to insoluble fibrils (34). In these seeding paradigms, mice are injected with lysates from human disease brain, transgenic mouse brain, or in vitro tau aggregates. The seeding can induce tau aggregation and pathology, which can be accelerated by amyloid pathology or age (35, 36). Pattern of distribution and affected cell type can be distinct between each tau strain, often mirroring findings of the initial disease (e.g., oligodendrocyte tau pathology in CBD mice) (37).

It is more difficult to connect tau burden with clinical presentation in "incidental" tauopathies, which are often subclinical in nature, noted as co-pathologies or contributing pathologies in brain autopsies, with phosphorylated tau (p-tau) aggregates, and also termed age-related tauopathies (25). They include patholo-



**Figure 2. Review of mechanisms for various anti-tau therapeutics.** 1. Genetically targeted therapies, such as antisense oligonucleotides (ASOs) and certain small molecules, can target tau production. 2. Small-molecule enzyme inhibitors can target posttranslational modifications (PTMs) such as acetylation (A), phosphorylation (P), and ubiquitination (Ub). 3. Methylene blue derivatives and other aggregation inhibitors were conceived of as targeting tau aggregation. 4. Tau clearance may be enhanced by molecules such as PROTACs (see above). 5. Immunotherapies (vaccines, anti-tau monoclonal antibodies) target extracellular tau. 6. Neuroprotective agents, including antiinflammatory agents, could limit the downstream impacts of tau pathology. Figure adapted with permission from *Neuroscience Letters* (95) and from Martin Kampmann (UCSF) with permission.

gies akin to primary age-related tauopathy (PART), aging-related astroglial pathology (ARTAG), and argyrophilic grain disease (AGD). For example, AGD is often comorbid with AD, and has been associated with a prolonged period of amnesic mild cognitive impairment. However, in a significant proportion of cases it may be asymptomatic (38). The existence of these disorders challenges the dogma that NFTs are always necessarily pathogenic, rather than reactive or protective, as neurons with NFTs can survive for decades (39, 40).

### Alternative conceptualizations of tau pathogenicity

Though toxic gain of function has been hypothesized to cause tauopathies, loss of tau physiological function could also contribute (12). Tau protein interacts with more than a hundred targets, including presynaptic, postsynaptic, and mitochondrial proteins (5, 41). Depletion of tau in cells with drug-induced DNA damage increases cell senescence (42). Further, missense mutations in the *MAPT* gene reduce tau's ability to bind microtubules and promote microtubule assembly, causing an FTD with Parkinsonism phenotype (43).

Other possible mechanisms of pathogenicity relate to downstream effects of tau dysfunction. One potential unifying hypothesis is that age- and/or neurodegeneration-related loss of protein homeostasis leads to an inability to clear soluble tau species that may be pathogenic (44). There is evidence that tau acetylation leads to failed tau clearance by chaperone-mediated autophagy (45, 46). Nucleocytoplasmic and mitochondrial transport may also be impaired by AD-related tau (47, 48).

Neuroinflammation related to tauopathy may also be an important mechanism leading to the development or progression of neurodegenerative disease (49, 50). Tau transgenic mice demonstrate colocalization of tau oligomers with astrocytes, microglia, and inflammatory cytokines (51). Moreover, it was recently shown that tauopathy mouse models have increased parenchymal cytotoxic T cells and microglia, and that depletion of either cell population prevents tau-mediated brain atrophy (50). Autophagy, mitophagy (the specific or selective removal of mitochondria), and neuroinflammation could have a synergistic effect in the development of tauopathy, particularly in AD (52).

Other possible routes of pathogenicity include the interaction of tau with other proteins involved in neurodegenerative disease. Amyloid- $\beta$  (A $\beta$ ) and tau in AD have a pathogenic interaction in human disease (53). In an AD mouse model expressing both human pathologies, tau and A $\beta$  had opposite effects on cortical hyperactivity, and tau gene suppression was ineffective in rescuing neuronal impairments, suggesting a complex interaction (54). Phase III trials in AD have suggested some efficacy of the A $\beta$ -targeting antibodies lecanemab and aducanumab in slowing rates of cognitive decline. Preliminary phase II trials with donanemab also demonstrate lowering of plasma p-tau, suggesting a downstream effect of these agents on AD tau pathology (55–57). Parallels between changes in plasma p-tau species and glial fibrillary acidic protein (GFAP) species in recent anti-amyloid antibody trials (phase II in donanemab and phase III in lecanemab) raise the possibility that astroglial activation may mediate the interaction between A $\beta$  plaques and soluble p-tau accumulation in AD. Further evidence from a human presenilin-1 (*PSEN1*) mutation car-

**Table 1. Summary of clinical trials of potential therapeutic agents targeting tau**

Agent	Mechanism	Population	Phase	Trial identifier	Status
Lithium	Anti-GSK-3 $\beta$	PSP, CBS	I/II	NCT00703677	Negative, not tolerated
Valproate	Anti-GSK-3 $\beta$	PSP	II	NCT00385710	Negative, harmful
Tideglusib	Anti-GSK-3 $\beta$	Mild-moderate AD, PSP	II	NCT01350362, NCT01049399	Negative, safe
Saracatinib	Fyn inhibitor	Mild AD	II	NCT02167256	Negative
LY3372689	OGA-targeting agent	AD	II	NCT05063539	Active
Salsalate	Acetylation inhibitor	PSP, mild-moderate AD	II	NCT02422485, NCT03277573	Negative, pending
Methylene blue	Tau aggregation inhibitor	bvFTD, AD	III	NCT03446001, NCT01626378	Negative
Davunetide	Microtubule stabilization	PSP, AD (MCI)	II/III	NCT01110720	Negative
Abeotaxane (TPI-287)	Microtubule stabilization	AD, 4R tauopathies	I	NCT02133846	Negative, caused harm
Levetiracetam	Hyperexcitability reduction	AD	II	NCT02002819	Completed, exploratory benefit; results pending in MCI
BIBD80	Tau ASO	PSP, mild AD	I/II	NCT05399888, NCT04539041	Pending, safe/well tolerated
NI0752	Tau ASO	PSP, early AD	I/Ib	NCT04539041, NCT05469360	Pending
AADvac1	Tau-directed vaccine	AD, nfvPPA	II/I	NCT03174886	Negative in AD, running in nfvPPA
ACI-35	Liposomal vaccine	AD	I/II	NCT04445831	Active
Gosuranemab	Tau N-terminal antibody	PSP, early AD, CBS	II	NCT03352557, NCT03068468, NCT03658135	Negative
Tilavonemab	Tau N-terminal antibody	PSP, AD	II	NCT03413319	Negative
Zagotenemab	Tau N-terminal antibody	AD	II	NCT03518073	Negative
Semorinemab	Tau N-terminal antibody	AD	II	NCT03289143	Negative (see text)
Various mid-region-, MTBR-, and C-terminal-targeting antibodies (bepranemab [UCB0107], E2814, LuAF87908, JNJ-63733657)	See text	PSP, AD	I/II	NCT04658199, NCT03375697, NCT04149860, NCT04971733	Active

bvFTD, behavioral variant frontotemporal dementia; CBS, corticobasal syndrome; MCI, mild cognitive impairment; nfvPPA, non-fluent variant primary progressive aphasia.

rier with a protective apolipoprotein E (apoE) mutation who had reduced tau accumulation and preserved cognition also implicates apoE in this process (58).

Tau,  $\alpha$ -synuclein, and TAR DNA-binding protein 43 (TDP-43) appear to have synergistic neurotoxic effects, based on their colocalization in humans at autopsy and in vivo model data (59, 60). Based on spectroscopic analysis, there may be synergistic aggregation between tau and  $\alpha$ -synuclein molecules that contributes to neural cytotoxicity (61). Co-pathology of various proteinopathies is very common in neurodegenerative disease and increases with age. This has therapeutic implications in tauopathies since the presence of co-pathologies could mask tau-specific therapeutic effects (62).

## Tau genetics

Mutations in *MAPT* are the cause of autosomal dominant forms of frontotemporal lobar degeneration (FTLD) that present most commonly with behavioral variant FTD, but sometimes with movement disorders. Other mutations, including R406W and V337M, produce mixed 3R/4R tau pathology similar to AD, presenting with an amnesic AD-like syndrome and tau that binds AD tau PET tracers. Overall, nearly 60 mutations in *MAPT* have been identified as pathogenic (63, 64).

A chromosomal inversion in the *MAPT* region defines two major tau haplotypes, H1 and H2. Various reports have mentioned different possible effects of H1 and H2 haplotypes on age of onset of or risk for different neurodegenerative diseases, either alone or

in combination with other genes. For example, the combination of H1 haplotype and apolipoprotein E (*APOE*)  $\epsilon$ 4 allele may increase risk of earlier-onset FTD (65). Many patients with PSP carry the H1 haplotype (66). A subhaplotype of H1, H1c, is linked to PSP and CBD (67, 68). It is also possible that the H2 haplotype may be protective against PSP and CBD, although the mechanisms are not well defined (69, 70). GWAS have also identified shared risk between CBD and PSP at different gene loci that do not involve the *MAPT* gene, including *MOBP*, *CXCR4*, *GLDC*, and *EGFR* (71).

Importantly, tau mutations can be influenced by other genetic and epigenetic factors and may result in heterogeneous clinical syndromes that cannot be well replicated in nonclinical models (72). Because the tau protein sequence is not different between the H1 and H2 haplotypes, pathogenic effects may relate to differences in gene expression or post-transcriptional changes (73). In addition, the association between H1 haplotype and PSP is of somewhat uncertain global significance given the variable haplotype expression in different groups; for example, the H2 haplotype is not present in many Asian populations (74). Further, the H1 haplotype associated with PSP in non-Latinx White populations was not associated with these symptoms in Guadeloupean patients (75).

## Nonclinical (cell culture and animal) tauopathy models

Most evidence in support of tau-targeting therapies is based on experiments in nonclinical models. Historically, tau transgenic mice have been used because they have CNS cell types similar to those

of humans and allow for in vivo manipulation of cellular systems. Unfortunately, the predictive value of therapeutic efficacy in mouse models is limited, as large therapeutic effects seen in tau transgenic mouse models have not been replicated in human clinical trials.

Mouse models largely do not develop the neurodegeneration and insoluble tau pathology seen in humans (76, 77). There is no evidence of murine tau fibril formation with age, and very few mouse models accumulate endogenous murine A $\beta$  (78, 79). As a result, transgenic mouse models to study neurodegeneration must express mutant human proteins that lead to rare, severe early-onset disease in humans. A further discrepancy, specifically for AD, is that even the most aggressive mouse models of A $\beta$  accumulation and early plaque development (e.g., the 5xFAD mouse) do not develop secondary murine tau tangle formation or substantial neuronal loss, as seen in human AD (80).

The most common transgenic tau mouse models express familial FTLN-associated (but not AD-associated) *MAPT* mutations (e.g., P301L, P301S) (81–83). These models accumulate hyperphosphorylated tau fibrils and develop phenotypically variable age-dependent synaptic dysfunction, cognitive impairment, and neurodegeneration. Restricting expression of mutated human tau to entorhinal cortex via genetic manipulations results in propagation of tau along connected limbic structures, supporting the “prion-like” hypothesis of tau spread (35). However, there are numerous limitations with these mouse models. First, transgenic mice often express much higher levels of mutated tau throughout the lifespan, which might induce compensatory changes that could either mask pathology or cause phenotypes irrelevant to human disease. Additionally, transgenic tau models only express one of six potential tau isoforms, typically a 4R tau, thus eliminating any potential contribution of alternative splicing or 3R/4R ratios to disease processes. As well, many tau models produce specific tau aggregate strains, which may not be relevant to the human disease in which a particular therapy is eventually tested.

With this knowledge, close attention should be paid to recent tau-directed antibody failures. For example, nonclinical testing of semorinemab was in a mouse model expressing P301L tau, a mutation found in FTLN (usually a behavioral variant FTD phenotype), with subsequent clinical testing in AD patients (84). Conversely, tilavonemab was tested in the P301S model, which expresses an FTLN-only mutation, and then tested in mild-to-moderate PSP and AD patients (4, 85). If tau strain or aggregate structure is key to the development of a particular human disease, failure to accurately target the relevant tau strain could result in a lack of efficacy in human trials.

Other key differences exist between mice and humans that may explain poor translation of mouse tau biology to therapeutics. Murine tau lacks 11 N-terminal amino acids that are present in the human version. These differences in the N-terminus affect tau secretion, protein interaction, and tau phosphorylation (recently reviewed in ref. 86) and may limit the ability of mice to recapitulate nuanced features of disease critical to the development of therapeutics (86). On an organismal level, there are key and relatively unexplored differences in CNS function between mice and humans. For example, microglia may contribute to neurodegenerative disease and show transcriptomic differences between mouse and human, particularly with age (87). Similarly, the blood-brain barrier transcriptome differs

between mice and humans, which may impact both disease pathophysiology and the action of peripherally administered drugs (88). In consequence, nonclinical models of tauopathy have at best partially approximated human neurodegeneration — they represent models of possibly relevant disease mechanisms.

Potential alternative models to study tauopathies are in development, including the seeding-based mouse models referenced above. Narasimhan et al. injected pathologic tau from postmortem brains into non-transgenic mouse brains and observed differences in tau strain potency and pathologic localization between AD-tau, CBD-tau, and PSP-tau, such that only PSP-tau and CBD-tau produced glial inclusions, and PSP-tau produced much more extensive tau pathology (89). These models may provide a tool in our arsenal to study the effects of tau treatments on specific aspects of pathology, and tau monoclonal antibodies (mAbs), such as gosuranemab, have been tested in induced pluripotent stem cell (iPSC) cultures seeded with disease-specific tau (90). To address contributions of multiple cell types and aging in vitro, tissue culture methods are becoming increasingly sophisticated. Organoids allow the coculture of multiple human cell types derived from iPSCs, and thus can model the interactions of human microglia, astrocytes, and neurons in vitro (91). Similarly, newer techniques to directly convert patient-derived skin fibroblasts into neurons (iNeurons) bypass the need for an iPSC step and maintain the aging signature of the sample patient skin biopsy (92).

A few rat models of AD also exist that more closely resemble human disease. Specifically, rats expressing mutated human APP develop age-dependent tau pathology and neurodegeneration (93, 94). More research is needed to understand why rats more accurately recapitulate human disease. While the increased cost of housing rats limits their use in many laboratories, rat models may prove a more useful tool for testing therapeutics.

## Therapeutics targeting tau

Tau therapies have attempted to disrupt toxic gain of function (antisense oligonucleotides/gene therapy), modulate posttranslational modification (PTM), disrupt tau aggregation, passively clear tau, and vaccinate against tau — see Figure 2 for a summary of the classes of therapeutic approaches (95). Conversely, approaches to replace loss of tau physiologic function (microtubule stabilizers) have also been assessed. Though these multiple classes of therapies have been evaluated as disease-modifying agents in human clinical trials (Table 1), therapeutically relevant mechanisms have not been validated.

Notably, the pathogenic tau species has not been definitively identified in living humans. Soluble tau, in the form of oligomers (including dimers), is being explored as a possible source of key neurotoxic species. Alternatively, insoluble tau in the form of both NFTs and other aggregates might represent the toxic species (96). In support of oligomeric soluble tau being important, injection of soluble tau oligomers into wild-type mouse brains, but not injection of tau fibrils or monomers, impaired memory (97).

In general, given the heterogeneity of tau isoforms, tau PTMs, and aggregate structures in tauopathies, some diseases may respond better than others to specific tau-targeting agents. There is cryo-electron microscopic evidence for differences in the structures of tau filaments in different diseases, including Pick's disease, AD, chronic traumatic encephalopathy (CTE), CBD, globular

glial tauopathy (GGT), AGD, and PSP. Particularly, a three-layered fold is noted in PSP and GGT, while a four-layer fold is noted in CBD and AGD (98, 99). Tau seeding models, mentioned above, support the idea that there are differences in tau conformers between pathologies. Experiments involving inoculation of human brain lysates from various tauopathies have revealed brain (neuronal or glial) lesions in mouse models or cell culture that differentially resemble the original human pathology (100). These differences might contribute to differences in efficacy, safety, and tolerability in treatments across tauopathies, as seen in a recent basket trial testing a single intervention in multiple disease groups expressing a common biomarker of a microtubule stabilizer (101).

**Small-molecule PTM inhibitors.** Agents targeting tau PTMs, particularly hyperphosphorylation, have included protein kinase inhibitors that aim to reduce tau aggregation. All of the agents discussed below demonstrated signal in nonclinical models. Concerns with these agents have included potential lack of target specificity and potential for off-target effects. Over 90 phosphorylation sites for tau exist, and specific interventions balancing efficacy with tolerability may be difficult to achieve. Glycogen synthase kinase 3 $\beta$  (GSK-3 $\beta$ ) hyperactivity contributes to hyperphosphorylation, which has been considered the major target for pathologic aggregation (102). Lithium inhibits GSK-3 $\beta$  and was evaluated in 17 patients with PSP and corticobasal syndrome; however, it was poorly tolerated due to increased falls, and therefore the trial was stopped (ClinicalTrials.gov NCT00703677). Valproate was also assessed because of anti-GSK-3 $\beta$  activity, but did not improve PSP Rating Scale scores in 28 PSP patients over the course of 2 years (ClinicalTrials.gov NCT00385710) (103). Tideglusib, a novel small-molecule GSK-3 $\beta$  inhibitor, did not demonstrate evidence of efficacy in mild to moderate AD (ARGO, NCT01350362) or in PSP (TAUROS, NCT01049399) (104, 105). Another kinase implicated in tau hyperphosphorylation, Fyn, has been targeted by a small-molecule inhibitor (saracatinib) in a phase II trial of patients with mild AD (CONNECT, NCT02167256), which was stopped for lack of clinical efficacy and concern for gastrointestinal side effects (106).

O-GlcNAcylation (OGA) targeting may decrease hyperphosphorylation, and a small-molecule inhibitor (MK-8719) showed nonclinical mouse model signal in decreasing tau aggregation, but did not advance to phase II clinical trials in humans (107). Other OGA-targeting agents also await evaluation in phase II studies; however, LY3372689 is currently in a phase II AD trial (NCT05063539).

Tau acetylation can prevent physiologic clearance; salsalate, a small-molecule acetylation inhibitor, did not show a treatment effect in a futility study of 10 patients with PSP, nor was there evidence of efficacy in a small randomized, placebo-controlled trial in mild to moderate AD (presented in abstract form at the Clinical Trials on Alzheimer's Disease conference in 2022) (108, 109).

Tau aggregation disruption aims to prevent the paired helical filament conformation observed in NFTs, and a derivative of methylene blue (LMTM), which prevents this in mouse models, was evaluated in a phase III trial of behavioral variant FTD without evidence of efficacy (110). Multiple phase III trials in AD, most recently LUCIDITY (NCT03446001), have also been negative based on prespecified analyses.

Microtubule stabilization designed to ameliorate putative loss of physiologic function has been attempted. Davunetide is derived from activity-dependent neurotrophic protein (ADNP), a neuroprotective agent that decreased hyperphosphorylated tau in non-clinical models through an unclear mechanism. It did not demonstrate any clear benefits in randomized trials in 144 patients with mild cognitive impairment nor in 313 patients with PSP (111, 112). Abeotaxane (TPI-287), a microtubule stabilizer, produced anaphylactoid reactions in patients with AD but not PSP in a basket-design clinical trial with patients with AD and 4R tauopathies; it also led to a dose-related worsening of function and more frequent falls in 4R tauopathies (101).

Antisense oligonucleotides (ASOs) are directed against *MAPT* mRNA to reduce tau expression. This strategy is based on data in mouse models showing that reducing human tau expression improves hippocampal volume loss and cognitive deficits (113). In this same work, CNS penetration was demonstrated in primate models. Results are pending in a study of tau lowering with ASO (BIIB080) in 64 patients with mild AD, but preliminarily ASO therapy reduced tau levels in cerebrospinal fluid (CSF), reduced MK-6240 tau PET uptake, and was well tolerated (114). A phase II trial in AD is now enrolling (NCT05399888), and a similar phase I trial with a different tau ASO is ongoing in PSP (NCT04539041).

Improvement of tau clearance has been assayed using proteolysis-targeting chimera (PROTAC) molecules to selectively enhance ubiquitination and proteolysis of tau proteins, as demonstrated in nonclinical models, including patient-derived neural cell models (115, 116). There are no currently running human clinical trials.

**Immune therapies.** Both active (vaccine) and passive (mAb-mediated) immune therapies are being investigated in tauopathies.

AADvac1 was the first tau-directed vaccine tested in trials, employing a truncated version of the tau protein that was thought to be the pathogenic fragment in the MTBR triggering aggregation. Immunogenicity was demonstrated in a phase I trial, but unfortunately in a phase II trial versus placebo in mild AD dementia, slowing of cognitive and functional decline was not demonstrated, although it slowed the increase in blood neurofilament light chain (117). Another trial of a liposome-based vaccine (ACI-35) targeted toward pathologic phosphorylation residues is under way (118).

Monoclonal antibodies targeting the N-terminal tau domain have been tested in multiple phase II trials, largely without clinical benefit despite evidence for target engagement via reduction of N-terminal CSF tau. These trials have included gosuranemab (in PSP and early AD), tilavonemab (in PSP and AD), and zagotenemab (119). Notably, in a trial of mild-to-moderate AD (in contrast to prodromal to mild AD), semorinemab, also an N-terminal IgG4 antibody, led to a 43.6% slowing of decline on the Alzheimer's Disease Assessment Scale-Cognitive Subscale (ADAS-Cog) co-primary outcome measure, in the absence of benefit for the other cognitive or functional outcomes. Whether this was due to chance or a true therapeutic effect is a topic of debate (phase II LAURINET trial, NCT03828747). If true, it is unclear why a therapeutic effect was absent in an earlier phase of AD, but it could be hypothesized that different species of tau more amenable to semorinemab engagement predominate in later stages of the disease; there may be higher concentrations of N-terminal tau fragments in later-stage disease if they are related to the overall amount of cortical

tau pathology (soluble or insoluble). In addition, the mid-region, MTBR, and C-terminal tau-targeting antibodies bepranemab, E2814, LuAF87908, and JNJ-63733657 are in phase I-II trials and may have better clinical effect given the importance of the MTBR and C-terminus in tau aggregate structure.

Another possible reason for the lack of observed clinical benefit in trials is that mAbs have targeted extracellular tau. This mechanism was thought to be valuable on the basis that extracellular tau may undergo spread to other neurons (as demonstrated in nonclinical models). However, it is unknown whether recent tau mAbs have reached a high enough concentration in the brain parenchyma to affect these species, since there are no human biomarkers to measure soluble tau levels in the brain parenchyma. By analogy to anti-amyloid antibodies, it may be necessary to activate immune-mediated clearance for efficacy, but most anti-tau mAbs tested have been IgG4 with reduced effector domain, which is the least effective isotype to promote microglia phagocytosis (120). Further, to bypass systemic circulation and ensure cerebral delivery at correct levels, adeno-associated viral antibody delivery may be an avenue (121). Interestingly, recent work demonstrated that tau immunotherapy may rely on the intracellular antibody receptor TRIM21 (122). Mice lacking expression of TRIM21 were nonresponsive to tau-targeting immunotherapy both at an early stage of tau pathogenesis and during prolonged treatment, which may have implications for tau mAb treatment in human disease. Optimization of antibody characteristics, including isotype, epitope, charge, affinity, size, vehicle, and timing of delivery, may also be important for identifying an efficacious approach (120).

It is also important to consider patient effects, including aging, on changes in effectiveness of immune therapies such as vaccination and antibody therapies, related to alterations in the B and T cell compartments termed immunosenescence (123). Decreased tau clearance related to aging, such as through the glymphatic system, may also be therapeutically relevant; even if tau is targeted appropriately by therapies, it may still not be cleared (124). These phenomena should be accounted for in designing such therapies in tauopathies, perhaps with dose and schedule differences (125).

## Tau biomarkers

In recent years, multiple *in vivo* biomarkers for tau pathology have been evaluated; these are key to detection of tau pathology, clinical trial enrollment, and assessment of the efficacy of tau therapeutics. These are overall better validated in AD than in other tauopathies. Currently, no biomarkers are approved for diagnosing non-AD tauopathies or for following the clinical course of any tauopathies.

The first tau PET tracer, [<sup>18</sup>F]flortaucipir, was approved for clinical use for the detection of AD by the US Food and Drug Administration (FDA) in May 2020 (126). This tracer was less sensitive to tau related to FTL spectrum disorders (127). Newer tau PET tracers, including [<sup>18</sup>F]PI2620, likely bind more selectively to hippocampal tau related to AD but may also have utility for identifying 4R tauopathies (128, 129).

Fluid biomarkers for tau pathology include serum and CSF tau assays. Plasma p-tau181, p-tau217, and p-tau231 are promising and potentially more easily accessible biomarkers (130). In particular,

plasma p-tau217 has shown utility in combination with tau PET for staging AD pathology (131, 132). Elevated CSF total tau and p-tau (most commonly p-tau181) are also suggestive of an AD pathology (133). Combining different markers, including A $\beta$  and neurofilament light chain, can yield better discriminability of CSF tau for FTL spectrum disorders (134). CBD may be distinguished from other tauopathies by incorporation of differences in specific CSF MTBR tau fragments, a finding that should be further explored (135).

Distinguishing AD from other tauopathies or identifying when they co-occur is important, as co-occurrence is common and may have therapeutic implications. One goal for future research is to design tau biomarkers with increased sensitivity and specificity for the early differential diagnosis of tauopathies and their longitudinal progression.

## Next steps in designing tau-targeting therapies

In this Review, we have outlined multiple potential reasons for the lack of success to date in the tau-targeting therapies that have come to human clinical trials. These include poorly predictive nonclinical models, an inability to relate specific models to specific human diseases, targeting of the wrong tau species (N-terminal tau) or pathogenic mechanism (phosphorylation), difficulty in designing the optimal immunologic approach, lack of biomarkers to diagnose early-stage tauopathies and to measure treatment response, the possibility that recent trials have started too late in the course of disease, and insufficient numbers of clinical trials in different human tauopathies that could respond differently to the same tau therapy.

It is clear that a novel approach to identifying and testing therapies in humans is needed. We know that nonclinical models are imperfect and that some phenomena studied in these models may not be therapeutically relevant to humans. Further, secondary effects of tau pathology, such as aggregation of other pathogenic proteins and neuroinflammation, may not be addressed by therapies that solely target tau. Timing is also critical: it is possible that even mild cognitive impairment is too late with regard to the development of pathology leading to neurodegeneration in humans. Or perhaps, as suggested by semorinemab's failure to slow progression of early and mild AD, it is too early?

There is an urgent need to bring therapies to the clinic setting for all patients with neurodegenerative disease, including tauopathies. We believe it is time to refocus on "interventional" human research, as a departure from the current focus on therapeutic design, which entails years of expensive work on nonclinical models. Once therapeutic safety is established in early-phase trials, new approaches will be necessary to efficiently and effectively evaluate multiple therapeutic mechanisms in parallel. Training and departmental support for academic clinical trialists to carry out this work should be prioritized. Therapeutic classes and agents targeting different mechanisms can be tested in basket trials to enhance drug development efficiency by evaluating the effects of one therapy in multiple tauopathies (136). Umbrella trials of multiple agents in one disease also have utility in tauopathies, as exemplified by the current combination trial of anti-amyloid (lecanemab) and anti-tau (E2814) treatments in dominantly inherited AD (DIAN-TU) (137). Pragmatic trials, which assess effectiveness in the real-world clinic setting, of existing, repurposable drugs,



such as symptom-targeting medications, have been successfully conducted in other neurologic conditions and might also be prioritized (138). Disease progression models of existing data have been applied in rare familial FTD (*MAPT* mutation carriers) to leverage surrogate biomarker endpoints (neurofilament light chain and MRI) to select the optimal inclusion criteria and endpoints to maximize power to detect treatment effects (139).

There are gaps in our understanding of the pathobiology of tauopathies, but regardless, an overwhelming amount of circumstantial evidence implicates tau protein as a driver of human disease, particularly in the primary tauopathies. As the science of tau therapy and clinical trials advances, there are likely to be important and unexpected insights into the pathogenic mechanisms of tauopathies that will identify novel agents that should be efficiently tested in clinical trials. Overall, we are optimistic about the future of tau-targeted therapies and our ability as a field to bring them to patients, as we continue to refine our understanding of tau biology and drug development.

## Acknowledgments

NS is supported by the NIH (training grant T32AG023481). CLD receives support from National Institute of Neurological Disorders and Stroke (R25NS070680). LV reports research support from the Alzheimer's Association, the American Academy of Neurology, the American Brain Foundation, and the NIH (K23AG073514). ALB has received research support from the National Institutes on Aging (NIH U19AG063911, R01AG073482, R56AG075744, R01AG038791, RF1AG077557, R01AG071756, U24AG057437), Rainwater Charitable Foundation, Bluefield Project to Cure FTD, GHR Foundation, Alzheimer's Association, Association for Frontotemporal Degeneration, Gates Ventures, Alzheimer's Drug Discovery Foundation, UCSF Parkinson's Spectrum Disorders Center, and the University of California Cures AD Program.

Address correspondence to: Adam Boxer, UCSF Memory and Aging Center, Box 1207, 675 Nelson Rising Lane, Suite 190, San Francisco, California 94143, USA. Email: adam.boxer@ucsf.edu.

- Orr ME, et al. A brief overview of tauopathy: causes, consequences, and therapeutic strategies. *Trends Pharmacol Sci.* 2017;38(7):637–648.
- Olfati N, et al. Clinical spectrum of tauopathies. *Front Neurol.* 2022;13:944806.
- Arriagada PV, et al. Neurofibrillary tangles but not senile plaques parallel duration and severity of Alzheimer's disease. *Neurology.* 1992;42(3 pt 1):631–639.
- Yanamandra K, et al. Anti-tau antibodies that block tau aggregate seeding in vitro markedly decrease pathology and improve cognition in vivo. *Neuron.* 2013;80(2):402–414.
- Tracy TE, et al. Tau interactome maps synaptic and mitochondrial processes associated with neurodegeneration. *Cell.* 2022;185(4):712–728.
- Weingarten MD, et al. A protein factor essential for microtubule assembly. *Proc Natl Acad Sci U S A.* 1975;72(5):1858–1862.
- Cleveland DW, et al. Purification of tau, a microtubule-associated protein that induces assembly of microtubules from purified tubulin. *J Mol Biol.* 1977;116(2):207–225.
- Brozakis ZF, et al. A structural ensemble of a Tau-microtubule complex reveals regulatory Tau phosphorylation and acetylation mechanisms. *ACS Cent Sci.* 2021;7(12):1986–1995.
- Wang Y, Mandelkow E. Tau in physiology and pathology. *Nat Rev Neurosci.* 2016;17(1):22–35.
- Kretschmar H. Brain banking: opportunities, challenges and meaning for the future. *Nat Rev Neurosci.* 2009;10(1):70–78.
- Williams DR, et al. Pathological tau burden and distribution distinguishes progressive supranuclear palsy-parkinsonism from Richardson's syndrome. *Brain.* 2007;130(pt 6):1566–1576.
- Medina M, et al. New features about Tau function and dysfunction. *Biomolecules.* 2016;6(2):21.
- Soto C, Estrada LD. Protein misfolding and neurodegeneration. *Arch Neurol.* 2008;65(2):184–189.
- Ossenkoppele R, et al. Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. *Brain.* 2016;139(pt 5):1551–1567.
- Imbimbo BP, et al. Initial failures of anti-tau antibodies in Alzheimer's disease are reminiscent of the amyloid- $\beta$  story. *Neural Regen Res.* 2023;18(1):117–118.
- Silva MC, Haggarty SJ. Tauopathies: deciphering disease mechanisms to develop effective therapies. *Int J Mol Sci.* 2020;21(23):8948.
- Bachmann S, et al. Differential effects of the six human TAU isoforms: somatic retention of 2N-TAU and increased microtubule number induced by 4R-TAU. *Front Neurosci.* 2021;15:643115.
- Goedert M, et al. Multiple isoforms of human microtubule-associated protein tau: sequences and localization in neurofibrillary tangles of Alzheimer's disease. *Neuron.* 1989;3(4):519–526.
- Kovacs GG. Tauopathies. *Handb Clin Neurol.* 2017;145:355–368.
- Wilson DM, Binder LI. Polymerization of microtubule-associated protein Tau under near-physiological conditions. *J Biol Chem.* 1995;270(41):24306–24314.
- Ferrer I, et al. Glial and neuronal tau pathology in tauopathies: characterization of disease-specific phenotypes and tau pathology progression. *J Neuropathol Exp Neurol.* 2014;73(1):81–97.
- Duquette A, et al. Similarities and differences in the pattern of Tau hyperphosphorylation in physiological and pathological conditions: impacts on the elaboration of therapies to prevent Tau pathology. *Front Neurol.* 2020;11:607680.
- Alquezar C, et al. Tau post-translational modifications: dynamic transformers of tau function, degradation, and aggregation. *Front Neurol.* 2020;11:595532.
- Josephs KA. Current understanding of neurodegenerative diseases associated with the protein Tau. *Mayo Clin Proc.* 2017;92(8):1291–1303.
- Castellani RJ. The significance of Tau aggregates in the human brain. *Brain Sci.* 2020;10(12):972.
- Kwok JB, et al. The complex relationship between genotype, pathology and phenotype in familial dementia. *Neurobiol Dis.* 2020;145:105082.
- Morris HR, et al. Tau exon 10 +16 mutation FTDP-17 presenting clinically as sporadic young onset PSP. *Neurology.* 2003;61(1):102–104.
- Myers AJ, et al. The *MAPT* H1c risk haplotype is associated with increased expression of tau and especially of 4 repeat containing transcripts. *Neurobiol Dis.* 2007;25(3):561–570.
- Paudel YN, et al. Revisiting the impact of neurodegenerative proteins in epilepsy: focus on alpha-synuclein, beta-amyloid, and Tau. *Biology (Basel).* 2020;9(6):122.
- Mudher A, et al. What is the evidence that tau pathology spreads through prion-like propagation? *Acta Neuropathol Commun.* 2017;5(1):99.
- Braak H, Braak E. Staging of Alzheimer's disease-related neurofibrillary changes. *Neurobiol Aging.* 1995;16(3):271–278.
- Stancu IC, et al. Templated misfolding of Tau by prion-like seeding along neuronal connections impairs neuronal network function and associated behavioral outcomes in Tau transgenic mice. *Acta Neuropathol.* 2015;129(6):875–894.
- Kaufman SK, et al. Tau prion strains dictate patterns of cell pathology, progression rate, and regional vulnerability in vivo. *Neuron.* 2016;92(4):796–812.
- Mirbaha H, et al. Seed-competent tau monomer initiates pathology in a tauopathy mouse model. *J Biol Chem.* 2022;298(8):102163.
- de Calignon A, et al. Propagation of tau pathology in a model of early Alzheimer's disease. *Neuron.* 2012;73(4):685–697.
- He Z, et al. Amyloid- $\beta$  plaques enhance Alzheimer's brain tau-seeded pathologies by facilitating neuritic plaque tau aggregation. *Nat Med.* 2018;24(1):29–38.
- He Z, et al. Transmission of tauopathy strains is independent of their isoform composition. *Nat Commun.* 2020;11(1):7.
- Rodriguez RD, Grinberg LT. Arglyophilic grain disease: an underestimated tauopathy. *Dement Neuropsychol.* 2015;9(1):2–8.
- Morsch R, et al. Neurons may live for decades with neurofibrillary tangles. *J Neuropathol Exp Neurol.* 1999;58(2):188–197.
- Harada A, et al. Altered microtubule organization in small-calibre axons of mice lacking tau protein. *Nature.* 1994;369(6480):488–491.
- Klein C, et al. Process outgrowth of oligodendro-

- cytes is promoted by interaction of fyn kinase with the cytoskeletal protein tau. *J Neurosci.* 2002;22(3):698–707.
42. Sola M, et al. Tau affects P53 function and cell fate during the DNA damage response. *Commun Biol.* 2020;3(1):245.
  43. D'Souza I, et al. Missense and silent tau gene mutations cause frontotemporal dementia with parkinsonism-chromosome 17 type, by affecting multiple alternative RNA splicing regulatory elements. *Proc Natl Acad Sci U S A.* 1999;96(10):5598–5603.
  44. Krishnan S, et al. Activate or inhibit? Implications of autophagy modulation as a therapeutic strategy for Alzheimer's Disease. *Int J Mol Sci.* 2020;21(18):6739.
  45. Caballero B, et al. Acetylated tau inhibits chaperone-mediated autophagy and promotes tau pathology propagation in mice. *Nat Commun.* 2021;12(1):2238.
  46. Alquezar C, et al. TSC1 loss increases risk for tauopathy by inducing tau acetylation and preventing tau clearance via chaperone-mediated autophagy. *Sci Adv.* 2021;7(45):eabg3897.
  47. Eftekharzadeh B, et al. Tau protein disrupts nucleocytoplasmic transport in Alzheimer's disease. *Neuron.* 2018;99(5):925–940.
  48. Shahpasand K, et al. Regulation of mitochondrial transport and inter-microtubule spacing by tau phosphorylation at the sites hyperphosphorylated in Alzheimer's disease. *J Neurosci.* 2012;32(7):2430–2441.
  49. Wang C, et al. The effects of microglia-associated neuroinflammation on Alzheimer's disease. *Front Immunol.* 2023;14:1117172.
  50. Chen X, et al. Microglia-mediated T cell infiltration drives neurodegeneration in tauopathy. *Nature.* 2023;615(7953):668–677.
  51. Nilson AN, et al. Tau oligomers associate with inflammation in the brain and retina of tauopathy mice and in neurodegenerative diseases. *J Alzheimers Dis.* 2017;55(3):1083–1099.
  52. Liu X, et al. The emerging role of autophagy and mitophagy in tauopathies: from pathogenesis to translational implications in Alzheimer's disease. *Front Aging Neurosci.* 2022;14:1022821.
  53. Pickett EK, et al. Amyloid beta and tau cooperate to cause reversible behavioral and transcriptional deficits in a model of Alzheimer's disease. *Cell Rep.* 2019;29(11):3592–3604.
  54. Busche MA, et al. Tau impairs neural circuits, dominating amyloid- $\beta$  effects, in Alzheimer models in vivo. *Nat Neurosci.* 2019;22(1):57–64.
  55. McDade E, et al. Lecanemab in patients with early Alzheimer's disease: detailed results on biomarker, cognitive, and clinical effects from the randomized and open-label extension of the phase 2 proof-of-concept study. *Alzheimers Res Ther.* 2022;14(1):191.
  56. Pontecorvo MJ, et al. Association of donanemab treatment with exploratory plasma biomarkers in early symptomatic Alzheimer disease: a secondary analysis of the TRAILBLAZER-ALZ Randomized Clinical Trial. *JAMA Neurol.* 2022;79(12):1250–1259.
  57. van Dyck CH, et al. Lecanemab in early Alzheimer's disease. *N Engl J Med.* 2022;388(1):9–21.
  58. Arboleda-Velasquez JF, et al. Resistance to autosomal dominant Alzheimer's disease in an APOE3 Christchurch homozygote: a case report. *Nat Med.* 2019;25(11):1680–1683.
  59. Vacchi E, et al. Tau and alpha synuclein synergistic effect in neurodegenerative diseases: when the periphery is the core. *Int J Mol Sci.* 2020;21(14):5030.
  60. Latimer CS, Liachko NF. Tau and TDP-43 synergy: a novel therapeutic target for sporadic late-onset Alzheimer's disease. *Geroscience.* 2021;43(4):1627–1634.
  61. Dasari AKR, et al. Tau interacts with the C-terminal region of  $\alpha$ -synuclein, promoting formation of toxic aggregates with distinct molecular conformations. *Biochemistry.* 2019;58(25):2814–2821.
  62. Robinson JL, et al. Neurodegenerative disease concomitant proteinopathies are prevalent, age-related and APOE4-associated. *Brain.* 2018;141(7):2181–2193.
  63. Zhang CC, et al. The role of MAPT in neurodegenerative diseases: genetics, mechanisms and therapy. *Mol Neurobiol.* 2016;53(7):4893–4904.
  64. Dujardin S, et al. Different tau species lead to heterogeneous tau pathology propagation and misfolding. *Acta Neuropathol Commun.* 2018;6(1):132.
  65. Ingelson M, et al. Increased risk for frontotemporal dementia through interaction between tau polymorphisms and apolipoprotein E epsilon4. *Neuroreport.* 2001;12(5):905–909.
  66. Ghidoni R, et al. The H2 MAPT haplotype is associated with familial frontotemporal dementia. *Neurobiol Dis.* 2006;22(2):357–362.
  67. Pastor P, et al. Further extension of the H1 haplotype associated with progressive supranuclear palsy. *Mov Disord.* 2002;17(3):550–556.
  68. Höglinger GU, et al. Identification of common variants influencing risk of the tauopathy progressive supranuclear palsy. *Nat Genet.* 2011;43(7):699–705.
  69. Pittman AM, et al. Linkage disequilibrium fine mapping and haplotype association analysis of the tau gene in progressive supranuclear palsy and corticobasal degeneration. *J Med Genet.* 2005;42(11):837–846.
  70. Valentino RR, et al. MAPT subhaplotypes in corticobasal degeneration: assessing associations with disease risk, severity of tau pathology, and clinical features. *Acta Neuropathol Commun.* 2020;8(1):218.
  71. Yokoyama JS, et al. Shared genetic risk between corticobasal degeneration, progressive supranuclear palsy, and frontotemporal dementia. *Acta Neuropathol.* 2017;133(5):825–837.
  72. Ikeda A, et al. Clinical heterogeneity of frontotemporal dementia and Parkinsonism linked to chromosome 17 caused by MAPT N279K mutation in relation to tau positron emission tomography features. *Mov Disord.* 2019;34(4):568–574.
  73. Wolfe MS. The role of tau in neurodegenerative diseases and its potential as a therapeutic target. *Scientifica (cairo).* 2012;2012:796024.
  74. Conrad C, et al. Differences in a dinucleotide repeat polymorphism in the tau gene between Caucasian and Japanese populations: implication for progressive supranuclear palsy. *Neurosci Lett.* 1998;250(2):135–137.
  75. Camuzat A, et al. The PSP-associated MAPT H1 subhaplotype in Guadeloupean atypical parkinsonism. *Mov Disord.* 2008;23(16):2384–2391.
  76. Weber M, et al. Cognitive deficits, changes in synaptic function, and brain pathology in a mouse model of normal aging. *eNeuro.* 2015;2(5):ENEURO.0047-15.2015.
  77. Radulescu CI, et al. The aging mouse brain: cognition, connectivity and calcium. *Cell Calcium.* 2021;94:102358.
  78. Ahlemeyer B, et al. Endogenous murine amyloid- $\beta$  peptide assembles into aggregates in the aged C57BL/6J mouse suggesting these animals as a model to study pathogenesis of amyloid- $\beta$  plaque formation. *J Alzheimers Dis.* 2018;61(4):1425–1450.
  79. Suire CN, et al. Cathepsin D regulates cerebral A $\beta$ 42/40 ratios via differential degradation of A $\beta$ 42 and A $\beta$ 40. *Alzheimers Res Ther.* 2020;12(1):80.
  80. Oakley H, et al. Intraneuronal  $\beta$ -amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: potential factors in amyloid plaque formation. *J Neurosci.* 2006;26(40):10129–101240.
  81. Allen B, et al. Abundant tau filaments and nonapoptotic neurodegeneration in transgenic mice expressing human P301S tau protein. *J Neurosci.* 2002;22(21):9340–9351.
  82. Maeda S, et al. Expression of A152T human tau causes age-dependent neuronal dysfunction and loss in transgenic mice. *EMBO Rep.* 2016;17(4):530–551.
  83. Lewis J, et al. Neurofibrillary tangles, amyotrophy and progressive motor disturbance in mice expressing mutant (P301L) tau protein. *Nat Genet.* 2000;25(4):402–405.
  84. Lee SH, et al. Antibody-mediated targeting of tau in vivo does not require effector function and microglial engagement. *Cell Rep.* 2016;16(6):1690–1700.
  85. Yanamandra K, et al. Anti-tau antibody reduces insoluble tau and decreases brain atrophy. *Ann Clin Transl Neurol.* 2015;2(3):278–288.
  86. Hernández F, et al. Differences between human and murine tau at the N-terminal end. *Front Aging Neurosci.* 2020;12:11.
  87. Galatro TF, et al. Transcriptomic analysis of purified human cortical microglia reveals age-associated changes. *Nat Neurosci.* 2017;20(8):1162–1171.
  88. Song HW, et al. Transcriptomic comparison of human and mouse brain microvessels. *Sci Rep.* 2020;10(1):12358.
  89. Narasimhan S, et al. Pathological tau strains from human brains recapitulate the diversity of tauopathies in nontransgenic mouse brain. *J Neurosci.* 2017;37(47):11406–11423.
  90. Bright J, et al. Human secreted tau increases amyloid-beta production. *Neurobiol Aging.* 2015;36(2):693–709.
  91. Grenier K, et al. Three-dimensional modeling of human neurodegeneration: brain organoids coming of age. *Mol Psychiatry.* 2020;25(2):254–274.
  92. Mertens J, et al. Directly reprogrammed human neurons retain aging-associated transcriptomic signatures and reveal age-related nucleocytoplasmic defects. *Cell Stem Cell.* 2015;17(6):705–718.
  93. Cohen RM, et al. A transgenic Alzheimer rat with

- plaques, tau pathology, behavioral impairment, oligomeric A $\beta$ , and frank neuronal loss. *J Neurosci*. 2013;33(15):6245–6256.
94. Pang K, et al. An App knock-in rat model for Alzheimer's disease exhibiting A $\beta$  and tau pathologies, neuronal death and cognitive impairments. *Cell Res*. 2022;32(2):157–175.
  95. VandeVrede L, et al. Targeting tau: clinical trials and novel therapeutic approaches. *Neurosci Lett*. 2020;731:134919.
  96. Bolós M, et al. Soluble Tau has devastating effects on the structural plasticity of hippocampal granule neurons. *Transl Psychiatry*. 2017;7(12):1267.
  97. Lasagna-Reeves CA, et al. Tau oligomers impair memory and induce synaptic and mitochondrial dysfunction in wild-type mice. *Mol Neurodegener*. 2011;6:39.
  98. Shi Y, et al. Structure-based classification of tauopathies. *Nature*. 2021;598(7880):359–363.
  99. Fitzpatrick AWP, et al. Cryo-EM structures of tau filaments from Alzheimer's disease. *Nature*. 2017;547(7662):185–190.
  100. Woerman AL, et al. Tau prions from Alzheimer's disease and chronic traumatic encephalopathy patients propagate in cultured cells. *Proc Natl Acad Sci U S A*. 2016;113(50):E8187–E8196.
  101. Tsai RM, et al. Reactions to multiple ascending doses of the microtubule stabilizer TPI-287 in patients with Alzheimer disease, progressive supranuclear palsy, and corticobasal syndrome: a randomized clinical trial. *JAMA Neurol*. 2020;77(2):215–224.
  102. Guo T, et al. Roles of tau protein in health and disease. *Acta Neuropathol*. 2017;133(5):665–704.
  103. Leclair-Visonneau L, et al. Randomized placebo-controlled trial of sodium valproate in progressive supranuclear palsy. *Clin Neurol Neurosurg*. 2016;146:35–39.
  104. Lovestone S, et al. A phase II trial of tideglusib in Alzheimer's disease. *J Alzheimers Dis*. 2015;45(1):75–88.
  105. Tolosa E, et al. A phase 2 trial of the GSK-3 inhibitor tideglusib in progressive supranuclear palsy. *Mov Disord*. 2014;29(4):470–478.
  106. van Dyck CH, et al. Effect of AZD0530 on cerebral metabolic decline in Alzheimer disease: a randomized clinical trial. *JAMA Neurol*. 2019;76(10):1219–1229.
  107. Li X, et al. Structure-based discovery and development of novel O-GlcNAcase inhibitors for the treatment of Alzheimer's disease. *Eur J Med Chem*. 2022;238:114444.
  108. VandeVrede L, et al. Open-label phase 1 futility studies of salsalate and young plasma in progressive supranuclear palsy. *Mov Disord Clin Pract*. 2020;7(4):440–447.
  109. Ljubenkova PA. 15th Conference Clinical Trials Alzheimer's Disease, November 29–December 2, 2022, San Francisco, CA, USA: Posters (Clinical Trial Alzheimer's Disease). *J Prev Alzheimers Dis*. 2022;9:51–248. <https://link.springer.com/article/10.14283/jpad.2022.97>. Accessed June 7, 2023.
  110. Wilcock GK, et al. Potential of low dose leuco-methylthionium bis(hydromethanesulphonate) (LMTM) monotherapy for treatment of mild Alzheimer's disease: cohort analysis as modified primary outcome in a phase III clinical trial. *J Alzheimers Dis*. 2018;61(1):435–457.
  111. Morimoto BH, et al. A double-blind, placebo-controlled, ascending-dose, randomized study to evaluate the safety, tolerability and effects on cognition of AL-108 after 12 weeks of intranasal administration in subjects with mild cognitive impairment. *Dement Geriatr Cogn Disord*. 2013;35(5–6):325–336.
  112. Boxer AL, et al. Davunetide in patients with progressive supranuclear palsy: a randomised, double-blind, placebo-controlled phase 2/3 trial. *Lancet Neurol*. 2014;13(7):676–685.
  113. DeVos SL, et al. Tau reduction prevents neuronal loss and reverses pathological tau deposition and seeding in mice with tauopathy. *Sci Transl Med*. 2017;9(374):eaag0481.
  114. Mummery CJ, et al. Results of the first-in-human, randomized, double-blind, placebo-controlled phase 1b study of lumbar intrathecal bolus administrations of antisense oligonucleotide (ISIS 814907; BIIB080) targeting tau mRNA in patients with mild Alzheimer's disease. *Alzheimers Dement*. 2021;17(s9):e051871.
  115. Wang W, et al. A novel small-molecule PROTAC selectively promotes tau clearance to improve cognitive functions in Alzheimer-like models. *Theranostics*. 2021;11(11):5279–5295.
  116. Silva MC, et al. Targeted degradation of aberrant tau in frontotemporal dementia patient-derived neuronal cell models. *Elife*. 2019;8:e45457.
  117. Novak P, et al. ADAMANT: a placebo-controlled randomized phase 2 study of AADvac1, an active immunotherapy against pathological tau in Alzheimer's disease. *Nat Aging*. 2021;1(6):521–534.
  118. Hickman DT, et al. Sequence-independent control of peptide conformation in liposomal vaccines for targeting protein misfolding diseases. *J Biol Chem*. 2011;286(16):13966–13976.
  119. Vaz M, Silvestre S. Alzheimer's disease: recent treatment strategies. *Eur J Pharmacol*. 2020;887:173554.
  120. Ji C, Sigurdsson EM. Current status of clinical trials on tau immunotherapies. *Drugs*. 2021;81(10):1135–1152.
  121. Liu W, et al. Vectored intracerebral immunization with the anti-tau monoclonal antibody PHF1 markedly reduces tau pathology in mutant tau transgenic mice. *J Neurosci*. 2016;36(49):12425–12435.
  122. Mukadam AS, et al. Cytosolic antibody receptor TRIM21 is required for effective tau immunotherapy in mouse models. *Science*. 2023;379(6639):1336–1341.
  123. Allen JC, et al. Understanding immunosenescence and its impact on vaccination of older adults. *Vaccine*. 2020;38(52):8264–8272.
  124. Ishida K, et al. Glymphatic system clears extracellular tau and protects from tau aggregation and neurodegeneration. *J Exp Med*. 2022;219(3):e20211275.
  125. Fulop T, et al. Immunosenescence and cancer. *Crit Rev Oncog*. 2013;18(6):489–513.
  126. Barthel H. First Tau PET Tracer approved: toward accurate in vivo diagnosis of Alzheimer disease. *J Nucl Med*. 2020;61(10):1409–1410.
  127. Soleimani-Meigooni DN, et al. 18F-flortaucipir PET to autopsy comparisons in Alzheimer's disease and other neurodegenerative diseases. *Brain*. 2020;143(11):3477–3494.
  128. Yushkevich PA, et al. Three-dimensional mapping of neurofibrillary tangle burden in the human medial temporal lobe. *Brain*. 2021;144(9):2784–2797.
  129. Mueller A, et al. Tau PET imaging with <sup>18</sup>F-PI-2620 in patients with Alzheimer disease and healthy controls: a first-in-humans study. *J Nucl Med*. 2020;61(6):911–919.
  130. Ossenkoppele R, et al. Tau biomarkers in Alzheimer's disease: towards implementation in clinical practice and trials. *Lancet Neurol*. 2022;21(8):726–734.
  131. Leuzy A, et al. Biomarker-based prediction of longitudinal tau positron emission tomography in Alzheimer disease. *JAMA Neurol*. 2022;79(2):149–158.
  132. Thijssen EH, et al. Plasma phosphorylated tau 217 and phosphorylated tau 181 as biomarkers in Alzheimer's disease and frontotemporal lobar degeneration: a retrospective diagnostic performance study. *Lancet Neurol*. 2021;20(9):739–752.
  133. Jack CR Jr, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *Lancet Neurol*. 2010;9(1):119–128.
  134. Mattsson-Carlgen N, et al. Cerebrospinal fluid biomarkers in autopsy-confirmed Alzheimer disease and frontotemporal lobar degeneration. *Neurology*. 2022;98(11):e1137–e1150.
  135. Horie K, et al. CSF tau microtubule-binding region identifies pathological changes in primary tauopathies. *Nat Med*. 2022;28(12):2547–2554.
  136. Cummings J, et al. The role of basket trials in drug development for neurodegenerative disorders. *Alzheimers Res Ther*. 2022;14(1):73.
  137. Bateman RJ, et al. The DIAN-TU Next Generation Alzheimer's prevention trial: adaptive design and disease progression model. *Alzheimers Dement*. 2017;13(1):8–19.
  138. Marquis-Gravel G, et al. Streamlining the institutional review board process in pragmatic randomized clinical trials: challenges and lessons learned from the Aspirin Dosing: A Patient-centric Trial Assessing Benefits and Long-Term Effectiveness (ADAPTABLE) trial. *Trials*. 2021;22(1):90.
  139. Staffaroni AM, et al. Temporal order of clinical and biomarker changes in familial frontotemporal dementia. *Nat Med*. 2022;28(10):2194–2206.