Title
Interacting symbionts and immunity in the amphibian skin mucosome predict disease risk and probiotic effectiveness

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Introduction

Probiotic therapies often aim to extend or shape the immune function of hosts by altering the symbiotic microbial community. Probiotics are used in human and veterinary medicine, agriculture and aquaculture, and have been proposed for treatment of emerging wildlife diseases such as those occurring on corals and amphibians [1,2]. Microbiota can mediate pathogenesis through a wide range of mechanisms [3,4], and disease ecology studies demonstrate that parasitic and non-parasitic microbes interact with each other and with the host immune system such that pathogenicity is often influenced by environmental conditions [5–8]. Thus, the environment affects the risk of disease to individuals, populations, and species, and assessing disease risk under changing conditions is vital to conservation and infectious disease mitigation and can direct the allocation of resources for most effective interventions.
The microbiota inhabiting skin and mucosal surfaces has a profound impact on host health and immunity [7,13,14], and may be predictive of risk for some diseases [15–17]. Amphibian skin is a model system for diseases affecting vertebrate mucosa. The mucosae, or micro-ecosystem of the mucus, as defined here contains interdependent host factors (mucosal antibodies, antimicrobial peptides, lysozyme, alkaloids) and microbial-community factors (microbiota, antibiotic metabolites). The mucosae has various functions potentially including communication, and predator and pathogen defense. Here, we develop a non-lethal assay and holistic measure referred to as "mucosome function" to describe the effect of amphibian skin mucus on pathogen viability. We examine how environmental and immunological contexts may impact the outcome of host-microbe symbioses, and how mucosome function captures the in vivo complexity of the micro-ecosystem and can thus accurately predict susceptibility to infection. We focus on probiotic bacteria and fungi applied to the skin mucosae as biocontrol agents against the emerging amphibian disease chytridiomycosis.

Chytridiomycosis is a major cause of global amphibian population declines and species extinctions [18,19]. The disease is caused by the chytrid fungus Batrachochytrium dendrobatidis, or Bd, and is strongly influenced by climatic conditions [20]. Climate-linked changes to the entire microbiota, not just Bd, may influence disease susceptibility [5]. Current efforts to mitigate chytridiomycosis in wildlife populations have turned to bioaugmentation, or the use of probiotic therapies [1,21]. The successful prophylactic use of Janthinobacterium lividum was demonstrated against chytridiomycosis in mountain yellow-legged frogs, Rana muscosa [22]. However, when tested on the endangered Panamanian golden frog, Atelopus zeteki, the probiotic survived briefly on the skin, but did not protect the amphibians from disease [23]. Similarly, the probiotic Pedobacter cryoconitis temporarily reduced infection loads of heavily infected R. muscosa [24]. Each target host may thus require probiotic therapy tailored to that species, population, or life-history stage. Screening the various bacteria associated with hosts or their environment to identify effective probiotics is challenging [25,26]. Thus, probiotic therapies for amphibians must be optimized, and an understanding of which candidate bacteria can establish and persist on the host in its natural environmental context is urgently needed.

To date, all attempts to apply probiotic therapy against chytridiomycosis have used simple selection criteria for choosing candidate probiotics. Selection of the most efficient probiotic is challenging because there are hundreds of culturable phylotypes to choose from, either from environmental sources, or more typically, from tolerant host populations that can persist with nonlethal Bd infections [1]. However, simple co-culture assays to determine antifungal capacity have been insufficient to ensure probiotic effectiveness [23,24]. Co-factors including interactions of the probiotic with the microbial community already present on the amphibian skin, as well as interactions with host immune defenses, and effects of environmental conditions, may complicate the outcome of biotherapy. Here, we experimentally test the impact of immunological and environmental context on potential probiotic bacteria both in vitro and in vivo. The tested conditions are illustrative rather than comprehensive for potential environmental conditions, community and immunological interactions. Because it is impractical to test all potential interactions before testing probiotics on amphibians for a disease resistance effect, we suggest a protocol for selecting probiotics with the highest potential benefit, and to test whether the probiotics will likely be effective in the range of foreseeable conditions on the host. Our non-lethal susceptibility assay of mucosome function can help assess disease risk and treatment effects in rare amphibians including relict populations or captive populations of endangered species intended for reintroduction.

Typical approaches to compare species susceptibility and to assess disease risk include pathogen exposure experiments [27], or field surveys to compare infection prevalence and monitor disease and population trajectories [20], or modeling environmental and biogeographic risk factors [10,29]. Deficits of conventional pathogen exposure experiments include lack of environmental context when amphibians are exposed under clean laboratory conditions. Biodiversity including microbiota and macrobiota can influence disease outcome [30], and bacterial community diversity is reduced through time in captivity without natural sources such as soil for re-inoculating the skin [31]. The exposure history, population genetics, and life-history stage of the amphibians used in the experiment, as well as the strain and dose of the pathogen can all affect experimental outcomes, and many threatened species are not suitable for such experiments. In addition, growth of Bd is often inhibited by skin microbiota of amphibians [32,33]. However, little is known about how protective microbiota differs among host populations or regions, or how mucosome function is altered by enrichment with potential probiotics.

Our aims in this study were (1) to develop a holistic, simple, non-invasive, and non-lethal method to measure mucosome function against Bd. Using this tool, we aimed (2) to test whether mucosome function can predict Bd infection prevalence of amphibians in the field and survival in Bd exposure experiments. While we show that probiotics are influenced by a variety of factors including competition, temperature, and innate immunity when tested in vitro, we aimed (3) to use mucosome function as an ecologically-integrated predictor of probiotic therapy effect so that future research can test probiotic strategies for conservation and not lose hope in the potential of probiotic therapy in the face of immunological and ecological complexity. We provide a detailed protocol for measuring mucosome function in File S1.

Materials and Methods

Ethics statement

Permits to conduct fieldwork were obtained from the Swiss cantonal conservation authorities, and from Germany - German federal licence (Rheinland-Pfalz) no. 425-104.143.0904 Struktur- und Genehmigungsdirektion Nord, Koblenz. All animal procedures were approved by the Veterinary Authority of Zurich (110/2007 and 227/2007) and the Federal Office for the Environment. Fieldwork conformed to standard decontamination practices to avoid transport of pathogens between sites. All animals in experiments were monitored daily for animal welfare and to ameliorate suffering. During experiments, any individual demonstrating clinical signs of disease including lethargy, abnormal skin shedding, and loss of righting reflex were humanely euthanized. At the end of the experiment, all animals were humanely euthanized by overdose of tricaine methanesulfonate.

Survey of Bd infection prevalence

To compare Bd infection prevalence among species and life-history stages, we combine previously unpublished results from field studies in Switzerland with Bd surveys from amphibians across Europe collated by Bd-Maps (www.bd-maps.net, accessed September 1, 2013). In addition to data from 5939 sampled amphibians available from Bd-maps, skin swabs were collected from 2591 amphibians from 12 species and from 66 Bd-positive populations from the northern parts of Switzerland and tested for Bd between 2007 and 2009 (Table 1). Amphibians were caught by
dip-netting and swabbed with a sterile cotton swab (Copan Italia S.p.A., Brescia, Italy). Field material was cleaned and disinfected before moving between different sites to avoid contamination and spread of Bd and other pathogens. Extraction and analysis for Bd-DNA were done following the qPCR protocol by Boyle et al. [34] using Bd-specific primers and standards to quantify the amount of DNA. We ran each sample twice and the PCR was repeated if the two wells returned dissimilar results. Reactions below 1 genomic equivalent were scored Bd-negative to avoid false positives. Mean infection prevalence with 95% binomial confidence interval was calculated for each species and life stage sampled, and calculated for both Europe and Switzerland.

**Bd infection prevalence predicted by skin defenses**

Skin defense peptides and mucosome samples were tested against Bd for comparison of anti-Bd activity with infection prevalence in natural populations by logistic regression in R. Amphibians sampled for skin peptides and mucosome function (Table 1) were sampled in Switzerland and compared to field infection prevalence from Switzerland and across Europe in separate analyses. Skin peptides were collected upon induction by subcutaneous injection of metamorphosed amphibians with 40 nmol/g body mass norepinephrine (bitartrate salt, Sigma) or immersion of larval amphibians in 100 µM norepinephrine, and tested for Bd growth inhibition as previously described [35,36]. Skin peptide samples from post-metamorphic amphibians only were used in the logistic regression analyses because different methods of peptide induction were used on larval stages. Mucosome samples from multiple life-history stages of the same species were included and matched to life-history stages sampled for Bd diagnostics (Table 1). Detailed methods for measuring mucosome function against Bd using a fluorescence assay of Bd viability adapted from Stockwell et al. [37] (Fig S1 in File S1) and comparisons of mucosome function and skin peptide defenses against Bd are presented in Supporting Information (Figs S4, S5 in File S1).

**Survival predicted by mucosome function**

To examine the relationship between mucosome function against Bd and susceptibility to infection and subsequent survival we performed experimental exposures to Bd on four species. All animals were exposed to zoospores from Swiss lineage Bd TG 739 isolated from a moribund *A. obstetricans* in Gamlikon, Switzerland in 2007 [38] and cryopreserved until use. Egg clutches were obtained from *P. esculentus* in 2007 [38] and cryopreserved until use. Egg clutches were obtained from *P. esculentus* (n = 8), *B. variegata* (n = 8), *R. temporaria* (n = 45), and *A. obstetricans* (n = 13) in northern Switzerland or southern Germany. *Rana temporaria* were raised in outdoor mesocosms through metamorphosis before experimental exposure of metamorphs to Bd (N = 92 exposed, 94 control). Other species were exposed to Bd as tadpoles (N = 80 exposed, 40 control per species). All animals were kept in the same laboratory at 19°C during the experiments. We measured the proportion of infected metamorphs by qPCR, and determined relative survival (survival of infected/survival of uninfected controls) at the end of the experiments (50–90 d after metamorphosis). Kaplan Meier curves are presented in the Supporting Information for each species. We examined the relationship between mucosome function against Bd and relative survival and proportion infected using logistic regression analyses in R.

**Host ecological context and skin defenses**

The in vivo effects of ambient temperature and skin microbiota on mucosome function against Bd and skin peptide defenses were tested on a focal amphibian species, *A. obstetricans*. In Europe, the common midwife toad, *A. obstetricans*, is a species of conservation concern [39] and is particularly sensitive to Bd early in life-history [40]. Host-associated bacteria and fungi were surveyed by culturing from populations of midwife toads near Basel, Switzerland in May, 2009, including samples from 19 adults, 32 larvae, and 9 egg clutches. Although many diverse antifungal bacteria have been described in association with skin of some amphibian hosts [32,33], we chose eleven bacterial isolates isolated from *A. obstetricans* for the environmental context experiments described below based on potency against Bd in culture and high prevalence in the populations sampled (L. Davis, unpublished). Two bacterial isolates with high in vitro potency against Bd and the ability to withstand host skin defense peptides, and one fungal isolate, were chosen for applications on recently metamorphosed *A. obstetricans*.

All metamorphs used in the study were raised in captivity from wild-caught tadpoles that were naturally exposed to the fungus in their pond of origin, near Zunzgen, Switzerland, but negative for Bd by qPCR at the time of the experiment. Toadlets were of similar size (mean±SD: 2.1±0.3 g; ANOVA F_{1,179} = 1.179, P = 0.332) and treated at the same time with one exception. Toadlets in the Bd-exposure group were exposed to Bd approximately 2 months prior to the microbial exposure treatments, and the toadlets were smaller (1.5±0.3 g), and no longer infected at the time of sampling based on qPCR.

We treated recently metamorphosed common midwife toads, *Alytes obstetricans* (N = 70; 10 per treatment group, 7 treatments), by housing them individually at 5°C, 18°C, or 25°C with no microbes added, or at 18°C with exposure to Bd zoospores (8.5×10^{6} zoospores of global panzootic lineage isolated from a *Bufo bufo* in the UK [38]), a probiotic fungus *Penicillium expansum*, or a probiotic bacterium *F. fluorescens* or *F. johnsoniae*. Toadlets were bathered individually for one hour in water containing the microbes and after 2 weeks, toadlets from all treatments were sampled on the same day for mucosome function and subsequently skin peptides, sampled as described above.

**Temperature, competition of probiotic strains, and co-culture with Bd**

To determine the effects of competitive interactions and temperature on probiotic potential, 11 common host-associated isolates were chosen. These included two isolates of *Serratia plymuthica* and one isolate of *Janthinobacterium lividum* from egg clutches of midwife toads, three isolates of *Flavobacterium johnsoniae* and five species of *Pseudomonas* isolated from the skin of adults. Based on 16S rRNA gene sequences, all 11 isolates were considered unique operational taxonomic units (OTUs) at 99%, but clustered into 7 OTUs at 97% similarity as determined by the UCLUST algorithm in QIIME. The 16S rRNA gene sequences of all isolates were deposited in the European Nucleotide Archive (Table S1 in File S1).

In one set of experiments, bacterial isolates were freshly grown at 18°C on RIIA agar media supplemented with 1% tryptone then transferred to experimental conditions. Bacteria and Bd (Swiss isolate TG 739) readily grew on the same media. Plate experiments were performed in duplicate. Both isolates of *Serratia plymuthica* were grown separately at 18 and 25°C, or at 18°C on media inoculated with Bd and allowed to dry before streaking the bacteria. Two isolates of *F. johnsoniae* were grown separately or combined on media inoculated with Bd, and grown at 18°C. When combined, each isolate was streaked across the entire plate. Three *Pseudomonas* isolates were grown either separately, combined, or combined on media inoculated with Bd, and grown at 18°C. Control plates of sterile media or Bd-only were also tested. All plates were incubated for 3 days, and then rinsed with 2 ml sterile...
Table 1. Amphibians from Switzerland sampled for skin peptide effectiveness and mucosome function against *Bd*, and *Bd* infection prevalence at different life-history stages.

<table>
<thead>
<tr>
<th>Species</th>
<th>Life-history stage#</th>
<th>Peptide effectiveness* (N)</th>
<th>SE</th>
<th>Mean mucosome function against Swiss <em>Bd</em> (N)</th>
<th>SE</th>
<th>Switzerland: Percent infected (N)</th>
<th>95% binomial confidence interval</th>
<th>Europe: Percent infected (N)</th>
<th>95% binomial confidence interval</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Alytes obstetricans</em></td>
<td>Adult/Subadult</td>
<td>15.92 (8)</td>
<td>6.21</td>
<td>0.012 (10)</td>
<td>0.00</td>
<td>4.9 (41)</td>
<td>0.6–16.5</td>
<td>29.7 (209)$</td>
<td>23.5–36.4</td>
</tr>
<tr>
<td><em>Alytes obstetricans</em></td>
<td>Metamorph</td>
<td>37.75 (5)</td>
<td>12.15</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Alytes obstetricans</em></td>
<td>Larvae</td>
<td>48.76 (5)</td>
<td>24.23</td>
<td>2.963 (10)</td>
<td>0.681</td>
<td>45.4 (2111)</td>
<td>43.3–47.6</td>
<td>38.0 (3008)</td>
<td>36.3–39.8</td>
</tr>
<tr>
<td><em>Bombina variegata</em></td>
<td>Adult/Subadult</td>
<td>1.075 (4)</td>
<td>0.081</td>
<td>20.0 (150)</td>
<td>13.9–27.3</td>
<td>21.1 (227)</td>
<td>16.0–27.0</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Bufo bufo</em></td>
<td>Adult</td>
<td>16.34 (15)</td>
<td>3.7753</td>
<td>0.117 (9)</td>
<td>0.082</td>
<td>0.0 (22)</td>
<td>0.0–15.4</td>
<td>0.9 (3606)</td>
<td>0.6–1.2</td>
</tr>
<tr>
<td><em>Bufo bufo</em></td>
<td>Larvae</td>
<td>1.284 (5)</td>
<td>0.404</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>6.7 (45)</td>
<td>1.4–18.3</td>
</tr>
<tr>
<td><em>Hyla arborea</em></td>
<td>Adult</td>
<td>11.42 (7)</td>
<td>2.15210</td>
<td></td>
<td></td>
<td>3.8 (26)</td>
<td>0.1–19.6</td>
<td>12.5 (32)</td>
<td>3.5–29.0</td>
</tr>
<tr>
<td><em>Ichthyosaura alpestris</em></td>
<td>Adult</td>
<td>0.94 (7)</td>
<td>0.52546</td>
<td>1.361 (20)</td>
<td>0.062</td>
<td>24.8 (629)</td>
<td>21.5–28.4</td>
<td>21.5 (775)</td>
<td>18.7–24.6</td>
</tr>
<tr>
<td><em>Lissotriton vulgaris</em></td>
<td>Adult</td>
<td>1.85 (4)</td>
<td>1.02506</td>
<td></td>
<td></td>
<td>27.3 (22)</td>
<td>10.7–50.2</td>
<td>17.0 (47)</td>
<td>7.7–30.8</td>
</tr>
<tr>
<td><em>Pelophylax lessonae/esculentus</em></td>
<td>Adult</td>
<td>27.27 (10)</td>
<td>3.18</td>
<td></td>
<td></td>
<td>22.4 (170)</td>
<td>16.3–29.4</td>
<td>15.6 (275)</td>
<td>11.6–20.5</td>
</tr>
<tr>
<td><em>Pelophylax lessonae/esculentus</em></td>
<td>Metamorph</td>
<td>5.34 (5)</td>
<td>1.88685</td>
<td>0.545 (10)</td>
<td>0.042</td>
<td>13.0 (69)</td>
<td>6.1–23.3</td>
<td>13.2 (76)</td>
<td>6.5–22.9</td>
</tr>
<tr>
<td><em>Rana temporaria</em></td>
<td>Adult/Subadult</td>
<td>1.97 (13)</td>
<td>0.62111</td>
<td>0.251 (10)</td>
<td>0.128</td>
<td>0.0 (10)</td>
<td>0.0–30.9</td>
<td>3.1 (129)</td>
<td>0.9–7.8</td>
</tr>
<tr>
<td><em>Rana temporaria</em></td>
<td>Larvae</td>
<td>0.220 (5)</td>
<td>0.120</td>
<td></td>
<td></td>
<td>0.0 (20)</td>
<td>0.0–16.8</td>
<td>0.0 (23)</td>
<td>0.0–14.8</td>
</tr>
<tr>
<td><em>Salamandra salamandra</em></td>
<td>Adult</td>
<td>4.92 (9)</td>
<td>1.32654</td>
<td></td>
<td></td>
<td>11.1 (9)</td>
<td>0.3–48.3</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Salamandra salamandra</em></td>
<td>Larvae</td>
<td>42.78 (5)</td>
<td>13.35528</td>
<td></td>
<td></td>
<td>23.2 (69)</td>
<td>13.9–34.9</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Skin peptide effectiveness is the percent inhibition of *Bd* zoospore growth caused by 50 μg/ml peptide multiplied by the quantity of peptides (mg) per g amphibian according to Woodhams et al. [11]. The mucosome function against *Bd* (Swiss isolate TG 739) is a measure of zoospore viability quantified by the ratio of green:red fluorescence as described above. Infection prevalence is the mean from all amphibians in each group from multiple sites and seasons.

*Peptide effectiveness = % inhibition of *Bd* growth at 50 μg/ml * mg peptides/g frog mass.

*Includes samples from chytridiomycosis outbreak sites in Spain (S. Walker, unpubl.), not included in logistic regression.

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Mili-Q water. Rinse water was then filtered through a 0.22 μm syringe filter.

Bacteria were also grown in liquid RIIA media for 4 d at 14, 19, and 22°C, and metabolites filtered as above. Metabolites from liquid cultures were added to Bd zoospores (Global panzootic lineage VMV 013 from a bullfrog, Lithobates catesbeianus tadpole) to test for inhibitory effects on pathogen growth. To determine the effect of bacterial filtrate on Bd growth, Bd zoospores were harvested in 1% tryptone and counted under a hemocytometer. Wells of a 96-well plate were inoculated with 50 μl zoospores at 8×10^5 zoospores per ml. Then, 50 μl of filtrate (or filtrate diluted 1:10) from each of the experimental or control plates, or liquid cultures, was added to the wells in replicates of four. In addition, 6 positive control wells contained Bd and 50 μl sterile water or RIIA media, and 6 negative control wells contained heat-killed Bd and 50 μl sterile water or RIIA media. The change in optical density measured at 490 nm absorbance over 7 days growth at 19°C was recorded using a Victor3 multilabel plate reader (PerkinElmer).

Standard statistical testing was carried out in IBM SPSS Statistics 22. Significant Bd growth inhibition (or enhancement) caused by bacterial filtrate was determined by t-test, and a repeatable result (Table S2 in File S1). Percent inhibition depended on filtrate dose (see Results) and was not considered comparable among bacterial isolates.

Effects of host skin peptides and Bd metabolites on probiotics in culture

To test for the response of bacterial growth upon culture with either Bd filtrate or host skin peptides, bacteria were grown in RIIA liquid media on 96 well plates. Supernatant from a 2-week old culture of Bd (type isolate JEL 197) growing in 0.5% tryptone was filtered through a 0.22 μm syringe filter. An equal volume of Bd filtrate or sterile media was added to bacterial cultures. To test effects of peptides, we added an equal volume of sterile water or natural mixtures of partially-purified skin peptides from A. obstetricans metanmorphs at a final concentration of 100 μg/ml in water. Growth after 48 hr was measured as change in optical density measured at 480 nm. Differences between experimental and control bacterial growth were tested by t-tests using a density measured at 480 nm. Differences between experimental and control bacterial growth were tested by t-tests using a density measured at 480 nm absorbance over 7 days growth at 19°C was recorded using a Victor3 multilabel plate reader (PerkinElmer).

Survival predicted by mucosome function

Pathogen exposure experiments were conducted on four host species with a Swiss isolate of Bd, and relative survival post-metamorphosis of infected tadpoles differed among species (% relative survival, mean±SD days survived: A. obstetricans (0%, 24±17.5 d), Bombina variegata (39.0%, 32±23.9 d), and Pelophylax esculentus (30.4%, 12±12.8 d; Fig. S2 in File S1). Relative survival of recently metamorphosed Rana temporaria exposed to Bd was 100% (Fig. S3 in File S1), and no colonization by Bd was detected by qPCR (n = 92). Success of Bd colonization of tadpoles also differed among species (Pearson χ² = 13.102, P = 0.004): A. obstetricans (13.9% infected, n = 36), B. variegata (10.7%, n = 75), and P. esculentus (7.9%, n = 76). Mucosome function predicted survival (logistic regression, P<0.0001; Fig. 2a) and infection with Bd in these species (P = 0.0106; Fig. 2b). The odds of infection increased by 1.751 with each unit change in mucosome function, and the odds of survival decreased by 0.0454.

Host ecological context and skin defenses

Midwife toads, A. obstetricans, were treated with various temperature and probiotic therapies and tested for mucosome function. Host context significantly affected mucosome permisiveness or lethality towards Bd (Fig. 3a; ANOVA, F5,60 = 41.606, P<0.001). Bd viability was similar following incubation with mucosome samples from toads at temperatures ranging from 5–25°C. Mucosome samples from toads previously exposed to Bd were less effective at killing Bd zoospores, while those from toads treated with probiotics Flavobacterium johnsoniae and Penicillium expansum were most effective at killing zoospores (Fig. 3a). While Pseudomonas in general, and the P. fluorescens isolate (76.5%) used in this study were often effective at inhibiting Bd in co-culture and produced antifungal metabolites across a range of temperatures, ideal for Bd growth (Fig. 4a, Table S2 in File S1), there was no significant benefit of this probiotic when applied on hosts in terms of increasing mucosome function and reducing Bd viability (Fig. 3a).

Because one significant antimicrobial component of A. obstetricans skin mucus is antimicrobial peptides (AMPs) [44], we collected peptide skin secretions, quantified them per surface area of the toads and measured their ability to inhibit Bd growth at a standardized concentration of 100 μg/ml. On average, toads produced 0.25 mg peptide per cm² surface area, and at 100 μg/ml these peptides inhibited Bd growth by 48.7%. These values did not differ significantly among treatment groups, nor did a combined measure of skin peptide effectiveness against Bd (% mg/cm², Fig. 3b; Kruskal-Wallis tests, P>0.05). Thus, skin peptides stored in granular glands were not significantly affected by the 2-week temperature and microbe treatments including previous exposure.
to *Bd*. There was not a significant correlation between peptide effectiveness and mucosome function against *Bd* (Fig. S5 in File S1; Pearson, $\chi^2 = -0.102, P = 0.827$). Zoospore viability after exposure to mucosome samples was significantly higher in the *Bd*-exposure treatment compared to other treatments (Fig. 3a). However, skin peptides induced from hosts in the *Bd*-exposure treatment were effective at inhibiting *Bd* growth, and not significantly different than peptides from toads in other treatments (Fig. 3b).

Environmental conditions affected the capacity of probiotic bacteria to inhibit the fungal pathogen *Bd* (Table S2 in File S1). Two *Serratia plymuthica* isolates (isolates 27 and 28) were capable of inhibiting *Bd* growth when incubated at 18°C. Isolate 27 was inhibitory under all tested conditions: 18°C, 25°C, and 18°C co-cultured with *Bd*. Isolate 28 significantly enhanced *Bd* growth at 25°C, and was neither enhancing nor inhibitory at 18°C when co-cultured with *Bd* (Fig. 4c, Table S2 in File S1). A dose-response of *Bd* growth inhibition was found such that filtrate diluted 1/10 was

**Figure 1. Infection prevalence (mean, 95% binomial CI) of amphibians sampled across Europe and within Switzerland predicted by mucosome function and skin defense peptide activity against *Batrachochytrium dendrobatidis (Bd)* zoospores.** Mucosome function (mean, SE) indicates *Bd* viability after a 1 hr exposure to amphibian mucus (a,b) and units represent green:red fluorescence. Peptide efficiency (mean, SE) indicates quantity of natural mixtures of skin peptides induced from granular glands multiplied by activity of a standard concentration of peptides against *Bd* zoospore growth. Only post-metamorphic amphibians sampled upon subcutaneous injection with norepinephrine are plotted in (c) and (d). Amphibian skin mucosome function is a better predictor of infection prevalence than induced skin peptide efficiency (logistic regression, see text). Summary data for all species and life-history stages are presented in Table 1.

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**Temperature, competition of probiotic strains, and co-culture with *Bd***

Environmental conditions affected the capacity of probiotic bacteria to inhibit the fungal pathogen *Bd* (Table S2 in File S1). Two *Serratia plymuthica* isolates (isolates 27 and 28) were capable of inhibiting *Bd* growth when incubated at 18°C. Isolate 27 was inhibitory under all tested conditions: 18°C, 25°C, and 18°C co-cultured with *Bd*. Isolate 28 significantly enhanced *Bd* growth at 25°C, and was neither enhancing nor inhibitory at 18°C when co-cultured with *Bd* (Fig. 4c, Table S2 in File S1). A dose-response of *Bd* growth inhibition was found such that filtrate diluted 1/10 was
Mucosome Function Predicts Disease Risk

Effects of host skin peptides and Bd metabolites on probiotics in culture

Amphibian skin defense peptides may regulate the skin microbiota. We found that natural mixtures of skin peptides from A. obstetricans at a concentration of 100 μg/ml significantly inhibited growth of Pseudomonas migulae (73c1) and significantly enhanced growth of P. filiscindens (73c1), Flavobacterium johnsoniae (70d1), and Janthinobacterium lividum (73c1), while significantly enhancing the growth of J. lividum (73b1), and P. filiscindens (5/27b2, 5/28a3), P. plymuthica (76c1), and Flavobacterium johnsoniae (70d1), and Janthinobacterium lividum (73c1), while significantly enhancing the growth of J. lividum (77.5b; t-test, Bonferroni corrected P's<0.05; Fig. S6 in File S1).

We tested for a direct effect of Bd metabolites on bacterial growth, and found that filtrate from two-week old cultures of Bd in 0.5% tryptone significantly inhibited the growth of Serratia plymuthica (5/27b2, 5/28a3), F. johnsoniae (81a1, 70d1), and P. filiscindens (73c1), while significantly enhancing the growth of J. lividum (77.5b1; t-test, Bonferroni corrected P's<0.05; Fig. S6 in File S1).

Discussion

We found that a holistic measure of mucosome function against Bd is predictive of infection risk in natural populations of amphibians and survival in laboratory exposure experiments. While induced antimicrobial peptides may explain some variation in infection risk (Fig. 1b,d), mucosome function can be altered through probiotic therapy (Fig. 3a), and thus microbial communities play a major role in determining susceptibility to infection with Bd. In particular, tadpoles of the endangered midwife toad, A. obstetricans may be most at risk of both infection and subsequent disease-induced mortality upon metamorphosis (Fig. 2), even though adult toads are well protected by the mucosome and perhaps resistant to colonization with Bd. Similarly, the common frog R. temporana has strong mucosome activity against Bd, shows Bd colonization resistance, but has relatively poor skin defense peptides. This suggests that this common species has protective microbial communities. Adaptive defenses are not suspected because frogs were raised from eggs and had no history of exposure to Bd.

In this study, we provide several striking examples showing that probiotic capacity depends on immunological and environmental context. These examples lead to recommendations for choosing probiotics based on predictable host conditions. Temperature is known to influence amphibian host immune function [41] and

Figure 2. Relative survival (95% binomial CI; a) and Proportion of infected frogs (95% binomial CI; b) predicted by Mucosome function. Post-metamorphosis survival was measured from four Swiss amphibian species after exposure to zoospores of a Swiss Bd isolate, TG 739. Survival curves for each species are presented in Supporting Information (Figs. S2, S3 in File S1) and relative survival was calculated as the proportion of infected frogs surviving/proportion of unexposed control frogs surviving. Alytes obstetricans showed the highest infection and mortality, and Rana temporaria the lowest, with Bombina variegata and Pelophylax esculentus intermediate. All frogs were raised in captivity from egg clutches and had no history of natural exposure to Bd. Mucosome function (mean, SE) indicates Bd viability after exposure to amphibian mucus and is a significant predictor of both survival (binomial logistic regressions, P<0.0001) and infection prevalence (P=0.0106).

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significantly less inhibitory than undiluted filtrate (paired t-test, t95 = 9.836, P<0.001), and filtrate from control plates with or without Bd significantly enhanced Bd growth (Table S2 in File S1). Testing metabolites of the bacteria growing at 14, 19, and 22°C in liquid culture against the global panzootic lineage of Bd showed similar results including a dose-response (Fig. 4a,b), paired t-test, t95 = −10.607, P<0.001). In several cases, Bd growth was enhanced with addition of diluted bacterial metabolites in comparison to positive control growth with RIIA media only (>100%, Fig. 4b). Most cultures were more inhibitory of Bd at the lower temperatures, except for J. lividum, (isolate 77.5b) which was most inhibitory at 22°C (Fig. 4a,b).

While all bacteria were unique based on 16S rRNA gene sequencing when clustered at 99% similarity, probiotic physiology and function against Bd did not always correspond to OTU clustering at 97% similarity (Table S1 in File S1). In other words, bacterial isolates considered to be the same “species” based on 16S rRNA could have different antifungal function. Here, only one of two Flavobacterium johnsoniae isolates inhibited Bd growth. When grown together, the filtrate remained inhibitory. However, when grown together and co-cultured with Bd, the filtrate was no longer inhibitory. Three Pseudomonas isolates were capable of inhibiting Bd growth, and were inhibitory when combined with or without co-culture with Bd. The above mentioned growth inhibition of Bd caused by bacterial filtrate was significantly different from control bacterial growth with water only added (independent t-tests, P<0.05 and replicated result; all data shown in Table S2 in File S1).

These conditions represent infected or uninfected hosts and are illustrative rather than comprehensive of all possible environmental conditions and competitive interactions.
bacterial growth, metabolism, pigment and antibiotic production [45]. However, it was surprising that a shift from 18 to 25°C, a typical natural range for midwife toads, caused a common bacterial symbiont of the eggs and skin, *Serratia plymuthica*, to change from inhibiting *Bd* to enhancing *Bd* growth (Fig. 4c).

Testing metabolites of the bacteria growing at 14, 19, and 22°C in liquid culture against the global panzootic lineage of *Bd* showed similar results (Fig. 4b). Functional changes in probiotic activity with shifts in temperature have not previously been reported. Our results provide an alternative mechanistic explanation for patterns of susceptibility related to climate, which have previously been limited to empirical observation and pathogen-centered effects [46–49].

The microbial interactions we tested also altered antifungal effects relative to what would be predicted from individual isolates. For example, co-culture of *Flavobacterium johnsoniae* with *Bd* caused cultures of the bacterium that normally produce antifungal metabolites to switch off antifungal activity: when grown together with *Bd*, *F. johnsoniae* filtrate was benign, and indeed *Bd* filtrate inhibited the growth of two out of three *F. johnsoniae* isolates (Fig. 4c).
Mucosome Function Predicts Disease Risk

![Graph showing growth of Bd upon exposure to bacterial metabolites at 14°C, 19°C, and 22°C.](image)

**Isolate 27:**
- Supernatant effect on Bd growth:
  - Inhibitory

**Isolate 28:**
- Supernatant effect on Bd growth:
  - Inhibitory
  - Enhancing
Figure 4. Environmental context determines antifungal capacity of probiotics. Tested temperatures (14, 19, 22 °C) significantly affected the production of bacterial metabolites in liquid media that could inhibit B. dendrobatidis (Bd; GPL isolate VMW 813) zoospore growth in a dose-dependent fashion (a = full strength metabolites, b = 1:10 dilution). * indicates that Bd growth differed among metabolite temperature treatments (ANOVA, Bonferroni-corrected P's < 0.05). (c) Representative replicates are shown of two isolates of Serratia plymuthica isolated from egg clutches of common midwife toads, Alytes obstetricans, grown on solid media under different temperature conditions. Filtrate from isolate 27 always inhibited growth of Bd, but filtrate from isolate 28 inhibited Bd growth at 18°C, and enhanced Bd growth at 25°C. Filtrate from sterile media (R2A agar supplemented with 1% tryptone) caused enhanced growth of Bd. Note that colony color can be an indication of antifungal metabolites such as prodigines from red Serratia spp. [45,67], but are produced only under certain growth conditions.

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S6 in File S1). Co-evolution of Bd and amphibian hosts is a postulated driver of pathogenicity factors including compounds suppressing host immune defenses [43,50,51]. These factors may extend to inhibiting certain antifungal symbionts or altering their function.

Myriad microbial and immune interactions occur once probiotics are added to living hosts. Thus, testing probiotics in vivo is critical for testing the intended antifungal effect of probiotic therapy under realistic environmental conditions. We found that previous exposure to Bd may have a negative effect on host immunity or the ability of the mucosome to kill zoospores (Fig 3A). This result is consistent with a study on Australia green-eyed tree frogs, Litoria serratia, showing inhibition of ambient skin peptides with Bd infection but no inhibition of inducible stored skin peptides [43]. Because stored skin defense peptides can have potent activity against Bd, yet not be active on the skin, induced skin peptides may not accurately predict infection susceptibility. This mystery of how seemingly well-defended species can be affected by chytridiomycosis [52] deserves careful study on the conditions under which host skin defense peptides are activated. Induced skin defense peptides were previously used to predict disease susceptibility in Panama [11] and New Zealand [53]. In Panama, most species had weak peptide defenses and declined after disease emergence while only two species had strong peptide defenses against Bd compared to reference species of known disease resistance. Of these two species, the one with the highest levels of skin peptide defenses persisted at the field site (Espadarana prosoblepon) [54], and the other species (Agalychnis lemur) disappeared, but a relict population has been detected nearby [Julie Ray, pers. comm.]. In New Zealand, all native species demonstrated high levels of skin peptide defenses and appear to resist chytridiomycosis [53], although populations are in decline [55].

We found that a bacterium F. johnsoniae and a fungal probiotic P. expansum can increase the Bd killing function of the mucosome. The bacterium P. fluorescens did not show this effect. Because host AMPs did not appear to be affected by these treatments (Fig. 3B), the observed effects are most likely caused by antifungal metabolites produced by the microbes growing on the amphibian skin [56]. Upregulation of host mucosal immunity excluding AMPs is an untested alternative mechanism, and potentially a beneficial host response to probiotics. A non-responsive immune system when given probiotics may be preferred from a conservation management standpoint in order for the probiotics to colonize the host, establish within the microbiota and persist. However, this is not necessarily common and immune stimulation in response to probiotics occurs in other systems [57,58].

An ideal probiotic would produce metabolites that inhibit Bd growth as shown above, and also be uninhibited by host skin defense peptides. A literature review demonstrates that skin peptides can inhibit the growth of some bacteria, but not others, and suggests that skin defense peptides may be critical in structuring the symbiont community on amphibian skin [52]. Rollins-Smith et al. [35] showed that Aeromonas hydrophila, a common resident on amphibian skin and also an opportunistic pathogen, could tolerate high levels of host antimicrobial peptides.

This organism shows antifungal characteristics including activity against Bd growth [33]. The ability of extracellular products of A. hydrophila to inhibit amphibian antimicrobial peptides indicates a co-evolutionary relationship between host and symbionts [59]. In addition, Pseudomonas mirabilis and Serratia liquefaciens were found to be resistant to antimicrobial peptides from several host frog species [60]. Here we used probiotics that largely resisted low concentrations of natural mixtures of host defense peptides (Fig. S6 in File S1). Thus, to increase the likelihood of probiotic establishment, use of probiotics with a co-evolutionary relationship with the target host may be advantageous.

While easily cultured, the isolates tested here may not be dominant community members based on culture-independent analyses [31,61,62]. Therefore, future studies will benefit by examining the effects of probiotic treatments on the natural microbial communities on host amphibians using culture-independent techniques such as next-generation sequencing. While community interactions are difficult to test in vitro and before probiotics are applied to a host, our results affirm that testing probiotics under certain foreseeable contexts may increase the pace of biotherapy development.

Because potential probiotics that inhibit the growth of Bd only do so under certain conditions, we recommend the following screening criteria (Fig. 5): (1) Candidates for probiotic development should be chosen from among the culturable microbiota locally present on tolerant hosts or populations that are able to persist with Bd [32,33]. (2) Candidates should have the capacity to inhibit Bd growth when grown in isolation, in co-culture with Bd, and in an environmental context relevant to the amphibian lifecycle, and (3) the ability to resist immune defenses on host skin, establish within the microbiota, and contribute to antifungal defenses in vivo. Resistance to mucosal immune defenses may be critical for establishment within the microbial community associated with the skin, and critical for long-term persistence. Some symbionts appear to be assisted in surviving on the host by thriving on skin mucosal products. Mucosal oligosaccharides, for example, differ among hosts and life-history stages, and may be a selective force in structuring the microbiota [63,64]. Amphibian skin provides a useful model of host-microbiota interactions to better understand mechanisms of microbial community assembly and maintenance within vertebrate mucosa. Indeed, these mechanisms underlie strategies to promote human health by manipulating microbial communities - a long-term goal of the Human Microbiome Project [7,65].

While screening for candidate probiotics, some beneficial organisms may be inadvertently discarded based on tests of bacterial filtrate on Bd growth. Microbes producing antifungal metabolites such as bacteriocins [66] or small molecule antibiotics [56,67] will be detected by this method. However, microbes may also compete directly for space or resources, and may exclude pathogenic fungi by other mechanisms [26,68]. Furthermore, microbial secondary metabolites such as prodigines produced by Serratia spp. can be immunosuppressive [67]. Probiotics may strongly influence host immunity through interactions with host Toll-like receptors or NOD-like receptors, or through interactions...
with epithelial cells and immune system cells modulating both local and systemic immune responses [69]. The immunomodulatory effect of probiotics cannot be tested with in vitro Bd growth assays and host trials are necessary to test for these emergent properties of probiotics.

Antimicrobial peptides and a range of other defenses protect amphibian skin by synergizing or interacting with microbes [41,70]. Thus, a better indication of antifungal effect of probiotics against amphibian chytridiomycosis. Candidate probiotic bacteria (or fungi) are isolated from populations of amphibians that are able to persist in the presence of B. dendrobatidis (Bd) [1]. To increase the chances of successful prophylactic biotherapy, candidate probiotics should be tested for at least three characteristics: (a) capacity to inhibit Bd growth as a pure isolate without specific competitive interactions to induce antifungal metabolites, (b) capacity to inhibit Bd at a temperature range consistent with host habitat, and (c) resistance to host skin immune defenses that would complicate probiotic establishment. Remedial biotherapy of already infected individuals should maintain antifungal capacity when grown in competition with Bd and withstand the sometimes lethal effects of Bd metabolites (Fig. S6 in File S1). Testing probiotic effect in vivo can be accomplished without resorting to pathogen exposure experiments by using the mucosome function assay described here. doi:10.1371/journal.pone.0096375.g005

Supporting Information

File S1 Protocol for determining Bd viability, supplementary tables and figures. (PDF)

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Author Contributions

Conceived and designed the experiments: DCW SB JK EK UT. Performed the experiments: DCW HB SB JK, EK UT LRD CB SH. Analyzed the data: DCW SB JK EK UT BRS. Wrote the paper: DCW BRS RK VM. Performed field work: DCW JK UT LRD.

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