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1998

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Functional Characterization of the HIV Genome  
by Genetic Footprinting

by

Louise Chang Laurent

DISSERTATION

Submitted in partial satisfaction of the requirements for the degree of

DOCTOR OF PHILOSOPHY

in

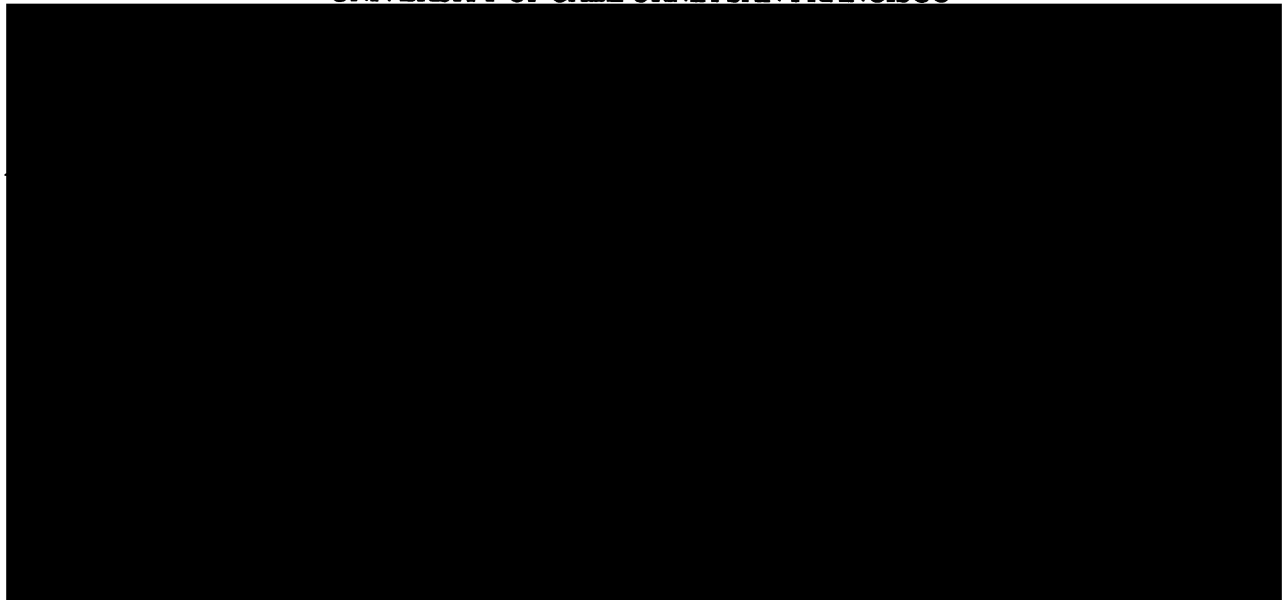
Biochemistry

in the

GRADUATE DIVISION

of the

UNIVERSITY OF CALIFORNIA SAN FRANCISCO



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**To Marc**

## Acknowledgements

Many thanks are due to the many members of the Brown lab, past and present. Foremost among these are fellow genetic footprinters Rachel Adams Crowley and Mari Olsen. Working with Rachel has made the struggle through graduate school a much more rewarding (and yes, even enjoyable) experience. I have benefited from her many abilities and personal strengths as we wrestled with the myriad difficulties, technical and ideological, involved in genetic footprinting. I will never be able to express how much I appreciate all of the help Mari has provided over the last few, very busy, years. I must thank Brian Scottoline and Liz Wilson for sharing their benches with me when I was orphaned by the Varmus lab. Thanks to Pat Brown, who has provided a stimulating environment in which to learn and taught me to true meaning of independence. Thanks to the other "virus people," including Richard.

Many thanks are due to the many mentors I have found in the scientific world. Thanks to Mrs. McPherson, my ninth-grade chemistry teacher. Thanks to Mr. Hozinsky, my tenth-grade biology teacher and the one who suggested that I roam the halls at the University of Chicago asking for someone to teach me about lab stuff. Thanks to Vikas Sukhatme, who took me into his lab and accepted the challenge of trying to teach a high school student molecular biology. Thanks to Jeremy Knowles and Betsy Komives, who were my mentors during my undergraduate years. Thanks to Mike Bishop and Harold Varmus for

convincing me to come to UCSF for my graduate training and playing large parts in that experience.

Many thanks are due to my friends. Here I tread lightly, as I am sure to leave someone out. I will restrict myself to thanking the members of my MSTP class and their spouses for making the past several years more than just school.

Many thanks are due to Jana and Sue, who have made life incredibly easy.

Many thanks are due to the many members of my family. My husband, Marc, has provided much support, moral and computer. My daughter, Clara, has made the last eight months a joy and a challenge. This challenge has been greatly decreased by help received from our friend and guardian angel, Agnes Leturgie, and advice received from my elder sister and mother of four, Cindy. My family-in-law, and especially my mother-in-law, has been steadfastly supportive. And finally, and most importantly, infinite thanks to my parents.

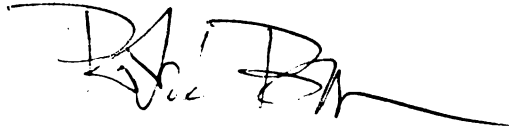
# **Abstract**

## **Functional Characterization of the HIV Genome**

### **by Genetic Footprinting**

In this report, I present a detailed analysis of the functional characteristics of the 1000 nucleotides at the 5' end of the HIV RNA genome. The effects of one hundred and thirty-four independent insertions mutations were examined in a quantitative manner at three points in the viral replication cycle. I studied the abilities of mutants 1) to make stable viral RNA, 2) to assemble and release viral-RNA-containing viral particles, 3) to enter host cells, complete reverse transcription, enter the nuclei of host cells, and generate proviruses in the host genome by integration. In order to carry out a thorough investigation on a large number of mutations, a modification of the genetic footprinting technique was employed. Using this method, all of the mutants were constructed and analyzed en masse, greatly decreasing the labor typically involved in mutagenesis studies. The presence of several functional features previously assigned to the region of the HIV genome under investigation was confirmed, and evidence for a number of novel features was found. Among these new features were cis-acting sequences that appeared to contribute to formation of stable viral transcripts, viral RNA packaging, or an early step in viral replication. These sequences were distinct from previously identified sequences that have been shown to be important for these steps in the viral life cycle. An unanticipated trans-acting role for sequences near the N-terminus of matrix in the formation of stable viral RNA transcripts was also seen. Finally, in contrast to previous reports, the results of

this study suggested that mutations detrimental to viral replication in sequences encoding the matrix and capsid proteins principally interfered with assembly.

A handwritten signature in black ink, appearing to be 'D. H. B.', with a long horizontal stroke extending to the right.



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# Chapter 1

## General Overview of Retrovirology

The first reports describing the disease that is now called AIDS (acquired immunodeficiency syndrome) were made in 1981 (Gottlieb et al. 1981; Masur et al. 1981; Siegal et al. 1981). The afflicted patients displayed an unusual series of symptoms and findings. They were all young, previously healthy men, a disproportionate number of whom were homosexual and/or substance abusers. They suffered from fulminant herpes simplex, candida, and cytomegalovirus infections and pneumocystis carinii pneumonia. At the time, these types of infections were very rare and found only in severely immunocompromised persons, such as premature infants and transplant patients undergoing immunosuppressive therapy. In the subsequent months and years, many more patients with the same constellation of findings were described. The pathogenic agent responsible for this epidemic was isolated in 1984 (Popovic et al. 1984) and in very short order the genome of this pathogen, the human immunodeficiency virus (HIV), was cloned and sequenced (Shaw et al. 1984). In the intervening years, HIV has been the focus of much attention and progress in understanding HIV has been rapid on many fronts.

HIV is a retrovirus, a class of viruses previously called RNA tumor viruses. This older and somewhat inaccurate nomenclature refers to two characteristics of retroviruses: the genomes of these diploid viruses consist of two molecules of linear single-stranded RNA; and many (though not all) retroviruses are associated with neoplasms. The current appellation "retrovirus" is the result of

the discovery of reverse transcriptase in 1970 (Baltimore 1970, Temin and Mizutani 1970). All retroviruses encode a reverse transcriptase, or RNA-directed DNA polymerase, that converts the single-stranded RNA viral genome found in virions into a double-stranded DNA form. Another virally-encoded enzyme, integrase, then integrates this linear DNA into the genome of a host cell, where it is called a provirus.

The rapid progress in understanding the biology of HIV was largely due to the work done on other retroviruses in the seventy or so years before AIDS was described. During the first decade of the twentieth century, two studies on tumors in chickens led to the discovery of the avian sarcoma/leukosis viruses (ASLV) (Ellerman and Bang 1908, Rous 1911). From the time of these initial reports until the 1960's, many more retroviruses were discovered and described in terms of their host range and the natural histories of the diseases they caused. Due to advances in biochemical, structural, cell culture, and molecular techniques, the emphasis in retrovirology in recent decades has been on cellular pathogenesis and a molecular description of retroviruses and retroviral replication. By the time HIV was isolated and shown to be the pathogenic agent in AIDS, the major protein components of retroviruses, their basic replication cycle, the general structure of the retroviral genome, and the nucleotide sequences of certain specific retroviruses had been described. Since the discovery of HIV, the field of retrovirology has expanded and progress has accelerated even more.

Mutants have been very useful tools in the study of retroviruses. With the advent of molecular biology, directed mutations have been made and screens of random mutations have been done to determine the functions of various parts of the retroviral genome. For HIV, the combination of knowing the nucleotide sequence of the genome and having structural information on most of the proteins makes it very interesting to have a detailed description of the phenotypes of mutants throughout the genome. Ideally, one would be able to study the effects of different types of mutations (point mutations, deletions, insertions, replacements) at every position in the genome on different points in the replication cycle (transcription, translation, packaging, budding, host-cell attachment, entry, uncoating, reverse transcription, nuclear entry, integration). Making individual mutants and studying their phenotypes one by one is now possible, but not experimentally feasible. Therefore, we have developed a method to study large numbers of mutants in parallel. This technique is restricted to two classes of mutations (insertions and replacements), but allows the collection of quantitative information on the effects of many mutations on several steps of the viral replication cycle in a highly parallel manner.

## **Overview of Mutagenesis**

Mutagenesis is a versatile and powerful tool in studying the function of nucleic acids. Mutagenesis can be performed either *in vivo* or *in vitro*, on a small piece of cloned DNA or on the intact genome of an organism, randomly or in a directed fashion. There are three types of mutagens in common use: chemicals (e.g. alkylating agents), radiation (e.g. X-rays or UV radiation), and enzymes (e.g. Taq polymerase in PCR mutagenesis or transposases in transposon-mediated mutagenesis). Standard methods of random mutagenesis involve subjecting the nucleic acid of interest to mutagenesis and either selecting/screening the resulting mutant population for a particular phenotype (e.g. resistance to an antibiotic) or isolating mutant clones and testing the characteristics of individual mutants one at a time (e.g. rate of replication). Directed mutagenesis involves constructing and testing mutants one at a time. These two basic strategies are useful in certain circumstances: the isolation of mutants where one has a good positive selection and the examination of a limited number of individual mutants. However, these techniques quickly become tedious if the goal is to quantitatively determine the behavior of large numbers of mutants. Recently, several methods aimed at assessing large numbers of mutants in parallel have been reported, including signature-tagged transposon mutagenesis (Hensel et al. 1995), genome-scale genetic footprinting (Smith et al. 1996), and high-resolution genetic footprinting (Singh et al. 1997).

The objective of this study was to obtain a detailed functional map of a portion of the HIV genome by examining between 100 and 200 insertional

mutations distributed over a 1000 nucleotide region, which encompassed the 5'-LTR (Long Terminal Repeat), the p17 (matrix, MA) gene, and p24 (capsid, CA) gene of HIV. Each mutant was to be mapped at single-nucleotide resolution and quantitatively assessed for its affect on viral replication. Given these criteria, the high-resolution genetic footprinting technique was the method of choice.



## **High-resolution Genetic Footprinting**

High resolution genetic footprinting was developed as a method to make and gather quantitative information on large numbers of mutants en masse. The basic concept consists of constructing a library of insertion or replacement mutants, where the different mutants contain the same insertion or replacement sequence, differing only in the position of the mutation. The idea for using an integrase or transposase enzyme to make the mutations and analyzing the population of mutants by PCR originated from a paper by Pryciak and Varmus (Pryciak and Varmus 1992). These authors were actually studying the effect of DNA conformation on target site preferences of retroviral integrases. However, their work showed that large numbers of integration events could be tracked in parallel and mapped to single-nucleotide resolution.

Moloney Murine Leukemia Virus integrase can be used to integrate short double-stranded oligonucleotides in a concerted fashion into a circular double-stranded DNA target in vitro. In this concerted reaction, the terminal two nucleotides of the upper strands of two double-stranded oligonucleotides are clipped off, leaving two-nucleotide 5' overhangs. The newly exposed terminal 3' hydroxyl groups of these oligonucleotides are then used to attack 5' phosphates in the target DNA staggered by 4 base-pairs, producing a linear target DNA with an oligonucleotide covalently joined to each end. There are 4-nucleotide gaps in the target DNA and an extra 2-nucleotide 5' extension on the oligonucleotides at each end. Both of these features can be eliminated by doing a run-off reaction using a DNA polymerase such as Taq polymerase. MLV integrase is relatively

insensitive to the sequence of the target DNA, resulting in integration events at many different sites.

In 1997, Singh, Crowley, and Brown demonstrated the utility of MLV integrase as a tool for genetic footprinting in high-resolution functional mapping of the SupF gene, which encodes an amber suppressor tRNA. The oligonucleotide used for integration contained three types of sequences: a viral end sequence that allowed MLV integrase to recognize the oligonucleotide as a substrate; a Bsg I restriction enzyme site; and a Not I restriction enzyme site. Insertion mutants were made by digesting the products of the concerted integration reaction with Not I, creating complementary cohesive ends, and recircularizing the target DNA by ligation. The resulting insertions included a 4-base-pair duplication in the target DNA and a central Not I site. Replacement mutants were generated by digesting with Bsg I, a type IIs restriction enzyme that cuts 16/14 nucleotides away from its recognition sequence, allowing cleavage within the target DNA sequence. The 12 base-pairs deleted from the target DNA sequence in this way were replaced by ligating in a 12-base-pair oligonucleotide containing an Nde I site. Both the insertion and replacement libraries were subjected to a selection that required the function of the SupF gene. The libraries before and after selection were analyzed and compared using PCR-based assays. To analyze the insertion library samples, PCR was performed using one oligonucleotide primer complementary to the sequence of the insert oligonucleotide and a second, <sup>32</sup>P-labelled, primer complementary to a fixed position in the target DNA. Each mutant in the library gave a product of unique

size that depended on the position of the insertion. Since the library consisted of mutants at many different positions, subjecting the products of the PCR reactions to electrophoresis through a denaturing polyacrylamide gel resulted in a ladder of bands. Bands that represented clones defective in SupF function were present in the pre-selection library and absent in the post-selection library, giving a functional footprint of the SupF gene. The oligonucleotide used for the replacement library was too short for efficient priming for PCR. Therefore, an alternative PCR strategy (which I will refer to as the “flanking PCR/restriction digestion” method) was designed to analyze the replacement library samples. Two fixed-position primers to target DNA sequences were used, one of which was labelled with  $^{32}\text{P}$ . The PCR products were digested with Nde I, which cleaved within the replacement sequence, yielding a unique-sized radioactively-labeled product for each mutant, the size of which again depended on the position of the replacement.

## **Overview of the Genetic Footprinting Aspect of the Current system**

Several modifications to the method reported by Singh, Crowley, and Brown (1997) were required to adapt it for the study of the HIV genome. Most significantly, the enzyme used for mutagenesis was changed from MLV integrase to MuA transposase. The basic strategy for introducing insertions into a target sequence remained the same, including a concerted integration reaction followed by gap-repair, restriction endonuclease digestion, and ligation reactions (figure 1). MLV integrase performs the concerted reaction inefficiently, requiring amplification of the integration products by PCR. Since the mixture of integration products is composed of circularly permuted linear pieces of DNA, troublesome PCR side-reactions tend to occur, with template DNAs priming off of one another. These reactions occur less frequently when smaller template DNAs are used, limiting the target DNA size to approximately 1000 base-pairs. This size limitation was unduly restrictive for the experiments on HIV, which involved mutagenizing stretches of the genome of up to 1.5 kilobases cloned into a 2.5 kilobase vector. MuA transposase is a much more efficient and robust enzyme, allowing the intermediate PCR amplification step to be eliminated. The most serious drawback to MuA transposase is that it is more finicky about the sequence of the target DNA. This property leads to an uneven representation of mutants, such that fewer mutants can be conveniently analyzed. An incidental difference between MLV integrase and MuA transposase is that MuA transposase produces a 5 base-pair rather than a 4 base-pair duplication. A

second modification was the optimization of the analysis procedure. The insertions made in HIV contained only 10 unique base-pairs, too short for efficient priming. However, the insertions contained a Not I site, permitting the use of the flanking PCR/restriction enzyme digestion analysis method. The samples in the HIV experiment were more complex than those in the SupF studies, leading to higher background from incomplete PCR extension products. The level of these background products was greatly reduced by performing the PCR using one <sup>32</sup>P-labelled target DNA primer and one biotinylated target DNA primer, treating the products of the PCR reactions with a single-stranded binding resin, adsorbing the PCR products to streptavidin-agarose beads, and digesting the products off of the beads with Not I (figure 2). Using this technique, bands visible on the denaturing polyacrylamide gel result from PCR products containing both a radioactive and a biotinylated primer, eliminating incomplete extension products.

The proviral HIV clone used in the work described here is approximately 9000 base-pairs long, and is carried in an approximately 2500 base-pair vector, making a total of 11500 base-pairs in the plasmid. During the mutagenesis procedure, it is necessary to separate the products of concerted integration by MuA transposase (linear) from the unintegrated target molecules (supercoiled circular) and products of single integration events (branched circular) by agarose gel electrophoresis. It is difficult to cleanly separate these species if the target molecule is more than 5000 base-pairs in length. Moreover, during the analysis step, only 200 to 300 base-pairs are examined at any given time. If 11500 base-

pairs are mutagenized, the fraction of PCR products containing insertions in a average 300 base-pair segment would be  $300/11500$ , or 2.6%. This value would result in an unacceptably low signal-to-noise ratio on the footprinting gel.

Therefore, segments of the HIV genome ranging from 500 to 1600 base-pairs were subcloned for mutagenesis (corresponding to a plasmid size of 3000 to 4100 base-pairs). In order to ensure a good representation of mutant clones in the libraries, we wanted to achieve an average of at least 100 “hits” per base-pair. For a 4100 base-pair construct, therefore, we would aim for a library with at least 410,000 elements, a number which we found to be experimentally feasible to attain.

The mutagenized proviral segments were recloned into a plasmid containing the complete sequence of the provirus. Since a fraction of the “hits” were in vector sequences (the fraction being approximately proportional to the percentage of the entire plasmid composed of vector sequences), approximately 60% to 80% of the clones in the resulting libraries contained no insertions. This situation was to our advantage, since the wild-type clones did not interfere with testing mutations in cis-acting elements, and were actually desired to provide helper functions during the first round of infection for testing mutations in coding sequences. Fewer undesired side-products were obtained during cloning if the mutagenesis was done on proviral fragments carried in an ampicillin-selectable vector and the intact provirus was carried in a kanamycin-selectable vector. The principle potential troublemakers resulted from ligations between two insert-containing vector fragments (i.e. vector fragments that had been “hit” during

mutagenesis), which could then homologously recombine using the insert sequences, generating very small plasmids which replicate very quickly and take over the culture.

A library of 15-nucleotide insertion mutants was made using MuA transposase in a replication-defective HIV background carrying the puromycin resistance gene in place of the env gene. The insertions contained a Not I restriction enzyme site, which was used in the analysis phase of the experiments. This library of replication-defective mutagenized proviruses was introduced into producer cells (details on the design of specific experiments are given in Chapter 4). Pseudotyping with VSV-G, a single round of viral production and infection was carried out. Nucleic acid samples were collected at various steps (figure 3), and footprinting these samples allowed us to examine the effect of different mutations on several steps in the replication cycle in parallel (figure 4). For example, samples of producer cell RNA ("cellular RNA") contained lower proportions of transcripts from proviruses containing mutations that interfere with transcription, mRNA stability, or polyadenylation than samples of producer cell genomic DNA. Similarly, RNAs with mutations that preclude efficient translation, dimerization and packaging of viral RNA, assembly of viral particles, or viral budding were underrepresented in pools of RNA in extracellular virions ("virion RNA") compared with pools of viral RNA in producer cells ("cellular RNA"). After infection of a fresh population of host cells with these virions, mutants defective in such processes as packaging of the tRNA primer, entry, uncoating, reverse transcription, nuclear entry, or integration were less well represented in pools of

integrated viral DNA ("infected cell genomic DNA ") than in pools of virion RNA. This scheme permitted the assignment of defects in viral replication caused by individual mutants to phases in the viral life cycle without the necessity of testing each mutant alone.

One could isolate interesting mutant clones in one of two ways. First, if one identified specific clones by footprinting, one could PCR those clones out of the mutant library using primers that would prime only from clones with an insert at the desired location (see figure 5). Second, if one were interested in isolating clones that were enriched by a selection scheme, one could PCR a region out of a sample of post-selection nucleic acid and clone the PCR products en masse. To eliminate wild-type clones, one could digest the population of plasmids with Not I and purify the linearized plasmids (those that have a Not I-containing insert).

In the experiments described here, mutants were selected for their ability to perform various steps in the viral replication cycle. Selection strategies other than the one described here can be easily used. For example, to study viral resistance to therapeutic agents one could subject a library of mutants in protease to a protease inhibitor and use footprinting to identify regions where insertions lead to resistant mutants.



## **Chapter 2**

### **Introduction to High-resolution Genetic Footprinting of HIV**

The retroviral life cycle is fairly well understood mechanistically and genetically. Mechanistically, the molecular events involved in virion production and infection are known in outline. Various processes, particularly transcription, assembly, reverse transcription, and integration, have been investigated and described in some detail (reviewed recently in Coffin et al. 1997). The genomes of several retroviruses have been subjected to extensive mutagenesis, both natural and experimental. As a result, functional regions of the viral genome, such as the long terminal repeats (LTRs) and sequences encoding the viral proteins, have been mapped. However, the mutations that have been made thus far are unevenly distributed across the genome and diverse (e.g. point mutations, insertions, and deletions of different sizes and sequences). Moreover, the effects of many of these mutations have not been studied in a uniform or comprehensive manner.

In the experiments described in this report, the goal was to create a high-resolution map of a one kilobase segment near the 5' end of the HIV RNA genome defining features essential for major steps in the viral replication cycle. This region of the HIV genome contains several previously identified functional elements (see figure 6), including several cis-acting elements and sequences encoding the matrix and capsid proteins. By studying a large number of mutants of uniform construction in a thorough and quantitative manner, we strove to gain

detailed insight into known elements in the viral genome and to define novel features.

Many of the cis-acting sequences overlap with each other or with coding sequences. The multifunctional nature of certain sequences in the HIV genome can create difficulties in assigning unambiguous functions to these sequences. The TAR stem-loop structure is important in transcription of the viral genome (Berkhout et al. 1989; Selby et al. 1989; Roy et al. 1990a; Roy et al. 1990b; Feng and Holland 1988; Dingwall et al. 1989; Cordingley et al. 1990; Gait and Kam 1993) and overlaps with the sequences in R that are used during the first strand-transfer event in reverse transcription (Coffin and Haseltine 1977; Haseltine et al. 1977; Schwartz et al. 1977; Stoll et al. 1977; Coffin et al. 1978). R also contains a polyadenylation signal. At the 3' end of U5 resides the sequence encoding the 3' att site, a short (~15 base-pair) sequence required by integrase for efficient integration of the viral genome into host cell genomic DNA (Bushman and Craigie 1991; LaFemina et al. 1991; Leavitt et al. 1992; Sherman et al. 1992; van den Ent et al. 1994; Vicenzi et al. 1994). Adjacent to the att site is the primer binding site, an eighteen nucleotide sequence complementary to the eighteen terminal nucleotides of tRNA-Lys, which is used to prime the negative strand during reverse transcription. This sequence also plays a role in the second strand-transfer step of reverse transcription (Rhim et al. 1991). The region of the genome from the end of the LTR into the beginning of the matrix coding sequence contains an AP-1/AP-3 site, a DBF-1 site, and a SP-1 site (Verdin et al. 1990; Van Lint et al. 1991), a splice donor sequence used to produce the

mRNA for the envelope protein, and sequences that contribute to dimerization and packaging of the viral single-stranded RNA genome (Lever et al. 1989; Luban and Goff 1994; McBride and Panganiban 1996; Laughrea et al. 1997a; Laughrea et al. 1997b; Clever and Parslow 1997).

The matrix and capsid proteins of retroviruses are translated as part of the gag polyprotein and subsequently cleaved from the polyprotein by a retrovirally-encoded protease. Matrix contains a N-terminal myristoyl group and a nearby basic region, both of which assist in targeting the unprocessed gag polyprotein to the host cell plasma membrane during assembly (Gottlinger et al. 1989; Bryant and Ratner 1990; Zhou et al. 1994). Matrix also interacts with the cytoplasmic tail of the viral envelope protein (Yu et al. 1992b; Facke et al. 1993). In some, but not all, experiments, HIV matrix has been demonstrated to assist in nuclear entry of the HIV pre-integration complex (Bukrinsky et al. 1993; Gallay et al. 1995a; Gallay et al. 1995b; von Schwedler et al. 1994; Fouchier et al. 1997; Freed et al. 1995). There are indications that the C terminus of matrix may play a role in uncoating (Yu et al. 1992a), and it has been suggested that matrix can bind to RNA (Bukrinskaya et al. 1992). HIV capsid is thought to be the major structural protein making up the viral core. Mutations in capsid have been shown to be defective in viral assembly or in an early step in viral replication, between entry and reverse transcription (Mammano et al. 1994; Wang and Barklis 1993; Reicin et al. 1995; Reicin et al. 1996; Dorfman et al. 1994a). Capsid interacts with a host protein, cyclophilin A, which is specifically incorporated into viral

particles and seems to play a role in uncoating (Luban et al. 1993; Braaten et al. 1996; Franke et al. 1994; Thali et al. 1994).

High-resolution genetic footprinting has been used to map functionally important domains in the SupF gene (Singh et al. 1997). We have employed a modification of this method to define functional domains in a portion of the HIV genome. A library of insertion mutants was made in a region of the HIV genome using MuA transposase and selected en masse for the ability to undergo various phases of the viral life cycle. Each mutant contained a single insertion, which included a restriction endonuclease recognition sequence at a "random" position (in fact the MuA transposase demonstrates preferences for certain target sequences). An assay involving a PCR reaction and a restriction endonuclease digestion was then performed on nucleic acid samples of the library taken before and after each phase to assess the recovery of each mutant through that phase. This assay generated a product of unique length for each mutation; the length depended on the position of the insertion in the HIV sequence. Therefore, the nucleic acid samples analyzed, which were mixtures of mutants, produced mixtures of products of different lengths, which were resolved as bands on denaturing polyacrylamide gels. Mutants defective for a given phase of the viral life cycle were eliminated at that step, leading to a depletion of the corresponding bands. This scheme permitted the assignment of defects in viral replication caused by individual mutants to phases in the viral life cycle without the necessity of testing each mutant alone.

# Chapter 3

## Materials and Methods

### Plasmids

The HIV replication-defective proviral clone mutagenized in this report (HIV puro) was derived from pHIV-AP $\Delta$ env $\Delta$ Vif $\Delta$ Vpr (Sutton et al. 1998) and subcloned into either Bluescript KS+ (Stratagene) or pBS -Kan (a Bluescript KS+-derived vector where the ampicillin-resistance gene was replaced by the kanamycin-resistance gene). pHIV-AP $\Delta$ env $\Delta$ Vif $\Delta$ Vpr was constructed from HIV-AP, an HIV proviral clone containing the human placental alkaline phosphatase in place of nef (He and Landau 1995), by making a large deletion to eliminate most of env, vif, and vpr. To make HIV puro, the human placental alkaline phosphatase gene was replaced by the puromycin resistance gene driven by the SV40 promoter (Morgenstern and Land 1990). In addition, host DNA sequences flanking the proviral sequences were eliminated. PCR mutagenesis was used to eliminate the five Bsg I sites originally present in the plasmid (G  $\rightarrow$  C at position 1222, C  $\rightarrow$  G at position 2574, A  $\rightarrow$  C at position 4856, A  $\rightarrow$  T at position 5755, and A  $\rightarrow$  C at position 5884. These changes did not detectably affect viral replication.

Fragments of HIV puro were subcloned into Bluescript KS+, mutagenized (see below) in the context of these smaller plasmids, and subsequently cloned back into HIV puro to generate libraries of mutant proviruses.

## **Mutagenesis**

The mutagenesis procedure was a modification of the method described by Singh et al. 1997. MuA transposase was a generous gift from Kiyoshi Mizuuchi and Harri Savilahti. The double-stranded oligonucleotide (Not15) used for mutagenesis was made by annealing Not15A (5'-TGCGGCCGCGCACGAAAAACGCGAAAGCGTTTCACGATAAATGCGAAAAC-3') and Not15B (5'-GTTTTCGCATTTATCGTGAAACGCTTTCGCGTTTTTCGTGCGCGGCCGCA-3') in 50 mM NaCl. The integration reaction was performed by incubating 25 pmol of Not15, 5 µg target plasmid, and 50 pmol MuA transposase (the volume of MuA transposase used was determined by a series of titration experiments) with 25 mM Tris pH 8.0, 100 µg/ml BSA, 15% glycerol (w/v), 144 mM NaCl, 0.1% Triton X-100 (v/v), 10 mM MgCl<sub>2</sub>, and 15% DMSO (v/v) in a 0.5 ml reaction volume at 30 °C for 1 hour (Savalahti et al. 1995). Reaction products were phenol/chloroform extracted once, chloroform extracted once, precipitated in 0.3 M NaOAc pH 5.2 and 70% ethanol, washed with 70% ethanol, dried briefly under vacuum, and resuspended in 10 mM Tris.HCl/1 mM EDTA pH 8.0. Plasmids linearized by concerted integration events were separated from plasmids that had undergone single-ended integrations events or no integration events by agarose gel electrophoresis. The products of concerted integration events were purified (Qiaquick gel extraction kit) and the 5-nucleotide gaps resulting from the integration events were repaired by Taq DNA polymerase-mediated nick translation (incubation in 1x Taq DNA polymerase buffer (Perkin Elmer), 2.5 mM

MgCl<sub>2</sub>, 2.5 mM dATP, 2.5 mM dCTP, 2.5 mM dGTP, 2.5 mM dTTP, and 2 units Taq DNA polymerase at 72 °C for 10 minutes in a 100 µl reaction volume). The products of these nick translation reactions were purified (Qiaquick PCR purification kit), then digested with Not I (New England Biolabs). Recircularization of the linear plasmids by ligation of the cohesive ends resulted in 15 base-pair insertions.

### **Cell culture**

293, 293T, and HOS cells were grown in Dulbecco's Modified Eagle's Medium containing 4.5 g/l glucose and 10% Defined Fetal Calf Serum (Hyclone). 293T cells were used for all transient transfection experiments. 293 cells were used for all stable transfection experiments and infections by virions produced by transient transfection. HOS cells were used for infections by virions produced from infected or stably transfected cells. Cells were grown at 37 °C in 5% CO<sub>2</sub> in a water-jacketed incubator. Puromycin selection was performed using 2.5 µg/ml puromycin (Sigma) for 293 cells and 5 µg/ml puromycin for HOS cells.

### **Transfections**

Transient and stable transfections using the Lipofectamine Plus kit (Gibco/BRL) were performed according to the recommended protocol. 30 µg total plasmid DNA, 60 µl Plus reagent, and 40 µl Lipofectamine were used for each 15 cm tissue culture dish. For stable transfections, puromycin selection was initiated 48

hours post-transfection. For transient transfections, the media was changed 48 hours post-transfection and virus was harvested 72 hours post-transfection.

## **Infection**

Viral stocks were diluted to the desired concentration in media containing 4 µg/ml polybrene (Sigma) and used to infect cells for 2 hours at 37 °C. Puromycin selection was initiated 48 hours post-infection.

## **Nucleic acid preparation**

Plasmid DNA: Plasmid DNA was purified using the Qiagen plasmid DNA kit and subsequently banded in a cesium chloride gradient (Sambrook et al. 1989).

Genomic DNA: The Qiagen Blood and Cell Culture Genomic DNA kit was used to prepare genomic DNA from tissue culture samples.

Total cellular RNA: Total cellular RNA was prepared using the Qiagen RNeasy total RNA kit.

Viral RNA: Viral RNA was prepared by pelleting virions by ultracentrifugation (28,000 rpm for 2 hours at 4 °C in a Beckman SW 28 rotor), pouring off the supernatant, resuspending the viral pellet in the residual media, and using the Qiagen Oligotex direct mRNA kit.

## **Sequencing reactions**

Sequencing reactions were performed using the Sequenase sequencing kit from USB.



## **Reverse transcription**

Reverse transcription of cellular RNA and virion RNA samples was performed using 100 ng template RNA with the HIV-specific oligonucleotides HIV521 (5'-GGGAGCTCTCTGGCTAACTAGGG -3') and HIV1573r (5'-CATCCTATTTGTTTCCTGAAGGG -3') according to the manufacturer's instructions (Titan reverse transcription kit (Boehringer-Mannheim)).

## **PCR**

PCR was performed in 20 mM Tris.HCl pH 8.55, 150 ng/ml BSA, 16 mM  $(\text{NH}_4)_2\text{SO}_4$ , 3.5 mM  $\text{MgCl}_2$ , 625  $\mu\text{M}$  each dNTP, 0.25  $\mu\text{M}$  each primer, and 1 unit per 50  $\mu\text{l}$  reaction Taq DNA polymerase (AmpliTaq from Perkin-Elmer). "Cold" PCR conditions consisted of 2 minutes at 94 °C followed by 30 cycles of 30 seconds at 94 °C, 30 seconds at 55 °C, and 2 minutes at 72 °C. "Hot" PCR conditions consisted of 2 minutes at 94 °C followed by 25 cycles of 30 seconds at 94 °C, 30 seconds at 55 °C, and 1 minute at 72 °C.

## **Pretreatment of streptavidin-agarose beads**

Streptavidin agarose beads (Sigma) were incubated in the presence of poly dI-dC (200  $\mu\text{g}$  per ml streptavidin agarose slurry) in 1x binding buffer (12% glycerol (v/v), 12 mM HEPES pH 7.9, 4 mM Tris.HCl pH 8.0, 60 mM KCl, 1 mM EDTA, 1 mM DTT) for one hour at 25 °C. The beads were then washed four times in 1x binding buffer (1 ml buffer/ml slurry) and finally resuspended in 1x binding buffer to reconstitute the initial volume of slurry.

### **Single-stranded Affinity Matrix (SSAM) treatment of PCR reactions**

8  $\mu$ l of 8M Lithium chloride and 10  $\mu$ l of SSAM (Clontech) were added to each 50  $\mu$ l PCR reaction. The mixture was incubated for 10 minutes at room temperature with agitation every two minutes. The SSAM resin was then removed by passing the mixture through a 0.45  $\mu$ m spin filter (Millipore). Alternatively, BNDC resin (Sigma) was suspended in 1M Lithium chloride (0.5 g resin in 2.5 ml 1M Lithium chloride) for 60 minutes at room temperature. 50  $\mu$ l of this suspension was used per PCR reaction.

### **Footprinting**

Initial amplification of nucleic acid samples was done according to the "cold" PCR protocol using HIV-specific primers HIV37 (5'-TGGAAGGGCTAATTCACCTCCCAAAG -3'), HIV493 (5'-TCTCTCTGGTTAGACCAGATCTG -3'), HIV521(5'-GGGAGCTCTCTGGCTAACTAGGG -3') and HIV1573r (5'-CATCCTATTTGTTCTGAAGGG -3'). 10 ng of plasmid samples, one-tenth of the products of reverse transcription reactions (equivalent to 10 ng input RNA), or 0.5  $\mu$ g genomic DNA samples were used as templates. 10 ng of "cold" PCR products were used for "hot" PCR reactions. For "hot" PCR reactions, one HIV-specific primer was labeled with  $^{32}$ P (T4 polynucleotide kinase, New England Biolabs) while the other primer was biotinylated (Operon). High-specific-activity  $^{32}$ P-gamma-ATP (160  $\mu$ Ci/mmol, 23 pmol/ $\mu$ l, ICN) was used for radiolabelling at a stoichiometry of 1 pmol ATP/1 pmol oligonucleotide. "Hot" PCR products were

treated with SSAM, purified (Qiaquick PCR purification kit), then adsorbed to 50  $\mu$ l pretreated streptavidin-agarose beads (Sigma) in 1x binding buffer for one hour at 25 °C. The beads were then washed twice with 0.5 ml 1x binding buffer for 15 minutes at 25 °C, washed once with 0.5 ml 1x restriction enzyme buffer 3 (New England Biolabs), and incubated in 50  $\mu$ l 1x restriction enzyme buffer 3 (New England Biolabs) containing 20 units Not I restriction enzyme (New England Biolabs) for 1 hour at 37 °C. The supernatant from this digestion step was separated from the beads by centrifugation through a Micro Bio-spin column (Bio-rad) at 3,000 rpm for 1 minute at room temperature in a tabletop microfuge. The supernatant was then precipitated in 0.3 M NaOAc pH 5.2 and 70% ethanol in the presence of 5  $\mu$ g linear acrylamide, washed with 70% ethanol, dried briefly under vacuum, and resuspended in 3  $\mu$ l 10 mM Tris.HCl/1 mM EDTA pH 8.0 + 3  $\mu$ l 2x formamide loading dye (95% deionized formamide/25 mM EDTA pH 8.0/0.25% bromophenol blue/0.25% xylene cyanol). Samples were heated at 95 °C for 2 minutes, placed immediately onto ice, and analyzed by electrophoresis through 6% acrylamide (19:1 acrylamide:bis-acrylamide)/1x TBE/7 M urea sequencing gels (2  $\mu$ l sample per lane). Gels were dried for 1.5 hours at 80 °C under vacuum and exposed to Biomax MR film (Kodak).

### **Quantitation**

Autoradiographs were scanned using a flatbed scanner (Hewlett-Packard) at 300 dpi resolution, with brightness and contrast set at 125 (50%). Scanned images were read into a Matlab-based application (see Appendices D and E) by which

individual bands were selected and quantitated for peak intensity values. Data from different footprinting reactions and different gels were normalized by fitting profiles of the relative intensities of bands within each run using an algorithm that minimizes the sum of the coefficients of variance for the mutants weighted for the number of measurements for each mutant (figure 7). The normalized data were then averaged. Data from triplicate experiments were normalized using the same algorithm and averaged (see Appendices D and E).

# Chapter 4

## Results

### Creation of a library of insertion mutants

The objective was to make a large number of mutations of the same type at diverse positions in a one kilobase stretch of the HIV genome and to assess the performance of each mutant at several points in the viral replication cycle. A library of 15-base-pair insertion mutants was constructed by in vitro transposition in a replication-defective HIV provirus containing the puromycin acetyltransferase gene driven by an internal promoter in place of the env gene. The mutations were made specifically in the segment of the HIV genome (positions 37-1550) including the 5'-LTR, the 5' untranslated region, the complete matrix gene and the 5' half of the capsid gene. Mutants are numbered according to the nucleotide position immediately 5' to the insertion.

The MuA enzyme was used to perform an in vitro transposition reaction, introducing a pair of double-stranded DNA oligonucleotides into a double-stranded circular target DNA molecule (figure 1). The oligonucleotides contained both sequences necessary for recognition by MuA and sequences recognized by the Not I restriction endonuclease. MuA inserts the oligonucleotides into the target DNAs in a staggered fashion, such that the products of the transposition reaction were gapped linear double-stranded DNA molecules, with an oligonucleotide located at either end. After filling in the gaps by nick translation, the reaction products were digested Not I, generating compatible cohesive ends, which were ligated. The final products were circular DNA molecules containing

the inserted sequence, 5'-TGCGGCCGCA-3', flanked by five base-pair duplications of the target sequence. The insertions retained the NotI recognition sequence, which was used during the analysis procedure. Insertional mutants were generated using MuA transposase rather than MLV integrase, the enzyme used in the original footprinting experiments, since MuA transposase executes the necessary *in vitro* concerted integration reaction more robustly (Crowley et al., manuscript in preparation). Sixteen individual mutant clones, at positions 189, 238, 268, 358, 557, 622, 776, 926, 1012, 1045, 1067, 1175, 1264, 1267, 1277, and 1399, were isolated and sequenced. These clones were used as markers to determine the location of insertions during analysis.

Insertions were designed such that mutations in coding sequences would be in-frame insertions of five codons. The identity of the amino acids encoded by the insertions depended on both the reading frame and the sequences in the target DNA adjacent to the insertion site.

The positions of insertion mutants for which data were obtained are indicated in figure 6. Although the collection of mutants is extensive, the sequence space was not saturated, since MuA transposase does not make insertions at the same frequency at all sites. Moreover, since transcription starts at R in the 5'-LTR, the effects of mutations in U3 could not be assessed. Examination of nucleic acid samples before and after a single round of transcription by genetic footprinting confirmed this loss of mutants in U3 at transcription, indicating that nucleic acid samples were not contaminated with plasmid DNA from the initial transfections.

## **Sampling populations of mutants at different steps in the viral replication cycle**

To study the effects of insertions on cis-acting elements (e.g. transcriptional modulators, the packaging sequence, and the viral att site), the library was either transiently or stably transfected into producer cells. A plasmid encoding VSV-G was transiently transfected into the producer cells to pseudotype the env-defective virions. A single round of infection was then performed. Nucleic acid samples were collected at various steps during this experiment (see figure 3). Depletion of mutants at different steps in the viral replication cycle was followed by analyzing these nucleic acid samples by genetic footprinting.

A similar strategy was utilized to determine the effects of insertions in trans-acting sequences. Since more than one piece of DNA often enters a given cell during transfection, complementation can occur in a mixed population between trans-acting elements in a transfection experiment (see figure 8). In order to study the functions of trans-acting factors in the absence of complementation, a first round of transient transfection was conducted, cotransfecting the mutant library with a VSV-G expression construct. The goal was to produce a VSV-G pseudotyped, phenotypically mixed population in which mutants with defective trans-acting functions were rescued by complementation. Since approximately half of the clones in the library were wild-type (i.e. did not contain an insertion), this complementation was easy to achieve. These virions were then used to infect fresh host cells at a low multiplicity of infection (1

infectious unit for every 20 cells) such that each cell would receive only one viral genome. According to a Poisson distribution, 95.12% of the cells would receive 0 virions, 4.76% of the cells would receive 1 virion, and 0.12% of the cells would receive more than 1 virion. Hence, of the cells that received at least one virion, approximately 2.5% received more than one virion. The infected cells were selected using puromycin, and this pool of cells was used as the starting population of producer cells for a single round of infection. Nucleic acids were purified at various steps during this experiment and analyzed by genetic footprinting, allowing us to study the effects of mutations on the functions of trans-acting sequences.

From results obtained in these studies, it is now clear that complementation of trans-acting factors occurred very efficiently during the first round of infection in our transient transfection experiments but not to any appreciable degree in our stable transfection experiments. The number of proviruses per cell has not been directly measured. However, if the number of proviruses per cell is  $T$  for our transient transfection experiments and  $S$  for our stable transfection experiments, our results suggest that  $T$  is greater than  $S$ . In the simple case where the wild-type version of a gene is dominant and a mutant version is recessive, we would expect  $T$  to be greater than or equal to two and  $S$  to be equal to one. However, the viral proteins studied in our experiments probably function as oligomers, such that mutants might display dominant negative phenotypes. Thus, in our experiments,  $S$  might be larger than one, with  $T$  significantly larger than  $S$ . In fact, as mentioned above, multiple pieces of DNA



can enter a single cell during transfection, leading us to expect S to be larger than one.

### **Description of footprinting analysis procedure (figure 2)**

Nucleic acid samples collected at various points in the viral life cycle were subjected to an initial round of amplification by either PCR (for DNA samples) or RT-PCR (for RNA samples). PCR was then performed on these pre-amplified samples using one <sup>32</sup>P-labelled DNA primer and one biotinylated DNA primer. The primers were complementary to HIV sequences and flanked the region to be analyzed. The products of this second PCR reaction were first treated with a single-stranded binding resin to remove incomplete extension products and then bound to streptavidin-agarose beads. The radioactively labeled portions of the PCR products containing Not I sites were digested off the beads with Not I, concentrated, and subjected to electrophoresis on denaturing polyacrylamide-urea gels. A typical gel is shown in figure 9.

### **Cis-acting versus trans-acting elements**

Mutations in cis-acting and trans-acting features can often be distinguished by differential behavior in complemented versus uncomplemented infection cycles. One would expect mutations in cis-acting sequences to show their phenotypes in the presence or absence of complementation, while mutations in trans-acting sequences should be apparent only when uncomplemented. Trans-acting sequences are typically considered to be coding

sequences. However, due to the pseudodiploid nature of retroviruses and peculiarities in certain steps of viral life cycle (such as assembly and reverse transcription), there can conceivably be trans-acting sequences in the HIV genome that act at the nucleic acid level.

Data showing the behavior of individual mutants in single-cycle infections are given in figure 10. Figure 10A shows survival of mutants through one round of complemented infection (first round transient transfection), while figure 10B shows the behavior of mutants through one round of uncomplemented infection (second round transient transfection). Mutations that affect replication in both complemented and uncomplemented infections to a significant degree (greater than 55% depletion during one round of infection) appear to be localized to the region 5' to position 847. Since most of this region appears to be composed of noncoding sequences (up to position 828, where the matrix coding sequence begins), it is not surprising that we found cis-acting elements in this area of the genome. Insertions in sequences between positions 847 and 1524 display no clearly discernible effects in complemented infections, while many of these insertions interfere with infection in the uncomplemented situations. Since coding sequences for matrix and capsid lie in this stretch of the genome, one might have expected to find trans-acting functions here.

## **Mutants in non-coding sequences defective in transcript formation or stability**

The six mutants in the TAR region (492-542) were severely compromised in their ability to replicate. The primary deficiency was in transcript production or stability (only qualitative data is given as the region surveyed is too close to the end of the viral RNA for accurate quantitation). Most likely, these mutants are defective for tat binding, which would result in a low efficiency of transcription. The phenotype appears in the presence of complementation, reconfirming the cis-acting nature of the affected noncoding sequences.

In the transient transfection experiment, insertions at positions 564, 573, and 583 had detrimental effects on the second, but not the first, round of infection (figure 10). For the second round of the transient transfection experiment, the effects of the insertions at all three positions were most pronounced during transcript formation (figure 11). However, in the stable transfection experiment, the mutations at positions 564 and 583 resulted in decreases in fitness in the phase of the life cycle occurring between collection of the cellular RNA and viral RNA samples (figure 12). The most probable explanation for these observations is that these mutations, which are in and around the polyadenylation consensus sequence (563-568), interfered with polyadenylation. This defect would not be observed in the first round of infection since only the 5'-LTR was mutagenized and it was not until the first round of reverse transcription that mutations were transferred to the 3'-LTR, where the operative polyadenylation signal lies. In addition, the mutations at positions 564 and 583 might interrupt partially trans-

complementable sequences that contribute to packaging of the viral RNA genome (see below for further discussion of packaging sequences). The dramatic depletion at a previous step (i.e. transcript formation) might be masking the same effect on viral assembly during the second round of the transient transfection experiment.

The other cis-acting mutations that appeared to affect transcript abundance (at positions 578, 727, 728, 730, 758, and 791) manifested moderately to severely decreased transcript levels in producer cells under all conditions tested. These mutations may affect the performance of cis-acting transcriptional enhancer elements.

### **Mutations in cis-acting sequences that affect viral assembly**

A cis-acting RNA packaging signal has been previously mapped to the few hundred base pairs around the 5' splice donor site and the 5' end of gag. Here, we have observed that mutations in the interval between positions 739 and 846 were depleted between transcription and release of cell-free virus in all (complemented and uncomplemented) experiments (figure 12). This region encompasses the "kissing loop" dimerization and packaging signal, the 5' splice donor site, and two stem-loop structures which have been found to bind in vitro to gag and nucleocapsid proteins (Berkowitz and Goff 1994; Berkowitz et al. 1993; Clever et al. 1995; Sakaguchi et al. 1993).

The existence of a supplementary packaging signal is implied by a report by Vicenzi et al. 1994, where a deletion of the 5' one-third of U5 results in a 10-

fold decrease in RNA packaging. In our turn, we have found additional mutations in U5 (at positions 564, 583, 607, 621, and 640) that appear to be defective in viral RNA packaging (figure 12).

In general, the phenotypes of these packaging mutants were more severe in the absence of complementation (figure 10 and data not shown). If viral genomes with insertions at these positions are still able to form dimers, dimerization with wild-type viral genomes may partially rescue the packaging defect of these mutant genomes.

### **Cis-acting mutants defective in late replication events**

Mutations at positions 607-654 and 758-791 resulted in a reduction in recovery during the early part of the viral life cycle, which includes viral entry, uncoating, reverse transcription, nuclear entry, and integration (figure 12). The mutations between positions 607-654 are located in U5, just 5' to the att site. Although no specific function for this region of U5 has been previously defined, its proximity to the att site raises the possibility that sequences in this region contribute to recognition of the viral genome by integrase. These mutations are also reasonably close to the primer binding site, and may interfere with initiation of reverse transcription (Leis et al. 1993). The second group of mutations, between positions 758-791, is in the "kissing loop" motif and the 5' splice donor sequence. A function for sequences in this area in early replication events has not been previously described.

The paucity of mutants displaying significant and specific defects in early steps of viral replication is probably due to the design of our experimental system. It is likely that elimination of mutants at steps in the viral life cycle occurring earlier in our series of experiments (e.g. transcription or assembly) prevents our recognition of additional defects in entry, reverse transcription, or integration. For example, two mutations (at positions 683 and 684) located in the primer binding site were severely depleted in the virion RNA sample (transient transfection experiment, data not shown), such that it was not possible to distinguish further reductions in the infected cell genomic DNA sample. Due to the sequence preferences of MuA transposase and the introduction of five base-pair duplications during the mutagenesis procedure, our pool of insertion mutants did not include any detectable mutations that destroyed the att site in U5, another feature in this segment of the genome known to be essential for integration.

### **Mutations in matrix**

Sequences at the 5' end of the matrix coding sequence (positions 827-838) appeared to contribute to viral RNA packaging in cis (see above). A few mutations near the 5' end of the matrix gene (positions 876-929) appeared to result in defects in the production of stable transcripts (figure 13). These trans-acting mutations, which could be rescued by complementation, were located in the sequences that encode the C-terminal end of helix 1, a loop between helix 1 and helix 2, and the N-terminal half of helix 2. This region contains many basic residues, and is at the edge of the globular domain of matrix that faces away

from the trimer interfaces. The phenotype of these mutants suggests that matrix might have a role in enhancing transcription or stabilizing the viral RNA genome in the producer cell prior to budding. Supporting this possibility, it has been proposed that matrix has RNA-binding activity (Bukrinskaya et al.1992).

Most of the mutants with insertions from positions 937-1131 demonstrated primary losses in fitness in the portion of the life cycle from translation through assembly to budding (figures 13 and 15). These mutations could be rescued in trans and were in the portion of the matrix gene encoding the core of the globular domain of matrix. Mutations in this region are likely to interfere with the proper folding of the matrix protein and thus produce defects in viral assembly, as seen in our results.

As reported previously (Freed et al. 1994; Dorfman et al. 1994a), mutations in the C-terminal domain of matrix (positions 1141-1211 in this study), which consists of a long alpha-helical tail that extends away from the globular domain, were well-tolerated (figures13 and 15).

### **Mutations in capsid**

In accordance with other reports (Dorfman et al. 1994b; Mammano et al. 1994), we found that mutations in the sequence encoding the N-terminal half of the capsid protein were severely detrimental to viral replication (figure 10B). Capsid mutants with insertions between positions 1276 and 1464 were defective both at a step in viral production (assembly or release) and at an early step in replication (figures 14 and 15). Preliminary results indicate that the defect in

early replication occurs before the completion of reverse transcription. This result is quite striking, particularly in comparison to the bulk of our matrix mutants, which appeared to be specifically defective at the assembly/budding step (figure 15).

Mutants with insertions in and immediately adjacent to the N-terminal  $\beta$  hairpin (1244-1264) and the cyclophilin A binding region (1479-1508) of capsid were able to form viral particles, but were defective in a step in early replication (figures 14 and 15). X-ray crystallographic studies (Wlodawer and Erickson 1993; Gitti et al. 1996) support the theory that the  $\beta$  hairpin structure forms only after proteolytic maturation of the viral particle. An extended, relatively disordered conformation during assembly may account for the fact that insertions in this region do not cause a drop in viral particle formation. The  $\beta$  hairpin and the cyclophilin A binding regions are the only regions in the N-terminal domain of capsid that protrude from a tightly packed helical core. These structural differences might explain the differential effects of insertions in these regions on assembly. It has been suggested that the disassembly of the viral core (uncoating) that occurs after viral entry and before the initiation of reverse transcription depends on an interaction between capsid and cyclophilin A (Braaten et al. 1996; Gamble et al. 1996). Therefore, one might expect insertions in the cyclophilin A binding region that interfere with this interaction to affect uncoating.



## **Discussion**

### **Expected results and novel observations**

In the course of these experiments, we have identified several features in the HIV genome, some of which have not been previously described. We mapped three types of cis-acting sequences: those that function in transcript formation/stability, those that are involved in viral RNA packaging, and those that are important for an early step in viral replication. Some of these sequences were found in areas previously mapped for these functions (e.g. TAR, the "kissing loop" motif) and others were found in novel locations. Several mutations near the N-terminus of matrix suggest an unforeseen trans-acting function for matrix in transcript formation or stabilization. In contrast to previous reports, we have observed that many mutations in the globular core of matrix have marked effects on assembly, and mutations in the helical core of the N-terminal domain of capsid cause defects in both assembly and an early step (perhaps disassembly) in viral replication (see below). Finally, mutations in the  $\beta$  hairpin and cyclophilin A binding regions of capsid primarily result in early replication defects. The behavior of the mutations in the cyclophilin A binding region are consistent with the postulated function of this region in uncoating.

How can the same mutation in capsid cause defects in both assembly and disassembly? The answer to this question may lie in the fact that assembly and disassembly are not simply reverse processes. Most obviously, assembly involves the aggregation of gag and gag-pol polyproteins whereas disassembly normally occurs after proteolysis of these polyproteins into several smaller

entities. A mutation that decreases the efficiency of assembly may cause the viral proticles that do form to be aberrant in some way. This notion is supported by observations of abnormal viral core morphology in viruses with mutations in the N-terminal half of capsid (Dorfman et al. 1994b; Reicin et al. 1996). These particles may have problems that interfere with steps that are prerequisites to uncoating. For example, perhaps a decreased ratio of gag-pol to gag compromises proteolytic maturation. Alternatively, essential host factors such as cyclophilin A might be inefficiently incorporated.

### **Sources of variability in the data**

Approximately one kilobase of the HIV genome was analyzed using ten primer pairs. Each primer pair was used to examine an interval of 200 to 300 base-pairs. Each mutant was examined with at least two primer pairs. Moreover, each series of transfection/infection experiments was carried out in triplicate. It was therefore necessary to develop a normalization procedure (see Materials and Methods) so that data from separate gels and different replicates could be combined in determining the quantitative effect of each mutation. Data were normalized based on previous findings that certain areas of the viral genome, such as the C-terminus of matrix, are consistently tolerant to small, in-frame insertions.

The normalized data was examined to determine whether the variability in the data arose primarily from variability in the transfection/infection experiments (which could result from sampling error) or variability in the genetic footprinting

procedure. Data for thirty mutants from the stable transfection experiment cellular RNA sample were tabulated and the weighted average of the variances were calculated for the complete data set, data "within replicates," and data "within gels." The normalized intensity measurements for these thirty samples ranged between 7.1 and 111.6. The abundance of each mutant was measured four or five times per replicate. Data within a replicate were derived from separate genetic footprinting reactions using different primer pairs performed on the same nucleic acid sample and run on separate gels. Data within a gel were derived from separate genetic footprinting reactions using the same primer pairs performed on different nucleic acid samples and run on the same gel. The weighted average of the variances was 63.8 for the complete data set, 65.9 "within replicates", and 16.7 "within gels". Therefore, most of the variability appears to arise from differences between gels or primers rather than sampling error incurred during the selection procedure, variability between PCR reactions, or inconsistencies in other nucleic acid manipulations.

### **Mutations that appear to confer an increase in replication-competence**

Mutations at a few positions appear to result in proviruses with an enhanced ability to carry out certain step in viral replication. This finding is somewhat unexpected, as one might expect the wild-type virus to be optimized for replication. However, the system used for the experiments described here is significantly different from the environment in which wild-type HIV evolved. Viral production and infection was carried out in a tissue culture system, rather than in

the context of a whole organism. In this context, the virus does not need to contend with the same complexity of virus-host interactions, such as evasion of the host immune defenses. The sequences encoding env and the accessory factors vif, vpr, vpu, and nef were removed from the proviral clone used, and VSV-G protein was used to pseudotype this defective proviral construct. The use of a pseudotyping system removes several constraints on the viral genome, including the preservation of a functional 5' splice donor sequence and retention of the env-interacting function of matrix. In summary, since the same constraints do not apply in the system used here and in the environment in which HIV evolved, mutations detrimental in one case may be beneficial in the other case.

### **Incomplete depletion of mutant proviruses**

For mutations that severely compromise viral replication, the system presented here may overestimate the ability of these mutants to replicate. This error may stem from three sources. First, there is a certain amount of error in the analysis and quantitation procedures used. Second, the insertion sequence used in these experiments includes a 10 base-pair palindrome, which may form a nucleic acid hairpin structure. This type of mutation may be less disruptive to cis-acting sequences that depend on nucleic acid secondary structure than other types of mutations, such as deletions, substitutions, or non-palindromic insertions. Third, in the selection strategy, the uncomplemented infection cycles were carried out using either stably transfected cells or cells that had been infected at low m.o.i as producer cells. As discussed above, it is possible (and

even likely) that some degree of complementation occurred in the stably transfected cells. As for the cells infected at low m.o.i., the measured m.o.i was 0.05. If the infection followed a Poisson distribution, approximately 2.5% of the cells that were infected by one virus were actually infected by more than one virus, allowing complementation to occur in those cells. Hence, for trans-acting factors, one would expect a background reading of approximately 2.5% of wild-type for recessive mutations.

### **Observed discrepancies with previously published results**

The inconsistencies between our results and those found in other reports can be grouped into two classes. First, mutations at certain positions in the matrix gene resulted in severe defects in replication in our study, while it has been reported elsewhere that mutations at the same positions were tolerated (Freed et al. 1994). Second, we found that many mutations in the N-terminal half of capsid were defective in viral assembly. In contrast, others have reported that residues important for gag multimerization and viral assembly reside in the C-terminal domain of capsid (Jowett et al. 1992; Dorfman et al. 1994b; Von Poblitzki et al. 1993; Reicin et al. 1995), while viruses with mutations in the N-terminal domain of capsid were competent for viral assembly, although many formed viral particles with abnormal core morphologies (Dorfman et al. 1994b; Wang and Barklis 1993; Franke et al. 1994; Reicin et al. 1995, Reicin et al. 1996). The discrepancies between our results and those found in other reports may result from differences in experimental method or interpretation.

First, the precise locations and types of mutations differ between all the reports. Different point mutations at the same position in a given gene can lead to different phenotypes. The types of mutations employed vary widely between (and even within) reports, and include point mutations, small deletions, large deletions, and insertions in various combinations.

Second, the methods used to assess replication-competence differ between reports. Wang and Barklis (1993) performed single-round infectivity assays by measuring infection of a marker gene. Other groups followed exogenous RT activities or production of viral proteins in spreading infections over the course of several weeks (Freed et al. 1994 and Reicin et al. 1995; Dorfman et al. 1994a; Dorfman et al. 1994b). We looked at data from two types of experiments: one single-round infection without complementation and two single-round infections in series, the first of which was complemented and the second of which was not complemented. This approach stands in contrast to experiments with spreading infections, where it is difficult to know how many rounds of infection have occurred, which in turn makes it difficult to measure infectivity quantitatively.

Third, viral assembly has been measured in a variety of ways, including exogenous RT assays, RNase protection, western blotting for viral proteins, and electron micrography. These methods do not always assess whether the viral particles contain viral RNA; some are qualitative or yield highly variable results. We believe that none of these methods is as rigorous as the method employed in

this report, where we assessed the relative representation of mutants in the viral RNA sample itself.

Fourth, in all of the other reports the viral particles studied were generated by transient transfection, while the viral RNA samples we footprinted were purified from virions produced from either stably transfected or cells infected at low m.o.i. In our experiments, the titer of virus produced by transient transfection was 10- to 100-fold higher than the titer of virus produced from stably transfected or infected cells. If this difference in titer reflected a difference in expression of the viral genome, the requirements for viral assembly and packaging of the viral RNA genome in our experiments were 10- to 100-fold more stringent than in the experiments described in the other reports. The mutants that have quantitative defects in assembly or packaging might appear to be competent for these functions by less stringent methods.

Finally, in the strategy presented here, a large number of mutants with insertions at diverse positions were followed en masse through two rounds of replication. This strategy permits a comprehensive examination of viral replication. Selection and analysis of the mutants in parallel provided built-in internal controls for variables such as sample recovery and efficiency of analysis procedures.

### **Future directions**

Our examination of three nucleic acid samples per round of replication yielded a relatively crude breakdown of the HIV life cycle. Refinement of our

picture of viral replication can be achieved by footprinting samples from more finely differentiated steps. For example, we could study the effects of mutations on nuclear export of viral RNA by comparing nuclear and cytoplasmic RNA samples from producer cells. Other interesting steps in the viral life cycle amenable to clarification by genetic footprinting are reverse transcription and nuclear entry. We could investigate these steps by collecting additional nucleic acid samples, such as intermediates in the reverse transcription reaction (minus-strand strong stop DNA and plus-strand strong stop DNA), full-length unintegrated viral DNA in the host cell cytoplasmic fraction, and full-length unintegrated viral DNA in host cell nuclear fraction. Of course, genetic footprinting can be performed on the rest of the HIV genome. In addition, we have developed methods to introduce and analyze a variety of mutations, including insertions of different lengths and sequences and substitutions of various types (Singh et al. 1997 and unpublished results).

### **Generalizability of the genetic footprinting technique**

In the original report describing the genetic footprinting technique, this method was used to generate a high-resolution functional map of a small (200 base-pair) gene encoding an RNA molecule (Singh et al. 1997). A functional selection was carried out in a prokaryotic system, and the footprinted nucleic acid samples consisted of purified plasmids. Here we present modifications to genetic footprinting that permitted us to analyze a much larger (1000 base-pair) stretch of nucleic acid including both cis- and trans-acting sequences. The



experiments presented here involved the isolation and analysis of complex nucleic acid samples, including cellular RNA, virion RNA, and genomic DNA samples from a selection scheme in eukaryotic cells. Thus, genetic footprinting can be used to map the functional features in any DNA sequence if an appropriate selection scheme exists. In such a scheme, the abundance of the sequence encoding a given mutant in a nucleic acid sample collected after selection varies directly with the ability of that mutant to survive the selection. In addition, we have developed methods to analyze genetic footprinting data in a quantitative manner. These tools not only reduce the labor involved in analysis of such data, but also allow a more objective assessment of the data.

# Chapter 5

## The Future of Genetic Footprinting

It is evident that the genetic footprinting method as it exists offers many advantages over traditional methods of mutagenesis and analysis of mutants. Genetic footprinting enables an investigator to perform the mutagenesis, functional selection, and analysis steps en masse, collecting quantitative data on hundreds of mutants at once. There are four major limitations to the present genetic footprinting technology.

First, the distribution of measurable mutants in a gene is largely limited by the sequence bias displayed by the enzyme utilized for mutagenesis. The current favorite enzyme, MuA transposase performs the desired concerted integration event robustly, but displays a sequence selectivity for integration that spans at least three orders of magnitude. The current analysis method covers two orders of magnitude and data can be obtained for only one out of six base-pair positions on average.

Second, the existing repertoire of enzymatic functions limits the design of mutations. The genesis of any mutant library must begin with the construction of a library of insertion mutants, where the palindromic insertions contain a five base-pair duplication in the target sequence and the recognition sequence for a restriction enzyme. This sequence must not occur anywhere else in the vector used for mutagenesis, and must be tolerated by MuA transposase. The range of mutants has been expanded by introducing a linker containing a type IIs restriction enzyme recognition sequence at each insertion. Type IIs restriction

enzymes cleave some number of nucleotides away from their recognition sites, the number being specific to the enzyme. The linker can be designed such that digestion with the type II restriction enzyme either precisely excises the insertion or creates a deletion. Finally, a new linker of desired sequence is inserted into the gap. The most significant drawback to this approach is the lack of type II restriction enzymes that cut more than 16/14 nucleotides away from their recognition sequences.

Third, there are several restrictions imposed by the current analysis method. Two strategies for analysis have been employed, both of which rely on the polymerase chain reaction. The initial strategy used, the "direct PCR" approach, involved using one fixed primer complementary to a sequence in the target gene outside the region under inspection and one "mobile" primer complementary to the insertion sequence. The lengths of the products of this type of PCR reaction correspond to the positions of the inserts relative to the position of the fixed sequence. However, if one wishes to examine the effects of a short insertion or replacement, the direct PCR method proves to be unsatisfactory. A fifteen base-pair insertion contains a unique sequence of only ten base-pairs, too short for sufficiently specific priming. The solution to this problem has been to use the "flanking PCR/restriction digestion" technique. Here, a PCR reaction is performed using two fixed primers complementary to sequences in the target gene flanking the region of interest. The products of this reaction are digested with a restriction enzyme that recognizes a site in the insertion sequence. The obvious limitation to this method is that the insertion

must be palindromic and contain a restriction site (if the sequence is not palindromic, each position of insertion will yield two products of different size, depending on the orientation of the insertion in the gene).

Finally, the gel-based detection method is cumbersome and introduces a significant amount of error into our results. In fact, using the current system, the analysis procedure appears to be responsible for much more variation than the selection scheme.

I believe there is a technically feasible alternate approach that eliminates the problems enumerated above. The investigator would be able to specify the positions and relative abundances of mutants, assuring more uniform coverage of the gene of interest. A much wider variety of mutations would be available, including insertions or replacements of as few as three base-pairs, with no limitations on the content of the introduced sequence. Even deletions can be examined, as long as one is willing to introduce a few unique base-pairs at the deletion site (see figure 16 for sketches of possible types of mutations). The analysis method would involve a flanking PCR step followed by hybridization of the PCR products to an array of oligonucleotides and scanning using a fluorescence detection system. A specific description of this approach follows.

First, one must make a library of mutants (see figure 17). This step involves the synthesis of two unique oligonucleotides for each mutant desired, in addition to two common oligonucleotides complementary to sequences flanking the region under mutagenesis. For example, in order to replace all the residues

in a 300 amino acid protein with alanine, one would need  $600 + 2$  oligonucleotides. These mutagenic oligonucleotides are also used in the analysis process. The two unique oligonucleotides for a given mutant are complementary to each other, and contain two types of sequences. At the edges, the oligonucleotides are complementary to target gene sequences on either side of the site of mutagenesis. The middle of each oligonucleotide contains the insertion or replacement sequence. The lengths of the "edge" sequences are adjusted such that the oligonucleotides for all of the mutants have approximately the same melting temperature. Now all the mutagenic oligonucleotides complementary to the top strand of the target gene are mixed together in one pot and all the mutagenic oligonucleotides complementary to the bottom strand are mixed together in another pot, adjusting the ratios of the individual oligonucleotides according to the desired proportions of each mutant in our starting library. For instance, in order to start with twice as many mutants at position A as at position B, one would add twice as many oligonucleotides for position A as for position B. Then, two PCR reactions are performed. The template for both reactions is the gene to be mutagenized. One reaction contains the fixed oligonucleotide complementary to the bottom strand of the template and the mixture of mutagenic oligonucleotides complementary to the top strand of the template, and the other reaction contains the other set of oligonucleotide primers. In order to minimize the introduction of unwanted mutations due to misincorporation by the enzyme used for PCR, a proofreading polymerase is used and the number of cycles of PCR is minimized. Suppose

that one starts with approximately 12.5 pmol of each primer and 0.015 pmol of template. After five cycles of PCR with an annealing temperature corresponding to the predicted melting temperature for the "edge" sequences of the mutagenic oligonucleotides, about one pmol of mutants should be present. Then, ten cycles of PCR with an annealing temperature corresponding to the predicted melting temperature for the complete mutagenic oligonucleotides are performed. After a purification step to eliminate any unincorporated primers, the products of the two PCR reactions are mixed together. Another PCR reaction including only the fixed, flanking oligonucleotides is performed, using an annealing temperature corresponding to the predicted melting temperature for the complete mutagenic oligonucleotides. The products of the initial round of PCR will prime off of each other if they overlap precisely, as they will when they correspond to the same mutant. Then, the flanking primers will amplify the population of mutants, which can be cloned into an appropriate vector for selection.

After a functional selection is performed, the relevant nucleic acid sample is purified. Both the original library of mutants and the selected nucleic acid sample are subjected separately to PCR using the flanking oligonucleotides. During this amplification step, fluorescent labels are incorporated, one color for the pre-selection sample and another color for the post-selection sample. The products of these two PCR reactions are then mixed and used as a probe to hybridize to an oligonucleotide array. The elements of this array are the original mutagenic oligonucleotides (one can of course use the fixed oligonucleotides as positive controls and normalization standards). PCR products containing

mutations hybridize to the corresponding mutagenic oligonucleotides. It has been shown that existing array hybridization technology allows discrimination of one mismatch in an oligonucleotide octomer. This level of specificity should be adequate for the purposes of this type of experiment. The arrays are then scanned and quantitated. The ratios of the two colors at each spot on the array reflect the ability of the corresponding mutant to survive the selection.

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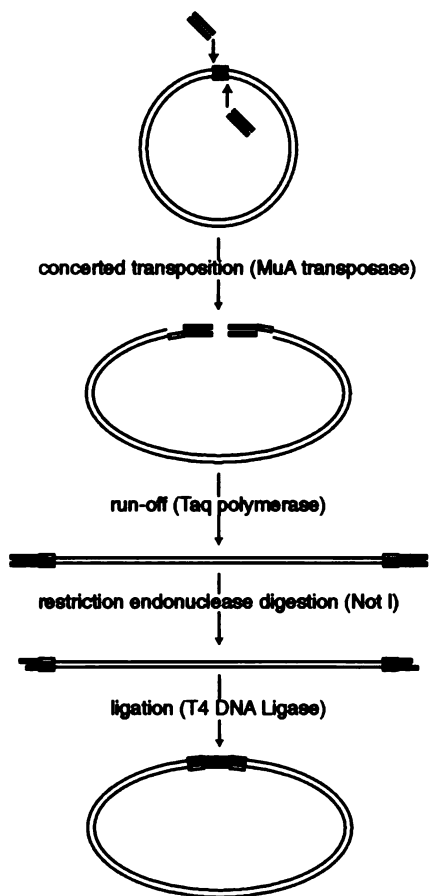


Figure 1. Mutagenesis scheme using MuA transposase. Oligonucleotides used for mutagenesis are bold lines (■), the target DNA plasmid is drawn as thin lines, and sequences in the target DNA duplicated during mutagenesis are empty lines (□).



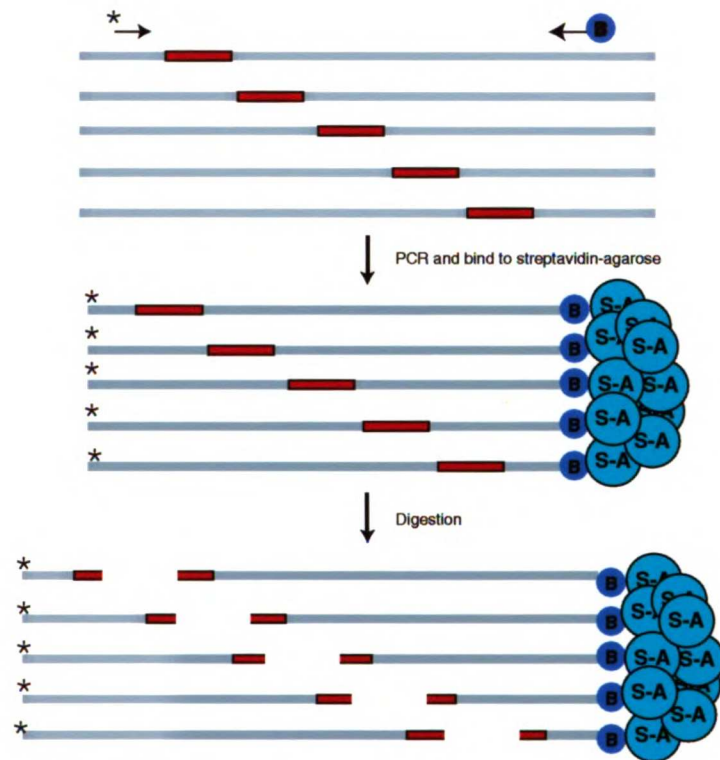


Figure 2. Genetic footprinting scheme using flanking PCR and restriction digestion. A collection of insertion mutants is subjected to PCR using one radioactively labelled primer ( $\ast \rightarrow$ ) and one biotinylated primer ( $\leftarrow \bullet$ ). The PCR products are bound to streptavidin-agarose resin ( $\bullet$ -S-A) and digested with a restriction enzyme that recognizes a site in the insertion sequence. The radioactively labelled ends of the PCR products are released.

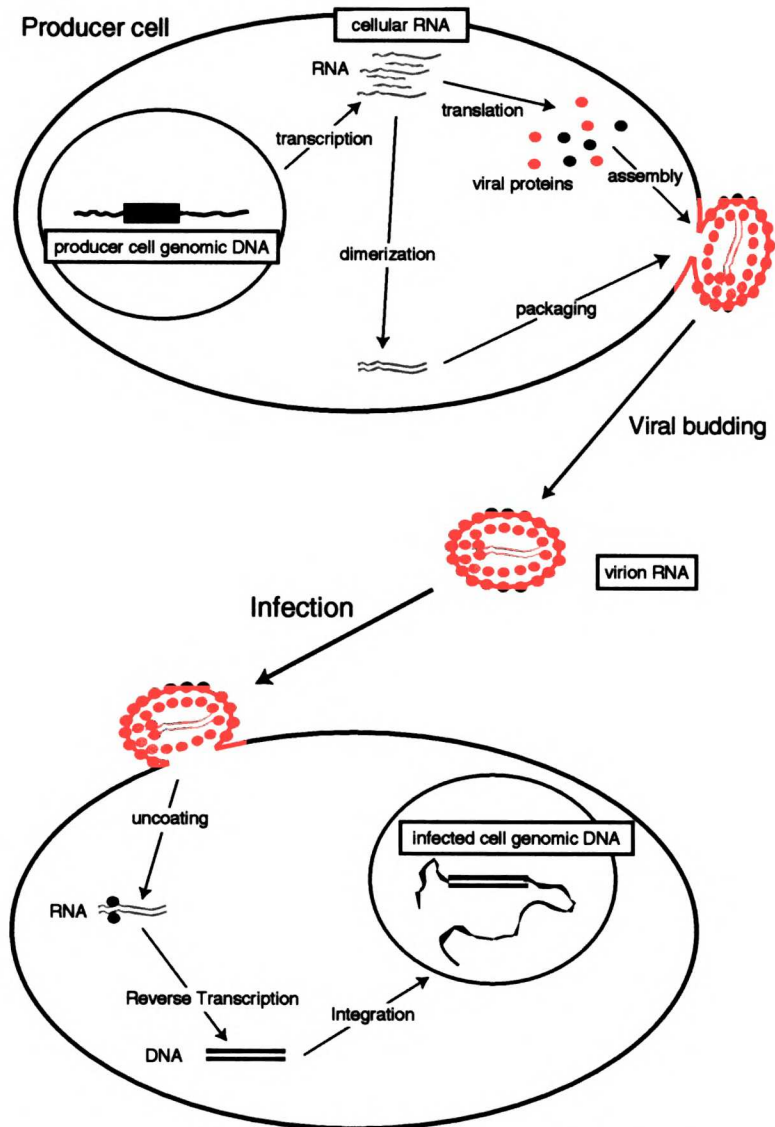


Figure 3. The retroviral life cycle. Nucleic acid samples analyzed in this study are boxed.

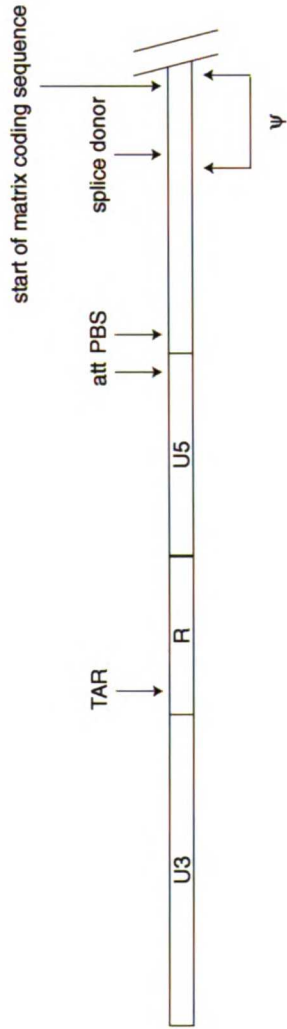


Figure 4A. Schematic representation of the 5' end of the HIV genome, which includes TAR, the U5 att site, the primer binding site (PBS), the splice donor for the env message, the start of the matrix coding sequence, and the packaging sequence ( $\psi$ ).

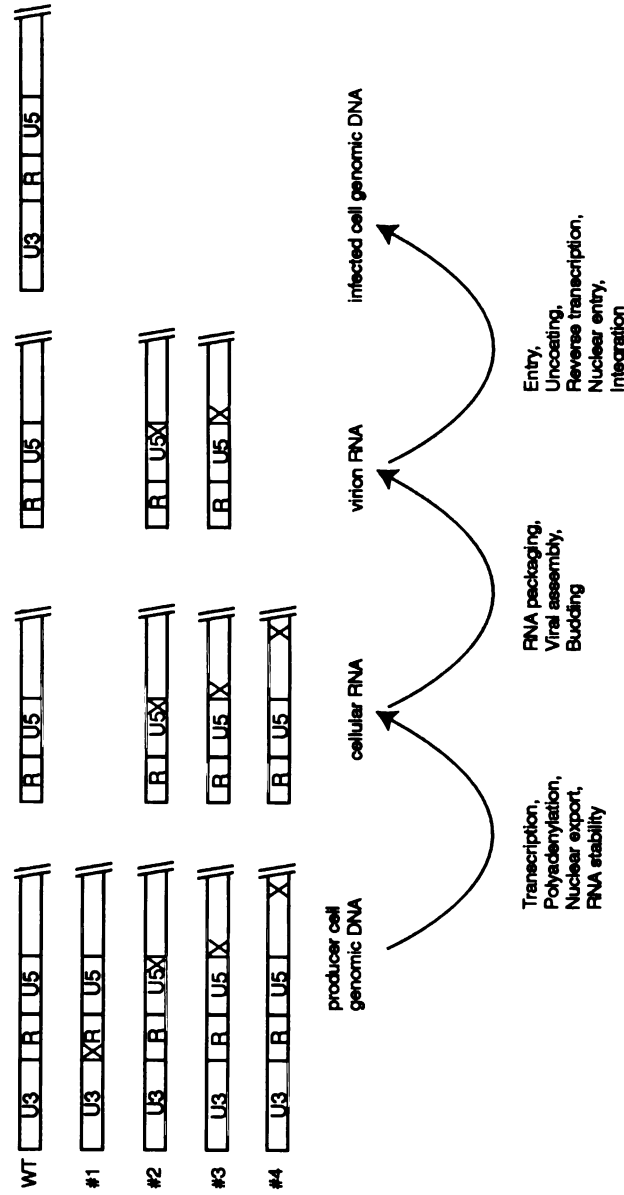


Figure 4B. Selection of mutants defective in TAR (#1), att site(#2), primer binding site (#3), and packaging sequence (#4) functions at different steps during the viral life cycle and the nucleic acid samples where one would observe these selections. The illustrated mutations and the phenotypes depicted for them are based on results from previous reports. "WT" indicates a wild-type, replication-competent viral genome.

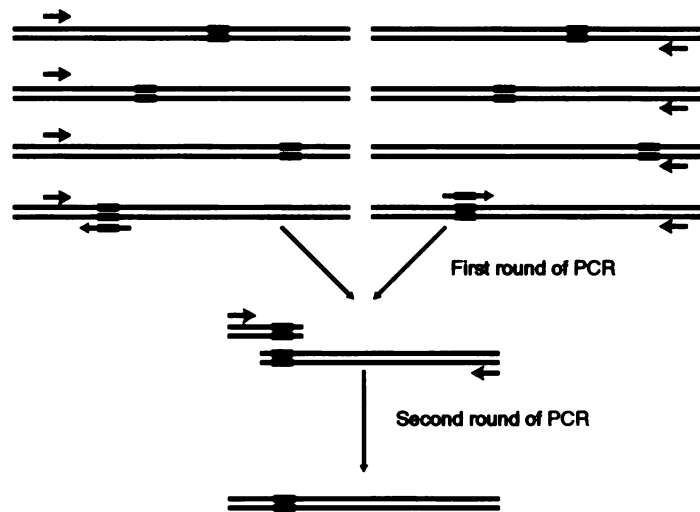


Figure 5. Isolating a specific mutant by PCR. Sequences introduced during the mutagenesis procedure are in bold lines (■). Primers are indicated by arrows. Note that primers that contains both wild-type and mutant sequences (←■→) will selectively prime off of template DNAs that have a mutation at the selected site.

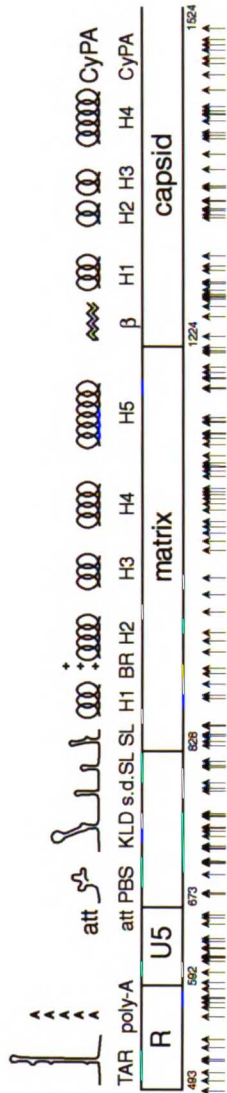


Figure 6. Diagram of mutations evaluated in this study mapped onto the HIV genome. Features described previously are TAR (TAR), the polyadenylation signal (poly-A), the att site (att), the primer binding site (PBS), the kissing loop domain (KLD), the splice donor (s.d.) for the env message, two gag-binding stem-loop structures (SL), helices 1-5 (H1-H5) of matrix, the basic region of matrix (BR), the beta hairpin of capsid ( $\beta$ ), helices 1-4 (H1-H4) of capsid, and the cyclophilin A binding region of capsid (CyPA). Numbers indicate nucleotide positions at the borders of major regions in the HIV genome. Arrows indicate positions of insertional mutations evaluated in this study.

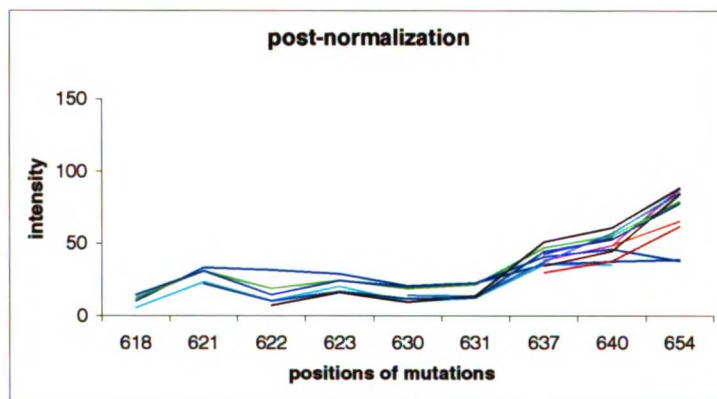
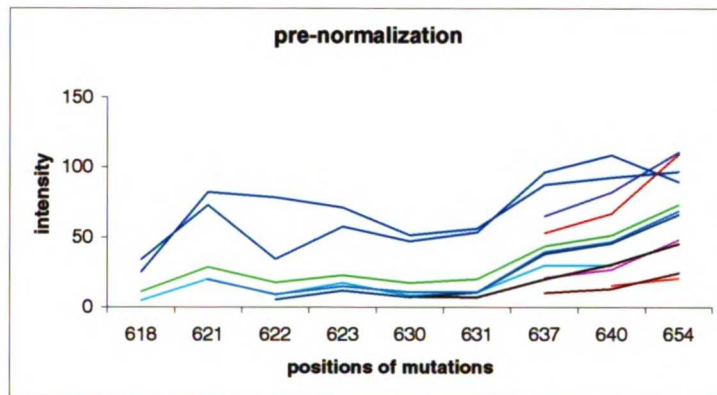


Figure 7. Example of quantitative data before and after normalization. Data shown is from genetic footprinting experiments on the library of mutagenized proviruses. Nucleotide positions of mutations for which data were obtained are given on the X-axis. Intensity of bands measured from autoradiograms is given in arbitrary units on the Y-axis. The different colored traces represent data measured from different gels.

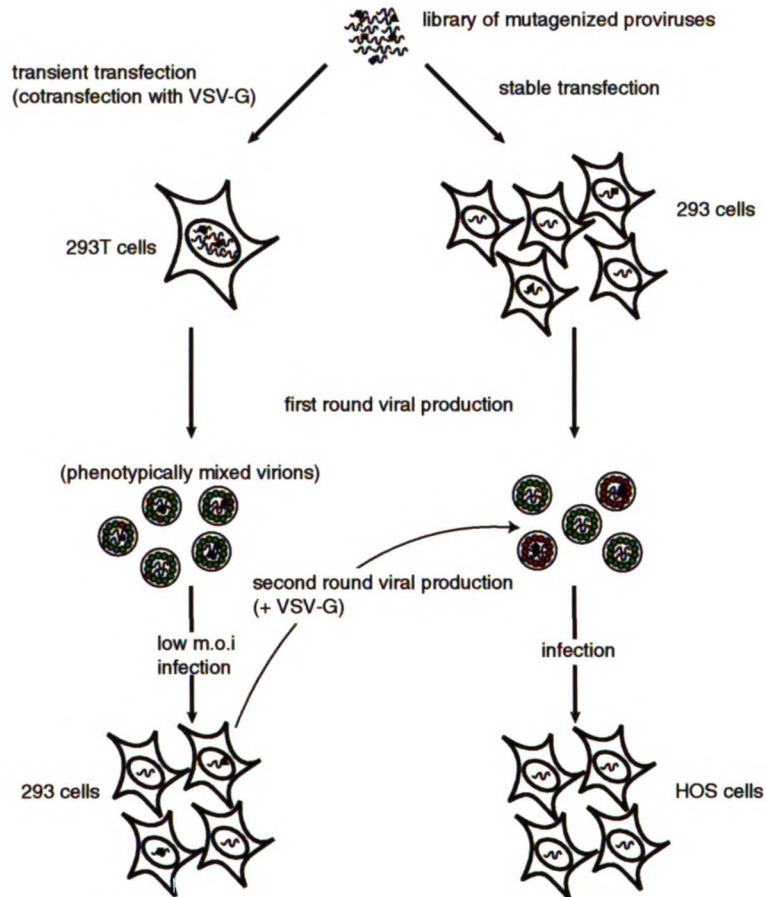


Figure 8. Diagram of transfection and infection experiments. The library of mutagenized proviruses was either transiently or stably transfected into cells to produce populations of mutant virions. In our experiments, the viral genomes of mutants defective in trans-acting factors were efficiently rescued during the transient transfection by phenotypic mixing, but were not detectably rescued during the stable transfection. The virus produced in the transient transfection experiment was used to infect fresh, uninfected cells at a low multiplicity, resulting in a population of producer cells that contained a single provirus per cell. Wild-type viral genome (✓), replication-defective viral genome with mutation in trans-acting factor (✓/), wild-type viral protein (●), mutant viral protein (●/).



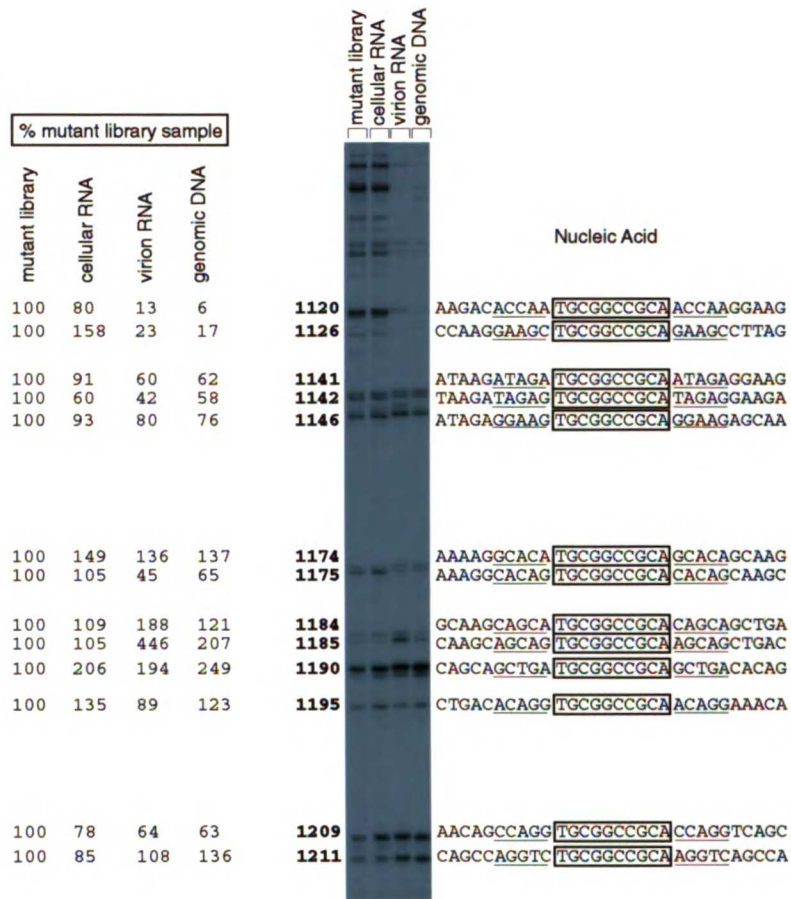


Figure 9. Genetic footprinting of library of mutagenized proviruses and nucleic acid samples from the transient transfection experiment, second round (the uncomplemented round) of viral production and infection (cellular RNA, viral RNA, and infected cell genomic DNA). Numbers directly to the left of the gel indicate exact positions of insertions. The first nucleotide of the HIV provirus is at position 37. Quantitative data averaged from normalized measurements are also given to the left of the gel. The nucleic acid sequences of the mutants are written to right of the gel. The sequences derived from the insertion oligonucleotide are boxed, while the target sequence duplications are underlined.

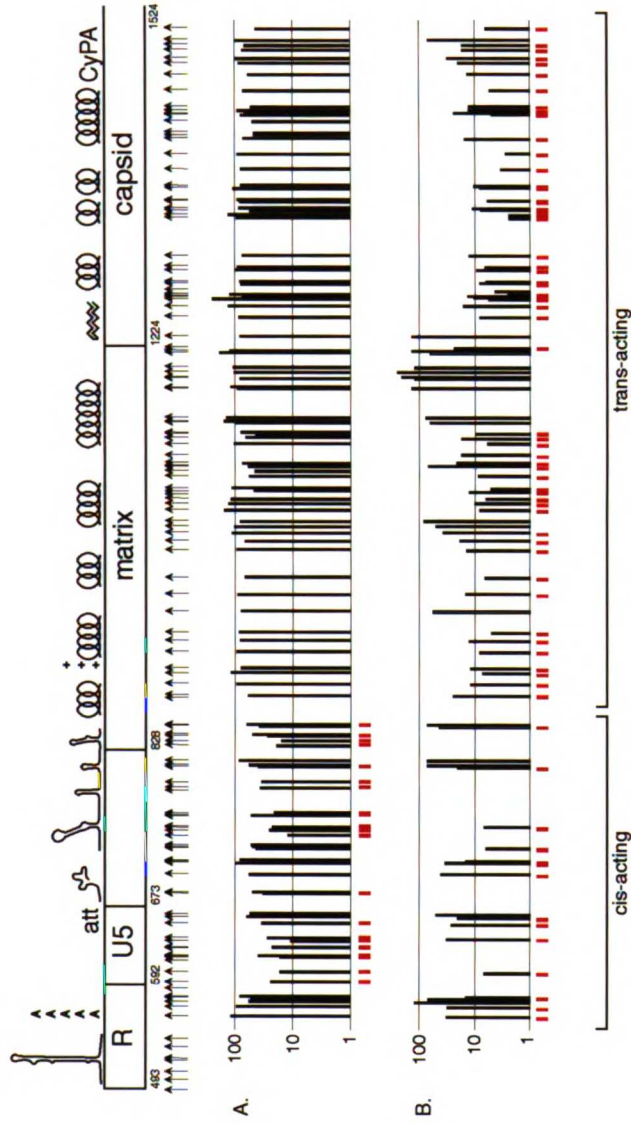


Figure 10. Percent recovery of mutants through single-cycle infections. Data are not shown for mutants where the coefficient of variation between triplicate experiments was greater than 0.5, except in cases where the observed phenotypes were confirmed by re-analysis. Graphs are plotted on a log scale. Red bars indicate mutants that display significant depletions (<45% recovery). Mutations that compromise replication both in the presence and the absence of complementation are considered to be located in cis-acting sequences, while those that affect only uncomplemented replication cycles are considered to be located in trans-acting sequences. A. Percent recovery of mutants after a single round of infection in the presence of complementation. Data are from the transient transfection experiment, first round of infection. B. Percent recovery of mutants after a single round of infection in the absence of complementation. Data are from the transient transfection experiment, second round of infection. Data are not given for mutants whose abundance was very low after the first round of infection.

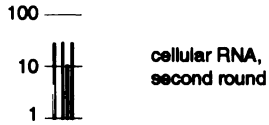
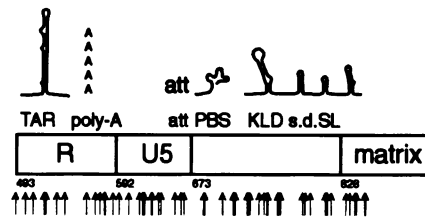


Figure 11. Behavior of selected mutants in cis-acting sequences in the transient transfection experiment. These mutants are replication competent in the first round of infection, but defective for transcript formation in the second round, possibly indicating that the cis-acting element interrupted by the insertions is active in the 3' LTR.

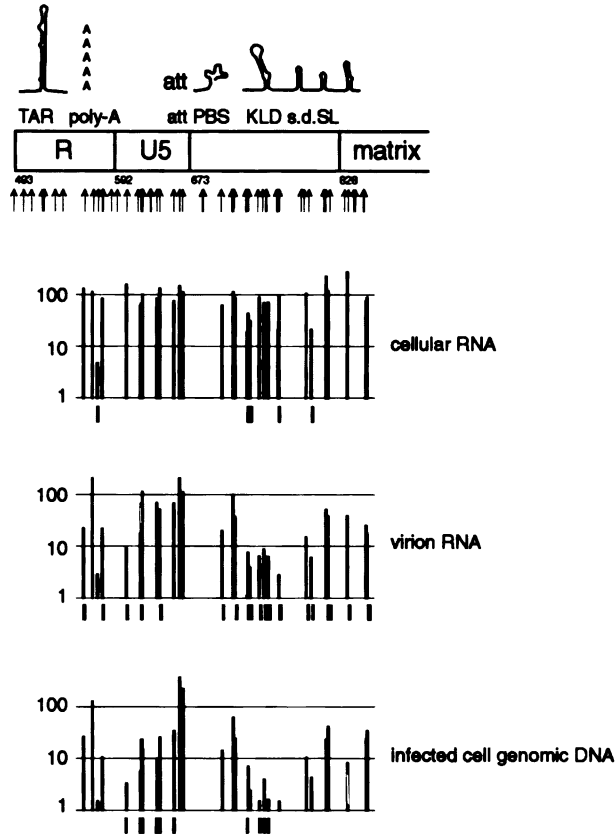


Figure 12. Percent recovery of mutations in cis-acting sequences at several steps of a single-cycle uncomplemented infection. Samples were collected from the stable transfection experiment, second round of viral production and infection. Percent recovery was calculated by dividing the abundance of a mutant in a given nucleic acid sample by the abundance of that mutant in the genomic DNA sample from the stable transfection. Data are not shown for points where the abundance of a particular mutant was very low in the preceding nucleic acid sample, or the coefficient of variation between triplicate experiments was greater than 0.5. Graphs are plotted on a log scale. Red bars indicate mutants that display significant depletions (<50% of preceding nucleic acid sample). Mutants which were depleted in the cellular RNA sample are considered to be defective in transcript formation, mutants which were depleted in the virion RNA sample are considered to be defective in packaging, and mutants which were depleted in the infected cell genomic DNA sample are considered to be defective in an early step in viral replication.

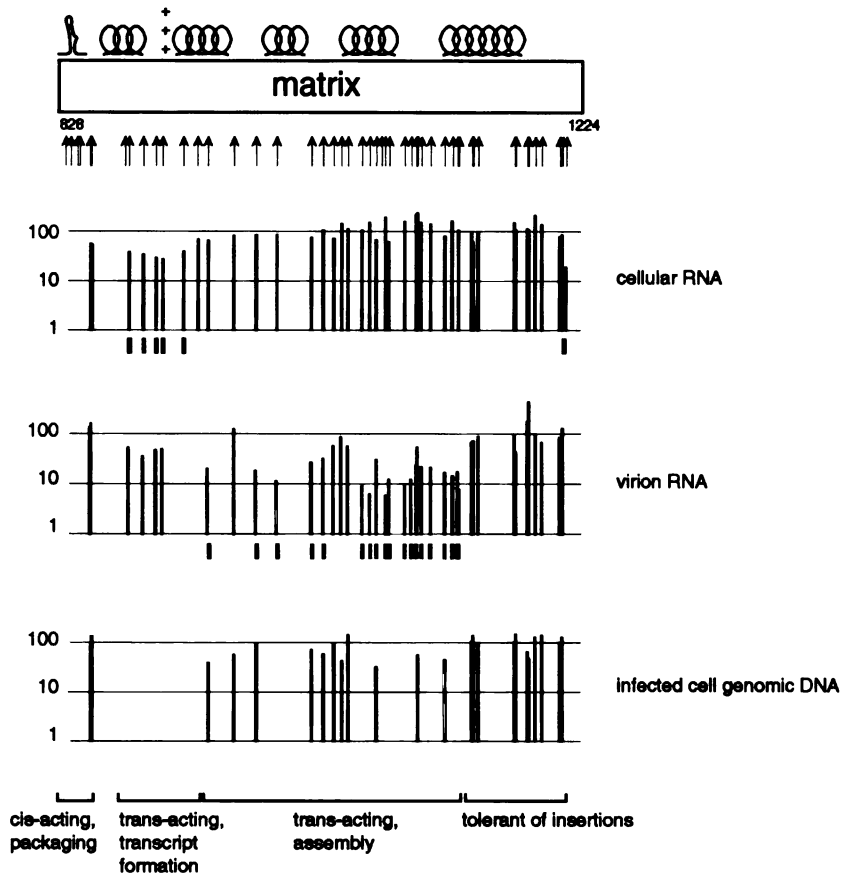


Figure 13. Percent recovery of mutations in matrix at several steps of a single-cycle uncomplemented infection. Samples were collected from the transient transfection experiment, second round of viral production and infection. Percent recovery was calculated by dividing the abundance of a mutant in a given nucleic acid sample by the abundance of that mutant in the infected cell genomic DNA sample from the first round of infection. Data are not shown for points where the abundance of a particular mutant was very low in the preceding nucleic acid sample, or the coefficient of variation between triplicate experiments was greater than 0.5. Graphs are plotted on a log scale. Red bars indicate mutants that display significant depletions (<45% of preceding nucleic acid sample). Mutants which were depleted in the cellular RNA sample are considered to be defective in transcript formation, mutants which were depleted in the virion RNA sample are considered to be defective in assembly, and mutants which were depleted in the infected cell genomic DNA sample are considered to be defective in an early step in viral replication.

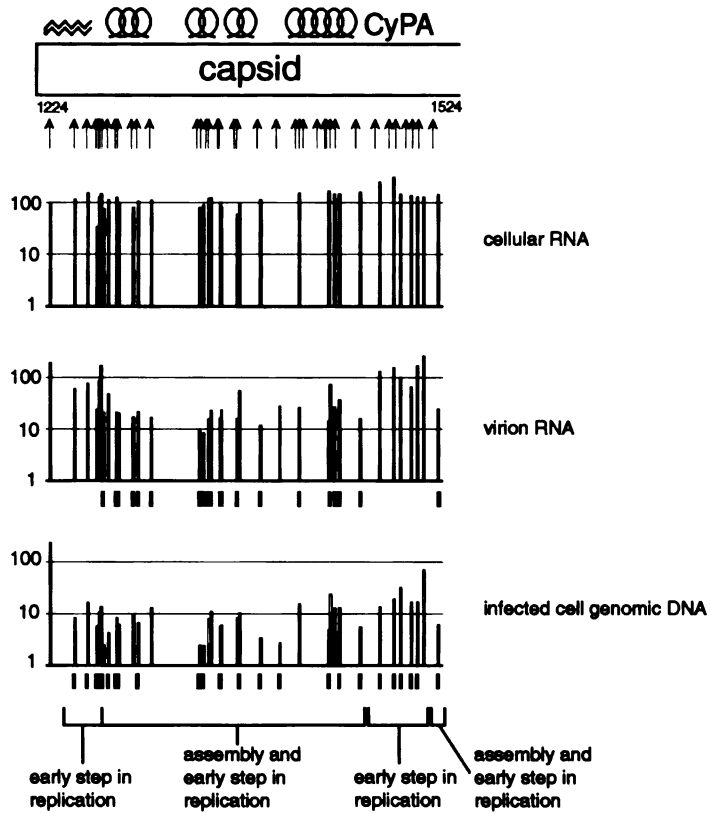
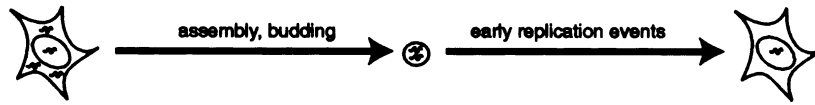
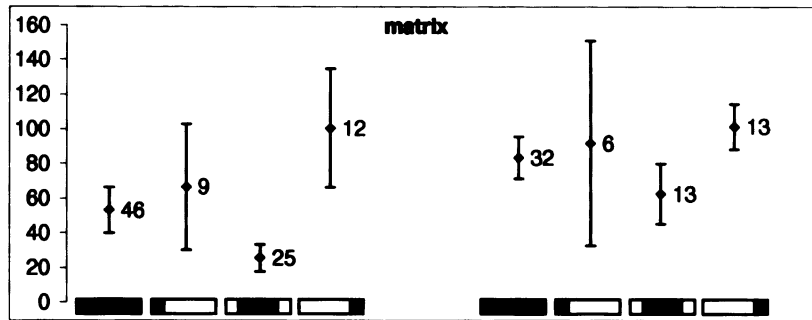


Figure 14. Percent recovery of mutations in capsid at several steps of a single-cycle uncomplemented infection. Samples were collected from the transient transfection experiment, second round of viral production and infection. Percent recovery was calculated by dividing the abundance of a mutant in a given nucleic acid sample by the abundance of that mutant in the infected cell genomic DNA sample from the first round of infection. Data are not shown for points where the abundance of a particular mutant was very low in the preceding nucleic acid sample, or the coefficient of variation between triplicate experiments was greater than 0.5. Graphs are plotted on a log scale. Red bars indicate mutants that display significant depletions (<45% of preceding nucleic acid sample). Mutants which were depleted in the cellular RNA sample are considered to be defective in transcript formation, mutants which were depleted in the virion RNA sample are considered to be defective in assembly, and mutants which were depleted in the infected cell genomic DNA sample are considered to be defective in an early step in viral replication. All mutations in capsid that affected viral replication showed their effects in trans.



A.



B.

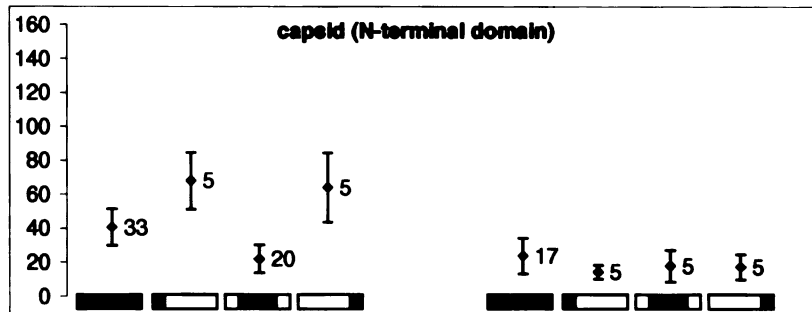


Figure 15. Percent recovery of matrix and capsid mutants through the viral assembly process and the early steps of the viral life cycle. Data are derived from single-round uncomplemented viral production and infection cycles. A schematic of the phases of the life cycle tested is drawn above the graphs. Numbers to the right of each point indicate the number of mutants from which data were averaged. Below each data point is a schematic of the region of matrix or capsid evaluated. Error bars indicate 95% confidence intervals. A. Data for the matrix protein. Data are shown for insertions between amino acid positions 1-132 (complete matrix protein), 1-35 (N-terminal region), 36-102 (central region), and 104-130 (C-terminal region). B. Data for the N-terminal half of the capsid protein. Data are shown for insertions between amino acid positions 1-101 (N-terminal half), 1-15 (N-terminal beta hairpin), 17-81 (central helical region), and 85-96 (Cyclophilin A binding region).

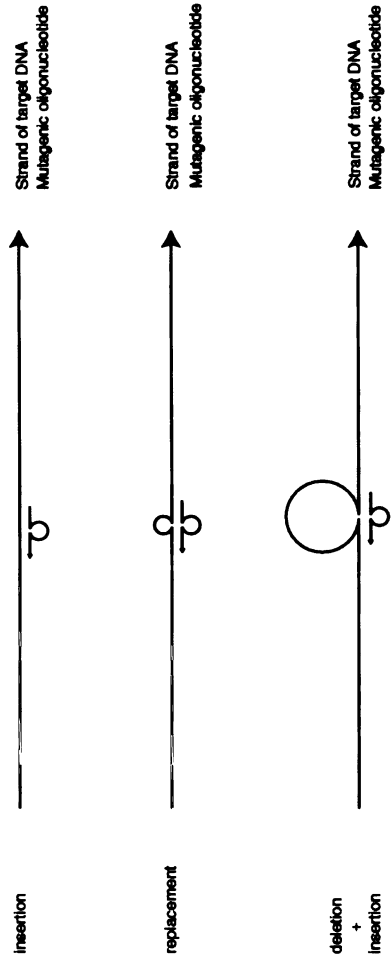


Figure 16. Design of oligonucleotides used to make different types of mutations by PCR.



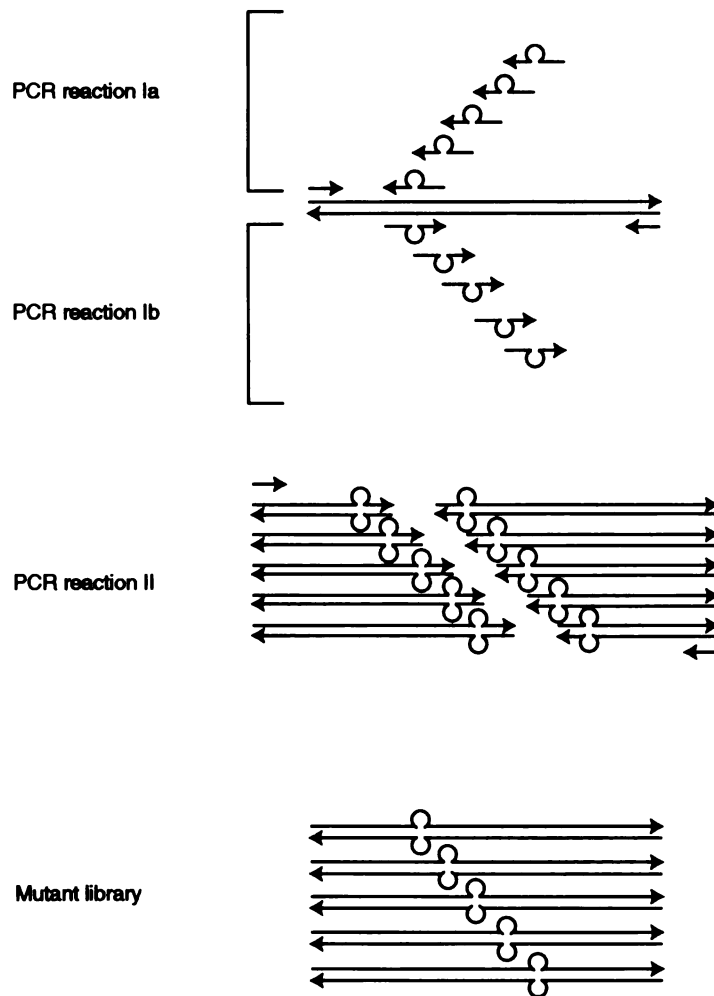


Figure 17. Strategy for achieving saturating mutagenesis of a stretch of DNA using PCR.

**Appendix A.**  
**Complete nucleotide sequence of HIV puro**  
**(the plasmid used for mutagenesis).**

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7501 GCAAGGTGTG GGTCGCGGAC GACGGCGCCG CCGTGGCGGT CTGGACCACG CCGGAGAGCG  
7561 TCGAAGCGGG GGCGGTGTTT GCCGAGATCG GCCCGCGCAT GGCCGAGTTG AGCGGTTCCC  
7621 GGCTGGCCGC GCAGCAACAG ATGGAAGGCC TCCTGGCGCC GCACCGGCC AAGGAGCCCC  
7681 CGTGGTTCCCT GGCCACCGTC GGCGTCTCGC CCGACCACCA GGGCAAGGGT CTGGGCAGCG  
7741 CCGTCGTGCT CCCCAGAGTG GAGGCGGCCG AGCGCGCCGG GGTGCCCGCC TTCCTGGAGA  
7801 CCTCCGCGCC CCGCAACCCT CCCTTCTACG AGCGGCTCGG CTTACCCGTC ACCGCCGACG  
7861 TCGAGGTGCC CGAAGGACCG CGCACCTGGT GCATGACCCG CAAGCCCGGT GCCTGACGCC  
7921 CGCCCCACGA CCCGCAGCGC CCGACCAAA GGAGCGCACG ACCCATCGCT CGAGACCTAG  
7981 AAAACATGG AGCAATCACA AGTAGCAATA CAGCAGCTAA CAATGCTGCT TGTGCCTGGC  
8041 TAGAAGCACA AGAGGAGGAA GAGGTGGGTT TTCCAGTCAC ACCTCAGGTA CTTTAAAGAC  
8101 CAATGACTTA CAAGGCAGCT GTAGATCTTA GCCACTTTTT AAAAGAAAAG GGGGGACTGG  
8161 AAGGGCTAAT TCAC'TCCAA AGAAGACAAG ATATCCTTGA TCTGTGGATC TACCACACAC  
8221 AAGGCTACTT CCTGTATTGG CAGAATACA CACCAGGGCC AGGGGTACAG TATCCACTGA  
8281 CCTTTGGATG GTGCTACAAG CTAGTACCAG TTGAGCCAGA TAAGGTAGAA GATGCCAATA  
8341 AAGGAGAGAA CACCAGCTTG TTACACCCTG TGAGCCTGCA TGGAAATGGAT GACCCTGAGA  
8401 GAGAAGTGTT AGAGTGGAGG TTTGACAGCC GCCTAGCATT TCATCACGTG GCCCGAGAGC  
8461 TGCATCCGGA GTACTTCAAG AACTGCTGAC ATCGAGCTTG CTACAAGGGA CTTTCCGCTG  
8521 GGGACTTTCC AGGGAGGCGT GGCTTGGGCG GGACTGGGGA GTGGCGAGCC CTCAGATGCT  
8581 GCATATAAGC AGCTGCTTTT TGCTGTACT GGGTCTCTCT GGTTAGACCA GATCTGAGCC  
8641 TGGGAGCTCT CTGGCTAACT AGGGAACCCA CTGCTTAAGC CTCAATAAAG CTTGCCTTGA  
8701 GGGAGTGCTT CAAGTAGTGT GTGCCCGTCT GTTGTGACTC TGGTAACTAG AGATCCCTCA  
8761 GACCTTTTTA GTCAGTGTGG AAAATCTCTA GCACCCAGGA GGTAGAGGTT GCAGTGAGCC  
8821 AAGATCGCGC CACTGCATTC CAGCCTGGGC AAGAAAACAA GACTGTTTTAA AATAATAATA  
8881 ATAAGTTAAG GGTATTAAAT ATATTTATAC ATGGAGGTCA TAAAAATATA TATATTTGGG  
8941 CTGGGCGCAG TGGCTCACAC ATGCGCCCGG CCCTTTGGGA GGCCGAGGCA GGTGGATCAC  
9001 CTGAGTTTGG GAGTTCCAGA CCAGCCTGAC CAACATGGAG AAACCCCTTC TCTGTGTATT  
9061 TTTAGTAGAT TTTATTTTAT GTGTATTTTA TTCACAGGTA TTTCTGGAAA ACTGAAACTG  
9121 TTTTCTTCT ACTCTGATAC CACAAGAATC ATCAGCACAG AGGAAGACTT CTGTGATCAA  
9181 ATGTGGTGGG AGAGGGAGGT TTTACCAGC ACATGAGCAG TCAGTTCTGC CGCAGACTCG  
9241 GCGGGTGTCC TTCGGTTCAG TTCCAACACC GCCTGCCTGG AGAGAGGTCA GACCACAGGG  
9301 TGAGGGCTCA GTCCCCAAGA CATAAACACC CAAGACATAA ACACCCAACA GGTCCACCCC  
9361 GCCTGCTGAG CAGGCAGAGC CGATTCACCA AGACGGGAAT TAGGATAGAG AAAGAGTAAG  
9421 TCACACAGAG CCGGCTTTCC CCGTCAAGCT CTAATCGGG GGCTCCCTTT AGGGTCCGA  
9481 TTTAGTGCCT TACGGCACCT CGACCCAAA AAACCTGATT AGGGTGATGG TTCACGTAGT  
9541 GGGCCATCGC CCTGATAGAC GGTTTTTCGC CCTTTGACGT TGGAGTCCAC GTTCTTTAAT  
9601 AGTGGACTCT GTTCCAAC TGGAACAACA CTCAACCCTA TCTCGGTCTA TTCTTTTGAT  
9661 TTATAAGGGA TTTTGCCGAT TTCGGCCTAT TGGTTAAAA ATGAGCTGAT TTAACAAAAA  
9721 TTTAACGCGA ATTTTAACAA AATATTAACG TTTACAATTT CAGGTGGCAC TTTTCGGGGA  
9781 AATGTGCGCG GAACCCCTAT TTGTTTTATT TTCTAAATAC ATTCAAATAT GTATCCGCTC

9841 ATGAGACAAT AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG TATGAGTATT  
 9901 CAACATTTCC GTGTCGCCCT TATTCCTTTT TTTGCGGCAT TTTGCCTTCC TGTTTTTGCT  
 9961 CACCCAGAAA CGCTGGTGAA AGTAAAAGAT GCTGAAGATC AGTTGGGTGC ACGAGTGGGT  
 10021 TACATCGAAC TGATCTCAA CAGCGGTAAG ATCCTTGAGA GTTTTCGCCC CGAAGAACGT  
 10081 TTTCCAATGA TGAGCACTTT TAAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTATTGAC  
 10141 GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT GGTTGAGTAC  
 10201 TCACCAGTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG TAAGAGAATT ATGCAGTGCT  
 10261 GCCATAAGCA TGAGTGATAA CACTGCGGCC AACTTACTTC TGACAACGAT CGGAGGACCG  
 10321 AAGGAGCTAA CCGCTTTTTT TCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG  
 10381 GAACCGGAGC TGAATGAAGC CATAACAAAC GACGAGCGTG ACACCACGAT GCCTGTAGCA  
 10441 ATGGCAACAA CGTTGCGCAA ACTATTAAC GCGCAACTAC TTACTCTAGC TTCCCAGCAA  
 10501 CAATTAATAG GGTGATGGA GCGGATAAAA GTTGCAGGAC CACTTCTGCG CTCGGCCCTT  
 10561 CCGCTGGCT GGTTTATGTC TGATAAATCT GGAGCCGGTG AGCGTGGGTC TCGCGGTATC  
 10621 ATTGCAGCAC TGGGGCCAGA TGGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGC  
 10681 AGTCAGGCAA CTATGGATGA ACGAAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT  
 10741 AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA TTTAAAACCT  
 10801 CATTTTTAAT TAAAAAGGAT CTAGGTGAAG ATCCTTTTTG ATAATCTCAT GACCAAAATC  
 10861 CCTTAACGTG AGTTTTCGTT CCACTGAGCG TCAGACCCCG TAGAAAAGAT CAAAGGATCT  
 10921 TCTTGAGATC CTTTTTTTCT GCGCGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA  
 10981 CCAGCGGTGG TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTTTCCGAA GGTAACTGGC  
 11041 TTCAGCAGAG CGCAGATACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT AGGCCACCAC  
 11101 TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC TAATCCTGTT ACCAGTGGCT  
 11161 GCTGCCAGTG GCGATAAGTC GTGTCTTACC GGGTTGGACT CAAGACGATA GTTACCGGAT  
 11221 AAGGCGCAGC GGTCCGGGCTG AACGGGGGGT TCGTGACAC AGCCCAGCTT GGAGCGAACG  
 11281 ACCTACACCG AACTGAGATA CCTACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCAGAA  
 11341 GGGAGAAAAG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA GCGCACGAGG  
 11401 GAGCTTCCAG GGGGGAACGC CTGGTATCTT TATAGTCCTG TCGGGTTTCG CCACCTCTGA  
 11461 CTTGAGCGTC GATTTTTGTG ATGCTCGTCA GGGGGGCCGA GCCTATGGAA AAACGCCAGC  
 11521 AACGCGCCT TTTTACGGTT CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GT

## Appendix B. Local nucleotide and peptide sequences of mutants

		left flanking nucleotide sequence	insertion	right flanking nucleotide sequence	peptide sequence
1	A	A	TGCGGCCGCA	ACATGTAGCCCCAGTT	AAATCSPS
2	C	AC	TGCGGCCGCA	ACATGTAGCCCCAGTTC	CGRNM*POF
3	A	ACA	TGCGGCCGCA	ACATGTAGCCCCAGTTCT	MRPQHVAPV
4	T	ACAT	TGCGGCCGCA	ACATGTAGCCCCAGTTCTA	IAAATCSPSS
5	G	ACATG	TGCGGCCGCA	ACATGTAGCCCCAGTTCTAC	MCGRNM*POFY
6	T	ACATGT	TGCGGCCGCA	CATGTAGCCCCAGTTCTACT	MLRPHM*POFY
7	A	ACATGTGA	TGCGGCCGCA	ATGTAGCCCCAGTTCTACTT	MYAAAM*POFY
8	G	ACATGTAG	TGCGGCCGCA	TGTAGCCCCAGTTCTACTTA	M*CGRM*POFY
9	C	ACATGTAGC	TGCGGCCGCA	GTAGCCCCAGTTCTACTTAC	M*LRPQ*POFY
10	C	ACATGTAGCC	TGCGGCCGCA	TAGCCCCAGTTCTACTTACA	M*PAAA*POFY
11	C	ACATGTAGCCC	TGCGGCCGCA	AGCCCCAGTTCTACTTACAC	M*PCGRKPQFYLH
12	C	ACATGTAGCCCC	TGCGGCCGCA	GCCCCAGTTCTACTTACACC	M*PLRPQPOFYLH
13	A	ACATGTAGCCCCA	TGCGGCCGCA	CCCCAGTTCTACTTACACCA	M*PHAAAPQFYLH
14	G	ACATGTAGCCCCAG	TGCGGCCGCA	CCCAGTTCTACTTACACCAA	M*PQCGRTOFYLHQ
15	T	ACATGTAGCCCCAGT	TGCGGCCGCA	CCAGTTCTACTTACACCAAG	M*PQLRPHQFYLHQ
16	T	ACATGTAGCCCCAGTT	TGCGGCCGCA	CAGTTCTACTTACACCAAGA	M*PQFAAAGFYLHQ
17	C	ACATGTAGCCCCAGTTC	TGCGGCCGCA	AGTTCTACTTACACCAAGAA	M*PQFCGRKFYLHQE
18	T	ACATGTAGCCCCAGTTCT	TGCGGCCGCA	GTTCTACTTACACCAAGAAA	M*PQFLRPQFYLHQE
19	A	ACATGTAGCCCCAGTTCTA	TGCGGCCGCA	TCTACTTACACCAAGAAAAG	M*POFYAAAFYLHQE
20	C	ACATGTAGCCCCAGTTCTACT	TGCGGCCGCA	TCTACTTACACCAAGAAAAGG	M*PQFYCGRIFYLHQER
21	T	CATGTAGCCCCAGTTCTACT	TGCGGCCGCA	CTACTTACACCAAGAAAAGGC	M*PQFYLRPHYLHQER
22	T	ATGTAGCCCCAGTTCTACTT	TGCGGCCGCA	TACTTACACCAAGAAAAGGCT	M*PQFYFAAAYLHQER
23	A	TGTAGCCCCAGTTCTACTTA	TGCGGCCGCA	ACTTACACCAAGAAAAGGCTG	*PQFYLCGRNLHQERL
24	C	GTAGCCCCAGTTCTACTTAC	TGCGGCCGCA	CTTACACCAAGAAAAGGCTGG	*PQFYLLRPHLHQERL
25	A	TAGCCCCAGTTCTACTTACA	TGCGGCCGCA	TTACACCAAGAAAAGGCTGGA	*PQFYLHAAALHQERL
26	C	AGCCCCAGTTCTACTTACAC	TGCGGCCGCA	TACACCAAGAAAAGGCTGGA	PQFYLHCGRIHQERLE
27	C	GCCCCAGTTCTACTTACACC	TGCGGCCGCA	ACACCAAGAAAAGGCTGGAAG	PQFYLHLRPHQERLE
28	A	CCCCAGTTCTACTTACACCA	TGCGGCCGCA	CACCAAGAAAAGGCTGGAAGG	PQFYLHAAAHQERLE
29	A	CCCAGTTCTACTTACACCAA	TGCGGCCGCA	ACCAAGAAAAGGCTGGAAGGG	QFYLHQCGRNRQLEGL
30	G	CCAGTTCTACTTACACCAAG	TGCGGCCGCA	CCAAGAAAAGGCTGGAAGGGC	QFYLHQVRPHQERLEGL
31	A	CAGTTCTACTTACACCAAGA	TGCGGCCGCA	CAAGAAAAGGCTGGAAGGGCT	QFYLHQDAAAHQERLEGL
32	A	AGTTCTACTTACACCAAGAA	TGCGGCCGCA	AAGAAAAGGCTGGAAGGGCTA	FYLHQECGRKRYLHQEGL
33	A	GTCTACTTACACCAAGAAA	TGCGGCCGCA	AGAAAAGGCTGGAAGGGCTAA	FYLHQEMRPERLEGL
34	G	TTCTACTTACACCAAGAAAAG	TGCGGCCGCA	GAAAAGGCTGGAAGGGCTAAT	FYLHQESAAAERLEGL
35	G	TCTACTTACACCAAGAAAAGG	TGCGGCCGCA	AAAGGCTGGAAGGGCTAATT	YLHQECGRKRYLHQEGL
36	C	CTACTTACACCAAGAAAAGGC	TGCGGCCGCA	AAGGCTGGAAGGGCTAATTCT	YLHQERLRRPQLEGL
37	T	TACTTACACCAAGAAAAGGCT	TGCGGCCGCA	AGGCTGGAAGGGCTAATTCA	YLHQERLCAARLEGL
38	G	ACTTACACCAAGAAAAGGCTG	TGCGGCCGCA	GGCTGGAAGGGCTAATTAC	LHQERLRCRRLEGLIH
39	G	CTTACACCAAGAAAAGGCTGG	TGCGGCCGCA	GCTGGAAGGGCTAATTCACT	LHQERLVRPQLEGLIH
40	A	TTACACCAAGAAAAGGCTGGA	TGCGGCCGCA	CTGGAAGGGCTAATTCACTC	LHQERLDAALQLEGLIH
41	A	TACACCAAGAAAAGGCTGGA	TGCGGCCGCA	TGGAAGGGCTAATTCACTCC	HQERLECGRMEGLIHS
42	G	ACACCAAGAAAAGGCTGGAAG	TGCGGCCGCA	GGAAGGGCTAATTCACTCCC	HQERLEVRPQLEGLIHS
43	G	CACCAAGAAAAGGCTGGAAGG	TGCGGCCGCA	GAAAGGGCTAATTCACTCCCA	HQERLEGAAGLEGLIHS
44	G	ACCAGAAGAAAAGGCTGGAAGGG	TGCGGCCGCA	AAGGGCTAATTCACTCCCAA	QERLEGCGRKGLIHSQ
45	C	CCAAGAAAAGGCTGGAAGGGC	TGCGGCCGCA	AGGGCTAATTCACTCCCAAAG	QERLEGLAAAGLIHSQ
46	T	CAAGAAAAGGCTGGAAGGGCT	TGCGGCCGCA	GGGCTAATTCACTCCCAAAG	QERLEGLAAAGLIHSQ
47	A	AAGAAAAGGCTGGAAGGGCTA	TGCGGCCGCA	GGCTAATTCACTCCCAAAGA	ERLEGLCGRRLIHSQR
48	A	AGAAGGCTGGAAGGGCTAA	TGCGGCCGCA	GCTAATTCACTCCCAAAGAA	ERLEGLMRPQIHSQR
49	T	GAAAGGCTGGAAGGGCTAAT	TGCGGCCGCA	CTAATTCACTCCCAAAGAAG	ERLEGLIAALHSQR
50	T	AAAGGCTGGAAGGGCTAATT	TGCGGCCGCA	TAATTCACTCCCAAAGAAGA	RLEGLICGRIIHSQR
51	C	AAGGCTGGAAGGGCTAATTC	TGCGGCCGCA	AATTCACTCCCAAAGAAGAC	RLEGLILRPQIHSQR
52	A	AGGCTGGAAGGGCTAATTCA	TGCGGCCGCA	ATTCACTCCCAAAGAAGACA	RLEGLIHAAIHSQR
53	C	GGCTGGAAGGGCTAATTCAC	TGCGGCCGCA	TTCACTCCCAAAGAAGACAA	LEGLIHCGRISQR
54	T	GCTGGAAGGGCTAATTCACT	TGCGGCCGCA	TCACTCCCAAAGAAGACAAG	LEGLIHLRPHHSQR
55	C	CTGGAAGGGCTAATTCACTC	TGCGGCCGCA	CACTCCCAAAGAAGACAAGA	LEGLIHSAAHSQR
56	C	TGGAAGGGCTAATTCACTCC	TGCGGCCGCA	ACTCCCAAAGAAGACAAGAT	EGLIHSGRNSQR
57	C	GGAAGGGCTAATTCACTCCC	TGCGGCCGCA	CTCCCAAAGAAGACAAGATA	EGLIHSRPHSQR
58	A	GAAGGGCTAATTCACTCCCA	TGCGGCCGCA	TCCCAAAGAAGACAAGATAT	EGLIHSAAAQR
59	A	AAGGGCTAATTCACTCCCAA	TGCGGCCGCA	CCCAAAGAAGACAAGATATC	GLIHSQCGRTQR
60	A	AGGGCTAATTCACTCCCAA	TGCGGCCGCA	CCAAAGAAGACAAGATATCC	GLIHSQMRPQR
61	G	GGCTAATTCACTCCCAAAG	TGCGGCCGCA	CAAGAAGACAAGATATCCT	GLIHSQSAARQR
62	A	GGCTAATTCACTCCCAAAGA	TGCGGCCGCA	AAAGAAGACAAGATATCCTT	LIHSQRGRKRQR
63	A	GCTAATTCACTCCCAAAGAA	TGCGGCCGCA	AAGAAGACAAGATATCCTTG	LIHSQRMRPQR
64	G	CTAATTCACTCCCAAAGAAG	TGCGGCCGCA	AGAAGACAAGATATCCTTGA	LIHSQRSAAARQR
65	A	TAATTCACTCCCAAAGAAGA	TGCGGCCGCA	GAAGACAAGATATCCTTGAT	IHSQRRCGRQR
66	C	AATTCACTCCCAAAGAAGAC	TGCGGCCGCA	AAGACAAGATATCCTTGATC	IHSQRRLRPQR
67	A	ATTCACTCCCAAAGAAGACA	TGCGGCCGCA	AGACAAGATATCCTTGATCT	IHSQRHAAARQR
68	A	TTCACTCCCAAAGAAGACAA	TGCGGCCGCA	GACAAGATATCCTTGATCTG	HSQRRCGRQR
69	G	TCACTCCCAAAGAAGACAAAG	TGCGGCCGCA	CAAGATATCCTTGATCTGTG	HSQRRCGRQR
70	A	CACTCCCAAAGAAGACAAAGA	TGCGGCCGCA	CAAGATATCCTTGATCTGTG	HSQRRCGRQR
71	T	ACTCCCAAAGAAGACAAAGAT	TGCGGCCGCA	AAGATATCCTTGATCTGTGG	SQRRCGRQR
72	A	CTCCCAAAGAAGACAAAGATA	TGCGGCCGCA	AGATATCCTTGATCTGTGGA	SQRRCGRQR
73	T	TCCCAAAGAAGACAAAGATAT	TGCGGCCGCA	GATATCCTTGATCTGTGGAT	SQRRCGRQR
74	C	CCCAAAGAAGACAAAGATATC	TGCGGCCGCA	ATATCCTTGATCTGTGGATC	QRRCGRQR
75	C	CCAAAGAAGACAAAGATATCC	TGCGGCCGCA	TATCCTTGATCTGTGGATCT	QRRCGRQR
76	T	CAAAGAAGACAAAGATATCCT	TGCGGCCGCA	ATCCTTGATCTGTGGATCTA	QRRCGRQR
77	T	AAAGAAGACAAAGATATCCTT	TGCGGCCGCA	TCCTTGATCTGTGGATCTAC	QRRCGRQR
78	G	AAGAAGACAAAGATATCCTTG	TGCGGCCGCA	CCTTGATCTGTGGATCTACC	RRQDLVRPHDLWI
79	A	AGAAGACAAAGATATCCTTGA	TGCGGCCGCA	CTTGATCTGTGGATCTACCA	RRQDLDAALDLWI
80	T	GAAGACAAAGATATCCTTGAT	TGCGGCCGCA	TTGATCTGTGGATCTACCAC	RQDLDCGRIDLWI

81	C	AAGACAAGATATCCTTGATC	TGCGGCCGCA	TGATCTGTGGATCTACCACA	RQDILDLRPHDLWIYH
82	T	AGACAAGATATCCTTGATCT	TGCGGCCGCA	GATCTGTGGATCTACCACAC	RQDILDAAADLWIYH
83	G	GACAAGATATCCTTGATCTG	TGCGGCCGCA	ATCTGTGGATCTACCACACA	QDILDLCGRNLWIYHT
84	T	ACAAGATATCCTTGATCTGT	TGCGGCCGCA	TCTGTGGATCTACCACACAC	QDILDLLRPHLWIYHT
85	G	CAAGATATCCTTGATCTGTG	TGCGGCCGCA	CTGTGGATCTACCACACACA	QDILDLCAAALWIYHT
86	G	AAGATATCCTTGATCTGTGG	TGCGGCCGCA	TGTGGATCTACCACACACAA	DILDLCGRMWWIYHTQ
87	A	AGATATCCTTGATCTGTGGA	TGCGGCCGCA	GTGGATCTACCACACACAAG	DILDLMWRPWIYHTQ
88	T	GATATCCTTGATCTGTGGAT	TGCGGCCGCA	TGGATCTACCACACACAAGG	DILDWIAAAWIYHTQ
89	C	ATATCCTTGATCTGTGGATC	TGCGGCCGCA	GGATCTACCACACACAAGGC	ILDWLICGRRIYHTQG
90	T	TATCCTTGATCTGTGGATCT	TGCGGCCGCA	GATCTACCACACACAAGGCT	ILDWLILRPOIYHTQG
91	A	ATCCTTGATCTGTGGATCTA	TGCGGCCGCA	ATCTACCACACACAAGGCTA	ILDWLYAAAIYHTQG
92	C	TCCTTGATCTGTGGATCTAC	TGCGGCCGCA	TCTACCACACACAAGGCTAC	LDLWIYCGRIYHTQGY
93	C	CCTTGATCTGTGGATCTACC	TGCGGCCGCA	CTACCACACACAAGGCTACT	LDLWIYLRPHYHTQGY
94	A	CTTGATCTGTGGATCTACCA	TGCGGCCGCA	TACCACACACAAGGCTACTT	LDLWIYHAAAIYHTQGY
95	C	TTGATCTGTGGATCTACCAC	TGCGGCCGCA	ACCACACACAAGGCTACTTC	DLWIYHGRNHTQGYF
96	A	TGATCTGTGGATCTACCACA	TGCGGCCGCA	CCACACACAAGGCTACTTCC	DLWIYHMRPHHTQGYF
97	C	GATCTGTGGATCTACCACAC	TGCGGCCGCA	CACACACAAGGCTACTTCCC	DLWIYHTAAAHTQGYF
98	A	ATCTGTGGATCTACCACACA	TGCGGCCGCA	ACACACAAGGCTACTTCCCT	LWIYHTCGRNHTQGYF
99	C	TCTGTGGATCTACCACACAC	TGCGGCCGCA	CACACAAGGCTACTTCCCTG	LWIYHTLRPHHTQGYF
100	A	CTGTGGATCTACCACACACA	TGCGGCCGCA	ACACAAGGCTACTTCCCTGA	LWIYHTHAAATQGYF
101	A	TGTGGATCTACCACACACAA	TGCGGCCGCA	CACAAGGCTACTTCCCTGAT	WIYHTQVRRPQGYFPD
102	G	GTGGATCTACCACACACAAG	TGCGGCCGCA	ACAAGGCTACTTCCCTGATT	WIYHTQVRRPQGYFPD
103	G	TGGATCTACCACACACAAGG	TGCGGCCGCA	CAAGGCTACTTCCCTGATTG	WIYHTQVRRPQGYFPD
104	C	GGATCTACCACACACAAGGC	TGCGGCCGCA	AAGGCTACTTCCCTGATTGG	IYHTQCGRKGYPFDW
105	T	GATCTACCACACACAAGGCT	TGCGGCCGCA	AGGCTACTTCCCTGATTGGC	IYHTQGLRPPQGYFPD
106	A	ATCTACCACACACAAGGCTA	TGCGGCCGCA	GGCTACTTCCCTGATTGGCA	IYHTQGYFAAIYFPD
107	C	TCTACCACACACAAGGCTAC	TGCGGCCGCA	GTACTTCCCTGATTGGCAG	YHTQGYCGRSYPFDWQ
108	T	CTACCACACACAAGGCTACT	TGCGGCCGCA	CTACTTCCCTGATTGGCAGA	YHTQGYLRPHYFPD
109	T	TACCACACACAAGGCTACTT	TGCGGCCGCA	TACTTCCCTGATTGGCAGAA	YHTQGYFAAIYFPD
110	C	ACCACACACAAGGCTACTTCC	TGCGGCCGCA	ACTTCCCTGATTGGCAGAAC	HTQGYFCGRNFPD
111	C	CCACACACAAGGCTACTTCCC	TGCGGCCGCA	CTTCCCTGATTGGCAGAACT	HTQGYFLRPHYFPD
112	C	CACACACAAGGCTACTTCCC	TGCGGCCGCA	TTCCCTGATTGGCAGAACTA	HTQGYFPAAIYFPD
113	T	ACACACAAGGCTACTTCCCT	TGCGGCCGCA	TCCCTGATTGGCAGAACTAC	TQGYFPCGRIPD
114	G	CACACAAGGCTACTTCCCTG	TGCGGCCGCA	CCCTGATTGGCAGAACTACA	TQGYFVRRPDPD
115	A	ACACAAGGCTACTTCCCTGA	TGCGGCCGCA	CCTGATTGGCAGAACTACAC	TQGYFPDAAAPD
116	T	CACAAGGCTACTTCCCTGAT	TGCGGCCGCA	CTGATTGGCAGAACTACACA	QGYFPDGRD
117	T	ACAAGGCTACTTCCCTGATT	TGCGGCCGCA	TGATTGGCAGAACTACACAC	QGYFPDLRPHD
118	G	CAAGGCTACTTCCCTGATTG	TGCGGCCGCA	GATTGGCAGAACTACACACC	QGYFPDCAAAD
119	G	AAGGCTACTTCCCTGATTGG	TGCGGCCGCA	ATTGGCAGAACTACACACCA	GYFPDWCGRN
120	C	AGGCTACTTCCCTGATTGGC	TGCGGCCGCA	TTGGCAGAACTACACACCAG	GYFPDWRPHW
121	A	GGCTACTTCCCTGATTGGCA	TGCGGCCGCA	TGGCAGAACTACACACCAGG	GYFPDWHAAAW
122	G	GCTACTTCCCTGATTGGCAG	TGCGGCCGCA	GGCAGAACTACACACCAGGG	YFPDWCGRN
123	A	CTACTTCCCTGATTGGCAGA	TGCGGCCGCA	GCAGAACTACACACCAGGGC	YFPDWCGRN
124	A	TACTTCCCTGATTGGCAGAA	TGCGGCCGCA	CAGAACTACACACCAGGGCC	YFPDWCGRN
125	C	ACTTCCCTGATTGGCAGAAC	TGCGGCCGCA	AGAACTACACACCAGGGCCA	FPDWCGRN
126	T	CTTCCCTGATTGGCAGAACT	TGCGGCCGCA	GAACTACACACCAGGGCCAG	FPDWCGRN
127	A	TTCCCTGATTGGCAGAACTA	TGCGGCCGCA	ACTACACACCAGGGCCAGG	FPDWCGRN
128	C	TCCCTGATTGGCAGAACTAC	TGCGGCCGCA	ACTACACACCAGGGCCAGGG	PDWCGRN
129	A	CCCTGATTGGCAGAACTACA	TGCGGCCGCA	CTACACACCAGGGCCAGGGG	PDWCGRN
130	C	CCTGATTGGCAGAACTACAC	TGCGGCCGCA	TACACACCAGGGCCAGGGGT	PDWCGRN
131	A	CTGATTGGCAGAACTACACA	TGCGGCCGCA	ACACACCAGGGCCAGGGGTC	DWCGRN
132	C	TGATTGGCAGAACTACACAC	TGCGGCCGCA	CACACCAGGGCCAGGGGTCA	DWCGRN
133	C	GATTGGCAGAACTACACACC	TGCGGCCGCA	ACACCAGGGCCAGGGGTGAG	DWCGRN
134	A	ATTGGCAGAACTACACACCA	TGCGGCCGCA	CACCAGGGCCAGGGGTGAGA	WQNYTPCGR
135	G	TTGGCAGAACTACACACCAG	TGCGGCCGCA	ACCAGGGCCAGGGGTGAGAT	WQNYTPVRR
136	G	TGGCAGAACTACACACCAGG	TGCGGCCGCA	CCAGGGCCAGGGGTGAGATA	WQNYTPGAA
137	G	GGCAGAACTACACACCAGGG	TGCGGCCGCA	CAGGGCCAGGGGTGAGATAT	QNYTPGCR
138	C	GCAGAACTACACACCAGGGC	TGCGGCCGCA	AGGGCCAGGGGTGAGATATC	QNYTPGCR
139	C	CAGAACTACACACCAGGGCC	TGCGGCCGCA	GGGCCAGGGGTGAGATATCC	QNYTPGAA
140	A	AGAACTACACACCAGGGCCA	TGCGGCCGCA	GGCCAGGGGTGAGATATCCA	NYTPGCR
141	G	GAACTACACACCAGGGCCAG	TGCGGCCGCA	GCCAGGGGTGAGATATCCCA	NYTPGCR
142	G	AACTACACACCAGGGCCAGG	TGCGGCCGCA	CCAGGGGTGAGATATCCACT	NYTPGCR
143	G	ACTACACACCAGGGCCAGGG	TGCGGCCGCA	CAGGGGTGAGATATCCACTG	YTPGCR
144	G	CTACACACCAGGGCCAGGGG	TGCGGCCGCA	AGGGGTGAGATATCCACTGA	YTPGCR
145	T	TACACACCAGGGCCAGGGGT	TGCGGCCGCA	GGGGTGAGATATCCACTGAC	YTPGCR
146	C	ACACACCAGGGCCAGGGGTC	TGCGGCCGCA	GGGTGAGATATCCACTGACC	TPGCR
147	A	CACACCAGGGCCAGGGGTCA	TGCGGCCGCA	GGTCAAGATATCCACTGACCT	TPGCR
148	G	ACACCAGGGCCAGGGGTGAG	TGCGGCCGCA	GTCAGATATCCACTGACCTT	TPGCR
149	A	CACCAGGGCCAGGGGTGAGA	TGCGGCCGCA	TCAGATATCCACTGACCTTT	PGCR
150	T	ACCAGGGCCAGGGGTGAGAT	TGCGGCCGCA	CAGATATCCACTGACCTTTG	PGCR
151	A	CCAGGGCCAGGGGTGAGATA	TGCGGCCGCA	AGATATCCACTGACCTTTGG	PGCR
152	T	CAGGGCCAGGGGTGAGATAT	TGCGGCCGCA	GATATCCACTGACCTTTGGA	PGCR
153	C	AGGGCCAGGGGTGAGATATC	TGCGGCCGCA	ATATCCACTGACCTTTGGAT	PGCR
154	C	GGCCAGGGGTGAGATATCCC	TGCGGCCGCA	TATCCACTGACCTTTGGATG	PGCR
155	A	GGCCAGGGGTGAGATATCCA	TGCGGCCGCA	ATCCACTGACCTTTGGATGG	PGCR
156	C	GCCAGGGGTGAGATATCCAC	TGCGGCCGCA	TCCACTGACCTTTGGATGGT	PGCR
157	T	CCAGGGGTGAGATATCCACT	TGCGGCCGCA	CCAAGGGGTGAGATATCCACT	PGCR
158	G	CAGGGGTGAGATATCCACTG	TGCGGCCGCA	CAAGGGGTGAGATATCCACTG	PGCR
159	A	AGGGGTGAGATATCCACTGA	TGCGGCCGCA	ACTGACCTTTGGATGGTGTCT	PGCR
160	C	GGGGTCAAGATATCCACTGAC	TGCGGCCGCA	CTGACCTTTGGATGGTGTCTA	PGCR
161	C	GGGTCAAGATATCCACTGACC	TGCGGCCGCA	TGACCTTTGGATGGTGTCTAC	PGCR
162	T	GGTCAAGATATCCACTGACCT	TGCGGCCGCA	GACCTTTGGATGGTGTCTACA	PGCR
163	T	GTCAGATATCCACTGACCTTT	TGCGGCCGCA	ACCTTTGGATGGTGTCTACAA	PGCR
164	T	TCAGATATCCACTGACCTTTG	TGCGGCCGCA	CCTTTGGATGGTGTCTACAAG	PGCR
165	G	CAGATATCCACTGACCTTTGG	TGCGGCCGCA	CTTTGGATGGTGTCTACAAGC	PGCR
166	G	AGATATCCACTGACCTTTGGG	TGCGGCCGCA	TTTGGATGGTGTCTACAAGCT	PGCR

167	A	GATATCCACTGACCTTTGGA	TGCGGCCGCA	TTGGATGGTGCTACAAGCTA	YPLTFGCGRIGWCYKL
168	T	ATATCCACTGACCTTTGGAT	TGCGGCCGCA	TGGATGGTGCTACAAGCTAG	YPLTFGLRPHGWYKL
169	G	TATCCACTGACCTTTGGATG	TGCGGCCGCA	GGATGGTGCTACAAGCTAGT	YPLTFGCAAGRWYKL
170	G	ATCCACTGACCTTTGGATGG	TGCGGCCGCA	GATGGTGCTACAAGCTAGTA	PLTFGWKGRRWYKLV
171	T	TCCTACTGACCTTTGGATGGT	TGCGGCCGCA	ATGGTGCTACAAGCTAGTAC	PLTFGWLRPQWCYKLV
172	G	CCACTGACCTTTGGATGGTG	TGCGGCCGCA	TGGTGCTACAAGCTAGTACC	PLTFGWCAAAWCYKLV
173	C	CACTGACCTTTGGATGGTGC	TGCGGCCGCA	GGTGCTACAAGCTAGTACCA	LTFGWCCGRRCYKLV
174	T	ACTGACCTTTGGATGGTGCT	TGCGGCCGCA	GTGCTACAAGCTAGTACCAG	LTFGWCLRPQCYKLV
175	A	CTGACCTTTGGATGGTGCTA	TGCGGCCGCA	TGCTACAAGCTAGTACCAGT	LTFGWCAAAACYKLV
176	C	TGACCTTTGGATGGTGCTAC	TGCGGCCGCA	GCTACAAGCTAGTACCAGTT	TFGWYCYGRSYKLV
177	A	GACCTTTGGATGGTGCTACA	TGCGGCCGCA	CTACAAGCTAGTACCAGTTG	TFGWYCYMRPHYKLV
178	A	ACCTTTGGATGGTGCTACAA	TGCGGCCGCA	TACAAGCTAGTACCAGTTGA	TFGWYCYNAAAYKLV
179	G	CCTTTGGATGGTGCTACAAG	TGCGGCCGCA	ACAAGCTAGTACCAGTTGAG	FGWCYKGRNKLV
180	C	CTTTGGATGGTGCTACAAGC	TGCGGCCGCA	CAAGCTAGTACCAGTTGAGC	FGWCYKLRPHKLV
181	T	TTTGGATGGTGCTACAAGCT	TGCGGCCGCA	AGCTAGTACCAGTTGAGCC	FGWCYKLAALKLV
182	A	TTGGATGGTGCTACAAGCTA	TGCGGCCGCA	AGCTAGTACCAGTTGAGCCA	GWYKLCGRKLV
183	G	TGGATGGTGCTACAAGCTAG	TGCGGCCGCA	GCTAGTACCAGTTGAGCCAG	GWYKLVRLV
184	T	GGATGGTGCTACAAGCTAGT	TGCGGCCGCA	CTAGTACCAGTTGAGCCAGA	GWYKLVRAALV
185	A	GATGGTGCTACAAGCTAGTA	TGCGGCCGCA	TAGTACCAGTTGAGCCAGAT	WCYKLVCGRIV
186	C	ATGGTGCTACAAGCTAGTAC	TGCGGCCGCA	AGTACCAGTTGAGCCAGATA	WCYKLVLRPQV
187	C	TGGTGCTACAAGCTAGTACC	TGCGGCCGCA	GTACCAGTTGAGCCAGATAA	WCYKLVPAAV
188	A	GGTGCTACAAGCTAGTACCA	TGCGGCCGCA	TACCAGTTGAGCCAGATAAG	CYKLVPCGRIP
189	G	GTGCTACAAGCTAGTACCAG	TGCGGCCGCA	ACCAGTTGAGCCAGATAAGG	CYKLVPRPQV
190	T	TGCTACAAGCTAGTACCAGT	TGCGGCCGCA	CCAGTTGAGCCAGATAAGGT	CYKLVPAVAA
191	T	GCTACAAGCTAGTACCAGTT	TGCGGCCGCA	CAGTTGAGCCAGATAAGGTA	YKLVPCGRTP
192	G	CTACAAGCTAGTACCAGTTG	TGCGGCCGCA	AGTTGAGCCAGATAAGGTAG	YKLVPRPQV
193	A	TACAAGCTAGTACCAGTTGA	TGCGGCCGCA	GTTGAGCCAGATAAGGTAGA	YKLVVDAAA
194	G	ACAAGCTAGTACCAGTTGAG	TGCGGCCGCA	TTGAGCCAGATAAGGTAGAA	KLVPVECGRI
195	C	CAAGCTAGTACCAGTTGAGC	TGCGGCCGCA	TGAGCCAGATAAGGTAGAA	KLVPVELRPH
196	C	AAGCTAGTACCAGTTGAGCC	TGCGGCCGCA	GAGCCAGATAAGGTAGAA	KLVPVEPAA
197	A	AGCTAGTACCAGTTGAGCCA	TGCGGCCGCA	AGCCAGATAAGGTAGAA	LVPVEPAA
198	G	GCTAGTACCAGTTGAGCCAG	TGCGGCCGCA	GCCAGATAAGGTAGAA	LVPVEPAA
199	A	CTAGTACCAGTTGAGCCAGA	TGCGGCCGCA	CCAGATAAGGTAGAA	LVPVEPAA
200	T	TAGTACCAGTTGAGCCAGAT	TGCGGCCGCA	CAGATAAGGTAGAA	VPVEPDMRP
201	A	AGTACCAGTTGAGCCAGATA	TGCGGCCGCA	AGATAAGGTAGAA	VPVEPDMRP
202	A	GTACCAGTTGAGCCAGATAA	TGCGGCCGCA	GATAAGGTAGAA	VPVEPDMRP
203	G	TACCAGTTGAGCCAGATAAG	TGCGGCCGCA	ATAAGGTAGAA	PVEPDKCGR
204	G	ACCAGTTGAGCCAGATAAAG	TGCGGCCGCA	TAAGGTAGAA	PVEPDKVPR
205	T	CCAGTTGAGCCAGATAAAGT	TGCGGCCGCA	AAGGTAGAA	PVEPDKVAA
206	A	CAGTTGAGCCAGATAAAGTA	TGCGGCCGCA	AGGTAGAA	VEPDKVCRK
207	G	AGTTGAGCCAGATAAAGTAG	TGCGGCCGCA	GGTAGAA	VEPDKVVRP
208	A	GTTGAGCCAGATAAAGTAGA	TGCGGCCGCA	TGAGAA	VEPDKVDA
209	A	TTGAGCCAGATAAAGGTAGAA	TGCGGCCGCA	TAGAA	EPDKVECGR
210	G	TGAGCCAGATAAAGGTAGAA	TGCGGCCGCA	AGAA	EPDKVEVRP
211	A	GAGCCAGATAAAGGTAGAA	TGCGGCCGCA	GAA	EPDKVEDAA
212	G	AGCCAGATAAAGGTAGAA	TGCGGCCGCA	AAG	PDKVEECGR
213	G	GCCAGATAAAGGTAGAA	TGCGGCCGCA	AGG	PDKVEEVRP
214	C	CCAGATAAAGGTAGAA	TGCGGCCGCA	GAG	PDKVEEAAA
215	C	CAGATAAAGGTAGAA	TGCGGCCGCA	AGG	DKVEEACGR
216	A	AGATAAAGGTAGAA	TGCGGCCGCA	GG	DKVEEAMRP
217	A	GATAAAGGTAGAA	TGCGGCCGCA	GA	DKVEEANA
218	T	ATAAGGTAGAA	TGCGGCCGCA	CA	KVEEANCGR
219	A	TAAGGTAGAA	TGCGGCCGCA	CA	KVEEANMRP
220	A	AAGGTAGAA	TGCGGCCGCA	AA	KVEEANNA
221	A	AGGTAGAA	TGCGGCCGCA	AA	VEEANKCRN
222	G	GGTAGAA	TGCGGCCGCA	AA	VEEANKVRP
223	G	GTAGAA	TGCGGCCGCA	AA	VEEANKGAA
224	A	TAGAA	TGCGGCCGCA	AA	EEANKGCRK
225	G	AGAA	TGCGGCCGCA	AA	EEANKGVRP
226	A	GAA	TGCGGCCGCA	AA	EEANKGDAA
227	G	AAG	TGCGGCCGCA	AA	EANKGECGR
228	A	AGG	TGCGGCCGCA	AA	EANKGEMRP
229	A	GAG	TGCGGCCGCA	AA	EANKGENAA
230	C	AGG	TGCGGCCGCA	AA	ANKGENCGR
231	A	GG	TGCGGCCGCA	AA	ANKGENMRP
232	C	GCA	TGCGGCCGCA	AA	ANKGENTAA
233	C	CA	TGCGGCCGCA	AA	NKGENTCGR
234	A	CA	TGCGGCCGCA	AA	NKGENMTRP
235	G	AA	TGCGGCCGCA	AA	NKGENSA
236	C	ATA	TGCGGCCGCA	AA	KGENTSCGR
237	T	TAA	TGCGGCCGCA	AA	KGENTSLR
238	T	AA	TGCGGCCGCA	AA	KGENTSF
239	G	AAG	TGCGGCCGCA	AA	GENTSLCGR
240	T	AGG	TGCGGCCGCA	AA	GENTSLRPH
241	T	GG	TGCGGCCGCA	AA	GENTSLF
242	A	GAG	TGCGGCCGCA	AA	ENTSLLCR
243	C	AG	TGCGGCCGCA	AA	ENTSLLRP
244	A	GAG	TGCGGCCGCA	AA	ENTSLHAA
245	C	AGA	TGCGGCCGCA	AA	NTSLHCGR
246	C	GA	TGCGGCCGCA	AA	NTSLHLR
247	C	A	TGCGGCCGCA	AA	NTSLHPAA
248	T	AC	TGCGGCCGCA	AA	TSLHPCGR
249	G	CA	TGCGGCCGCA	AA	TSLHPVRP
250	T	ACC	TGCGGCCGCA	AA	TSLHPVA
251	G	CC	TGCGGCCGCA	AA	SLLHPV
252	A	CA	TGCGGCCGCA	AA	SLLHPV



253	G	AGCTTGTACACCCTGTGAG	TGCGGCCGCA	GTGAGCCTGCATGGAATGGA	SLLHPVSAAAVSLHGM
254	C	GCTTGTACACCCTGTGAGC	TGCGGCCGCA	TGAGCCTGCATGGAATGGAT	LLHPVSCGRMSLHGMD
255	C	CTTGTACACCCTGTGAGCC	TGCGGCCGCA	GAGCCTGCATGGAATGGATG	LLHPVSLRPQSLHGMD
256	T	TTGTTACACCCTGTGAGCCT	TGCGGCCGCA	AGCCTGCATGGAATGGATGA	LLHPVSLAAASLHGMD
257	G	TGTTACACCCTGTGAGCCTG	TGCGGCCGCA	GCCTGCATGGAATGGATGAC	LHPVSLCGRSLHGMD
258	C	GTTACACCCTGTGAGCCTGC	TGCGGCCGCA	CCTGCATGGAATGGATGACC	LHPVSLRPHLHGMD
259	A	TTACACCCTGTGAGCCTGCA	TGCGGCCGCA	CTGCATGGAATGGATGACCC	LHPVSLHAAALHGMD
260	T	TACACCCTGTGAGCCTGCAT	TGCGGCCGCA	TGCATGGAATGGATGACCCT	HPVSLHCGRMHGMD
261	G	ACACCCTGTGAGCCTGCATG	TGCGGCCGCA	GCATGGAATGGATGACCCTG	HPVSLHVRPQHGMDDP
262	G	CACCCTGTGAGCCTGCATGG	TGCGGCCGCA	CATGGAATGGATGACCCTGA	HPVSLHGAAAHGMDDP
263	A	ACCCTGTGAGCCTGCATGGA	TGCGGCCGCA	ATGGAATGGATGACCCTGAG	PVSLHGCGRNGMDDPE
264	A	CCTGTGAGCCTGCATGGAA	TGCGGCCGCA	TGGAATGGATGACCCTGAGA	PVSLHGMFRPHGMDDPE
265	T	CCTGTGAGCCTGCATGGAAT	TGCGGCCGCA	GGAATGGATGACCCTGAGAG	PVSLHGIAAAGMDDPE
266	G	CTGTGAGCCTGCATGGAATG	TGCGGCCGCA	GAATGGATGACCCTGAGAGA	VSLHGMCGRMDDPER
267	G	TGTGAGCCTGCATGGAATGG	TGCGGCCGCA	AATGGATGACCCTGAGAGAG	VSLHGMVVRPQMDPPER
268	A	GTGAGCCTGCATGGAATGGA	TGCGGCCGCA	ATGGATGACCCTGAGAGAGA	VSLHGMDAAAMDDPER
269	T	TGAGCCTGCATGGAATGGAT	TGCGGCCGCA	TGGATGACCCTGAGAGAGAA	SLHGMDCGRMDDPERE
270	G	GAGCCTGCATGGAATGGATG	TGCGGCCGCA	GGATGACCCTGAGAGAGAA	SLHGMDVVRPQMDPERE
271	A	AGCCTGCATGGAATGGATGA	TGCGGCCGCA	GATGACCCTGAGAGAGAAAGT	SLHGMDDAADDDPERE
272	C	GCCTGCATGGAATGGATGAC	TGCGGCCGCA	ATGACCCTGAGAGAGAAAGTG	LHGMDDCGRNDPERE
273	C	CCTGCATGGAATGGATGACC	TGCGGCCGCA	TGACCCTGAGAGAGAAAGTGT	LHGMDLRPHDPERE
274	C	CTGCATGGAATGGATGACCC	TGCGGCCGCA	GACCCTGAGAGAGAAAGTGT	LHGMDPAAADPERE
275	T	TGCATGGAATGGATGACCCT	TGCGGCCGCA	ACCCTGAGAGAGAAAGTGTTA	HGMDDPVRPMPEREVL
276	G	GCATGGAATGGATGACCCTG	TGCGGCCGCA	CCCTGAGAGAGAAAGTGTTAG	HGMDDPVRPMPEREVL
277	A	CATGGAATGGATGACCCTGA	TGCGGCCGCA	CCTGAGAGAGAAAGTGTTAGA	HGMDDPDAAPEREVL
278	G	ATGGAATGGATGACCCTGAG	TGCGGCCGCA	CTGAGAGAGAAAGTGTAGAG	GMDDPDCGRREVL
279	A	TGGAATGGATGACCCTGAGA	TGCGGCCGCA	TGAGAGAGAAAGTGTAGAGT	GMDDPMPREREVL
280	G	GGAATGGATGACCCTGAGAG	TGCGGCCGCA	GAGAGAGAAAGTGTAGAGTG	GMDDPESAAEREVL
281	A	GAATGGATGACCCTGAGAGA	TGCGGCCGCA	AGAGAGAAAGTGTAGAGTGG	MDDPERCGRKREVL
282	G	AATGGATGACCCTGAGAGAG	TGCGGCCGCA	GAGAGAAAGTGTAGAGTGG	MDDPERVVRPQREVL
283	A	ATGGATGACCCTGAGAGAGA	TGCGGCCGCA	AGAGAAAGTGTAGAGTGGAG	MDDPERDAAAREVL
284	A	TGGATGACCCTGAGAGAGAA	TGCGGCCGCA	GAGAAGTGTAGAGTGGAGG	DDPERECGRREVL
285	G	GGATGACCCTGAGAGAGAA	TGCGGCCGCA	AGAAGTGTAGAGTGGAGGT	DDPEREVRPQREVL
286	T	GATGACCCTGAGAGAGAAAGT	TGCGGCCGCA	GAAGTGTAGAGTGGAGGTT	DDPEREVAAREVL
287	G	ATGACCCTGAGAGAGAAAGT	TGCGGCCGCA	AAGTGTAGAGTGGAGGTTT	DPEREVCGRKLWRF
288	T	TGACCCTGAGAGAGAAAGTGT	TGCGGCCGCA	AGTGTAGAGTGGAGGTTTG	DPEREVLRPQVLEWRF
289	T	GACCCTGAGAGAGAAAGTGT	TGCGGCCGCA	GTGTAGAGTGGAGGTTTGA	DPEREVFAAAVLEWRF
290	A	ACCCTGAGAGAGAAAGTGT	TGCGGCCGCA	TGTAGAGTGGAGGTTTGAC	PEREVLVGRMLEWRF
291	G	CCCTGAGAGAGAAAGTGTAG	TGCGGCCGCA	GTTAGAGTGGAGGTTTGACA	PEREVLVGRMLEWRF
292	A	CCTGAGAGAGAAAGTGTAGA	TGCGGCCGCA	TTAGAGTGGAGGTTTGACAG	PEREVLDAALWRF
293	G	CTGAGAGAGAAAGTGTAGAG	TGCGGCCGCA	TAGAGTGGAGGTTTGACAGC	EREVLECGRIEWRFS
294	T	TGAGAGAGAAAGTGTAGAGT	TGCGGCCGCA	AGAGTGGAGGTTTGACAGCC	EREVLELRPQEWRFDS
295	G	GAGAGAGAAAGTGTAGAGTG	TGCGGCCGCA	GAGTGGAGGTTTGACAGCCG	EREVLECAAAEWRFDS
296	G	AGAGAGAAAGTGTAGAGTGG	TGCGGCCGCA	AGTGGAGGTTTGACAGCCGC	REVLEWCGRWRFSR
297	A	GAGAGAAAGTGTAGAGTGG	TGCGGCCGCA	GTGGAGGTTTGACAGCCGCC	REVLEWVRPQWRFSR
298	G	AGAGAAAGTGTAGAGTGGAG	TGCGGCCGCA	TGGAGGTTTGACAGCCGCC	REVLEWAAAARWRFDS
299	G	GAGAAAGTGTAGAGTGGAGG	TGCGGCCGCA	GGAGGTTTGACAGCCGCCA	EVLEWRCGRRRFDSRL
300	T	AGAAGTGTAGAGTGGAGGT	TGCGGCCGCA	GAGGTTTGACAGCCGCCA	EVLEWRLRPQRFDSRL
301	T	GAAGTGTAGAGTGGAGGTT	TGCGGCCGCA	AGGTTTGACAGCCGCCA	EVLEWRFAAARFDSRL
302	T	AAGTGTAGAGTGGAGGTTT	TGCGGCCGCA	GGTTTGACAGCCGCCA	VLEWRFVGRRRFDSRL
303	G	AGTGTAGAGTGGAGGTTTG	TGCGGCCGCA	GTTTGACAGCCGCCA	VLEWRFVGRRRFDSRL
304	A	TGTTAGAGTGGAGGTTTGA	TGCGGCCGCA	TTTGACAGCCGCCA	VLEWRFDAAFDSRLA
305	C	GTGTAGAGTGGAGGTTTGAC	TGCGGCCGCA	TTGACAGCCGCCA	LEWRFDCGRIDSLAF
306	A	GTTAGAGTGGAGGTTTGACA	TGCGGCCGCA	TGACAGCCGCCA	LEWRFDMRPHDSLAF
307	G	TTAGAGTGGAGGTTTGACAG	TGCGGCCGCA	GACAGCCGCCA	LEWRFDSAAADSLAF
308	C	TAGAGTGGAGGTTTGACAGC	TGCGGCCGCA	ACAGCCGCCA	EWRFDSCGRNSLAFH
309	C	AGAGTGGAGGTTTGACAGCC	TGCGGCCGCA	CAGCCGCCA	EWRFDLRLPHSLAFH
310	G	GAGTGGAGGTTTGACAGCCG	TGCGGCCGCA	AGCCGCCA	EWRFDLRLPHSLAFH
311	C	AGTGGAGGTTTGACAGCCGC	TGCGGCCGCA	GCCGCCA	WRFDLRLPHSLAFH
312	C	GTGGAGGTTTGACAGCCGCC	TGCGGCCGCA	CCGCCA	WRFDLRLPHSLAFH
313	T	TGGAGGTTTGACAGCCGCCT	TGCGGCCGCA	CGCCA	WRFDLRLPHSLAFH
314	A	GGAGGTTTGACAGCCGCCTA	TGCGGCCGCA	GCCTAGCATTTCATCAGT	RFDLRLCGRSLAFH
315	G	GAGGTTTGACAGCCGCCTAG	TGCGGCCGCA	CCTAGCATTTCATCAGTGG	RFDLRLCGRSLAFH
316	C	AGGTTTGACAGCCGCCTAGC	TGCGGCCGCA	CTAGCATTTCATCAGTGGC	RFDLRLAAALAFH
317	A	GGTTTGACAGCCGCCTAGCA	TGCGGCCGCA	TAGCATTTCATCAGTGGCC	FDSLRLACGRIFHVA
318	T	GTTTGACAGCCGCCTAGCAT	TGCGGCCGCA	AGCATTTCATCAGTGGCCC	FDSLRLALRPQAFHVA
319	T	TTTGACAGCCGCCTAGCATT	TGCGGCCGCA	GCATTTCATCAGTGGCCC	FDSLRLAFAAAFHVA
320	T	TTGACAGCCGCCTAGCATT	TGCGGCCGCA	CATTTCATCAGTGGCCC	DSLRLACGRIFHVAR
321	C	TGACAGCCGCCTAGCATTTC	TGCGGCCGCA	ATTTCATCAGTGGCCC	DSLRLALRPQAFHVAR
322	A	GACAGCCGCCTAGCATTTC	TGCGGCCGCA	TTTCATCAGTGGCCC	DSLRLAFAAAFHVAR
323	T	ACAGCCGCCTAGCATTTCAT	TGCGGCCGCA	TCATCAGTGGCCC	SRLAFHCGRIHVAR
324	C	CAGCCGCCTAGCATTTCATC	TGCGGCCGCA	TCATCAGTGGCCC	SRLAFHLRPHHVAR
325	A	AGCCGCCTAGCATTTCATCA	TGCGGCCGCA	CATCAGTGGCCC	SRLAFHAAAHHVAR
326	C	GCCGCCTAGCATTTCATCAC	TGCGGCCGCA	ATCAGTGGCCC	RLAFHHCGRNHVAREL
327	G	CCGCCTAGCATTTCATCACG	TGCGGCCGCA	TCAGTGGCCC	RLAFHHCGRNHVAREL
328	T	CGCCTAGCATTTCATCACGT	TGCGGCCGCA	CAGTGGCCC	RLAFHAAAHHVAREL
329	G	GCCTAGCATTTCATCACGTG	TGCGGCCGCA	ACGTGGCCC	LAFHHCGRNVAREL
330	G	CCTAGCATTTCATCACGTGG	TGCGGCCGCA	CGTGGCCC	LAFHHCGRNVAREL
331	C	CTAGCATTTCATCACGTGGC	TGCGGCCGCA	GTGGCCC	LAFHAAAHHVAREL
332	C	TAGCATTTCATCACGTGGCC	TGCGGCCGCA	TGGCCC	AFHHCGRMARELHP
333	C	AGCATTTCATCACGTGGCCC	TGCGGCCGCA	GGCCC	AFHHCGRMARELHP
334	G	GCATTTCATCACGTGGCCC	TGCGGCCGCA	GCCC	AFHHCGRMARELHP
335	A	CATTTCATCACGTGGCCC	TGCGGCCGCA	CCCG	FHHVRCGRNHVAREL
336	G	ATTCATCACGTGGCCC	TGCGGCCGCA	CCGAGCTGCATCCGGAGT	FHHVRCGRNHVAREL
337	A	TTTCATCACGTGGCCC	TGCGGCCGCA	CGAGCTGCATCCGGAGTA	FHHVRCGRNHVAREL
338	G	TTCATCACGTGGCCC	TGCGGCCGCA	GAGCTGCATCCGGAGTAC	HVVARECGRNHVAREL

339	C	TCATCACGTGGCCCGAGAGC	TGCGGCCGCA	AGAGCTGCATCCGGAGTACT	HHVARELRPQELHPEY
340	T	CATCACGTGGCCCGAGAGCT	TGCGGCCGCA	GAGCTGCATCCGGAGTACTT	HHVARELAAAELHPEY
341	G	ATCACGTGGCCCGAGAGCTG	TGCGGCCGCA	AGCTGCATCCGGAGTACTTC	HVARELCGRKLHPEYF
342	C	TCACGTGGCCCGAGAGCTGC	TGCGGCCGCA	GCTGCATCCGGAGTACTTCA	HVARELLRPQLHPEYF
343	A	CACGTGGCCCGAGAGCTGCA	TGCGGCCGCA	CTGCATCCGGAGTACTTCAA	HVARELHAAALHPEYF
344	T	ACGTGGCCCGAGAGCTGCAT	TGCGGCCGCA	TGCATCCGGAGTACTTCAAG	VARELHCGRMHPEYFK
345	C	CGTGGCCCGAGAGCTGCATC	TGCGGCCGCA	GCATCCGGAGTACTTCAAGA	VARELHLRPQHPPEYFK
346	C	GTGGCCCGAGAGCTGCATCC	TGCGGCCGCA	CATCCGGAGTACTTCAAGAA	VARELHAAAHPPEYFK
347	G	TGGCCCGAGAGCTGCATCCG	TGCGGCCGCA	ATCCGGAGTACTTCAAGAAC	ARELHPCGRNPEYFKN
348	G	GGCCCGAGAGCTGCATCCGG	TGCGGCCGCA	TCCGGAGTACTTCAAGAACT	ARELHPVRPHPEYFKN
349	A	GCCCGAGAGCTGCATCCGGA	TGCGGCCGCA	CCGGAGTACTTCAAGAACTG	ARELHPDAAAPEYFKN
350	G	CCCGAGAGCTGCATCCGGAG	TGCGGCCGCA	CGGAGTACTTCAAGAACTGC	RELHPECGRTEYFKNC
351	T	CCGAGAGCTGCATCCGGAGT	TGCGGCCGCA	GGAGTACTTCAAGAACTGCT	RELHPELRPQPEYFKNC
352	A	CGAGAGCTGCATCCGGAGTA	TGCGGCCGCA	GAGTACTTCAAGAACTGCTG	RELHPEYAAAAYFKNC
353	C	GAGAGCTGCATCCGGAGTAC	TGCGGCCGCA	GTACTTCAAGAACTGCTGA	ELHPEYCGRKYFKNC*
354	T	AGAGCTGCATCCGGAGTACT	TGCGGCCGCA	GTACTTCAAGAACTGCTGAC	ELHPEYLRPQYFKNC*
355	T	GAGCTGCATCCGGAGTACTT	TGCGGCCGCA	TACTTCAAGAACTGCTGACA	ELHPEYFAAAAYFKNC*
356	C	AGCTGCATCCGGAGTACTTC	TGCGGCCGCA	ACTTCAAGAACTGCTGACAT	LHPEYFCGRNFKNC*H
357	A	GCTGCATCCGGAGTACTTCA	TGCGGCCGCA	CTTCAAGAACTGCTGACATC	LHPEYFMRPHFKNC*H
358	A	CTGCATCCGGAGTACTTCAA	TGCGGCCGCA	TTCAGAACTGCTGACATCG	LHPEYFNAAAFKNC*H
359	G	TGCATCCGGAGTACTTCAAG	TGCGGCCGCA	TCAAGAACTGCTGACATCGA	HPEYFKCGRIKNC*HR
360	A	GCATCCGGAGTACTTCAAGA	TGCGGCCGCA	CAAGAACTGCTGACATCGAG	HPEYFKMRPHKNC*HR
361	A	CATCCGGAGTACTTCAAGAA	TGCGGCCGCA	AAGAACTGCTGACATCGAGC	HPEYFKNAAAKNC*HR
362	C	ATCCGGAGTACTTCAAGAAC	TGCGGCCGCA	AGAACTGCTGACATCGAGCT	PEYFKNCGRKNC*HRA
363	T	TCCGGAGTACTTCAAGAACT	TGCGGCCGCA	GAAGTCTGACATCGAGCTT	PEYFKNLRPQNC*HRA
364	G	CCGGAGTACTTCAAGAACTG	TGCGGCCGCA	AACTGCTGACATCGAGCTTG	PEYFKNCAAAAC*HRA
365	C	CGGAGTACTTCAAGAACTGC	TGCGGCCGCA	ACTGCTGACATCGAGCTTGC	EYFKNCCGRNC*HRAC
366	T	GGAGTACTTCAAGAACTGCT	TGCGGCCGCA	CTGCTGACATCGAGCTTGT	EYFKNLRPHC*HRAC
367	G	GAGTACTTCAAGAACTGCTG	TGCGGCCGCA	TGCTGACATCGAGCTTGCTA	EYFKNCAAAAC*HRAC
368	A	AGTACTTCAAGAACTGCTGA	TGCGGCCGCA	GCTGACATCGAGCTTGCTAC	YFKNC*CGRS*HRACY
369	C	GTACTTCAAGAACTGCTGAC	TGCGGCCGCA	CTGACATCGAGCTTGCTACA	YFKNC*LRPH*HRACY
370	A	TACTTCAAGAACTGCTGACA	TGCGGCCGCA	TGACATCGAGCTTGCTACAA	YFKNC*HAAA*HRACY
371	T	ACTTCAAGAACTGCTGACAT	TGCGGCCGCA	GACATCGAGCTTGCTACAAG	FKNC*HCGRRHRACYK
372	C	CTTCAAGAACTGCTGACATC	TGCGGCCGCA	ACATCGAGCTTGCTACAAGG	FKNC*HLRPQHRACYK
373	G	TTCAGAACTGCTGACATCG	TGCGGCCGCA	CATCGAGCTTGCTACAAGGG	FKNC*HRAAAHRACYK
374	A	TCAAGAACTGCTGACATCGA	TGCGGCCGCA	ATCGAGCTTGCTACAAGGGA	KNC*HRCGRNHRACYK
375	G	CAAGAACTGCTGACATCGAG	TGCGGCCGCA	TCGAGCTTGCTACAAGGGAC	KNC*HRVRPHRACYK
376	C	AAGAACTGCTGACATCGAGC	TGCGGCCGCA	CGAGCTTGCTACAAGGGACT	KNC*HRAAAARACYK
377	T	AGAACTGCTGACATCGAGCT	TGCGGCCGCA	GAGCTTGCTACAAGGGACTT	NC*HRACGRNHRACYKGL
378	T	GAAGTCTGACATCGAGCTT	TGCGGCCGCA	AGCTTGCTACAAGGGACTTT	NC*HRALRPQACYKGL
379	G	AACTGCTGACATCGAGCTTG	TGCGGCCGCA	GCTTGCTACAAGGGACTTTT	NC*HRACAAAACYKGL
380	C	ACTGCTGACATCGAGCTTGC	TGCGGCCGCA	CTTGCTACAAGGGACTTTCC	C*HRACCGRTCYKGLS
381	T	CTGCTGACATCGAGCTTGT	TGCGGCCGCA	TTGCTACAAGGGACTTTCCG	C*HRACLRPHCYKGLS
382	A	TGCTGACATCGAGCTTGCTA	TGCGGCCGCA	TGCTACAAGGGACTTTCCG	C*HRACYAAACYKGLS
383	C	GCTGACATCGAGCTTGCTAC	TGCGGCCGCA	GCTACAAGGGACTTTCCGCT	*HRACYCGRSYKGLSA
384	A	CTGACATCGAGCTTGCTACA	TGCGGCCGCA	CTACAAGGGACTTTCCGCTG	*HRACYMRPHYKGLSA
385	A	TGACATCGAGCTTGCTACAA	TGCGGCCGCA	TACAAGGGACTTTCCGCTGG	*HRACYNAAAYKGLSA
386	G	GACATCGAGCTTGCTACAAG	TGCGGCCGCA	ACAAGGGACTTTCCGCTGGG	HRACYKCGRNKGLSAG
387	G	ACATCGAGCTTGCTACAAGG	TGCGGCCGCA	CAAGGGACTTTCCGCTGGGG	HRACYKVRPHKGLSAG
388	G	CATCGAGCTTGCTACAAGGG	TGCGGCCGCA	AAGGGACTTTCCGCTGGGGA	HRACYKGAAGKGLSAG
389	A	ATCGAGCTTGCTACAAGGGA	TGCGGCCGCA	AGGGACTTTCCGCTGGGGAC	RACYKCGCRKGLSAGD
390	C	TCGAGCTTGCTACAAGGGAC	TGCGGCCGCA	GGGACTTTCCGCTGGGGACT	RACYKGLRPQGLSAGD
391	T	CGAGCTTGCTACAAGGGACT	TGCGGCCGCA	GGACTTTCCGCTGGGGACTT	RACYKGLAAAGLSAGD
392	T	GAGCTTGCTACAAGGGACTT	TGCGGCCGCA	GACTTTCCGCTGGGGACTTT	ACYKGLCGRRLSAGDF
393	T	AGCTTGCTACAAGGGACTTT	TGCGGCCGCA	ACTTTCCGCTGGGGACTTTT	ACYKGLLRPQLSAGDF
394	C	GCTTGCTACAAGGGACTTTT	TGCGGCCGCA	CTTTCCGCTGGGGACTTTT	ACYKGLSAAALSAGDF
395	C	CTTGCTACAAGGGACTTTCC	TGCGGCCGCA	TTTCCGCTGGGGACTTTCCA	CYKGLSCGRISAGDFP
396	G	TTGCTACAAGGGACTTTCCG	TGCGGCCGCA	TTCCGCTGGGGACTTTCCAG	CYKGLSVRPHSAGDFP
397	C	TGCTACAAGGGACTTTCCGC	TGCGGCCGCA	TCCGCTGGGGACTTTCCAGG	CYKGLSAAAASAGDFP
398	T	GCTACAAGGGACTTTCCGCT	TGCGGCCGCA	CCGCTGGGGACTTTCCAGGG	YKGLSACRTAGDFPF
399	G	CTACAAGGGACTTTCCGCTG	TGCGGCCGCA	CGCTGGGGACTTTCCAGGGA	YKGLSASVRPHAGDFPF
400	G	TACAAGGGACTTTCCGCTGG	TGCGGCCGCA	GCTGGGGACTTTCCAGGGAG	YKGLSAGAAAAGDFPF
401	G	ACAAGGGACTTTCCGCTGGG	TGCGGCCGCA	CTGGGGACTTTCCAGGGAGG	KGLSAGCGRTGDFPGR
402	G	CAAGGGACTTTCCGCTGGGG	TGCGGCCGCA	TGGGGACTTTCCAGGGAGGC	KGLSAGVRPHGDFPGR
403	A	AAGGGACTTTCCGCTGGGGA	TGCGGCCGCA	GGGGACTTTCCAGGGAGGCG	KGLSAGDAAAAGDFPGR
404	C	AGGGACTTTCCGCTGGGGAC	TGCGGCCGCA	GGGACTTTCCAGGGAGGCGT	GLSAGDCGRNDFPGR
405	T	GGGACTTTCCGCTGGGGACT	TGCGGCCGCA	GGACTTTCCAGGGAGGCGTG	GLSAGDLRPQDFPGR
406	T	GGACTTTCCGCTGGGGACTT	TGCGGCCGCA	GACTTTCCAGGGAGGCGTGG	GLSAGDFAAADFPGR
407	T	GACTTTCCGCTGGGGACTTT	TGCGGCCGCA	ACTTTCCAGGGAGGCGTGGC	LSAGDFCGRNFPGR
408	C	ACTTTCCGCTGGGGACTTTC	TGCGGCCGCA	CTTTCCAGGGAGGCGTGGCC	LSAGDFLRPHFPGR
409	C	CTTTCCGCTGGGGACTTTCC	TGCGGCCGCA	TTTCCAGGGAGGCGTGGCCT	LSAGDFPAAAFPGRR
410	A	TTTCCGCTGGGGACTTTCCA	TGCGGCCGCA	TTCCAGGGAGGCGTGGCCTG	SAGDFPCGRIPGR
411	G	TTCCGCTGGGGACTTTCCAG	TGCGGCCGCA	TCCAGGGAGGCGTGGCCTGG	SAGDFPVRPHFPGR
412	G	TCCGCTGGGGACTTTCCAGG	TGCGGCCGCA	CCAGGGAGGCGTGGCCTGGG	SAGDFPFAAAGPGR
413	G	CCGCTGGGGACTTTCCAGGG	TGCGGCCGCA	CAGGGAGGCGTGGCCTGGGC	AGDFPFCGRTRGRGLG
414	A	CGTGGGGACTTTCCAGGGA	TGCGGCCGCA	AGGGAGGCGTGGCCTGGGCG	AGDFPGRMPPQGRGLG
415	G	GCTGGGGACTTTCCAGGGAG	TGCGGCCGCA	GGGAGGCGTGGCCTGGGCGG	AGDFPFGSAAAGRRGLG
416	G	CTGGGGACTTTCCAGGGAGG	TGCGGCCGCA	GGAGGCGTGGCCTGGGCGGG	GDFPGRCGRRRRLGG
417	C	TGGGGACTTTCCAGGGAGGC	TGCGGCCGCA	GAGGCGTGGCCTGGGCGGGA	GDFPGRRLRQRRRLGG
418	G	GGGGACTTTCCAGGGAGGCG	TGCGGCCGCA	AGGCGTGGCCTGGGCGGGAC	GDFPGRRAAARRLGG
419	T	GGGACTTTCCAGGGAGGCGT	TGCGGCCGCA	GGCGTGGCCTGGGCGGGACT	DFPGRRCGRRRRLGGT
420	G	GGACTTTCCAGGGAGGCGTG	TGCGGCCGCA	CGCTGGCCTGGGCGGGACTG	DFPGRRRRQRRRLGGT
421	G	GACTTTCCAGGGAGGCGTGG	TGCGGCCGCA	CGTGGCCTGGGCGGGACTGG	DFPGRRAAARRLGGT
422	C	ACTTTCCAGGGAGGCGTGGC	TGCGGCCGCA	TGGCCTGGGCGGGACTGGG	FPGRRCGRSGLGGT
423	C	CTTTCCAGGGAGGCGTGGCC	TGCGGCCGCA	TGGCCTGGGCGGGACTGGGG	FPGRRLRPHGLGGT
424	T	TTTTCCAGGGAGGCGTGGCCT	TGCGGCCGCA	GGCCTGGGCGGGACTGGGGA	FPGRRLAAAGLGGT

425	G	TTCCAGGGAGGCGTGGCCTG	TGCGGCCGCA	GCCTGGGCGGGACTGGGGAG	PGRRLCGRSLGGTGE
426	G	TCCAGGGAGGCGTGGCCTGG	TGCGGCCGCA	CCTGGGCGGGACTGGGGAGT	PGRRLVVRPHLGGTGE
427	G	CCAGGGAGGCGTGGCCTGGG	TGCGGCCGCA	CTGGGCGGGACTGGGGAGTG	PGRRLGAAALGGTGE
428	C	CAGGGAGGCGTGGCCTGGGC	TGCGGCCGCA	TGGGCGGGACTGGGGAGTGG	GRRRLGCGRMGGTGEW
429	G	AGGGAGGCGTGGCCTGGGCG	TGCGGCCGCA	GGGCGGGACTGGGGAGTGGC	GRRRLGVRPQGGTGEW
430	G	GGGAGGCGTGGCCTGGGCGG	TGCGGCCGCA	GGGCGGGACTGGGGAGTGGCG	GRRRLGAAASGGTGEW
431	G	GGAGGCGTGGCCTGGGCGGG	TGCGGCCGCA	GCGGGACTGGGGAGTGGCGA	RRLLGGCTGRRTGEWRA
432	A	GAGGCGTGGCCTGGGCGGGA	TGCGGCCGCA	CGGGACTGGGGAGTGGCGAG	RRLLGGMRPHGTGEWR
433	C	AGGCGTGGCCTGGGCGGGAC	TGCGGCCGCA	GGGACTGGGGAGTGGCGAGC	RRLLGGTAAAGTGEWR
434	T	GGCGTGGCCTGGGCGGGACT	TGCGGCCGCA	GGACTGGGGAGTGGCGAGCC	RGLGGTCGRRTGEWRA
435	G	GCCTGGCCTGGGCGGGACTG	TGCGGCCGCA	GACTGGGGAGTGGCGAGCC	RGLGGTVRPTGEWRA
436	G	CGTGGCCTGGGCGGGACTGG	TGCGGCCGCA	ACTGGGGAGTGGCGAGCCCT	RGLGGTAAATGEWRA
437	G	GTGGCCTGGGCGGGACTGGG	TGCGGCCGCA	CTGGGGAGTGGCGAGCCCTC	GLGGTGCGRRTGEWRAL
438	G	TGGCCTGGGCGGGACTGGGG	TGCGGCCGCA	TGGGGAGTGGCGAGCCCTCA	GLGGTVRPHGEWRAL
439	A	GGCCTGGGCGGGACTGGGGA	TGCGGCCGCA	GGGAGTGGCGAGCCCTCAG	GLGGTGDAAAGGEWRAL
440	G	GCCTGGGCGGGACTGGGGAG	TGCGGCCGCA	GGGAGTGGCGAGCCCTCAGA	LGGTGECGRREWRALR
441	T	CCTGGGCGGGACTGGGGAGT	TGCGGCCGCA	GGAGTGGCGAGCCCTCAGAT	LGGTGELRPQEWRALR
442	G	CTGGGCGGGACTGGGGAGTG	TGCGGCCGCA	GAGTGGCGAGCCCTCAGATG	LGGTGECAAAEWRALR
443	G	TGGGCGGGACTGGGGAGTGG	TGCGGCCGCA	AGTGGCGAGCCCTCAGATGC	GGTGEWCRPQWRALRC
444	C	GGGCGGGACTGGGGAGTGGC	TGCGGCCGCA	TGGCGAGCCCTCAGATGCT	GGTGEWLRRKWRALRC
445	G	GGCGGGACTGGGGAGTGGCG	TGCGGCCGCA	TGGCGAGCCCTCAGATGCTG	GGTGEWRAAAWRALRC
446	A	GCGGGACTGGGGAGTGGCGA	TGCGGCCGCA	GCGGAGCCCTCAGATGCTGC	GTGEWRCGRRRALRCC
447	G	CGGACTGGGGAGTGGCGAG	TGCGGCCGCA	GCGAGCCCTCAGATGCTGCA	GTGEWVRPQRALRCC
448	C	GGGACTGGGGAGTGGCGAGC	TGCGGCCGCA	GAGCCCTCAGATGCTGCAT	GTGEWRAAAARALRCC
449	C	GGACTGGGGAGTGGCGAGCC	TGCGGCCGCA	GAGCCCTCAGATGCTGCATA	TGEWRACRRRALRCCI
450	C	GACTGGGGAGTGGCGAGCCC	TGCGGCCGCA	AGCCCTCAGATGCTGCATAT	TGEWRALRPQALRCCI
451	T	ACTGGGGAGTGGCGAGCCCT	TGCGGCCGCA	GCCCTCAGATGCTGCATATA	TGEWRALAAALRCCI
452	C	CTGGGGAGTGGCGAGCCCTC	TGCGGCCGCA	CCTCAGATGCTGCATATAA	GEWRALCGRTLRCCI
453	A	TGGGGAGTGGCGAGCCCTCA	TGCGGCCGCA	CCTCAGATGCTGCATATAAG	GEWRALMRPHLRCCI
454	G	GGGGAGTGGCGAGCCCTCAG	TGCGGCCGCA	CTCAGATGCTGCATATAAGC	GEWRALSAALRCCI
455	A	GGGAGTGGCGAGCCCTCAGA	TGCGGCCGCA	TAGATGCTGCATATAAGCA	EWRALRCGRIRCCI
456	T	GGAGTGGCGAGCCCTCAGAT	TGCGGCCGCA	CAGATGCTGCATATAAGCAG	EWRALRLRPHRCCI
457	G	GAGTGGCGAGCCCTCAGATG	TGCGGCCGCA	AGATGCTGCATATAAGCAGC	EWRALRCAARCCI
458	C	AGTGGCGAGCCCTCAGATGC	TGCGGCCGCA	GATGCTGCATATAAGCAGCT	WRALRCCGRRCCI
459	T	GTGGCGAGCCCTCAGATGCT	TGCGGCCGCA	ATGCTGCATATAAGCAGCTG	WRALRCLRCCI
460	G	TGGCGAGCCCTCAGATGCTG	TGCGGCCGCA	TGCTGCATATAAGCAGCTGC	WRALRCCAAACCI
461	C	GGCGAGCCCTCAGATGCTGC	TGCGGCCGCA	GCTGCATATAAGCAGCTGCT	RALRCCGRSCI
462	A	GCGAGCCCTCAGATGCTGCA	TGCGGCCGCA	CTGCATATAAGCAGCTGCTT	RALRCCMRPHCI
463	T	CGAGCCCTCAGATGCTGCAT	TGCGGCCGCA	TGCATATAAGCAGCTGCTTT	RALRCCIAAACI
464	A	GAGCCCTCAGATGCTGCATA	TGCGGCCGCA	GCATATAAGCAGCTGCTTTT	ALRCCICGRSI
465	T	AGCCCTCAGATGCTGCATAT	TGCGGCCGCA	CATATAAGCAGCTGCTTTTT	ALRCCILRPHI
466	A	GCCTCAGATGCTGCATATA	TGCGGCCGCA	ATATAAGCAGCTGCTTTTTG	ALRCCIAAACI
467	A	CCCTCAGATGCTGCATATAA	TGCGGCCGCA	TATAAGCAGCTGCTTTTTGC	LRCCI
468	G	CCTCAGATGCTGCATATAAG	TGCGGCCGCA	ATAAGCAGCTGCTTTTTGCC	LRCCI
469	C	CTCAGATGCTGCATATAAGC	TGCGGCCGCA	TAAGCAGCTGCTTTTTGCCT	LRCCI
470	A	TCAGATGCTGCATATAAGCA	TGCGGCCGCA	AAGCAGCTGCTTTTTGCCTG	RCCI
471	G	CAGATGCTGCATATAAGCAG	TGCGGCCGCA	AGCAGCTGCTTTTTGCCTGT	RCCI
472	C	AGATGCTGCATATAAGCAGC	TGCGGCCGCA	GCAGCTGCTTTTTGCCTGTA	RCCI
473	T	GATGCTGCATATAAGCAGCT	TGCGGCCGCA	CAGCTGCTTTTTGCCTGTAC	CCI
474	G	ATGCTGCATATAAGCAGCTG	TGCGGCCGCA	AGCTGCTTTTTGCCTGTACT	CCI
475	C	TGCTGCATATAAGCAGCTGC	TGCGGCCGCA	GCTGCTTTTTGCCTGTACTG	CCI
476	T	GCTGCATATAAGCAGCTGCT	TGCGGCCGCA	CTGCTTTTTGCCTGTACTGG	CI
477	T	CTGCATATAAGCAGCTGCTT	TGCGGCCGCA	TGCTTTTTGCCTGTACTGGG	CI
478	T	TGCATATAAGCAGCTGCTTT	TGCGGCCGCA	GCTTTTTGCCTGTACTGGGT	CI
479	T	GCATATAAGCAGCTGCTTTT	TGCGGCCGCA	CTTTTTGCCTGTACTGGGTC	I
480	T	CATATAAGCAGCTGCTTTTT	TGCGGCCGCA	TTTTTGCCTGTACTGGGTCT	I
481	G	ATATAAGCAGCTGCTTTTTG	TGCGGCCGCA	TTTTTGCCTGTACTGGGTCTC	I
482	C	TATAAGCAGCTGCTTTTTGC	TGCGGCCGCA	TTTTTGCCTGTACTGGGTCTCT	I
483	C	ATAAGCAGCTGCTTTTTGCC	TGCGGCCGCA	TTTGCCTGTACTGGGTCTCTC	I
484	T	TAAGCAGCTGCTTTTTGCC	TGCGGCCGCA	TGCTGTACTGGGTCTCTCT	I
485	G	AAGCAGCTGCTTTTTGCCTG	TGCGGCCGCA	GCCTGTACTGGGTCTCTCTG	I
486	T	AGCAGCTGCTTTTTGCCTGT	TGCGGCCGCA	CCTGTACTGGGTCTCTCTGG	I
487	A	GCACTGCTTTTTGCCTGTA	TGCGGCCGCA	CTGTACTGGGTCTCTCTGGT	I
488	C	CAGCTGCTTTTTGCCTGTAC	TGCGGCCGCA	TGTACTGGGTCTCTCTGGTT	I
489	T	AGCTGCTTTTTGCCTGTACT	TGCGGCCGCA	GTACTGGGTCTCTCTGGTTA	I
490	G	GCTGCTTTTTGCCTGTACTG	TGCGGCCGCA	TACTGGGTCTCTCTGGTTAG	I
491	G	CTGCTTTTTGCCTGTACTGG	TGCGGCCGCA	ACTGGGTCTCTCTGGTTAGA	I
492	G	TGCTTTTTGCCTGTACTGGG	TGCGGCCGCA	CTGGGTCTCTCTGGTTAGAC	I
493	T	GCTTTTTGCCTGTACTGGGT	TGCGGCCGCA	TGGGTCTCTCTGGTTAGACC	I
494	C	CTTTTTGCCTGTACTGGGTC	TGCGGCCGCA	GGGTCTCTCTGGTTAGACCA	I
495	T	TTTTTGCCTGTACTGGGTCT	TGCGGCCGCA	GGTCTCTCTGGTTAGACCAG	I
496	C	TTTTGCCTGTACTGGGTCTC	TGCGGCCGCA	GTCCTCTCTGGTTAGACCAGA	I
497	T	TTTGCCTGTACTGGGTCTCT	TGCGGCCGCA	TCTCTCTCTGGTTAGACCAGAT	I
498	C	TTGCCTGTACTGGGTCTCTC	TGCGGCCGCA	CTCTCTGGTTAGACCAGATC	I
499	T	TGCTGTACTGGGTCTCTCTC	TGCGGCCGCA	TCTCTGGTTAGACCAGATCT	I
500	G	GCCTGTACTGGGTCTCTCTG	TGCGGCCGCA	CTCTGGTTAGACCAGATCTG	I
501	G	CCTGTACTGGGTCTCTCTGG	TGCGGCCGCA	TCTGGTTAGACCAGATCTGA	I
502	T	CTGTACTGGGTCTCTCTGGT	TGCGGCCGCA	CTGGTTAGACCAGATCTGAG	I
503	T	TGTACTGGGTCTCTCTGGTT	TGCGGCCGCA	TGGTTAGACCAGATCTGAGC	I
504	A	GTAAGTGGGTCTCTCTGGTTA	TGCGGCCGCA	GTTAGACCAGATCTGAGCC	I
505	G	TACTGGGTCTCTCTGGTTAG	TGCGGCCGCA	GTTAGACCAGATCTGAGCCT	I
506	A	ACTGGGTCTCTCTGGTTAGA	TGCGGCCGCA	TTAGACCAGATCTGAGCCTG	I
507	C	CTGGGTCTCTCTGGTTAGAC	TGCGGCCGCA	TAGACCAGATCTGAGCCTGG	I
508	C	TGGGTCTCTCTGGTTAGACC	TGCGGCCGCA	AGACCAGATCTGAGCCTGGG	I
509	A	GGGTCTCTCTGGTTAGACCA	TGCGGCCGCA	GACCAGATCTGAGCCTGGGA	I
510	G	GGTCTCTCTGGTTAGACCAG	TGCGGCCGCA	ACCAGATCTGAGCCTGGGAG	I

511	A	GTCTCTCTGGTTAGACCAGA	TGCGGCCGCA	CCAGATCTGAGCCTGGGAGC	VSLVRPDAAAPDLSLG
512	T	TCTCTCTGGTTAGACCAGAT	TGCGGCCGCA	CAGATCTGAGCCTGGGAGCT	SLVRPDCGRDLSLGA
513	C	CTCTCTGGTTAGACCAGATC	TGCGGCCGCA	AGATCTGAGCCTGGGAGCTC	SLVRPDLRPQDLSLGA
514	T	TCTCTGGTTAGACCAGATCT	TGCGGCCGCA	GATCTGAGCCTGGGAGCTCT	SLVRPDLAAADLSLGA
515	G	CTCTGGTTAGACCAGATCTG	TGCGGCCGCA	ATCTGAGCCTGGGAGCTCTC	LVRPDLRMRPHLSLGA
516	A	TCTGGTTAGACCAGATCTGA	TGCGGCCGCA	TCTGAGCCTGGGAGCTCTCT	LVRPDLRMRPHLSLGA
517	G	CTGGTTAGACCAGATCTGAG	TGCGGCCGCA	CTGAGCCTGGGAGCTCTCTG	LVRPDLRMRPHLSLGA
518	C	TGGTTAGACCAGATCTGAGC	TGCGGCCGCA	TGAGCCTGGGAGCTCTCTGG	VRPDLRMRPHLSLGA
519	C	GGTTAGACCAGATCTGAGCC	TGCGGCCGCA	GAGCCTGGGAGCTCTCTGGC	VRPDLRMRPHLSLGA
520	T	GTTAGACCAGATCTGAGCCT	TGCGGCCGCA	AGCCTGGGAGCTCTCTGGCT	VRPDLRMRPHLSLGA
521	G	TTAGACCAGATCTGAGCCTG	TGCGGCCGCA	GCCTGGGAGCTCTCTGGCTA	RPDLSLGRMSLALW
522	G	TAGACCAGATCTGAGCCTGG	TGCGGCCGCA	CCTGGGAGCTCTCTGGCTAA	RPDLSLGRMSLALW
523	G	AGACCAGATCTGAGCCTGGG	TGCGGCCGCA	CTGGGAGCTCTCTGGCTAAC	RPDLSLGRMSLALW
524	A	GACCAGATCTGAGCCTGGGA	TGCGGCCGCA	TGGGAGCTCTCTGGCTAACT	PDLSLGVRMSLALW
525	G	ACCAGATCTGAGCCTGGGAG	TGCGGCCGCA	GGGAGCTCTCTGGCTAACTA	PDLSLGVRMSLALW
526	C	CCAGATCTGAGCCTGGGAGC	TGCGGCCGCA	GGAGCTCTCTGGCTAACTAG	PDLSLGAAAALWLT
527	T	CAGATCTGAGCCTGGGAGCT	TGCGGCCGCA	GAGCTCTCTGGCTAACTAGG	DLSLGRMSLALWTR
528	C	AGATCTGAGCCTGGGAGCTC	TGCGGCCGCA	AGCTCTCTGGCTAACTAGGG	DLSLGRMSLALWTR
529	T	GATCTGAGCCTGGGAGCTCT	TGCGGCCGCA	GCTCTCTGGCTAACTAGGGA	DLSLGRMSLALWTR
530	C	ATCTGAGCCTGGGAGCTCTC	TGCGGCCGCA	CTCTCTGGCTAACTAGGGA	LSLGRMSLALWTR
531	T	TCTGAGCCTGGGAGCTCTCT	TGCGGCCGCA	TCTCTGGCTAACTAGGGAAC	LSLGRMSLALWTR
532	G	CTGAGCCTGGGAGCTCTCTG	TGCGGCCGCA	CTCTGGCTAACTAGGGAACC	LSLGRMSLALWTR
533	G	TGAGCCTGGGAGCTCTCTGG	TGCGGCCGCA	TCTGGCTAACTAGGGAACCC	SLGALWLRPHWLTR
534	C	GAGCCTGGGAGCTCTCTGGC	TGCGGCCGCA	CTGGCTAACTAGGGAACCCA	SLGALWLRPHWLTR
535	T	AGCCTGGGAGCTCTCTGGCT	TGCGGCCGCA	TGGCTAACTAGGGAACCCAC	SLGALWLRPHWLTR
536	A	GCCTGGGAGCTCTCTGGCTA	TGCGGCCGCA	GGCTAACTAGGGAACCCACT	LGALWLRMRPQLTREP
537	A	CCTGGGAGCTCTCTGGCTAA	TGCGGCCGCA	GCTAACTAGGGAACCCACTG	LGALWLRMRPQLTREP
538	C	CTGGGAGCTCTCTGGCTAAC	TGCGGCCGCA	CTAACTAGGGAACCCACTGC	LGALWLRMRPQLTREP
539	T	TGGGAGCTCTCTGGCTAACT	TGCGGCCGCA	TAAGCTAGGGAACCCACTGCT	GALWLRMRPQLTREP
540	A	GGGAGCTCTCTGGCTAACTA	TGCGGCCGCA	AACTAGGGAACCCACTGCTT	GALWLRMRPQLTREP
541	G	GGAGCTCTCTGGCTAACTAG	TGCGGCCGCA	ACTAGGGAACCCACTGCTTA	GALWLRMRPQLTREP
542	G	GAGCTCTCTGGCTAACTAGG	TGCGGCCGCA	CTAGGGAACCCACTGCTTAA	ALWLRMRPQLTREP
543	G	AGCTCTCTGGCTAACTAGGG	TGCGGCCGCA	TAGGGAACCCACTGCTTAA	ALWLRMRPQLTREP
544	A	GCTCTCTGGCTAACTAGGGA	TGCGGCCGCA	AGGGAACCCACTGCTTAA	ALWLRMRPQLTREP
545	A	CTCTCTGGCTAACTAGGGA	TGCGGCCGCA	GGGAACCCACTGCTTAA	LWLRMRPQLTREP
546	C	TCTCTGGCTAACTAGGGA	TGCGGCCGCA	GAACCCACTGCTTAA	LWLRMRPQLTREP
547	C	TCTGGCTAACTAGGGA	TGCGGCCGCA	GAACCCACTGCTTAA	LWLRMRPQLTREP
548	C	TCTGGCTAACTAGGGA	TGCGGCCGCA	AACCCACTGCTTAA	LWLRMRPQLTREP
549	A	CTGGCTAACTAGGGA	TGCGGCCGCA	ACCCACTGCTTAA	LWLRMRPQLTREP
550	C	TGGCTAACTAGGGA	TGCGGCCGCA	CCCCTGCTTAA	LWLRMRPQLTREP
551	T	GGCTAACTAGGGA	TGCGGCCGCA	CCCTGCTTAA	LWLRMRPQLTREP
552	G	GCTAACTAGGGA	TGCGGCCGCA	CACTGCTTAA	LWLRMRPQLTREP
553	C	CTAACTAGGGA	TGCGGCCGCA	ACTGCTTAA	LWLRMRPQLTREP
554	T	TAAGCTAGGGA	TGCGGCCGCA	CTGCTTAA	LWLRMRPQLTREP
555	T	AACTAGGGA	TGCGGCCGCA	TGCTTAA	LWLRMRPQLTREP
556	A	ACTAGGGA	TGCGGCCGCA	GCTTAA	LWLRMRPQLTREP
557	A	CTAGGGA	TGCGGCCGCA	CTTAA	LWLRMRPQLTREP
558	G	TAGGGA	TGCGGCCGCA	TTAA	LWLRMRPQLTREP
559	C	AGGGA	TGCGGCCGCA	TAAG	LWLRMRPQLTREP
560	C	GGGA	TGCGGCCGCA	AAG	LWLRMRPQLTREP
561	T	GGA	TGCGGCCGCA	AG	LWLRMRPQLTREP
562	C	GA	TGCGGCCGCA	AG	LWLRMRPQLTREP
563	A	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
564	A	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
565	T	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
566	A	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
567	A	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
568	A	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
569	G	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
570	C	T	TGCGGCCGCA	AG	LWLRMRPQLTREP
571	T	G	TGCGGCCGCA	AG	LWLRMRPQLTREP
572	T	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
573	G	T	TGCGGCCGCA	AG	LWLRMRPQLTREP
574	C	T	TGCGGCCGCA	AG	LWLRMRPQLTREP
575	C	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
576	T	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
577	T	G	TGCGGCCGCA	AG	LWLRMRPQLTREP
578	G	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
579	A	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
580	G	T	TGCGGCCGCA	AG	LWLRMRPQLTREP
581	G	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
582	G	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
583	A	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
584	G	T	TGCGGCCGCA	AG	LWLRMRPQLTREP
585	T	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
586	G	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
587	C	A	TGCGGCCGCA	AG	LWLRMRPQLTREP
588	T	G	TGCGGCCGCA	AG	LWLRMRPQLTREP
589	T	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
590	C	T	TGCGGCCGCA	AG	LWLRMRPQLTREP
591	A	T	TGCGGCCGCA	AG	LWLRMRPQLTREP
592	A	G	TGCGGCCGCA	AG	LWLRMRPQLTREP
593	G	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
594	T	C	TGCGGCCGCA	AG	LWLRMRPQLTREP
595	A	T	TGCGGCCGCA	AG	LWLRMRPQLTREP
596	G	T	TGCGGCCGCA	AG	LWLRMRPQLTREP

597	T	GAGGGAGTGCTTCAAGTAGT	TGCGGCCGCA	GTAGTGTGTGCCCGTCTGTT	RECFK*LRPQ*CVVPC
598	G	AGGGAGTGCTTCAAGTAGTG	TGCGGCCGCA	TAGTGTGTGCCCGTCTGTTG	RECFK*CAAA*CVVPC
599	T	GGGAGTGCTTCAAGTAGTGT	TGCGGCCGCA	AGTGTGTGCCCGTCTGTTGT	ECFK*CCGRKCVVPC
600	G	GGAGTGCTTCAAGTAGTGTG	TGCGGCCGCA	GTGTGTGCCCGTCTGTTGTG	ECFK*CVRPQCVVPC
601	T	GAGTGCTTCAAGTAGTGTGT	TGCGGCCGCA	TGTGTGCCCGTCTGTTGTGA	ECFK*CVAAACVPC
602	Q	AGTGCTTCAAGTAGTGTGTG	TGCGGCCGCA	GTGTGCCCGTCTGTTGTGAC	CFK*CVCGRSPVPCD
603	C	GTGCTTCAAGTAGTGTGTGC	TGCGGCCGCA	TGTGCCCGTCTGTTGTGACT	CFK*CVLRPHVPCD
604	C	TGCTTCAAGTAGTGTGTGCC	TGCGGCCGCA	GTGCCCGTCTGTTGTGACTC	CFK*CVPAAVVPCD
605	C	GCTTCAAGTAGTGTGTGCC	TGCGGCCGCA	TGCCCGTCTGTTGTGACTCT	FK*CVPCGRMPVCCDS
606	G	CTTCAAGTAGTGTGTGCCCG	TGCGGCCGCA	GCCCGTCTGTTGTGACTCTG	FK*CVVPRQPVCCDS
607	T	TCAAGTAGTGTGTGCCCGT	TGCGGCCGCA	CCCGTCTGTTGTGACTCTGG	FK*CVVPAAVVPCD
608	C	TCAAGTAGTGTGTGCCCGTC	TGCGGCCGCA	CCGCTGTTGTGACTCTGGT	K*CVPCGRMVCCDSG
609	T	CAAGTAGTGTGTGCCCGTCT	TGCGGCCGCA	CGTCTGTTGTGACTCTGGTA	K*CVPLRPHVCCDSG
610	G	AAGTAGTGTGTGCCCGTCTG	TGCGGCCGCA	GTCTGTTGTGACTCTGGTAA	K*CVPVCAAAVCCDSG
611	T	AGTAGTGTGTGCCCGTCTGT	TGCGGCCGCA	TCTGTTGTGACTCTGGTAACT	*CVPVCAAAVCCDSGN
612	T	GTAGTGTGTGCCCGTCTGTT	TGCGGCCGCA	CTGTTGTGACTCTGGTAACT	*CVPVCLRPHCCDSGN
613	G	TAGTGTGTGCCCGTCTGTTG	TGCGGCCGCA	TGTTGTGACTCTGGTAACTA	*CVPVCAAAVCCDSGN
614	T	AGTGTGTGCCCGTCTGTTGT	TGCGGCCGCA	GTGTTGTGACTCTGGTAACTAG	CVPVCCVRRHCCDSGN*
615	G	GTGTGTGCCCGTCTGTTGTG	TGCGGCCGCA	TTGTGACTCTGGTAACTAGA	CVPVCCVRRHCCDSGN*
616	A	TGTGTGCCCGTCTGTTGTGA	TGCGGCCGCA	TGTGACTCTGGTAACTAGAG	CVPVCCDAAVCCDSGN*
617	C	GTGTGCCCGTCTGTTGTGAC	TGCGGCCGCA	GTGACTCTGGTAACTAGAGA	VPVCCDGRSDSGN*R
618	T	TGTGCCCGTCTGTTGTGACT	TGCGGCCGCA	TGACTCTGGTAACTAGAGAT	VPVCCDLRPHDSGN*R
619	C	GTGCCCGTCTGTTGTGACTC	TGCGGCCGCA	GACTCTGGTAACTAGAGATC	VPVCCDGRSDSGN*R
620	T	TGCCCGTCTGTTGTGACTCT	TGCGGCCGCA	ACTCTGGTAACTAGAGATCC	PVCCDSCGRNSGN*RS
621	G	GCCCGTCTGTTGTGACTCTG	TGCGGCCGCA	CTCTGGTAACTAGAGATCCC	PVCCDSDVRPHSGN*RS
622	G	CCCGTCTGTTGTGACTCTGG	TGCGGCCGCA	TCTGGTAACTAGAGATCCCT	PVCCDSGAAASGN*RS
623	T	CCGTCTGTTGTGACTCTGGT	TGCGGCCGCA	CTGGTAACTAGAGATCCCTC	VCCDSGCGRTGN*RSL
624	A	CGTCTGTTGTGACTCTGGTA	TGCGGCCGCA	TGGTAACTAGAGATCCCTCA	VCCDSGMRPHGN*RSL
625	A	GTCTGTTGTGACTCTGGTAA	TGCGGCCGCA	GGTAACTAGAGATCCCTCAG	VCCDSGNAAAGN*RSL
626	C	TCTGTTGTGACTCTGGTAACT	TGCGGCCGCA	GTAAGTAGAGATCCCTCAGA	CCDSNGCRSN*RSLR
627	T	CTGTTGTGACTCTGGTAACT	TGCGGCCGCA	TAAGTAGAGATCCCTCAGAC	CCDSGNLRPHN*RSLR
628	A	TGTTGTGACTCTGGTAACTA	TGCGGCCGCA	AACTAGAGATCCCTCAGACC	CCDSGNAAAN*RSLR
629	G	GTGTTGACTCTGGTAACTAG	TGCGGCCGCA	ACTAGAGATCCCTCAGACCC	CDSGN*CGRN*RSLRP
630	A	TTGTGACTCTGGTAACTAGA	TGCGGCCGCA	CTAGAGATCCCTCAGACCCCT	CDSGN*MRPH*RSLRP
631	G	TGTGACTCTGGTAACTAGAG	TGCGGCCGCA	TAGAGATCCCTCAGACCCCTT	CDSGN*SAAA*RSLRP
632	A	GTGACTCTGGTAACTAGAGA	TGCGGCCGCA	AGAGATCCCTCAGACCCCTTT	DSGN*RCGRKRSRPF
633	T	TGACTCTGGTAACTAGAGAT	TGCGGCCGCA	GAGATCCCTCAGACCCCTTTT	DSGN*RLRQRSLRPF
634	C	GACTCTGGTAACTAGAGATC	TGCGGCCGCA	AGATCCCTCAGACCCCTTTTA	DSGN*RSAAARSLRPF
635	C	ACTCTGGTAACTAGAGATCC	TGCGGCCGCA	GATCCCTCAGACCCCTTTTAG	SGN*RCGRKRSRPF*
636	C	CTCTGGTAACTAGAGATCCC	TGCGGCCGCA	ATCCCTCAGACCCCTTTTAGT	SGN*RSLRQSLRPF*
637	T	TCTGGTAACTAGAGATCCCT	TGCGGCCGCA	TCCCTCAGACCCCTTTTAGTC	SGN*RSLAAASLRF*
638	C	CTGGTAACTAGAGATCCCTC	TGCGGCCGCA	CCCTCAGACCCCTTTTAGTCA	GN*RSLCGRTLRF*S
639	A	TGGTAACTAGAGATCCCTCA	TGCGGCCGCA	CCTCAGACCCCTTTTAGTCAG	GN*RSLMRPHLRF*S
640	G	GGTAACTAGAGATCCCTCAG	TGCGGCCGCA	CTCAGACCCCTTTTAGTCAGT	GN*RSLAAALRF*S
641	A	GTAAGTAGAGATCCCTCAGA	TGCGGCCGCA	TCAGACCCCTTTTAGTCAGTG	N*RSLRCAARLRF*SV
642	C	TAAGTAGAGATCCCTCAGAC	TGCGGCCGCA	CAGACCCCTTTTAGTCAGTGT	N*RSLRLRPHRPF*SV
643	C	AACTAGAGATCCCTCAGACC	TGCGGCCGCA	GACCCCTTTTAGTCAGTGTG	N*RSLRPAARPF*SV
644	C	ACTAGAGATCCCTCAGACCC	TGCGGCCGCA	GACCCCTTTTAGTCAGTGTGG	*RSLRPGRRPF*SVW
645	T	CTAGAGATCCCTCAGACCCCT	TGCGGCCGCA	ACCCCTTTTAGTCAGTGTGGA	*RSLRPLRQPF*SVW
646	T	TAGAGATCCCTCAGACCCCTT	TGCGGCCGCA	CCCTTTTAGTCAGTGTGAAA	*RSLRPFAAAPF*SVW
647	T	AGAGATCCCTCAGACCCCTTT	TGCGGCCGCA	CCTTTTAGTCAGTGTGAAAA	RSLRPFGRFT*SVWK
648	T	GAGATCCCTCAGACCCCTTTT	TGCGGCCGCA	CTTTTAGTCAGTGTGAAAAA	RSLRPFLLRPF*SVWK
649	A	AGATCCCTCAGACCCCTTTTA	TGCGGCCGCA	TTTTAGTCAGTGTGAAAAAT	RSLRPFYAAF*SVWK
650	G	GATCCCTCAGACCCCTTTTAG	TGCGGCCGCA	TTTAGTCAGTGTGAAAAATC	SLRPF*CGRI*SVWKI
651	T	ATCCCTCAGACCCCTTTTAGT	TGCGGCCGCA	TTAGTCAGTGTGAAAAATCT	SLRPF*LRPH*SVWKI
652	C	TCCCTCAGACCCCTTTTAGTC	TGCGGCCGCA	TAGTCAGTGTGAAAAATCTC	SLRPF*SAAA*SVWKI
653	A	CCCTCAGACCCCTTTTAGTCA	TGCGGCCGCA	AGTCAGTGTGAAAAATCTCT	LRPF*SCGRSVWKIS
654	G	CCTCAGACCCCTTTTAGTCAG	TGCGGCCGCA	GTCAGTGTGAAAAATCTCTA	LRPF*SVRPQSVWKIS
655	T	CTCAGACCCCTTTTAGTCAGT	TGCGGCCGCA	TCAGTGTGAAAAATCTCTAG	LRPF*SVAAASVWKIS
656	G	TCAGACCCCTTTTAGTCAGTG	TGCGGCCGCA	CAGTGTGAAAAATCTCTAGC	RPF*SVCGRTVWKISS
657	T	CAGACCCCTTTTAGTCAGTGT	TGCGGCCGCA	AGTGTGAAAAATCTCTAGCA	RPF*SVLRPQVWKISS
658	G	AGACCCCTTTTAGTCAGTGTG	TGCGGCCGCA	GTGTGAAAAATCTCTAGCAG	RPF*SVCAAVWKISS
659	G	GACCCCTTTTAGTCAGTGTGG	TGCGGCCGCA	TGTGAAAAATCTCTAGCAGT	PF*SVVCGRMWKISSS
660	A	ACCCCTTTTAGTCAGTGTGGA	TGCGGCCGCA	GTGAAAAATCTCTAGCAGTG	PF*SVWMRPQWKISSS
661	A	CCCTTTTAGTCAGTGTGAAA	TGCGGCCGCA	TGAAAAATCTCTAGCAGTGG	PF*SVWNAAAWKISSS
662	A	CCTTTTAGTCAGTGTGAAAA	TGCGGCCGCA	GGAAAAATCTCTAGCAGTGGC	F*SVWKCGRRKISSSG
663	A	CTTTTAGTCAGTGTGAAAAA	TGCGGCCGCA	GAAAAATCTCTAGCAGTGGCG	F*SVWKMRRPKISSSG
664	T	TTTTAGTCAGTGTGAAAAAT	TGCGGCCGCA	AAAAATCTCTAGCAGTGGCGC	F*SVWKIAAAKISSSG
665	C	TTTAGTCAGTGTGAAAAATC	TGCGGCCGCA	AAATCTCTAGCAGTGGCGCC	*SVWKICGRKISSSGA
666	T	TTAGTCAGTGTGAAAAATCT	TGCGGCCGCA	AATCTCTAGCAGTGGCGCCC	*SVWKILRPQISSSGA
667	C	TAGTCAGTGTGAAAAATCTC	TGCGGCCGCA	ATCTCTAGCAGTGGCGCCCG	*SVWKISAAIASSSGA
668	T	AGTCAGTGTGAAAAATCTCT	TGCGGCCGCA	TCTCTAGCAGTGGCGCCCGA	SVWKISCGRISSSGAR
669	A	GTAGTCAGTGTGAAAAATCTCTA	TGCGGCCGCA	CTCTAGCAGTGGCGCCCGAA	SVWKISMRPHSSSGAR
670	G	TCAGTGTGAAAAATCTCTAG	TGCGGCCGCA	TCTAGCAGTGGCGCCCGAAC	SVWKISAAASSSGAR
671	C	CAGTGTGAAAAATCTCTAGC	TGCGGCCGCA	CTAGCAGTGGCGCCCGAAC	VWKISSCGRTSSGART
672	A	AGTGTGAAAAATCTCTAGCA	TGCGGCCGCA	TAGCAGTGGCGCCCGAACAG	VWKISSMRPHSSSGART
673	G	GTGTGAAAAATCTCTAGCAG	TGCGGCCGCA	AGCAGTGGCGCCCGAACAGG	VWKISSAAASSSGART
674	T	TGTGTGAAAAATCTCTAGCAGT	TGCGGCCGCA	GACAGTGGCGCCCGAACAGGG	WKISSSCGRSSGARTG
675	G	GTGAAAAATCTCTAGCAGTG	TGCGGCCGCA	CAGTGGCGCCCGAACAGGGA	WKISSSVRPHSSGARTG
676	G	TGAAAAATCTCTAGCAGTGG	TGCGGCCGCA	AGTGGCGCCCGAACAGGGAC	WKISSSGAAASGARTG
677	C	GGAAAAATCTCTAGCAGTGGC	TGCGGCCGCA	GTGGCGCCCGAACAGGGACT	KISSSGCGRSGARTGT
678	G	GAAAAATCTCTAGCAGTGGCG	TGCGGCCGCA	TGGCGCCCGAACAGGGACTT	KISSSGVRPHSSGARTGT
679	C	AAAAATCTCTAGCAGTGGCGC	TGCGGCCGCA	GGCGCCCGAACAGGGACTTG	KISSSGAAAGARTGT
680	C	AAATCTCTAGCAGTGGCGCC	TGCGGCCGCA	CGCGCCCGAACAGGGACTTGA	ISSSGAGARSARTGT*
681	C	AAATCTCTAGCAGTGGCGCC	TGCGGCCGCA	CGCGCCCGAACAGGGACTTGA	ISSSGALRPHARTGT*
682	G	ATCTCTAGCAGTGGCGCCCG	TGCGGCCGCA	GCGCGAACAGGGACTTGAAA	ISSSGARAAAARTGT*

683	A	TCTCTAGCAGTGGCGCCGA	TGCGGCCGA	CCCGAACAGGGACTTGAAG	SSSGARCGRTRTGT*K
684	A	CTCTAGCAGTGGCGCCGAA	TGCGGCCGA	CCGAACAGGGACTTGAAGC	SSSGARMRPHRTGT*K
685	C	TCTAGCAGTGGCGCCGAAC	TGCGGCCGA	CGAACAGGGACTTGAAGCG	SSSGARTAAARTGT*K
686	A	CTAGCAGTGGCGCCCGAAC	TGCGGCCGA	GAACAGGGACTTGAAGCGA	SSGARTCGRRTGT*KR
687	G	TAGCAGTGGCGCCCGAACAG	TGCGGCCGA	AACAGGGACTTGAAGCGAA	SSGARTVRRPQTGT*KR
688	G	AGCAGTGGCGCCCGAACAGG	TGCGGCCGA	ACAGGGACTTGAAGCGAAA	SSGARTGAAATGT*KR
689	G	GCAGTGGCGCCCGAACAGGG	TGCGGCCGA	CAGGGACTTGAAGCGAAAG	SGARTGCGRRT*KRK
690	A	CAGTGGCGCCCGAACAGGGGA	TGCGGCCGA	AGGGACTTGAAGCGAAAGT	SGARTGMRPQGT*KRK
691	C	AGTGGCGCCCGAACAGGGAC	TGCGGCCGA	GGGACTTGAAGCGGAAAGT	SGARTGTAAGT*KRK
692	T	GTGGCGCCCGAACAGGGACT	TGCGGCCGA	GGACTTGAAGCGGAAAGTAA	GARTGTCGRRT*KRK*
693	T	TGGCGCCCGAACAGGGACTT	TGCGGCCGA	GACTTGAAGCGGAAAGTAAA	GARTGTLRPQT*KRK*
694	G	GGCGCCCGAACAGGGACTTG	TGCGGCCGA	ACTTGAAGCGGAAAGTAAAG	GARTGTCAAAAT*KRK*
695	A	GCGCCCGAACAGGGACTTGA	TGCGGCCGA	CTTGAAGCGGAAAGTAAAGC	ARTGT*CGRT*KRK*S
696	A	CGCCCGAACAGGGACTTGA	TGCGGCCGA	TTGAAGCGGAAAGTAAAGCC	ARTGT*MRPH*KRK*S
697	A	GCCCGAACAGGGACTTGA	TGCGGCCGA	TGAAGCGGAAAGTAAAGCCA	ARTGT*NAAA*KRK*S
698	G	CCCGAACAGGGACTTGAAG	TGCGGCCGA	GAAAGCGGAAAGTAAAGCCAG	RTGT*KGRRKRK*SQ
699	C	CCGAACAGGGACTTGAAGC	TGCGGCCGA	AAAGCGGAAAGTAAAGCCAGA	RTGT*KLRPQKRK*SQ
700	G	CGAACAGGGACTTGAAGCG	TGCGGCCGA	AAGCGGAAAGTAAAGCCAGAG	RTGT*KRAAAKRK*SQ
701	A	GAACAGGGACTTGAAGCGA	TGCGGCCGA	AGCGGAAAGTAAAGCCAGAGG	TGT*KRCGRKRK*SQ
702	A	AACAGGGACTTGAAGCGAA	TGCGGCCGA	GCGGAAAGTAAAGCCAGAGGA	TGT*KRMRPQRK*SQ
703	A	ACAGGGACTTGAAGCGAAA	TGCGGCCGA	CGAAAGTAAAGCCAGAGGAG	TGT*KRNAARK*SQ
704	G	CAGGGACTTGAAGCGAAA	TGCGGCCGA	GAAAGTAAAGCCAGAGGAGA	GT*KRKCGRKRK*SQRR
705	T	AGGGACTTGAAGCGAAA	TGCGGCCGA	AAAGTAAAGCCAGAGGAGAT	GT*KRKLRPQK*SQRR
706	A	GGGACTTGAAGCGAAA	TGCGGCCGA	AAGTAAAGCCAGAGGAGATC	GT*KRKYAAAK*SQRR
707	A	GGACTTGAAGCGAAA	TGCGGCCGA	AGTAAAGCCAGAGGAGATCT	T*KRK*CGRT*SQRRS
708	A	GACTTGAAGCGAAA	TGCGGCCGA	GTAAGCCAGAGGAGATCTC	T*KRK*MRPQ*SQRRS
709	G	ACTTGAAGCGAAA	TGCGGCCGA	TAAAGCCAGAGGAGATCTCT	T*KRK*SAAA*SQRRS
710	C	CTTGAAGCGAAA	TGCGGCCGA	AAAGCCAGAGGAGATCTCTC	*KRK*SCGRKSQRRSL
711	C	TTGAAGCGAAA	TGCGGCCGA	AAGCCAGAGGAGATCTCTCG	*KRK*SLRPQSQRRSL
712	A	TGAAAGCGAAA	TGCGGCCGA	AGCCAGAGGAGATCTCTCGA	*KRK*SHAAASQRRSL
713	G	GAAAGCGAAA	TGCGGCCGA	GCCAGAGGAGATCTCTCGAC	KRK*SQCRSQRRLD
714	A	AAAGCGAAA	TGCGGCCGA	CCAGAGGAGATCTCTCGACG	KRK*SQMRPHQRRSLD
715	G	AAGCGAAA	TGCGGCCGA	CAGAGGAGATCTCTCGACG	KRK*SQSAAAQRRSLD
716	G	AGCGAAA	TGCGGCCGA	AGAGGAGATCTCTCGACGCA	RK*SQRCGRKRRLDA
717	A	GCGAAA	TGCGGCCGA	GAGGAGATCTCTCGACGCG	RK*SQRMRPQRRSLDA
718	G	CGAAA	TGCGGCCGA	AGGAGATCTCTCGACGCGAG	RK*SQRSAAARRSLDA
719	A	GAAAGTAAAGCGAAGGAGA	TGCGGCCGA	GGAGATCTCTCGACGCGAGGA	K*SQRRCGRRRLDAG
720	T	AAAGTAAAGCGAAGGAGAT	TGCGGCCGA	GAGATCTCTCGACGCGAGGAC	K*SQRRLPQRSLDAG
721	C	AAGTAAAGCGAAGGAGATC	TGCGGCCGA	GATCTCTCTCGACGCGAGACT	K*SQRRSAAARRSLDAG
722	T	AGTAAAGCGAAGGAGATCT	TGCGGCCGA	GATCTCTCTCGACGCGAGACTC	*SQRRSCGRRSLDAGL
723	C	GTAAGCGAAGGAGATCTCT	TGCGGCCGA	ATCTCTCTCGACGCGAGACTCG	*SQRRSLRPQSLDAGL
724	T	TAAAGCCAGAGGAGATCTCT	TGCGGCCGA	TCTCTCGACGCGAGGACTCGG	*SQRRSLAAASLDAGL
725	C	AAAGCCAGAGGAGATCTCT	TGCGGCCGA	CTCTCGACGCGAGGACTCGGC	SQRRSLCGRSLDAGLG
726	G	AAGCCAGAGGAGATCTCTCG	TGCGGCCGA	TCTCGACGCGAGGACTCGGT	SQRRSLVPHLDAGLG
727	A	AGCCAGAGGAGATCTCTCGA	TGCGGCCGA	CTCGACGCGAGGACTCGGCTT	SQRRSLDAAALDAGLG
728	C	GCCAGAGGAGATCTCTCGAC	TGCGGCCGA	TCGACGCGAGGACTCGGCTTG	QRRSLDCGRIDAGLGL
729	G	CCAGAGGAGATCTCTCGACG	TGCGGCCGA	GACGCGAGGACTCGGCTTGC	QRRSLDVRPHDAGLGL
730	C	CAGAGGAGATCTCTCGACGC	TGCGGCCGA	GACGCGAGGACTCGGCTTGCT	QRRSLDAAAADAGLGL
731	A	AGAGGAGATCTCTCGACGCA	TGCGGCCGA	ACGCGAGGACTCGGCTTGCTG	RRSLDACGRNAGLGLL
732	G	GAGGAGATCTCTCGACGCG	TGCGGCCGA	GCGAGGACTCGGCTTGCTGA	RRSLDAVRPHAGLGLL
733	G	AGGAGATCTCTCGACGCGAG	TGCGGCCGA	GCGAGGACTCGGCTTGCTGAA	RRSLDAGAAAAGLGLL
734	A	GGAGATCTCTCGACGCGAGGA	TGCGGCCGA	CAGGACTCGGCTTGCTGAAG	RSLDAGCGRTGLGLLK
735	C	GAGATCTCTCGACGCGAGGAC	TGCGGCCGA	AGGACTCGGCTTGCTGAAGC	RSLDAGLRPQGLGLLK
736	T	AGATCTCTCGACGCGAGGACT	TGCGGCCGA	GGACTCGGCTTGCTGAAGCG	RSLDAGLAAAGLGLLK
737	C	GATCTCTCGACGCGAGGACTC	TGCGGCCGA	GACTCGGCTTGCTGAAGCGC	SLDAGLCRRRGLLKR
738	G	ATCTCTCGACGCGAGGACTCG	TGCGGCCGA	ACTCGGCTTGCTGAAGCGCG	SLDAGLVRPQLGLLKR
739	G	TCTCTCGACGCGAGGACTCGG	TGCGGCCGA	CTCGGCTTGCTGAAGCGCGC	SLDAGLGAALGLLKR
740	C	CTCTCGACGCGAGGACTCGGC	TGCGGCCGA	TCGGCTTGCTGAAGCGCGCA	LDAGLGCRRRGLLKR
741	T	TCTCGACGCGAGGACTCGGCT	TGCGGCCGA	CGGCTTGCTGAAGCGCGCAC	LDAGLGLRPHGLLKR
742	T	CTCGACGCGAGGACTCGGCTT	TGCGGCCGA	GGCTTGCTGAAGCGCGCACG	LDAGLGLFAAAGLKR
743	G	TCGACGCGAGGACTCGGCTTG	TGCGGCCGA	GCTTGCTGAAGCGCGCACGG	DAGLGLCGRSLLKRAR
744	C	CGACGCGAGGACTCGGCTTGC	TGCGGCCGA	CTTGCTGAAGCGCGCACGGC	DAGLGLLRPHLLKRAR
745	T	GACGCGAGGACTCGGCTTGT	TGCGGCCGA	TTGCTGAAGCGCGCACGGCA	DAGLGLLAAALLKRAR
746	G	ACGCGAGGACTCGGCTTGTG	TGCGGCCGA	TGCTGAAGCGCGCACGGCAA	AGLGLLRCRMLKRARQ
747	A	CGCAGGACTCGGCTTGTGA	TGCGGCCGA	GCTGAAGCGCGCACGGCAAG	AGLGLLMRPQLKRARQ
748	A	GCAGGACTCGGCTTGTGAA	TGCGGCCGA	CTGAAGCGCGCACGGCAAGA	AGLGLLNAALKRARQ
749	G	CAGGACTCGGCTTGTGAAG	TGCGGCCGA	TGAAGCGCGCACGGCAAGAG	GLGLLKCGRMKRARQE
750	C	AGGACTCGGCTTGTGAAGC	TGCGGCCGA	GAAAGCGCGCACGGCAAGAGG	GLGLLKLPRQKRARQE
751	G	GGACTCGGCTTGTGAAGCG	TGCGGCCGA	AAGCGCGCACGGCAAGAGGC	GLGLLKRAAAKRARQE
752	C	GACTCGGCTTGTGAAGCGC	TGCGGCCGA	AGCGCGCACGGCAAGAGGGC	LGLLKRCGRKRARQE
753	G	ACTCGGCTTGTGAAGCGCG	TGCGGCCGA	GCGCGCACGGCAAGAGGGCGA	LGLLKRVPRQARQE
754	C	CTCGGCTTGTGAAGCGCGC	TGCGGCCGA	CGCGCACGGCAAGAGGGCGAG	LGLLKRAAAARARQE
755	A	TCGGCTTGTGAAGCGCGCA	TGCGGCCGA	GCGCACGGCAAGAGGGCGAGG	GLLKRCGRSARARQE
756	C	CGGCTTGTGAAGCGCGCAC	TGCGGCCGA	GCGCACGGCAAGAGGGCGAGGG	GLLKRALRPHARARQE
757	G	GGCTTGTGAAGCGCGCACG	TGCGGCCGA	GACGGCAAGAGGGCGAGGGG	GLLKRARAAARARQE
758	G	GCTTGTGAAGCGCGCACGG	TGCGGCCGA	CACGGCAAGAGGGCGAGGGGC	LLKRARCRRRQEAR
759	C	CTTGTGAAGCGCGCACGGC	TGCGGCCGA	ACGGCAAGAGGGCGAGGGGCG	LLKRARLRPQRQEAR
760	A	TTGCTGAAGCGCGCACGGCA	TGCGGCCGA	CGGCAAGAGGGCGAGGGGCGG	LLKRARHAAARQEAR
761	A	TGCTGAAGCGCGCACGGCAA	TGCGGCCGA	GCGCAAGAGGGCGAGGGGCGGC	LKRARCRRRQEAR
762	G	GCTGAAGCGCGCACGGCAAG	TGCGGCCGA	GCAAGAGGGCGAGGGGCGGCG	LKRARQVRRPQEAR
763	A	CTGAAGCGCGCACGGCAAGA	TGCGGCCGA	CAAGAGGGCGAGGGGCGGCGA	LKRARQDAAQEAR
764	G	TGAAGCGCGCACGGCAAGAG	TGCGGCCGA	AAGAGGGCGAGGGGCGGCGAC	KRARQECRKEAR
765	G	GAAGCGCGCACGGCAAGAGG	TGCGGCCGA	AGAGGGCGAGGGGCGGCGACT	KRARQEVRRQEAR
766	C	AAGCGCGCACGGCAAGAGGC	TGCGGCCGA	GAGGCGAGGGGCGGCGACTG	KRARQEAAAQEAR
767	G	AGCGCGCACGGCAAGAGGCG	TGCGGCCGA	AGGCGAGGGGCGGCGACTGG	RARQEAACGRKARGGDW
768	A	GCGCGCACGGCAAGAGGCGA	TGCGGCCGA	GCGGAGGGGCGGCGACTGGT	RARQEAAMRPQARGGDW

769	G	CGCGCACGGCAAGAGGCGAG	TGCGGCCGCA	CGAGGGGGCGGCGACTGGTG	RARQEAASAAAARGDW
770	G	CGCGCACGGCAAGAGGCGAGG	TGCGGCCGCA	CGAGGGGGCGGCGACTGGTGA	ARQEARCGRTRGGDW*
771	G	CGCGCACGGCAAGAGGCGAGGG	TGCGGCCGCA	GAGGGGGCGGCGACTGGTGAG	ARQEARVRPQRGGDW*
772	G	GCACGGCAAGAGGCGAGGGG	TGCGGCCGCA	AGGGGGCGGCGACTGGTGAGT	ARQEARGAAARGGDW*
773	C	CACGGCAAGAGGCGAGGGGCG	TGCGGCCGCA	GGGGGGCGGCGACTGGTGAGTA	RQEARGCGRRRGGDW*V
774	G	ACGGCAAGAGGCGAGGGGCGG	TGCGGCCGCA	GGGGGGCGGCGACTGGTGAGTAC	RQEARGVRRPQGGDW*V
775	G	CGGCAAGAGGCGAGGGGCGG	TGCGGCCGCA	GGCGGCGACTGGTGAGTACG	RQEARGGAAGGDW*V
776	C	GGCAAGAGGCGAGGGGCGGC	TGCGGCCGCA	GCGGCGACTGGTGAGTACGC	QEARGGCGRSGDW*VR
777	G	GCAAGAGGCGAGGGGCGGGC	TGCGGCCGCA	CGGCGACTGGTGAGTACGCC	QEARGGVRRPHGDW*VR
778	A	CAAGAGGCGAGGGGCGGCGA	TGCGGCCGCA	GCGCGACTGGTGAGTACGCCA	QEARGGDAAGDW*VR
779	C	AAGAGGCGAGGGGCGGCGC	TGCGGCCGCA	GCGACTGGTGAGTACGCCAA	EARGGDCGRSDW*VRQ
780	T	AGAGGCGAGGGGCGGCGACT	TGCGGCCGCA	CGACTGGTGAGTACGCCAAA	EARGGDLRPHDW*VRQ
781	G	GAGGCGAGGGGCGGCGACTG	TGCGGCCGCA	GACTGGTGAGTACGCCAAAA	EARGGDCAAADW*VRQ
782	G	AGGCGAGGGGCGGCGACTGG	TGCGGCCGCA	ACTGGTGAGTACGCCAAAAA	ARGGDWCGRR*VRQK
783	T	GCGAGGGGCGGCGACTGGT	TGCGGCCGCA	CTGGTGAGTACGCCAAAAAT	ARGGDWRPHW*VRQK
784	G	GCGAGGGGCGGCGACTGGTG	TGCGGCCGCA	TGGTGAGTACGCCAAAAATT	ARGGDWCAAAW*VRQK
785	A	CGAGGGGCGGCGACTGGTGA	TGCGGCCGCA	GGTGAGTACGCCAAAAATTT	RGGDW*CGRR*VRQKF
786	G	GAGGGGCGGCGACTGGTGAG	TGCGGCCGCA	GTGAGTACGCCAAAAATTTT	RGGDW*VRPQ*VRQKF
787	T	AGGGGCGGCGACTGGTGAGT	TGCGGCCGCA	TGAGTACGCCAAAAATTTTG	RGGDW*VAAA*VRQKF
788	A	GGGGGCGGCGACTGGTGAGTA	TGCGGCCGCA	GAGTACGCCAAAAATTTTGA	GGDW*VCGRRVRQKF*
789	C	GGGCGGCGACTGGTGAGTAC	TGCGGCCGCA	AGTACGCCAAAAATTTGAC	GGDW*VLRPQVRQKF*
790	G	GGCGGCGACTGGTGAGTACG	TGCGGCCGCA	GTACGCCAAAAATTTTGACT	GGDW*VRAAAVRQKF*
791	C	GCGGCGACTGGTGAGTACGC	TGCGGCCGCA	TACGCCAAAAATTTTGACTA	GDW*VRCRRIQKF*L
792	C	CGGCGACTGGTGAGTACGCC	TGCGGCCGCA	ACGCCAAAAATTTTGACTAG	GDW*VRLRPQRQKF*L
793	A	GGCGACTGGTGAGTACGCCA	TGCGGCCGCA	CGCCAAAAATTTTGACTAGC	GDW*VRHAAARQKF*L
794	A	GCGACTGGTGAGTACGCCAA	TGCGGCCGCA	GCCAAAAATTTTGACTAGCG	DW*VRQCGRSQKF*LA
795	A	CGACTGGTGAGTACGCCAAA	TGCGGCCGCA	CCAAAAATTTTGACTAGCGG	DW*VRQMRPHQKF*LA
796	A	GACTGGTGAGTACGCCAAAA	TGCGGCCGCA	CAAAAAATTTTGACTAGCGGA	DW*VRQNAAAQKF*LA
797	A	ACTGGTGAGTACGCCAAAAA	TGCGGCCGCA	AAAAATTTTGACTAGCGGAG	W*VRQKCRKFK*LAE
798	T	CTGGTGAGTACGCCAAAAAT	TGCGGCCGCA	AAAAATTTTGACTAGCGGAGG	W*VRQKLRPQKF*LAE
799	T	TGGTGAGTACGCCAAAAATT	TGCGGCCGCA	AAATTTTGACTAGCGGAGGC	W*VRQKFAAAKF*LAE
800	T	GGTGAGTACGCCAAAAATTT	TGCGGCCGCA	AATTTTGACTAGCGGAGGCT	*VRQKFCGRKF*LAEA
801	T	GTGAGTACGCCAAAAATTTT	TGCGGCCGCA	ATTTTGACTAGCGGAGGCTA	*VRQKFLRPQF*LAEA
802	G	TGAGTACGCCAAAAATTTTG	TGCGGCCGCA	TTTTGACTAGCGGAGGCTAG	*VRQKFCAAAF*LAEA
803	A	GAGTACGCCAAAAATTTTGA	TGCGGCCGCA	TTTGACTAGCGGAGGCTAGA	VRQKF*CGRI*LAEAR
804	C	AGTACGCCAAAAATTTTACT	TGCGGCCGCA	TTGACTAGCGGAGGCTAGAA	VRQKF*LRPH*LAEAR
805	T	GTACGCCAAAAATTTTGACT	TGCGGCCGCA	TGACTAGCGGAGGCTAGAAG	VRQKF*LAEA*LAEAR
806	A	TACGCCAAAAATTTTGACTA	TGCGGCCGCA	GACTAGCGGAGGCTAGAAGG	ROKFL*CGRRLEARR
807	G	ACGCCAAAAATTTTGACTAG	TGCGGCCGCA	ACTAGCGGAGGCTAGAAGGA	ROKFL*VLRPQLEARR
808	C	CGCCAAAAATTTTGACTAGC	TGCGGCCGCA	CTAGCGGAGGCTAGAAGGAG	ROKFL*LAEALEARR
809	G	GCCAAAAATTTTGACTAGCG	TGCGGCCGCA	TAGCGGAGGCTAGAAGGAGA	QKF*LACGRLEARRR
810	G	CCAAAAATTTTGACTAGCGG	TGCGGCCGCA	AGCGGAGGCTAGAAGGAGAG	QKF*LAVRPQLEARRR
811	A	CAAAAAATTTTGACTAGCGGA	TGCGGCCGCA	GCGGAGGCTAGAAGGAGAGA	QKF*LAADAAAEARRR
812	G	AAAAATTTTGACTAGCGGAG	TGCGGCCGCA	CGGAGGCTAGAAGGAGAGAG	KF*LAECGRTEARRR
813	G	AAAATTTTGACTAGCGGAGG	TGCGGCCGCA	GGAGGCTAGAAGGAGAGAGA	KF*LAEVRRQEARRR
814	C	AAATTTTGACTAGCGGAGGC	TGCGGCCGCA	GAGGCTAGAAGGAGAGAGAT	KF*LEAAEAAREARRR
815	T	AATTTTGACTAGCGGAGGCT	TGCGGCCGCA	AGGCTAGAAGGAGAGAGATG	F*LAEACGRKARRR
816	A	ATTTTGACTAGCGGAGGCTA	TGCGGCCGCA	GGCTAGAAGGAGAGAGATGG	F*LAEAMRPQARRR
817	G	TTTTGACTAGCGGAGGCTAG	TGCGGCCGCA	GCTAGAAGGAGAGAGATGGG	F*LAEASAAAARRR
818	A	TTTGACTAGCGGAGGCTAGA	TGCGGCCGCA	CTAGAAGGAGAGAGATGGGT	*LAEARCTRARRR
819	A	TTGACTAGCGGAGGCTAGAA	TGCGGCCGCA	TAGAAGGAGAGAGATGGGTG	*LAEARMRPHRRR
820	G	TGACTAGCGGAGGCTAGAA	TGCGGCCGCA	AGAAGGAGAGAGATGGGTGC	*LAEARSAARRR
821	G	GACTAGCGGAGGCTAGAAG	TGCGGCCGCA	GAAGGAGAGAGATGGGTGCG	LAEARRCGRRRR
822	A	ACTAGCGGAGGCTAGAAGGA	TGCGGCCGCA	AAGGAGAGAGATGGGTGCGA	LAEARMRPQRREMG
823	G	CTAGCGGAGGCTAGAAGGAG	TGCGGCCGCA	AGGAGAGAGATGGGTGCGAG	LAEARSAARRR
824	A	TAGCGGAGGCTAGAAGGAGA	TGCGGCCGCA	GAGAGAGAGATGGGTGCGAGA	AEARRRCGRRRR
825	G	AGCGGAGGCTAGAAGGAGAG	TGCGGCCGCA	GAGAGAGATGGGTGCGAGAG	AEARRRVRPQREMG
826	A	GCGGAGGCTAGAAGGAGAGA	TGCGGCCGCA	AGAGAGATGGGTGCGAGAGC	AEARRRDAARREMG
827	G	CGGAGGCTAGAAGGAGAGAG	TGCGGCCGCA	GAGAGATGGGTGCGAGAGCG	EARRRRCGRRRR
828	A	GGAGGCTAGAAGGAGAGAGA	TGCGGCCGCA	AGAGATGGGTGCGAGAGCGT	EARRRMRPQREMG
829	T	GAGGCTAGAAGGAGAGAGAT	TGCGGCCGCA	GAGATGGGTGCGAGAGCGTC	EARRRREIAAEEMG
830	G	AGGCTAGAAGGAGAGAGATG	TGCGGCCGCA	AGATGGGTGCGAGAGCGTCG	ARRRRCGRKMGARAS
831	G	GGCTAGAAGGAGAGAGATGG	TGCGGCCGCA	GATGGGTGCGAGAGCGTCGG	ARRRMRPQMGARAS
832	G	GCTAGAAGGAGAGAGATGGG	TGCGGCCGCA	ATGGGTGCGAGAGCGTCGGT	ARRRMRPQMGARAS
833	T	CTAGAAGGAGAGAGATGGGT	TGCGGCCGCA	TGGGTGCGAGAGCGTCGGTA	RRRRCGRMGARASV
834	G	TAGAAGGAGAGAGATGGGTG	TGCGGCCGCA	GGGTGCGAGAGCGTCGGTAT	RRRRCGRVPQMGARASV
835	C	AGAAGGAGAGAGATGGGTGC	TGCGGCCGCA	GGTGCGAGAGCGTCGGTATT	RRRRCMGAAAARASV
836	G	GAAGGAGAGAGATGGGTGCG	TGCGGCCGCA	GTGCGAGAGCGTCGGTATTA	RRRRCMGAGRSARASV
837	A	AAGGAGAGAGATGGGTGCGA	TGCGGCCGCA	TGCGAGAGCGTCGGTATTA	RRRRCMGAMRPHARASV
838	G	AGGAGAGAGATGGGTGCGAG	TGCGGCCGCA	GCGAGAGCGTCGGTATTAAG	RRRRCMGASAAAARASV
839	A	GGAGAGAGATGGGTGCGAGA	TGCGGCCGCA	CGAGAGCGTCGGTATTAAGC	RRRRCMGARCTRASV
840	G	GAGAGAGATGGGTGCGAGAG	TGCGGCCGCA	GAGAGCGTCGGTATTAAGCG	RRRRCMGARVRRASV
841	C	AGAGAGATGGGTGCGAGAGC	TGCGGCCGCA	AGAGCGTCGGTATTAAGCGG	RRRRCMGARAAAARASV
842	G	GAGAGATGGGTGCGAGAGCG	TGCGGCCGCA	GAGCGTCGGTATTAAGCGGG	RRRRCMGARACRRASV
843	T	AGAGATGGGTGCGAGAGCGT	TGCGGCCGCA	AGCGTCGGTATTAAGCGGGG	RRRRCMGARALRPQASV
844	C	GAGATGGGTGCGAGAGCGTC	TGCGGCCGCA	GCGTCGGTATTAAGCGGGGG	RRRRCMGARASAAAARASV
845	G	AGATGGGTGCGAGAGCGTCG	TGCGGCCGCA	CGTCGGTATTAAGCGGGGGG	RRRRCMGARASCRTASV
846	G	GATGGGTGCGAGAGCGTCGG	TGCGGCCGCA	GTGCGTATTAAGCGGGGGGAG	RRRRCMGARASVRRASV
847	T	ATGGGTGCGAGAGCGTCGGT	TGCGGCCGCA	TCGGTATTAAGCGGGGGGAGA	RRRRCMGARASVAAAARASV
848	A	TGGGTGCGAGAGCGTCGGTA	TGCGGCCGCA	CGGTATTAAGCGGGGGGAGAA	RRRRCMGARASVCGRTASV
849	T	GGGTGCGAGAGCGTCGGTATT	TGCGGCCGCA	GGTATTAAGCGGGGGGAGAA	RRRRCMGARASVLRPQASV
850	T	GGTGGCGAGAGCGTCGGTATT	TGCGGCCGCA	GTATTAAGCGGGGGGAGAA	RRRRCMGARASVAAAARASV
851	A	GTGGAGAGCGTCGGTATTA	TGCGGCCGCA	TATTAAGCGGGGGGAGAA	RRRRCMGARASVLRPQASV
852	A	TGCGAGAGCGTCGGTATTA	TGCGGCCGCA	ATTAAGCGGGGGGAGAA	RRRRCMGARASVLRPQASV
853	G	GCGAGAGCGTCGGTATTAAG	TGCGGCCGCA	TTAAGCGGGGGGAGAA	RRRRCMGARASVLRPQASV
854	C	CGAGAGCGTCGGTATTAAGC	TGCGGCCGCA	TAAGCGGGGGGAGAA	RRRRCMGARASVLRPQASV

855	G	GAGAGCGTCGGTATTAAGCG	TGCGGCCGCA	AAGCGGGGAGAATTAGATA	RASVLSVRPQSGGELD
856	G	AGAGCGTCGGTATTAAGCGG	TGCGGCCGCA	AGCGGGGAGAATTAGATAA	RASVLSGAAASGGELD
857	G	GAGCGTCGGTATTAAGCGGG	TGCGGCCGCA	GCGGGGAGAATTAGATAAA	ASVLSGCGRRSGGELDK
858	G	AGCGTCGGTATTAAGCGGGG	TGCGGCCGCA	CGGGGAGAATTAGATAAAAT	ASVLSGVRPHGGELDK
859	G	CGCTCGGTATTAAGCGGGGG	TGCGGCCGCA	GGGGGAGAATTAGATAAAATG	ASVLSGGAAAGGELDK
860	A	CGTCGGTATTAAGCGGGGGGA	TGCGGCCGCA	GGGGAGAATTAGATAAAATGG	SVLSGGCGRRRGLDKW
861	G	GTCCGTATTAAGCGGGGGAG	TGCGGCCGCA	GGGAGAATTAGATAAAATGGG	SVLSGGVRRPQGGELDKW
862	A	TCCGTATTAAGCGGGGGAGA	TGCGGCCGCA	GGAGAATTAGATAAAATGGGA	SVLSGGDAAAGELDKW
863	A	CGGTATTAAGCGGGGGAGAA	TGCGGCCGCA	GAGAATTAGATAAAATGGGAA	VLSSGEGCRRRGLDKWE
864	T	GGTATTAAGCGGGGGAGAAT	TGCGGCCGCA	AGAATTAGATAAAATGGGAAA	VLSSGELRPQELDKWE
865	T	GTATTAAGCGGGGGAGAATT	TGCGGCCGCA	GAATTAGATAAAATGGGAAAA	VLSSGGEFAAAELDKWE
866	A	TATTAAGCGGGGGAGAATTA	TGCGGCCGCA	AATTAGATAAAATGGGAAAAA	LSGGELCGRKLDKWEK
867	G	ATTAAGCGGGGGAGAATTAG	TGCGGCCGCA	ATTAGATAAAATGGGAAAAAA	LSGGELVRRPQLDKWEK
868	A	TTAAGCGGGGGAGAATTAGA	TGCGGCCGCA	TTAGATAAAATGGGAAAAAAT	LSGGELDAAALDKWEK
869	T	TAAGCGGGGGAGAATTAGAT	TGCGGCCGCA	TAGATAAAATGGGAAAAAATT	SGGELDCGRIDKWEKI
870	A	AAGCGGGGGAGAATTAGATA	TGCGGCCGCA	AGATAAAATGGGAAAAAATTC	SGGELDMRPQDKWEKI
871	A	AGCGGGGGAGAATTAGATAA	TGCGGCCGCA	GATAAAATGGGAAAAAATTCG	SGGELDNAAANKWEKI
872	A	GCGGGGGAGAATTAGATAAA	TGCGGCCGCA	ATAAATGGGAAAAAATTCGG	GGELDKCGRNKWEKIR
873	T	CGGGGGAGAATTAGATAAAAT	TGCGGCCGCA	TAAATGGGAAAAAATTCGGT	GGELDKLRPHKWEKIR
874	G	GGGGGAGAATTAGATAAAATG	TGCGGCCGCA	AAATGGGAAAAAATTCGGTT	GGELDKCAAANKWEKIR
875	G	GGGGAGAATTAGATAAAATGG	TGCGGCCGCA	AATGGGAAAAAATTCGGTTA	GELDKWVRRPQWEKIRL
876	G	GGGAGAATTAGATAAAATGGG	TGCGGCCGCA	ATGGGAAAAAATTCGGTTAA	GELDKWVRRPQWEKIRL
877	A	GGAGAATTAGATAAAATGGGA	TGCGGCCGCA	TGGGAAAAAATTCGGTTAAG	GELDKWVRRPQWEKIRL
878	A	GAGAATTAGATAAAATGGGAA	TGCGGCCGCA	GGGAAAAAATTCGGTTAAGG	ELDKWECGRREKIRLR
879	A	AGAATTAGATAAAATGGGAAA	TGCGGCCGCA	GGAAAAAATTCGGTTAAGGC	ELDKWEMRRPQEKIRLR
880	A	GAATTAGATAAAATGGGAAAA	TGCGGCCGCA	GAAAAAATTCGGTTAAGGCC	ELDKWENAAEKIRLR
881	A	AATTAGATAAAATGGGAAAAA	TGCGGCCGCA	AAAAAATTCGGTTAAGGCCA	LDKWEKCGRKKIRLRP
882	A	ATTAGATAAAATGGGAAAAAA	TGCGGCCGCA	AAAAATTCGGTTAAGGCCAG	LDKWEKMRPQKIRLRP
883	T	TTAGATAAAATGGGAAAAAAT	TGCGGCCGCA	AAAAATTCGGTTAAGGCCAGG	LDKWEKIAAAKIRLRP
884	T	TAGATAAAATGGGAAAAAATTC	TGCGGCCGCA	AAAAATTCGGTTAAGGCCAGGG	DKWEKICGRKIRLRPG
885	C	AGATAAAATGGGAAAAAATTC	TGCGGCCGCA	AATTCGGTTAAGGCCAGGGG	DKWEKIRPQKIRLRPG
886	G	GATAAAATGGGAAAAAATTCG	TGCGGCCGCA	ATTCGGTTAAGGCCAGGGGG	DKWEKIRAAAIRLRPG
887	G	ATAAAATGGGAAAAAATTCGG	TGCGGCCGCA	TTCGGTTAAGGCCAGGGGGA	KWEKIRCGRIRLRPGG
888	T	TAAATGGGAAAAAATTCGGTT	TGCGGCCGCA	TCGGTTAAGGCCAGGGGGAA	KWEKIRLRPHLRPQGG
889	T	AAATGGGAAAAAATTCGGTT	TGCGGCCGCA	CGGTTAAGGCCAGGGGGAAA	KWEKIRFAAARLRPQGG
890	A	AATGGGAAAAAATTCGGTTA	TGCGGCCGCA	GGTTAAGGCCAGGGGGAAAG	WEKIRLCGRRLRPQGGK
891	A	ATGGGAAAAAATTCGGTTAA	TGCGGCCGCA	GTTAAGGCCAGGGGGAAAGA	WEKIRLMRPQLRPQGGK
892	G	TGGGAAAAAATTCGGTTAAG	TGCGGCCGCA	TTAAGGCCAGGGGGAAAGAA	WEKIRLSAAALRPQGGK
893	G	GGGAAAAAATTCGGTTAAGG	TGCGGCCGCA	TAAGGCCAGGGGGAAAGAAA	EKIRLCGRIRPQGGKK
894	C	GGAAAAAATTCGGTTAAGGC	TGCGGCCGCA	AAGGCCAGGGGGAAAGAAAC	EKIRLRPQRRPQGGKK
895	C	GAAAAAATTCGGTTAAGGCC	TGCGGCCGCA	AGGCCAGGGGGAAAGAAACA	EKIRLRPAARPPGGKK
896	A	AAAAAATTCGGTTAAGGCCA	TGCGGCCGCA	GGCCAGGGGGAAAGAAACAA	KIRLRPCGRRPQGGKKQ
897	G	AAAAATTCGGTTAAGGCCAG	TGCGGCCGCA	GCCAGGGGGAAAGAAACAAT	KIRLRPVRPQGGKKQK
898	G	AAAAATTCGGTTAAGGCCAGG	TGCGGCCGCA	CCAGGGGGAAAGAAACAATA	KIRLRPAAAPQGGKKQ
899	G	AAATTCGGTTAAGGCCAGGG	TGCGGCCGCA	CAGGGGGAAAGAAACAATAT	IRLRPQCGRTQGGKKQY
900	G	AATTCGGTTAAGGCCAGGGG	TGCGGCCGCA	AGGGGGAAAGAAACAATATA	IRLRPQVRRPQGGKKQY
901	G	ATTCGGTTAAGGCCAGGGGG	TGCGGCCGCA	GGGGAAAGAAACAATATAAA	IRLRPQGAAPQGGKKQY
902	A	TTCCGGTTAAGGCCAGGGGGA	TGCGGCCGCA	GGGAAAGAAACAATATAAA	RLRPQGGCRRRQGGKKQYK
903	A	TCCGGTTAAGGCCAGGGGGAA	TGCGGCCGCA	GGGAAAGAAACAATATAAACT	RLRPQGGMRPQGGKKQYK
904	A	CGGTTAAGGCCAGGGGGAAA	TGCGGCCGCA	GGAAAGAAACAATATAAACT	RLRPQGGNAAAGKKQYK
905	G	GGTTAAGGCCAGGGGGAAAAG	TGCGGCCGCA	GAAAGAAACAATATAAACTA	LRPQGGKCGRRKQYK
906	A	GTTAAGGCCAGGGGGAAAAGA	TGCGGCCGCA	AAAGAAACAATATAAACTAA	LRPQGGKMRPQGGKKQYK
907	A	TTAAGGCCAGGGGGAAAAGAA	TGCGGCCGCA	AAGAACAATATAAACTAAA	LRPQGGKNAAAKQYK
908	A	TAAGGCCAGGGGGAAAAGAAA	TGCGGCCGCA	AGAAACAATATAAACTAAAA	RPQGGKCGRRKQYK
909	C	AAGGCCAGGGGGAAAAGAAAC	TGCGGCCGCA	GAAACAATATAAACTAAAAC	RPQGGKLRPQGGKKQYK
910	A	AGGCCAGGGGGAAAAGAAACA	TGCGGCCGCA	AAACAATATAAACTAAAACA	RPQGGKHAQAQYK
911	A	GGCCAGGGGGAAAAGAAACA	TGCGGCCGCA	ACAATATAAACTAAAAACAT	PGGKKQCGRRKQYK
912	T	GCCAGGGGGAAAAGAAACAAT	TGCGGCCGCA	ACAATATAAACTAAAAACATA	PGGKKQLRQYK
913	A	CCAGGGGGAAAAGAAACAATA	TGCGGCCGCA	CAATATAAACTAAAAACATAT	PGGKKQYAAAQYK
914	T	CAGGGGGAAAAGAAACAATAT	TGCGGCCGCA	AATATAAACTAAAAACATATA	GGKKQYCGRRKQYK
915	A	AGGGGGAAAAGAAACAATATA	TGCGGCCGCA	ATATAAACTAAAAACATATAG	GGKKQYMRPQYK
916	A	GGGGAAAAGAAACAATATAA	TGCGGCCGCA	TATAAACTAAAAACATATAGT	GGKKQYNAAYK
917	A	GGGAAAAGAAACAATATAAAC	TGCGGCCGCA	ATAAACTAAAAACATATAGTA	GKKQYKCGRRKQYK
918	C	GGGAAAAGAAACAATATAAAC	TGCGGCCGCA	TAAACTAAAAACATATAGTAT	GKKQYKLRPHKQYK
919	T	GGAAAAGAAACAATATAAACT	TGCGGCCGCA	AAACTAAAAACATATAGTATG	GKKQYKLAALKQYK
920	A	GAAAAGAAACAATATAAACTA	TGCGGCCGCA	AACTAAAAACATATAGTATGG	KKQYKCGRRKQYK
921	A	AAAGAAACAATATAAACTAA	TGCGGCCGCA	ACTAAAAACATATAGTATGGG	KKQYKLMRPQKQYK
922	A	AAGAAACAATATAAACTAAA	TGCGGCCGCA	CTAAAAACATATAGTATGGGC	KKQYKLNAAKQYK
923	A	AGAACAATATAAACTAAAA	TGCGGCCGCA	TAAAAACATATAGTATGGGCA	KQYKLCGRKQYK
924	C	GAAACAATATAAACTAAAA	TGCGGCCGCA	AAAACATATAGTATGGGCAA	KQYKLRPQKQYK
925	A	AAACAATATAAACTAAAA	TGCGGCCGCA	AAACATATAGTATGGGCAAG	KQYKLAHAAKQYK
926	T	AACAATATAAACTAAAA	TGCGGCCGCA	AACATATAGTATGGGCAAGC	QYKLCGRKQYK
927	A	ACAATATAAACTAAAA	TGCGGCCGCA	ACATATAGTATGGGCAAGCA	QYKLCGRKQYK
928	T	CAATATAAACTAAAA	TGCGGCCGCA	CATATAGTATGGGCAAGCAG	QYKLCGRKQYK
929	A	AATATAAACTAAAA	TGCGGCCGCA	ATATAGTATGGGCAAGCAGG	YKLCGRKQYK
930	G	ATATAAACTAAAA	TGCGGCCGCA	TATAGTATGGGCAAGCAGGG	YKLCGRKQYK
931	T	TATAAACTAAAA	TGCGGCCGCA	ATAGTATGGGCAAGCAGGGA	YKLCGRKQYK
932	A	ATAAACTAAAA	TGCGGCCGCA	TAGTATGGGCAAGCAGGGAG	YKLCGRKQYK
933	T	TAAACTAAAA	TGCGGCCGCA	AGTATGGGCAAGCAGGGAGC	YKLCGRKQYK
934	G	AAACTAAAA	TGCGGCCGCA	GTATGGGCAAGCAGGGAGCT	YKLCGRKQYK
935	G	AACTAAAA	TGCGGCCGCA	TATGGGCAAGCAGGGAGCTA	YKLCGRKQYK
936	G	ACTAAAA	TGCGGCCGCA	ATGGGCAAGCAGGGAGCTAG	YKLCGRKQYK
937	C	CTAAAA	TGCGGCCGCA	TGGGCAAGCAGGGAGCTAGA	YKLCGRKQYK
938	A	TAAAA	TGCGGCCGCA	GGGCAAGCAGGGAGCTAGAA	YKLCGRKQYK
939	A	AAAA	TGCGGCCGCA	GGCAAGCAGGGAGCTAGAAC	YKLCGRKQYK
940	G	AAACATATAGTATGGGCAAG	TGCGGCCGCA	GCAAGCAGGGAGCTAGAACG	YKLCGRKQYK



941	C	AACATATAGTATGGGCAAGC	TGCGGCCGCA	CAAGCAGGGAGCTAGAACGA	HIVWASCGRTSRELER
942	A	ACATATAGTATGGGCAAGCA	TGCGGCCGCA	AAGCAGGGAGCTAGAACGAT	HIVWASMRPQSRELER
943	G	CATATAGTATGGGCAAGCAG	TGCGGCCGCA	AGCAGGGAGCTAGAACGATT	HIVWASAAAARELER
944	G	ATATAGTATGGGCAAGCAGG	TGCGGCCGCA	GCAGGGAGCTAGAACGATTC	IVWASRCGRSRELERF
945	G	TATAGTATGGGCAAGCAGGG	TGCGGCCGCA	CAGGGAGCTAGAACGATTCCG	IVWASRVPRHLERF
946	A	ATAGTATGGGCAAGCAGGGA	TGCGGCCGCA	AGGGAGCTAGAACGATTCCG	IVWASRDAAAARELERF
947	G	TAGTATGGGCAAGCAGGGAG	TGCGGCCGCA	GGGAGCTAGAACGATTCCGCA	VWASRECGRRLELRF
948	C	AGTATGGGCAAGCAGGGAGC	TGCGGCCGCA	GGAGCTAGAACGATTCCGAG	VWASRELRPQELERFA
949	T	GTATGGGCAAGCAGGGAGCT	TGCGGCCGCA	GAGCTAGAACGATTCCGAGT	VWASRELAALERLFA
950	A	TATGGGCAAGCAGGGAGCTA	TGCGGCCGCA	AGCTAGAACGATTCCGAGTT	WASRELCGRKLERFAV
951	G	ATGGGCAAGCAGGGAGCTAG	TGCGGCCGCA	GCTAGAACGATTCCGAGTTA	WASRELVPRQLERFAV
952	A	TGGGCAAGCAGGGAGCTAGA	TGCGGCCGCA	CTAGAACGATTCCGAGTTAA	WASRELDAAALERFAV
953	A	GGGCAAGCAGGGAGCTAGAA	TGCGGCCGCA	TAGAACGATTCCGAGTTAAT	ASRELECGRIERFAVN
954	C	GGCAAGCAGGGAGCTAGAAC	TGCGGCCGCA	AGAACGATTCCGAGTTAATC	ASRELELRPQERFAVN
955	G	GCAAGCAGGGAGCTAGAACG	TGCGGCCGCA	GAACGATTCCGAGTTAATCC	ASRELERAAAERFAVN
956	A	CAAGCAGGGAGCTAGAACGA	TGCGGCCGCA	AACGATTCCGAGTTAATCCT	SRELERCGRKRFAVNP
957	T	AAGCAGGGAGCTAGAACGAT	TGCGGCCGCA	ACGATTCCGAGTTAATCCTG	SRELERLRPQRFVAVNP
958	T	AGCAGGGAGCTAGAACGATT	TGCGGCCGCA	CGATTCCGAGTTAATCCTGG	SRELERFAAARFAVNP
959	C	GCAGGGAGCTAGAACGATT	TGCGGCCGCA	GATTCCGAGTTAATCCTGGC	RELERFCGRRFAVNP
960	G	CAGGGAGCTAGAACGATTCCG	TGCGGCCGCA	ATTCCGAGTTAATCCTGGCC	RELERFVRPQFAVNP
961	C	AGGGAGCTAGAACGATTCCG	TGCGGCCGCA	TTCCGAGTTAATCCTGGCCT	RELERFAAARFAVNP
962	A	GGGAGCTAGAACGATTCCGCA	TGCGGCCGCA	TCGCGAGTTAATCCTGGCCT	ELERFACGRIVAVNPGL
963	G	GGAGCTAGAACGATTCCGAG	TGCGGCCGCA	CGCAGTTAATCCTGGCCTTT	ELERFAVRPHAVNPGL
964	T	GAGCTAGAACGATTCCGAGT	TGCGGCCGCA	GCAGTTAATCCTGGCCTTTT	ELERFAVAAAANPGL
965	T	AGCTAGAACGATTCCGAGTT	TGCGGCCGCA	CAGTTAATCCTGGCCTTTTA	LERFAVCGRTVNPGLL
966	A	GCTAGAACGATTCCGAGTTA	TGCGGCCGCA	AGTTAATCCTGGCCTTTTAG	LERFAVMRQVNPVNPGLL
967	A	CTAGAACGATTCCGAGTTAA	TGCGGCCGCA	GTTAATCCTGGCCTTTTAGA	LERFAVNAAVNPVNPGLL
968	T	TAGAACGATTCCGAGTTAAT	TGCGGCCGCA	TTAATCCTGGCCTTTTAGAG	ERFAVNCGRINPGLLE
969	C	AGAACGATTCCGAGTTAATC	TGCGGCCGCA	TAATCCTGGCCTTTTAGAGA	ERFAVNLRPHNPGLLE
970	C	GAACGATTCCGAGTTAATCC	TGCGGCCGCA	AATCCTGGCCTTTTAGAGAC	ERFAVNPAAANPGLLE
971	T	AACGATTCCGAGTTAATCCT	TGCGGCCGCA	ATCCTGGCCTTTTAGAGACA	RFVAVNPCGRNPGLLET
972	G	ACGATTCCGAGTTAATCCTG	TGCGGCCGCA	TCCTGGCCTTTTAGAGACAT	RFVAVNPRPHNPGLLET
973	G	CGATTCCGAGTTAATCCTGG	TGCGGCCGCA	CCTGGCCTTTTAGAGACATC	RFVAVNPAAAAPGLLET
974	C	GATTCCGAGTTAATCCTGGC	TGCGGCCGCA	CTGGCCTTTTAGAGACATCA	FAVNPCCGRTGLLETS
975	C	ATTCCGAGTTAATCCTGGCC	TGCGGCCGCA	TGGCCTTTTAGAGACATCAG	FAVNPGLRPHGLLETS
976	T	TTCCGAGTTAATCCTGGCCT	TGCGGCCGCA	GGCCTTTTAGAGACATCAGA	FAVNPGLAAAAGLLETS
977	T	TCGAGTTAATCCTGGCCTTT	TGCGGCCGCA	GCCTTTTAGAGACATCAGAA	AVNPGLCGRSLLETSE
978	T	CGCAGTTAATCCTGGCCTTT	TGCGGCCGCA	CCTTTTAGAGACATCAGAA	AVNPGLLRPHLLETSE
979	T	GCAGTTAATCCTGGCCTTTT	TGCGGCCGCA	CTTTTAGAGACATCAGAA	AVNPGLFAAALLETSE
980	A	CAGTTAATCCTGGCCTTTTA	TGCGGCCGCA	TTTTAGAGACATCAGAA	VNPGLLCGRILETSEG
981	G	AGTTAATCCTGGCCTTTTAG	TGCGGCCGCA	TTTAGAGACATCAGAA	VNPGLLRPHLLETSEG
982	A	GTTAATCCTGGCCTTTTAGA	TGCGGCCGCA	TTAGAGACATCAGAA	VNPGLLVRPHLLETSEG
983	G	TTAATCCTGGCCTTTTAGAG	TGCGGCCGCA	TAGAGACATCAGAA	NPGLLECGRIETSEG
984	A	TAATCCTGGCCTTTTAGAGA	TGCGGCCGCA	AGAGACATCAGAA	NPGLLEMPAETSEG
985	C	AATCCTGGCCTTTTAGAGAC	TGCGGCCGCA	GAGACATCAGAA	NPGLLETRPQETSEG
986	A	ATCCTGGCCTTTTAGAGACA	TGCGGCCGCA	AGACATCAGAA	PGLLETGCRKLETSEG
987	T	TCCTGGCCTTTTAGAGACAT	TGCGGCCGCA	GACATCAGAA	PGLLETLPQETSEG
988	C	CCTGGCCTTTTAGAGACATC	TGCGGCCGCA	ACATCAGAA	PGLLETSAAETSEG
989	A	CTGGCCTTTTAGAGACATCA	TGCGGCCGCA	CATCAGAA	GLLETSCGRTSEGCR
990	G	TGGCCTTTTAGAGACATCAG	TGCGGCCGCA	ATCAGAA	GLLETSCGRTSEGCR
991	A	GGCCTTTTAGAGACATCAGA	TGCGGCCGCA	TCAGAA	GLLETSDAAASEGCR
992	A	GCCTTTTAGAGACATCAGAA	TGCGGCCGCA	CAGAA	LLETSECGRTSEGCR
993	G	CCTTTTAGAGACATCAGAA	TGCGGCCGCA	AGAAGGCTGTAGACAAATAC	LLETSEVRPQEGCRQI
994	G	CTTTTAGAGACATCAGAA	TGCGGCCGCA	GAAGGCTGTAGACAAATACT	LLETSEGAAAEGCRQI
995	C	TTTTAGAGACATCAGAA	TGCGGCCGCA	AAGGCTGTAGACAAATACTG	LETSEGCGRKQI
996	T	TTTAGAGACATCAGAA	TGCGGCCGCA	AGGCTGTAGACAAATACTGG	LETSEGLRPPQGRQI
997	G	TTAGAGACATCAGAA	TGCGGCCGCA	GGCTGTAGACAAATACTGGG	LETSEGCAAAGCRQI
998	T	TAGAGACATCAGAA	TGCGGCCGCA	GCTGTAGACAAATACTGGGA	ETSEGCGRGCRQILG
999	A	AGAGACATCAGAA	TGCGGCCGCA	CTGTAGACAAATACTGGGAC	ETSEGCMPRPHCRQILG
1000	G	GAGACATCAGAA	TGCGGCCGCA	TGTAGACAAATACTGGGACA	ETSEGCSSAACRQILG
1001	A	AGACATCAGAA	TGCGGCCGCA	GTAGACAAATACTGGGACAG	TSEGCRCGRSRQILGQ
1002	C	GACATCAGAA	TGCGGCCGCA	TAGACAAATACTGGGACAGC	TSEGCRLRPHRQILGQ
1003	A	ACATCAGAA	TGCGGCCGCA	AGACAAATACTGGGACAGCT	TSEGCRRHAARQILGQ
1004	A	CATCAGAA	TGCGGCCGCA	GACAAATACTGGGACAGCTA	TSEGCRCGRRQILGQL
1005	A	ATCAGAA	TGCGGCCGCA	ACAAATACTGGGACAGCTAC	TSEGCRCMRPQILGQL
1006	T	TCAGAA	TGCGGCCGCA	CAAATACTGGGACAGCTACA	TSEGCRCQIAAAILGQL
1007	A	CAGAA	TGCGGCCGCA	AAATACTGGGACAGCTACAA	TSEGCRCQICGRKILGQL
1008	C	AGAAGGCTGTAGACAAATAC	TGCGGCCGCA	AATACTGGGACAGCTACAAC	TSEGCRCQILRQILGQL
1009	T	GAAGGCTGTAGACAAATAC	TGCGGCCGCA	ATACTGGGACAGCTACAACC	TSEGCRCQILAAAAILGQL
1010	G	AAGGCTGTAGACAAATACTG	TGCGGCCGCA	TACTGGGACAGCTACAACCA	TSEGCRCQILCGRILGQL
1011	G	AGGCTGTAGACAAATACTGG	TGCGGCCGCA	ACTGGGACAGCTACAACCAT	TSEGCRCQILVRPQLGQL
1012	G	GGCTGTAGACAAATACTGGG	TGCGGCCGCA	CTGGGACAGCTACAACCATC	TSEGCRCQILGAAAILGQL
1013	A	GCTGTAGACAAATACTGGGA	TGCGGCCGCA	TGGGACAGCTACAACCATCC	TSEGCRCQILGCRMRQILGQ
1014	C	CTGTAGACAAATACTGGGAC	TGCGGCCGCA	GGGACAGCTACAACCATCCC	TSEGCRCQILGRPQGLQPS
1015	A	TGTAGACAAATACTGGGACA	TGCGGCCGCA	GGACAGCTACAACCATCCCT	TSEGCRCQILGHAAGQLQPS
1016	G	GTAGACAAATACTGGGACAG	TGCGGCCGCA	GACAGCTACAACCATCCCTT	TSEGCRCQILGCGRRQLGQPS
1017	C	TAGACAAATACTGGGACAGC	TGCGGCCGCA	ACAGCTACAACCATCCCTTC	TSEGCRCQILGRLPQGLQPS
1018	T	AGACAAATACTGGGACAGCT	TGCGGCCGCA	CAGCTACAACCATCCCTTCA	TSEGCRCQILGRLAAAILGQPS
1019	A	GACAAATACTGGGACAGCTA	TGCGGCCGCA	AGCTACAACCATCCCTTCAG	TSEGCRCQILGQLCGRKLPQSL
1020	C	ACAAATACTGGGACAGCTAC	TGCGGCCGCA	GCTACAACCATCCCTTCAGA	TSEGCRCQILGQLLRPQLQPS
1021	A	CAAATACTGGGACAGCTACA	TGCGGCCGCA	CTACAACCATCCCTTCAGAC	TSEGCRCQILGQLHAAAILGQPS
1022	A	AAATACTGGGACAGCTACAA	TGCGGCCGCA	TACAACCATCCCTTCAGACA	TSEGCRCQILGQLCGRKLPQSL
1023	C	AATACTGGGACAGCTACAAC	TGCGGCCGCA	ACAACCATCCCTTCAGACAG	TSEGCRCQILGQLLRPQPSLQ
1024	C	ATACTGGGACAGCTACAAC	TGCGGCCGCA	CAACCATCCCTTCAGACAGG	TSEGCRCQILGQLPAAAQPSLQ
1025	A	TACTGGGACAGCTACAACCA	TGCGGCCGCA	AACCATCCCTTCAGACAGGA	TSEGCRCQILGQLQPCGRKPSLQ
1026	T	ACTGGGACAGCTACAACCAT	TGCGGCCGCA	ACCATCCCTTCAGACAGGAT	TSEGCRCQILGQLQPLRPQPSLQ

1027	C	CTGGGACAGCTACAACCATC	TGCGGCCGCA	CCATCCCTTCAGACAGGATC	LGQLQPSAAAPSLQTG
1028	C	TGGGACAGCTACAACCATCC	TGCGGCCGCA	CATCCCTTCAGACAGGATCA	GQLQPSGCRSTSLQTGS
1029	C	GGGACAGCTACAACCATCCC	TGCGGCCGCA	ATCCCTTCAGACAGGATCAG	GQLQPSLRPQSLQTGS
1030	T	GGACAGCTACAACCATCCCT	TGCGGCCGCA	TCCCTTCAGACAGGATCAGA	GQLQPSLAAASLQTGS
1031	T	GACAGCTACAACCATCCCTT	TGCGGCCGCA	CCCTTCAGACAGGATCAGAA	QLQPSLCGRTLQTGSE
1032	C	ACAGCTACAACCATCCCTTC	TGCGGCCGCA	CCTTCAGACAGGATCAGAAG	QLQPSLLRPHLQTGSE
1033	A	CAGCTACAACCATCCCTTCA	TGCGGCCGCA	CTTCAGACAGGATCAGAAGA	QLQPSLHAAALQTGSE
1034	G	AGCTACAACCATCCCTTCAG	TGCGGCCGCA	TTCAGACAGGATCAGAAGAA	LQPSLQCGRITQGSSE
1035	A	GCTACAACCATCCCTTCAGA	TGCGGCCGCA	TCAGACAGGATCAGAAGAAC	LQPSLQMRPHQTGSEE
1036	C	CTACAACCATCCCTTCAGAC	TGCGGCCGCA	CAGACAGGATCAGAAGAACT	LQPSLQTAQAQTGSEE
1037	A	TACAACCATCCCTTCAGACA	TGCGGCCGCA	AGACAGGATCAGAAGAACTT	QPSLQTCGRKTGSEEL
1038	G	ACAACCATCCCTTCAGACAG	TGCGGCCGCA	GACAGGATCAGAAGAACTTA	QPSLQTVRPQTGSEEL
1039	G	CAACCATCCCTTCAGACAGG	TGCGGCCGCA	ACAGGATCAGAAGAACTTAG	QPSLQTAQAQTGSEEL
1040	A	AACCATCCCTTCAGACAGGA	TGCGGCCGCA	CAGGATCAGAAGAACTTAGA	PSLQTCGRITGSEELR
1041	T	ACCATCCCTTCAGACAGGAT	TGCGGCCGCA	AGGATCAGAAGAACTTAGAT	PSLQTLRPPQGSSEELR
1042	C	CCATCCCTTCAGACAGGATC	TGCGGCCGCA	GGATCAGAAGAACTTAGATC	PSLQTSAAAGSEELR
1043	A	CATCCCTTCAGACAGGATCA	TGCGGCCGCA	GATCAGAAGAACTTAGATCA	SLQTGSGRRSEELRS
1044	G	ATCCCTTCAGACAGGATCAG	TGCGGCCGCA	ATCAGAAGAACTTAGATCAT	SLQTGSRVPPQSEELRS
1045	A	TCCTTCAGACAGGATCAGA	TGCGGCCGCA	TCAGAAGAACTTAGATCATT	SLQTGSDAAAASEELRS
1046	A	CCCTTCAGACAGGATCAGAA	TGCGGCCGCA	CAGAAGAACTTAGATCATT	LQTGSECGRTEELRSL
1047	G	CCTTCAGACAGGATCAGAAG	TGCGGCCGCA	AGAAGAACTTAGATCATTAT	LQTGSEVRPQSEELRSL
1048	A	CTTCAGACAGGATCAGAAGA	TGCGGCCGCA	GAAGAACTTAGATCATTATA	LQTGSEDAAAAASEELRSL
1049	A	TTCAGACAGGATCAGAAGAA	TGCGGCCGCA	AAGAACTTAGATCATTATAT	QTGSEECRQKELRSLY
1050	C	TCAGACAGGATCAGAAGAAC	TGCGGCCGCA	AGAACTTAGATCATTATATA	QTGSEELRQELRSLY
1051	T	CAGACAGGATCAGAAGAACT	TGCGGCCGCA	GAACTTAGATCATTATATAA	QTGSEELAAAELRSLY
1052	T	AGACAGGATCAGAAGAACTT	TGCGGCCGCA	AACTTAGATCATTATATAAT	TGSEELCGRKLRSLYN
1053	A	GACAGGATCAGAAGAACTTA	TGCGGCCGCA	ACTTAGATCATTATATAATA	TGSEELMRPQLRSLYN
1054	G	ACAGGATCAGAAGAACTTAG	TGCGGCCGCA	CTTAGATCATTATATAATAC	TGSEELSAALRSLYN
1055	A	CAGGATCAGAAGAACTTAGA	TGCGGCCGCA	TTAGATCATTATATAATACA	GSEELRCGRIRSLYNT
1056	T	AGGATCAGAAGAACTTAGAT	TGCGGCCGCA	TAGATCATTATATAATACAA	GSEELRLRPHRSLYNT
1057	C	GGATCAGAAGAACTTAGATC	TGCGGCCGCA	AGATCATTATATAATACAAT	GSEELRSAAARSLYNT
1058	A	GATCAGAAGAACTTAGATCA	TGCGGCCGCA	GATCATTATATAATACAATA	SEELRSCGRRSLYNTI
1059	T	ATCAGAAGAACTTAGATCAT	TGCGGCCGCA	ATCATTATATAATACAATAG	SEELRSLRPQSLYNTI
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1061	A	CAGAAGAACTTAGATCATT	TGCGGCCGCA	CATTATATAATACAATAGCA	EELRSLCGRTLNTIA
1062	T	AGAAGAACTTAGATCATTAT	TGCGGCCGCA	ATTATATAATACAATAGCAG	EELRSLRPLQYNTIA
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1064	T	AAGAACTTAGATCATTATAT	TGCGGCCGCA	TATATAATACAATAGCAGTC	ELRSLYCGRIYNTIAV
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1068	A	ACTTAGATCATTATATAATA	TGCGGCCGCA	TAATACAATAGCAGTCTCT	LRLSYNMRPNTIAVL
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1070	A	TTAGATCATTATATAATACA	TGCGGCCGCA	ATACAATAGCAGTCTCTAT	RSLYNTCGRNTIAVLY
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1075	C	TCATTATATAATACAATAGC	TGCGGCCGCA	ATAGCAGTCTCTATTGTGT	SLYNTIAAAAIAVLYC
1076	A	CATTATATAATACAATAGCA	TGCGGCCGCA	TAGCAGTCTCTATTGTGTG	LYNTIACGRIVLYCV
1077	G	ATTATATAATACAATAGCAG	TGCGGCCGCA	AGCAGTCTCTATTGTGTGC	LYNTIIVRPQAVLYCV
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1079	C	TATATAATACAATAGCAGTC	TGCGGCCGCA	CAGTCTCTATTGTGTGCAT	YNTIAVCGRTVLYCVH
1080	C	ATATAATACAATAGCAGTCC	TGCGGCCGCA	AGTCTCTATTGTGTGCATC	YNTIAVLRPQVLYCVH
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1089	G	AATAGCAGTCTCTATTGTG	TGCGGCCGCA	TTGTGTGCATCAAAGGATAG	IAVLYCVRPHCVHQRI
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1092	C	AGCAGTCTCTATTGTGTGC	TGCGGCCGCA	TGTGCATCAAAGGATAGATG	AVLYCVLRPHVHQRID
1093	A	GCAGTCTCTATTGTGTGCA	TGCGGCCGCA	GTGCATCAAAGGATAGATGT	AVLYCVHAAAVHQRID
1094	T	CAGTCTCTATTGTGTGCAT	TGCGGCCGCA	TGCATCAAAGGATAGATGTA	VLYCVHCGRRMHQRIDV
1095	C	AGTCTCTATTGTGTGCATC	TGCGGCCGCA	GCATCAAAGGATAGATGTAA	VLYCVHLRPHQHQRIDV
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1097	A	TCCTCTATTGTGTGCATCAA	TGCGGCCGCA	ATCAAAGGATAGATGTAAAA	LYCVHQCGRRNQRIDVK
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1100	G	TCTATTGTGTGCATCAAAGG	TGCGGCCGCA	AAAGGATAGATGTAAAAAGC	YCVHQRCGRKRIDVKD
1101	A	CTATTGTGTGCATCAAAGGA	TGCGGCCGCA	AAGGATAGATGTAAAAAGACA	YCVHQRRMPQRIDVKD
1102	T	TATTGTGTGCATCAAAGGAT	TGCGGCCGCA	AGGATAGATGTAAAAAGCAC	YCVHQRIAAARIDVKD
1103	A	ATTGTGTGCATCAAAGGATA	TGCGGCCGCA	GGATAGATGTAAAAAGACACC	CVHQRCGRRIDVKDT
1104	G	TTGTGTGCATCAAAGGATAG	TGCGGCCGCA	GATAGATGTAAAAAGACCCA	CVHQRIVRPQIDVKDT
1105	A	TGTGTGCATCAAAGGATAGA	TGCGGCCGCA	ATAGATGTAAAAAGACCCAA	CVHQRIIDAAIDVKDT
1106	T	GTGTGCATCAAAGGATAGAT	TGCGGCCGCA	TAGATGTAAAAAGACCCAAAG	VHQRIDCGRIDVKDTK
1107	G	TGTGCATCAAAGGATAGATG	TGCGGCCGCA	AGATGTAAAAAGACCCAAAGG	VHQRIDVRPQDVVKDTK
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1110	A	GCATCAAAGGATAGATGTAA	TGCGGCCGCA	TGTAAAAAGACCCAAAGGAAG	HQRIDVMRPHVKDTKE
1111	A	CATCAAAGGATAGATGTAAA	TGCGGCCGCA	GTAAGAACCCAAAGGAAGCC	HQRIDVNAAAVKDTKE
1112	A	ATCAAAGGATAGATGTAAAA	TGCGGCCGCA	TAAAGAACCCAAAGGAAGCC	QRIDVKCGRIKDTKEA

1113	G	TCAAAGGATAGATGTAAGG	TGCGGCCGCA	AAAAGACACCAAGGAAGCCT	QRIDVKVRPQKDTKEA
1114	A	CAAAGGATAGATGTAAGG	TGCGGCCGCA	AAAGACACCAAGGAAGCCTT	QRIDVKDAAAKDTKEA
1115	C	AAAGGATAGATGTAAGGAC	TGCGGCCGCA	AAGACACCAAGGAAGCCTTA	RIDVKDCGRKDTKEAL
1116	A	AAGGATAGATGTAAGGACA	TGCGGCCGCA	AGACACCAAGGAAGCCTTAG	RIDVKDMRPQDTKEAL
1117	C	AGGATAGATGTAAGGACAC	TGCGGCCGCA	GACACCAAGGAAGCCTTAGA	RIDVKDAAAADTKEAL
1118	C	GGATAGATGTAAGGACACC	TGCGGCCGCA	ACACCAAGGAAGCCTTAGAT	IDVKDTCGRNTKEALD
1119	A	GATAGATGTAAGGACACCA	TGCGGCCGCA	CACCAAGGAAGCCTTAGATA	IDVKDTRMPPHTKEALD
1120	A	ATAGATGTAAGGACACCAA	TGCGGCCGCA	ACCAAGGAAGCCTTAGATAA	IDVKDNTAAATKEALD
1121	G	TAGATGTAAGGACACCAAG	TGCGGCCGCA	CCAAGGAAGCCTTAGATAAG	DVKDTCGRRTKEALDK
1122	G	AGATGTAAGGACACCAAGG	TGCGGCCGCA	CAAGGAAGCCTTAGATAAGA	DVKDTCVPRPHKALDK
1123	A	GATGTAAGGACACCAAGGA	TGCGGCCGCA	AAAGGAAGCCTTAGATAAGAT	DVKDTCDAAAKEALDK
1124	A	ATGTAAGGACACCAAGGAA	TGCGGCCGCA	AGGAAGCCTTAGATAAGATA	VKDTKECGRKEALDKI
1125	G	TGTAAGGACACCAAGGAAG	TGCGGCCGCA	GGAAGCCTTAGATAAGATAG	VKDTKEVPRQALDKI
1126	C	GTAAGGACACCAAGGAAGC	TGCGGCCGCA	GAAGCCTTAGATAAGATAGA	VKDTKEAAAALDKI
1127	C	TAAAGGACACCAAGGAAGCC	TGCGGCCGCA	AAGCCTTAGATAAGATAGAG	KDTCACGRKALDKIE
1128	T	AAAAGACACCAAGGAAGCCT	TGCGGCCGCA	AGCCTTAGATAAGATAGAGG	KDTCALRPPALDKIE
1129	T	AAAGACACCAAGGAAGCCTT	TGCGGCCGCA	GCCTTAGATAAGATAGAGGA	KDTCFAFAAALDKIE
1130	A	AAGACACCAAGGAAGCCTTA	TGCGGCCGCA	CCTTAGATAAGATAGAGGAA	DTKEALCGRHTDKIEE
1131	G	AGACACCAAGGAAGCCTTAG	TGCGGCCGCA	CTTAGATAAGATAGAGGAAG	DTKEALVRPHLDKIEE
1132	A	GACACCAAGGAAGCCTTAGA	TGCGGCCGCA	TTAGATAAGATAGAGGAAGA	DTKEALDAAALDKIEE
1133	T	ACACCAAGGAAGCCTTAGAT	TGCGGCCGCA	TAGATAAGATAGAGGAAGAG	TKEALDCGRIDKIEE
1134	A	CACCAAGGAAGCCTTAGATA	TGCGGCCGCA	AGATAAGATAGAGGAAGAGC	TKEALDMRPQDKIEE
1135	A	ACCAAGGAAGCCTTAGATAA	TGCGGCCGCA	GATAAGATAGAGGAAGAGCA	TKEALDAAAADKIEE
1136	G	CCAAGGAAGCCTTAGATAAG	TGCGGCCGCA	ATAAGATAGAGGAAGAGCAA	KEALDKCGRNKIEEEQ
1137	A	CAAGGAAGCCTTAGATAAGA	TGCGGCCGCA	TAAGATAGAGGAAGAGCAAA	KEALDKMRPHKIEEEQ
1138	T	AAGGAAGCCTTAGATAAGAT	TGCGGCCGCA	AAGATAGAGGAAGAGCAAAA	KEALDKIAAAKIEEEQ
1139	A	AGGAAGCCTTAGATAAGATA	TGCGGCCGCA	AGATAGAGGAAGAGCAAAAAC	EALDKICGRKIEEQN
1140	G	GGAAGCCTTAGATAAGATAG	TGCGGCCGCA	GATAGAGGAAGAGCAAAAACA	EALDKIVRPQIEEQN
1141	A	GAAGCCTTAGATAAGATAGA	TGCGGCCGCA	ATAGAGGAAGAGCAAAAACAA	EALDKIDAAAIEEQN
1142	G	AAGCCTTAGATAAGATAGAG	TGCGGCCGCA	TAGAGGAAGAGCAAAAACAAA	EALDKIEGRIEEQNK
1143	G	AGCCTTAGATAAGATAGAGG	TGCGGCCGCA	AGAGGAAGAGCAAAAACAAA	ALDKIEVRPQIEEQNK
1144	A	GCCTTAGATAAGATAGAGGA	TGCGGCCGCA	GAGGAAGAGCAAAAACAAAAG	ALDKIEDAAAIEEQNK
1145	A	CCTTAGATAAGATAGAGGAA	TGCGGCCGCA	AGGAAGAGCAAAAACAAAAGT	LDKIEECGRKEEQNKS
1146	G	CTTAGATAAGATAGAGGAAG	TGCGGCCGCA	GGAAGAGCAAAAACAAAAGTA	LDKIEEVRPQIEEQNKS
1147	A	TTAGATAAGATAGAGGAAGA	TGCGGCCGCA	GAAGAGCAAAAACAAAAGTAA	LDKIEEAAAIEEQNKS
1148	G	TAGATAAGATAGAGGAAGAG	TGCGGCCGCA	AAGAGCAAAAACAAAAGTAA	DKIEECCGRKEQNKSK
1149	C	AGATAAGATAGAGGAAGAGC	TGCGGCCGCA	AGAGCAAAAACAAAAGTAAAG	DKIEEELRPEQNKSK
1150	A	GATAAGATAGAGGAAGAGCA	TGCGGCCGCA	GAGCAAAAACAAAAGTAAAGAA	DKIEEHAIAIEEQNKSK
1151	A	ATAAGATAGAGGAAGAGCAA	TGCGGCCGCA	AGCAAAAACAAAAGTAAAGAAA	KIEEECCGRKQNKSKK
1152	A	TAAGATAGAGGAAGAGCAAA	TGCGGCCGCA	GCAAAAACAAAAGTAAAGAAA	KIEEEQMRPQNKSKK
1153	A	AAGATAGAGGAAGAGCAAAA	TGCGGCCGCA	CAAAAACAAAAGTAAAGAAAA	KIEEEQNAAAQNKSKK
1154	C	AGATAGAGGAAGAGCAAAAAC	TGCGGCCGCA	AAAACAAAAGTAAAGAAAAAG	IEEEQNCRKQNKSKK
1155	A	GATAGAGGAAGAGCAAAAACA	TGCGGCCGCA	AAACAAAAGTAAAGAAAAAGG	IEEEQNMRPQNKSKK
1156	A	ATAGAGGAAGAGCAAAAACAA	TGCGGCCGCA	AACAAAAGTAAAGAAAAAGGC	IEEEQNAAAANKSKK
1157	A	TAGAGGAAGAGCAAAAACAAA	TGCGGCCGCA	ACAAAAGTAAAGAAAAAGGCA	EEEQNKCGRNKSKKKA
1158	A	AGAGGAAGAGCAAAAACAAA	TGCGGCCGCA	CAAAAAGTAAAGAAAAAGGCAC	EEEQNKMRPHKSKKKA
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1160	T	AGGAAGAGCAAAAACAAAAGT	TGCGGCCGCA	AAAGTAAAGAAAAAGGCACAG	EEQNKSCGRKSKKKAQ
1161	A	GGAAGAGCAAAAACAAAAGTA	TGCGGCCGCA	AAGTAAAGAAAAAGGCACAGC	EEQNKSMRPQSKKKAQ
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1170	G	AAACAAAAGTAAAGAAAAAGG	TGCGGCCGCA	AAAGGCACAGCAAGCAGCAG	NKSKKVRPQKAQQAQA
1171	C	ACAAAAGTAAAGAAAAAGGC	TGCGGCCGCA	AGGCACAGCAAGCAGCAGC	NKSKKAAAQAQQAQA
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1173	C	CAAAAGTAAAGAAAAAGGCAC	TGCGGCCGCA	GGCACAGCAAGCAGCAGCTG	KSKKALRPQAQQAQA
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1175	G	AAAGTAAAGAAAAAGGCACAG	TGCGGCCGCA	CACAGCAAGCAGCAGCTGAC	SKKKAQCGRTQAQAAD
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1190	C	CACAGCAAGCAGCAGCTGAC	TGCGGCCGCA	CTGACACAGGAAACAACAGC	QQAADCGRTDTGNNS
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1192	C	CAGCAAGCAGCAGCTGACAC	TGCGGCCGCA	GACACAGGAAACAACAGCCA	QQAADTAAADTGNNS
1193	A	AGCAAGCAGCAGCTGACACA	TGCGGCCGCA	ACACAGGAAACAACAGCCAG	QQAADTCGRNTGNNSQ
1194	G	GCAAGCAGCAGCTGACACAG	TGCGGCCGCA	CACAGGAAACAACAGCCAGG	QQAADTVRPHDTGNNSQ
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1196	A	AAGCAGCAGCTGACACAGGA	TGCGGCCGCA	CAGGAAACAACAGCCAGGTC	AAADTGCGRNTGNNSQV
1197	A	AGCAGCAGCTGACACAGGAA	TGCGGCCGCA	AGGAAACAACAGCCAGGTCA	AAADTGMRPQGNNSQV
1198	A	GCAGCAGCTGACACAGGAAA	TGCGGCCGCA	GGAACAACAGCCAGGTGAC	AAADTGNAAAQGNNSQV

1199	C	CAGCAGCTGACACAGGAAAC	TGCGGCCGCA	GAACAACAGCCAGGTCAGC	AADTGNCRRNNSQVS
1200	A	AGCAGCTGACACAGGAAACA	TGCGGCCGCA	AAACAACAGCCAGGTCAGCC	AADTGNMRPQNSQVS
1201	A	GCAGCTGACACAGGAAACA	TGCGGCCGCA	AACAACAGCCAGGTCAGCCA	AADTGNNAANNSQVS
1202	C	CAGCTGACACAGGAAACAAC	TGCGGCCGCA	ACAACAGCCAGGTCAGCCAA	ADTGNCCGRNNSQVSQ
1203	A	AGCTGACACAGGAAACAACA	TGCGGCCGCA	CAACAGCCAGGTCAGCCAAA	ADTGNMRPHNSQVVSQ
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1205	C	CTGACACAGGAAACAACAGC	TGCGGCCGCA	ACAGCCAGGTCAGCCAAAAT	DTGNNSCGRNSQVVSQN
1206	C	TGACACAGGAAACAACAGCC	TGCGGCCGCA	CAGCCAGGTCAGCCAAAATT	DTGNNSLRPHSQVVSQN
1207	A	GACACAGGAAACAACAGCCA	TGCGGCCGCA	AGCCAGGTCAGCCAAAATTA	DTGNNSHAAASQVVSQN
1208	G	ACACAGGAAACAACAGCCAG	TGCGGCCGCA	GCCAGGTCAGCCAAAATTAC	TGNNSQVRRPHQVVSQNY
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1210	T	ACAGGAAACAACAGCCAGGT	TGCGGCCGCA	CAGGTCAGCCAAAATTACCC	GNNQVCGRKNYQVSNYP
1211	C	CAGGAAACAACAGCCAGGTC	TGCGGCCGCA	AGGTCAGCCAAAATTACCTT	GNNQVVMRPQVVSQNY
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1213	G	GGAAACAACAGCCAGGTCAG	TGCGGCCGCA	GTCAGCCAAAATTACCTTAT	NNSQVSCGRISQVSNYPI
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1234	A	CAAAAATTACCTTATAGTCCA	TGCGGCCGCA	GTCCAGAACCTCCAGGGGGCA	SQVSNLQVRRPHQVVSQNY
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1303	T	AATGCATGGGTA	TGCGGCCGCA	TAGTAGAAGAGAAGGCTTT	NAWVKVAAAANAWVKVVEEK
1304	A	ATGCATGGGTA	TGCGGCCGCA	TAGTAGAAGAGAAGGCTTTC	AWVKVVCGRIVVEEKAF
1305	G	TGCATGGGTA	TGCGGCCGCA	AGTAGAAGAGAAGGCTTTC	AWVKVVRPQVVEEKAF
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1307	A	CATGGGTA	TGCGGCCGCA	TAGAAGAGAAGGCTTCAGC	WVKVVECGRIEKEKAFS
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1327	A	GAGA	TGCGGCCGCA	AGCCAGAAGTAA	VVEEKAFSAAAFSPEVI
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1329	G	GA	TGCGGCCGCA	AGCCAGAAGTAA	VVEEKAFSAAAFSPEVI
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1332	A	GGCTTCAGCCAGA	TGCGGCCGCA	AGTAA	VVEEKAFSAAAFSPEVI
1333	T	GCTTCAGCCAGA	TGCGGCCGCA	GTAATACCCATGTTTCAGC	VVEEKAFSAAAFSPEVI
1334	A	CTTCAGCCAGA	TGCGGCCGCA	TAATACCCATGTTTCAGCA	VVEEKAFSAAAFSPEVI
1335	C	TTTCAGCCAGA	TGCGGCCGCA	AATACCCATGTTTCAGCAT	VVEEKAFSAAAFSPEVI
1336	C	TTCAGCCAGA	TGCGGCCGCA	ATACCCATGTTTCAGCATT	VVEEKAFSAAAFSPEVI
1337	C	TTCAGCCAGA	TGCGGCCGCA	TACCCATGTTTCAGCATT	VVEEKAFSAAAFSPEVI
1338	A	CAGCCAGA	TGCGGCCGCA	ACCCATGTTTCAGCATT	VVEEKAFSAAAFSPEVI
1339	T	AGCCAGA	TGCGGCCGCA	CCCATGTTTCAGCATT	VVEEKAFSAAAFSPEVI
1340	G	GCCAGA	TGCGGCCGCA	CCATGTTTCAGCATT	VVEEKAFSAAAFSPEVI
1341	T	CCAGA	TGCGGCCGCA	CATGTTTCAGCATT	VVEEKAFSAAAFSPEVI
1342	T	CCAGA	TGCGGCCGCA	ATGTTTCAGCATT	VVEEKAFSAAAFSPEVI
1343	T	CAGA	TGCGGCCGCA	TGTTTCAGCATT	VVEEKAFSAAAFSPEVI
1344	T	AGA	TGCGGCCGCA	GTTTCAGCATT	VVEEKAFSAAAFSPEVI
1345	C	GAAGTAATACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1346	A	AAGTAATACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1347	G	AGTAATACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1348	C	GTAATACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1349	A	TAATACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1350	T	AATACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1351	T	ATACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1352	A	TACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1353	T	ACCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1354	C	CCCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1355	A	CCATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1356	G	CATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1357	A	ATGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1358	A	TGTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1359	G	GTTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1360	G	TTTT	TGCGGCCGCA	TTTCAGCATT	VVEEKAFSAAAFSPEVI
1361	A	TTTCAGCATT	TGCGGCCGCA	AAGGAGCCACCCACAAGAT	SALSEGCGRKGATPQD
1362	G	TTTCAGCATT	TGCGGCCGCA	AGGAGCCACCCACAAGAT	SALSEVRPQGGATPQD
1363	C	TCAGCATT	TGCGGCCGCA	GGAGCCACCCACAAGATTT	SALSEGAAGGATPQD
1364	C	CAGCATT	TGCGGCCGCA	GAGCCACCCACAAGATTTA	ALSEGACGRRATPQDL
1365	A	AGCATT	TGCGGCCGCA	AGCCACCCACAAGATTTAA	ALSEGAMRPQATPQDL
1366	C	GCATT	TGCGGCCGCA	CCACCCACAAGATTTAAA	ALSEGATAAAATPQDL
1367	C	CATT	TGCGGCCGCA	CCACCCACAAGATTTAAAT	LSEGATCGRTTPQDLN
1368	C	ATT	TGCGGCCGCA	CACCCACAAGATTTAAATA	LSEGATLRPHTPQDLN
1369	C	TAT	TGCGGCCGCA	ACCCACAAGATTTAAATAC	LSEGATPAATPQDLN
1370	A	TAT	TGCGGCCGCA	CCCCACAAGATTTAAATACC	SEGATPCGRRTPQDLN

1371	C	ATCAGAAGGAGCCACCCAC	TGCGGCCGCA	CCCACAAGATTTAAATACCA	SEGATPLRHPQDLNT
1372	A	TCAGAAGGAGCCACCCACA	TGCGGCCGCA	CCACAAGATTTAAATACCAT	SEGATPHAAAPQDLNT
1373	A	CAGAAGGAGCCACCCACAA	TGCGGCCGCA	CACAAGATTTAAATACCATG	EGATPQCGRTQDLNTM
1374	G	AGAAGGAGCCACCCACAAG	TGCGGCCGCA	ACAAGATTTAAATACCATGC	EGATPQVRRPQDLNTM
1375	A	GAAGGAGCCACCCACAAGA	TGCGGCCGCA	CAAGATTTAAATACCATGCT	EGATPQDAAAQDLNTM
1376	T	AAGGAGCCACCCACAAGAT	TGCGGCCGCA	AAGATTTAAATACCATGCTA	GATPQDCGRKDLNTML
1377	T	AGGAGCCACCCACAAGATT	TGCGGCCGCA	AGATTTAAATACCATGCTAA	GATPQDLRPQDLNTML
1378	T	GGAGCCACCCACAAGATT	TGCGGCCGCA	GATTTAAATACCATGCTAAA	GATPQDFAAADLNTML
1379	A	GAGCCACCCACAAGATTTA	TGCGGCCGCA	ATTTAAATACCATGCTAAAC	ATPQDLCGRNLNTMLN
1380	A	AGCCACCCACAAGATTTAA	TGCGGCCGCA	TTTAAATACCATGCTAAACA	ATPQDLMRPHLNTMLN
1381	A	GCCACCCACAAGATTTAAA	TGCGGCCGCA	TTAAATACCATGCTAAACAC	ATPQDLNAAALNTMLN
1382	T	CCACCCACAAGATTTAAAT	TGCGGCCGCA	TAAATACCATGCTAAACACA	TPQDLNCGRINTMLNT
1383	A	CACCCACAAGATTTAAATA	TGCGGCCGCA	AAATACCATGCTAAACACAG	TPQDLNMRPQDLNTMLNT
1384	C	ACCCACAAGATTTAAATAC	TGCGGCCGCA	AATACCATGCTAAACACAGT	TPQDLNTAAANTMLNT
1385	C	CCCCACAAGATTTAAATACC	TGCGGCCGCA	ATACCATGCTAAACACAGTG	PQDLNLCGRINTMLNTV
1386	A	CCCACAAGATTTAAATACCA	TGCGGCCGCA	TACCATGCTAAACACAGTGG	PQDLNMRPHTMLNTV
1387	T	CCACAAGATTTAAATACCAT	TGCGGCCGCA	ACCATGCTAAACACAGTGGG	PQDLNTIAAATMLNTV
1388	G	CACAAGATTTAAATACCATG	TGCGGCCGCA	CCATGCTAAACACAGTGGGG	QDLNLMCGRHTMLNTVG
1389	C	ACAAGATTTAAATACCATGC	TGCGGCCGCA	CATGCTAAACACAGTGGGGG	QDLNLMRPHLNTMLNTVG
1390	T	CAAGATTTAAATACCATGCT	TGCGGCCGCA	ATGCTAAACACAGTGGGGGG	QDLNLMRPHLNTMLNTVG
1391	A	AAGATTTAAATACCATGCTA	TGCGGCCGCA	TGCTAAACACAGTGGGGGGA	DLNLMCGRHTMLNTVG
1392	A	AGATTTAAATACCATGCTAA	TGCGGCCGCA	GCTAAACACAGTGGGGGGAC	DLNLMRPHLNTMLNTVG
1393	A	GATTTAAATACCATGCTAAA	TGCGGCCGCA	CTAAACACAGTGGGGGGACA	DLNLMRPHLNTMLNTVG
1394	C	ATTTAAATACCATGCTAAAC	TGCGGCCGCA	TAAACACAGTGGGGGGACAT	LNTMLNCGRINTMLNTV
1395	A	TTTAAATACCATGCTAAACA	TGCGGCCGCA	AAACACAGTGGGGGGACATC	LNTMLNMRPONTVGGH
1396	C	TTAAATACCATGCTAAACAC	TGCGGCCGCA	AAACACAGTGGGGGGACATCA	LNTMLNAAANTVGGH
1397	A	TAAATACCATGCTAAACACA	TGCGGCCGCA	ACACAGTGGGGGGACATCAA	NMLNLCGRINTMLNTVG
1398	G	AAATACCATGCTAAACACAG	TGCGGCCGCA	CACAGTGGGGGGACATCAAG	NMLNLCGRINTMLNTVG
1399	T	AATACCATGCTAAACACAGT	TGCGGCCGCA	ACAGTGGGGGGACATCAAGC	NMLNLCGRINTMLNTVG
1400	G	ATACCATGCTAAACACAGTG	TGCGGCCGCA	CAGTGGGGGGACATCAAGCA	TMLNTVCAATVGGHQA
1401	G	TACCATGCTAAACACAGTGG	TGCGGCCGCA	AGTGGGGGGACATCAAGCAG	TMLNTVRRPQVGGHQA
1402	G	ACCATGCTAAACACAGTGGG	TGCGGCCGCA	GTGGGGGGACATCAAGCAGC	TMLNTVGAAVGGHQA
1403	G	CCATGCTAAACACAGTGGGG	TGCGGCCGCA	TGGGGGGACATCAAGCAGCC	MLNTVGCGRMGGHQA
1404	G	CATGCTAAACACAGTGGGGG	TGCGGCCGCA	GGGGGGACATCAAGCAGCCA	MLNTVGRPQGGHQA
1405	G	ATGCTAAACACAGTGGGGGG	TGCGGCCGCA	GGGGGGACATCAAGCAGCCAT	MLNTVGGAAAGGHQA
1406	A	TGCTAAACACAGTGGGGGGA	TGCGGCCGCA	GGGGACATCAAGCAGCCATG	LNTVGGCRRRHQAAM
1407	C	GCTAAACACAGTGGGGGGGAC	TGCGGCCGCA	GGGACATCAAGCAGCCATGC	LNTVGLRPQGHQAAM
1408	A	CTAAACACAGTGGGGGGACA	TGCGGCCGCA	GGACATCAAGCAGCCATGCA	LNTVGGHAAAGHQAAM
1409	T	TAAACACAGTGGGGGGACAT	TGCGGCCGCA	GACATCAAGCAGCCATGCAA	NTVGGHCGRRHQAAMQ
1410	C	AAACACAGTGGGGGGACATC	TGCGGCCGCA	ACATCAAGCAGCCATGCAA	NTVGGHLPQGHQAAMQ
1411	A	AAACACAGTGGGGGGACATCA	TGCGGCCGCA	CATCAAGCAGCCATGCAAAT	NTVGGHAAAHQAAMQ
1412	A	ACACAGTGGGGGGACATCAA	TGCGGCCGCA	ATCAAGCAGCCATGCAAATG	TVGGHCGRRHQAAMQM
1413	G	CACAGTGGGGGGACATCAAG	TGCGGCCGCA	TCAAGCAGCCATGCAAATGT	TVGGHQRPHQAAMQM
1414	C	ACAGTGGGGGGACATCAAGC	TGCGGCCGCA	CAAGCAGCCATGCAAATGTT	TVGGHQAAMQAAMQM
1415	A	CAGTGGGGGGACATCAAGCA	TGCGGCCGCA	AAGCAGCCATGCAAATGTTA	VGGHQAACGRKAAMQML
1416	G	AGTGGGGGGACATCAAGCAG	TGCGGCCGCA	AGCAGCCATGCAAATGTTAA	VGGHQAACGRKAAMQML
1417	C	GTGGGGGGACATCAAGCAGC	TGCGGCCGCA	GCAGCCATGCAAATGTTAAA	VGGHQAAMQAAMQML
1418	C	TGGGGGGACATCAAGCAGCC	TGCGGCCGCA	CAGCCATGCAAATGTTAAAA	GGHQAACGRKAAMQML
1419	A	GGGGGGACATCAAGCAGCCA	TGCGGCCGCA	AGCCATGCAAATGTTAAAAG	GGHQAAMRPAAMQMLK
1420	T	GGGGGACATCAAGCAGCCAT	TGCGGCCGCA	GCCATGCAAATGTTAAAAGA	GGHQAAMQAAMQMLK
1421	G	GGGGACATCAAGCAGCCATG	TGCGGCCGCA	CCATGCAAATGTTAAAAGAG	GHQAAMCGRTPQMLKE
1422	C	GGGACATCAAGCAGCCATGC	TGCGGCCGCA	CATGCAAATGTTAAAAGAGA	GHQAAMLRPHMQMLKE
1423	A	GGACATCAAGCAGCCATGCA	TGCGGCCGCA	ATGCAAATGTTAAAAGAGAC	GHQAAMHAAAMQMLKE
1424	A	GACATCAAGCAGCCATGCAA	TGCGGCCGCA	TGCAAATGTTAAAAGAGACC	HQAAMCGRTPQMLKET
1425	A	ACATCAAGCAGCCATGCAAA	TGCGGCCGCA	GCAAATGTTAAAAGAGACCA	HQAAMMRPQMLKET
1426	T	CATCAAGCAGCCATGCAAAT	TGCGGCCGCA	CAATGTTAAAAGAGACCAT	HQAAMQAAMQMLKET
1427	G	ATCAAGCAGCCATGCAAATG	TGCGGCCGCA	AAATGTTAAAAGAGACCATC	QAAMQMCGRKMLKETI
1428	T	CAAGCAGCCATGCAAATGT	TGCGGCCGCA	AATGTTAAAAGAGACCATCA	QAAMQMLRPQMLKETI
1429	T	CAAGCAGCCATGCAAATGTT	TGCGGCCGCA	ATGTTAAAAGAGACCATCAA	QAAMQMFAAAMMLKETI
1430	A	AAGCAGCCATGCAAATGTTA	TGCGGCCGCA	TGTTAAAAGAGACCATCAAT	AAMQMLCGRMLKETIN
1431	A	AGCAGCCATGCAAATGTTAA	TGCGGCCGCA	GTTAAAAGAGACCATCAATG	AAMQMLMRPQLKETIN
1432	A	GCAGCCATGCAAATGTTAAA	TGCGGCCGCA	TTAAAAGAGACCATCAATGA	AAMQMLNAAALKETIN
1433	A	CAGCCATGCAAATGTTAAAA	TGCGGCCGCA	TAAAAGAGACCATCAATGAG	AMQMLKCGRIKETINE
1434	G	AGCCATGCAAATGTTAAAAG	TGCGGCCGCA	AAAAGAGACCATCAATGAGG	AMQMLKVRPQKETINE
1435	A	GCCATGCAAATGTTAAAAGA	TGCGGCCGCA	AAAGAGACCATCAATGAGGA	AMQMLKDAAAKETINE
1436	G	CCATGCAAATGTTAAAAGAG	TGCGGCCGCA	AAGAGACCATCAATGAGGAA	MQMLKECGRKETINEE
1437	A	CATGCAAATGTTAAAAGAGA	TGCGGCCGCA	AGAGACCATCAATGAGGAAG	MQMLKEMRPQETINEE
1438	C	ATGCAAATGTTAAAAGAGAC	TGCGGCCGCA	GAGACCATCAATGAGGAAGC	MQMLKETAAETINEE
1439	C	TGCAAATGTTAAAAGAGACC	TGCGGCCGCA	AGACCATCAATGAGGAAGCT	QMLKETCGRKTINEEA
1440	A	GCAAATGTTAAAAGAGACCA	TGCGGCCGCA	GACCATCAATGAGGAAGCTG	QMLKETMRPQTINEEA
1441	T	CAAATGTTAAAAGAGACCAT	TGCGGCCGCA	ACCATCAATGAGGAAGCTGC	QMLKETIAAATINEEA
1442	C	AAATGTTAAAAGAGACCATC	TGCGGCCGCA	CCATCAATGAGGAAGCTGCA	MLKETICGRKTINEEA
1443	A	AATGTTAAAAGAGACCATCA	TGCGGCCGCA	CATCAATGAGGAAGCTGCAG	MLKETIMRPHINEEA
1444	A	ATGTTAAAAGAGACCATCAA	TGCGGCCGCA	ATCAATGAGGAAGCTGCAGA	MLKETINAAINEEA
1445	T	TGTTAAAAGAGACCATCAAT	TGCGGCCGCA	TCAATGAGGAAGCTGCAGAA	LKETINCGRINEEA
1446	G	GTTAAAAGAGACCATCAATG	TGCGGCCGCA	CAATGAGGAAGCTGCAGAA	LKETINVRPHINEEA
1447	A	TTAAAAGAGACCATCAATGA	TGCGGCCGCA	AATGAGGAAGCTGCAGAAATG	LKETINDAAINEEA
1448	G	TAAAAGAGACCATCAATGAG	TGCGGCCGCA	ATGAGGAAGCTGCAGAAATGG	KETINECGRINEEA
1449	G	AAAAGAGACCATCAATGAGG	TGCGGCCGCA	TGAGGAAGCTGCAGAAATGGG	KETINEVRPHEEA
1450	A	AAAGAGACCATCAATGAGGA	TGCGGCCGCA	GAGGAAGCTGCAGAAATGGGA	KETINEAAAINEEA
1451	A	AAGAGACCATCAATGAGGAA	TGCGGCCGCA	AGGAAGCTGCAGAAATGGGAT	ETINEECGRKEAAEWD
1452	G	AGAGACCATCAATGAGGAAAG	TGCGGCCGCA	GGAAGCTGCAGAAATGGGATA	ETINEEVRPHEEA
1453	C	GAGACCATCAATGAGGAAAGC	TGCGGCCGCA	GAAGCTGCAGAAATGGGATAG	ETINEEAAAINEEA
1454	T	AGACCATCAATGAGGAAAGCT	TGCGGCCGCA	AAGCTGCAGAAATGGGATAGT	TINEEACGRKAAEWD
1455	G	GACCATCAATGAGGAAAGCTG	TGCGGCCGCA	AGCTGCAGAAATGGGATAGAT	TINEEAIVRPAAEWDR
1456	C	ACCATCAATGAGGAAAGCTGC	TGCGGCCGCA	GCTGCAGAAATGGGATAGATT	TINEEAAAINEEA

1457	A	CCATCAATGAGGAAGCTGCA	TGCGGCCGCA	CTGCAGAATGGGATAGATTG	INEEAACGRTEAWDRL
1458	G	CATCAATGAGGAAGCTGCAG	TGCGGCCGCA	TGCAGAATGGGATAGATTGC	INEEAAVRPHAEWDRL
1459	A	ATCAATGAGGAAGCTGCAGA	TGCGGCCGCA	GCAGAATGGGATAGATTGCA	INEEAADAAAAEWDRL
1460	A	TCAATGAGGAAGCTGCAGAA	TGCGGCCGCA	CAGAATGGGATAGATTGCAT	NEEAACEGRTEWDRLH
1461	T	CAATGAGGAAGCTGCAGAAAT	TGCGGCCGCA	AGAATGGGATAGATTGCATC	NEEAALRPQEWDRLH
1462	G	AATGAGGAAGCTGCAGAAATG	TGCGGCCGCA	GAATGGGATAGATTGCATCC	NEEAEECAAAEWDRLH
1463	G	ATGAGGAAGCTGCAGAAATGG	TGCGGCCGCA	AATGGGATAGATTGCATCCA	EAAEAWCGRDRLHPV
1464	G	TGAGGAAGCTGCAGAAATGGG	TGCGGCCGCA	ATGGGATAGATTGCATCCAG	EAAEAWVRPQWDRLHP
1465	A	GAGGAAGCTGCAGAAATGGGA	TGCGGCCGCA	TGGGATAGATTGCATCCAGT	EAAEWDAAAADWDRLHP
1466	T	AGGAAGCTGCAGAAATGGGAT	TGCGGCCGCA	GGGATAGATTGCATCCAGTG	EAAEWDRCRRDRLHPV
1467	A	GAAGCTGCAGAAATGGGATA	TGCGGCCGCA	GGATAGATTGCATCCAGTGC	EAAEWDMPRQDRLHPV
1468	G	GAAGCTGCAGAAATGGGATAG	TGCGGCCGCA	GATAGATTGCATCCAGTGC	EAAEWDSSAADRLHPV
1469	A	AAGTGCAGAAATGGGATAGATA	TGCGGCCGCA	ATAGATTGCATCCAGTGCAT	AAEWDRCRRNRLHPVH
1470	T	AGTGCAGAAATGGGATAGAT	TGCGGCCGCA	TAGATTGCATCCAGTGCATG	AAEWDRLRPHRLHPVH
1471	T	GCTGCAGAAATGGGATAGATT	TGCGGCCGCA	AGATTGCATCCAGTGCATGC	AAEWDRLRPHRLHPVH
1472	G	CTGCAGAAATGGGATAGATTG	TGCGGCCGCA	GATTGCATCCAGTGCATGCA	AEWDRLCRRRLHPVHA
1473	C	TGCAGAAATGGGATAGATTGC	TGCGGCCGCA	ATTGCATCCAGTGCATGCA	AEWDRLLRPLQHPVHA
1474	A	GCAGAAATGGGATAGATTGCA	TGCGGCCGCA	TTCATCCAGTGCATGCAGG	AEWDRLHAAALHPVHA
1475	T	CAGAAATGGGATAGATTGCAT	TGCGGCCGCA	TGCATCCAGTGCATGCAGGG	EWDRLHCGRMHPVHAG
1476	C	AGAAATGGGATAGATTGCATC	TGCGGCCGCA	GCATCCAGTGCATGCAGGGC	EWDRLHLRPLQHPVHAG
1477	C	GAATGGGATAGATTGCATCC	TGCGGCCGCA	CATCCAGTGCATGCAGGGCC	EWDRLHPAAAHVPVHAG
1478	A	AATGGGATAGATTGCATCCA	TGCGGCCGCA	ATCCAGTGCATGCAGGGCCT	WDRLHPCGRNPVHAGP
1479	G	ATGGGATAGATTGCATCCAGT	TGCGGCCGCA	TCCAGTGCATGCAGGGCCTA	WDRLHPVRPHPVHAGP
1480	T	TGGGATAGATTGCATCCAGT	TGCGGCCGCA	CCAGTGCATGCAGGGCCTAT	WDRLHPVAAAPVHAGP
1481	G	GGGATAGATTGCATCCAGTG	TGCGGCCGCA	CAGTGCATGCAGGGCCTATT	DRLHPVCGRTVHAGPI
1482	C	GGATAGATTGCATCCAGTGC	TGCGGCCGCA	AGTGCATGCAGGGCCTATTG	DRLHPVLRPQVHAGPI
1483	A	GATAGATTGCATCCAGTGCAT	TGCGGCCGCA	TGTCATGCAGGGCCTATTGC	DRLHPVHAAAVHAGPI
1484	T	ATAGATTGCATCCAGTGCATG	TGCGGCCGCA	TGCATGCAGGGCCTATTGCA	RLHPVHCGRMHAGPIA
1485	G	TAGATTGCATCCAGTGCATG	TGCGGCCGCA	GCATGCAGGGCCTATTGCAC	RLHPVHVPRQVHAGPIA
1486	C	AGATTGCATCCAGTGCATGC	TGCGGCCGCA	CATGCAGGGCCTATTGCACC	RLHPVHAAAHAAGPIA
1487	A	GATTGCATCCAGTGCATGCA	TGCGGCCGCA	ATGCAGGGCCTATTGCACCA	LHPVHACGRNAGPIAP
1488	G	ATTGCATCCAGTGCATGCAG	TGCGGCCGCA	TGCAGGGCCTATTGCACCAG	LHPVHAVRPHAGPIAP
1489	G	TTCATCCAGTGCATGCAGG	TGCGGCCGCA	GCAGGGCCTATTGCACCAGG	LHPVHAGAAAAGPIAP
1490	G	TGCATCCAGTGCATGCAGGG	TGCGGCCGCA	CAGGGCCTATTGCACCAGGC	HPVHAGCGRTGPIAPG
1491	C	GCATCCAGTGCATGCAGGGC	TGCGGCCGCA	AGGGCCTATTGCACCAGGCC	HPVHAGLRPQGGPIAPG
1492	C	CATCCAGTGCATGCAGGGCC	TGCGGCCGCA	GGGCCTATTGCACCAGGCCA	HPVHAGPAAAGPIAPG
1493	T	ATCCAGTGCATGCAGGGCCT	TGCGGCCGCA	GGCCTATTGCACCAGGCCAG	PVHAGPCGRRRIAPGQ
1494	A	TCCAGTGCATGCAGGGCCTA	TGCGGCCGCA	GCCTATTGCACCAGGCCAGA	PVHAGPMRQPIAPGQ
1495	T	CCAGTGCATGCAGGGCCTAT	TGCGGCCGCA	CCTATTGCACCAGGCCAGAT	PVHAGPIAAPIAPGQ
1496	T	CAGTGCATGCAGGGCCTATT	TGCGGCCGCA	CTATTGCACCAGGCCAGATG	VHAGPICGRPIAPGQM
1497	G	AGTGCATGCAGGGCCTATTG	TGCGGCCGCA	TATTGCACCAGGCCAGATGA	VHAGPIVRPIAPGQM
1498	C	GTGCATGCAGGGCCTATTGC	TGCGGCCGCA	ATTGCACCAGGCCAGATGAG	VHAGPIAAAIAPIGQM
1499	A	TGCATGCAGGGCCTATTGCA	TGCGGCCGCA	TTGCACCAGGCCAGATGAGA	HAGPIACGRPIAPGQMR
1500	C	GCATGCAGGGCCTATTGCAC	TGCGGCCGCA	TGCACCAGGCCAGATGAGAG	HAGPIALRPHAPGQMR
1501	C	CATGCAGGGCCTATTGCACC	TGCGGCCGCA	GCACCAGGCCAGATGAGAGA	HAGPIAPAAAAPGQMR
1502	A	ATGCAGGGCCTATTGCACCA	TGCGGCCGCA	CACCAGGCCAGATGAGAGAA	AGPIAPCGRTPGQMR
1503	G	TGCAGGGCCTATTGCACCAG	TGCGGCCGCA	ACCAGGCCAGATGAGAGAA	AGPIAPVRPQPGQMR
1504	G	GCAGGGCCTATTGCACCAGG	TGCGGCCGCA	CCAGGCCAGATGAGAGAA	AGPIAPGAAAPGQMR
1505	C	CAGGGCCTATTGCACCAGGC	TGCGGCCGCA	CAGGCCAGATGAGAGAA	GPIAPCGRTPGQMR
1506	C	AGGGCCTATTGCACCAGGCC	TGCGGCCGCA	AGGCCAGATGAGAGAA	GPIAPGLRPQPGQMR
1507	A	GGGCCTATTGCACCAGGCCA	TGCGGCCGCA	GGCCAGATGAGAGAA	GPIAPGHAAGQMR
1508	G	GGCCTATTGCACCAGGCCAG	TGCGGCCGCA	CCAGATGAGAGAA	PIAPGQGRSQRGMR
1509	A	GCCTATTGCACCAGGCCAGA	TGCGGCCGCA	CCAGATGAGAGAA	PIAPGQMRPHQMR
1510	T	CCTATTGCACCAGGCCAGAT	TGCGGCCGCA	CAGATGAGAGAA	PIAPGQIAAQMREPR
1511	G	CTATTGCACCAGGCCAGATG	TGCGGCCGCA	AGATGAGAGAA	IAPGQMCGRKMRPRG
1512	A	TATTGCACCAGGCCAGATGA	TGCGGCCGCA	GATGAGAGAA	IAPGQMMRPMRPRG
1513	G	ATTGCACCAGGCCAGATGAG	TGCGGCCGCA	ATGAGAGAA	IAPGQMSAAAMRPRG
1514	A	TTGCACCAGGCCAGATGAGA	TGCGGCCGCA	TGAGAGAA	APGQMRGRMRPRGS
1515	G	TGCACCAGGCCAGATGAGAG	TGCGGCCGCA	GAGAGAA	APGQMRVRPQREPRGS
1516	A	GCACCAGGCCAGATGAGAGA	TGCGGCCGCA	AGAGAA	APGQMRDAAAREPRGS
1517	A	CACCAGGCCAGATGAGAGAA	TGCGGCCGCA	GAGAA	PGQMRGCRREPRGSD
1518	C	CCAGGCCAGATGAGAGAAC	TGCGGCCGCA	AGA	PGQMRRLRPEPRGSD
1519	C	ACAGGCCAGATGAGAGAA	TGCGGCCGCA	GA	PGQMRPAAAEPGSD
1520	A	CAGGCCAGATGAGAGAA	TGCGGCCGCA	AA	GQMRPEPCRPKPRGSDI
1521	A	AGGCCAGATGAGAGAA	TGCGGCCGCA	AA	GQMRPRPQPRGSDI
1522	G	GGCCAGATGAGAGAA	TGCGGCCGCA	CA	GQMRPESAAAPRGSDI
1523	G	GCCAGATGAGAGAA	TGCGGCCGCA	CA	QMRPEPCGRTRGSDIA
1524	G	CCAGATGAGAGAA	TGCGGCCGCA	AA	QMRPRVRPQPRGSDIA
1525	G	CAGATGAGAGAA	TGCGGCCGCA	AG	QMRPRGAAARGSDIA
1526	A	AGATGAGAGAA	TGCGGCCGCA	GG	MREPRGCGRRGSDIAG
1527	A	GATGAGAGAA	TGCGGCCGCA	GG	MREPRGMRPQSGDIAG
1528	G	ATGAGAGAA	TGCGGCCGCA	GG	MREPRGSAAGSDIAG
1529	T	TGAGAGAA	TGCGGCCGCA	GA	REPRGSCRRSDIAGT
1530	G	GAGAGAA	TGCGGCCGCA	AG	REPRGSSVRPQSDIAGT
1531	A	AGAGAA	TGCGGCCGCA	AG	REPRGSDAAASDIAGT
1532	C	GAGA	TGCGGCCGCA	AG	EPRGSDCGRSDIAGT
1533	A	AGA	TGCGGCCGCA	AG	EPRGSDMRPHDIAGT
1534	T	GA	TGCGGCCGCA	AG	EPRGSDIAAADIAGT
1535	A	A	TGCGGCCGCA	AG	PRGSDICGRNIAGTTS
1536	G	AA	TGCGGCCGCA	AG	PRGSDIVRPHIAGTTS
1537	C	AA	TGCGGCCGCA	AG	PRGSDIAAAIAGTTS
1538	A	AA	TGCGGCCGCA	AG	RGSDIACGRIAGTTS
1539	G	AA	TGCGGCCGCA	AG	RGSDIAVRPQAGTTS
1540	G	AA	TGCGGCCGCA	AG	RGSDIAGAAAAGTTS
1541	A	AA	TGCGGCCGCA	AG	GSDIACGRTGTTS
1542	A	AA	TGCGGCCGCA	AG	GSDIAGMRPQGTTS

1543	C	GGAAGTGACATAGCAGGAAC	TGCGGCCGCA	GGAACTACTAGT	GSDIAGTAAAGTTS
1544	T	GAAGTGACATAGCAGGAAC	TGCGGCCGCA	GAACTACTAGT	SDIAGTCGRRITS
1545	A	AAGTGACATAGCAGGAAC	TGCGGCCGCA	AACTACTAGT	SDIAGTMRPQTS
1546	C	AGTGACATAGCAGGAAC	TGCGGCCGCA	ACTACTAGT	SDIAGTTAAATTS
1547	T	GTGACATAGCAGGAAC	TGCGGCCGCA	CTACTAGT	DIAGTTCGRITS
1548	A	TGACATAGCAGGAAC	TGCGGCCGCA	TACTAGT	DIAGTTMRPHTS
1549	G	GACATAGCAGGAAC	TGCGGCCGCA	ACTAGT	DIAGTTSAAATS
1550	T	ACATAGCAGGAAC	TGCGGCCGCA	CTAGT	IAGTTSCGRITS



## Appendix C. Oligonucleotides used for genetic footprinting.

<u>Oligo Name</u>	<u>Sequence</u>	<u>T<sub>m</sub> (°C)</u>
HIV1	5'-ACATGTAGCCCCAGTTCTACTTACACC	80
HIV37	5'-TGGAAGGGCTAATTCACTCCCAAAG	74
HIV251	5'-GAGCCTGCATGGAATGGATG	62
HIV270r	5'-CATCCATTCCATGCAGGCTC	62
HIV361	5'-ACTGCTGACATCGAGCTTGC	62
HIV400r	5'-CCAGCGGAAAGTCCCTTGATGC	66
HIV492r	5'-CCCAGTACAGGCAAAAAGCAGC	65
HIV493	5'-TCTCTCTGGTTAGACCAGATCTG	63
HIV501	5'-GTTAGACCAGATCTGAGCCTGGG	66
HIV521	5'-GGGAGCTCTCTGGCTAACTAGGG	68
HIV591r	5'-TGAAGCACTCCCTCAAGGCAAGC	66
HIV592	5'-AGTAGTGTGTGCCCGTCTGTTG	65
HIV644r	5'-GGGTCTGAGGGATCTCTAGTTACC	74
HIV672r	5'-TGCTAGAGATTTTCCCACTGAC	66
HIV751	5'-GCGCACGGCAAGAGGCGAGG	71
HIV770r	5'-CCTCGCCTCTTGCCGTGCGC	71
HIV905r	5'-CTTTCCCCCTGGCCTTAACCG	68
HIV1027	5'-CCCTTCAGACAGGATCAGAAGAAC	65
HIV1224	5'-CCTATAGTCCAGAACCTCCAG	64
HIV1244r	5'-CTGGAGGTTCTGGACTATAGG	64
HIV1539	5'-GGA ACTACTAGTACCCTTCAGG	66
HIV1573r	5'-CATCCTATTTGTTCCCTGAAGGG	64

## Appendix D.

### Documentation for software developed for genetic footprinting



# Help for Louise and Marc's Footprinting Utilities



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## Overview

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The Footprinting Utilities are a set of tools to gather, manipulate, and present quantitative data from scanned gels using Excel and Matlab. From a set of footprinting gels and the sequence of the mutagenized DNA, you will be able to quantitatively assess band intensities, normalize data gathered from different gels, consolidate data from many spreadsheets into a single spreadsheet, and color code this data.

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## Scanning a Gel

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Your gel must be scanned:

- at 300 DPI.
- without auto levelling (contrast and brightness at 50%, or 125), and no enhancement (e.g. use DeskScan's "Black and White Photo," NOT "Sharp Black and White Picture").
- in 256 gray scale.

Save the gel image as a TIFF file. Include no more than 8 characters in the filename and make sure that the file has a ".tif" extension.

The gel should be scanned vertically. Matlab takes care of rotating the image to better fit the screen. Since gels are often larger than the window of the scanner, for better image quality, a weight should be put on top of the scanner's lid to properly push the gel against the glass. It is a good idea to crop the gel image as much as possible, as it will speed up the program. In cases where you have extremely large gel images, it may be worth saving half of the gel (i.e. the top half) in one file and the other half (i.e. the bottom half) in another file in the interest of speed. Using a gel image that is twice as big may slow down the program much more than two-fold.

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## Excel Utilities

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— **Tools:** The Excel utilities enable you to:

- Create Excel sheets to hold your data.
- Start the Matlab Quantitation Utility.
- Correct mistakes in band assignments from previous Matlab Quantitation Utility sessions.
- Consolidate data from many spreadsheets into a single spreadsheet.
- Color-code numerical data in Excel spreadsheets.
- Format data for the Normalization Utility.


— **Excel Sheets:** A Footprinting spreadsheet is composed of 2 sheets. Sheet 1 is **the summary sheet**. It displays the sequence of the target gene vertically. The position of each nucleotide is indicated in the column to the left of the target gene sequence. To the right of each nucleotide in the target sequence is written the structure of a mutant at that position, both in nucleotide form and in peptide form. Sheet 2 is **the data sheet**. It contains only the position numbers and target sequence until Matlab sends more data. Do not rename the sheets since Matlab sends the data to Sheet 2. Footprinting spreadsheets can be generated, saved, and reopened at a later time to add data using Matlab. Data can be entered to the same spreadsheet over several sessions. Simply open the appropriate


spreadsheet before starting the Matlab Quantitation Utility at the beginning of each session and save your data at the end of each session. A Footprinting spreadsheet must be open before using the Matlab Quantitation Utility.


—  **Help:** Launch a HTML browser with this help file.


—  **Make Footprinting Sheets:** To create a Footprinting spreadsheet, first open a blank Excel Workbook, then click on the icon. It will take you through the entire process using several prompts. You may choose to start the Matlab Quantitation Utility directly after creating this spreadsheet in order to select bands on a gel image.


—  **Select Peptide Reading Frame:** This icon allows you to change the translation frame for the peptides displayed on the summary sheet.

—  **Make Data Sheet:** This icon allows you to create only the second sheet (the data sheet) in case you don't want the summary sheet. You can enter data using Matlab Utilities in this data sheet.


—  **Start the Matlab Quantitation Utility:** This icon starts Matlab Quantitation Utility. You will be using the Matlab Quantitation Utility to select the bands you want to quantitate and to send the resulting data to the data sheet. Each time you click on this icon, you will start a new Matlab session. It is a good idea to have only one Matlab session open at a given time, so close the current session before opening a new one. Matlab Quantitation Utility are explained more fully below. The Matlab Quantitation Utility can be started directly from Matlab by typing "foot" in the Matlab Command Window.


—  **Change Number:** If you want to change the number of a band after it has been entered by Matlab, open the appropriate data sheet, select the cell with the nucleotide number you want to change from and click on this icon. You will be asked to enter the nucleotide number to which you want the data to move. This operation will move the data on the Excel data sheet from the old nucleotide position to the new position, as well as change the label of the band on the gel image.


—  **Regroup Data:** This icon allows you to import data from many spreadsheets into one spreadsheet. Open the destination spreadsheet (a new, blank sheet) and click on the icon. You will be asked to select a source file (Excel spreadsheet). The rows holding data will be imported into the destination spreadsheet. If you want to import data from several source files, do not click anywhere except on this icon to repeat the operation. This tool will only work with unmodified data sheets generated by Matlab Quantitation Utility as the source files.

—  **Paste Column:** This icon allows you to import columns from many spreadsheets into one spreadsheet. For example, a given nucleic acid sample (called

George) would typically be analyzed using several different primer pairs. The data for George would therefore end up on several data sheets. If you made sure that the data for George was always entered in a specific column (e.g. column B), you could use this tool to consolidate all of the data for George into a single spreadsheet. Open the destination spreadsheet and click anywhere in the column into which you wish to import data. Any data in this column will be erased. You will be asked which column number you wish to copy. In Excel, column headings are letters, so you must convert the letter of the desired column into a number (e.g. A=1, B=2, C=3,...). Next, you will be asked how many columns you wish to skip between pastes. If you wish to paste data into consecutive columns, enter "0" here. Finally, select the spreadsheet from which you wish to copy a column of data. You can select several source spreadsheets in succession. The same column number will be accessed for each spreadsheet. When you have finished, click the 'Cancel' button. This tool will work using any type of Excel spreadsheet as the source file.

 **Color Code Numerical Data:** This icon color codes cell values on a 56-shade grayscale. White is assigned to the number 1 and black is assigned to the numbers 100 and higher. You may scale your data as you wish to fit this scale. Highlight the cells holding the data you wish to color code and click on the icon. Unfortunately, due to a Microsoft bug, using this tool will change the entire color scheme of the current spreadsheet!

 **Formatting for the Normalization Utility:** To use the Matlab Normalization Utility, you must get your data into the proper format. Paste the values for your data into the upper left-hand corner of a blank Excel worksheet (to paste only values and not formulae, use Edit | Paste Special... | "as Text"). Eliminate any non-data information (e.g. data labels, nucleotide position numbers, column headings) -- the normalization program will try to normalize anything you give it. Your data should be organized such that a given column contains data from a single gel and a given row contains data for a given nucleotide position. Highlight the region of the spreadsheet where you have data. Hit the "NaN" function icon. NaN ("Not a Number") will appear in all the blank cells in the area where you have your data. Save your worksheet as "Text -- Tab-delimited." A ".txt" extension should automatically appear on your file. This specific extension is required by the Matlab Normalization Utility. Put the files you want to normalize into a dedicated folder. Do not put extraneous files into this folder -- the normalization utility will try to normalize them. You will not mess up your files, but the utility will crash.

 **Start the Matlab Normalization Utility:** This icon starts the Matlab Normalization Utility. The Matlab Normalization Utility can be started directly from Matlab by typing "normalize" in the Matlab Command Window.

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## The Matlab Quantitation Utility

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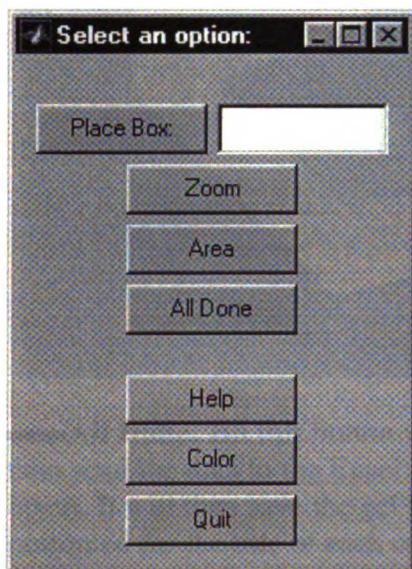
Number of Bands per Set  
Main Dialog Screen  
Figure Menu

Create or open the appropriate spreadsheet in Excel and click on this icon .

### Number of Bands per Set

Enter the number of bands that you wish quantitate per nucleotide position, or hit Cancel to simply view the gel (the Zoom will then automatically be turned on).

### Main Dialog Screen



**Place Box:** Enter the nucleotide position number in the edit box. Then press 'Return', or click 'Place Box'. The pointer becomes a long crosshair, indicating that you can select the first box of a set. Click on the band you are interested in. Two boxes and a number will appear:

- The first box around the band will remain; it indicates the area where data is taken.
- The second box shows the area used to determine the background and will disappear.
- A number will temporarily appear; this number is the value of the data read (with background subtracted). It will disappear when you select the next band.

At this point you may decide to keep this data by selecting the next band, or delete it (giving you a chance to re-select it) by pressing 'Enter'.

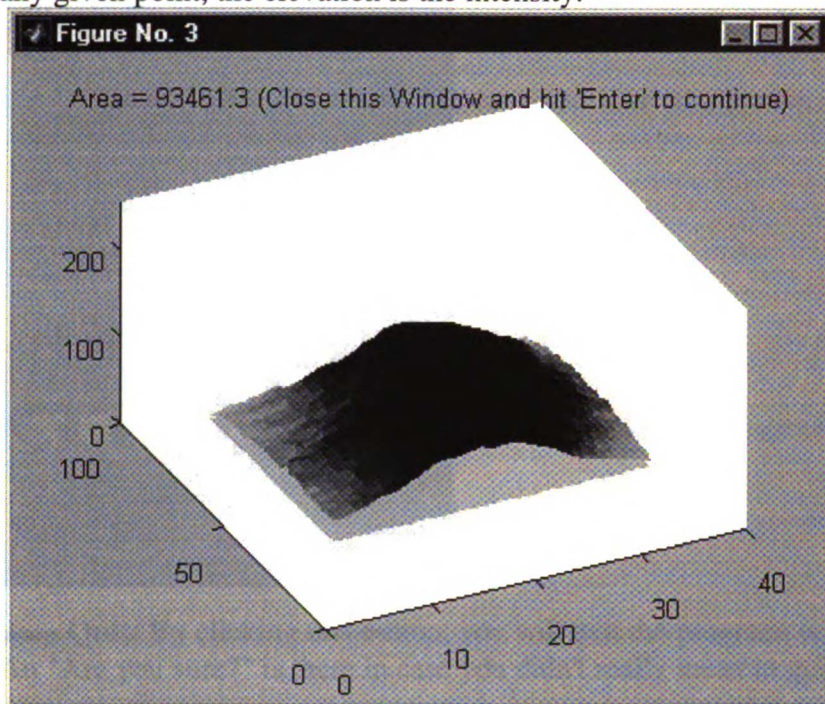
When you reach the last band at a given position, you will receive a "last band" prompt. You can either keep the data by clicking anywhere on the figure, or delete it and re-select it by pressing 'Enter'.

Every time you hit the 'Enter' key, you will delete the data from one band, starting from the most recent band selected and proceeding backwards in time. However, you can only delete data at the nucleotide position where you are placing bands. Once you make the final click after the "last band" prompt, you cannot go back and delete data for that position using the 'Enter' key. You can always "Quit" without saving and start all over again.

**Zoom:** When the zoom is enabled, the cursor icon changes and you can Left-click to zoom in, Right-click to zoom out, or drag a box around the area you want to zoom. When you have zoomed to the level of magnification you want to work with, press 'Enter'.

**Area:** You probably won't be using this tool. It calculates the volume that lies

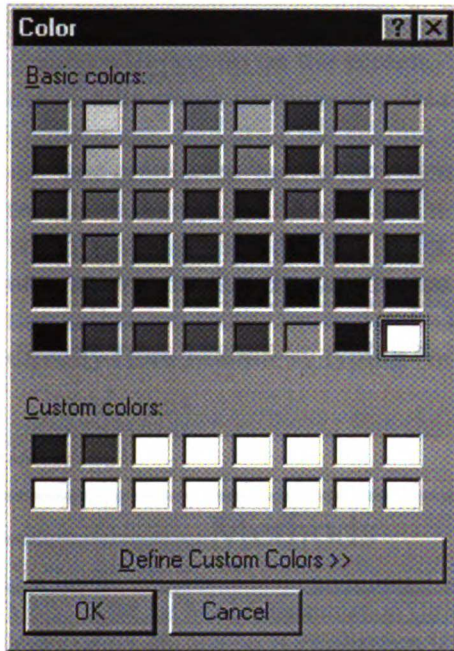
between the plane defined by the background and the surface defined by the band. At any given point, the elevation is the intensity.



— **All Done:** Hit this button when you are finished entering data for your gel. It will then send the data to the Excel spreadsheet (the correct spreadsheet should already be open). It will then save the gel file with the boxes and position numbers. You will be prompted for the title of each column (if no titles are desired, or the titles are already in the spreadsheet, just hit 'Cancel'). Make sure to save the Excel spreadsheet with your new data before closing it.

— **Help:** This will start a Web Browser if one is not already running and display this document.

— **Color:** Click here to set the color you wish for the boxes and the text on the gel.



— **Quit:** By clicking this button, you will exit the program, without saving anything. An "Are you sure?" is there in case you didn't really mean to quit without saving.

## — Figure Menu

— **Print:** Click here to print the current gel image, including boxed and labelled bands. To resume or finish your quantitation session, find the "Select an option" button on the taskbar at the bottom of your screen and click on it to reactivate the Main Dialog Screen.

— **Colormap:** Click here to open another window displaying the current gel image next to a colorbar showing the color scheme used to pseudocolor the gel image. To resume or finish your quantitation session, close this colorbar window, find the "Select an option" button on the taskbar at the bottom of your screen, and click on it to reactivate the Main Dialog Screen.

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## — The Matlab Normalization Utility

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This tool allows you to merge data for the same nucleic acid sample between different gels or different exposures of the same gel.

First, make sure your data is in the proper format and in a dedicated folder, as described in the section on Formating for the Normalization Utility. Then start the Matlab Normalization Utility. You will be asked to select any file in the dedicated folder. The Normalization Utility will proceed to normalize every file in that folder. The program will run for a while, perhaps a long while if you are comparing many gels (we have



normalized up to sixty gels in one .txt file). When the program is finished, a series of graphs will pop up on the screen. The top graph displays the data before normalization, the middle graph displays the data after one round of normalization, and the bottom graph displays the data after the second round of normalization. The highlighted blue curves, one per graph, represents the calculated average of the curves in that graph. Your real results will consist of a series of normalization factors, and are saved in ".res" files which will appear in the dedicated folder. To get your normalized data, you multiply the original values (i.e. the values you read off of the gel image) for a given gel by the normalization factor for that gel. The Matlab Normalization Utility can be started directly from Matlab by typing "normalize" in the Matlab Command Window.

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## Methods

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[Methods for the Matlab Quantitation Utility](#)

[Methods for the Matlab Normalization Utility](#)

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### Methods for the Matlab Quantitation Utility

**Pseudocolors:** The scanned gel is a large 2-dimensional matrix, where each element of the matrix represents a pixel location and holds a number between 1 and 256 indicating the intensity of the gray. For pseudocolor, the image (= matrix) is searched for the biggest of the 10 smallest values of gray and the smallest of the 10 biggest values of gray. This gives us a good low and high boundaries for the gray present in the image. Then, a custom-made colormap (= set of 256 different colors) is stretched to fit exactly between the two boundaries. The use of this tailored pseudocoloring scheme helps when selecting bands on the gel. To see the colormap of a given gel, start the Matlab Utility and at the "Number of Bands per Set" prompt hit 'Cancel'. The menu should then have a 'ColorMap' option. Values that are outside the dynamic range of the film are colored bright red to indicate saturation.

#### Data Values:



For each band, an area around the band (60 X 10 pixels) is considered. In this area, the darkest 50 pixels are averaged to give the raw reading. The background value (see below) is then subtracted to give the data. It is a number between 1 (light) and 256 (dark). This value is temporarily displayed on the gel as the user selects bands.

#### Background Subtraction:



For background subtraction, a longer and narrower area than the one used for collecting the data is considered (30 X 160 pixels). This area is divided into 16 vertical strips (30 X 10 pixels). For each strip, the darkest 50 pixels are averaged. The lowest of these 16 averages is considered the value of the background. The background value is subtracted from the value read in the Data Area (above).

## Methods for the Matlab Normalization Utility

You may wish to merge data for the same nucleic acid sample between different gels or different exposures of the same gel. As you might expect, the normalization algorithm works better if you have more positions in common between gels. The goal of the algorithm is to minimize the weighted sum of the coefficients of variation for each position. First, the Normalization Utility pre-processes the data in three ways:

- It eliminates values that exceed the "maxgrey" value. Values that are beyond the dynamic range of the film are meaningless.
- It eliminates nucleotide positions that contain only one value.
- It eliminates gels that contain only one value.

Next, the average value at each nucleotide position is calculated. These average values (the Starting Averages for round 1) are kept throughout the first round of normalization. At a given nucleotide position, a weighted coefficient of variation is calculated using the values for the data points as well as the Starting Average. The Starting Average is assigned a weight that is equal to the total number of data points at this position. For example, suppose you have three data points ( $x_1$ ,  $x_2$ ,  $x_3$ ) at position 637, with a Starting Average of AV.  $x_1$ ,  $x_2$ , and  $x_3$  are each given a weight of 1, while AV is given a weight of 3. A weighted sum of the weighted coefficients of variation is taken. The weighted coefficient of variation at a given position is assigned a weight according to the number of data points (not including the synthetic "average value" data points) present at that position. A position with two data points is given a weight of 2, a position with three data points is given a weight of 3, and so on. The Normalization Utility tries by iteration to minimize this weighted sum by adjusting each gel. The adjustment is achieved by multiplying the data from each gel by a factor (a different factor for each gel, but the same factor for all data points within a gel). We stop the iteration process when the variation in the weighted sum is less than the "precision" value or after a defined number ("iteration") of iterations has been performed.

A new set of average values is calculated using the normalization factors from the first round of normalization. These average values (the Starting Averages for round 2) are kept throughout the second round of normalization. The second round of normalization is carried out exactly like the first round.

Your results will consist of a series of normalization factors, one for each gel (note that if a gel was eliminated during pre-processing, it will not receive a normalization factor). To

get your normalized data, you multiply the original values (i.e. the values you read off of the gel image) for a given gel by the normalization factor for that gel.

The default values for "**maxgrey**", "**precision**", and "**iteration**" are 110, 0.001, and 10000, respectively. You can modify these values. For example, type "**maxgrey = 120**" in the Matlab Command Window to set "**maxgrey**" to 120. Your modifications will not be saved between Matlab sessions. To verify the current values for these properties, type "**maxgrey**", "**precision**", or "**iteration**" in the Matlab Command Window (case-sensitive).

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## Appendix E. Code for software developed for genetic footprinting

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CODE FOR MATLAB FOORPRINTING UTILITIES

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FILE FOOT.M

---

```
function Foot
%
% Utility for gathering data on scanned gels for foot printing.
%

% Initialize
global Dir

try
    cd(Dir.GelDir)
catch
    h = msgbox('The Default Gel Directory path is wrong.  Edit
    ''Startup.m'' to correct the path. ');
    drawnow
    waitfor(h)
end
clear S
global S
S.Data = [];
S.Image = [];
O = NaN;
ZoomPointer = [ ...
0 0 0 1 1 1 1 1 0 0 0 0 0 0 0 0;...
0 1 1 2 2 2 2 2 1 1 0 0 0 0 0 0;...
0 1 2 2 1 1 1 2 2 1 0 0 0 0 0 0;...
1 2 2 1 0 0 0 1 2 2 1 0 0 0 0 0;...
1 2 1 0 0 0 0 0 1 2 1 0 0 0 0 0;...
1 2 1 0 0 1 0 0 1 2 1 0 0 0 0 0;...
1 2 1 0 0 0 0 0 1 2 1 0 0 0 0 0;...
1 2 2 1 0 0 0 1 2 2 1 0 0 0 0 0;...
1 1 2 2 1 1 1 2 2 1 0 0 0 0 0 0;...
0 1 1 2 2 2 2 2 1 1 1 0 0 0 0 0;...
0 0 0 1 1 1 1 1 0 1 1 1 0 0 0 0;...
0 0 0 0 0 0 0 0 0 0 1 1 1 0 0 0;...
0 0 0 0 0 0 0 0 0 0 0 1 1 1 0;...
0 0 0 0 0 0 0 0 0 0 0 0 1 1 1;...
0 0 0 0 0 0 0 0 0 0 0 0 0 1 1];

% Open Gel
% Get a file
```

```

[ImFile, ImPath] = uigetfile('*.tif');
if max(ImFile) == 0
    return
end
warning off
ImFile = [ImPath, ImFile];
im = imread(ImFile, 'tiff');
warning on
% Put it horizontal and show it
S.Image = rot90(im);
imshow(S.Image)

S.FigH = gcf;
S.AxeH = gca;

SetBestColorMap

% Add print to menu, and add setup for pretty printing
set(S.FigH, 'NumberTitle', 'off')
set(S.FigH, 'Name', ImFile)
set(S.FigH, 'Color', [1 1 1], ...
    'PaperOrientation', 'Landscape')
set(S.FigH, 'PaperUnits', 'inches');
PaperSize = get(S.FigH, 'PaperSize');
PaperPos=[0.3 0.3 (PaperSize(1)-.3) (PaperSize(2)-0.3)];
set(S.FigH, 'PaperPosition', PaperPos);
h1 = uimenu('Parent',S.FigH, ...
    'Callback','prindlg', ...
    'Label','&Print', ...
    'Tag','MnPrint');
h1 = uimenu('Parent',S.FigH, ...
    'Callback','Colors', ...
    'Label','&ColorMap', ...
    'Tag','MnColors');

% Try to load previous lines if needed
[LineFile,rem] = strtok(ImFile, '.');
LineFile = strcat(LineFile, '.fig');
EvalStr = ['hgload(''',LineFile, ''');'];
eval(EvalStr, '');

% Get general info about gel
figure(S.FigH)
drawnow
% Play sound
try
    [a,b,c] = wavread('Utopia Windows Start.wav');
    playsnd(a,b,c);
catch
end

List = {'2','3','4','5','6','7','8','9',...
    '10','11','12','13','14','15','16','17','18','19',...
    '20','21','22','23','24','25','26','27','28','29',...
    '30','31','32','33','34','35','36','37','38','39','40'};
[NumCol,v] = listdlg('PromptString','How many lines per set?',...
    'SelectionMode','single',...

```

```

        'ListString',List);
S.NumCol = NumCol+1;
if v == 0
    set(S.FigH, 'Pointer', 'custom', 'PointerShapeCData', ZoomPointer,
'PointerShapeHotSpot', [6 6])
    zoom
    return
end

i = 0;
while(1)
    drawnow
    S.Ret = 'PBQuit';
    DlgH = FgWhatNext;
    CenterFigure(S.FigH, DlgH);
    waitfor(DlgH)
    if S.Ret == 'PBDone'
        % Get out
        break
    elseif S.Ret == 'PBQuit'
        % Make sure and quit
        ButtonName=questdlg('Do you really want to quit WITHOUT saving
?', 'Are you nuts?', ...
        'Quit','No','No');
        if strcmp(ButtonName, 'Quit')
            set(S.FigH, 'Pointer', 'arrow')
            return
        end
    elseif S.Ret == 'PBColo'
        TheColor = uisetcolor;
        if size(TheColor,2) == 3
            Child = get(gca, 'Children');
            for i = 1 : size(Child)-1
                set(Child(i), 'Color', TheColor)
            end
        end
    elseif S.Ret == 'PBArea'
        GetArea
    elseif S.Ret == 'PBHelp'
        % show help
        set(S.FigH, 'Pointer', 'watch')
        try
            web(Dir.HelpPage)
        catch
            h = msgbox('The Help Page path is wrong. Edit ''Startup.m''
to correct the path. ');
            drawnow
            waitfor(h)
        end
        set(S.FigH, 'Pointer', 'arrow')
    elseif S.Ret == 'PBZoom'
        % Zoom
        set(S.FigH, 'Name', [ImFile, ' - Zoom'])
        set(S.FigH, 'Pointer', 'custom', 'PointerShapeCData',
ZoomPointer, 'PointerShapeHotSpot', [6 6])
        zoom on
        Key = 0;

```

```

while Key == 0
    Key = waitforbuttonpress;
end
zoom off
set(S.FigH, 'Pointer', 'arrow')
set(S.FigH, 'Name', ImFile)
elseif S.Ret == 'EdNumb'
    % Do it!
    i = i+1;
    iMax = i;
    S.Data(i).Place = str2num(S.EdNumber);
    set(S.FigH, 'Name', [ImFile, ' - ', S.EdNumber])

    j = 1;
    LastAction = 'Put';
    while j <= S.NumCol+1
        [x, y] = ginput(1);
        if isempty(x)
            if (j > 1)
                if (j == S.NumCol+1)
                    % Remove 2 boxes and text
                    NumChildren = 10;
                elseif (LastAction ~= 'Rmv')
                    % Remove 2 boxes
                    NumChildren = 9;
                else
                    % Remove 1 box
                    NumChildren = 4;
                end
                end
                % Last was an error, erase (Enter was hit)
                Children = get(S.AxeH, 'children');
                delete(Children(1: NumChildren))
                LastAction = 'Rmv';
                j = j -1;
            else
                S.Data(i) = [];
                i = i -1;
                break
            end
        else
            if (j ~= S.NumCol+1)
                if (j ~= 1)
                    % Remove line across from previous
                    if (LastAction ~= 'Rmv')
                        Children = get(S.AxeH, 'children');
                        delete(Children(1:5))
                    end
                end
                LastAction = 'Put';
                if (j == S.NumCol)
                    text(x+2, y-30, S.EdNumber, ...
                        'Color', 'w', ...
                        'Rotation', 90, ...
                        'FontWeight', 'bold');
                end
                % Start getting data
                x = round(x);
            end
        end
    end
end

```

```

        y = round(y);
        S.Data(i).BoxVal(j) = BoxTopAverage(S.Image, x, y);
        text(x+10, y, num2str(S.Data(i).BoxVal(j)), ...
            'Color', 'w', ...
            'FontWeight', 'bold');
        if (S.Data(i).BoxVal(j) < 0.0)
            h = msgbox('Value < 0 !!!');
            drawnow
            waitfor(h)
            drawnow
            S.Data(i).BoxVal(j) = 0;
        end
    else
        % Remove line across from previous
        Children = get(S.AxeH, 'children');
        delete(Children(1:5))
    end
    j = j+1;
    % Tell that this was the last one to give a chance to
delete it
    if j == S.NumCol+1
        h = msgbox('Last one.', '', 'custom', S.Image, S.ColorMap);
        drawnow
        pause(1)
        try
            delete(h)
        catch
        end
        drawnow
    end
end
end
end
[SFile,rem] = strtok(ImFile, '.');
SFile = strcat(SFile, '.mat');
save(SFile, 'S')
end

% Save the picture under bitmap format with colormap
%[BmpFile,rem] = strtok(ImFile, '.');
%BmpFile = strcat(BmpFile, '.bmp');
%imwrite(S.Image, S.ColorMap, BmpFile, 'bmp')

if ~isempty(S.Data)
    %Check that spreadsheet is open
    try
        Channel = ddeinit('excel', 'book1.xls:Sheet2');
        DNASTart = ddereq(Channel, 'r3c1:r3c1');
        ddeterm(Channel);
    catch
        msg = sprintf('Your Excel Spreadsheet doesn't seem to be open.
\nOpen it FIRST and THEN press OK. ');
        h = msgbox(msg, 'Error', 'Error');
        drawnow
        waitfor(h)
    end
end

```



```

% Save the S structure
[SFile,rem] = strtok(ImFile, '.');
SFile = strcat(SFile, '.mat');
save(SFile, 'S')

% Save the lines
ChildH = get(S.AxeH, 'Children');
LinesH = ChildH(1 : size(ChildH, 1)-1);
hgsave(LinesH, LineFile);

% Send it to excel
% Get the right page in Excel
Channel = ddeinit('excel', 'book1.xls:Sheet2');
DNASStart = ddereq(Channel, 'r3c1:r3c1');
ddeterm(Channel);
% Put file name at top of spreadsheet
Channel = ddeinit('excel', 'book1.xls:Sheet2');
ddepoke(Channel, 'r1c4:r1c4', ImFile);
ddeterm(Channel);

% Send the data, first
for i = 1 : iMax
    disp(S.Data(i).Place)
    for j = 1 : S.NumCol
        Row = num2str(S.Data(i).Place - DNASStart + 3);
        Col = num2str(2 + (4*(j-1)) + 2);
        ColPlus1 = num2str(2 + (4*(j-1)) + 2 + 1);
        CellStr = ['r', Row, 'c', Col, ':r', Row, 'c', Col];
        Channel = ddeinit('excel', 'book1.xls:Sheet2');
        ddepoke(Channel, CellStr, S.Data(i).BoxVal(j));
        ddeterm(Channel);
    end
end
% Then format Spread sheet column
for i = 1 : S.NumCol
    Tmp = inputdlg(['Enter the title for column ', num2str(i), ' on
the Excel Spreadsheet'], ...
    'Cool Title', 1);
    if isempty(Tmp)
        Tmp = '';
    elseif isempty(Tmp{1,:})
        Tmp = '';
    else
        Tmp = Tmp{1,:};
    end
    Channel = ddeinit('excel', 'book1.xls:Sheet2');
    ddepoke(Channel, 'r1c1:r1c1', i);
    ddeterm(Channel);
    Channel = ddeinit('excel', 'book1.xls:Sheet2');
    ddepoke(Channel, 'r1c2:r1c2', Tmp);
    ddeterm(Channel);
    Channel = ddeinit('excel', 'book1.xls:Sheet2');
    ddeexec(Channel, '[run("''Foot Printing.xls''!FormatGelCol")]');
    ddeterm(Channel);
end
end
warning off

```

```
[im, map] = imread('face.tif');
warning on
msgbox('All done','Clara dit:','custom',im, map)
set(S.FigH, 'Name', ImFile)
```

---

## FILE STARTUP.M

---

```
iptsetpref('ImshowBorder', 'tight')
iptsetpref('ImshowTruesize', 'manual')
set(0, 'DefaultFigureMenuBar','none')
set(0, 'DefaultFigurePosition',[2, 70, 1022, 657])
set(0, 'DefaultFigureInvertHardCopy', 'on')
global S
global Dir

% Edit following if you change the directories
Dir.GelDir = 'd:\data';
Dir.HelpPage = 'd:\foot printing\help\foothelp.htm';
Dir.CodeDir = 'd:\foot printing\code matlab';

try
    cd(Dir.CodeDir);
catch
    h = msgbox('The Footprinting Code path is wrong. Edit ''Startup.m''
to correct the path.');
```

```
    drawnow
    waitfor(h)
end

disp(' ')
disp(' Type ''Foot'' to start the Footprinting utility.');
```

```
disp(' ')

Setup % for gel curve fitting
```

---

## FILE SETUP.M

---

```
global V
global sV
global Curves
global Points

precision = 1.e-3;
iteration = 10000;
maxgrey = 110;
```

---

## FILE FGWHATNEXT.M

---

```
function fig = FgWhatNext()
% This is the machine-generated representation of a Handle Graphics
object
% and its children. Note that handle values may change when these
objects
% are re-created. This may cause problems with any callbacks written to
% depend on the value of the handle at the time the object was saved.
%
% To reopen this object, just type the name of the M-file at the MATLAB
% prompt. The M-file and its associated MAT-file must be on your path.

load FgWhatNext

h0 = figure('Color',[0.8 0.8 0.8], ...
    'Colormap',mat0, ...
    'MenuBar','none', ...
    'Name','Select an option:', ...
    'NumberTitle','off', ...
    'PointerShapeCData',mat1, ...
    'Position',[503 205 195 254], ...
    'Tag','Fig1');
h1 = uicontrol('Parent',h0, ...
    'Units','points', ...
    'BackgroundColor',[1 1 1], ...
    'Callback','FgWhatNextGUI EdNumber', ...
    'HorizontalAlignment','left', ...
    'ListboxTop',0, ...
    'Position',[75 153.75 63.75 18.75], ...
    'Style','edit', ...
    'Tag','EdNumber');
h1 = uicontrol('Parent',h0, ...
    'Units','points', ...
    'Callback','FgWhatNextGUI PBNumber', ...
    'ListboxTop',0, ...
    'Position',[7.5 153.75 63.75 18.75], ...
    'String','Place Box:', ...
    'Tag','PBNumber');
h1 = uicontrol('Parent',h0, ...
    'Units','points', ...
    'Callback','FgWhatNextGUI PBZoom', ...
    'ListboxTop',0, ...
    'Position',[41.25 131.25 63.75 18.75], ...
    'String','Zoom', ...
    'Tag','PBZoom');
h1 = uicontrol('Parent',h0, ...
    'Units','points', ...
    'Callback','FgWhatNextGUI PBDone', ...
    'ListboxTop',0, ...
    'Position',[41.25 86.25 63.75 18.75], ...
    'String','All Done', ...
    'Tag','PBDone');
h1 = uicontrol('Parent',h0, ...
```

```

    'Units','points', ...
    'Callback','FgWhatNextGUI PBQuit', ...
    'ListboxTop',0, ...
    'Position',[41.25 7.5 63.75 18.75], ...
    'String','Quit', ...
    'Tag','PBQuit');
h1 = uicontrol('Parent',h0, ...
    'Units','points', ...
    'Callback','FgWhatNextGUI PBHelp', ...
    'ListboxTop',0, ...
    'Position',[41.25 52.5 63.75 18.75], ...
    'String','Help', ...
    'Tag','PBHelp');
h1 = uicontrol('Parent',h0, ...
    'Units','points', ...
    'Callback','FgWhatNextGUI PBColor', ...
    'ListboxTop',0, ...
    'Position',[41.25 30 63.75 18.75], ...
    'String','Color', ...
    'Tag','PBColor');
h1 = uicontrol('Parent',h0, ...
    'Units','points', ...
    'Callback','FgWhatNextGUI PBArea', ...
    'ListboxTop',0, ...
    'Position',[41.25 108.75 63.75 18.75], ...
    'String','Area', ...
    'Tag','PBArea');
if nargin > 0, fig = h0; end

```

---

## FILE FGWHATNEXTGUI.M

---

```

function FgWhatNextGUI(action)
global S
%
% Callback's for FgWhatNext GUI page
%

FigH =(gcf;

switch action

case {'EdNumber', 'PBNumber'}
    S.EdNumber = get(findobj('tag', 'EdNumber'), 'string');
    if isempty(S.EdNumber) | max(isletter(S.EdNumber))
        MsgBox('Please enter a number.');
```

```

        set(findobj('tag', 'EdNumber'), 'string', '');
    else
        S.Ret = 'EdNumb';
        delete(FigH)
    end
end

case 'PBZoom'
```

```

        S.Ret = 'PBZoom';
        delete(FigH)

case 'PBDone'
    S.Ret = 'PBDone';
    delete(FigH)

case 'PBArea'
    S.Ret = 'PBArea';
    delete(FigH)

case 'PBColor'
    S.Ret = 'PBColo';
    delete(FigH)

case 'PBHelp'
    S.Ret = 'PBHelp';
    delete(FigH)

case 'PBQuit'
    S.Ret = 'PBQuit';
    delete(FigH)

otherwise
    msgbox('ERROR...')
    S.Ret = 'ERROR_';
end

```

---

## FILE WAITSCREEN.M

---

```

function fig = WaitScreen()
% This is the machine-generated representation of a Handle Graphics
object
% and its children. Note that handle values may change when these
objects
% are re-created. This may cause problems with any callbacks written to
% depend on the value of the handle at the time the object was saved.
%
% To reopen this object, just type the name of the M-file at the MATLAB
% prompt. The M-file and its associated MAT-file must be on your path.

load WaitScreen

h0 = figure('Color',[0.8 0.8 0.8], ...
    'Colormap',mat0, ...
    'MenuBar','none', ...
    'Name','Please wait while MATLAB updates its data...', ...
    'NextPlot','replacechildren', ...
    'NumberTitle','off', ...
    'PointerShapeCData',mat1, ...
    'Position',[273 265 377 257], ...
    'Tag','Fig1');

```

```

h1 = axes('Parent',h0, ...
'Box','on', ...
'CameraUpVector',[0 1 0], ...
'Color',[1 1 1], ...
'ColorOrder',mat2, ...
'DataAspectRatioMode','manual', ...
'Layer','top', ...
'Position',[0 0 1 1], ...
'Tag','Axes1', ...
'TickDir','out', ...
'TickDirMode','manual', ...
'Visible','off', ...
'WarpToFill','off', ...
'XColor',[0 0 0], ...
'XLim',[0.5 417.5], ...
'XLimMode','manual', ...
'YColor',[0 0 0], ...
'YDir','reverse', ...
'YLim',[0.5 284.5], ...
'YLimMode','manual', ...
'ZColor',[0 0 0]);
h2 = image('Parent',h1, ...
'BusyAction','cancel', ...
'CData',mat3, ...
'Interruptible','off', ...
'Tag','Axes1Image1', ...
'XData',[1 417], ...
'YData',[1 284]);
h2 = text('Parent',h1, ...
'Color',[0 0 0], ...
'HandleVisibility','off', ...
'HorizontalAlignment','center', ...
'Position',[207.890625 -7.265625 2523.127598358505], ...
'Tag','Axes1Text4', ...
'VerticalAlignment','bottom');
set(get(h2,'Parent'),'Title',h2);
h2 = text('Parent',h1, ...
'Color',[0 0 0], ...
'HandleVisibility','off', ...
'HorizontalAlignment','center', ...
'Position',[207.890625 315.5625 2523.127598358505], ...
'Tag','Axes1Text3', ...
'VerticalAlignment','cap');
set(get(h2,'Parent'),'XLabel',h2);
h2 = text('Parent',h1, ...
'Color',[0 0 0], ...
'HandleVisibility','off', ...
'HorizontalAlignment','center', ...
'Position',[-39.50000000000001 143.609375 2523.127598358505], ...
'Rotation',90, ...
'Tag','Axes1Text2', ...
'VerticalAlignment','baseline');
set(get(h2,'Parent'),'YLabel',h2);
h2 = text('Parent',h1, ...
'Color',[0 0 0], ...
'HandleVisibility','off', ...
'HorizontalAlignment','right', ...

```

```

    'Position',[-0.67187500000001 1.609375 2523.127598358505], ...
    'Tag','Axes1Text1', ...
    'Visible','off');
set(get(h2,'Parent'),'ZLabel',h2);
if nargin > 0, fig = h0; end

```

---

## FILE CENTERFIGURE.M

---

```

function CenterFigure(MainFgH, NewFgH)
global S
%
% Centers the new figure in the previous(MainFgH) figure
% or center on screen if no previous figure(MainFgH = 0)
%
MainUnits = get(MainFgH, 'Units');
NewUnits = get(NewFgH, 'Units');
set(NewFgH, 'Units', MainUnits);
if (MainFgH == 0)
    % Center on screen
    MainPos = get(MainFgH, 'ScreenSize');
else
    MainPos = get(MainFgH, 'Position');
end
NewPos = get(NewFgH, 'Position');
set(NewFgH, 'Position', ...
    [MainPos(1) + (MainPos(3)/2) - (NewPos(3)/2) ...
    MainPos(2) + (MainPos(4)/2) - (NewPos(4)/2) ...
    NewPos(3) ...
    NewPos(4)])
set(NewFgH, 'Units', NewUnits);

```

---

## FILE ZOOM.M

---

This Matlab file (Revision: 5.34 Date: 1997/12/02 21:08:55) was modified:

```

Line 226:
if isempty(state),
    %ML added
    LocPointer = get(fig, 'Pointer');
    %ML end Added
    state = uisuspend(fig);
    setuprop(fig,'ZOOMfigureState',state);
end
%ML changed
%set(fig,'windowbuttondownfcn','zoom down', ...
% 'windowbuttonupfcn','ones;', ...

```

```

% 'windowbuttonmotionfcn','', 'buttondownfcn','', ...
% 'interruptible','on');
set(fig,'windowbuttondownfcn','zoom down', ...
    'windowbuttonupfcn','ones;', ...
    'windowbuttonmotionfcn','', 'buttondownfcn','', ...
    'Pointer',LocPointer, ...
    'interruptible','on');
%ML end Changed

```

Line 371:

```

%
% Actual zoom operation
%
%ML added
LocPointer = get(fig, 'Pointer');
set(fig, 'Pointer', 'watch')
%ML end Changed

```

Line 445 (end of function):

```

%ML Added
drawnow
set(fig, 'Pointer', LocPointer)
%ML end added

```

---

## FILE SETBESTCOLORMAP.M

---

```

function SetBestColorMap
global S
%
% Sets the best color map based on the picture
%

%Min = double(min(min(S.Image)));
%Max = double(max(max(S.Image)));
Tmp = sort(double(min(S.Image)));
Min = Tmp(5);
Tmp = sort(double(max(S.Image)));
Max = Tmp(size(Tmp,2)-5);
Total = Max-Min;

Half = round(Total/2);
OtherHalf = Total - Half;

OneQuarter = round(OtherHalf/2);
OtherQuarter = OtherHalf - OneQuarter;

Bleu2Black = [];
for i =1 : OtherQuarter
    Bleu2Black(i) = (i-1)*(.5625/OtherQuarter);
end
Bleu2Black = Bleu2Black';
Bleu2Black = [zeros(OtherQuarter,1), zeros(OtherQuarter,1),
Bleu2Black];

```



```

Satur = 10;
TmpMap = bone(OneQuarter-Satur);
for i =1 : OneQuarter-Satur
    Black2White(i,:) = TmpMap(OneQuarter-Satur-i+1,:);
end

BeforePad = [ones(Min, 1), ones(Min, 1), ones(Min, 1)];
AfterPad = [ones(256-Max, 1)*.5, zeros(256-Max, 1), zeros(256-Max, 1)];
Saturation = [ones(Satur, 1)*1, ones(Satur, 1)*.0, ones(Satur, 1)*0];
%for i = 1 : Satur
%   Saturation(i,:) = [1, ((i-1)/(Satur-1))^2, ((i-1)/(Satur-1))^2];
%end

GelMap = [BeforePad; Saturation; Black2White; Bleu2Black; jet(Half);
AfterPad];

set(S.FigH, 'ColorMap', GelMap)

S.ColorMap = GelMap;

```

---

## FILE COLORS.M

---

```

global S
figure
imshow(S.Image)
colormap(S.ColorMap)
colorbar

```

---

## FILE RECT.M

---

```

function rect(x, y)
%
% draw a rectangle from x(1),y(1) to x(2),y(2)
%

line([x(1), x(1)],[y(1), y(2)], 'Color', 'w')
line([x(1), x(2)],[y(2), y(2)], 'Color', 'w')
line([x(2), x(2)],[y(2), y(1)], 'Color', 'w')
line([x(2), x(1)],[y(1), y(1)], 'Color', 'w')

```

---

## FILE BOXTOPAVERAGE.M

---

```

function BoxRet = BoxTopAverage(im, x,y)

```

```

global S
%
% Return the average of the 10 most dark points in a rectangle around
x,y
% minus the background color (== 5 brightest points on line across)

% Average and box
HalfH = 30;
HalfW = 5;
rect([x+HalfW, x-HalfW], [y+HalfH, y-HalfH])

Tmp = im(y-HalfH : y+HalfH, x-HalfW : x+HalfW);
Tmp = reshape(Tmp, size(Tmp,1)*size(Tmp,2),1);
Tmp = sort(double(Tmp));
BoxAverage = 255 -(sum(Tmp(1:50))/50);

% Background and line
HalfW = 80;
HalfH = 15;
XMinus = x-HalfW;
XPlus = x+HalfW;
YMinus = y-HalfH;
YPlus = y+HalfH;
Size1 = size(im,1);
Size2 = size(im,2);
if XMinus < 0
    XPlus = XPlus - XMinus;
    XMinus = 1;
elseif XPlus > Size2
    XMinus = XMinus - (XPlus - Size2);
    XPlus = Size2;
end
if YMinus < 0
    YPlus = XPlus - XMinus;
    YMinus = 1;
elseif YPlus > Size1
    YMinus = YMinus - (YPlus - Size1);
    YPlus = Size1;
end

rect([XPlus, XMinus], [YPlus, YMinus])
Incr = HalfW*2/16;
for i = 1 : 16
    Tmp1(i).Tmp = im(YMinus : YPlus, XMinus +((i-1)*Incr) : XMinus
+(i*Incr));
    Tmp1(i).Tmp = reshape(Tmp1(i).Tmp,
size(Tmp1(i).Tmp,1)*size(Tmp1(i).Tmp,2),1);
    Tmp1(i).Tmp = sort(double(Tmp1(i).Tmp));
    BackGround(i) = 255 -(sum(Tmp1(i).Tmp(1:50))/50);
end

BackGround = min(BackGround);

BoxRet = BoxAverage - BackGround;

```

## FILE GETAREA.M

---

```
function GetArea
global S
%
% Calculates the integral under the curve
%

%clear c3
%clear P3
%clear P1
%clear BackGround
%clear Tmp1

[x,y] = ginput(1);

% Average and box
HalfH = 30;
HalfW = 5;
%rect([x+HalfW, x-HalfW], [y+HalfH, y-HalfH])
warning off
Tmp = S.Image(y-HalfH : y+HalfH, x-HalfW : x+HalfW);
Tmp = reshape(Tmp, size(Tmp,1)*size(Tmp,2),1);
Tmp = sort(double(Tmp));
BoxAverage = 255 - (sum(Tmp(1:50))/50);
warning on
% Background and line
HalfW = 80;
HalfH = 15;
XMinus = x-HalfW;
XPlus = x+HalfW;
YMinus = y-HalfH;
YPlus = y+HalfH;
Size1 = size(S.Image,1);
Size2 = size(S.Image,2);
if XMinus < 0
    XPlus = XPlus - XMinus;
    XMinus = 1;
elseif XPlus > Size2
    XMinus = XMinus - (XPlus - Size2);
    XPlus = Size2;
end
if YMinus < 0
    YPlus = XPlus - XMinus;
    YMinus = 1;
elseif YPlus > Size1
    YMinus = YMinus - (YPlus - Size1);
    YPlus = Size1;
end

warning off
%rect([XPlus, XMinus], [YPlus, YMinus])
Incr = HalfW*2/16;
```

```

for i = 1 : 16
    Tmp1(i).Tmp = S.Image(YMinus : YPlus, XMinus +((i-1)*Incr) : XMinus
+(i*Incr));
    Tmp1(i).Tmp = reshape(Tmp1(i).Tmp,
size(Tmp1(i).Tmp,1)*size(Tmp1(i).Tmp,2),1);
    Tmp1(i).Tmp = sort(double(Tmp1(i).Tmp));
    BackGround(i) = 255 -(sum(Tmp1(i).Tmp(1:50))/50);
end
warning on
BackGround = min(BackGround);

BoxRet = BoxAverage - BackGround;

ii = 0;
xx = 15;
x = round(x);
y = round(y);
TheArea = 0;
for i = y-30 : y+30
    ii = ii + 1;
    c3t(ii).Data = 255-double(S.Image(i, x-xx:x+xx));
    c3tB(ii).Data = 255-double(S.Image(i, x-xx:x+xx))-BackGround;
    for j = 1 : size(c3tB(ii).Data,2)
        if c3tB(ii).Data(j) < 0
            c3tB(ii).Data(j) = 0;
        end
    end
    TheArea = TheArea + trapz(c3tB(ii).Data);
end
rect([x+xx, x-xx], [y+30, y-30])
TheArea
for i = 1 : ii
    P3(i,:) = c3t(i).Data;
end
scrsz = get(0,'ScreenSize');
figure('Position',[20 20 500 500])
surface(P3, 'linestyle', 'none')
set(gca, 'CLim', [1, 256])
set(gca, 'ZLim', [0,255])
title(['Area = ', num2str(TheArea), ' (Close this Window and hit
''Enter'' to continue)']);
try
    for i = 1 : size(S.ColorMap)
        TmpMap(i,:) = S.ColorMap(257-i,:);
    end
    set(gcf, 'ColorMap', TmpMap)
catch
end
view([-26, 46])
hold
P1 = ones(size(P3,1),size(P3, 2))*BackGround;
surface(P1, 'linestyle', 'none')

pause
Children = get(S.AxeH, 'children');
delete(Children(1:4))

```

---

FILE CHANGENUMBER.M

---

```
function ChangeNumber
global S
%
%
%

hh = WaitScreen;
drawnow

Data=1:64;Data=(Data'*Data)/64;

FigH = figure;
set(FigH, 'visible', 'off')
set(FigH, 'Pointer', 'watch')

%Get the file name
Channel = ddeinit('excel', 'book1.xls:Sheet2');
GelFile = ddereq(Channel, 'r1c4:r1c4', [1,1]);
ddeterm(Channel);
%Get the Old Number
Channel = ddeinit('excel', 'book1.xls:Sheet2');
OldNumber = ddereq(Channel, 'r1c1:r1c1');
ddeterm(Channel);
%Get the New Number
Channel = ddeinit('excel', 'book1.xls:Sheet2');
NewNumber = ddereq(Channel, 'r1c2:r1c2');
ddeterm(Channel);

% Try to load previous lines if needed
[LineFile,rem] = strtok(GelFile, '.');
LineFile = strcat(LineFile, '.fig');
Child = hgload(LineFile);

% Check that you are not overwriting a set of data
for i = 1 : size(Child)
    if strcmp(get(Child(i), 'Type'), 'text')
        Num = str2num(get(Child(i), 'String'));
        if Num == NewNumber
            close(hh);
            ButtonName=questdlg(['Do you really want to overwrite the ',
num2str(NewNumber), ' box ?'], ...
                'Yo!!!', ...
                'Yes', 'No', 'No');
            switch ButtonName,
            case 'Yes',
                hh = WaitScreen
                % Keep on going
                break
            case 'No',
                % Clean worksheet
```

```

        Channel = ddeinit('excel', 'book1.xls:Sheet2');
        ddepoke(Channel, 'r1c1:r1c1', '');
        ddeterm(Channel);
        Channel = ddeinit('excel', 'book1.xls:Sheet2');
        ddepoke(Channel, 'r1c2:r1c2', '');
        ddeterm(Channel);
        % End program
        exit
        return
    end
    break
end
end
end

Changed = 0;
for i = 1 : size(Child)
    if strcmp(get(Child(i), 'Type'), 'text')
        Num = str2num(get(Child(i), 'String'));
        if Num == OldNumber
            set(Child(i), 'String', num2str(NewNumber))
            Changed = 1;
            break
        end
    end
end
end

if Changed == 1
    hgsave(Child, LineFile);
    % Play sound
    try
        [a,b,c] = wavread('Utopia Critical Stop.wav');
        playsnd(a,b,c);
    catch
        end
        h = msgbox(['Changed : ', num2str(OldNumber), ' to ',
num2str(NewNumber), '.'], 'Yo!!!', 'custom', Data, hot(64));
        close(hh);
    else
        % Play sound
        try
            [a,b,c] = wavread('Robotz Error.wav');
            playsnd(a,b,c);
        catch
            end
            h = msgbox(['Did not find ', num2str(OldNumber), ' in file ',
LineFile, '.'], 'Yo!!!', 'custom', Data, hot(64));
            close(hh);
        end

% Clean worksheet
Channel = ddeinit('excel', 'book1.xls:Sheet2');
ddepoke(Channel, 'r1c1:r1c1', '');
ddeterm(Channel);
Channel = ddeinit('excel', 'book1.xls:Sheet2');
ddepoke(Channel, 'r1c2:r1c2', '');

```

```
ddeterm(Channel);  
uiwait(h);  
exit
```

---

#### FILE FOOTHELP.M

---

```
%  
% Start Netscape with help file  
%  
global Dir  
web(Dir.HelpPage)  
exit
```

---

#### FILE NORMALIZE.M

---

```
global V  
global File  
  
cd('D:\Data\excel sheets\source')  
[File, Path] = uigetfile('*..*', 'Select any file in the Directory');  
D = dir(Path);  
n = size(D, 1);  
for i = 3 : n  
    File = strcat(Path, D(i).name);  
    disp(['Working on ', File])  
    drawnow  
    V = load(File);  
    V = V';  
    h = msgbox('Keep Ctrl-C down for 10 sec. to stop.');
```

```
    drawnow  
    %pause(10)  
    try  
        delete(h)  
    catch  
    end  
    drawnow  
    NormCode  
end  
warning off  
[im, map] = imread('face.tif');  
warning on  
msgbox('All done', 'Clara dit:', 'custom', im, map)
```

---

## FILE NORMFUNC.M

---

```
function y = NormalizeFunc(v)
%
% Just if you try to read this code...
%   a(p) is the average at position p
%   V(c,p) is the value of curve c, position p
%   sV is a binary representation of V (NaN or Value => 0 or 1)
%   v is the coef to move curves up or down (changed to minimize this
function)
%

global V
global sV
global a
global Curves
global Points

Means = zeros(Points,1);
Sigma = zeros(Points,1);
n = sum(sV);
%vv = zeros(Points,Points);
for p = 1 : Points
    % Mean
    Means(p) = (sum(v'.*V(:,p)) + n(p)*a(p)) / (2*n(p));
end
for p = 1 : Points
    % Sigma
    Sigma(p) = sum((v'.*V(:,p)-ones(1)*Means(p)).*(v'.*V(:,p)-
ones(1)*Means(p))) ...
    + n(p)*(a(p)-Means(p))*(a(p)-Means(p));
    Sigma(p) = sqrt(Sigma(p)/(2*n(p)-1)) / Means(p);
end
y = sum(n'.*Sigma);
```

---

## FILE NORMCODE.M

---

```
global V
global sV
global File
global Curves
global Points
```

```
precision
iteration
maxgrey
```

```
tic
```



```

%Change NaN to zero, just in case
for p = 1 : size(V(1,:),2)
    for c = 1 : size(V(:,1),1)
        if isnan(V(c,p));
            V(c,p) = 0;
        end
    end
end

% remove anything bigger than maxgrey
for p = 1 : size(V(1,:),2)
    for c = 1 : size(V(:,1),1)
        if V(c,p) >= maxgrey;
            sprintf('V(%d,%d) = %d = 0', c,p,V(c,p));
            V(c,p) = 0;
        end
    end
end

% remove any DNA points that has only one point
for p = 1 : size(V(1,:),2)
    n = 0;
    for c = 1 : size(V(:,1),1)
        if V(c,p) > 0;
            n = n + 1;
        end
    end
    if n == 1
        V(:,p) = 0;
    end
end

% remove any curve that has only one point
for c = 1 : size(V(:,1),1)
    n = 0;
    for p = 1 : size(V(1,:),2)
        if V(c,p) > 0;
            n = n + 1;
        end
    end
    if n == 1
        V(c,:) = 0;
    end
end

% Clean V of 0 Column
V(:,all((V==0)')) = [];

% Clean V of 0 Row
V(all((V==0)',:)) = [];

% Create sV = NaN or not (binary matrix)
sV = V>0;

% Average at each point
clear global a;
clear a1;

```

```

clear a2;
global a
for p = 1 : size(V(1,:),2)
    a(p) = 0;
    n(p) = 0;
    for c = 1 : size(V(:,1),1)
        if V(c,p)>0
            n(p) = n(p) + 1;
            a(p) = a(p) + V(c,p);
        end
    end
    a(p) = a(p)/n(p);
end

% Coef to get each curve to average
clear global v;
clear v1;
clear v2;
global v
for c = 1 : size(V(:,1),1)
    v(c) = 0;
    n(c) = 0;
    for p = 1 : size(V(1,:),2)
        if V(c,p) > 0
            n(c) = n(c) + 1;
            v(c) = v(c) + a(p)/V(c,p);
        end
    end
    v(c) = v(c)/n(c);
end
a1 = a;
v1 = v;

v
clear vv
Curves = size(V(:,1),1);
Points = size(V(1,:),2);

options = [0, precision, precision, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
iteration];
[Res1, Opt] = fmins('NormFunc', v, options, a);
Res1
Opt(10)

V1 = V;
for i = 1 : size(V(:,1),1)
    V1(i,:) = Res1(i).*V(i,:);
end

% Average at each point
for p = 1 : size(V1(1,:),2)
    a(p) = 0;
    n(p) = 0;
    for c = 1 : size(V1(:,1),1)
        if V1(c,p)>0
            n(p) = n(p) + 1;
            a(p) = a(p) + V1(c,p);
        end
    end
end

```

```

        end
    end
    a(p) = a(p)/n(p);
end

a2 = a;

%v
options = [0, precision, precision, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0, 0,
iteration];
[Res2, Opt] = fmins('NormFunc', Res1, options, a);
Res2
Opt(10)

toc

for p = 1 : size(V(1,:),2)
    for c = 1 : size(V(:,1),1)
        if V(c,p) == 0;
            V(c,p) = NaN;
        end
    end
end
for p = 1 : size(V1(1,:),2)
    for c = 1 : size(V1(:,1),1)
        if V1(c,p) == 0;
            V1(c,p) = NaN;
        end
    end
end
end

% Plot
figure
subplot(3,1,1)
plot(V','*-')
warning off;
title(File);
warning on;
hold on;
plot(a1, 'o-', 'LineWidth',1.5)
hold off;

subplot(3,1,2)
plot(a1, 'o-', 'LineWidth',1.5)
hold on;
plot(V1','*-')
hold off;

subplot(3,1,3)
V2 = V;
for i = 1 : size(V(:,1),1)
    V2(i,:) = Res2(i).*V(i,:);
end
plot(a2, 'o-', 'LineWidth',1.5)
hold on;
plot(V2','*-')
hold off;

```

```
% set printing
set(gcf, 'Color',[1 1 1], ...
    'PaperOrientation', 'Landscape')
set(gcf, 'PaperUnits', 'inches');
PaperSize = get(gcf, 'PaperSize');
PaperPos=[0.3 0.3 (PaperSize(1)-.3) (PaperSize(2)-0.3)];
set(gcf, 'PaperPosition', PaperPos);
h1 = uimenu('Parent',gcf, ...
    'Callback','printdlg', ...
    'Label','&Print', ...
    'Tag','MnPrint');

% Save to file
NewFile = strcat(File, '.res');
fid = fopen(NewFile, 'wt');
fprintf(fid, '%s\n', File);
for i = 1 : size(V(:,1),1)
    fprintf(fid, '%s\n', num2str(Res2(i)));
end
fclose(fid);
```

---



# For reference

Not to be taken  
from the room.

