Lawrence Berkeley National Laboratory

LBL Publications

Title

kb_DRAM: annotation and metabolic profiling of genomes with DRAM in KBase

Permalink https://escholarship.org/uc/item/1bg5v2mr

Journal Bioinformatics, 39(4)

ISSN 1367-4803

Authors

Shaffer, Michael Borton, Mikayla A Bolduc, Ben <u>et al.</u>

Publication Date

2023-04-03

DOI

10.1093/bioinformatics/btad110

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <u>https://creativecommons.org/licenses/by/4.0/</u>

Peer reviewed

OXFORD

Genome analysis kb_DRAM: annotation and metabolic profiling of genomes with DRAM in KBase

Michael Shaffer ()¹, Mikayla A. Borton¹, Ben Bolduc², José P. Faria³, Rory M. Flynn¹, Parsa Ghadermazi¹, Janaka N. Edirisinghe³, Elisha M. Wood-Charlson⁴, Christopher S. Miller ()⁵, Siu Hung Joshua Chan ()¹, Matthew B. Sullivan ()², Christopher S. Henry³, Kelly C. Wrighton ()¹*

¹Colorado State University, Fort Collins, CO, USA

²The Ohio State University, Columbus, OH, USA

³Argonne National Laboratory, Lemont, IL, USA

⁴Lawrence Berkeley National Laboratory, Berkeley, CA, USA

⁵University of Colorado Denver, Denver, CO, USA

*Corresponding author. Colorado State University, 307 University Avenue, Fort Collins, CO 80523, USA. E-mail: kelly.wrighton@colostate.edu Associate Editor: Tobias Marschall

Received on 13 March 2022; revised on 30 December 2022; accepted on 28 February 2023

Abstract

Microbial genome annotation is the process of identifying structural and functional elements in DNA sequences and subsequently attaching biological information to those elements. DRAM is a tool developed to annotate bacterial, archaeal, and viral genomes derived from pure cultures or metagenomes. DRAM goes beyond traditional annotation tools by distilling multiple gene annotations to genome level summaries of functional potential. Despite these benefits, a downside of DRAM is the requirement of large computational resources, which limits its accessibility. Further, it did not integrate with downstream metabolic modeling tools that require genome annotation. To alleviate these constraints, DRAM and the viral counterpart, DRAM-v, are now available and integrated with the freely accessible KBase cyberinfrastructure. With kb_DRAM users can generate DRAM annotations and functional summaries from microbial or viral genomes in a point-and-click interface, as well as generate genome-scale metabolic models from DRAM annotations.

Availability and implementation: For kb_DRAM users, the kb_DRAM apps on KBase can be found in the catalog at https://narrative.kbase.us/#catalog/modules/kb_DRAM. For kb_DRAM users, a tutorial workflow with all documentation is available at https://narrative.kbase.us/narrative/129480. For kb_DRAM developers, software is available at https://github.com/shafferm/kb_DRAM.

1 Introduction

Genome annotation is gene prediction followed by the assignment of biological function to genes. Protein coding sequences are commonly assigned function via homology searches to protein databases which contain sequences with assigned or inferred functional content. A number of genome annotators targeting microbial genomes have been developed (Aziz et al. 2008; Seemann 2014; Tanizawa et al. 2018; Dong and Strous 2019; Zhou et al. 2020). We previously developed DRAM, a genome annotation tool which allows the user to compile annotations from multiple functionally divergent protein databases at one time, then synthesizes this content into functional profiles for each genome (Shaffer et al. 2020). This allows the user to rapidly understand the collection of biologically encoded functions in a set of microbial genomes.

Genome-scale metabolic models (GEMs) are representations of the metabolic reactions that occur within a bacterial cell. These

DRAM is limited by the high computational requirements of rapidly searching against large protein databases. It requires a minimum of 128 GB of RAM to set up and 64 GB of RAM to annotate. Thus, the use of DRAM is currently limited to those with access to large compute servers. Recently, cyberinfrastructure platforms have been built that provide access to computing resources as well as point-and-click interfaces to software that would usually require command-line access (Merchant et al. 2016; Afgan et al. 2018; Arkin et al. 2018).

[©] The Author(s) 2023. Published by Oxford University Press.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

reactions can be predicted from genome annotations, but many bacterial genome annotators do not generate output that is easily integrated into modeling frameworks. Additionally, recent research has shown value in combining annotations to improve genome function coverage (Griesemer et al. 2018), making frameworks with interoperable support of genome annotators and GEM construction more valuable.

We have built a DRAM KBase module (kb_DRAM). Here we show that using kb_DRAM, anyone with access to the KBase cyberinfrastructure (Arkin et al. 2018) can annotate microbial genomes with DRAM, distill these annotations into visualizations of predicted genomic functions, and use the annotations to build GEMs. Also, using bacterial genomes derived from phylogenetically distinct lineages, we show value added of including kb_DRAM annotation alongside an established KBase annotator, RAST (Aziz et al. 2008).

2 kb_DRAM

KBase is a cyberinfrastructure platform which allows users to use common bioinformatics tools to analyze public data or a user can upload their own. Within KBase users can process microbial genomics and metagenomics data from raw reads to assemblies and bins or imported data from other sources. kb_DRAM is a plugin that provides three KBase apps. These can (i) annotate microbial DNA sequences from assemblies, isolate genomes or metagenomeassembled genomes (KBase assembly objects), (ii) annotate predicted coding sequences from microbial genomes (KBase genome objects), or (iii) annotate viral genomes identified from metagenomes using DRAM-v.

The kb_DRAM apps use the same databases as the default DRAM installation [KOfam (Aramaki et al. 2020), dbCAN2 (Zhang et al. 2018), PFAM (El-Gebali et al. 2019), and MERPOS (Rawlings et al. 2018)] to annotate predicted microbial proteincoding genes as well as barrnap (https://github.com/tseemann/barr nap) for rRNA identification and tRNA-scanSE (Chan and Lowe 2019) for tRNA detection. Like DRAM, kb_DRAM generates and summarizes gene annotations across genomes into three levels of refinement: (i) Raw, (ii) Distillate, and (iii) Product (Supplementary Information). The raw is a synthesized annotation of all genes in a dataset across multiple databases, the distillate assigns many of these genes to specific functional categories, and the product visualizes the presence of key functional genes across genomes. Notably, the product is an interactive heatmap shown in the KBase browser that enables users to visually profile the functional potential of input genomes or metagenomes. All files generated by kb_DRAM (e.g. raw annotations, distillate, and genome completion) are available to download so users can understand their genomes more deeply. kb_DRAM apps also generate annotated KBase genome objects, which can be used in downstream analyses in KBase including building GEMs, not previously possible in DRAM alone. Each output from kb_DRAM is described in detail in the Supplementary Information.

DRAM-v is designed to annotate viral genomes that are identified using VirSorter (Roux et al. 2015). DRAM-v uses the same functional databases as DRAM with the addition of RefSeq viral. To annotate with DRAM-v within KBase users can start with metagenomic assemblies and identify potential viral contigs from metagenomes using the VirSorter app in KBase. The output of the VirSorter app is then passed to the DRAM-v app for auxiliary metabolic gene (AMG) annotation. The DRAM-v app shows the interactive product heatmap, which highlights potential AMGs identified in the dataset along with confidence scores for each and allows the user to download all other DRAM-v files.

3 DRAM annotations of microbial genomes can generate quality GEMs in KBase

To demonstrate DRAM in KBase and show its compatibility with downstream applications, we annotated two genomes with the RAST and DRAM apps. *Escherichia coli* strain K-12 was chosen as

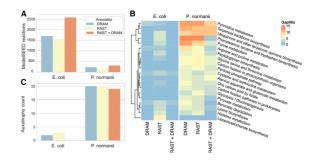


Figure 1 Annotation and modeling performance on three genomes. (A) Number of modelSEED reactions assigned by each tool on each genome. (B) Heatmap showing number of gap filled reactions required for growth on glucose minimal media per pathway. Only pathways with ≥ 5 gap fills required by at least one annotation set shown. (C) Number of auxotrophies predicted present based on annotations from each tool for each genome

a well-characterized bacterial genome, while Paceibacter normanii AAA255-P19, a member of the candidate phyla radiation, represents a genome obtained from uncultivated microbes through metagenomics with limited functional curation (Castelle et al. 2018). Full workflows with these data are available on KBase (E.coli: https:// kbase.us/n/103341/23/, P.normanii: https://kbase.us/n/128174/5/), and Supplementary Fig. S1 displays the methods workflow for this analysis in KBase. For both genomes, when comparing the outputs of DRAM to RAST, DRAM annotations yielded more ModelSEED reactions, a measure of how many unique gene annotations could be converted to metabolic reactions (Fig. 1A, Supplementary Table S1). As expected, due to the depth of study of E.coli, we obtained more reaction-specific annotations for E.coli and much fewer for P.normanii. Ultimately, as others have shown (Griesemer et al. 2018), there was value in using more than one annotator, as merged DRAM + RAST annotations yielded $1.5 \times$ and $1.4 \times$ more reactions than DRAM and 1.6× and 2.3× for RAST in E.coli and P.normanii, respectively.

Next, we constructed GEMs using the RAST, DRAM, and DRAM + RAST annotations and GEMs were gap filled using glucose minimal media to bridge gaps in metabolic pathway reconstruction leading to biomass production (see Supplementary Information). The kb_DRAM app represents a significant advance, as DRAM annotations can be directly integrated into GEMs, a capability not previously available in DRAM.

Escherichia coli RAST had more reactions (n = 1681) in the model than DRAM but fewer in *P.normanii* (n = 488). In both cases, DRAM + RAST outperformed each annotator alone (Supplementary Table S1). We failed to find a clear pattern of better performance by DRAM or RAST in any particular metabolic pathway (Fig. 1B). Subsequently, the GEMs were characterized to predict auxotrophies. In all genomes the merged annotation model showed the least number of auxotrophies (Fig. 1C). Interestingly for the E.coli model both RAST and DRAM predicted auxotrophies, while merging the annotations removed these auxotrophies, yielding a final GEM more consistent with expected experimental evidence (Tao et al. 1999). We note that a large number of auxotrophies are still predicted for P.normanii, even when merging annotations. This finding may be biological, reflecting the symbiotic lifestyle predicted for members of this species, as many of these reactions could be provided by the host (He et al. 2021). However, there is no experimental data for P.normanii to validate this inference at this time.

4 Conclusion

Here, we present a KBase module with apps for running DRAM and DRAM-v. This resource enables computationally intensive genome annotation by broader audiences. We highlight that both DRAM-v and DRAM are integrated into the KBase cyberinfrastructure with the ability to ingest data from and pass data to other KBase applications. We show that DRAM can be applied to generate gene

annotations from phylogenetically distinct genomes derived from pure cultures and metagenomics. Using *E.coli*, we demonstrated the addition of DRAM annotations yielded a GEM that was consistent with experimental evidence. Thus, the kb_DRAM app will enhance user analyses of genome function beyond DRAM, enabling seamless integration of DRAM gene annotations into modeling frameworks.

Supplementary data

Supplementary data are available at Bioinformatics online.

Conflict of interest: None declared.

Funding

This work was supported by an Early Career Award from the U.S. Department of Energy Office of Science, Office of Biological and Environmental Research (DOE BER), under award number [DE-SC0019746 to K.C.W.] as well as two additional DOE BER grants [DE-SC0021350, DE-SC0023084] and a National Institutes of Health [R01AI143288].

References

- Afgan E, Baker D, Batut B et al. The galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. Nucleic Acids Res 2018;46:W537-44.
- Aramaki T, Blanc-Mathieu R, Endo H et al. KofamKOALA: KEGG Ortholog assignment based on profile HMM and adaptive score threshold. *Bioinformatics* 2020;36:2251–2.
- Arkin AP, Cottingham RW, Henry CS et al. KBase: the United States Department of Energy Systems Biology Knowledgebase. Nat Biotechnol 2018;36:566–9.
- Aziz RK, Bartels D, Best AA et al. The RAST Server: rapid annotations using subsystems technology. BMC Genomics 2008;9:1–15.
- Castelle CJ, Brown CT, Anantharaman K *et al.* Biosynthetic capacity, metabolic variety and unusual biology in the CPR and DPANN radiations. *Nat Rev Microbiol* 2018;16:629–45.

- Chan PP, Lowe TM. tRNAscan-SE: searching for tRNA genes in genomic sequences. Nucleic Acids Research 2019:1–14.
- Dong X, Strous M. An integrated pipeline for annotation and visualization of metagenomic contigs. Front Genet 2019;10:999.
- El-Gebali S, Mistry J, Bateman A et al. The Pfam protein families database in 2019. Nucleic Acids Res 2019;47:D427-32.
- Griesemer M, Kimbrel JA, Zhou CE *et al.* Combining multiple functional annotation tools increases coverage of metabolic annotation. *BMC Genomics* 2018;19:1–11.
- He C, Keren R, Whittaker ML *et al.* Genome-resolved metagenomics reveals site-specific diversity of episymbiotic CPR bacteria and DPANN archaea in groundwater ecosystems. *Nat Microbiol* 2021;6:354–65.
- Merchant N, Lyons E, Goff S *et al.* The iPlant collaborative: cyberinfrastructure for enabling data to discovery for the life sciences. *PLoS Biol* 2016;14: e1002342.
- Rawlings ND, Barrett AJ, Thomas PD *et al.* The MEROPS database of proteolytic enzymes, their substrates and inhibitors in 2017 and a comparison with peptidases in the PANTHER database. *Nucleic Acids Res* 2018;46: D624–32.
- Roux S, Enault F, Hurwitz BL et al. VirSorter: mining viral signal from microbial genomic data. PeerJ 2015;3:e985.
- Seemann T. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 2014;30:2068–9.
- Shaffer M, Borton MA, McGivern BB et al. DRAM for distilling microbial metabolism to automate the curation of microbiome function. Nucleic Acids Res 2020;48:8883–900.
- Tanizawa Y, Fujisawa T, Nakamura Y *et al.* DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. *Bioinformatics* 2018;34:1037–9.
- Tao H, Bausch C, Richmond C et al. Functional genomics: expression analysis of *Escherichia coli* growing on minimal and rich media. J Bacteriol 1999; 181:6425–40.
- Zhang H, Yohe T, Huang L *et al.* DbCAN2: a meta server for automated carbohydrate-active enzyme annotation. *Nucleic Acids Res* 2018;46: W95–101.
- Zhou Z, Tran PQ, Breister AM *et al.* METABOLIC: High-throughput profiling of microbial genomes for functional traits, biogeochemistry, and community-scale metabolic networks. Microbiome, 2020; 10:33.