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Species differences in tumour responses to cancer chemotherapy

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Despite advances in chemotherapy, radiotherapy and targeted drug development, cancer remains a disease of high morbidity and mortality. The treatment of human cancer patients with chemotherapy has become commonplace and accepted over the past 100 years. In recent years, and with a similar incidence of cancer to people, the use of cancer chemotherapy drugs in veterinary patients such as the dog has also become accepted clinical practice. The poor predictability of tumour responses to cancer chemotherapy drugs in rodent models means that the standard drug development pathway is costly, both in terms of money and time, leading to many drugs failing in Phase I and II clinical trials. This has led to the suggestion that naturally occurring cancers in pet dogs may offer an alternative model system to inform rational drug development in human oncology. In this review, we will explore the species variation in tumour responses to conventional chemotherapy and highlight our understanding of the differences in pharmacodynamics, pharmacokinetics and pharmacogenomics between humans and dogs. Finally, we explore the potential hurdles that need to be overcome to gain the greatest value from comparative oncology studies.

1. Introduction

The German chemist, Paul Ehrlich, who devised the term 'chemotherapy', was also one of the first people to document the utility of animal models in screening chemicals for their potential anti-cancer activity. Chemotherapy is now widely known as the use of chemicals to treat disease and it has been widely employed to treat cancer since the beginning of the twentieth century. Despite pessimism about the utility of chemotherapy to cure cancer, the success of combination chemotherapy in curing paediatric acute leukaemia and diffuse large B-cell lymphoma in the 1960s spurred the formation of medical oncology as a specialty. Since the 1970s, extensive development of anti-cancer drug screening programmes occurred with animal models supporting early assessment of therapeutic and toxicity potential. These animal models are often the final investigation in the development of drug candidates for human clinical trials following promising *in vitro* activity.

Animal models are diverse and often distinguished on the basis of the origin of the tumour—namely, spontaneous or inducible tumour development versus tumours that are transplanted. While human tumour xenografts have been the most extensively used model to predict anti-tumour efficacy, several studies have indicated variable correlation between xenograft models and clinical activity [1–10]. Mouse models tend to suffer from a number of limitations that impact their predictive behaviour for human tumour types; while not an exhaustive list, some primary reasons for failure of xenograft models include biological differences between species (for example, telomerase is active in almost all murine cells contrary to in human cells), altered downstream signalling in the mouse compared with known human cancer signalling pathways such as Ras, altered metabolism of and sensitivity to DNA-damaging agents, variations in immune competency and altered tumour microenvironment [1,11–14]. The progression of mouse models from syngeneic to sophisticated genetically engineered mouse (GEM) models and recently to non-germline GEM models has vastly improved current understanding of cancer biology

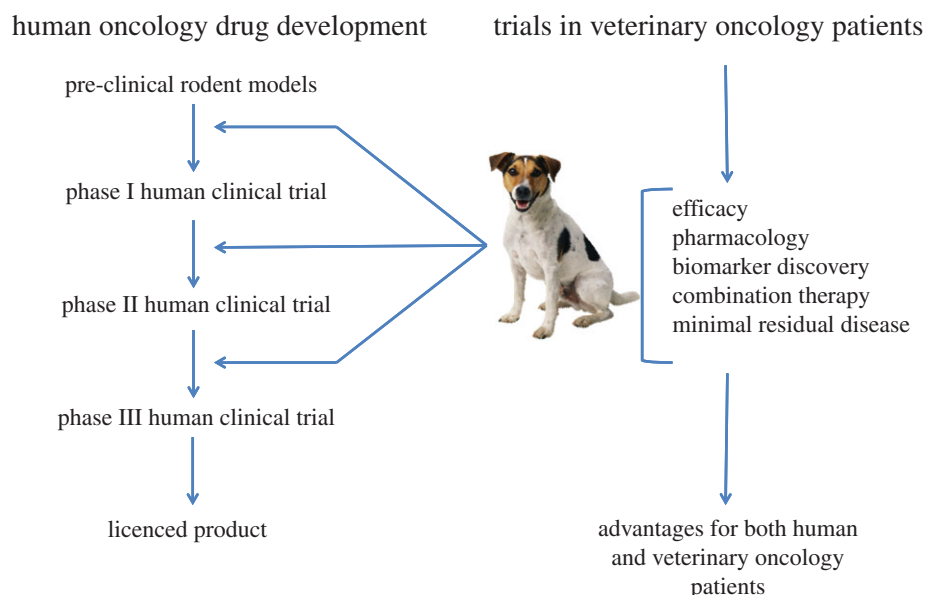


Figure 1. A unique collaboration between human oncology drug development and trials in veterinary oncology patients has the potential to increase the speed with which new human drugs reach the clinic. (Online version in colour.)

but has modestly improved throughput, expense and the practicality of preclinical drug testing [1,13–15]. This latter aspect is predominantly due to the complexity of cancer, as it is difficult to design an animal model that preserves the same genotypic and phenotypic characteristics of the tumours from which they were derived [12,16,17].

Preclinical studies of the anti-tumour activity of novel or modified existing chemotherapy agents requires the use of a model system capable of answering specific questions regarding the drug's efficacy. A failure to consider an appropriate animal model may contribute to the failure of a compound to achieve approval for use or hinder further investigation of new drugs. The integration of comparative oncology approaches using pet dogs with spontaneously occurring tumours as models in drug development pathways has garnered much attention in recent years due to numerous clinical and molecular similarities between common canine and human cancers [18–21] (figure 1). Whilst implementing a combined use model, wherein information from mouse and large animal models both provide input into rational development of chemotherapy drugs, it is important to address interspecies similarities and differences in drug metabolism, physiology, absorption and distribution. Given that in some clinical scenarios, even human beings are not predictive models of cancer in distinct human groups, consideration must also be given to emerging inter- and intra-species pharmacogenetic differences in order to maximize the human predictive potential of data obtained from various screening systems [11,22].

Tumour responsiveness to various chemotherapy drugs is dependent on a multitude of factors that go far beyond simple dosage and frequency of chemotherapy administration. Responsiveness is also dependent on tumour histology, growth rate, tumour heterogeneity and mechanisms of drug resistance. Tumour sensitivity is therefore a dynamic and complicated issue, particularly when acquired resistance is taken into account. Additional factors such as immunologic responses to tumour antigens, serum protein content and natural barriers (e.g. the blood-brain barrier) also play integral roles in chemotherapy distribution to tumour and normal tissues. This article is not intended to provide an overview of all mechanisms of tumour

response, but rather aims to summarize some of the challenges and opportunities that present when considering animal models in predicting tumour response to chemotherapy. A general overview of chemotherapy indications and evidence for clinical use are presented using comparative human and canine histologies; the question as to whether or not there are significant differences in tumour response between species is also posed. Subsequently, species differences in pharmacokinetic, pharmacodynamics, pharmacogenomics and immunologic factors are also discussed. This article focuses on increasing interest in the dog as a model for comparative oncology approaches but also references common laboratory species that continue to serve as the mainstay for chemotherapy research. The purpose of the review is to stimulate discussion on the underlying question as to whether an appreciation of interspecies differences can alert researchers and clinicians to deviations in tumour response to chemotherapy between species.

2. What is the rationale for chemotherapy?

The overall objective of chemotherapy is to reduce the population of tumour cells to zero in order to effect a cure. Following studies of murine leukaemia, the fractional cell kill hypothesis was generally accepted as a method by which to approximate tumour response; specifically, the hypothesis stated that a given drug concentration applied for a defined time period will kill a constant fraction of the cell population, regardless of the initial, absolute number of tumour cells [23]. This provided the rationale for many chemotherapy protocols, particularly leukaemia and lymphoma protocols, as the outcome of chemotherapy is dependent on the drug dosage and the number and frequency of drug administrations. Indeed, both human and canine non-Hodgkin's lymphoma (NHL) are treated primarily with multidrug chemotherapy with remission rates approaching 90% in both species [24,25]. When applied to solid tumours, multiple confounding factors wreak havoc with the fractional cell kill hypothesis, such as decelerating growth with a small fraction of cycling cells, intrinsic resistance (as postulated with tumour stem

cells), impaired tumour vascularity and variations within the tumour heterogeneity and the tumour microenvironment. Selection pressure following chemotherapy treatment provides an entirely unique tumour population to work with, also invalidating the fractional cell kill model [26–28]. While acknowledging that chemotherapy alone rarely induces a durable response for solid tumours, adjuvant systemic treatment has made an impact on several solid tumour histologies in humans [29,30]. Post-operative adjuvant therapy, when indicated, generally significantly reduces the risk of recurrence compared with surgery alone although the degree of benefit varies from conservative to remarkable. The benefit of adjuvant chemotherapy has been recognized in many tumour histologies in veterinary patients as well, even though the principles of chemotherapy are altered with one of the primary goals centring on the maintenance of good-to-excellent quality-of-life measures [31–34].

3. Comparative aspects of chemotherapy indications for selected tumours

(a) Non-Hodgkin's lymphoma

The principles of treatment for NHL in humans are complex and must take into account patient factors, subtype of lymphoma, stage and biologic behaviour of the disease [35]. However, multidrug chemotherapy protocols, and often CHOP-based (Cyclophosphamide, Hydroxydaunorubicin, Oncovin, Prednisone) protocols, form the mainstay of treatment for high-grade, multifocal follicular lymphoma and diffuse large B-cell lymphoma in humans, the two most common lymphomas to affect adults in the United States [35]. While the inclusion of rituximab (R) to chemotherapy combinations has improved response rates and outcome, response rates to R-CHOP are approximately 85–90% and 70–80%, respectively, with median progression-free survival of approximately 5–6 years and 3–4 years, respectively [35–38]. While histological classification is not routinely sought following diagnosis of canine NHL, the majority are diffuse large-cell lymphoma and are subsequently treated with CHOP-based chemotherapy [24,39–41]. Remission rates vary depending on several factors but average 80–95% with CHOP-based therapy, historically providing median survival times of approximately 11–12 months with 25% of dogs alive at 2 years [24]. As many pet owners choose to proceed with rescue chemotherapy following first relapse, median survival times for dogs tend to be greater than 1 year [42].

(b) Sarcomas

Sarcomas are biologically complex mesenchymal tumours requiring multi-disciplinary management due to the variation in location and behaviour. In osteosarcoma (OSA) of the extremity, the most common primary malignant bone cancer in people primarily affecting young children or adolescents, treatment prior to 1970 centred on amputation, with most patients experiencing a 5-year survival rate of only 20–30% due to distant metastatic disease [25,43,44]. While surgical and radiation therapy approaches have been consistently instrumental in controlling localized disease, the therapeutic benefit of chemotherapy at delaying or reducing metastasis was first observed over 40 years ago [30,45].

Subsequent randomized controlled clinical trials provided the evidence needed to justify incorporating chemotherapy into standard therapeutic protocols [25,44,46,47]. Since the addition of cisplatin-based adjuvant therapy in the 1980s, adjuvant or neoadjuvant chemotherapy protocols have provided 5-year survival rates of 60–70% [25,46,47]. While the survival benefit of chemotherapy has been well established in human OSA, the use of adjuvant chemotherapy for soft tissue sarcomas (STS), a vastly heterogeneous group of tumours, has taken a more circuitous path. Several factors are predictive of outcome in human STS, including tissue of origin, histological grade, tumour size, anatomic site, degree of invasion or depth into underlying tissues and patient performance score [29,48–51]. While surgery with or without radiation therapy is the mainstay of treatment for loco-regional control of STS, many patients—approximately 50% of patients with high-grade STS—will develop and succumb to recurrent and/or metastatic disease. Despite the recognition of many prognostic factors predictive of disease recrudescence, adjuvant chemotherapy has failed to unequivocally provide clinical benefit in patients with high-risk disease. The standard first-line treatment tends to be single agent doxorubicin, although some literature suggests there is added overall clinical benefit in some patients to the addition of ifosfamide despite the associated increase in toxicity [52,53]. In a landmark meta-analysis of individual patient data, data analysis from 1568 patients from 14 clinical studies with a median follow-up of 9.4 years demonstrated evidence that adjuvant chemotherapy significantly improved local and distant recurrence-free intervals; however, there was no significant benefit in overall survival [54]. Subsequent meta-analyses confirmed the suggestion that doxorubicin-based chemotherapy improved recurrence-free survival at 10 years with a non-significant trend towards improved survival [52,55]. However, most current guidelines suggest that patient selection is paramount in order to justify the chemotherapy-associated toxicities associated with treatment [29,52,53].

Veterinary studies, despite being less numerous and under-powered in comparison to human studies, have predominantly paralleled the findings in support of adjuvant therapy for sarcomas. Because of salient clinical and molecular similarities between OSA in humans and dogs, canine appendicular OSA has been considered a valid model for human OSA [18,56–59]. Similar to humans, amputation alone yields a poor prognosis with most dogs euthanized within four to five months due to metastatic disease [60–62]. The addition of adjuvant chemotherapy to amputation or limb-sparing surgery has demonstrated clear benefit in the disease-free survival and overall survival, similar to human OSA [60–65]. Various protocols have been assessed with platinum agents or doxorubicin forming the basis for most protocols; to date, there has been no clear benefit with the use of combination chemotherapy but rather single agent carboplatin or doxorubicin are most often used due to ease of administration, acceptable toxicity profile and comparable outcomes [60–66]. Unfortunately, while there is clear short-term benefit to adjuvant chemotherapy, long-term survival for canine OSA remains poor with 1-year survival estimated at approximately 35–45% [66]. Paralleling the case in human OSA, few improvements in outcome have been made in the last 20 years with local control with peri-operative chemotherapy remaining the standard approach for optimal

outcome. While there has been a clear benefit to the use of chemotherapy in canine OSA, the utility of chemotherapy following local control for canine high-grade STS is unclear. Similar to human studies, there is no evidence to support a significant role for chemotherapy in the management of low-risk disease and surgery with or without radiation therapy is considered the standard-of-care. Doxorubicin alone and doxorubicin-based protocols have shown the most promise with advanced measurable STS and therefore are most often elected for dogs at risk for metastasis [67,68]. However, in a report of 39 dogs with high-grade STS, there was no improvement in either disease-free interval or overall survival in dogs treated with surgery and doxorubicin compared with surgery alone [69]. This report was small and included uncommon (visceral) STS in the analysis thus results may have represented type II error; nonetheless, use of chemotherapy for high-grade STS remains controversial.

4. Is there a variation in response to chemotherapy in dogs versus humans?

Given the similar trends in response and indications for chemotherapy in some veterinary patients, as illustrated above, some (often pet owners) query the seeming lack of equivalent response to chemotherapy in their pet dogs. For example, why do dogs with lymphoma benefit from 11 to 12 months of survival with chemotherapy, whereas humans often achieve 3–4 years? The domestic dog is an interesting study of age-specific mortality evolution, possibly associated with selection for body size. Body size in dogs varies by almost two orders of magnitude and a longevity factor of two; this implies that, on average, small breed dogs die at approximately 10–15 years, while large breed dogs die at approximately 5–8 years [70,71]. While paradoxical to the common notion that there is a positive relationship between lifespan and body size, as is obvious when considering survival of a rat (5 years) compared with a whale (often more than 100 years), there is an inverse relationship in occasional species such as humans and dogs [72–74]. The commonly touted ‘7-year rule’ that defines 1 ‘human year’ as equivalent to 7 ‘dog years’ is mythical, with research suggesting that dogs indeed age faster than humans and that after 2 years of age for a dog (equivalent to approx. 24 human years), each year of a dog’s life is equivalent to approximately 4–5 human years [75]. Extrapolating from this, comparable remission durations could be deemed reasonable with a 10–11 month remission on CHOP comparable with approximately 4 years of remission in a human. Using similar extrapolation, a dog with OSA may only achieve one-quarter of the remission duration of a paediatric human patient with OSA, suggesting either differences in tumour response, drug sensitivity or biologic behaviour as most dogs die of metastatic disease rather than co-morbid factors associated with age. It must be acknowledged, however, that given the vast variation in breed size, it would seem impossible to develop a single factor to account for translating ‘dog years’ to ‘human years’ [71]. Given that lifespan in dogs is inversely related to body size, breed and intra-breed variability needs to be worked into an appropriate model. The biologic basis for the inverse relationship between size and lifespan is not understood although some investigators have suggested that the insulin-like growth factor 1 (IGF-1) signalling cascade

plays a role, as smaller dogs have lower levels of IGF-1 compared with large breed dogs [76–78]. While small breed dogs with OSA are postulated to have a better prognosis than large breed dogs treated with local control and chemotherapy, this potential difference could be explained by the size-to-lifespan relationship. A recent report evaluating 26 small breed dogs with appendicular OSA treated with surgery and chemotherapy indicated the median survival time was 415 days, longer than reports including dogs of all sizes (predominantly large breed dogs) [60–66,79]. However, recognizing that small breed dogs take a longer time to ‘age’, 415 days in a small breed dog may be comparable to 330 days in a large breed dog [70,76]. The more accepted hypothesis, however, is that there is a relative difference in dosing of chemotherapy in small breed dogs, with large breed dogs receiving a lower dosing than small breed dogs. Alternatively, the biologic behaviour of OSA in small breed dogs is truly altered in comparison to large breed dogs, as suggested by lower mitotic indices and grade [79].

With respect to comparative aspects of STS, there is the suggestion of a small yet consistent improved recurrence-free interval in humans with the use of adjuvant doxorubicin-based chemotherapy following local control; this has not been realized in dogs, although only one small study has addressed the issue. The disease-free survival for human STS increased from 45 to 55% at 10 years with the use of adjuvant chemotherapy but there was an insignificant improvement in overall survival; the study was not powered to detect a small change (less than 4-year improvement) in survival [54]. Given the relationship of dog aging to human aging and presuming equivalent STS response to doxorubicin, it is possible that doxorubicin only induced an undetectable short (months) improvement in disease-free interval [69].

5. What impact do dose and dose intensity have on tumour response in dogs versus humans?

Chemotherapy drugs are considered some of the most dangerous within the medical arsenal due to their narrow therapeutic index and the desire to use them near their maximally tolerated dose. Most chemotherapy drugs are currently dosed in both companion animals and humans on the basis of the patient’s body surface area (BSA), which tends to correlate poorly with drug pharmacokinetics [80,81]. BSA is proportional to both blood volume and glomerular filtration rate (GFR), despite neither contributing to chemotherapy efficacy or toxicity as much as liver function or other metabolic variations [82–85]. Interestingly, BSA was initially derived as a mathematical approach to estimating tolerable starting doses in humans for phase I trials based on preclinical data in animals; BSA dosing essentially normalizes the maximum tolerated dose of many chemotherapy drugs in humans, dogs, rats and mice [83–86]. There has been no clear relationship between pharmacokinetic parameters and BSA for common chemotherapy drugs, and in people, up to 20-fold variation in pharmacokinetics is routinely observed in patients receiving BSA-calculated doses [82]. In dogs, there has been empirical evidence that smaller dogs experience increased toxicity compared with larger dogs when administered chemotherapy dosed based on BSA [87–90]. For non-metabolized drugs, the use of BSA may be effective, but when tumour effects and side effects are based on complex

systems such as metabolism and genetics, there are too many size-independent factors that can affect a generalized BSA approach to dosing [83,91]. It is important to perform studies that relate drug exposure to tumour response in species commonly used in drug development, whether looking at animal models as predictive of efficacy or toxicity. Generally speaking, these data are lacking in companion animals for many drugs despite the recognition that there are many limitations to BSA dosing of chemotherapy in dogs [92,93]. One pivotal study in cats demonstrated a clear relationship between drug exposure and neutrophil nadir, clearance and GFR, permitting calculation of individual animal dosing [94,95]. A recent study in dogs attempted to develop a simple strategy for measuring doxorubicin exposure in dogs in order to improve the study of the correlation between pharmacokinetics and both toxicity and tumour response [96]. It is not yet clear if pharmacokinetic-based dosing improves outcome in companion animals, but several studies in humans have demonstrated beneficial effects both in terms of reducing toxicity and improving disease-free intervals for various chemotherapy drugs and protocols [97–102]. In a phase II study of metastatic colorectal cancer, both efficacy and tolerability of pharmacokinetic-adjusted fluorouracil as part of a multidrug protocol were higher than BSA dosing [103]. An earlier phase III study comparing pharmacokinetically adjusted fluorouracil to conventional dosing in metastatic colorectal cancer patients demonstrated that personalized dosing improved the response rate, decreased severe toxicity and led to a trend in improved survival [102]. Importantly, the mean fluorouracil dose was higher in the phase III trial with personalized dosing, which was also the group with decreased occurrence of severe toxicity [102]. As quality-of-life measures are important when considering any chemotherapy regimen, efforts to improve outcome while decreasing toxicity are paramount to advancing cancer care.

Despite efforts to investigate drug exposure in companion animals and the assumptions made to define the relationship between chemotherapy drug exposure and effect, conflicting results exist in the veterinary literature. In canine lymphoma, one study showed that dogs that developed grade III or IV neutropenia after chemotherapy demonstrated improved survival, leading to the suggestion that neutropenia was associated with more optimal drug exposure [104]. A separate study showed similar results: dogs requiring dose delays and dosage reductions during chemotherapy for lymphoma demonstrated improved outcomes compared with those without adjustments [105].

6. What role do interspecies differences in pharmacokinetics, pharmacodynamics and pharmacogenomics play in tumour response to chemotherapy?

The pharmacologic treatment of cancer, regardless of human or pet origin, is a challenging endeavour, as medical oncologists must choose and use drugs with relatively narrow efficacy profiles while being aware of serious toxicities and while monitoring tumour response. Clinical pharmacology is defined as the study of drugs, and the application of clinical pharmacology attempts to predict and explain variable drug actions and interactions. Chemosensitivity depends

heavily on factors such as drug uptake into the cell, interaction within the cell and the cellular response to damage; exposure of tumour cells to chemotherapy effects is heavily dependent on pharmacologic effects. As the quantitative study of drug absorption, distribution, elimination and drug interactions, pharmacokinetics is often termed ‘what the body does to the drug’ and plays an integral role early in clinical study design [106]. Many methods of scaling have been developed to predict pharmacokinetic parameters from animals to humans, however little research has addressed scaling within different animal species. In humans, clearance is considered the most important pharmacokinetic parameter as it is directly linked to area under the curve [107]. Clearance of any drug from the body involves multiple organ systems and there are several allometric models that can be used to predict clearance in humans from animals (and vice versa). There are several excellent reviews highlighting numerous interspecies differences in drug pharmacokinetics, with an emphasis on drug development and the use of preclinical models [108–110]. Contrary to pharmacokinetics, pharmacodynamics, as the study of the drug dose and kinetics in relation to clinical effects, is often redefined simply as ‘what the drug does to the body’. Pharmacodynamic differences across species are often reported as differences in toxicity profiles for a specific drug in question. Quite prominently lacking in the veterinary literature is information on pharmacogenomic differences across species, in spite of the fact that the field of pharmacogenomics has erupted as a major area of advancement in humans. Pharmacogenomics, or the study of the role genetics plays in drug response, offers a host of additional reasons for altered responses to drugs such as those used in chemotherapy and an integrative systems pharmacology approach including pharmacokinetics, pharmacodynamics and pharmacogenomics is now proposed as an ideal method to approach drug regimen design [80,81,111].

6-Mercaptopurine (6-MP) is a core purine antimetabolite chemotherapy drug in maintenance protocols in childhood acute lymphoblastic leukaemia. 6-MP is inactive and undergoes activation to form 6-thioguanine (6-TG), which exerts cytotoxicity by incorporation into DNA and RNA, which is linked to cytotoxicity. 6-MP is cleared by either oxidation to the inactive 6-thiouric acid by xanthine oxidase or by S-methylation by thiopurine methyltransferase (TPMT) to yield 6-methyl mercaptopurine [80,112]. Haematopoietic cells do not have xanthine oxidase activity, thus leaving TPMT as the primary mechanism of metabolism [80,113]. In the absence of TPMT, 6-MP is metabolized by haematopoietic cells to produce high levels of 6-TG, causing profound haematologic toxicity. It is now recognized that there is significant variability in red blood cell TPMT activity in humans, with approximately 11% encoding for a nucleotide polymorphism associated with low TPMT activity [114,115]. This recognition has altered current practice paradigms as myelosuppression following treatment is directly related to TPMT phenotype. Greater than 60–65% of human patients experiencing extreme toxicity have TPMT deficiency, most of which can be detected by genetic testing for *TPMT*2*, *TPMT*3A* and *TPMT*3C* alleles [80,116–118]. Clinical guidelines incorporating this pharmacogenetic information for 6-MP in leukaemia are now recommended in order to manage both efficacy and toxicity [116–118]. Although 6-MP is not widely used in veterinary oncology, its prodrug

Table 1. Selected examples of species differences in drug pharmacokinetics, pharmacodynamics and pharmacogenomics that may influence response to cancer chemotherapy or targeted drug therapy.

parameter	species	feature/example
serum albumin binding	variable	canine and human albumin site II binding were very similar while albumin derived from rabbits, rats and cows were markedly different [127]
plasma protein binding	variable	total plasma protein content was highest in the dog compared with mouse, rat, rabbit, monkey and human [128]
protein binding affinity: alendronate	dog versus rats	alendronate demonstrated little binding in the dog as opposed to high binding in the rat [129]
immune response: macrophages	humans versus dogs, rats and mice	pulmonary alveolar macrophages in humans have highest phagocytic ability compared with rat, mouse or dogs suggesting some targeted drugs (liposomes) may effect species differences in response [109,130]
immune response: hypersensitivity and anaphylactic responses [109]	humans	shock organs include lung, larynx and vasculature
	dog	shock organ is classically considered the liver but includes all splanchnic circulation
	rat	shock organs include liver and intestine
	mouse	shock organs include vasculature and intestine
immune response: opsonization via complement proteins and immunoglobulins—Cremophor EL and polysorbate 80	dog	dogs display a much greater hypersensitivity to both Cremophor EL and polysorbate 80 compared with other species such as mice and pigs [131,132]
immune response: opsonization of liposomes—liposome encapsulated doxorubicin	rats versus dogs, pigs and humans	rats were markedly less sensitive to liposomal phospholipids compared with dogs, pigs and humans [131]
drug absorption from interstitial tissue	variable	macromolecules absorbed via capillaries in rats, whereas macromolecules often dependent on lymphatic absorption in dogs, sheep and humans [108,109,133,134]
drug delivery to tumour: colloidal osmotic pressure	variable	interstitial fluid pressure at the periphery of a tumour likely differs significantly between dogs, cats, rats and humans [135–137]
drug delivery to tumour: transport across blood-brain barrier	variable	active transporter expression is highly variable between humans, rodents, cows, pigs and dogs. Despite P-glycoprotein (ABCB1A) homology across species, significant differences in substrate recognition and transport efficiency have been noted between human and mouse [138–141]
breed-related physiologic differences: Sighthounds	dogs	lower volume of distribution of lipophilic compounds in Sighthounds compared with other breeds [142]
breed-related physiologic differences: dog size	dogs	gastrointestinal transit, fecal quality, intestinal permeability and GFR related to body size in dogs [143,144]

(Continued.)

Table 1. (Continued.)

parameter	species	feature/example
breed-related metabolic differences: CYP2D15 (similar to human CYP2D6)	Beagle dog versus humans	polymorphisms in CYP2D15 greatly affected metabolism of celecoxib across purebred Beagles [145]
breed-related pharmacogenetic differences	dogs—namely herding breeds (Collie, Australian shepherd, long-haired Whippet, Shetland Sheepdog, Old English Sheepdog, White Swiss Shepherd) [146,147]	polymorphisms in ABCB1 altered response associated with P-glycoprotein substrates [148–150]

azathioprine is commonly prescribed for various diseases. Dogs have variable red blood cell TPMT levels and while some breed tendencies were noted (Labrador Retrievers with high TPMT activity and Cocker Spaniels with low activity), there was a considerable range (ninefold) of activity across dogs [119]. Cats are recognized as being extremely sensitive to azathioprine and have lower red blood cell TPMT activity compared with dogs and humans [120,121]. Additional research in companion animals and other non-human species will build on preliminary research and explore the functional and clinical impact of TPMT polymorphisms to help identify altered drug responses, ultimately improving animal models in drug development [119,120,122].

While the TPMT story provides the best example of applied pharmacogenetics in human oncology, much work needs to be done to identify the impact of other known genetic and metabolic differences across species. It is widely recognized that cats can respond vastly differently to several drugs compared with other companion animals although the underlying reasons are not always clear. Generally speaking, drugs that are metabolized via conjugation are cleared slowly in cats compared with dogs and humans due to a lack of many conjugation enzymes besides TPMT, including UDP-glucuronosyltransferase (UGT) enzymes and *N*-acetyltransferase 2 (NAT2). The human UGT family consists of 19 different isoforms that are primarily expressed in liver, kidney and intestinal mucosa, which are primary sites of drug metabolism, thus highlighting a substantial difference between cats and humans, making drug dosing and response comparisons inherently difficult [123,124]. *N*-acetylation of amines in humans occurs via *N*-acetyltransferase enzymes NAT1 and NAT2 activity; dogs and related canids are deficient in NAT genes, emphasizing another primary difference in metabolism among companion animal species and humans [125]. Interestingly, NATs have been widely studied in humans due to their importance in xenobiotic metabolism while NAT polymorphism has been linked to population differences in drug metabolism [126] (figure 1).

There are many other examples of altered parameters affecting drug absorption and protein binding, drug delivery to the target tissue and toxicities that are beyond the scope of this review; table 1 provides additional examples of variables that may affect response to chemotherapy. Despite the overwhelming range of potential factors that can influence drug efficacy and tumour response, it is remarkable that correlations can be made across species in support of the field of comparative oncology.

7. Concluding remarks

Therapeutic response of a particular cancer to chemotherapy is very difficult to predict across the species. The use of rodent models to dissect the biology of cancer has proved invaluable in supporting the exponential growth of our understanding of this disease but rodents still prove to be poor models for predicting therapeutic responses leading to an incredibly costly linear drug development pathway. Naturally occurring cancer in dogs has been suggested as an alternative therapeutic model system that could prove to be more cost effective, with greater predictability and potentially allowing enormous savings in drug development costs. However, as we have seen, even with natural models we need to have a greater understanding of pharmacodynamics, pharmacokinetics and pharmacogenomics in natural models such as the dog. Publication of the canine genome and the development of a toolbox of reagents to study canine pharmacology and cancer biology will help to underpin progress in this area. However, to gain the optimal clinical benefit from comparative studies, we need to:

- obtain a greater understanding of the comparative biology of cancer between dogs and humans;
- develop a toolbox of reagents that can be used to dissect the biology of cancer in both species and underpinned by genomic, proteomic, metabolomic studies with appropriate bioinformatics;
- gain wider acceptance among the medical and scientific community that we must use the best model for a particular biological question. While rodent models have many benefits, they are not necessarily the best models for rational drug development;
- gain wider acceptance from the approvals agencies (FDA/EMA) that studies done in species other than the mouse may offer greater predictability of use of a drug in people; and
- conduct well-designed, statistically appropriate studies in veterinary patients.

As a final consideration, it may be important to adopt a more holistic systems biology approach to cancer chemotherapy across the species. In this approach, the whole patient and networks are considered rather than a 'reductionist' type study where 'cause and effect' are the only parameters. Reductionism focuses on the disease rather than on the individualization of treatment or on a multidimensional use of

drugs. To this notion of reductionism, the true benefit of chemotherapy for many human solid tumours has been brought into question by some researchers who suggest that chemotherapy neither improves survival nor provides a higher quality of life [91,151–153]. While chemotherapy for some cancers has decreased tumour size, tumour response comes at the expense of an increased risk of chemotherapy-induced neoplasia and an adversely affected lifestyle. At least for the foreseeable future, for the veterinary oncologist, and irrespective of apparent differences in tumour responses across species, the focus for veterinary patients such as the dog is maintaining or

improving an excellent quality-of-life, as perceived by the owner and/or attending veterinary clinician.

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References

- Langdon SP. 2012 Animal modeling of cancer pathology and studying tumor response to therapy. *Curr. Drug Targets* **13**, 1535–1547. (doi:10.2174/138945012803530152)
- Bailey MJ, Gazet JC, Smith IE, Steel GG. 1980 Chemotherapy of human breast-carcinoma xenografts. *Br. J. Cancer* **42**, 530–536. (doi:10.1038/bjc.1980.276)
- Favre R, Marotia L, Drancourt M, Jaquemier J, Delpero JR, Guerinel G, Carcassone Y. 1986 6-day subrenal capsule assay (SRCA) as a predictor of the response of advanced cancers to chemotherapy. *Eur. J. Cancer Clin. Oncol.* **22**, 1171–1178. (doi:10.1016/0277-5379(86)90318-4)
- Fiebig HH, Maier A, Burger AM. 2004 Clonogenic assay with established human tumour xenografts: correlation of *in vitro* to *in vivo* activity as a basis for anticancer drug discovery. *Eur. J. Cancer* **40**, 802–820. (doi:10.1016/j.ejca.2004.01.009)
- Inoue K, Fujimoto S, Ogawa M. 1983 Antitumor efficacy of seventeen anticancer drugs in human breast cancer xenograft (MX-1) transplanted in nude mice. *Cancer Chemother. Pharmacol.* **10**, 182–186. (doi:10.1007/BF00255758)
- Johnson JI *et al.* 2001 Relationships between drug activity in NCI preclinical *in vitro* and *in vivo* models and early clinical trials. *Br. J. Cancer* **84**, 1424–1431. (doi:10.1054/bjoc.2001.1796)
- Matter J, Bak M, Hahn EW, Volm M. 1988 Human tumor xenografts as model for drug testing. *Cancer Metastasis Rev.* **7**, 263–284. (doi:10.1007/BF00047755)
- Taetle R, Rosen F, Abramson I, Venditti J, Howell S. 1987 Use of nude mouse xenografts as preclinical drug screens: *in vivo* activity of established chemotherapeutic agents against melanoma and ovarian carcinoma xenografts. *Cancer Treat. Rep.* **71**, 297–304.
- Voskoglou-Nomikos T, Pater JL, Seymour L. 2003 Clinical predictive value of the *in vitro* cell line, human xenograft, and mouse allograft preclinical cancer models. *Clin. Cancer Res.* **9**, 4227–4239.
- Peterson JK, Houghton PJ. 2004 Integrating pharmacology and *in vivo* cancer models in preclinical and clinical drug development. *Eur. J. Cancer* **40**, 837–844. (doi:10.1016/j.ejca.2004.01.003)
- Gordon IK, Khanna C. 2010 Modeling opportunities in comparative oncology for drug development. *ILAR* **51**, 214–220. (doi:10.1093/ilar.51.3.214)
- Rangarajan A, Weinberg RA. 2003 Comparative biology of mouse versus human cells: modelling human cancer in mice. *Nat. Rev. Cancer* **3**, 952–959. (doi:10.1038/nrc1235)
- Frese KK, Tuveson DA. 2007 Maximizing mouse cancer models. *Nat. Rev. Cancer* **7**, 645–658. (doi:10.1038/nrc2192)
- Talmadge JE, Singh RK, Fidler IJ, Raz A. 2007 Murine models to evaluate novel and conventional therapeutic strategies for cancer. *Am. J. Pathol.* **170**, 793–804. (doi:10.2353/ajpath.2007.060929)
- Heyer J, Kwong LN, Lowe SW, Chin L. 2010 Non-germline genetically engineered mouse models for translational cancer research. *Nat. Rev. Cancer* **10**, 470–480. (doi:10.1038/nrc2877)
- Hanahan D, Weinberg RA. 2000 The hallmarks of cancer. *Cell* **100**, 57–70. (doi:10.1016/S0092-8674(00)81683-9)
- Hanahan D, Weinberg RA. 2011 Hallmarks of cancer: the next generation. *Cell* **144**, 646–674. (doi:10.1016/j.cell.2011.02.013)
- Mueller F, Fuchs B, Kaser-Hotz B. 2007 Comparative biology of human and canine osteosarcoma. *Anticancer Res.* **27**, 155–164.
- Paoloni M, Khanna C. 2008 Translation of new cancer treatments from pet dogs to humans. *Nat. Rev. Cancer* **8**, 147–156. (doi:10.1038/nrc2273)
- Porrello A, Cardelli P, Spugnini EP. 2006 Oncology of companion animals as a model for humans. An overview of tumor histotypes. *J. Exp. Clin. Cancer Res.* **25**, 97–105.
- Vail DM, MacEwen EG. 2000 Spontaneously occurring tumors of companion animals as models for human cancer. *Cancer Invest.* **18**, 781–792. (doi:10.3109/07357900009012210)
- Fleischer S, Sharkey M, Mealey K, Ostrander EA, Martinez M. 2008 Pharmacogenetic and metabolic differences between dog breeds: their impact on canine medicine and the use of the dog as a preclinical animal model. *AAPS J.* **10**, 110–119. (doi:10.1208/s12248-008-9011-1)
- Skipper HE, Schabel Jr FM, Mellett LB, Montgomery JA, Wilkoff LJ, Lloyd HH, Brockman RW. 1970 Implications of biochemical, cytokinetic, pharmacologic, and toxicologic relationships in the design of optimal therapeutic schedules. *Cancer Chemother. Rep. Part 1* **54**, 431–450.
- Vail DM, Pinkerton ME, Young KM. 2013 Hematopoietic tumors. In *Small animal clinical oncology* (eds SJ Withrow, DM Vail, RL Page), pp. 608–678, 5th edn. St Louis, MO: Saunders.
- Smith MA, Seibel NL, Altekruze SF, Ries LA, Melbert DL, O'Leary M, Smith FO, Reaman GH. 2010 Outcomes for children and adolescents with cancer: challenges for the twenty-first century. *J. Clin. Oncol.* **28**, 2625–2634. (doi:10.1200/JCO.2009.27.0421)
- Deininger MW, Druker BJ. 2004 SRCircumventing imatinib resistance. *Cancer Cell* **6**, 108–110. (doi:10.1016/j.ccr.2004.08.006)
- Gerrie AS, Power MM, Shepherd JD, Savage KJ, Sehn LH, Connors JM. 2014 Chemoresistance can be overcome with high-dose chemotherapy and autologous stem cell transplantation for relapsed and refractory Hodgkin lymphoma. *Ann. Oncol.* **25**, 2218–2223. (doi:10.1093/annonc/mdu387)
- Maxwell SA, Mousavi-Fard S. 2013 Non-Hodgkin's B-cell lymphoma: advances in molecular strategies targeting drug resistance. *Exp. Biol. Med.* **238**, 971–990. (doi:10.1177/1535370213498985)
- Kirkwood JM, Tarhini A, Sparano JA, Patel P, Schiller JH, Vergo MT, Benson III AB, Tawbi H. 2013 Comparative clinical benefits of systemic adjuvant therapy for paradigm solid tumors. *Cancer Treatment Rev.* **39**, 27–43. (doi:10.1016/j.ctrv.2012.03.007)
- Rosen G, Marcove RC, Huvos AG, Caparros BI, Lane JM, Nirenberg A, Cacavio A, Groshen S. 1983 Primary osteogenic sarcoma: eight-year experience with adjuvant chemotherapy. *J. Cancer Res. Clin. Oncol.* **106**, 55–67. (doi:10.1007/BF00625054)
- Gustafson DL PR. 2013 Cancer chemotherapy. In *Small animal clinical oncology* (eds SJ Withrow, DM Vail, RL Page), pp. 157–179, 5th edn. St Louis, MO: Saunders.
- Thamm DH, Vail DM. 2007 Aftershocks of cancer chemotherapy: managing adverse effects. *J. Am. Anim. Hosp. Assoc.* **43**, 1–7. (doi:10.5326/0430001)
- Bowles DB, Robson MC, Galloway PE, Walker L. 2010 Owner's perception of carboplatin in

- conjunction with other palliative treatments for cancer therapy. *J. Small Anim. Pract.* **51**, 104–112. (doi:10.1111/j.1748-5827.2009.00891.x)
34. Bronden LB, Rutteman GR, Flagstad A, Teske E. 2003 Study of dog and cat owners' perceptions of medical treatment for cancer. *Vet. Rec.* **152**, 77–80. (doi:10.1136/vr.152.3.77)
 35. Friedberg JW MP, Rimsza L, Fisher RI. 2011 Non-Hodgkin lymphomas. In *Cancer principles and practice of oncology* (eds VJ DeVita, TS Lawrence, SA Rosenberg), pp. 1855–1893, 9th edn. Philadelphia, PA: Lippincott Williams & Wilkins.
 36. Nastoupil LJ *et al.* 2014 Comparison of the effectiveness of frontline chemoimmunotherapy regimens for follicular lymphoma used in the United States. *Leuk. Lymphoma* **2014**, 1–28. (doi:10.3109/10428194.2014.953144)
 37. Jung SH *et al.* 2014 Weekly rituximab consolidation following four cycles of R-CHOP induction chemotherapy in very elderly patients with diffuse large B-cell lymphoma: Consortium for improving survival of lymphoma study (CISL). *Eur. J. Haematol.* (doi:10.1111/ejh.12459)
 38. Plosker GL, Figgitt DP. 2003 Rituximab: a review of its use in non-Hodgkin's lymphoma and chronic lymphocytic leukaemia. *Drugs* **63**, 803–843. (doi:10.2165/00003495-200363080-00005)
 39. Sueiro FA, Alessi AC, Vassallo J. 2004 Canine lymphomas: a morphological and immunohistochemical study of 55 cases, with observations on p53 immunoprotein expression. *J. Comp. Pathol.* **131**, 207–213. (doi:10.1016/j.jcpa.2004.04.002)
 40. Valli VE *et al.* 2011 Classification of canine malignant lymphomas according to the World Health Organization criteria. *Vet. Pathol.* **48**, 198–211. (doi:10.1177/0300985810379428)
 41. Vezzali E, Parodi AL, Marcato PS, Bettini G. 2010 Histopathologic classification of 171 cases of canine and feline non-Hodgkin lymphoma according to the WHO. *Vet. Comp. Oncol.* **8**, 38–49. (doi:10.1111/j.1476-5829.2009.00201.x)
 42. Flory AB, Rassnick KM, Erb HN, Garrett LD, Northrup NC, Selting KA, Phillips BS, Locke JE, Chretien JD. 2011 Evaluation of factors associated with second remission in dogs with lymphoma undergoing retreatment with a cyclophosphamide, doxorubicin, vincristine, and prednisone chemotherapy protocol: 95 cases (2000–2007). *J. Am. Vet. Med. Assoc.* **238**, 501–506. (doi:10.2460/javma.238.4.501)
 43. Allison DC, Carney SC, Ahlmann ER, Hendifar A, Chawla S, Fedenko A, Angeles C, Menendez LR. 2012 A meta-analysis of osteosarcoma outcomes in the modern medical era. *Sarcoma* **2012**, 704872. (doi:10.1155/2012/704872)
 44. Meyers PA, Heller G, Healey J, Huvos A, Lane J, Marcove R, Applewhite A, Vlamis V, Rosen G. 1992 Chemotherapy for nonmetastatic osteogenic sarcoma: the Memorial Sloan–Kettering experience. *J. Clin. Oncol.* **10**, 5–15.
 45. Cortes EP, Holland JF, Wang JJ, Sinks LF, Blom J, Senn H, Bank A, Glidewell O. 1974 Amputation and adriamycin in primary osteosarcoma. *N. Engl. J. Med.* **291**, 998–1000. (doi:10.1056/NEJM197411072911903)
 46. Link MP *et al.* 1986 The effect of adjuvant chemotherapy on relapse-free survival in patients with osteosarcoma of the extremity. *N. Engl. J. Med.* **314**, 1600–1606. (doi:10.1056/NEJM198606193142502)
 47. Winkler K *et al.* 1988 Neoadjuvant chemotherapy of osteosarcoma: results of a randomized cooperative trial (COSS-82) with salvage chemotherapy based on histological tumor response. *J. Clin. Oncol.* **6**, 329–337.
 48. Sleijfer S, Ouali M, van Glabbeke M, Krarup-Hansen A, Rodenhuis S, Le Cesne A, Hogendoorn PCW, Verweij J, Blay J-Y. 2010 Prognostic and predictive factors for outcome to first-line ifosfamide-containing chemotherapy for adult patients with advanced soft tissue sarcomas: an exploratory, retrospective analysis on large series from the European Organization for Research and Treatment of Cancer-Soft Tissue and Bone Sarcoma Group (EORTC-STBSG). *Eur. J. Cancer* **46**, 72–83. (doi:10.1016/j.ejca.2009.09.022)
 49. Patrikidou A, Domont J, Cioffi A, Le Cesne A. 2011 Treating soft tissue sarcomas with adjuvant chemotherapy. *Curr. Treat. Options Oncol.* **12**, 21–31. (doi:10.1007/s11864-011-0145-5)
 50. Gronchi A, Casali PG. 2013 Adjuvant therapy for high-risk soft tissue sarcoma in the adult. *Curr. Treat. Options Oncol.* **14**, 415–424. (doi:10.1007/s11864-013-0243-7)
 51. Scurr M. 2011 Histology-driven chemotherapy in soft tissue sarcomas. *Curr. Treat. Options Oncol.* **12**, 32–45. (doi:10.1007/s11864-011-0140-x)
 52. Pervaiz N, Colterjohn N, Farrokhlyar F, Tozer R, Figueredo A, Ghert M. 2008 A systematic meta-analysis of randomized controlled trials of adjuvant chemotherapy for localized resectable soft-tissue sarcoma. *Cancer* **113**, 573–581. (doi:10.1002/cncr.23592)
 53. Verma S, Younus J, Stys-Norman D, Haynes AE, Blackstein M. 2008 Meta-analysis of ifosfamide-based combination chemotherapy in advanced soft tissue sarcoma. *Cancer Treat. Rev.* **34**, 339–347. (doi:10.1016/j.ctrv.2008.01.005)
 54. Collaboration SM-A. 1997 Adjuvant chemotherapy for localised resectable soft-tissue sarcoma of adults: meta-analysis of individual data. (Sarcoma meta-analysis collaboration.) *Lancet* **350**, 1647–1654. (doi:10.1016/S0140-6736(97)08165-8)
 55. Sarcoma Meta-analysis Collaboration (SMAC). 2000 Adjuvant chemotherapy for localised resectable soft tissue sarcoma in adults. *Cochrane Database Syst. Rev.* 2000(4): CD001419. Available at <http://www.ncbi.nlm.nih.gov/pubmed/11034717>.
 56. Paoloni M *et al.* 2009 Canine tumor cross-species genomics uncovers targets linked to osteosarcoma progression. *BMC Genomics* **10**, 625. (doi:10.1186/1471-2164-10-625)
 57. Fenger JM, London CA, Kisseberth WC. 2014 Canine osteosarcoma: a naturally occurring disease to inform pediatric oncology. *ILAR J.* **55**, 69–85. (doi:10.1093/ilar/flu009)
 58. Withrow SJ, Wilkins RM. 2010 Cross talk from pets to people: translational osteosarcoma treatments. *ILAR J.* **51**, 208–213. (doi:10.1093/ilar.51.3.208)
 59. Rankin KS, Starkey M, Lunec J, Gerrard CH, Murphy S, Biswas S. 2012 Of dogs and men: comparative biology as a tool for the discovery of novel biomarkers and drug development targets in osteosarcoma. *Pediatr. Blood Cancer* **58**, 327–333. (doi:10.1002/pbc.23341)
 60. Thompson JP, Fugent MJ. 1992 Evaluation of survival times after limb amputation, with and without subsequent administration of cisplatin, for treatment of appendicular osteosarcoma in dogs: 30 cases (1979–1990). *J. Am. Vet. Med. Assoc.* **200**, 531–533.
 61. Spodnick GJ *et al.* 1992 Prognosis for dogs with appendicular osteosarcoma treated by amputation alone: 162 cases (1978–1988). *J. Am. Vet. Med. Assoc.* **200**, 995–999.
 62. Mauldin GN, Matus RE, Withrow SJ, Patnaik AK. 1988 Canine osteosarcoma. Treatment by amputation versus amputation and adjuvant chemotherapy using doxorubicin and cisplatin. *J. Vet. Intern. Med.* **2**, 177–180. (doi:10.1111/j.1939-1676.1988.tb00313.x)
 63. Selmic LE, Burton JH, Thamm DH, Withrow SJ, Lana SE. 2014 Comparison of carboplatin and doxorubicin-based chemotherapy protocols in 470 dogs after amputation for treatment of appendicular osteosarcoma. *J. Vet. Intern. Med.* **28**, 554–563. (doi:10.1111/jvim.12313)
 64. Phillips B, Powers BE, Dernel WS, Straw RC, Khanna C, Hogge GS, Vail DM. 2009 Use of single-agent carboplatin as adjuvant or neoadjuvant therapy in conjunction with amputation for appendicular osteosarcoma in dogs. *J. Am. Anim. Hosp. Assoc.* **45**, 33–38. (doi:10.5326/0450033)
 65. Bacon NJ, Ehrhart NP, Dernel WS, Lafferty M, Withrow SJ. 2008 Use of alternating administration of carboplatin and doxorubicin in dogs with microscopic metastases after amputation for appendicular osteosarcoma: 50 cases (1999–2006). *J. Am. Vet. Med. Assoc.* **232**, 1504–1510. (doi:10.2460/javma.232.10.1504)
 66. Ehrhart NP, Ryan SP, Fan TM. 2013 Tumors of the skeletal system. In *Small animal clinical oncology* (eds SJ Withrow, DM Vail, RL Page), pp. 463–503, 5th edn. St Louis, MO: Saunders.
 67. Ogilvie GK, Reynolds HA, Richardson RC, Withrow SJ, Norris AM, Henderson RA, Klausner JS, Fowler JD, McCaw D. 1989 Phase II evaluation of doxorubicin for treatment of various canine neoplasms. *J. Am. Vet. Med. Assoc.* **195**, 1580–1583.
 68. Liptak JM FL. 2013 Soft tissue sarcomas. In *Small animal clinical oncology* (eds SJ Withrow, DM Vail, RL Page), pp. 356–380, 5th edn. St Louis, MO: Saunders.
 69. Selting KA PB, Thompson LJ, Mittleman E, Tyler JW, Lafferty MH, Withrow SJ. 2005 Outcome of dogs

- with high-grade soft tissue sarcomas treated with and without adjuvant doxorubicin chemotherapy: 39 cases (1996–2004). *J. Am. Vet. Med. Assoc.* **227**, 1442–1448. (doi:10.2460/javma.2005.227.1442)
70. Kraus C, Pavard S, Promislow DE. 2013 The size-life span trade-off decomposed: why large dogs die young. *Am. Nat.* **181**, 492–505. (doi:10.1086/669665)
71. Patronek GJ, Waters DJ, Glickman LT. 1997 Comparative longevity of pet dogs and humans: implications for gerontology research. *J. Gerontol. A Biol. Sci. Med. Sci.* **52**, B171–B178. (doi:10.1093/gerona/52A.3.B171)
72. de Magalhaes JP, Costa J. 2009 A database of vertebrate longevity records and their relation to other life-history traits. *J. Evol. Biol.* **22**, 1770–1774. (doi:10.1111/j.1420-9101.2009.01783.x)
73. Caulin AF, Maley CC. 2011 Peto's paradox: evolution's prescription for cancer prevention. *Trends Ecol. Evol.* **26**, 175–182. (doi:10.1016/j.tree.2011.01.002)
74. Samaras TT, Storms LH. 1992 Impact of height and weight on life span. *Bull. World Health Organ.* **70**, 259–267.
75. Lebeau A. 1956 L'age du chien et celui de l'homme. Essai de statistique sur la mortalité canine. *Bull. Acad. Vet. Fr.* **26**, 229–232.
76. Greer KA, Hughes LM, Masternak MM. 2011 Connecting serum IGF-1, body size, and age in the domestic dog. *Age* **33**, 475–483. (doi:10.1007/s11357-010-9182-4)
77. Sutter NB *et al.* 2007 A single IGF1 allele is a major determinant of small size in dogs. *Science* **316**, 112–115. (doi:10.1126/science.1137045)
78. Rimbault M, Beale HC, Schoenebeck JJ, Hoopes BC, Allen JJ, Kilroy-Glynn P, Wayne RK, Sutter NB, Ostrander EA. 2013 Derived variants at six genes explain nearly half of size reduction in dog breeds. *Genome Res.* **23**, 1985–1995. (doi:10.1101/gr.157339.113)
79. Amsellem PM, Selmic LE, Wypij JM, Bacon NJ, Culp WT, Ehrhart NP, Powers BE, Stryhn H, Farese JP. 2014 Appendicular osteosarcoma in small-breed dogs: 51 cases (1986–2011). *J. Am. Vet. Med. Assoc.* **245**, 203–210. (doi:10.2460/javma.245.2.203)
80. Walko CM, Ikediobi O. 2012 Pharmacogenomic applications in oncology. *J. Pharm. Pract.* **25**, 439–446. (doi:10.1177/0897190012448308)
81. Walko CM, McLeod H. 2009 Pharmacogenomic progress in individualized dosing of key drugs for cancer patients. *Nat. Clin. Pract. Oncol.* **6**, 153–162. (doi:10.1038/nncponc1303)
82. Gurney H. 1996 Dose calculation of anticancer drugs: a review of the current practice and introduction of an alternative. *J. Clin. Oncol.* **14**, 2590–2611.
83. Gao B, Klumpen HJ, Gurney H. 2008 Dose calculation of anticancer drugs. *Expert Opin. Drug Metab. Toxicol.* **4**, 1307–1319. (doi:10.1517/17425255.4.10.1307)
84. Freireich EJ, Gehan EA, Rall DP, Schmidt LH, Skipper HE. 1966 Quantitative comparison of toxicity of anticancer agents in mouse, rat, hamster, dog, monkey, and man. *Cancer Chemother. Rep.* **1** **50**, 219–244.
85. Goldsmith MA, Slavik M, Carter SK. 1975 Quantitative prediction of drug toxicity in humans from toxicology in small and large animals. *Cancer Res.* **35**, 1354–1364.
86. Pinkel D. 1958 The use of body surface area as a criterion of drug dosage in cancer chemotherapy. *Cancer Res.* **18**, 853–856.
87. Page RL, Macy DW, Thrall DE, Dewhirst MW, Allen SL, Heidner GL, Sim DA, McGee ML, Gillette EL. 1988 Unexpected toxicity associated with use of body surface area for dosing melphalan in the dog. *Cancer Res.* **48**, 288–290.
88. Arrington KA, Legendre AM, Tabeling GS, Frazier DL. 1994 Comparison of body surface area-based and weight-based dosage protocols for doxorubicin administration in dogs. *Am. J. Vet. Res.* **55**, 1587–1592.
89. Ogilvie GK, Moore AS, Curtis CR. 1989 Evaluation of cisplatin-induced emesis in dogs with malignant neoplasia: 115 cases (1984–1987). *J. Am. Vet. Med. Assoc.* **195**, 1399–1403.
90. Ogilvie GK *et al.* 1989 Acute and short-term toxicoses associated with the administration of doxorubicin to dogs with malignant tumors. *J. Am. Vet. Med. Assoc.* **195**, 1584–1587.
91. Greek R, Rice MJ. 2012 Animal models and conserved processes. *Theor. Biol. Med. Model.* **9**, 40. (doi:10.1186/1742-4682-9-40)
92. Price GS, Frazier DL. 1998 Use of body surface area (BSA)-based dosages to calculate chemotherapeutic drug dose in dogs: I. Potential problems with current BSA formulae. *J. Vet. Intern. Med.* **12**, 267–271. (doi:10.1111/j.1939-1676.1998.tb02121.x)
93. Frazier DL, Price GS. 1998 Use of body surface area to calculate chemotherapeutic drug dose in dogs: II. Limitations imposed by pharmacokinetic factors. *J. Vet. Intern. Med.* **12**, 272–278. (doi:10.1111/j.1939-1676.1998.tb02122.x)
94. Bailey DB, Rassnick KM, Dykes NL, Pendyala L. 2009 Phase I evaluation of carboplatin by use of a dosing strategy based on a targeted area under the platinum concentration-versus-time curve and individual glomerular filtration rate in cats with tumors. *Am. J. Vet. Res.* **70**, 770–776. (doi:10.2460/ajvr.70.6.770)
95. Bailey DB, Rassnick KM, Erb HN, Dykes NL, Hoopes PJ, Page RL. 2004 Effect of glomerular filtration rate on clearance and myelotoxicity of carboplatin in cats with tumors. *Am. J. Vet. Res.* **65**, 1502–1507. (doi:10.2460/ajvr.2004.65.1502)
96. Wittenburg LA, Thamm DH, Gustafson DL. 2014 Development of a limited-sampling model for prediction of doxorubicin exposure in dogs. *Vet. Comp. Oncol.* **12**, 114–119. (doi:10.1111/j.1476-5829.2012.00340.x)
97. Engels FK, Loos WJ, van der Bol JM, de Bruijn P, Mathijssen RH, Verweij J, Mathot RAA. 2011 Therapeutic drug monitoring for the individualization of docetaxel dosing: a randomized pharmacokinetic study. *Clin. Cancer Res.* **17**, 353–362. (doi:10.1158/1078-0432.CCR-10-1636)
98. Kline CL *et al.* 2014 Personalized dosing via pharmacokinetic monitoring of 5-fluorouracil might reduce toxicity in early- or late-stage colorectal cancer patients treated with infusional 5-fluorouracil-based chemotherapy regimens. *Clin. Colorectal Cancer* **13**, 119–126. (doi:10.1016/j.clcc.2013.11.001)
99. Rousseau A, Marquet P. 2002 Application of pharmacokinetic modelling to the routine therapeutic drug monitoring of anticancer drugs. *Fundam. Clin. Pharmacol.* **16**, 253–262. (doi:10.1046/j.1472-8206.2002.00086.x)
100. Salas S *et al.* 2006 Therapeutic drug monitoring for dose individualization of Cisplatin in testicular cancer patients based upon total platinum measurement in plasma. *Ther. Drug Monit.* **28**, 532–539. (doi:10.1097/00007691-200608000-00008)
101. Mercier C *et al.* 2006 Dose individualization of carboplatin after a 120-hour infusion schedule: higher dose intensity but fewer toxicities. *Ther. Drug Monit.* **28**, 212–218. (doi:10.1097/01.ftd.0000198646.32128.ef)
102. Gamelin E *et al.* 2008 Individual fluorouracil dose adjustment based on pharmacokinetic follow-up compared with conventional dosage: results of a multicenter randomized trial of patients with metastatic colorectal cancer. *J. Clin. Oncol.* **26**, 2099–2105. (doi:10.1200/JCO.2007.13.3934)
103. Capitain O, Asevoaia A, Boisdron-Celle M, Poirier AL, Morel A, Gamelin E. 2012 Individual fluorouracil dose adjustment in FOLFOX based on pharmacokinetic follow-up compared with conventional body-area-surface dosing: a phase II, proof-of-concept study. *Clin. Colorectal Cancer* **11**, 263–267. (doi:10.1016/j.clcc.2012.05.004)
104. Vaughan A, Johnson JL, Williams LE. 2007 Impact of chemotherapeutic dose intensity and hematologic toxicity on first remission duration in dogs with lymphoma treated with a chemoradiotherapy protocol. *J. Vet. Intern. Med.* **21**, 1332–1339. (doi:10.1111/j.1939-1676.2007.tb01956.x)
105. Sorenmo K, Overley B, Krick E, Ferrara T, LaBlanc A, Shofer F. 2010 Outcome and toxicity associated with a dose-intensified, maintenance-free CHOP-based chemotherapy protocol in canine lymphoma: 130 cases. *Vet. Comp. Oncol.* **8**, 196–208. (doi:10.1111/j.1476-5829.2010.00222.x)
106. Ratain MJ, Mick R. 1996 Principles of pharmacokinetics and pharmacodynamics. In *Principles of antineoplastic drug development and pharmacology basic and clinical oncology* (eds RL Schilsky, GA Milano, MJ Ratain), pp. 123–142. New York, NY: Marcel Dekker.
107. Mahmood I. 2007 Application of allometric principles for the prediction of pharmacokinetics in human and veterinary drug development. *Adv. Drug Deliv. Rev.* **59**, 1177–1192. (doi:10.1016/j.addr.2007.05.015)

108. Martinez MN. 2009 Interspecies differences in physiology and pharmacology: extrapolating preclinical data to human populations. In *Preclinical drug development* (eds MC Rogge, D Taft), pp. 35–70. Boca Raton, FL: Taylor and Francis Group.
109. Martinez MN. 2011 Factors influencing the use and interpretation of animal models in the development of parenteral drug delivery systems. *AAPS J.* **13**, 632–649. (doi:10.1208/s12248-011-9303-8)
110. Martignoni M, Groothuis GM, de Kanter R. 2006 Species differences between mouse, rat, dog, monkey and human CYP-mediated drug metabolism, inhibition and induction. *Expert Opin. Drug Metab. Toxicol.* **2**, 875–894. (doi:10.1517/17425255.2.6.875)
111. Gumbo T. 2008 Integrating pharmacokinetics, pharmacodynamics and pharmacogenomics to predict outcomes in antibacterial therapy. *Curr. Opin. Drug Discov. Dev.* **11**, 32–42.
112. Eilon GB. 1989 The purine path to chemotherapy. *Science* **244**, 41–47. (doi:10.1126/science.2649979)
113. Parks DA, Granger DN. 1986 Xanthine oxidase: biochemistry, distribution and physiology. *Acta Physiol. Scand. Suppl.* **548**, 87–99.
114. McLeod HL, Relling MV, Liu Q, Pui CH, Evans WE. 1995 Polymorphic thiopurine methyltransferase in erythrocytes is indicative of activity in leukemic blasts from children with acute lymphoblastic leukemia. *Blood* **85**, 1897–1902.
115. Weinshilboum RM, Sladek SL. 1980 Mercaptopurine pharmacogenetics: monogenic inheritance of erythrocyte thiopurine methyltransferase activity. *Am. J. Hum. Genet.* **32**, 651–662.
116. Lopez-Lopez E, Gutierrez-Camino A, Bilbao-Aldaiturriaga N, Pombar-Gomez M, Martin-Guerrero I, Garcia-Orad A. 2014 Pharmacogenetics of childhood acute lymphoblastic leukemia. *Pharmacogenomics* **15**, 1383–1398. (doi:10.2217/pgs.14.106)
117. Swen JJ *et al.* 2011 Pharmacogenetics: from bench to byte—an update of guidelines. *Clin. Pharmacol. Ther.* **89**, 662–673. (doi:10.1038/clpt.2011.34)
118. Relling MV *et al.* 2011 Clinical pharmacogenetics implementation consortium guidelines for thiopurine methyltransferase genotype and thiopurine dosing. *Clin. Pharmacol. Ther.* **89**, 387–391. (doi:10.1038/clpt.2010.320)
119. Salavaggione OE, Kidd L, Prondzinski JL, Szumlanski CL, Pankratz VS, Wang L, Trepanier L, Weinshilboum RM. 2002 Canine red blood cell thiopurine S-methyltransferase: companion animal pharmacogenetics. *Pharmacogenetics* **12**, 713–724. (doi:10.1097/00008571-200212000-00005)
120. Salavaggione OE, Yang C, Kidd LB, Thomae BA, Pankratz VS, Trepanier LA, Weinshilboum RM. 2004 Cat red blood cell thiopurine S-methyltransferase: companion animal pharmacogenetics. *J. Pharmacol. Exp. Ther.* **308**, 617–626. (doi:10.1124/jpet.103.059055)
121. Court MH. 2013 Feline drug metabolism and disposition: pharmacokinetic evidence for species differences and molecular mechanisms. *Vet. Clin. N. Am. Small Anim. Pract.* **43**, 1039–1054. (doi:10.1016/j.cvsm.2013.05.002)
122. Salavaggione OE, Wang L, Wiepert M, Yee VC, Weinshilboum RM. 2005 Thiopurine S-methyltransferase pharmacogenetics: variant allele functional and comparative genomics. *Pharmacogen. Genomics* **15**, 801–815. (doi:10.1097/01.fpc.0000174788.69991.6b)
123. Court MH, Zhang X, Ding X, Yee KK, Hesse LM, Finel M. 2012 Quantitative distribution of mRNAs encoding the 19 human UDP-glucuronosyltransferase enzymes in 26 adult and 3 fetal tissues. *Xenobiotica* **42**, 266–277. (doi:10.3109/00498254.2011.618954)
124. Court MH, Greenblatt DJ. 1997 Molecular basis for deficient acetaminophen glucuronidation in cats. An interspecies comparison of enzyme kinetics in liver microsomes. *Biochem. Pharmacol.* **53**, 1041–1047. (doi:10.1016/S0006-2952(97)00072-5)
125. Trepanier LA, Ray K, Winand NJ, Spielberg SP, Cribb AE. 1997 Cytosolic arylamine N-acetyltransferase (NAT) deficiency in the dog and other canids due to an absence of NAT genes. *Biochem. Pharmacol.* **54**, 73–80. (doi:10.1016/S0006-2952(97)00140-8)
126. Sim E, Abuhammad A, Ryan A. 2014 Arylamine N-acetyltransferases: from drug metabolism and pharmacogenetics to drug discovery. *Br. J. Pharmacol.* **171**, 2705–2725. (doi:10.1111/bph.12598)
127. Kosa T, Maruyama T, Otagiri M. 1997 Species differences of serum albumins: I. Drug binding sites. *Pharm. Res.* **14**, 1607–1612. (doi:10.1023/A:1012138604016)
128. Davies B, Morris T. 1993 Physiological parameters in laboratory animals and humans. *Pharm. Res.* **10**, 1093–1095. (doi:10.1023/A:1018943613122)
129. Lin JH, Chen IW, deLuna FA. 1994 Nonlinear kinetics of alendronate. Plasma protein binding and bone uptake. *Drug Metab. Dispos.* **22**, 400–405.
130. Haley PJ. 2003 Species differences in the structure and function of the immune system. *Toxicology* **188**, 49–71. (doi:10.1016/S0300-483X(03)00043-X)
131. Szebeni J, Alving CR, Rosivall L, Bunger R, Baranyi L, Bedocs P, Tóth M, Barenholz Y. 2007 Animal models of complement-mediated hypersensitivity reactions to liposomes and other lipid-based nanoparticles. *J. Liposome Res.* **17**, 107–117. (doi:10.1080/08982100701375118)
132. Poirier VJ, Hershey AE, Burgess KE, Phillips B, Turek MM, Forrest LJ, Beaver L, Vail DM. 2004 Efficacy and toxicity of paclitaxel (Taxol) for the treatment of canine malignant tumors. *J. Vet. Intern. Med.* **18**, 219–222. (doi:10.1111/j.1939-1676.2004.tb00164.x)
133. Trevaskis NL, Caliph SM, Nguyen G, Tso P, Charman WN, Porter CJ. 2013 A mouse model to evaluate the impact of species, sex, and lipid load on lymphatic drug transport. *Pharm. Res.* **30**, 3254–3270. (doi:10.1007/s11095-013-1000-0)
134. Porter CJ, Edwards GA, Charman SA. 2001 Lymphatic transport of proteins after s.c. injection: implications of animal model selection. *Adv. Drug Deliv. Rev.* **50**, 157–171. (doi:10.1016/S0169-409X(01)00153-3)
135. Jain RK. 1987 Transport of molecules in the tumor interstitium: a review. *Cancer Res.* **47**, 3039–3051.
136. Netti PA *et al.* 1999 Enhancement of fluid filtration across tumor vessels: implication for delivery of macromolecules. *Proc. Natl Acad. Sci. USA* **96**, 3137–3142. (doi:10.1073/pnas.96.6.3137)
137. Thomas LA, Brown SA. 1992 Relationship between colloid osmotic pressure and plasma protein concentration in cattle, horses, dogs, and cats. *Am. J. Vet. Res.* **53**, 2241–2244.
138. Gerhart DZ, Leino RL, Borson ND, Taylor WE, Gronlund KM, McCall AL, Drewes LR. 1995 Localization of glucose transporter GLUT 3 in brain: comparison of rodent and dog using species-specific carboxyl-terminal antisera. *Neuroscience* **66**, 237–246. (doi:10.1016/0306-4522(94)00544-F)
139. Westerhout J, Danhof M, De Lange EC. 2011 Preclinical prediction of human brain target site concentrations: considerations in extrapolating to the clinical setting. *J. Pharm. Sci.* **100**, 3577–3593. (doi:10.1002/jps.22604)
140. Borst P, Schinkel AH. 2013 P-glycoprotein ABCB1: a major player in drug handling by mammals. *J. Clin. Invest.* **123**, 4131–4133. (doi:10.1172/JCI70430)
141. Syvanen S, Lindhe O, Palner M, Kornum BR, Rahman O, Langstrom B, Knudsen GM, Hammarlund-Udenaes M. 2009 Species differences in blood-brain barrier transport of three positron emission tomography radioligands with emphasis on P-glycoprotein transport. *Drug Metab. Dispos.* **37**, 635–643. (doi:10.1124/dmd.108.024745)
142. Hay Kraus BL, Greenblatt DJ, Venkatakrishnan K, Court MH. 2000 Evidence for propofol hydroxylation by cytochrome P4502B11 in canine liver microsomes: breed and gender differences. *Xenobiotica* **30**, 575–588. (doi:10.1080/004982500406417)
143. Hernot DC, Biourge VC, Martin LJ, Dumon HJ, Nguyen PG. 2005 Relationship between total transit time and faecal quality in adult dogs differing in body size. *J. Anim. Physiol. Anim. Nutr.* **89**, 189–193. (doi:10.1111/j.1439-0396.2005.00544.x)
144. Randell SC, Hill RC, Scott KC, Omori M, Burrows CF. 2001 Intestinal permeability testing using lactulose and rhamnose: a comparison between clinically normal cats and dogs and between dogs of different breeds. *Res. Vet. Sci.* **71**, 45–49. (doi:10.1053/rvsc.2001.0483)
145. Paulson SK, Engel L, Reitz B, Bolton S, Burton EG, Maziasz TJ, Yan B, Schoenhard GL. 1999 Evidence for polymorphism in the canine metabolism of the cyclooxygenase 2 inhibitor, celecoxib. *Drug Metab. Dispos.* **27**, 1133–1142.
146. Gramer I, Leidolf R, Doring B, Klintzsch S, Kramer EM, Yalcin E, Petzinger E, Geyer J. 2011 Breed distribution of the nt230(del4) MDR1 mutation in dogs. *Vet. J.* **189**, 67–71. (doi:10.1016/j.tvjl.2010.06.012)

147. Mealey KL, Meurs KM. 2008 Breed distribution of the ABCB1-1Delta (multidrug sensitivity) polymorphism among dogs undergoing ABCB1 genotyping. *J. Am. Vet. Med. Assoc.* **233**, 921–924. (doi:10.2460/javma.233.6.921)
148. Mealey KL. 2008 Canine ABCB1 and macrocyclic lactones: heartworm prevention and pharmacogenetics. *Vet. Parasitol.* **158**, 215–222. (doi:10.1016/j.vetpar.2008.09.009)
149. Mealey KL, Fidel J, Gay JM, Impellizeri JA, Clifford CA, Bergman PJ. 2008 ABCB1-1Δ polymorphism can predict hematologic toxicity in dogs treated with vincristine. *J. Vet. Intern. Med.* **22**, 996–1000. (doi:10.1111/j.1939-1676.2008.0122.x)
150. Munana KR, Nettifee-Osborne JA, Bergman Jr RL, Mealey KL. 2012 Association between ABCB1 genotype and seizure outcome in Collies with epilepsy. *J. Vet. Intern. Med.* **26**, 1358–1364. (doi:10.1111/j.1939-1676.2012.01006.x)
151. Heng HH. 2008 The conflict between complex systems and reductionism. *JAMA* **300**, 1580–1581. (doi:10.1001/jama.300.13.1580)
152. Bear HD. 2003 Earlier chemotherapy for breast cancer: perhaps too late but still useful. *Ann. Surg. Oncol.* **10**, 334–335. (doi:10.1245/ASO.2003.02.023)
153. Mitra I. 2007 The disconnection between tumor response and survival. *Nat. Clin. Pract. Oncol.* **4**, 203. (doi:10.1038/ncponc0772)