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Article

Cytoskeletal Network Morphology Regulates Intracellular Transport Dynamics

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ABSTRACT Intracellular transport is essential for maintaining proper cellular function in most eukaryotic cells, with perturbations in active transport resulting in several types of disease. Efficient delivery of critical cargos to specific locations is accomplished through a combination of passive diffusion and active transport by molecular motors that ballistically move along a network of cytoskeletal filaments. Although motor-based transport is known to be necessary to overcome cytoplasmic crowding and the limited range of diffusion within reasonable timescales, the topological features of the cytoskeletal network that regulate transport efficiency and robustness have not been established. Using a continuum diffusion model, we observed that the time required for cellular transport was minimized when the network was localized near the nucleus. In simulations that explicitly incorporated network spatial architectures, total filament mass was the primary driver of network transit times. However, filament traps that redirect cargo back to the nucleus caused large variations in network transport. Filament polarity was more important than filament orientation in reducing average transit times, and transport properties were optimized in networks with intermediate motor on and off rates. Our results provide important insights into the functional constraints on intracellular transport under which cells have evolved cytoskeletal structures, and have potential applications for enhancing reactions in biomimetic systems through rational transport network design.

INTRODUCTION

The transport of intracellular cargos occurs in virtually all eukaryotic cells and is essential for many cellular functions. Whereas diffusion is sufficient for the transport of small (~nm) molecules, diffusion becomes prohibitively slow for large cargos traversing extremely crowded and relatively large cellular distances; these cargos thus require an active transport system (1,2). Such intracellular transport is carried out by molecular motors that move along a complex system of cytoskeletal polymers such as actin filaments, actin bundles, and microtubules (3). The three classes of molecular motors (myosin, kinesin, dynein) that transport cargo along the cytoskeleton are broadly categorized by their transport behavior and by the filament type upon which they move. Most myosins travel toward the barbed (+) end of actin filaments (4), which tend to form random networks (5). Most kinesins move toward the plus end of microtubules, which are typically radially oriented and polarized with the (+)end pointed away from the nucleus (toward the cell membrane) (6), whereas dyneins transport cargo toward the minus end (-) of their microtubule track (7). Despite a wealth of information about the biophysical properties of motors (8) and the filaments upon which they walk (9), we have little knowledge of the relative importance of various transport variables in the vast phase space of network, motor, and cargo properties, especially in the context of explicit models of physical network architecture. Computational models provide the means to identify how these transport variables influence outcomes such as the mean and variation in transport time between cellular locations that may impact cellular fitness.

Conceptual frameworks of active intracellular transport (6,10) have considered a cytoskeletal network with organized microtubules that serve as tracks for long-distance transport, whereas random actin-filament networks are utilized for local transport. Microtubules are polarized and radially oriented away from the nucleus and the centrosome (9), enabling transport from the nucleus to the periphery or vice versa in an approximately linear manner. In conjunction, actin filaments are thought to underlie local transport from the microtubule network to the remainder of the cell (11) via a randomly oriented network that is distributed throughout the cytoplasm. Although there have been careful studies of motor properties (8) and of the structure of individual network filaments (9), we still lack understanding of how motor properties coordinate with the topology of cytoskeletal networks to yield efficient, reliable transport.

Previous computational studies of intracellular transport mainly relied on reaction-hyperbolic partial differential equation modeling and so-called virtual network models that are based on the theory of intermittent dynamics in



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the context of search processes (2,12-15). Virtual network models assume that cargos repeatedly bind and unbind motors (5,15), and as a result exhibit phases of diffusion for random lengths of time that are interrupted by ballistic excursions of constant velocity and direction. Because the ballistic motion representing motor binding is assumed to be uniform in space and direction, heterogeneity in the network architecture is usually ignored. A previous study used an explicit filament network model to determine the effect of motor processivity on intracellular transport times (16). However, it has not yet been determined how the morphological properties of the network itself dictate transport outcomes relative to motor properties.

In this study, we use simulations to determine how cells regulate transport via changes in network and motor properties, such as filament localization, polarity, and orientation and motor mobility and binding. We first use a simple continuum model of increased bulk diffusion in a shell representing the cytoskeletal network to show that transport time from the nucleus to the periphery is minimized when the shell is localized near the nucleus, demonstrating the potential importance of the overall spatial configuration of the network. Simulations of ballistic motion along polarized filaments with explicit spatial extent reveal that particular filament arrangements near the nucleus can constitute traps (regions in which cargos spend increased amounts of time) that lead to large variations in the time required to achieve transport from the nucleus to the cell membrane. This variability, which can be mitigated by distributing the network mass over more filaments with shorter lengths, may be an important constraint on network organization. We also demonstrate that polarizing filaments with the direction of transport facing away from the nucleus is significant in reducing transit time, although the precise distribution of the angles of orientation has little effect. Finally, we determine that high on rates and/or low off rates, potentially representative of the binding properties of cargo complexes with multiple motors, actually slow cargo transport and increase variability, suggesting that single motors can more robustly transport cargo than a collection of multiple motors over random networks.

MATERIALS AND METHODS

For our continuum model diffusion simulations, cargo complexes diffused by taking random steps of fixed length (100 nm), with the time required per step determined by the bulk diffusion constant of the region in which the cargo was located. For explicit networks, simulations were initialized by placing circular cargo-motor complexes 100 nm in radius at random locations on the nuclear boundary. These complexes subsequently diffused randomly in steps of fixed length (100 nm) until they bound a filament with which the cargo overlapped at rate k_{on} . Filaments were represented as line segments with polarity. During network initialization, the centers of mass of filaments with a portion lying outside the cell or inside the nucleus were shifted inward or outward, respectively, along the radial direction until the filament was completely inside the cell, thus ensuring that all filaments were of equal length. Active motor transport occurred in 100-nm steps, with displacement in the direction of the filament's (+) end at a velocity of 1 μ m/s. During active transport, cargos dissociated at the motor off rate k_{off} or when they encountered the end of the filament. After unbinding, cargos were moved precisely adjacent to but not overlapping the filament. An effective cytoplasmic viscosity for cargos of 0.05 Pa s (1,17,18) was assumed, resulting in a diffusion constant of $D = 0.051 \mu$ m²/s for a cargo-motor complex with radius of 100 nm. Additional methods details appear in the Supporting Material.

RESULTS

Localizing transport systems near the nucleus minimizes cargo-transport time

The endoplasmic reticulum, an organelle that is located near and is continuous with the outer nuclear membrane, plays a central role in membrane and protein biosynthesis (19). We therefore focused our modeling on the transport of cargos that originate from the surface of an impenetrable nuclear membrane and terminate at the cell (cytoplasmic) membrane. Because imaging studies generally visualize cells adhered to a substrate, which tends to produce a flattened morphology, we focused our modeling on transport in two dimensions. Although eukaryotic cell shape and size can vary across a broad range, we simulated a circular, twodimensional cell with radius $R = 10 \ \mu$ m, the typical size of well-studied eukaryotic cell lines (20).

To investigate whether the spatial localization of the cytoskeletal network affects the time required to transport cargo from the nucleus to the membrane, we initially implemented a simple continuum diffusion model of cargos that originate at the precise center of a cell (in the absence of a nucleus) and are transported solely by diffusion with constant *D*; transport terminates when the cargo reaches the cell membrane (Fig. 1 *A*). The following diffusion equation describes the concentration of cargos c(x, y, t) diffusing in two dimensions:

$$\frac{\partial c}{\partial t} = D\left(\frac{\partial^2 c}{\partial x^2} + \frac{\partial^2 c}{\partial y^2}\right).$$
 (1)

For the transport problems that we consider, we assume a constant source at either the center of the cell or at the nuclear boundary, and zero cargo concentration along the cell boundary.

At steady state, Eq. 1. transformed into polar coordinates becomes the following:

$$D\left(\frac{\partial^2 c}{\partial x^2} + \frac{\partial^2 c}{\partial y^2}\right) = \left(\frac{\partial^2 c}{\partial r^2} + \frac{1}{r}\frac{\partial c}{\partial r} + \frac{1}{r^2}\frac{\partial^2 c}{\partial \theta^2}\right) = 0.$$
 (2)

Because the system is symmetric with respect to θ , the solution is the following:

$$c(r,\theta) = C_1 + C_2 \log r. \tag{3}$$

The spatial distribution of cargo residence time can then be calculated by applying the appropriate boundary conditions; see the Supporting Material for full derivation and analytical solutions.



FIGURE 1 Localization of a cytoskeletal network near the nuclear surface minimizes transport time to the membrane in a continuum diffusion model. (A) In these simulations, cargo-motor complexes move through the cytoplasm, which is modeled by bulk (*blue*) and network (*green*) regions. Our simulations are typically initialized with cargo on the surface of an impenetrable nucleus (*pink*), and transport ends when the cargo reaches the cell membrane (*red circle*). The cytoskeletal network is represented by an annulus of width w located at an inner radius R_a within which the diffusion constant is increased to $D_a =$ $1.1 \ \mu m^2/s$ from $D = 0.011 \ \mu m^2/s$ in the bulk (54). The annulus is placed outside of a nucleus with radius R_n . (B) Average cargo residence time peaked ~3.5 μm from the cell center for simulations of 10,000 cargos that terminate transport at the cell membrane in a cell with no nucleus ($R_n = 0$) and only bulk diffusion with constant D. The gray line indicates the analytical calculation of residence times (see Supporting Material). (C) Mean first passage time (MFPT) after addition of an annulus with diffusion constant $D_a = 1.1 \ \mu m^2/s$, width $w = 3 \ \mu m$. The annulus position was shifted from the nuclear surface to the cell membrane, and each result is the mean over 1,000 cargos. Black lines indicate analytically determined residence times. For $R_n \leq 2 \ \mu m$, MFPT was minimized at an intermediate R_a ; for larger R_n , MFPT was minimized with the annulus positioned adjacent to the nucleus. To see this figure in color, go online.

In this simple scenario, cargo residence time as a function of distance from the cell center peaked at an intermediate radius (Fig. 1 B). To introduce the effects of active transport in a simplified manner, we represented a cytoskeletal network as an annulus of fixed width w, whose inner radius is R_a , within which the diffusion constant is increased to $D_a = 100D$ to account for rapid motor transport along a random network. This annulus was placed outside a nucleus with radius R_n . For small nuclear radii ($R_n \leq 2 \mu m$) and an annulus width of $w = 3 \mu m$, the mean first passage time (MFPT) of cargo transport from the nuclear surface to the cell membrane was minimized when the inner boundary of the annulus was placed away from the nucleus, with the center of the annulus at a radius of ~3.5 μ m (Fig. 1 C). This minimal MFPT position corresponds approximately to the location of the peak residence time in the absence of a network (Fig. 1 B), suggesting that localization of enhanced diffusion around regions of peak residence times provides the greatest reduction in MFPT. Annuli closer to the cell membrane led to higher MFPTs (Fig. 1 C), even though the total annulus area increased linearly with R_a . When we instead fixed the area of the annulus such that w was inversely proportional to R_a , the MFPT increased monotonically as the annulus was shifted away from the nucleus, regardless of R_n (Fig. S1). Thus, for cells with $R_n \ge R/4$, a range encompassing typical nuclear to membrane size ratios, accelerating diffusion closer to the nucleus generally reduces MFPT.

Filament network architecture affects transport-time variability

To determine whether specific network architectures affect transport properties, we moved away from continuum diffusion simulations and introduced active transport via ballistic motion of cargo-motor complexes along explicitly modeled filaments. In our model, as in vivo, the filaments are characterized by their length, orientation, and polarity. Orientation and polarity of the filaments are defined relative to the cellular geometry. The filament angle θ is defined as the angle between the filament's direction vector (from the filament midpoint to its plus end) and a vector pointing radially outward. θ modulo π radians is defined as the orientation of a filament. Filament polarity is defined by the sign of the projection of the filament's direction vector on the vector pointing radially outward, which is the sign of $\cos(\theta)$. Thus, a positive or negative value corresponds to outward or inward polarity, respectively. For example, for a network in which all filaments have an orientation of $\theta = 0$ or π , all filaments would be radially oriented with their polarities defined by whether the (+) or (-) end is closest to the cell membrane. By contrast, a network could have outwardly polarized filaments with random orientations, which would correspond to values of θ uniformly distributed between $-\pi/2$ and $+\pi/2$. Motor-cargo complexes bind a filament with a specified on rate k_{on} , walk ballistically in the direction determined by the filament polarity, and unbind from the filament with an off rate k_{off} or fall off the end of the filament. The cargo then returns to the cytoplasm and diffuses with constant D. In vivo, many individual molecular motors such as kinesin, myosin V, and dynein have processivities of ~1 to 2 μ m during transport (8,18,21-23); we start by assuming a mean processivity of 1 μ m.

We first focused on randomly oriented and positioned filament networks (see Materials and Methods; Fig. 2 A), which can approximate disordered actin-filament or micro-tubule networks as observed in vivo (5,24). We assumed a



FIGURE 2 MFPT is conserved with respect to total filament mass, but MFPT variability increases with fewer filaments. (*A*) Example of an explicit-filament representation of a cytoskeletal network for simulations in which cargo-motor complexes diffuse through the cytoplasmic cell bulk or move ballistically along transport filaments. Complexes are assumed to originate at the nuclear surface and transport terminates at the cell membrane. Red and green circles represent the (+) and (-) ends of randomly oriented filaments, respectively. The white circle indicates a 0.2- μ m buffer between the nucleus and the filaments. (*B*) MFPTs in explicit networks with a range of filament lengths and numbers; $v = 1 \mu m/s$, $D = 0.051 \mu m^2/s$, $k_{on} = 5/s$, and $k_{off} = 1/s$. MFPT is approximately constant along the black lines, which represent conserved network mass. (*C*) The standard deviation in MFPT over network realizations, normalized by MFPT, typically increases for smaller numbers of longer filaments. To see this figure in color, go online.

nuclear radius of 4.8 μ m, which is approximately consistent with the observed ratio between nuclear and membrane size (25,26). The filaments were located in the outer 5 μ m of a two-dimensional cell 10 μ m in radius. The 0.2- μ m buffer between the nuclear membrane and the region occupied by filaments allowed cargos to reach the ends of filaments near the nuclear membrane without interference from the nuclear surface, which was treated as a reflecting boundary condition. Up to 500 filaments were simulated, which is equivalent to ~10-fold the microtubule density in monkey kidney epithelia cells (27). The average microtubule length in human foreskin fibroblast cells is 5.45 μ m (28), which is roughly comparable with our maximum simulated filament length of 5 μ m. Similarly, the range of parameters simulated for filament number and filament length are representative of actin filaments in the cytoplasmic bulk, which have lengths of ~1 to 3 μ m (29,30). The density of actin filaments in flattened BSC-1 epithelial cells can be estimated via STORM imaging to be ~3 to 10 μ m of filament per μ m² (31), which is roughly comparable to 140 to 470 5- μ m filaments present in our simulation geometry.

In our model, a motor-cargo complex was represented by a circle with radius 100 nm, which is roughly consistent with the typical size range of a molecular motor (30 to 50 nm (32)) bound to a vesicle (20 to 600 nm (33)). Given that cells are ≥ 200 nm in height (34), we assumed that these particles experience no excluded-volume interactions with the network filaments, enabling our model to more effectively represent the three-dimensional nature of living cells. For simplicity, there were no interactions between cargo-motor complexes in our simulations. When a cargo spatially overlapped with a filament, binding within a time step dt occurred with probability $k_{on}dt$. Binding was followed by ballistic movement at a constant velocity in the direction indicated by the filament's polarity, until the cargo either dissociated from its track or reached the end of the filament.

We first determined the MFPT of cargo-motor complexes moving on random networks as a function of the primary network-topology parameters of filament length and number (Fig. 2 B). For each combination of filament length and number, we simulated 3,000 cargos on each of 10,000 network realizations. We used physiologically relevant parameters motivated by in vitro experiments on kinesin/ myosin (21) (see Materials and Methods), with motor velocity $v = 1 \ \mu \text{m/s}$, bulk diffusion $D = 0.051 \ \mu \text{m}^2/\text{s}$, $k_{on} = 5/\text{s}$, and $k_{\text{off}} = 1/s$. The MFPT averaged over network realizations and cargos was approximately constant over a wide range of filament lengths and numbers in networks with fixed total filament mass (Fig. 2 B). However, the standard deviation of MFPT over network realizations was high relative to the mean along curves of constant mass in networks with fewer filaments (Fig. 2 C). Thus, specific topological features of a random transport network can dramatically affect transport rates such that architectures with a higher number of shorter filaments have lower variation in MFPT.

Network-transport variability results from rare cargo traps

Over many network realizations the distribution of MFPTs was much broader for networks with fewer filaments than for networks with many filaments (Figs. 2 C and 3 A). To determine the origin of the large standard deviation in



FIGURE 3 High MFPTs result from traps near the nucleus. (A) Histograms of average MFPTs for 3,000 cargos for each of 500,000 network realizations consisting of random networks of 150 filaments with length of 3.5 µm (pink) and 500 filaments with length of 4.5 µm (blue). (B) Cargo residence times reveal traps near the nuclear surface for a network with a relatively large MFPT (220 s) relative to the mean over all networks (82 s) from the pink distribution in (A). (C) Cargo residence times of the network in (B) with the polarity of all filaments reversed: this reversal vielded a below-average MFPT of 62 s. (D) Total network trap area is strongly connected with MFPT across networks. (E) Reversing the polarity of all filaments in a network decreases the MFPT of slow networks. (F) Polarity reversal for 100% of the filaments that previously faced inward dramatically decreases MFPT. (G) The MFPT of a network typically increases substantially after polarity reversal for 25% of the filaments that previously faced outward. All simulations in (D)-(G) were carried out with 3,000 cargos on 1,000 networks of 150 3.5 μ m filaments. Red lines in (E)-(G) indicate the line y = x. To see this figure in color, go online.

MFPT in networks with relatively few longer filaments, we visually inspected networks that had particularly large MFPTs. We spatially binned the residence time averaged over cargos across the cytoplasm and noticed that for a network with large MFPT (220 s) relative to the average MFPT (82 s), there was a hotspot of >10-fold higher residence times near the nucleus in a region with multiple filaments pointed inward that acted as a sink for cargo (Fig. 3 B). To verify that this architecture was directly responsible for the large range of MFPTs, we reversed the polarity of each filament in the network and observed a drop in the MFPT to 62 s, without any residence-time hotspots (Fig. 3) C). We binned space into 0.04 μ m² regions and defined traps using a residence-time threshold of 10 s/ μ m². This choice of residence time threshold guaranteed that there were no spurious traps in the absence of filaments over a wide range of spatial bin sizes (Fig. S7). We scanned 1,000 network realizations and found that the total area of trap regions was strongly related to the MFPT of a network (Fig. 3 D), further supporting the hypothesis that traps are a major underlying cause of increases in MFPT for a given number of filaments of fixed length. Because a reasonable fraction (79%) of network realizations do not have traps, reversing filament polarity within a given network architecture generally removed traps from networks with large trap area, reducing MFPT (Fig. 3 E).

With a $0.2-\mu m$ buffer surrounding the nucleus, traps generally occurred near the nucleus, with 86% of the total trap area in traps with a centroid within 1 μ m of the nuclear boundary (Fig. S2A). We sought to understand the origin of this localization by increasing the size of the buffer region around the nucleus in our simulations from 0.2 to 1.0 μ m, which we accomplished by shrinking the nuclear radius by 0.8 μ m. To make MFPTs comparable across simulations with small and large buffers, we ignored cargo-residence times in the extended buffer region. In simulations with the larger buffer, both overall trap number and MFPT variability over network realizations decreased strongly (Fig. S2, B and C), whereas the concentration of traps near the nucleus declined (Fig. S2 A). Thus, the barrier formed by the nuclear membrane is an important factor in trap generation.

Polarity, not filament orientation, dominates transport times

Because reversing the polarity of all filaments in a network with traps was sufficient to cause an overall decrease in MFPTs (Fig. 3 E), we hypothesized that filament polarity could generally be utilized to regulate transport dynamics. We therefore fixed the orientations and positions of all filaments in a random network and reoriented all filament polarities such that they pointed outward (away from the nucleus). This global polarization resulted in an average fivefold reduction in MFPT (Fig. 3 F). In contrast, polarizing only 25% of the filaments in a random network such that they pointed inward (toward the nucleus) approximately doubled the average MFPT (Fig. 3 G). Interestingly, average MFPT was not affected when we reoriented filament directions from a random network to one in which all filaments were radially aligned, keeping the polarities and center of masses of the filaments fixed (Fig. S3).

Enhanced filament binding by multiple motors can have negative effects on transport efficiency

Transport via multiple motors bound to a single cargo is common in vivo (35,36) and has been the subject of numerous in vitro studies (37,38). Previous studies (39,40)modeled the effects of adding motors to cargo-motor complexes by increasing the on rate and decreasing the off rate relative to a single-motor scenario, increasing the time spent on the network during any binding event. To study how binding and unbinding rates affect MFPT statistics, we examined a wide range of values for random networks with 150 filaments of 3 μ m in length, a set of network parameters that yields an intermediate MFPT (Fig. 2 B). For high on and off rates, increases in either rate resulted in increases in MFPT (Fig. 4 A), although MFPT also increased for small k_{on} or k_{off} below ~2/s (Fig. 4 A), indicating that there is an optimal time spent on the network and processivity along filaments to minimize MFPT. MFPT variability over networks also increased substantially with increasing k_{on} , but decreased somewhat with increasing k_{off} (Fig. 4 B). For a collection of four motors bound to a cargo, each with a typical filament binding on rate of 5/s and an off rate of 0.005/step, a previous study, which simplistically assumed that the motors act completely independent of each other, predicted an effective collective motor-cargo on rate of 20/s and an off rate of 0.0001/step (39). Given these values, our simulations (Fig. 4) predict that the action of multiple motors could result in an average MFPT that is twice as long as the average MFPT in a scenario with just one motor, with a large increase in the variability of MFPTs across network realizations. More realistic models of multiple motor transport that address the effective interactions between processive motor



FIGURE 4 MFPT and variability over networks are minimized for intermediate on rate. (*A*) MFPT for random networks of 150 filaments with length of 3 μ m, measured for 3,000 motors on 60,000 network realizations for each combination of k_{on} and k_{off} , has a minimum at a motor on rate of k_{on} ~5/s. A motor step length of 8 nm, corresponding to the distance between subunits on a microtubule filament, was assumed; larger step length magnitudes merely result in a rescaling of the motor off rates in per step terms. (*B*) Standard deviation of MFPTs across network realizations normalized by average MFPT decreases with decreasing on rate and increasing off rate. To see this figure in color, go online.

proteins also predict the optimality of single-motor transport based primarily on geometric and kinetic constraints (41,42). Nonetheless, these models still predict an increase in the effective on rate and a decrease in the off rate for multiple motor cargos similar to the independent motor model studied, in which scenario our simulations show that the lowest MFPT for cargo transport is achieved by single-processive motors given their optimal network residence time (Fig. 4 *A*).

DISCUSSION

Given the importance of transport for many fundamental cellular processes, minimizing both the mean and variation in transport time from the nucleus to the cell membrane may result in increased cellular fitness. Our initial continuum model simulations (Fig. 1) suggest that the time of transport from the nucleus to the cell surface is minimized when transport systems with a constrained volume are localized adjacent to the nuclear surface in cells with nuclear radius greater than ~35% of the cell radius. In vivo, the ratio between the nuclear and cellular dimensions in most eukaryotic cells has been empirically observed to be 0.4 to 0.5 (25,26). The physical mechanism underlying the minimum in transport time is attributable to the prediction from the diffusion equation for a circular geometry that cargo residence time because of transport via only diffusion through a cytoplasmic bulk is greatest at ~35% of the cell radius. Because the nuclear surface generally lies in or near this region of the cell, placing a transport system closer to the cell nucleus results in speeding up transport in the region where the cargo would otherwise spend more time, thus resulting in the greatest reduction in MFPT. Although many cellular,

motor, and cytoskeletal network properties may affect transport efficiency, our simulations reveal that MFPT variability is a key quantity that is generally sensitive to specific details of the network architecture (Figs. 2 C and 3). High MFPTs relative to the average result from small subsets of filaments that form traps near the nucleus, the occurrence of which can be minimized in systems with a larger number of shorter filaments (Fig. 2 C) and in systems with a buffer region between the nucleus and the transport network (Fig. S2). Our results suggest that in situations in which highly variable transport times can be detrimental to cellular function, eukaryotic cells may have evolved mechanisms to protect cargos against becoming trapped in the nucleus; these mechanisms could include establishing a buffer region around the nuclear region that lacks polymerized, random, cytoskeletal filament networks (Fig. S2). Relatively lower actin network densities near the nucleus have generally been reported, especially in the context of crawling cells (43,44) as well as during nuclear positioning (45,46). Although in these cases different functional roles or causes for actin depletion have been postulated, our results suggest that an additional benefit of this arrangement is an avoidance of trapping in transport. Additionally, cellular filaments are likely to be dynamic; our transport results, which were obtained for fixed networks, can be understood as instantaneous characterizations of dynamic networks. Network dynamics and intracellular transport could be coupled in vivo (47), potentially allowing for feedbackregulated searches of the network parameter space resulting in beneficial outcomes such as MFPT reduction and responsiveness to changes in the external environment.

Because transport behavior is more strongly affected by filament polarity than by filament orientation (Figs. 3 and S3), adding a few outwardly polarized filaments dramatically affected transport times (Fig. S4), suggesting a potential benefit of the microtubule nucleation at the nucleus that occurs in many eukaryotic cells. With regards to future experimental research with transport networks, these results can provide a framework for the measurement and exploration of in vivo networks properties that act to functionally reduce MFPT and MFPT variability.

In previous models (2,5,12,15) that ignored the explicit spatial architecture of the network, the durations of each alternating phase of diffusive and ballistic motion during cargo transport were typically assumed to be exponentially distributed. However, in our simulations with explicit networks, the distribution of ballistic phases was complex and often strongly divergent from exponential (Fig. S5). Previous virtual network models could not yield information on variations in MFPT over network realizations, as the networks themselves were not explicitly modeled. Differences between our explicit physical modeling and previous virtual filament models can be substantial because of cargo rebinding and trapping effects, which can introduce errors in measuring MFPT of up to ~60% (Fig. S6).

Context is important when considering whether strong binding between the motor-cargo complex and the filament track is beneficial to the cell. For random networks, our simulations revealed that higher on rates and lower off rates than those typical for individual kinesin/myosin motors increase MFPT (Fig. 4 A), suggesting that binding of cargos to multiple motors, which has been observed in vivo (35,36), is not necessarily beneficial for transport. In these cases, periods of bulk diffusion enable cargos to disengage from regions of the transport network that are nonoptimally oriented, such as traps near the nucleus (Fig. 3), and to explore neighboring regions of the cytoplasm. This result is consistent with recent multiple-motor models and experiments, which predict that processive motors are unlikely to collaborate, predominantly because of geometric and kinetic constraints (41,42). However, along outwardly polarized filaments, stronger binding is likely to always decrease MFPT given that filaments point directly toward the target of transport.

Recently, experiments in vivo and in vitro have demonstrated the existence of anomalous features in intracellular transport (48-50), observing cargo motion as combinations of continuous time random walk (CTRW) and fractional Brownian-Langevin motion (fBM). Some of these experiments observed intracellular transport as primarily ergodic without aging (49,51), with active network transport fundamentally modeled as an fBM process. Statistically, we find cargo motion in our simulations to be consistent with fBM, which is ergodic (50,52). Firstly, the time-averaged mean squared displacement (MSD) for cargo transport as a function of lag time showed superdiffusive motion with an exponent $\alpha = 1.44$ (where MSD ~ t^{α}), which is consistent with the exponent measured with microspheres in cell extracts (49). Secondly, the ensemble-averaged MSD for cargo transport as a function of time shows convergence after ~100 s, implying that the system obeys ergodic behavior without aging (48). We do not observe a contribution from a CTRW-like process, which would be nonergodic, wherein the ensemble-averaged MSD continues to decrease with time (48). Experiments that show aging and nonergodicity are performed in vivo or with crowded cytosol extract, which in of itself can induce nonergodic motion (48,49), whereas our simulations are the result of a simple coupling of pure bulk diffusion and ballistic motion on fixed explicit filaments.

Overall, our results suggest a diverse range of mechanisms by which molecular transport over filaments can be tuned and regulated. We hypothesize that, in addition to reducing MFPT, reducing the variability in MFPT across network realizations promotes transport robustness over time. For example, having low variance in MFPT between network realizations among individual cells in a population, even at the expense of having a higher MFPT, is likely to promote fitness at the population level by virtue of having many fewer instances of cells with particularly poor network transport. Although we observed that MFPT was more sensitive to filament polarity (Fig. 3, E–G) than to filament orientation (Fig. S4), using transport constraints to design the polarity of a network may be challenging because a single network is used to transport cargos in multiple directions (9) and because cytoskeletal filaments also have other important structural roles (53). For biomimetic and other applications, however, we have demonstrated that several network properties, such as localization, filament length and number, and on and off rates, can be readily and rationally tuned.

SUPPORTING MATERIAL

Supporting Materials and Methods and eight figures are available at http://www.biophysj.org/biophysj/supplemental/S0006-3495(15)00869-3.

AUTHOR CONTRIBUTIONS

A.G., K.C.H., D.A., and N.C. designed the research; D.A. and N.C. performed the research; A.G., K.C.H., D.A., and N.C. analyzed the data; and A.G., K.C.H., and D.A. wrote the article.

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