UCLA

UCLA Previously Published Works

Title

Genetic risk models: Influence of model size on risk estimates and precision.

Permalink

<https://escholarship.org/uc/item/1d64z98k>

Journal

Genetic epidemiology, 41(4)

ISSN

0741-0395

Authors

Shan, Ying
Tromp, Gerard
Kuivaniemi, Helena
et al.

Publication Date

2017-05-01

DOI

10.1002/gepi.22035

Peer reviewed

Genetic risk models: Influence of model size on risk estimates and precision

Ying Shan¹ | Gerard Tromp^{2,3} | Helena Kuivaniemi^{2,3} | Diane T. Smelser² |
Shefali S. Verma⁴ | Marylyn D. Ritchie⁴ | James R. Elmore⁵ | David J. Carey² |
Yvette P. Conley^{6,7} | Michael B. Gorin^{8,9} | Daniel E. Weeks^{1,7}

¹Department of Biostatistics, Graduate School of Public Health, University of Pittsburgh, Pittsburgh, Pennsylvania, United States of America

²The Sigfried and Janet Weis Center for Research, Geisinger Health System, Danville, Pennsylvania, United States of America

³Division of Molecular Biology and Human Genetics, Department of Biomedical Sciences, Faculty of Medicine and Health Sciences, Stellenbosch University, Tygerberg, South Africa

⁴Department of Biomedical and Translational Informatics, Geisinger Health System, Danville, Pennsylvania, United States of America

⁵Department of Vascular and Endovascular Surgery, Geisinger Health System, Danville, PA

⁶Department of Health Promotion and Development, School of Nursing, University of Pittsburgh, Pittsburgh, Pennsylvania, United States of America

⁷Department of Human Genetics, Graduate School of Public Health, University of Pittsburgh, Pittsburgh, Pennsylvania, United States of America

⁸Departments of Ophthalmology and Human Genetics, David Geffen School of Medicine, University of California Los Angeles, Los Angeles, California, United States of America

⁹Stein Eye Institute, Los Angeles, California, United States of America

Correspondence

Daniel E. Weeks, Departments of Human Genetics and Biostatistics, Graduate School of Public Health, University of Pittsburgh, 130 De Soto Street, Pittsburgh, PA 15261, USA.
Email: weeks@pitt.edu

Grant sponsor: The Commonwealth Universal Research Enhancement (CURE) program of the Commonwealth of Pennsylvania; Grant number: 1120101; Grant sponsor: National Institutes of Health; Grant number: R01 EY009859; Grant sponsor: National Institutes of Health American Recovery and Reinvestment Act; Grant number: R01 EY009859-14S1.

ABSTRACT

Disease risk estimation plays an important role in disease prevention. Many studies have found that the ability to predict risk improves as the number of risk single-nucleotide polymorphisms (SNPs) in the risk model increases. However, the width of the confidence interval of the risk estimate is often not considered in the evaluation of the risk model. Here, we explore how the risk and the confidence interval width change as more SNPs are added to the model in the order of decreasing effect size, using both simulated data and real data from studies of abdominal aortic aneurysms and age-related macular degeneration. Our results show that confidence interval width is positively correlated with model size and the majority of the bigger models have wider confidence interval widths than smaller models. Once the model size is bigger than a certain level, the risk does not shift markedly, as 100% of the risk estimates of the one-SNP-bigger models lie inside the confidence interval of the one-SNP-smaller models. We also created a confidence interval-augmented reclassification table. It shows that both more effective SNPs with larger odds ratios and less effective SNPs with smaller odds ratios contribute to the correct decision of whom to screen. The best screening strategy is selected and evaluated by the net benefit quantity and the reclassification rate. We suggest that individuals whose upper bound of their risk confidence interval is above the screening threshold, which corresponds to the population prevalence of the disease, should be screened.

KEYWORDS

disease risk estimation, confidence interval, model size, reclassification

1 | INTRODUCTION

Personalized genomics is currently a widely discussed topic (Bloss, Darst, Topol, & Schork, 2011). Personalized genomics companies and many publications (Evans,

Visscher, & Wray, 2009; Morrison et al., 2007; Wray, Goddard, & Visscher, 2007) have provided disease risk prediction models based on genetic predictors. However, these risk reports seldom take the confidence interval of the risk estimate into account (Kalf et al., 2014). For example,

23andMe presented to its customers a point estimate of the risk and the average risk of the disease in the population, as well as how much higher the estimated risk was than the average risk. 23andMe did not present confidence intervals of the provided risk estimates. And in many publications, especially when risk estimates are based on odds ratios derived by meta-analysis, the confidence intervals of the risk estimates are not presented nor considered in the evaluation of the risk model. Many studies have applied regression models to a set of risk single-nucleotide polymorphisms (SNPs) to make predictions. Using the area under the curve (AUC) metric to evaluate their risk models, they conclude that the more risk SNPs in the risk model, the larger the AUC will be, thus, the better the ability to predict the risk (De Jager et al., 2009; van Dieren et al., 2012). However, as we illustrate here, as the number of risk SNPs in the model increases, the confidence interval of the risk estimate can widen. In fact, a risk estimate with a larger confidence interval from a larger model with more SNPs may not be practically better than a similar risk estimate with a smaller confidence interval from a smaller model based on fewer SNPs. When presenting and evaluating risk estimates, it is important to consider the level of uncertainty in the risk estimate.

In this study, we explore the changes of risk estimates and their 95% confidence interval widths as more SNPs, in the order of decreasing effect size, are added into the model, based on both simulated and real data. We also created a reclassification table to evaluate the effect of the added SNPs predictors, taking the confidence interval of the risk estimate into account. Finally, we selected the best screening strategy based on the net benefit quantity and the reclassification rate.

2 | METHODS

2.1 | Data description

In this study, we use three data sets to evaluate and compare our risk models. The first data set is a simulated one. We simulated a data set of 100,000 people assuming a genetic model based on 19 independent risk SNPs with odds ratios and allele frequencies matching those observed in a large meta-analysis of age-related macular degeneration (AMD; Fritsche et al., 2013), using the Multiple Gene Risk Prediction Performance (mgrp) R package (Pepe, Gu, & Morris, 2010). In the large meta-analysis of AMD, the 19 SNPs were shown to be highly associated with AMD. AMD is a progressive neurodegenerative disease that constitutes one of the primary causes of visual impairment and irreversible blindness in the elderly of western countries (Klein et al., 2011). In our simulation, we assumed that the disease is dichotomous with a prevalence of 0.055, which is similar to the prevalence of AMD.

The second data set is from a study of abdominal aortic aneurysms (AAA). AAA is the most common form of

aortic aneurysm. In general, the prevalence of AAA (2.9–4.9 cm in diameter) ranges from 1.3% for men aged 45–54 to up to 12.5% for men aged 75–84. Comparable prevalence figures for women are 0% and 5.2%, respectively (Rooke et al., 2012). Up to 10% of the male population who are more than 65 years old has AAA, and 80–90% of ruptures lead to sudden death (Assar, & Zarins, 2009). Our goal was to classify the population into high- and low-risk categories, where “high risk” is defined as having a risk higher than the population prevalence. Our motivation was to identify people with high AAA risk for targeted ultrasound screening. The samples were genotyped at 731K SNPs using the Illumina OmniExpress platform (dbGaP Study Accession numbers: phs000381.v1.p1, phs000408.v1.p1, and phs000387.v1.p1). AAA cases and controls were identified by electronic phenotyping (Borthwick et al., 2015). After imputation and quality control (Verma et al., 2014), 2,626 samples (733 cases and 1,893 controls) were available. The imputed data are part of the eMERGE Network Imputed GWAS data for 41 phenotypes (the dbGaP eMERGE phase 1 and 2 merged data submission with accession number phs000888.v1.p1). By modeling in a much larger electronic medical record (EMR)-based clinical data set, seven easy-to-measure clinical predictors (age, smoking status, sex, systolic blood pressure, diastolic blood pressure, height, and weight) were chosen for use in our risk models (Smelser et al., 2014). Based on prior literature (Biros et al., 2011; Bown et al., 2011; Elmore et al., 2009; Galora et al., 2013; Giusti et al., 2008; Harrison et al., 2013; Helgadottir et al., 2012; Jones et al., 2008, 2013; Saracini, et al., 2012; Thompson, Drenos, Hafez, & Humphries, 2008), 15 SNPs present in the imputed data were selected with odds ratios in the literature ranging from 0.41 to 2.16 (Supplementary Table S1).

The third data set is from a study of the genetics of AMD (Weeks et al., 2000, 2004). In our analysis, for 1,015 unrelated individuals (882 cases and 133 controls), high quality genotypes were available at 14 of the 19 SNPs mentioned above, and these 14 were used as predictors in the AMD data analysis. The cases in our study were defined according to the diagnosis criteria of “Model C” in Weeks et al. (2004). Under Model C, cases are those who are definitely or probably affected with AMD or with a related maculopathy. Model C also included individuals with end-stage disease, in the absence of any other documentation of macular pathology. The controls had no AMD symptoms with an age at last eye examination ≥ 65 .

2.2 | Data analysis

First, for all the three data sets, we used logistic regression to fit the risk models. To avoid over fitting, we used four-fifths of the data as the training data set and the rest of the data as the testing data set. The training and testing data sets are not only used to recommend a single best prediction model, but are

also applied to explore the behavior of models with different sizes, when each model is built using an ordered set of risk SNPs. We did not include any covariates besides the SNPs when analyzing the AMD and simulated data sets (because the simulated data are set up to be similar to the AMD data, this makes the results from these more comparable). When analyzing the AAA data set, we included seven easy-to-measure clinical predictors (age, smoking status, sex, systolic blood pressure, diastolic blood pressure, height, and weight). We let M be the total number of risk SNPs. Using the training data set, we fit the largest model using logistic regression with all of the M SNPs to estimate an odds ratio for each SNP. We ordered the M SNPs in decreasing order of effect size using odds ratios as an estimate of effect size after inverting any odds ratios < 1 . So, SNP 1 has the largest estimated effect size, SNP 2 has the next largest estimated effect size, etc. After ordering the SNPs in this manner, starting by fitting a model of size 1 using SNP 1, we then fit successively larger models using the training data set, increasing the model size K by adding in the next SNP from our ordered list (Supplementary Tables S1 and S2). For each model, all of the effect sizes were reestimated—these are the natural logarithm of odds ratios: β parameters. Then, we estimated risks for each individual in the testing data set by plugging in the β 's as estimated from the training set using K SNPs. When estimating risks from a case/control sample using logistic regression, the resulting risk estimate is not the absolute risk, but rather depends on the case/control ratio in the sample itself. Accordingly, for the case/control data sets, risk estimates were adjusted using the methods described in Pyke et al. (1979). For each person in the testing data set, we recorded the risk estimate, its 95% confidence interval, the model size, and the SNP genotypes.

We then explored how the risk shifts as the model size increases using bean plots and risk trajectory plots. To quantify the magnitudes of the risk shifts, we recorded the maximum of the absolute risk shifts (MRS) between model k and all bigger models for each individual. We recorded the maximum, across all individuals, of the MRS when additional SNPs were added to the model k that we refer to as the “maxMRS”; and the 95th percentile of the MRS that we refer to as the “95PMRS.” To investigate the relationship of the confidence interval width and the model size, we used Spearman's rank correlation test and bean plots.

For the AAA and AMD data sets, we evaluated the risk models using reclassification tables, taking the confidence interval into account, classifying individuals into high- and low-risk groups based on a threshold T corresponding to the population prevalence (we assumed the prevalence was 0.033 for AAA and 0.055 for AMD). In the traditional reclassification tables (which do not take the confidence intervals into account), assignment to either the low- or high-risk classes is defined solely based on the chosen risk threshold T . In order to take the risk confidence intervals into account, we created confidence interval augmented (CI-augmented) reclassifica-

FIGURE 1 Illustration of the risk estimates falling in the four LOW*, $\{-T\}$, $\{+T\}$, and HIGH* risk classes, which are defined as a function of the risk estimate value (gray dot) and its confidence interval. The horizontal line indicates the threshold T

tion tables where we defined the LOW*/HIGH* risk classes to contain individuals whose risk estimates were lower/higher than T and whose confidence interval did not overlap T . Individuals in these two classes had risk estimates that were unambiguously either below or above T (Fig. 1). Then, we added two more classes, denoted as “ $\{-T\}$ ” and “ $\{+T\}$,” which contain individuals with risk estimates with confidence intervals that overlap the threshold T . The individuals in the $\{-T\}$ class had risk estimates $< T$, while those in the $\{+T\}$ class had risk estimates $\geq T$. For individuals in these two classes, it is not clear if their true risk is above or below T . As the CI-augmented approach classifies the individuals into four categories (LOW*, $\{-T\}$, $\{+T\}$, HIGH*), there are three possible screening strategies: (1) screen the individuals in HIGH* risk class only (defined as $\{T, 1\}$); (2) screen the individuals in both $\{+T\}$ class and HIGH* risk class (defined as $\{+T, 1\}$); (3) screen the individuals in $\{-T\}$, $\{+T\}$, and HIGH* risk class (defined as $\{-T, 1\}$). We calculated the net benefit (McGeechan, Macaskill, Irwig, & Bossuyt, 2014), which provides a measure of the number of people correctly screened as having the outcome, adjusted for the number of people incorrectly screened as having the outcome. The net benefit formula is:

$$\text{Net benefit} = \frac{\text{True positives}}{n} - \frac{\text{False positives}}{n} \left(\frac{T}{1-T} \right),$$

where n is the sample size, and T is the threshold as indicated above. Then, we calculated the reclassification rate of $[0, -T] \Leftrightarrow \{+T, 1\}$ and the reclassification rate of $\text{LOW}^* \Leftrightarrow \{-T, 1\}$ according to the screening strategies 2 and 3. The reclassification rate of lower risk group $\Leftrightarrow$ higher risk group means the proportion of individuals reclassified from the lower risk group to the higher risk group or from the higher risk group to the lower risk group. We also evaluated the rate of correct reclassifications for the three screening strategies. Correct reclassification means reclassifying cases from the lower risk group to the higher risk group, or reclassifying

FIGURE 2 Risk trajectories as categorized by the initial risk for (a) the simulation study, (b) the AAA data set, and (c) the AMD data set. Each part contains the trajectories of 30 individuals randomly chosen from the testing data set. The odds ratios of the added SNPs are shown on the top of each subfigure. The horizontal black line is the disease prevalence (0.055 for the simulated data set and the AMD data set; 0.033 for the AAA data set). The maxMRS_{mi} and 95PMRS_{mi} are based on the model with i SNPs, where MRS = max(absolute risk shifts between the current model and all bigger models for a given individual); maxMRS = max(MRS) across all individuals; and 95PMRS = the 95th percentile of the MRS

controls from the higher risk group to the lower risk group. We used the net benefit and the rates of correct reclassification to select the best screening strategy. Furthermore, in order to explore the influence of model size on the confidence interval width, we recorded how many confidence interval widths increased and decreased when more SNPs were added to the initial model.

3 | RESULTS

First, in each data set, we examined how much the risk shifted when one more SNP with the next largest odds ratio (after all odds ratios were inverted to be >1) was added to the model. To explore the risk shift at the individual level, we plotted representative risk trajectories as SNPs were added to the model in the order of decreasing effect sizes (Fig. 2). As expected, the risks shift less when SNPs with the smaller odds ratios are added. Figure 2 shows, at the individual level, movement in risk among the smaller models has a marked flattening of the risk trajectories as the models get larger. We also found that individuals with higher initial risks tend to have their risks shift more than those with lower initial risks as the model size increases. In the simulated data set (Fig. 2A), when the three initial risk are 0.027, 0.068, and 0.161, the maxMRS's based on the smallest model are 0.15, 0.27, and 0.39, and the 95PMRS's based on the smallest model are 0.05, 0.11, and 0.21, respectively. In AAA and AMD data sets, the 95PMRS's

based on the smallest model are also bigger when the initial risks are bigger (Fig. 2B and C).

We then explored the risk shift at the population level, as more SNPs are added into the risk model. Table 1 shows that the risks do not shift markedly once the model size is bigger than a certain level. For example, if we let the “maxMRS-selected model” be the smallest model with a maxMRS < 0.06 , then in all the three data sets, the 95PMRS of the models bigger than the maxMRS-selected model were all smaller than 0.025. Furthermore, if we let M_i represent the model with i SNPs, in all the three data sets, when the model size is bigger than the maxMRS-selected model, 100% of the M_{i+1} risk estimates lay inside the corresponding M_i confidence interval (Fig. 3A–C) and 100% of the M_{i+1} confidence intervals overlap with the corresponding M_i confidence interval (Fig. 3D–E). In addition, when the model size is bigger than the maxMRS-selected model, all the M_{i+1} confidence intervals overlapped more than 50%, 90%, and 95% with the corresponding M_i confidence intervals, in the simulation data set, AAA data set, and AMD data set, respectively (data not shown). Consistent with these observations, Figure 4 shows that when the model size was greater than the maxMRS-selected model, the risk shift distributions did not change markedly as the model sizes grew.

We then explored the influence of the model size on the confidence interval width. Figure 5 shows that the confidence interval width was positively correlated with model size in all the three data sets. For all the three data sets, the

TABLE 1 The maxMRS and 95PMRS measures^a in the simulation data set, the AAA data set and the AMD data set

# of SNPs in Model	Simulation data set		AAA data set		AMD data set	
	maxMRS	95PMRS	maxMRS	95PMRS	maxMRS	95PMRS
0	-	-	0.221	0.075	-	-
1	0.395	0.113	0.222	0.070	0.381	0.227
2	0.311	0.076	0.182	0.063	0.322	0.158
3	0.286	0.067	0.133	0.051	0.307	0.136
4	0.214	0.060	0.147	0.041	0.297	0.128
5	0.209	0.053	0.098	0.038	0.222	0.104
6	0.185	0.050	0.070	0.027	0.164	0.079
7	0.176	0.046	0.058	0.022	0.142	0.066
8	0.163	0.041	0.042	0.016	0.096	0.050
9	0.150	0.036	0.039	0.014	0.064	0.035
10	0.119	0.032	0.030	0.011	0.032	0.021
11	0.103	0.029	0.021	0.009	0.022	0.010
12	0.099	0.025	0.020	0.006	0.007	0.004
13	0.087	0.022	0.017	0.005	0.002	0.001
14	0.069	0.019	0.002	0.001		
15	0.060	0.016				
16	0.043	0.012				
17	0.025	0.007				
18	0.011	0.003				

The bold values indicate the “maxMRS-selected model” which is the smallest model with maxMRS less than 0.06.

^aMRS = max(absolute risk shifts between the current model and all bigger models for a given individual); maxMRS = max(MRS) across all individuals; 95PMRS = the 95th percentile of the MRS.

Spearman's rank correlation test gives *P* values smaller than 0.001, indicating positive correlation between the confidence interval width and model size. Table 2 also shows the influence of the model size on the confidence interval width. More estimates have wider confidence intervals in the updated model than in the initial model. For the AAA data set, comparing M_0 with M_7 , 84.8% of the estimates have wider confidence intervals in the updated model compared to the initial model; while comparing the M_7 with M_{15} , 96.0% of the estimates have wider confidence intervals in the updated model compared to the initial model. For the AMD data set, comparing M_1 to M_{10} , 90.0% of the estimates have wider confidence intervals in the updated model compared to the initial model; while comparing M_{10} to M_{14} , 100% of the estimates have wider confidence intervals in the updated model compared to the initial model.

Furthermore, we determined the reclassification rates based on the screening strategies 2 and 3. The reclassification rates with bigger-effect SNPs in Table 2a and c are higher than that with smaller-effect SNPs in Table 2b and d. But the small-effect SNPs can still affect the reclassifications. Table 2b shows that in the AAA data set, adding eight less effective SNPs to the maxMRS-selected model, 19.0% of cases and 0.6% controls were correctly reclassified; while 0% of cases and 3.5% of controls were mistakenly reclassified. Table 2d shows that in the AMD data set, adding four less effective

SNPs to the maxMRS-selected model, 21.1% of the cases and 0% of the controls were correctly reclassified; while 0% of the cases and 13.0% of the controls were mistakenly reclassified. We also found the correctly reclassified rate of $LOW^* \Leftrightarrow \{-T, 1\}$ is much higher than $[0, -T] \Leftrightarrow \{+T, 1\}$ for cases, and the correctly reclassified rate of $LOW^* \Leftrightarrow \{-T, 1\}$ is lower than $[0, -T] \Leftrightarrow \{+T, 1\}$ for controls, in both AAA and AMD data sets.

Finally, we evaluated the net benefit quantities of the three screening strategies. Table 3 shows that in both of the two data sets, the screening strategy of screening the individuals in the $\{-T, 1\}$ category provides the biggest net benefit quantity among the three strategies. The full models of both AAA and AMD data sets with $\{-T, 1\}$ screening strategy have the biggest net benefit quantity.

4 | DISCUSSION

Due to rapid progress and advancements in sequencing technology, it is now feasible, yet still expensive, to accurately type all genetic variants for an individual. To construct a risk estimate from these variants, we could attempt to use all of them or we could order them by estimated effect size, and use only the strongest predictors. But then the question is how many of these should be used. Clearly, as the effect

FIGURE 3 Percentages of M_{i+1} risks inside the M_i confidence intervals (a–c), and percentages of M_{i+1} confidence intervals that overlap with the M_i confidence intervals (d–f), where model M_{i+1} is one SNP larger than model M_i . The gray bar shows the maxMRS-selected models

size shrinks, adding a single small effect predictor to the risk model will not shift the risk by much. We explored here how the risk estimate and its certainty change as variants of decreasing effect size are added into the risk model, using simulated data and real data of two different complex diseases (AAA and AMD).

If we order SNPs by decreasing effect sizes and build risk models of increasing size by adding in the next SNP, we first observe that the risk shifts between successive models become more and more modest (Figs. 2 and 4, Table 1) and the confidence intervals of the risk estimates tend to become larger (Fig. 5, Table 2). Then, we observe that when the model size is large enough, if one more variant is added, the majority of the updated risk estimates will lie within the confidence interval of the preceding estimate and the confidence intervals of the new and old estimates will overlap substantially (Fig. 3). However, as we add multiple small-effect SNPs to the model simultaneously, these SNPs can still affect the reclassifications (Tables 2 and 3).

Our data also suggest that models with slightly larger AUCs are not necessarily better than those with smaller AUCs, if one takes into consideration risk shifts and confi-

dence interval widths. Tables 1, 2b, and 2d show that when the model size is bigger than “MaxMRS-selected model,” the risks shifts become modest and the confidence intervals become wider. Thus, with similar risk estimates and wider confidence interval widths, full models are not necessarily superior to maxMRS-selected models. However, Table 4 shows that the AUCs of the full models are only slightly larger than the AUCs of the maxMRS-selected models. This suggests that only considering the AUC but ignoring risk shifts and confidence intervals may not be adequate.

We recommend that all individuals with risk estimates above the threshold T or who have risk estimates with confidence intervals that overlap T (e.g., those in the $\{-T, 1\}$ category) should be screened. There are two reasons for this. First, the strategy of screening the individuals in the $\{-T, 1\}$ category gives the biggest net benefit among all three screening strategies. Second, for the cases, the correctly reclassified rate of $LOW^* \Leftrightarrow \{-T, 1\}$ is much higher than $[0, -T] \Leftrightarrow \{+T, 1\}$, although for the controls, the correctly reclassified rate of $LOW^* \Leftrightarrow \{-T, 1\}$ is lower than $[0, -T] \Leftrightarrow \{+T, 1\}$, in both AAA and AMD data sets. Where screening costs much less compared to failing to detect the disease,

FIGURE 4 The distribution of risk shifts as a function of the number of SNPs in the updated model. The plots were generated by the beanplot command in the R package of the same name (Kampstra, 2008). The dark horizontal lines show individual observations, and the red line indicates the mean. The label above the plot is the added SNP's odds ratio in the model. The red circle indicates the risk shift distribution where the updated model is the maxMRS-selected model

screening the individuals in $\{-T, 1\}$ is the most appropriate strategy. However, it is important to remember that clinical cost-benefit analyses are complex and the assumption here is that screening is beneficial, although it is not necessarily so (for various diseases) if the “cost” of intervention risks are taken into account.

The results (Table 3) are based on setting the threshold (T) to the population disease prevalence. The purpose of setting T to the population disease prevalence is to recommend screen-

ing for anyone whose risk was higher than what it would be if they were sampled from the general population. However, for many diseases, people may not undergo screening unless their estimated risk is relatively high. So we reevaluated the net benefit, setting T to higher values of 10% and 20%. Supplementary Table S3 shows that when the thresholds are 10% and 20%, for both AAA and AMD data sets, the strategy of screening individuals in the $\{-T, 1\}$ category still provides the biggest net benefit quantity among the three strategies, and

FIGURE 5 The distribution of the confidence interval widths by model size. The confidence interval width axis uses the log scale. The label above the bean plot is the added SNP's odds ratio in the model. The horizontal line in the middle of each bean plot shows the mean value

the full models with $\{-T, 1\}$ screening strategy still have the biggest net benefit quantity.

In our study, all the results were generated by one single split with 80% individuals in the training data set and 20% individuals in the testing data set. We then generated five more 80/20 random splits of the training and testing data sets to illustrate the results change. Table 5 shows the maxMRS-selected models of each split. In the simulation data set, the maxMRS-selected models in the five testing data sets are similar; while in the AAA and AMD data sets, the maxMRS-selected models in the five testing data sets are variant. This is because the sample size in the simulation data set is large (100,000), while the sample sizes in the AAA and AMD data

sets are small (2,626 and 1,015, respectively). Therefore, the max-MRS selected models should be built using data sets with large sample sizes. Otherwise, the max-MRS selected models may be greatly affected by the splitting of the training and testing data sets. When the sample size is small and the maxMRS-selected model sizes are variant, we would recommend using the median value of the maxMRS-selected model sizes as the final model.

In our results, the relationship of the risks and the confidence interval widths is consistent with the binomial distribution property that the confidence interval width increases as the risk estimate rises to 0.50 and decreases as the risk estimate increases beyond 0.5. Because the disease prevalence

TABLE 2 CI-Augmented reclassification tables for the AAA data set and the AMD data set

(a) CI-Augmented Reclassification Table for the AAA Data Set, When the Initial Model Only Has the Clinical Predictors (Model 0) and the Updated Model Added Seven Most Effective SNPs (Model 7)										
Outcome: unaffected with AAA										
Initial model: clinical predictors	Updated model: clinical predictors plus seven SNPs								[0, −0.033] ⇔ {+0.033, 1} % Reclassified	LOW* ⇔ {−0.033, 1} % Reclassified
	LOW* [0,0.033]		{−0.033}		{+0.033}		HIGH* {0.033,1}			
	−=	+	−=	+	−=	+	−=	+		
LOW* [0,0.033]	99	323	0	19	0	1	0	0	2.1	4.5
{−0.033}	5	1	0	12	0	8	0	1		
{+0.033}	0	0	5	7	0	9	0	4	11.9	3.7
HIGH* {0.033,1}	0	0	0	4	3	10	1	92		
Outcome: affected with AAA										
Initial model: clinical predictors	Updated model: clinical predictors plus seven SNPs								[0, −0.033] ⇔ {+0.033, 1} % Reclassified	LOW* ⇔ {−0.033, 1} % Reclassified
	LOW* [0,0.033]		{−0.033}		{+0.033}		HIGH* {0.033,1}			
	−=	+	−=	+	−=	+	−=	+		
LOW* [0,0.033]	2	17	0	4	0	0	0	0	13.9	17.4
{−0.033}	0	2	1	5	0	5	0	0		
{+0.033}	0	0	1	4	0	8	0	3	4.3	1.0
HIGH* {0.033,1}	0	0	1	2	2	9	6	152		
(b) CI-Augmented Reclassification Table for the AAA Data Set, When the Initial Model Has Clinical Predictors Plus the Seven Most Effective SNPs (Model 7) and the Updated Model Has the Clinical Predictors Plus all the 15 SNPs (Model 15)										
Outcome: unaffected with AAA										
Initial model: clinical predictors plus seven SNPs	Updated model: clinical predictors plus 15 SNPs								[0, −0.033] ⇔ {+0.033, 1} % Reclassified	LOW* ⇔ {−0.033, 1} % Reclassified
	LOW* [0,0.033]		{−0.033}		{+0.033}		HIGH* {0.033,1}			
	−=	+	−=	+	−=	+	−=	+		
LOW* [0,0.033]	29	379	0	15	0	0	0	0	0.6	3.5
{−0.033}	0	1	3	40	0	3	0	0		
{+0.033}	0	0	0	4	0	26	0	0	3.1	0.6
HIGH* {0.033,1}	0	0	0	0	0	10	0	88		

(continues)

TABLE 2 (Continued)

(d) CI-Augmented Reclassification Table for the AMD Data Set, When the Initial Model Has 10 Most Effective SNPs (Model 10) and the Updated Model Has the Clinical Predictors Plus all the 14 SNPs (Model 14)												
Outcome: unaffected with AMD												
Initial Model: 10 SNP	Updated model: 14 SNPs											
	LOW* [0,0.055]			{-0.055}			{+0.055}			HIGH* {0.055,1}		
	--	+		--	+		--	+		--	+	
LOW* [0,0.055]	0	20		0	3		0	0		0	0	
{-0.055}	0	0		0	13		0	0		0	0	
{+0.055}	0	0		0	1		0	11		0	0	
HIGH* {0.055,1}	0	0		0	0		0	1		0	3	
Outcome: affected with AMD												
Initial Model: 10 SNP	Updated model: 14 SNPs											
	LOW* [0,0.055]			{-0.055}			{+0.055}			HIGH* {0.055,1}		
	--	+		--	+		--	+		--	+	
LOW* [0,0.055]	0	30		0	8		0	0		0	0	
{-0.055}	0	0		0	47		0	2		0	0	
{+0.055}	0	0		0	1		0	68		0	0	
HIGH- {0.055,1}	0	0		0	0		0	23		0	108	
											LOW* ⇔ {-0.055, 1} % Reclassified	
											13.0	
											0.0	
											6.2	
											LOW* ⇔ {-0.055, 1} % Reclassified	
											21.1	
											2.3	
											0.5	
											0.0	

“LOW*/“HIGH*” class records the number of samples with both risk estimates and the two confidence interval bounds lower/higher than the threshold, which is the prevalence of the corresponding disease. “{-threshold}” class records the number of samples with risk estimates lower than the threshold, but the higher confidence interval bounds above the threshold. “{+threshold}” class records the number of samples with risk estimates higher than the threshold, but the lower confidence interval bounds below the threshold. “% reclassified” is the percentage of samples that are reclassified from LOW*/HIGH* risk class to HIGH*/LOW* class. “-” means the confidence interval width in the updated model is narrower than or equal to the width in the initial model. “+” means the confidence interval width in the updated model is wider than the initial width.

TABLE 3 Net benefit of the classification of each model in the AAA data set and the AMD data set for the three screening strategies

Screening strategies	AAA			AMD		
	Model 0	Model 7	Model 15	Model 1	Model 10	Model 14
Screen individuals in $\{T,1\}$	0.201	0.191	0.180	0.601	0.386	0.318
Screen individuals in $\{+T,1\}$	0.220	0.218	0.216	0.601	0.587	0.590
Screen individuals in $\{-T,1\}$	0.236	0.238	0.242	0.601	0.730	0.753

The threshold T is the population disease prevalence.

TABLE 4 AUCs (Area Under the Curve) of the maxMRS-selected model and the full model in the simulation data set, AAA data set, and AMD data set

	Simulation	AAA	AMD
MaxMRS-selected model	0.758 (0.006)	0.871 (0.017)	0.741 (0.021)
Full model	0.760 (0.006)	0.873 (0.016)	0.742 (0.022)

The numbers in each cell are the mean value and the standard deviation (shown in the parenthesis) of the AUCs in the 10 replicates.

TABLE 5 The number of SNPs in the maxMRS-selected models of five times 80/20 random splits in simulation data set, AAA data set, and AMD data set

Cross-Validation	Simulation	AAA	AMD
1	16	7	10
2	16	7	11
3	17	8	12
4	17	12	—
5	17	13	—

The “—” symbol indicates that the maxMRS based on the full model is bigger than 0.06. The numbers of SNPs in the maxMRS-selected model of the five splits are sorted by an increasing order in each data set.

in the simulation study, AAA study, and AMD study were 0.055, 0.033, and 0.055, respectively, most of the risk estimates were much lower than 0.5 in all three data sets. In the simulation, AAA, and AMD data sets, only three, one, and eight individuals had risk estimates bigger than 0.5, respectively. In all the three data sets, most of the confidence intervals increased as the risk increased, or decreased as the risk decreased, when one more SNP with the next largest effect size was added to the model. But there were still some confidence intervals that increased as the risks decreased in the three data sets and some confidence intervals that decreased as the risks increased in the AAA data set only. These two scenarios are because of two reasons. The first one is that the confidence interval widths are not only related to the risk size, but are also related to the model size. Even though the risks estimated by larger models are smaller, the confidence intervals can still become bigger if the model sizes are bigger. The second reason is that when the risk estimate exceeds 0.50, the confidence interval width decreases as the risk increases, and vice versa.

The risk trajectory plot (Fig. 2) shows that the higher-initial-risk individuals have their risks shifted more than the

lower-initial-risk individuals as more SNPs are added to the model. This observation is mainly because of two reasons. First, the risk trajectories that start with a low initial risk suffer from a lower bound effect—they cannot move very far in the downward direction. Second, because the disease prevalence in the three data sets is as low as 0.055, 0.033, and 0.055, respectively, the majority of people must be in the low risk category.

Other previous studies classified individuals using both the risks and the confidence intervals. Goddard and Lewis (2010) developed a strategy, which has been implemented in the R package REGENT (Crouch, Goddard, & Lewis, 2013), to classify individuals into risk classes using the risk and the confidence interval of an average individual to anchor the classification. With N SNPs, there are 3^N genotypes. The “average individual” is the individual with a genotype relative risk closest to the average risk, which is the sum across all the 3^N genotypes of the products of their frequencies and relative risks of disease. An estimate with confidence interval overlapping the confidence interval of the “average individual” is classified as “Average” risk. An estimate with confidence interval below the confidence interval of the “average individual” is categorized as “low” risk. In a similar manner, they also define “moderate” and “high” risk categories. Scott et al. (2013) applied the reclassification method and the REGENT R package to predict the risk of rheumatoid arthritis and its age of onset with smoking. In Goddard and Lewis (2010), they observed that when one uses confidence interval-based risk classification, one can run into the situation where an individual with a lower risk is classified into the high risk group because their confidence interval was larger than an individual with a slightly higher risk who had a narrower confidence interval. This phenomenon also happens in our AAA and AMD data sets. We recorded the smallest risk estimate among those whose upper bounds of the confidence intervals are higher than the threshold. Then, we counted the number of estimates that are higher than this smallest risk estimate, but with confidence intervals that do not cross the threshold. Using the smallest model (model 0) and the biggest model (model 15) of the AAA data set, models 1, 11, and 14 of the AMD data set, there are 12, 24, 0, 19, and 9 estimates that meet these criteria, respectively.

Hart et al. (2013) also built a logistic regression model for risk estimation and took confidence intervals into account.

They used logistic regression to create a new actuarial risk assessment instrument (ARAI). They categorized the individuals to two groups based on the ARAI score. They evaluate the ARAI at both group and the individual levels. Their results at the individual level are similar to our results. The mean width of the 95% confidence intervals for individual risk estimates in the high risk score category was much bigger than that of subjects in the low risk category. Confidence intervals for individual risk estimates overlapped completely within groups, and almost completely across groups.

In our study, the numbers of SNPs used in the simulation data set, AAA data set, and AMD data set are small (19, 14, and 15 risk SNPs, respectively), based on using the subset of significantly associated risk SNPs. Of course, in genome-wide studies, it might be of interest to use more or all of the available SNPs. In such a case, statistical methods, such as penalized regression equipped with variable selection (Austin, Pan, & Shen, 2013) and Bayesian Alphabet methods (Gianola, 2013), can be applied to the SNP-selection; prediction is then based on the selected SNPs. Wimmer et al. (2013) compared methods performing variable selection to methods that retain all predictors in the model, for example, ridge regression best linear unbiased prediction (RR-BLUP). They concluded that when the sample size is much larger than the number of causal mutations contributing to the trait, SNP selection based prediction outperforms RR-BLUP. However, when the number of SNPs is big compared to sample size, each of small to modest effect size, such as in many complex disease scenarios, RR-BLUP is superior to the SNP selection based prediction. Thus, under the situation where the sample size is smaller than the number of causal mutations contributing to the traits, methods distributing effects across the genome would provide more precise predictions than those that perform model selection, and our proposed SNP-selection based prediction according to width of confidence intervals might be less than optimal. It would be of interest to extend this work to the context of penalized shrinkage models and traits with larger numbers of established risk SNPs.

Consideration of risk estimate uncertainty is important because if the disease risk estimates, as well as the confidence intervals are provided, people can make more informed decisions regarding their screening decisions (Weeks, & Ott, 1990). For example, suppose an individual has a risk estimate below the threshold, but the upper bound of the confidence interval is much higher than the threshold. If only the risk estimate is provided, there will be an unfounded confidence in the estimate and the individual may feel safe, and therefore may choose to not undergo screening. But if both the risk estimate and its confidence interval are provided, the individual may no longer feel safe, and probably will undergo screening. For another example, consider an individual with a risk estimate slightly higher than the threshold and the lower bound of the confidence interval also above the threshold. If only the risk estimate is provided,

this individual may not undergo screening, because the risk estimate is only slightly higher than the threshold. However, if the confidence interval shows that it has 95% certainty that the individual has high risk of getting the disease, then this individual may decide to undergo screening. On the other hand, because it is difficult to clearly convey risk estimates in such a way that they are understood and interpreted correctly, it may be even more difficult to clearly communicate the information embodied in the confidence intervals around those risk estimates (Lautenbach, Christensen, Sparks, & Green, 2013). Careful consideration of how to best communicate these measures of risk estimate uncertainty is merited, lest such communications lead to increased disease-related anxieties and poorer risk perceptions (Han, 2013; Han et al., 2011).

ACKNOWLEDGMENTS

This work was supported by a Collaborative Research Award (#1120101) on Translational Genomics, part of The Commonwealth Universal Research Enhancement (CURE) program of the Pennsylvania Department of Health, and Geisinger Health System. The AMD data were generated under support of the National Institutes of Health grants R01 EY009859 (to M.B.G.) and American Recovery and Reinvestment Act supplement R01 EY009859-14S1 (to M.B.G.) and additional funding from an unrestricted grant from Research to Prevent Blindness, N.Y., N.Y. and the Harold and Pauline Price Foundation (to M.B.G.).

CONFLICT OF INTEREST

D.E.W., Y.P.C., and M.B.G. are coinventors on licensed patents held by the University of Pittsburgh for the chromosome 10q26 PLEKHA1/ARMS2/HTRA1 loci for AMD.

REFERENCES

- Assar, A. N., & Zarins, C. K. (2009). Ruptured abdominal aortic aneurysm: A surgical emergency with many clinical presentations. *Postgraduate Medical Journal*, 85(1003), 268–273.
- Austin, E., Pan, W., & Shen, X. (2013). Penalized regression and risk prediction in genome-wide association studies. *Statistical Analysis and Data Mining*, 6(4), 315–328.
- Biros, E., Norman, P. E., Jones, G. T., van Rij, A. M., Yu, G., Moxon, J. V., ... Gollidge, J. (2011). Meta-analysis of the association between single nucleotide polymorphisms in TGF-beta receptor genes and abdominal aortic aneurysm. *Atherosclerosis*, 219(1), 218–223.
- Bloss, C. S., Darst, B. F., Topol, E. J., & Schork, N. J. (2011). Direct-to-consumer personalized genomic testing. *Human Molecular Genetics*, 20(R2), R132–R141.
- Borthwick, K. M., Smelser, D. T., Bock, J. A., Elmore, J. R., Ryer, E. J., Ye, Z., ... Kuivaniemi, H. (2015). ePhenotyping for abdominal aortic aneurysm in the electronic medical records and genomics (eMERGE) network: Algorithm development and konstanz information miner workflow. *International Journal of Biomedical Data Mining*, 4(1), 113.
- Bown, M. J., Jones, G. T., Harrison, S. C., Wright, B. J., Bumpstead, S., Baas, A. F., ... Samani, N. J. (2011). Abdominal aortic aneurysm is associated with

- a variant in low-density lipoprotein receptor-related protein 1. *American Journal of Human Genetics*, 89(5), 619–627.
- Crouch, D. J., Goddard, G. H., & Lewis, C. M. (2013). REGENT: A risk assessment and classification algorithm for genetic and environmental factors. *European Journal of Human Genetics*, 21(1), 109–111.
- De Jager, P. L., Chibnik, L. B., Cui, J., Reischl, J., Lehr, S., Simon, K. C., ... Karlson, E. W. (2009). Integration of genetic risk factors into a clinical algorithm for multiple sclerosis susceptibility: A weighted genetic risk score. *Lancet Neurology*, 8(12), 1111–1119.
- Elmore, J. R., Obmann, M. A., Kuivaniemi, H., Tromp, G., Gerhard, G. S., Franklin, D. P., ... Carey, D. J. (2009). Identification of a genetic variant associated with abdominal aortic aneurysms on chromosome 3p12.3 by genome wide association. *Journal of Vascular Surgery*, 49(6), 1525–1531.
- Evans, D. M., Visscher, P. M., & Wray, N. R. (2009). Harnessing the information contained within genome-wide association studies to improve individual prediction of complex disease risk. *Human Molecular Genetics*, 18(18), 3525–3531.
- Fritsche, L. G., Chen, W., Schu, M., Yaspan, B. L., Yu, Y., Thorleifsson, G., ... Abecasis, G. R. (2013). Seven new loci associated with age-related macular degeneration. *Nature Genetics*, 45(4), 433–439.
- Galora, S., Saracini, C., Palombella, A. M., Pratesi, G., Pulli, R., Pratesi, C., ... Giusti, B. (2013). Low-density lipoprotein receptor-related protein 5 gene polymorphisms and genetic susceptibility to abdominal aortic aneurysm. *Journal of Vascular Surgery*, 58(4), 1062–1068.e1.
- Gianola, D. (2013). Priors in whole-genome regression: The Bayesian alphabet returns. *Genetics*, 194(3), 573–596.
- Giusti, B., Saracini, C., Bolli, P., Magi, A., Sestini, I., Sticchi, E., ... Abbate, R. (2008). Genetic analysis of 56 polymorphisms in 17 genes involved in methionine metabolism in patients with abdominal aortic aneurysm. *Journal of Medical Genetics*, 45(11), 721–730.
- Goddard, G. H., & Lewis, C. M. (2010). Risk categorization for complex disorders according to genotype relative risk and precision in parameter estimates. *Genetic Epidemiology*, 34(6), 624–632.
- Han, P. K. (2013). Conceptual, methodological, and ethical problems in communicating uncertainty in clinical evidence. *Medical Care Research and Review*, 70(1 Suppl), 14S–36S.
- Han, P. K., Klein, W. M., Lehman, T., Killam, B., Massett, H., & Freedman, A. N. (2011). Communication of uncertainty regarding individualized cancer risk estimates: Effects and influential factors. *Medical Decision Making*, 31(2), 354–366.
- Harrison, S. C., Smith, A. J., Jones, G. T., Swerdlow, D. I., Rampuri, R., Bown, M. J., ... Humphries, S. E. (2013). Interleukin-6 receptor pathways in abdominal aortic aneurysm. *European Heart Journal*, 34(48), 3707–3716.
- Hart, S. D., & Cooke, D. J. (2013). Another look at the (Im-) precision of individual risk estimates made using actuarial risk assessment instruments. *Behavioral Sciences & the Law*, 31(1), 81–102.
- Helgadottir, A., Gretarsdottir, S., Thorleifsson, G., Holm, H., Patel, R. S., Gudnason, T., ... Stefansson, K. (2012). Apolipoprotein(a) genetic sequence variants associated with systemic atherosclerosis and coronary atherosclerotic burden but not with venous thromboembolism. *Journal of the American College of Cardiology* 60(8), 722–729.
- Jones, G. T., Bown, M. J., Gretarsdottir, S., Romaine, S. P., Helgadottir, A., Yu, G., ... van Rij, A. M. (2013). A sequence variant associated with sortilin-1 (SORT1) on 1p13.3 is independently associated with abdominal aortic aneurysm. *Human Molecular Genetics*, 22(14), 2941–2947.
- Jones, G. T., Thompson, A. R., van Bockxmeer, F. M., Hafez, H., Cooper, J. A., Gollidge, J., ... van Rij, A. M. (2008). Angiotensin II type 1 receptor 1166C polymorphism is associated with abdominal aortic aneurysm in three independent cohorts. *Arteriosclerosis Thrombosis and Vascular Biology*, 28(4), 764–770.
- Kalf, R. R., Mihaescu, R., Kundu, S., de Knijff, P., Green, R. C., Janssens, A. C. (2014). Variations in predicted risks in personal genome testing for common complex diseases. *Genetics in Medicine*, 16(1), 85–91.
- Kampstra, P. (2008). Beanplot: A boxplot alternative for visual comparison of distributions. *Journal of Statistical Software*, 28(Code Snippet 1), 1–9.
- Klein, R., Chou, C. F., Klein, B. E., Zhang, X., Meuer, S. M., & Saaddine, J. B. (2011). Prevalence of age-related macular degeneration in the US population. *Archives of Ophthalmology*, 129(1), 75–80.
- Lautenbach, D. M., Christensen, K. D., Sparks, J. A., & Green, R. C. (2013). Communicating genetic risk information for common disorders in the era of genomic medicine. *Annual Review of Genomics and Human Genetics*, 14, 491–513.
- McGeechan, K., Macaskill, P., Irwig, L., & Bossuyt, P. M. (2014). An assessment of the relationship between clinical utility and predictive ability measures and the impact of mean risk in the population. *BMC Medical Research Methodology*, 14, 86.
- Morrison, A. C., Bare, L. A., Chambless, L. E., Ellis, S. G., Malloy, M., Kane, J. P., ... Boerwinkle, E. (2007). Prediction of coronary heart disease risk using a genetic risk score: The atherosclerosis risk in communities study. *American Journal of Epidemiology*, 166(1), 28–35.
- Pepe, M. S., Gu, J. W., & Morris, D. E. (2010). The potential of genes and other markers to inform about risk. *Cancer Epidemiology, Biomarkers & Prevention*, 19(3), 655–665.
- Pyke, R., & Prentice, R. L. (1979). Logistic disease incidence models and case-control studies. *Biometrika*, 66(3), 403–4011.
- Rooke, T. W., Hirsch, A. T., Misra, S., Sidawy, A. N., Beckman, J. A., Findeiss, L. K., ... White, J. V. (2012). 2011 ACCF/AHA focused update of the guideline for the management of patients with peripheral artery disease (updating the 2005 guideline): A report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines—Developed in collaboration with the Society for Cardiovascular Angiography and Interventions, Society of Interventional Radiology, Society for Vascular Medicine, and Society for Vascular Surgery. *Catheterization and Cardiovascular Interventions*, 79(4), 501–531.
- Saracini, C., Bolli, P., Sticchi, E., Pratesi, G., Pulli, R., Sofi, F., ... Giusti, B. (2012). Polymorphisms of genes involved in extracellular matrix remodeling and abdominal aortic aneurysm. *Journal of Vascular Surgery*, 55(1), 171–179 e172.
- Scott, I. C., Seegobin, S. D., Steer, S., Tan, R., Forabosco, P., Hinks, A., ... Lewis, C. M. (2013). Predicting the risk of rheumatoid arthritis and its age of onset through modelling genetic risk variants with smoking. *PLoS Genet* 9(9), e1003808.
- Smelser, D. T., Tromp, G., Elmore, J. R., Kuivaniemi, H., Franklin, D. P., Kirchner, H. L., & Carey, D. J. (2014). Population risk factor estimates for abdominal aortic aneurysm from electronic medical records: A case control study. *BMC Cardiovascular Disorders*, 14, 174.
- Thompson, A. R., Drenos, F., Hafez, H., & Humphries, S. E. (2008). Candidate gene association studies in abdominal aneurysm disease: A review and meta-analysis. *European Journal of Vascular and Endovascular Surgery*, 38(1), 764–770.
- van Dieren, S., Beulens, J. W., Kengne, A. P., Peelen, L. M., Rutten, G. E., Woodward, M., ... Moons, K. G. (2012). Prediction models for the risk of cardiovascular disease in patients with type 2 diabetes: A systematic review. *Heart*, 98(5), 360–369.
- Verma, S. S., de Andrade, M., Tromp, G., Kuivaniemi, H., Pugh, E., Namjou-Khales, B., ... Ritchie, M. D. (2014). Imputation and quality control steps for combining multiple genome-wide datasets. *Frontiers in Genetics* 5, 370.
- Weeks, D. E., Conley, Y. P., Mah, T. S., Paul, T. O., Morse, L., Ngo-Chang, J., ... Gorin, M. B. (2000). A full genome scan for age-related maculopathy. *Human Molecular Genetics*, 9(9), 1329–1349.
- Weeks, D. E., Conley, Y. P., Tsai, H. J., Mah, T. S., Schmidt, S., Postel, E. A., ... Gorin, M. B. (2004). Age-related maculopathy: A genomewide scan with

continued evidence of susceptibility loci within the 1q31, 10q26, and 17q25 regions. *Journal of Human Genetics* 75(2), 174–189.

Weeks, D. E., & Ott, J. (1990). Reply to Dr. Carothers: Support intervals for genetic risks. *Journal of Human Genetics*, 47, 166.

Wimmer, V., Lehermeier, C., Albrecht, T., Auinger, H. J., Wang, Y., & Schon, C. C. (2013). Genome-wide prediction of traits with different genetic architecture through efficient variable selection. *Genetics*, 195(2), 573–587.

Wray, N. R., Goddard, M. E., & Visscher, P. M. (2007). Prediction of individual genetic risk to disease from genome-wide association studies. *Genome Research*, 17(10), 1520–1528.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

How to cite this article: Shan Y, Tromp G, Kuivaniemi H, Smelser DT, Verma SS, Ritchie MD, Elmore JR, Carey DJ, Conley YP, Gorin MB, and Weeks DE. (2017). Genetic risk models: Influence of model size on risk estimates and precision. *Genetic Epidemiology*, 00:1–15. doi:10.1002/gepi.22035.