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Gene regulatory mechanisms underlying sex differences in brain development and psychiatric disease

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Abstract

The sexual differentiation of the mammalian nervous requires the precise coordination of the temporal and spatial regulation of gene expression in diverse cell types. Sex hormones act at multiple developmental time points to specify sex-typical differentiation during embryonic and early development and to coordinate subsequent responses to gonadal hormones later in life by establishing sex-typical patterns of epigenetic modifications across the genome. Thus, mutations associated with neuropsychiatric conditions may result in sexually dimorphic symptoms by acting on different neural substrates or chromatin landscapes in males and females. Finally, as stress hormone signaling may directly alter the molecular machinery that interacts with sex hormone receptors to regulate gene expression, the contribution of chronic stress to the pathogenesis or presentation of mental illness may be additionally different between the sexes. Here, we review the mechanisms that contribute to sexual differentiation in the mammalian nervous system and consider some of the implications of these processes for sex differences in neuropsychiatric conditions.

Keywords

epigenetic mechanisms; sex differences; steroid hormones; development; neuropsychiatric illness

Many psychiatric conditions display a sex bias in incidence, onset, or symptoms; however, the underlying mechanisms that lead to these sex differences are still obscure. Here, we consider some of the levels of development and mechanisms of gene regulation in which sex-specific processes may contribute to sexual dimorphisms observed in mental illness, with a focus on gene regulation and epigenetic mechanisms. We first describe early behavioral sex differences in humans, focusing on two early-presenting disorders that are diagnosed more frequently in males: autism spectrum disorder (ASD) and attention-deficit/ hyperactivity disorder (ADHD). We next consider the developmental events that give rise to sex differences in the brain and discuss gene regulatory mechanisms that may underlie the

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Competing interests

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persistent effects of these processes on brain function. We then explore how similar genetic risks for ASD and schizophrenia could be invoked in either early life or adolescence, resulting in the distinct trajectories seen in these two conditions. We propose that the intersection between the gene programs that regulate sex-specific development and function in the brain and those that are disrupted in the context of psychiatric illness can significantly influence the pathogenesis and presentation of most neuropsychiatric conditions.

Sex differences in human behavior and neurodevelopmental disorders

Before considering the molecular mechanisms through which pathways for sexual differentiation may intersect with genetic and environmental factors that contribute to psychiatric illness, we first consider a few examples in which early sexual differentiation of the nervous system manifests in early differences in behavior between boys and girls. Not surprisingly, these differences appear in the context of both normal development and in the early presentation of neuropsychiatric conditions.

Little debate exists as to the effects of cultural influences on the early aspects of sex-typical behavior in children. Indeed, the dichotomy of nature versus nurture and their distinct contributions to sex-typical development and behavior is generally acknowledged as false, and human sex-typical behaviors are considered to be the result of dynamic interactions between pre- and postnatural biological factors as well as a child's social milieu (reviewed in Refs. 1–3). At the earliest stages of life, consistent differences in behavior and neurologic development appear between boys and girls. Infant girls appear to have more intense and expressive reactions to painful stimuli.^{1,2} Male infants appear to track objects more consistently in an event-mapping task.^{2,3} Intriguingly, while both infant boys and girls attend more to female faces, no significant difference in eye-tracking of social stimuli appears between in the sexes in healthy infants.^{$3-5$} Such differences reflect only a few of the likely innate differences in neural function between infant boys and girls. Nevertheless, they illustrate the perhaps intuitive conclusion that, like other mammals, embryonic and early developmental programs regulated by sex hormones result in innate differences between the sexes that are present at the earliest stages of life.

While the consequences of such differences for normal development remain unknown, though actively explored, they likely contribute to differences in presentation seen in pediatric neuropsychiatric conditions. Two examples serve to illustrate how, in addition to likely sex differences in the molecular pathogenesis of such disorders (discussed below), innate sex differences in behavior likely also contribute to the differences in symptoms observed between boys and girls. ASD is diagnosed nearly four times more often in boys than in girls.^{6–9} In addition to this overall sex difference, the presentation of ASD also differs between boys and girls. While many studies have documented a higher incidence of intellectual impairment in girls with ASD compared with boys, some debate exists as to whether the overall severity of ASD differs between them.¹⁰ Differences between the presentation of boys and girls with ASD reflect biological differences in symptoms, which may also contribute to biases in reporting and attention. Externalizing behaviors, such as aggression, repetitive behaviors, restricted interests, reduced prosocial behavior, and hyperactivity tend to occur more prominently in males, while females with ASD have more

internalizing symptoms, including anxiety, depression, and self-oriented emotional disruption, that are often only reported by parents.^{10–12} As a consequence of these potential differences in presentation, ASD in boys is more likely to come to attention due to disruptive behaviors in school or at home, thereby skewing the bias in prevalence even further. Sex differences in innate responses to stimuli or patterns of emotional expressivity likely contribute to distinct presentation of these disorders. Indeed, sex differences in patterns of cognition, in particular specific dimensions of social cognition and abstract reasoning associated with identifying rules and patterns within systems, led to the hypothesis that domains of autistic symptoms may arise from an extreme masculinization of the brain in autistic patients.13 Nevertheless, despite many efforts to determine if prenatal exposure to increased levels of androgens is associated with ASD, no causal mechanisms have been established linking sex hormone signaling to $ASD^{14,15}$ However, it is worth noting that such a relationship has been repeatedly supported, linking elevated levels of umbilical cord testosterone to language delays in children, suggesting that, in specific contexts likely determined by diverse genes affecting sensitive aspects of CNS development, sex differences in developmental pathways may contribute to patterns of symptoms that children present.¹⁶ Additional mechanisms may mediate sex differences in the sensitivity to genomic disruption or the consequences of ASD mutations for neural development.

Given the innate differences between the sexes in the presentation or pathogenesis of most if not all psychiatric conditions, it is worth considering whether a re-evaluation of the criteria for those particularly diagnosed in childhood may be warranted. For example, in ASD, aside from specific core deficits in social cognition and emotion processing, other patterns of symptoms used to establish diagnostic criteria may vary between the sexes. Further investigation of sexual dimorphisms in phenotypes resulting from disease-associated mutations will be central to such refinements in clinical criteria.

Extreme sex biases in the diagnosis of ADHD have been observed, such that boys can outnumber girls by up to 10:1, though meta-analyses and population-based studies suggest that this ratio is likely closer to $4:1.^{17-19}$ Intriguingly, sex differences in the prevalence of ADHD appear highest during childhood and become significantly less pronounced later in life.20,21 As with ASD, given that the pattern of symptoms of ADHD manifested by boys more strongly correlates with conduct disorder and disruptive behaviors, parents or teachers are more likely to bring these cases to attention. Consistent with a model in which females are more likely to manifest comorbid distress or disorders with internalizing symptoms, girls are typically diagnosed later in childhood than their male counterparts.22 However, in addition to sex differences in overall prevalence rate, the distribution of subtypes or patterns of associated phenotypes also appears to differ between the sexes, with a larger percentage of females (45–60%) presenting as inattentive compared with males (35–50%).^{17,23} Thus, innate sex differences in the circuits regulating attention or behavioral inhibition may underlie the presentation of ADHD, as well as other neuropsychiatric conditions. Indeed, it has been observed that boys with ADHD exhibit deficits in behavioral inhibition, while girls tend to display impairments in planning.²⁴ Given that the extent and pattern of sex differences in ADHD-associated symptoms appear to decrease with age into adulthood, it is interesting to speculate that, despite innate differences in neural circuits underlying attention and behavioral inhibition, the developmental pathways activated during adolescence

(discussed below) normalize these differences and restructure such circuits to participate in behavioral processes common to both sexes in adults, while distinct pathways elaborate the sexual dimorphisms in behavior and pathology seen in later in development.

Independent of the societal factors that likely skew the diagnoses of these two syndromes, the male bias of ASD and ADHD prevalence and the distinct patterns in which boys versus girls present with these disorders suggest that innate sex differences in the brain are likely programmed during the earliest stages of development and contribute to sex differences in the symptoms observed. We now turn to the molecular mechanisms that mediate the sexual differentiation of the mammalian nervous system and discuss how these pathways may be altered to give rise to sex differences in the pathogenesis of neuropsychiatric conditions.

Sexual differentiation of the brain is regulated by gonadal hormones

Much of our knowledge about the cellular and molecular differences between the sexes in the mammalian brain has been obtained through studies of the hormonal regulation of the differentiation and function of neural circuits underlying innate, sex-typical behaviors and physiology in rodents, particularly sexual behavior and territorial aggression. The neural circuitry that controls these behaviors develops under the control of gonadal hormones.^{25–29} Male mice undergo a surge of testosterone at birth that subsides within hours.^{30,31} This circulating testosterone is directly converted to estradiol in the brain by aromatase.^{32,33} Estradiol is the primary endogenous estrogen, although estrone and estriol also bind estrogen receptors; here, we primarily use the general term estrogen for simplicity. Pharmacological and genetic experiments have demonstrated that this brain-derived perinatal estrogen is the primary driver of sexual differentiation of the brain and permanently establishes sex-typical differences in the structure and function of the neural circuitry that mediates sex-specific behaviors in the adult.^{28,34–36} Females given estradiol at birth display male-typical fighting behavior as adults with no additional hormone supplementation.³⁷ This sensitivity to estradiol is lost by the second postnatal week. $38-40$ Although sex differences in neural circuitry are specified during this postnatal critical period, sex-typical behaviors are not displayed until puberty, when the male testes produce testosterone and female ovaries make estrogens and progesterone. These hormones are acutely required in adult life: gonadectomy abolishes mating and aggression, but the circuit structure remains intact and behaviors can be restored by exogenous hormones. Although testosterone is the primary driver of adult male-typical behaviors, estradiol alone can restore some mating and territorial behaviors. 41–44 Therefore, estrogen acts to both modulate postnatal male-typical circuit development and to "activate" circuits for sex-typical behaviors in adulthood.

In addition to its masculinizing effects on behavior, perinatal estrogen is known to give rise to anatomic and molecular sex differences. Many excellent reviews have summarized findings on cellular and neuroanatomic sex differences, including cell number, neural projections, and spine number.27,34,45–48 Sex differences in gene expression in the brain have also been described in both rodents and humans.^{49–56} Although estrogen is the primary driver of sexual differentiation in rodents, both estrogen and testosterone signaling are required for full masculinization of adult behaviors. Male mice mutant for androgen receptor (AR), the receptor for testosterone, show decreased spatial memory, increased anxiety,

In humans, however, it appears that brain masculinization occurs largely through testosterone signaling, rather than estrogen. Human males with mutations in CYP19A1, the gene for aromatase, cannot synthesize estrogen, yet present as normal males. Men with aromatase deficiency experience sustained linear growth rather than a pubertal growth spurt and epiphyseal closure, demonstrating that estrogen is required in males for proper skeletal maturation.61 In contrast, AR function is essential for phenotypic and behavioral masculinization of human males. Patients with an XY karyotype and a complete loss of AR function have complete androgen insensitivity syndrome (CAIS), present as women, and have female-typical brain morphology.62 Humans also experience developmental testosterone surges; though consistent with the scaling of natal development, they are much more prolonged than those in rodents. The testes begin to secrete testosterone around week 7 of gestation, with maximal levels between weeks 8 and 24.63,64 Human brain at midgestation is similar to mouse brain at birth with regard to staging of cortical development. 65,66 The timing of developmental hormone surges is thus somewhat conserved between rodents and humans: the mid-gestation testosterone surge in humans is concordant with the perinatal surge in mice and rats. Human males also experience an additional surge in infancy that peaks between months 1 and $3.67,68$ Female ovaries are also known to be active during infancy, but the levels of estradiol are variable, and the time course of its secretion has not been well described.^{68–70} The timing of these increases in gonadal hormone levels intersects with neural development so that male and female brains have very different internal states during neurogenesis, neuronal migration and synaptogenesis (Fig. 1). Consequently, it is not surprising that, like the myriad dimorphisms observed in rodents due to the postnatal testosterone surge, humans also demonstrate innate differences during fetal and infant development between the sexes.71,72

Sex chromosomes and brain development

Sex chromosomes also contribute to sexual differentiation of the brain, both directly through their own genetic content and indirectly through regulation of gonadal development (reviewed in Refs. $^{73-76}$). Sex chromosome aneuploidies are some of the most common genetic disorders in humans, affecting nearly 1/400 live births.77 These disorders are associated with cognitive and behavioral symptoms, particularly social skills and motor abilities.78 Notably, language and spatial abilities appear to correlate with sex chromosome dosage; females with X monosomy show normal or increased verbal and lexical abilities and visuospatial deficits, while individuals with sex chromosome polysomy have language impairments that increase with the number of chromosomes, while spatial skills are often enhanced.77–79 Brain imaging studies have identified a relationship between sex chromosome dosage and brain volume⁷⁷ and highlight specific chromosomal effects in cortical⁸⁰ and subcortical^{81,82} brain areas. Mouse models of sex chromosome aneuploidies have been used to discern the effects of sex chromosomes on specific behaviors, including social behaviors, anxiety, feeding, and nociception.⁷⁴ The most widely used model is that of the "four core genotypes." This system employs two modified alleles of the testis-

determining Sry gene; one where Sry has been deleted from the Y chromosome, resulting in genetic males that resemble females, and another where Sry has been inserted on an autosome to generate XX animals that develop testes.⁸³ Comparison of these mutants with wild-type XX and XY animals thereby permits the dissociation of sex chromosome complement and gonadal development.

Similar to Sry, the few other genes on the Y chromosome are primarily specialized for testis determination and spermatogenesis.84 Gene products such as DDX3Y, UTY, and KDM5D are abundant in the brain, but their neural functions have not been elucidated.^{51,85–87} In contrast, the X chromosome is enriched for genes that control brain function; 40% of the genes on the X are expressed in the brain, and X-linked gene expression is higher in the brain of both sexes compared with other tissues. $88-91$ As the X chromosome is also the most rapidly evolving mammalian chromosome, it has been suggested that this enrichment of neural-expressed genes is a consequence of natural selection for increased cognitive abilities.88,92–94 Consequently, X-linked mutations are a leading cause of intellectual disability, a condition that is more prevalent in boys than girls. $89,93,94$ As female cells achieve dosage compensation through stochastic inactivation of one X chromosome (XCI), females are buffered from the effects of deleterious X mutations.^{95,96} A recent analysis of XCI in diverse human cell types and tissues established that the extent of X inactivation varies within and between individuals and is often incomplete.⁹⁷ In brain, excitatory and inhibitory neurons show different patterns of clonal XCI mosaicism within a cortical column owing to their different developmental migration patterns. Excitatory neuron XCI is heterogeneous with high variance within and between individuals, while inhibitory neurons show equal, fine-grained inactivation patterns, suggesting that any heterozygosity for an Xlinked gene would affect excitatory circuitry only in specific clonal areas, but all inhibitory circuits would be affected equally.⁹⁸

What genes on the X or Y chromosomes are contributing to sex differences in brain development and function? Histone lysine demethylases are intriguing candidates, as they can induce broad, persistent effects on gene expression. Kdm6a/Utx and Kdm5c/SMCX are known escapers of X-inactivation, resulting in increased expression of these epigenetic regulators in the brains of females. Mutations in KDM5C have been implicated in a variety of neurodevelopmental disorders, including intellectual disability, ASD, and cerebral palsy. $99,100$ Mice lacking *Kdm5c* display increased aggression, decreased anxiety, impaired motor coordination, and decreased dendritic arborization in the amygdala, although these phenotypes are dramatically reduced with a forebrain-specific deletion in adulthood.^{100,101} This finding provides evidence that these enzymes have distinct genomic targets in specific cellular or developmental contexts. In the next section, we will explore current questions and approaches regarding regulation of gene expression in the brain.

Gene regulation in the brain: unique strategies and new methodologies

From a general perspective, epigenetic regulation refers to mechanisms that mediate persistent changes in gene expression in response to transient events, such as developmental programs, experience, or environmental cues, although the stringency of this definition is much debated.^{102–105} Thus, the activity of a specific gene or sets of genes across the genome

is regulated by changes in chromatin structure that occur via interactions with the gene's local environment (*cis*) or other regions in the genome (*trans*) and changes in nuclear structure. Such chromatin remodeling involves dynamic processes that include the movement of histones along DNA, covalent modifications to histone proteins (including acetylation, methylation, and phosphorylation), the binding of transcription factors to regulatory regions and the subsequent recruitment of coactivators and corepressors that may be responsible for such modifications, or the covalent modification of DNA itself. These processes work to integrate developmental and environmental signals over time to determine levels and patterns of gene expression.

Several recent reviews discuss general principles of epigenetics and gene regulation in neurons.104,106–111 While the details of the mechanisms regulating neural gene expression are beyond the scope of this review, we wish to emphasize the unique developmental trajectory of chromatin state in neurons and to highlight recent insights obtained from highthroughput sequencing approaches. Chromatin structure is known to reorganize during the peak of synaptogenesis in postnatal life. This was first observed through analysis of chromatin repeat length, which reflects the length of the linker DNA between nucleosomes and is a proxy for the density of chromatin packing.112 Chromatin repeat length in cortical neurons is shorter than that of neighboring glia or other cell types,¹¹³ which suggests a unique pattern of higher-order chromatin organization in neurons. Furthermore, shortening of repeat length coincides with region-specific dynamics of neuronal maturation, suggesting that regions undergo chromatin reorganization in response to neural activity and circuit formation.114–116 Epigenomic analyses are now elaborating on these pioneering findings, particularly through studies on higher-order chromatin organization and DNA methylation. 117–122

The extraordinary heterogeneity of the mammalian nervous system raises challenges in dissecting the role of a given factor or genetic variant in precise spatial and developmental contexts. The advent of single-cell sequencing has made it possible to identify and classify neurons from their transcriptomes in addition to morphological or electrophysiological characteristics.123–127 The cis-regulatory elements that coordinate the specification and function of discrete neuronal types are being revealed through technological refinements in chromatin immunoprecipitation sequencing (ChIP-seq) and ATAC-seq (assay for transposase-accessible chromatin using sequencing) methods.125,128–131 Future studies will describe the dynamics of these elements across development or in disease. In postnatal development, experience-dependent neural activity induces transcriptional programs that sculpt neural circuits by regulating synapse development and plasticity.^{132,133} The complex dialogue between the synapse and the nucleus involves diverse adhesion molecules, scaffolding proteins, and chromatin regulators, many of which have been implicated in neurodevelopmental disorders, such as ASD .^{134–136} Thus, perturbation of the mechanisms that regulate gene expression at a genomic level may affect the development of the nervous system during the earliest stages, causing global disruption in neuronal differentiation and wiring or, at later time-points, causing abnormalities in synaptic function or activitydependent processes that underlie learning or more complex aspects of information processing in the developed brain.

Epigenetic mechanisms can define developmental trajectories

Gonadal hormones, such as estrogen and testosterone, bind steroid hormone receptors (SRs): nuclear receptor transcription factors that can recruit chromatin remodeling machinery to activate or repress gene expression. Accordingly, it has been proposed that developmental exposure to hormones organizes sexual differentiation of the brain in part through long-term effects on gene expression.137–141 Differential exposure to hormones in males and females during a neurodevelopmental critical period is likely to result in sex-specific patterns of gene expression by SRs that can lead to persistent sexually dimorphic chromatin patterning (Fig. 2). Consequently, when the gonads begin to secrete hormones at puberty, the same SRexpressing neurons would possess a differential capacity to respond to the same hormone stimulus, as a consequence of the previously established chromatin landscape. However, it has proven difficult to test this hypothesis, as sex differences in gene expression are subtleeven key drivers of sex differences, such as estrogen receptor α (ERα) and AR vary by only a few folds in expression levels and are rarely detected in genome-wide screens for sex differences. Identifying sites of hormone action and the mechanisms by which hormones regulate gene expression in the brain is essential for understanding which neural processes are likely to differ between the sexes. Given the dynamics of sex hormone levels throughout development, it becomes apparent that disruptions in specific neurodevelopmental programs may intersect with sex-specific gene regulation at various points in development or adulthood to cause differences in the symptoms experienced by males and females.

Regulation of gene expression by steroid hormone receptors

Studies on sex differences in SR function in the brain have focused primarily on the fundamental differences in circulating ligands: female ovaries primarily produce estrogen and progesterone, while male testes release testosterone that can be converted into local estradiol in the brain. However, there are many additional factors that can fine-tune specificity in hormone-regulated gene expression, such as neural production of diverse steroid hormones (neurosteroids), transcription co-factor expression, and local chromatin context. Extensive details of SR mechanisms have been elucidated over the past 20 years; however, the role of these receptors in regulating gene expression in distinct, behaviorally relevant populations of neurons remains poorly characterized. This is largely due to the technical challenges of isolating and purifying sufficient numbers of the sparse populations that express hormone receptors. Here, we discuss mechanisms of gene regulation by SRs, emphasizing insights obtained from biochemical and genomic approaches in non-neural systems, to better understand how SR signaling can give rise to sex differences in gene expression in the brain. We then discuss regions of the mammalian brain in which steroid hormones are known to mediate sex-specific differentiations.

In the classic description of gene activation by SRs, circulating steroid hormones diffuse through the cell membrane and bind to receptors, which then undergo a ligand-dependent conformational change, dimerization, and association with cognate recognition sequences on DNA.142,143 Estrogen receptor β (ERβ) is highly homologous to ERα, with 95% homology in the DNA-binding domain and 55% homology in the ligand-binding domain and similar ligand-binding affinity for physiological estrogens.¹⁴⁴ ER α and ER β bind the same

recognition sequence and can heterodimerize.145 As these receptors are co-expressed in many brain regions, they therefore have the potential to both cooperate and compete with one another. In addition to sex differences in levels of hormones and their receptors, sexually dimorphic transcription programs can emerge from over 350 transcriptional cofactors that provide contextual specificity and tuning of gene expression.^{146,147} Nuclear receptor coactivators are large multi-unit complexes that link sequence-specific hormone receptors to the general transcription machinery, as well as enzymatic factors that can covalently modify histone tails or invoke ATP-dependent chromatin remodeling machinery.¹⁴⁸ Corepressors generally bind unliganded receptors and recruit histone deacetylase (HDAC)-containing enzymatic complexes that maintain a repressive chromatin state.¹⁴⁷ Of note, nuclear receptor corepressor 1 (NCoR) interacts with MeCP2¹⁴⁹ and is also an ASD risk gene.¹⁵⁰ Coexpression analysis of Allen Brain Atlas data recently identified the SR cofactors, including NCoR, that are likely to play key roles in the mouse brain.¹⁵¹ It will be interesting to determine whether these factors contribute to sex differences in gene expression or to disease susceptibility. Characterization of neuron-specific or cell-type specific SR coregulators could lead to the development of neural selective estrogen receptor modulators (SERMs) that potentiate or attenuate ERα/β transcriptional output in a defined population of neurons to provide neuroprotective or mood benefits.

Steroid hormones are also known to produce rapid changes in neuronal function or behavior that occur on the order of minutes, rather than hours or days.^{152–154} Notably, many of these non-genomic effects may in fact converge at the level of gene regulation. Studies in diverse tissues and cell types have demonstrated that estrogen and testosterone can act through intracellular signaling cascades to mobilize calcium stores, causing CREB phosphorylation. ¹⁵⁵ Phospho-CREB can then activate a number of immediate early genes, including FOS, and this cascade is required for estrogen-dependent dendritic spine formation.156,157 An additional level of cross talk can occur at the level of signaling through Fos itself. Fos protein heterodimerizes with Jun to form the AP-1 transcription factor, which directly binds ERα and recruits it to DNA via a tethering mechanism.158,159 Thus, steroid hormones can regulate gene expression both through their own consensus DNA-binding sites and through the actions of phospho-CREB and immediate early genes. Such cross talk may explain why social experience can enhance or replace the effects of gonadal hormones on innate behaviors in rodents, as seen in maternal behaviors demonstrated by reproductively naive females exposed to pups.^{160–162}

SR expression patterns in the brains of rodents and humans

All four gonadal hormone receptors (AR/PR/ERα/ERβ) are expressed most abundantly in limbic and hypothalamic areas that regulate innate reproductive behaviors, including the bed nucleus of the stria terminalis (BNST), the medial preoptic hypothalamus (MPOA), the medial amygdala (MeA), and the ventrolateral nucleus of the ventromedial hypothalamus. 151,163–168 All but ERβ are expressed in the arcuate nucleus, which regulates homeostasis including feeding and energy balance.¹⁶⁹ Extensive analysis of ER α , ER β , and PR expression describes signal throughout the cortex and in midbrain areas, such as the ventral tegmental area (VTA), substantia nigra (SNc), periaqueductal gray (PAG), and raphe. 163,164,170–173 Thus, sex differences in reward processing and reward-seeking behavior may

be controlled by the effects of SR function in VTA- and SNc-associated dopaminergic pathways, while expression in the PAG may underlie sex differences in pain processing and analgesia. Similarly, serotonergic projections from raphe nuclei have ramifications throughout the brain, and widespread effects of such neuromodulation may underlie sex differences in fear and anxiety behaviors, as well as stress-sensitivity and the activation of the hypothalamic–pituitary–adrenal axis.¹⁷⁴ AR is also expressed in the cortex, particularly in the primary visual cortex and prefrontal cortex.¹⁷⁵ These four receptors are present in the suprachiasmatic nucleus of the hypothalamus, which regulates circadian rhythm, allowing gonadal hormones to directly influence daily fluctuations in the adrenal output, sleep and mood.¹⁷⁶ Finally, ER α , ER β , and AR are found in astrocytes and endothelial cells,^{176,177} while $ER\beta$ has anti-inflammatory effects in microglia.¹⁷⁸ There are still few studies quantifying the co-expression of these receptors or detailing the ontogeny of their expression. RNA-seq experiments and fluorescent double *in situ* hybridization will provide further insight into the shared sites of action of SRs, pinpoint novel hormone-responsive populations, and classify the cell types that express gonadal hormone receptors.

In humans and monkeys, ERα and ERβ are expressed in similar areas as in mice, including high levels in the hypothalamus and amygdala and lower expression in the hippocampus and cortex, particularly the temporal cortex.¹⁷⁹ ER β is more prominent in the cortex compared to the hypothalamus, with significantly higher levels of expression in the deep layers of the temporal and entorhinal cortex, suggesting that ERβ is the principal modulator of estrogen effects on cognition in humans.¹⁸⁰ AR is similarly expressed in the hypothalamus, amygdala, and temporal cortex, as well as the diagonal band of Broca.¹⁸¹

Sex differences in SR expression and the onset of SR expression during brain development have not been well described in humans, 182 and indeed these receptors are only minimally detected in human brain transcriptome studies. Focused gene expression analyses in subcortical areas rich in SRs may therefore identify downstream genes regulated by hormone receptors that show a sex bias in expression. One such region, the accessory basal nucleus of the amygdala, is larger in primates compared with rodents and sends projections to the hippocampus and entorhinal cortex.¹⁷⁹ At a more general level, sex-specific processes in the neural circuits underlying dimorphic behavior can arise from the differentiation of these pathways at different points¹⁸³ (Fig. 3). Better understanding of where hormone receptors are expressed in humans, when expression is initiated during development, and the connectivity of these regions is necessary to understand how gonadal hormones can specify unique developmental trajectories in males and females.

As discussed above, surges in testosterone during early development mediate male-specific aspects of early brain development and differentiation. Lasting changes in gene expression programmed by such developmental hormone surges or perturbations during hormonemediated sexual differentiation of the brain may therefore contribute to the male bias observed in some neuropsychiatric disorders. Despite limited knowledge of the extent of SR expression in the human brain, sex differences in specific syndromes may be linked to hypothalamic dysfunction stemming from abnormal activity or development in individual nuclei. For example, sex differences in impulsivity in the form of aggression and hypersexual behavior seen in neurodevelopmental conditions such as ASD are thought to

arise, in part, as a consequence of SR-regulated sex dimorphisms in hypothalamic nuclei that control reproductive drive and contribute to affective state.¹⁸⁴ Similarly, it has been well documented that males and females differ in patterns of activation in the amygdala in response to acute stress, and sex differences in the sensitivity of the amygdala to chronic stress, as well as in the connectivity and function of corticoamygadlar pathways, is thought to underlie aspects of the dramatic sex differences in the incidence and presentation of depression and anxiety.^{185–187} In addition, sex differences in the function and connectivity of serotonergic projections from the raphe nuclei throughout the brain also likely contribute to the dimorphisms seen in mood and anxiety disorders, as well as the response to medications that act on these pathways.188–190 Finally, the dense interconnectivity of reward pathways with frontal cortical circuitry is involved in attention, planning, and other aspects of cognition and may contribute to sex differences in the incidence and presentation of ADHD, as well as schizophrenia.^{191–193} In a similar manner, it is likely that sex differences in midbrain dopaminergic signaling underlies some of the sex differences in patterns of drug abuse and addiction, as well as the difference between the sexes in their response to antipsychotic medications.194–196

Integrating patient sequencing studies and patient symptoms to link genes to pathology

As the list of syndromic neuropsychiatric disorders––those caused by mutations in specific genes––continues to grow, deep phenotypic analyses of the discrete patterns of deficits caused by disruption of individual genes will help to elucidate the developmental and molecular programs that underlie specific and shared neural processes. Most neuropsychiatric conditions are thought to occur as a consequence of the interactions of environmental risk factors with genetically specified developmental sensitivity. The advent of high-throughput sequencing led to an explosion of patient-sequencing studies that provided fundamental insight into the developmental origins of psychiatric disorders. We will now examine the findings in ASD and schizophrenia to discuss the possible mechanisms by which epigenetic regulation of neural development may interact with pathways for sexual differentiation in the nervous system.

Using a wide range of techniques to examine patterns of mutations associated with neuropsychiatric conditions, culminating recently with whole-exome sequencing of patients and their parents and siblings to identify de novo and rare coding mutations, many studies have led to the identification of genes whose mutation correlates with the occurrence of ASD and schizophrenia, as well as depression and bipolar disorder (reviewed in Refs. $197-201$). Whole-genome sequencing is now extending this work to identify noncoding mutations associated with disease. $202,203$ These studies are likely to identify regulatory elements that direct specific aspects of brain development and function and may provide insight into the majority of disease cases, which have no currently known genetic origin. With regards to ASD in particular, in addition to implicating neural specific processes, including synaptic structure and neuronal excitability, these studies revealed a critical role for genes implicated in chromatin and transcriptional regulation in the pathogenesis of these disorders^{66,197,200,204} (Fig. 4). Integrating patient genome–sequencing studies with human

gene expression and epigenomic data has identified key regulatory nodes and pathways highlighting the importance of chromatin regulation in brain development and function. 66,197,200,204–213

ASD and schizophrenia: shared genes but distinct trajectories?

Recent evidence suggests an overlap in the genetic risks for ASD and schizophrenia with regard to rare copy number variants and *de novo* mutation events, particularly for genes involved in synaptic function and immune regulation, though the contribution of shared common genetic risk between these disorders is less clear. $214-216$ Like ASD, the incidence and presentation of schizophrenia shows a significant difference between males and females. ²¹⁷ Males typically present earlier in adolescence with more severe symptoms and poorer outcomes, suggesting that fundamental sex differences in the developing brain contribute to the onset and pattern of deficits. However, despite some overlap in the mechanisms contributing to ASD and schizophrenia and aspects of the behavioral and cognitive deficits with which they present, the dramatic difference in their trajectories illustrates the central role of distinct developmental periods.

As discussed above, ASD is typically diagnosed early in childhood, suggesting disruptions of early developmental programs in the brain that result in deficits at the earliest stages of life. Consistent with this hypothesis and the identification of multiple genes involved in chromatin regulation and structure as risk alleles, recent work has identified genome-wide alterations in the regulation of noncoding RNAs, patterns of histone modification, and higher-order chromatin structure in tissue from patients with ASD.^{211,213,218,219} In contrast, the majority of symptoms in schizophrenia evolve during adolescence, suggesting that distinct developmental programs or mechanisms that are activated by the onset of puberty may play roles in the pathogenesis.220 The initiation of developmental programs following the onset of puberty occurs via SR-dependent and SR-independent pathways, both of which contribute to the restructuring and development of the nervous system for adult and sexually dimorphic behaviors. A hallmark of these processes that occurs following the initiation of these pathways at the onset of puberty is, for example, the elimination of synapses in regions, including the prefrontal cortex, that continues throughout adolescence and into early adulthood.221,222 Consistent with these observations at the cellular level, functional studies demonstrate that cortical gray matter volume peaks before adolescence and then slowly declines until reaching its adult volume.223 Such synaptic pruning is thought to achieve the balance of excitatory and inhibitory activity in adult cortical regions, with adult patterns of inhibitory activity in prefrontal cortical regions implicated as an essential part of network dynamics and synchronized activity thought to underlie cognition and cortical processing. 224,225 An intriguing additional mechanism for sex differences in the incidence and presentation of psychiatric illnesses during adolescence is the role of microglia in the maturation and pruning that occurs during this adolescent period and the sex differences in gene expression that suggest additional dimorphisms downstream of their activity.^{226,227} Thus, the developmental programs initiated by the onset of puberty that persist throughout adolescence and into early adulthood represent fundamental programs of reorganization that may unmask or create new substrates for the neuropathology that underlies adult presentations of psychiatric illnesses.²²⁸

Intriguingly, recent work suggests shared common genetic overlap in social communication difficulties and both ASD and schizophrenia but distinct patterns in the relationship between genetic traits and each disorder, largely consistent with the onset of clinical symptoms.²¹⁶ Thus, while the sex differences in ASD may be a consequence of the early sex differentiation of the brain during embryonic and perinatal development, those observed in schizophrenia are likely a consequence of the regulatory epigenetic landscape established during this period but only activated by the onset of puberty. In this context, sex differences observed in the onset and presentation of schizophrenia could arise either from the direct activation of genes in a sexually dimorphic manner by gonadal hormones or through signals in adolescence common to both sexes acting on poised sexually dimorphic programs established early in development (Fig. 2).

Consistent with both models, sequencing studies of sex-biased gene expression in regions of the human brain at distinct developmental time points reveals limited overlap between sexbiased genes early in development and in adolescence and an enrichment for genes implicated in neuropsychiatric illness in those with a male expression bias.²²⁹ Future studies will undoubtedly reveal whether additional sets of mutations associated with schizophrenia are enriched in regulatory regions that mediate aspects of gene regulation specifically during adolescence, thus contributing to the developmental window during which symptoms and sex differences manifest.202,230–235

Sex differences and stress: intersection at the level of DNA

As a final example of how such regional regulation of sexually dimorphic differentiation of the nervous system may contribute to sex differences in neuropsychiatric illness, we consider role of early life stress in the later onset of depression. Women have twice the lifetime risk of developing depression compared with men and appear to experience symptoms that are more severe and diverse.^{236,237} Like many other major neuropsychiatric conditions, major depressive disorder typically evolves during early adolescence, with an earlier and distinct presentation in girls compared with boys.^{238,239} This suggests that sexspecific pathways that regulate mood are activated by the onset of puberty and may intersect with stress response pathways to contribute to gender-specific symptoms.^{240,241}

Increasing evidence supports a role for epigenetic changes in the pathogenesis of depression, particularly in response to early-life stress. The interactions with stress signaling and the pathways for sex differentiation suggest a mechanism by which the activation of stress pathways may act sex-specifically to control the response to chronic stress.184,239,242 These findings are supported at the gene regulatory level by recent studies on cross talk between SRs and glucocorticoid receptor (GR). In response to stress, the adrenal glands release glucocorticoid hormones: cortisol in humans and corticosterone (CORT) in rodents. In the brain, CORT binds to both glucocorticoid (GR) and mineralocorticoid (MR) receptors. MR and GR have a highly homologous DNA-binding domain but possess different affinities for CORT: MR has a very high affinity of 0.5 nM, while GR affinity is about 10-fold lower.²⁴³ Thus, MR is thought to respond to the onset of stress, while GR responds to increasing levels of CORT to end the stress reaction and promote memory consolidation,²⁴⁴ although a recent *in vivo* study found that these receptors heterodimerize in response to acute stress.²⁴⁵

Both receptors are expressed in the hippocampus and lateral septum, while GR is found in many other areas, including the central amygdala, paraventricular hypothalamus, and prefrontal cortex.246,247

The DNA-binding domains of GR and MR are homologous to those of AR and PR; therefore, all four of these receptors can bind the same recognition sequences.¹⁴³ ChIP-seq data from castration-resistant prostate cancer (CRPC) has shown that half of AR occupancy sites are bound by GR in dexamethasone-treated cells and that GR can substitute for AR to regulate a subset of AR transcriptional programs.²⁴⁸ These observations reveal a mechanism by which stress can alter sexual differentiation of the brain. For example, FK506-binding protein 5 (FKBP5) is a GR target gene and co-factor that that has been implicated in depression, posttraumatic stress disorder (PTSD), and anxiety.249 AR directly increases FKBP expression in the prostate through multiple distal and intronic enhancers, $250,251$ but regulation of FKBP5 by testosterone signaling in the brain has not yet been explored. GR recruitment can also modify chromatin to facilitate binding by other transcription factors in a process known as *assisted loading*.²⁵² ChIP-seq studies have demonstrated that GR increases chromatin accessibility to modulate ERa binding to DNA through AP-1 sites.^{253,254} This could potentially lead to new estrogen-responsive gene programs after a period of stress. Taken together, these studies suggest that stress can directly alter regulation of gene expression by gonadal hormones in the brain (Fig. 5).

Intriguingly, hypermethylation of the GR locus has been observed in hippocampal tissues from males with a history of abuse who completed suicide.255,256 Both sexes show changes in DNA methylation in immune-related genes in the context of PTSD, while some sex differences are observed in non-neuronal cell types in the setting of depression, 257, 258 highlighting a role for epigenetic changes in mediating long-term pathology via cell type– specific mechanisms. Indeed, multiple studies have already identified heterogeneity in patterns of methylation across neuronal populations.259,260 Thus, pathways induced by chronic stress are likely to interact both with mechanisms that mediate long term, sexspecific patterns of gene regulation across the genome, as well as direct activation of targets of sex hormone regulation to mediate the pathogenesis of depression and PTSD.

Conclusions

In summary, epigenetic mechanisms likely mediate sex-specific differentiation in the nervous system at every stage of development. As these pathways elaborate, the impact of genetic and environmental factors that contribute to psychiatric illness can thus have distinct effects in either sex, contributing to sex differences in the time of presentation, pattern of symptoms, or severity of illness. As we continue to understand the specific developmental programs and neural process that mediate sex-specific differentiation and function in the brain at particular developmental time-points, we will gain deeper insights into how specific mutations sensitize individuals to distinct neuropsychiatric conditions. Although we are not yet capable of developing treatments that target specific epigenetic mechanisms or sexspecific developmental processes to ameliorate the symptoms of any psychiatric illness, understanding how these mechanisms contribute to these conditions in both sexes is critical to the future of treatment in mental health.

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References

- 1. Alexander GM, Wilcox T. Sex Differences in Early Infancy. Child Dev Perspect. 2012; 6:400–406.
- 2. Guinsburg R, et al. Differences in pain expression between male and female newborn infants. Pain. 2000; 85:127–133. [PubMed: 10692611]
- 3. Wilcox T, Alexander GM, Wheeler L, Norvell JM. Sex differences during visual scanning of occlusion events in infants. Dev Psychol. 2012; 48:1091–1105. [PubMed: 22148944]
- 4. Gluckman M, Johnson SP. Attentional capture by social stimuli in young infants. Front Psychol. 2013; 4:527. [PubMed: 23966966]
- 5. Chawarska K, Macari S, Powell K, DiNicola L, Shic F. Enhanced Social Attention in Female Infant Siblings at Risk for Autism. J Am Acad Child Adolesc Psychiatry. 2016; 55:188–195.e1. [PubMed: 26903252]
- 6. Werling DM. The role of sex-differential biology in risk for autism spectrum disorder. Biol Sex Differ. 2016; 7:58. [PubMed: 27891212]
- 7. Werling DM, Geschwind DH. Sex differences in autism spectrum disorders. Curr Opin Neurol. 2013; 26:146–153. [PubMed: 23406909]
- 8. Elsabbagh M, et al. Global prevalence of autism and other pervasive developmental disorders. Autism Res Off J Int Soc Autism Res. 2012; 5:160–179.
- 9. Mattila ML, et al. Autism spectrum disorders according to DSM-IV-TR and comparison with DSM-5 draft criteria: an epidemiological study. J Am Acad Child Adolesc Psychiatry. 2011; 50:583–592.e11. [PubMed: 21621142]
- 10. Bölte S, Duketis E, Poustka F, Holtmann M. Sex differences in cognitive domains and their clinical correlates in higher-functioning autism spectrum disorders. Autism Int J Res Pract. 2011; 15:497– 511.
- 11. Mandy W, et al. Sex differences in autism spectrum disorder: evidence from a large sample of children and adolescents. J Autism Dev Disord. 2012; 42:1304–1313. [PubMed: 21947663]
- 12. Solomon M, Miller M, Taylor SL, Hinshaw SP, Carter CS. Autism symptoms and internalizing psychopathology in girls and boys with autism spectrum disorders. J Autism Dev Disord. 2012; 42:48–59. [PubMed: 21442362]
- 13. Baron-Cohen. The extreme male brain theory of autism. Trends Cogn Sci. 2002; 6:248–254. [PubMed: 12039606]
- 14. Kung KTF, et al. No relationship between prenatal androgen exposure and autistic traits: convergent evidence from studies of children with congenital adrenal hyperplasia and of amniotic testosterone concentrations in typically developing children. J Child Psychol Psychiatry Allied Discip. 2016; 57:1455–1462.
- 15. Mitra I, et al. Pleiotropic Mechanisms Indicated for Sex Differences in Autism. PLoS Genet. 2016; 12:1–27.
- 16. Whitehouse AJO, et al. Sex-specific associations between umbilical cord blood testosterone levels and language delay in early childhood. J Child Psychol Psychiatry Allied Discip. 2012; 53:726– 734.
- 17. Biederman J, et al. Influence of Gender on Attention Deficit Hyperactivity Disorder in Children Referred to a Psychiatric Clinic. Am J Psychiatry. 2002; 159:36–42. [PubMed: 11772687]
- 18. Catalá-López F, et al. Prevalence of attention deficit hyperactivity disorder among children and adolescents in Spain: a systematic review and meta-analysis of epidemiological studies. BMC Psychiatry. 2012; 12:168. [PubMed: 23057832]
- 19. Willcutt EG. The Prevalence of DSM-IV Attention-Deficit/Hyperactivity Disorder: A Meta-Analytic Review. Neurotherapeutics. 2012; 9:490–499. [PubMed: 22976615]

- 20. Nussbaum NL. ADHD and Female Specific Concerns: A Review of the Literature and Clinical Implications. J Atten Disord. 2012; 16:87–100. [PubMed: 21976033]
- 21. Simon V, Czobor P, Bálint S. Mészáros, Á. & Bitter, I. Prevalence and correlates of adult attentiondeficit hyperactivity disorder: meta-analysis. Br J Psychiatry. 2009; 194:204–211. [PubMed: 19252145]
- 22. Rucklidge JJ. Gender Differences in Attention-Deficit/Hyperactivity Disorder. Psychiatr Clin North Am. 2010; 33:357–373. [PubMed: 20385342]
- 23. Hinshaw SP, Owens EB, Sami N, Fargeon S. Prospective follow-up of girls with attention-deficit/ hyperactivity disorder into adolescence: Evidence for continuing cross-domain impairment. J Consult Clin Psychol. 2006; 74:489. [PubMed: 16822106]
- 24. O'Brien JW, Dowell LR, Mostofsky SH, Denckla MB, Mahone EM. Neuropsychological Profile of Executive Function in Girls with Attention-Deficit/Hyperactivity Disorder. Arch Clin Neuropsychol. 2010; 25:656–670. [PubMed: 20639299]
- 25. Phoenix CH, Goy RW, Gerall AA, Young WC. Organizing action of prenatally administered testosterone propionate on the tissues mediating mating behavior in the female guinea pig. Endocrinology. 1959; 504:369–382.
- 26. Arnold AP. The organizational-activational hypothesis as the foundation for a unified theory of sexual differentiation of all mammalian tissues. Horm Behav. 2009; 55:570–578. [PubMed: 19446073]
- 27. McCarthy MM, Wright CL, Schwarz JM. Hormones and Behavior New tricks by an old dogma : Mechanisms of the Organizational / Activational Hypothesis of steroid-mediated sexual differentiation of brain and behavior. Horm Behav. 2009; 55:655–665. [PubMed: 19682425]
- 28. Wu MV, Shah NM. Control of masculinization of the brain and behavior. Curr Opin Neurobiol. 2011; 21:116–23. [PubMed: 20970320]
- 29. Simerly RB. WIRED FOR REPRODUCTION: Organization and Development of Sexually Dimorphic Circuits in the Mammalian Forebrain. Annu Rev Neurosci. 2002; 25:507–536. [PubMed: 12052919]
- 30. Motelica-Heino I, Castanier M, Corbier P, Edwards DA, Roffi J. Testosterone levels in plasma and testes of neonatal mice. J Steroid Biochem. 1988; 31:283–286. [PubMed: 3419158]
- 31. Corbier P, Edwards DA, Roffi J. The neonatal testosterone surge: a comparative study. Arch Int Physiol Biochim Biophys. 1992; 100:127–31. [PubMed: 1379488]
- 32. MacLusky NJ, Naftolin F. Sexual Differentiation of the Central Nervous System. Science (80-). 1981; 211:1294–1303.
- 33. Amateau SK, Alt JJ, Stamps CL, McCarthy MM. Brain estradiol content in newborn rats: Sex differences, regional heterogeneity, and possible de novo synthesis by the female telencephalon. Endocrinology. 2004; 145:2906–2917. [PubMed: 14988386]
- 34. McCarthy MM. Estradiol and the developing brain. Physiol Rev. 2008; 88:91–124. [PubMed: 18195084]
- 35. Honda S, Harada N, Ito S, Takagi Y, Maeda S. Disruption of sexual behavior in male aromatasedeficient mice lacking exons 1 and 2 of the cyp19 gene. Biochem Biophys Res Commun. 1998; 252:445–449. [PubMed: 9826549]
- 36. Rissman EF, Wersinger SR, Fugger HN, Foster TC. Sex with knockout models: Behavioral studies of estrogen receptor. Brain Res. 1999; 835:80–90. [PubMed: 10448199]
- 37. Wu MV, et al. Estrogen Masculinizes Neural Pathways and Sex-Specific Behaviors. Cell. 2009; 139:61–72. [PubMed: 19804754]
- 38. Gerall AA, Hendricks SE, Johnson LL, Bounds TW. Effects of Early Castration in Male Rats on Adult Sexual Behavior. J Comp Physiol Psychol. 1967; 64:206–212. [PubMed: 6050565]
- 39. Motelica-Heino I, Edwards DA, Roffi J. Intermale aggression in mice: Does hour of castration after birth influence adult behavior? Physiol Behav. 1993; 53:1017–1019. [PubMed: 8511195]
- 40. Toda K, Saibara T, Okada T, Onishi S, Shizuta Y. A loss of aggressive behaviour and its reinstatement by oestrogen in mice lacking the aromatase gene (Cyp19). J Endocrinol. 2001; 168:217–220. [PubMed: 11182758]

- 41. Södersten P. Mounting behavior and lordosis behavior in castrated male rats treated with testosterone propionate, or with estradiol benzoate or dihydrotestosterone in combination with testosterone propionate. Horm Behav. 1975; 6:109–126. [PubMed: 1221003]
- 42. Kimura T, Hagiwara Y. Regulation of urine marking in male and female mice: effects of sex steroids. Horm Behav. 1985; 19:64–70. [PubMed: 4038966]
- 43. Cross E, Roselli CE. 17beta -estradiol rapidly facilitates chemoinvestigation and mounting in castrated male rats. Am J Physiol Regul Integr Comp Physiol. 1999; 276:1346–1350.
- 44. Bakker J, Honda S, Harada N, Balthazart J. Restoration of male sexual behavior by adult exogenous estrogens in male aromatase knockout mice. Horm Behav. 2004; 46:1–10. [PubMed: 15215036]
- 45. Simerly RB. Wired for reproduction: organization and development of sexually dimorphic circuits in the mammalian forebrain. Annu Rev Neurosci. 2002; 25:507–36. [PubMed: 12052919]
- 46. Yang CF, Shah NM. Representing sex in the brain, one module at a time. Neuron. 2014; 82:261– 278. [PubMed: 24742456]
- 47. Bao AM, Swaab DF. Sexual differentiation of the human brain: relation to gender identity, sexual orientation and neuropsychiatric disorders. Front Neuroendocrinol. 2011; 32:214–26. [PubMed: 21334362]
- 48. Forger NG. Control of cell number in the sexually dimorphic brain and spinal cord. J Neuroendocrinol. 2009; 21:393–399. [PubMed: 19207822]
- 49. Armoskus C, et al. Identification of sexually dimorphic genes in the neonatal mouse cortex and hippocampus. Brain Res. 2014; 1562:23–38. [PubMed: 24661915]
- 50. Duclot F, Kabbaj M. The estrous cycle surpasses sex differences in regulating the transcriptome in the rat medial prefrontal cortex and reveals an underlying role of early growth response 1. Genome Biol. 2015; 16:1–20. [PubMed: 25583448]
- 51. Werling DM, Parikshak NN, Geschwind DH. Gene expression in human brain implicates sexually dimorphic pathways in autism spectrum disorders. Nat Commun. 2016; 7:10717. [PubMed: 26892004]
- 52. Dewing P, Shi T, Horvath S, Vilain E. Sexually dimorphic gene expression in mouse brain precedes gonadal differentiation. Mol Brain Res. 2003; 118:82–90. [PubMed: 14559357]
- 53. Kang HJ, et al. Spatio-temporal transcriptome of the human brain. Nature. 2011; 478:483–9. [PubMed: 22031440]
- 54. Xu X, et al. Modular genetic control of sexually dimorphic behaviors. Cell. 2012; 148:596–607. [PubMed: 22304924]
- 55. Hodes GE, et al. Sex Differences in Nucleus Accumbens Transcriptome Profiles Associated with Susceptibility versus Resilience to Subchronic Variable Stress. 2015; 35:16362–16376.
- 56. Walker DM, Kirson D, Perez LF, Gore AC. Molecular Profiling of Postnatal Development of the Hypothalamus in Female and Male Rats1. Biol Reprod. 2012; 87:1–12.
- 57. Zuloaga DG, Puts DA, Jordan CL, Breedlove SM. The role of androgen receptors in the masculinization of brain and behavior: What we've learned from the testicular feminization mutation. Horm Behav. 2008; 53:613–626. [PubMed: 18374335]
- 58. Juntti SA, et al. The androgen receptor governs the execution, but not programming, of male sexual and territorial behaviors. Neuron. 2010; 66:260–272. [PubMed: 20435002]
- 59. Handa RJ, Mani SK, Uht RM. Estrogen receptors and the regulation of neural stress responses. Neuroendocrinology. 2012; 96:111–118. [PubMed: 22538291]
- 60. Walf AA, Frye CA. A Review and Update of Mechanisms of Estrogen in the Hippocampus and Amygdala for Anxiety and Depression Behavior. Neuropsychopharmacology. 2006; 31:1097. [PubMed: 16554740]
- 61. Grumbach MM, Auchus RJ. Estrogen: Consequences and implications of human mutations in synthesis and action. J Clin Endocrinol Metab. 1999; 84:4677–4694. [PubMed: 10599737]
- 62. Van Hemmen J, et al. Sex Differences in White Matter Microstructure in the Human Brain Predominantly Reflect Differences in Sex Hormone Exposure. Cereb Cortex. 2017; 27:2994–3001. [PubMed: 27226438]

- 63. Reyes FI, Boroditsky RS, Winter JS, Faiman C. Studies on human sexual development. II. Fetal and maternal serum gonadotropin and sex steroid concentrations. J Clin Endocrinol Metab. 1974; 38:612–7. [PubMed: 4856555]
- 64. Hines M. Prenatal testosterone and gender-related behaviour. Eur J Endocrinol. 2006; 155(Suppl):S115–S121. [PubMed: 17074984]
- 65. Workman AD, Charvet CJ, Clancy B, Darlington RB, Finlay BL. Modeling transformations of neurodevelopmental sequences across mammalian species. J Neurosci. 2013; 33:7368–83. [PubMed: 23616543]
- 66. Willsey AJ, et al. Coexpression networks implicate human midfetal deep cortical projection neurons in the pathogenesis of autism. Cell. 2013; 155:997–1007. [PubMed: 24267886]
- 67. Hrabovszky Z, Hutson JM. Androgen imprinting of the brain in animal models and humans with intersex disorders: review and recommendations. J Urol. 2002; 168:2142–8. [PubMed: 12394744]
- 68. Winter JSD, Hughes IA, Reyes FI, Faiman C. Pituitary-gonadal relations in infancy: 2. Patterns of serum gonadal steroid concentrations in man from birth to two years of age. J Clin Endocrinol Metab. 1976; 42:679–686. [PubMed: 1262442]
- 69. Chellakooty M, et al. Inhibin A, inhibin B, follicle-stimulating hormone, luteinizing hormone, estradiol, and sex hormone-binding globulin levels in 473 healthy infant girls. J Clin Endocrinol Metab. 2003; 88:3515–3520. [PubMed: 12915629]
- 70. Thompson AL, Whitten PL, Johnson ML, Lampl M. Non-invasive methods for estradiol recovery from infant fecal samples. Front Physiol. Nov 1.2010 :1–8. [PubMed: 21522484]
- 71. Lombardo MV, et al. Fetal testosterone influences sexually dimorphic gray matter in the human brain. J Neurosci Off J Soc Neurosci. 2012; 32:674–680.
- 72. Li G, et al. Mapping longitudinal development of local cortical gyrification in infants from birth to 2 years of age. J Neurosci Off J Soc Neurosci. 2014; 34:4228–4238.
- 73. Arnold AP, et al. The importance of having two X chromosomes. Philos Trans R Soc Lond B Biol Sci. 2016; 371:20150113. [PubMed: 26833834]
- 74. Cox KH, Bonthuis PJ, Rissman EF. Mouse model systems to study sex chromosome genes and behavior: Relevance to humans. Front Neuroendocrinol. 2014; 35:405–419. [PubMed: 24388960]
- 75. Arnold AP. Sex chromosomes and brain gender. Nat Rev Neurosci. 2004; 5:701–8. [PubMed: 15322528]
- 76. Bramble MS, Lipson A, Vashist N, Vilain E. Effects of chromosomal sex and hormonal influences on shaping sex differences in brain and behavior: Lessons from cases of disorders of sex development. J Neurosci Res. 2017; 95:65–74. [PubMed: 27841933]
- 77. Lenroot RK, Lee NR, Giedd JN. Effects of sex chromosome aneuploidies on brain development: Evidence from neuroimaging studies. Dev Disabil Res Rev. 2009; 15:318–327. [PubMed: 20014372]
- 78. Hong DS, Reiss AL. Cognitive and neurological aspects of sex chromosome aneuploidies. Lancet Neurol. 2014; 13:306–318. [PubMed: 24556008]
- 79. Crespi B. Turner syndrome and the evolution of human sexual dimorphism. Evol Appl. 2008; 1:449–61. [PubMed: 25567727]
- 80. Lin A, et al. Mapping the Stability of Human Brain Asymmetry across Five Sex-Chromosome Aneuploidies. J Neurosci. 2015; 35:140–145. [PubMed: 25568109]
- 81. Reardon PK, et al. An Allometric Analysis of Sex and Sex Chromosome Dosage Effects on Subcortical Anatomy in Humans. J Neurosci. 2016; 36:2438–2448. [PubMed: 26911691]
- 82. Raznahan A, et al. Triangulating the sexually dimorphic brain through high-resolution neuroimaging of murine sex chromosome aneuploidies. Brain Struct Funct. 2015; 220:3581–3593. [PubMed: 25146308]
- 83. De Vries GJ, et al. A model system for study of sex chromosome effects on sexually dimorphic neural and behavioral traits. J Neurosci. 2002; 22:9005–14. [PubMed: 12388607]
- 84. Hughes JF, Page DC. The Biology Evolution of Mammalian Y Chromosomes. Annu Rev Genet. 2015; 49 annurev-genet-112414-055311.
- 85. Shimogori T, et al. A genomic atlas of mouse hypothalamic development. Nat Neurosci. 2010; 13:767–75. [PubMed: 20436479]

- 86. Wolstenholme JT, Rissman EF, Bekiranov S. Sexual differentiation in the developing mouse brain: Contributions of sex chromosome genes. Genes, Brain Behav. 2013; 12:166–180. [PubMed: 23210685]
- 87. Xu J, Burgoyne PS, Arnold AP. Sex differences in sex chromosome gene expression in mouse brain. 2002; 11:1409–1419.
- 88. Vicoso B, Charlesworth B. Evolution on the X chromosome: unusual patterns and processes. Nat Rev Genet. 2006; 7:645–653. [PubMed: 16847464]
- 89. Ropers HH, Hamel BCJ. X-linked mental retardation. Nat Rev Genet. 2005; 6:46–57. [PubMed: 15630421]
- 90. Nguyen DK, Disteche CM. Dosage compensation of the active X chromosome in mammals. Nat Genet. 2006; 38:47–53. [PubMed: 16341221]
- 91. Nguyen DK, Disteche CM. High expression of the mammalian X chromosome in brain. Brain Res. 2006; 1126:46–9. [PubMed: 16978591]
- 92. Vallender EJ, Lahn BT. How mammalian sex chromosomes acquired their peculiar gene content. BioEssays. 2004; 26:159–169. [PubMed: 14745834]
- 93. Skuse DH. X-linked genes and mental functioning. Hum Mol Genet. 2005; 14:27–32.
- 94. Zechner U, et al. A high density of X-linked genes for general cognitive ability: A run-away process shaping human evolution? Trends Genet. 2001; 17:697–701. [PubMed: 11718922]
- 95. Disteche CM. Dosage compensation of the sex chromosomes and autosomes. Semin Cell Dev Biol. 2016; 56:9–18. [PubMed: 27112542]
- 96. Jeon Y, Sarma K, Lee JT. New and Xisting regulatory mechanisms of X chromosome inactivation. Curr Opin Genet Dev. 2012; 22:62–71. [PubMed: 22424802]
- 97. Tukiainen T, et al. Landscape of X chromsome inactivation across human tissues. Nat Publ Gr. 2017; 550:244–248.
- 98. Wu H, et al. Cellular Resolution Maps of X Chromosome Inactivation: Implications for Neural Development, Function, and Disease. Neuron. 2014; 81:103–119. [PubMed: 24411735]
- 99. Xu J, Andreassi M. Reversible histone methylation regulates brain gene expression and behavior. Horm Behav. 2011; 59:383–392. [PubMed: 20816965]
- 100. Iwase S, et al. A Mouse Model of X-linked Intellectual Disability Associated with Impaired Removal of Histone Methylation. Cell Rep. 2016; 14:1000–1009. [PubMed: 26804915]
- 101. Scandaglia M, et al. Loss of Kdm5c Causes Spurious Transcription and Prevents the Fine-Tuning of Activity-Regulated Enhancers in Neurons. Cell Rep. 2017; 21:47–59. [PubMed: 28978483]
- 102. Bonasio R, Tu S, Reinberg D. Molecular Signals of Epigenetic States. Science (80-). 2010; 330:612–616.
- 103. Ptashne M. Epigenetics: core misconcept. Proc Natl Acad Sci U S A. 2013; 110:7101–3. [PubMed: 23584020]
- 104. Sweatt JD. The emerging field of neuroepigenetics. Neuron. 2013; 80:624–632. [PubMed: 24183015]
- 105. Deans C, Maggert KA. What do you mean, ? ??Epigenetic???? Genetics. 2015; 199:887–896. [PubMed: 25855649]
- 106. Cholewa-Waclaw J, et al. The Role of Epigenetic Mechanisms in the Regulation of Gene Expression in the Nervous System. J Neurosci. 2016; 36:11427–11434. [PubMed: 27911745]
- 107. Yao B, et al. Epigenetic mechanisms in neurogenesis. Nat Rev Neurosci. 2016; 17:537–49. [PubMed: 27334043]
- 108. Alberini CM, Kandel ER. The regulation of transcription in memory consolidation. Cold Spring Harb Perspect Biol. 2014; 7:a021741. [PubMed: 25475090]
- 109. Riccio A. Dynamic epigenetic regulation in neurons: enzymes, stimuli and signaling pathways. Nat Neurosci. 2010; 13:1330–1337. [PubMed: 20975757]
- 110. Lomvardas S, Maniatis T. Histone and DNA modifications as regulators of neuronal development and function. Cold Spring Harb Perspect Biol. 2016; 8
- 111. Vuong CK, Black DL, Zheng S. The neurogenetics of alternative splicing. Nat Rev Neurosci. 2016; 17:265–81. [PubMed: 27094079]
- 112. Felsenfeld G. Chromatin. Nature. 1978; 271:115–22. [PubMed: 340956]

- 113. Thomas JO, Thompson RJ. Variation in chromatin structure in two cell types from the same tissue: a short DNA repeat length in cerebral cortex neurons. Cell. 1977; 10:633–640. [PubMed: 862024]
- 114. Ermini M, Kuenzle CC. The chromatin repeat length of cortical neurons shortens during early postnatal development. FEBS Lett. 1978; 90:167–172. [PubMed: 658434]
- 115. Whatley, Sa, Hall, C., Lim, L. Chromatin organization in the rat hypothalamus during early development. Biochem J. 1981; 196:115–9. [PubMed: 7306065]
- 116. Wjaeger A, Kuenzle CC. The chromatin repeat length of brain cortex and cerebellar neurons changes concomitant with terminal differentiation. EMBO J. 1982; 1:811–816. [PubMed: 7188362]
- 117. Rajarajan P, Gil SE, Brennand KJ, Akbarian S. Spatial genome organization and cognition. Nat Rev Neurosci. 2016; 17:681–691. [PubMed: 27708356]
- 118. Watson LA, Tsai LH. In the loop: how chromatin topology links genome structure to function in mechanisms underlying learning and memory. Curr Opin Neurobiol. 2016; 43:48–55. [PubMed: 28024185]
- 119. Ciernia AV, LaSalle J. The landscape of DNA methylation amid a perfect storm of autism aetiologies. Nat Rev Neurosci. 2016; 17:411–423. [PubMed: 27150399]
- 120. Alexander JM, Lomvardas S. Nuclear architecture as an epigenetic regulator of neural development and function. Neuroscience. 2014; 264:39–50. [PubMed: 24486963]
- 121. Pohodich AE, Zoghbi HY. Rett syndrome: disruption of epigenetic control of postnatal neurological functions. Hum Mol Genet. 2015; 24:R10–16. [PubMed: 26060191]
- 122. Bonev B, et al. Multiscale 3D Genome Rewiring during Mouse Neural Development. Cell. 2017; 171:557.e1–557.e24. [PubMed: 29053968]
- 123. Poulin J, Tasic B, Hjerling-leffler J, Trimarchi JM, Awatramani R. Disentangling neural cell diversity using single-cell transcriptomics. 2016; 19
- 124. Harbom LJ, Chronister WD, Mcconnell MJ. Single neuron transcriptome analysis can reveal more than cell type classification. BioEssays. 2016; 38:157–161. [PubMed: 26749010]
- 125. Johnson MB, Walsh CA. Cerebral cortical neuron diversity and development at single-cell resolution. Curr Opin Neurobiol. 2017; 42:9–16. [PubMed: 27888678]
- 126. Zeng H, Sanes JR. Neuronal cell-type classification: challenges, opportunities and the path forward. Nat Rev Neurosci. 2017; 18:530–546. [PubMed: 28775344]
- 127. Paul A, Crow M, Raudales R, Gillis J, Huang ZJ. Transcriptional Architecture of Synaptic Communication Delineates Cortical GABAergic Neuron Identity. Cell. 2017; 171:1–18. [PubMed: 28938111]
- 128. Mo A, et al. Epigenomic Signatures of Neuronal Diversity in the Mammalian Brain. Neuron. 2015; 86:1369–1384. [PubMed: 26087164]
- 129. Nord AS, Pattabiraman K, Visel A, Rubenstein JLR. Genomic Perspectives of Transcriptional Regulation in Forebrain Development. Neuron. 2015; 85:27–47. [PubMed: 25569346]
- 130. Gray LT, et al. Layer-specific chromatin accessibility landscapes reveal regulatory networks in adult mouse visual cortex. Elife. 2017; :1–30. DOI: 10.7554/eLife.21883
- 131. Buenrostro JD, et al. Single-cell chromatin accessibility reveals principles of regulatory variation. Nature. 2015; 523:486–90. [PubMed: 26083756]
- 132. West AE, Griffith EC, Greenberg ME. Regulation of transcription factors by neuronal activity. Nat Rev Neurosci. 2002; 3:921–931. [PubMed: 12461549]
- 133. Hensch TK. Critical period plasticity in local cortical circuits. Nat Rev Neurosci. 2005; 6:877–88. [PubMed: 16261181]
- 134. Ebert DH, Greenberg ME. Activity-dependent neuronal signalling and autism spectrum disorder. Nature. 2013; 493:327–37. [PubMed: 23325215]
- 135. Bourgeron T. From the genetic architecture to synaptic plasticity in autism spectrum disorder. Nat Rev Neurosci. 2015; 16:551–563. [PubMed: 26289574]
- 136. Herbst WA, Martin KC. Regulated transport of signaling proteins from synapse to nucleus. Curr Opin Neurobiol. 2017; 45:78–84. [PubMed: 28502891]

- 137. Gagnidze K, Weil ZM, Pfaff DW. Histone modifications proposed to regulate sexual differentiation of brain and behavior. Bioessays. 2010; 32:932–9. [PubMed: 20836091]
- 138. Forger NG. Epigenetic mechanisms in sexual differentiation of the brain and behaviour. Philos Trans R Soc London B Biol Sci. 2016; 371
- 139. McCarthy MM, et al. The epigenetics of sex differences in the brain. J Neurosci. 2009; 29:12815– 23. [PubMed: 19828794]
- 140. Nugent BM, McCarthy MM. Epigenetic underpinnings of developmental sex differences in the brain. Neuroendocrinology. 2011; 93:150–8. [PubMed: 21411982]
- 141. Ratnu VS, Emami MR, Bredy TW. Genetic and epigenetic factors underlying sex differences in the regulation of gene expression in the brain. J Neurosci Res. 2017; 95:301–310. [PubMed: 27870402]
- 142. Yamamoto KR. Steroid Receptor Regulated Transcription of Specific Genes and Gene Networks. Annu Rev Genet. 1985; 19:209–252. [PubMed: 3909942]
- 143. Mangelsdorf DJ, et al. The nuclear receptor superfamily: the second decade. Cell. 1995; 83:835– 839. [PubMed: 8521507]
- 144. Kuiper GGJM, et al. Comparison of the Ligand Binding Site Specificity and Transcript Tissue Distribution of Estrogen Receptors a and b. Endocrinology. 1997; 138:863–870. [PubMed: 9048584]
- 145. Pettersson K, Grandien K, Kuiper GG, Gustafsson JA. Mouse estrogen receptor beta forms estrogen response element-binding heterodimers with estrogen receptor alpha. Mol Endocrinol. 1997; 11:1486–1496. [PubMed: 9280064]
- 146. McKenna NJ, O'Malley BW. Combinatorial control of gene expression by nuclear receptors and coregulators. Cell. 2002; 108:465–474. [PubMed: 11909518]
- 147. Perissi V, Jepsen K, Glass CK, Rosenfeld MG. Deconstructing repression: evolving models of corepressor action. Nat Rev Genet. 2010; 11:109–123. [PubMed: 20084085]
- 148. Lan X, Glass CK, Rosenfeld MG. Coactivator and corepressor complexes in nuclear receptor function. Curr Opin Genet Dev. 1999; 9:140–147. [PubMed: 10322133]
- 149. Ebert DH, et al. Activity-dependent phosphorylation of MeCP2 threonine 308 regulates interaction with NCoR. Nature. 2013; 499:341–345. [PubMed: 23770587]
- 150. Wang T, et al. De novo genic mutations among a Chinese autism spectrum disorder cohort. Nat Commun. 2016; 7:13316. [PubMed: 27824329]
- 151. Mahfouz A, et al. Genome-wide coexpression of steroid receptors in the mouse brain: Identifying signaling pathways and functionally coordinated regions. Proc Natl Acad Sci U S A. 2016; 113:2738–43. [PubMed: 26811448]
- 152. Micevych PE, Kelly MJ. Membrane estrogen receptor regulation of hypothalamic function. Neuroendocrinology. 2012; 96:103–110. [PubMed: 22538318]
- 153. Heimovics SA, Trainor BC, Soma KK. Rapid Effects of Estradiol on Aggression in Birds and Mice: The Fast and the Furious. Integr Comp Biol. 2015; 55:281–293. [PubMed: 25980562]
- 154. Vasudevan N, Pfaff DW. Non-genomic actions of estrogens and their interaction with genomic actions in the brain. Front Neuroendocrinol. 2008; 29:238–257. [PubMed: 18083219]
- 155. Björnström L, Sjöberg M. Mechanisms of estrogen receptor signaling: convergence of genomic and nongenomic actions on target genes. Mol Endocrinol. 2005; 19:833–842. [PubMed: 15695368]
- 156. Murphy DD, Segal M. Morphological plasticity of dendritic spines in central neurons is mediated by activation of cAMP response element binding protein. Proc Natl Acad Sci U S A. 1997; 94:1482–7. [PubMed: 9037079]
- 157. Abrahám IM, Herbison AE. Major sex differences in non-genomic estrogen actions on intracellular signaling in mouse brain in vivo. Neuroscience. 2005; 131:945–51. [PubMed: 15749347]
- 158. Kushner PJ, et al. Estrogen receptor pathways to AP-1. J Steroid Biochem Mol Biol. 2000; 74:311–317. [PubMed: 11162939]
- 159. Heldring N, et al. Multiple sequence-specific DNA-binding proteins mediate estrogen receptor signaling through a tethering pathway. Mol Endocrinol. 2011; 25:564–574. [PubMed: 21330404]

- 160. Stolzenberg DS, Rissman EF. Oestrogen-Independent, Experience-Induced Maternal Behaviour in Female Mice. J Neuroendocrinol. 2011; 23:345–354. [PubMed: 21276101]
- 161. Banerjee SB, Liu RC. Storing maternal memories: Hypothesizing an interaction of experience and estrogen on sensory cortical plasticity to learn infant cues. Front Neuroendocrinol. 2013; 34:300– 314. [PubMed: 23916405]
- 162. Rosenblatt JS. Nonhormonal basis of maternal behavior in the rat. Science (80-). 1967; 156:1512– 1513.
- 163. Shughrue PJ, Lane MV, Merchenthaler I. Comparative distribution of estrogen receptor-alpha and -beta mRNA in the rat central nervous system. J Comp Neurol. 1997; 388:507–525. [PubMed: 9388012]
- 164. Mitra SW, et al. Immunolocalization of estrogen receptor beta in the mouse brain: Comparison with estrogen receptor alpha. Endocrinology. 2003; 144:2055–2067. [PubMed: 12697714]
- 165. Yang CF, et al. Sexually dimorphic neurons in the ventromedial hypothalamus govern mating in both sexes and aggression in males. Cell. 2013; 153:896–909. [PubMed: 23663785]
- 166. Shah NM, et al. Visualizing sexual dimorphism in the brain. Neuron. 2004; 43:313–319. [PubMed: 15294140]
- 167. Quadros PS, Pfau JL, Wagner CK. Distribution of Progesterone Receptor Immunoreactivity in the Fetal and Neonatal Rat Forebrain. J Comp Neurol. 2007; 504:42–56. [PubMed: 17614295]
- 168. McAbee MD, DonCarlos LL. Regulation of Androgen Receptor Messenger Ribonucleic Acid Expression in the Developing Rat Forebrain *. Endocrinology. 1999; 140:1807–14. [PubMed: 10098519]
- 169. Andermann ML, Lowell BB. Toward a Wiring Diagram Understanding of Appetite Control. Neuron. 2017; 95:757–778. [PubMed: 28817798]
- 170. Creutz LM, Kritzer MF. Mesostriatal and mesolimbic projections of midbrain neurons immunoreactive for estrogen receptor beta or androgen receptors in rats. J Comp Neurol. 2004; 476:348–362. [PubMed: 15282710]
- 171. Quesada A, Romeo HE, Micevych P. Distribution and localization patterns of estrogen receptorbeta and insulin-like growth factor-1 receptors in neurons and glial cells of the female rat substantia nigra: localization of ERbeta and IGF-1R in substantia nigra. J Comp Neurol. 2007; 503:198–208. [PubMed: 17480015]
- 172. Purves-Tyson TD, et al. Testosterone regulation of sex steroid-related mRNAs and dopaminerelated mRNAs in adolescent male rat substantia nigra. BMC Neurosci. 2012; 13:95. [PubMed: 22867132]
- 173. Quadros PS, Schlueter LJ, Wagner CK. Distribution of progesterone receptor immunoreactivity in the midbrain and hindbrain of postnatal rats. Dev Neurobiol. 2008; 68:1378–1390. [PubMed: 18712784]
- 174. Goel N, Bale TL. Examining the intersection of sex and stress in modelling neuropsychiatric disorders. J Neuroendocrinol. 2009; 21:415–420. [PubMed: 19187468]
- 175. Nuñez JL, Huppenbauer CB, McAbee MD, Juraska JM, DonCarlos LL. Androgen receptor expression in the developing male and female rat visual and prefrontal cortex. J Neurobiol. 2003; 56:293–302. [PubMed: 12884268]
- 176. Kruijver FPM, Balesar R, Espila AM, Unmehopa UA, Swaab DF. Estrogen-receptor-beta distribution in the human hypothalamus: Similarities and differences with ER-alpha distribution. J Comp Neurol. 2003; 466:251–277. [PubMed: 14528452]
- 177. Kruijver FPM, Balesar R, Espila AM, Unmehopa UA, Swaab DF. Estrogen receptor alpha distribution in the human hypothalamus in relation to sex and endocrine status. J Comp Neurol. 2002; 454:115–139. [PubMed: 12412138]
- 178. Saijo K, Collier JG, Li AC, Katzenellenbogen Ja, Glass CK. An ADIOL-ERbeta-CtBP transrepression pathway negatively regulates microglia-mediated inflammation. Cell. 2011; 145:584–595. [PubMed: 21565615]
- 179. Österlund MK, Keller E, Hurd YL. The human forebrain has discrete estrogen receptor alpha messenger RNA expression: high levels in the amygdaloid complex. Neuroscience. 2000; 95:333–342. [PubMed: 10658612]

- 180. Österlund MK, Gustafsson JA, Keller E, Hurd YL. Estrogen receptor beta (ER beta) messenger ribonucleic acid (mRNA) expression within the human forebrain: Distinct distribution pattern to ER alpha mRNA. J Clin Endocrinol Metab. 2000; 85:3840–3846. [PubMed: 11061547]
- 181. Fernández-Guasti A, Kruijver FP, Fodor M, Swaab DF. Sex differences in the distribution of androgen receptors in the human hypothalmus. J Comp Neurol. 2000; 435:422–435.
- 182. Nguyen TV, Ducharme S, Karama S. Effects of Sex Steroids in the Human Brain. Mol Neurobiol. 2016; :1–13. DOI: 10.1007/s12035-016-0198-3 [PubMed: 25394384]
- 183. Yang CF, Shah NM. Representing sex in the brain, one module at a time. Neuron. 2014; 82:261– 78. [PubMed: 24742456]
- 184. Davis EP, Pfaff D. Sexually dimorphic responses to early adversity: Implications for affective problems and autism spectrum disorder. Psychoneuroendocrinology. 2014; 49:11–25. [PubMed: 25038479]
- 185. Vidal-Ribas P, Brotman MA, Valdivieso I, Leibenluft E, Stringaris A. The Status of Irritability in Psychiatry: A Conceptual and Quantitative Review. J Am Acad Child Adolesc Psychiatry. 2016; 55:556–570. [PubMed: 27343883]
- 186. Hammerslag LR, Gulley JM. Sex differences in behavior and neural development and their role in adolescent vulnerability to substance use. Behav Brain Res. 2016; 298:15–26. [PubMed: 25882721]
- 187. Lorenzetti V, Allen NB, Fornito A, Yücel M. Structural brain abnormalities in major depressive disorder: A selective review of recent MRI studies. Journal of Affective Disorders. 2009; 117:1– 17. [PubMed: 19237202]
- 188. Yohn CN, Gergues MM, Samuels BA. The role of 5-HT receptors in depression. Mol Brain. 2017; 10:28. [PubMed: 28646910]
- 189. Ionescu DF, Papakostas GI. Experimental medication treatment approaches for depression. Transl Psychiatry. 2017; 7:e1068. [PubMed: 28323287]
- 190. Marrocco J, Mcewen BS. Sex in the brain: hormones and sex differences. Dialogues Clin Neurosci. 2016:373–383. [PubMed: 28179809]
- 191. Schibli K, Wong K, Hedayati N, D'Angiulli A. Attending, learning, and socioeconomic disadvantage: developmental cognitive and social neuroscience of resilience and vulnerability. Ann N Y Acad Sci. 2017; 1396:19–38. [PubMed: 28548461]
- 192. Slifstein M, et al. Deficits in Prefrontal Cortical and Extrastriatal Dopamine Release in Schizophrenia. JAMA Psychiatry. 2015; 72:316. [PubMed: 25651194]
- 193. Grissom N, et al. Male-specific deficits in natural reward learning in a mouse model of neurodevelopmental disorders. Mol Psychiatry. 2017
- 194. Abel KM, Drake R, Goldstein JM. Sex differences in schizophrenia. Int Rev Psychiatry. 2010; 22:417–428. [PubMed: 21047156]
- 195. Crawford MB, DeLisi LE. Issues related to sex differences in antipsychotic treatment. Curr Opin Psychiatry. 2016; 29:211–217. [PubMed: 26906336]
- 196. Fattore L, Melis M, Fadda P, Fratta W. Sex differences in addictive disorders. Front Neuroendocrinol. 2014; 35:272–284. [PubMed: 24769267]
- 197. Jeste SS, Geschwind DH. Disentangling the heterogeneity of autism spectrum disorder through genetic findings. Nat Rev Neurol. 2014; 10:74–81. [PubMed: 24468882]
- 198. Girirajan S, Eichler EE. Phenotypic variability and genetic susceptibility to genomic disorders. Hum Mol Genet. 2010; 19:R176–187. [PubMed: 20807775]
- 199. Sebat J, Levy DL, McCarthy SE. Rare structural variants in schizophrenia: one disorder, multiple mutations; one mutation, multiple disorders. Trends Genet TIG. 2009; 25:528–535. [PubMed: 19883952]
- 200. De Rubeis S, Buxbaum JD. Genetics and genomics of autism spectrum disorder: Embracing complexity. Hum Mol Genet. 2015; 24:R24–R31. [PubMed: 26188008]
- 201. Genovese G, et al. Increased burden of ultra-rare protein-altering variants among 4,877 individuals with schizophrenia. Nat Neurosci. 2016; 19:1433–1441. [PubMed: 27694994]
- 202. Werling DM, et al. Limited contribution of rare, noncoding variation to autism spectrum disorder from sequencing of 2,076 genomes in quartet families. bioRxiv. 2017

- 203. Turner TN, et al. Genomic Patterns of De Novo Mutation in Simplex Resource Genomic Patterns of De Novo Mutation in Simplex Autism. Cell. 2017; :1–13. DOI: 10.1016/j.cell.2017.08.047
- 204. De Rubeis S, et al. Synaptic, transcriptional and chromatin genes disrupted in autism. Nature. 2014; 3
- 205. Akbarian S, et al. The PsychENCODE project. Nat Neurosci. 2015; 18:1707–1712. [PubMed: 26605881]
- 206. Doan RN, et al. Mutations in Human Accelerated Regions Disrupt Cognition and Social Behavior. Cell. 2016; 167:341–354.e12. [PubMed: 27667684]
- 207. Sanders SJ. First glimpses of the neurobiology of autism spectrum disorder. Curr Opin Genet Dev. 2015; 33:80–92. [PubMed: 26547130]
- 208. Mastrototaro G, Zaghi M, Sessa A. Epigenetic Mistakes in Neurodevelopmental Disorders. J Mol Neurosci. 2017; 61:590–602. [PubMed: 28255957]
- 209. Miller, Ja, et al. Transcriptional landscape of the prenatal human brain. Nature. 2014; 508:199– 206. [PubMed: 24695229]
- 210. Reilly SK, et al. Evolutionary changes in promoter and enhancer activity during human corticogenesis. Science (80-). 2015; 347:1155–1159.
- 211. Sun W, et al. Histone Acetylome-wide Association Study of Autism Spectrum Disorder. Cell. 2016; 167:1385–1397.e11. [PubMed: 27863250]
- 212. Voineagu I, et al. Transcriptomic analysis of autistic brain reveals convergent molecular pathology. Nature. 2011; 474:380–384. [PubMed: 21614001]
- 213. Won H, et al. Chromosome conformation elucidates regulatory relationships in developing human brain. Nature. 2016; 538:523–527. [PubMed: 27760116]
- 214. Fromer M, et al. De novo mutations in schizophrenia implicate synaptic networks. Nature. 2014; 506:179–184. [PubMed: 24463507]
- 215. Doherty JL, Owen MJ. Genomic insights into the overlap between psychiatric disorders: implications for research and clinical practice. Genome Med. 2014; 6:29. [PubMed: 24944580]
- 216. St Pourcain B, et al. ASD and schizophrenia show distinct developmental profiles in common genetic overlap with population-based social communication difficulties. Mol Psychiatry. 2017; doi: 10.1038/mp.2016.198
- 217. Tamminga CA. Gender and schizophrenia. J Clin Psychiatry. 1997; 58:33–37.
- 218. Parikshak NN, et al. Genome-wide changes in lncRNA, splicing, and regional gene expression patterns in autism. Nature. 2016; 540:423–427. [PubMed: 27919067]
- 219. Wu YE, Parikshak NN, Belgard TG, Geschwind DH. Genome-wide, integrative analysis implicates microRNA dysregulation in autism spectrum disorder. Nat Neurosci. 2016; 19:1463– 1476. [PubMed: 27571009]
- 220. Walder DJ, et al. Sexual dimorphisms and prediction of conversion in the NAPLS psychosis prodrome. Schizophr Res. 2013; 144:43–50. [PubMed: 23340377]
- 221. Rakic P, Bourgeois J, Eckenhoff M, Zecevic N, Goldman-Rakic P. Concurrent overproduction of synapses in diverse regions of the primate cerebral cortex. Science (80-). 1986; 232:232–235.
- 222. Petanjek Z, et al. Extraordinary neoteny of synaptic spines in the human prefrontal cortex. Proc Natl Acad Sci. 2011; 108:13281–13286. [PubMed: 21788513]
- 223. Giedd JN, et al. Brain development during childhood and adolescence: a longitudinal MRI study. Nat Neurosci. 1999; 2:861–863. [PubMed: 10491603]
- 224. Lee A, et al. Pyramidal neurons in prefrontal cortex receive subtype-specific forms of excitation and inhibition. Neuron. 2014; 81:61–68. [PubMed: 24361076]
- 225. Fries P. Neuronal Gamma-Band Synchronization as a Fundamental Process in Cortical Computation. Annu Rev Neurosci. 2009; 32:209–224. [PubMed: 19400723]
- 226. Notter T, Meyer U. Microglia and schizophrenia: where next? Mol Psychiatry. 2017; 22:788–789. [PubMed: 28348384]
- 227. Nelson LH, Lenz KM. The immune system as a novel regulator of sex differences in brain and behavioral development. J Neurosci Res. 2017; 95:447–461. [PubMed: 27870450]
- 228. Owens SJ, Murphy CE, Purves-Tyson TD, Weickert TW, Weickert CS. Considering the role of adolescent sex steroids in schizophrenia. J Neuroendocrinol. 2017; doi: 10.1111/jne.12538

- 229. Shi L, Zhang Z, Su B. Sex Biased Gene Expression Profiling of Human Brains at Major Developmental Stages. Sci Rep. 2016; 6:21181. [PubMed: 26880485]
- 230. Brandler WM, et al. Paternally inherited noncoding structural variants contribute to autism. bioRxiv. 2017
- 231. Gulsuner S, et al. Spatial and temporal mapping of de novo mutations in schizophrenia to a fetal prefrontal cortical network. Cell. 2013; 154:518–529. [PubMed: 23911319]
- 232. Rizzardi L, et al. Neuronal brain region-specific DNA methylation and chromatin accessibility are associated with neuropsychiatric disease heritability. bioRxiv. 2017
- 233. de Vries GJ, Fields CT, Peters NV, Whylings J, Paul MJ. Sensitive Periods for Hormonal Programming of the Brain. Curr Top Behav Neurosci. 2014; 16:79–108. [PubMed: 24549723]
- 234. Thompson BL, Levitt P. The clinical-basic interface in defining pathogenesis in disorders of neurodevelopmental origin. Neuron. 2010; 67:702–12. [PubMed: 20826303]
- 235. Skene NG, Roy M, Grant SGN. A genomic lifespan program that reorganises the young adult brain is targeted in schizophrenia. Elife. 2017; 6:e17915. [PubMed: 28893375]
- 236. Kessler RC, McGonagle KA, Swartz M, Blazer DG, Nelson CB. Sex and depression in the National Comorbidity Survey. I: Lifetime prevalence, chronicity and recurrence. J Affect Disord. 1993; 29:85–96. [PubMed: 8300981]
- 237. Martin LA, Neighbors HW, Griffith DM. The experience of symptoms of depression in men vs women: analysis of the National Comorbidity Survey Replication. JAMA psychiatry. 2013; 70:1100–1106. [PubMed: 23986338]
- 238. Altemus M, Sarvaiya N, Neill Epperson C. Sex differences in anxiety and depression clinical perspectives. Front Neuroendocrinol. 2014; 35:320–330. [PubMed: 24887405]
- 239. Hodes GE, Walker DM, Labonté B, Nestler EJ, Russo SJ. Understanding the epigenetic basis of sex differences in depression. J Neurosci Res. 2017; 95:692–702. [PubMed: 27870456]
- 240. Bangasser DA, Valentino RJ. Sex differences in stress-related psychiatric disorders: Neurobiological perspectives. Front Neuroendocrinol. 2014; 35:303–319. [PubMed: 24726661]
- 241. Handa RJ, Weiser MJ. Gonadal steroid hormones and the hypothalamo-pituitary-adrenal axis. Front Neuroendocrinol. 2014; 35:197–220. [PubMed: 24246855]
- 242. Binder EB, et al. Association of FKBP5 polymorphisms and childhood abuse with risk of posttraumatic stress disorder symptoms in adults. JAMA. 2008; 299:1291–1305. [PubMed: 18349090]
- 243. Reul JMHM, De Kloet ER. Two receptor systems for corticosterone in rat brain: Microdistribution and differential occupation. Endocrinology. 1985; 117:2505–2511. [PubMed: 2998738]
- 244. de Kloet ER, Joëls M, Holsboer F. Stress and the brain: from adaptation to disease. Nat Rev Neurosci. 2005; 6:463–75. [PubMed: 15891777]
- 245. Mifsud KR, Reul JMHM. Acute stress enhances heterodimerization and binding of corticosteroid receptors at glucocorticoid target genes in the hippocampus. Proc Natl Acad Sci U S A. 2016; 113:201605246.
- 246. Bao A, Meynen G, Swaab DF. The stress system in depression and neurodegeneration : Focus on the human hypothalamus. 2007; 57
- 247. McEwen BS, Nasca C, Gray JD. Stress Effects on Neuronal Structure: Hippocampus, Amygdala, and Prefrontal Cortex. Neuropsychopharmacology. 2015; 41:3–23. [PubMed: 26076834]
- 248. Arora VK, et al. Glucocorticoid receptor confers resistance to antiandrogens by bypassing androgen receptor blockade. Cell. 2013; 155:1309–1322. [PubMed: 24315100]
- 249. Binder EB. The role of FKBP5, a co-chaperone of the glucocorticoid receptor in the pathogenesis and therapy of affective and anxiety disorders. Psychoneuroendocrinology. 2009; 34:186–195.
- 250. Bolton EC, et al. Cell- and gene-specific regulation of primary target genes by the androgen receptor. Genes Dev. 2007; 21:2005–2017. [PubMed: 17699749]
- 251. Magee JA, Chang LW, Stormo GD, Milbrandt J. Direct, androgen receptor-mediated regulation of the FKBP5 gene via a distal enhancer element. Endocrinology. 2006; 147:590–598. [PubMed: 16210365]

- 252. Voss TC, et al. Dynamic exchange at regulatory elements during chromatin remodeling underlies assisted loading mechanism. Cell. 2011; 146:544–554. [PubMed: 21835447]
- 253. Miranda TB, et al. Reprogramming the chromatin landscape: Interplay of the estrogen and glucocorticoid receptors at the genomic level. Cancer Res. 2013; 73:5130–5139. [PubMed: 23803465]
- 254. Swinstead EE, et al. Steroid Receptors Reprogram FoxA1 Occupancy through Dynamic Chromatin Transitions. Cell. 2016; 165:593–605. [PubMed: 27062924]
- 255. McGowan, O Patrick, Sasaki, Aya, D'Alessio, Ana, Cymov, Sergit, Labonte, Benoit, Szyf, Moshe, Turecki, Gustavo, Meaney, JM. Epigenetic regulation of the glucocorticoid receptor in human brain associates with childhood abuse. Nat Neurosci. 2009; 12:342–348. [PubMed: 19234457]
- 256. Labonte B, et al. Differential Glucocorticoid Receptor Exon 1B, 1C, and 1H Expression and Methylation in Suicide Completers with a History of Childhood Abuse. Biol Psychiatry. 2012; 72:41–48. [PubMed: 22444201]
- 257. Uddin M, et al. Epigenetic and immune function profiles associated with posttraumatic stress disorder. Proc Natl Acad Sci U S A. 2010; 107:9470–9475. [PubMed: 20439746]
- 258. Nagy C, et al. Astrocytic abnormalities and global DNA methylation patterns in depression and suicide. Mol Psychiatry. 2015; 20:320–328. [PubMed: 24662927]
- 259. Nishioka M, Bundo M, Kasai K, Iwamoto K. DNA methylation in schizophrenia: progress and challenges of epigenetic studies. Genome Med. 2012; 4:96. [PubMed: 23234572]
- 260. Iwamoto K, et al. Neurons show distinctive DNA methylation profile and higher interindividual variations compared with non-neurons. Genome Res. 2011; 21:688–696. [PubMed: 21467265]

Figure 1.

Intersection of hormone surges with stages of brain development and maturation. Major events in neural development are depicted during human gestation (weeks) and into adulthood (years). Fluctuations in hormone levels intersect with these events in a sexspecific fashion; critical periods for sexual differentiation of the brain are boxed in orange. Male testosterone levels (blue line) begin to rise during the eighth week of gestation as the testes mature, peak around week 16, and decline after week 24. Testosterone levels spike again in infancy with a peak around 1–3 months and then remain flat until the onset of puberty. In contrast, in females, the ovaries are largely inactive during gestation and begin to secrete estradiol (red line) and progesterone during puberty. There is also evidence of estradiol secretion in female infants, but the exact levels and duration are not well described.

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Figure 2.

Gene regulatory strategies for achieving sex-biased gene expression. Sex differences in gene expression can be patterned by differential exposure to gonadal hormones during early life or puberty. Such differences can arise either from epigenetic events where lasting changes in chromatin state are established by transient exposure to hormone or through the induction of a trans-acting factor. (A) During developmental hormone exposure (mid-gestation or perinatal), ligand-bound steroid hormone receptors induce an active chromatin state (green peak) on an enhancer (grey oval), thereby causing a gene to be expressed more highly in males. (B) Alternatively, a chromatin modifier, co-regulator, or transcription factor may be present in one sex or the other; here, this scenario is depicted in females. The effects of early hormone signaling may not become apparent until puberty. (C) Male hormones prime a gene for later expression during adolescence (yellow peak indicates poised enhancer). (D) Cycling hormones in adolescent and adult females regulate gene expression acutely in the presence of estradiol or progesterone. A ligand-bound ERα homodimer is depicted in red.

Figure 3.

Sex-specific function in neural circuits and effects of mutations. Sex-specific differentiation in the nervous system may occur at different points in neural circuits to produce gendertypical behaviors. (A) A mutation may therefore alter expression of a gene that is expressed in a sex-specific manner in a brain region, thereby changing its output to cause phenotypes in only one sex. (B) Alternatively, a mutation may produce different effects in the two sexes non-cell autonomously by affecting the function of a region functionally upstream of sexspecific circuitry to cause distinct phenotypes in each sex. (C) Finally, functional disruption of a downstream behavioral output region that receives sexually dimorphic input can also produce a different effect in the two sexes; here, the mutation changes the response of neurons that receive more input in females compared with males.

Figure 4.

Mechanisms of gene regulation and effects of disease-associated mutations. Genes implicated in cell-specific neuronal function may (A) encode proteins that are expressed in distinct cell types or (B) regulate chromatin structure to control patterns of the expression of many genes in specify distinct cell types . (C) Profound phenotypes may arise from a lossof-function mutation (LoF) in a coding sequence of a broadly expressed gene. (D) Mutations in a chromatin regulatory factor affect gene expression in trans by changing chromatin state at multiple target genes, thereby resulting in dysregulation of gene expression but not complete LoF. (E) Alternatively, a mutation in a regulatory element could alter spatial or temporal aspects of gene expression, effectively causing a LoF in a specific context, such as a distinct cell type. Active chromatin states are depicted in green in wild-type individuals and attenuated to a yellow-green when the chromatin regulator is not functional. (F) Finally, a mutation in an enhancer of a chromatin regulator can lead to aberrant chromatin patterning

in only a subset of neurons or developmental stages, resulting in a comparatively mild phenotype.

Figure 5.

Cross talk between gonadal hormone signaling and stress response. (A) In the classical model of nuclear receptor action, there is no binding of AR to its consensus androgen response elements (ARE) in the absence of ligand. (B) When testosterone is present, AR homodimerizes and binds AREs, but can also bind glucocorticoid response elements (GREs) that are typically responsive to CORT. (C) Conversely, in the presence of CORT, GR can both activate its own GRE targets and bind AREs to activate testosterone-responsive genes. GR can also open (inaccessible chromatin (D) through transient binding to non-consensus

sites, and €recruitment of chromatin-remodeling machinery. (F) This action reveals additional binding sites that can later be bound by ERα tethered to AP-1.