Lawrence Berkeley National Laboratory

LBL Publications

Title

The living interface between synthetic biology and biomaterial design

https://escholarship.org/uc/item/1f9343w2

Journal

Permalink

Nature Materials, 21(4)

ISSN

1476-1122

Authors

Liu, Allen P Appel, Eric A Ashby, Paul D <u>et al.</u>

Publication Date 2022-04-01

DOI 10.1038/s41563-022-01231-3

Copyright Information

This work is made available under the terms of a Creative Commons Attribution-NonCommercial License, available at <u>https://creativecommons.org/licenses/by-nc/4.0/</u>

Peer reviewed

The living interface between synthetic biology and biomaterial design

Allen P. Liu^{*,1}, Eric A. Appel², Paul D. Ashby³, Brendon M. Baker⁴, Elisa Franco⁵, Luo Gu⁶, Karmella Haynes⁷, Neel S. Joshi⁸, April M. Kloxin⁹, Paul H. J. Kouwer¹⁰, Jeetain Mittal¹¹, Leonardo Morsut¹², Vincent Noireaux¹³, Sapun Parekh¹⁴, Rebecca Schulman¹⁵, Sindy K.Y. Tang¹⁶, Megan T. Valentine¹⁷, Sebastián L. Vega¹⁸, Wilfried Weber¹⁹, Nicholas Stephanopoulos^{*,20}, Ovijit Chaudhuri^{*,16}

¹ University of Michigan, Department of Mechanical Engineering, Ann Arbor, MI 48109, USA

² Stanford University, Department of Materials Science & Engineering, Stanford, CA 94305, USA

³ Lawrence Berkeley National Laboratory, Molecular Foundry, Berkeley, CA 94720, USA

⁴ University of Michigan, Department of Biomedical Engineering, Ann Arbor, MI 48109, USA

⁵ University of California – Los Angeles, Department of Mechanical and Aerospace Engineering, Los Angeles, CA 90095, USA

⁶ Johns Hopkins University, Department of Materials Science and Engineering, Baltimore, MD 21218, USA

⁷ Emory University, Wallace H. Coulter Department of Biomedical Engineering, Atlanta, GA 30322, USA

^e Northeastern University, Department of Chemistry and Chemical Biology, Boston, MA 02115, USA

⁹ University of Delaware, Department of Chemical and Biomolecular Engineering and Materials Science and Engineering, Newark, DE 19716, USA

¹⁰ Radboud University, Institute for Molecules and Materials, Nijmegen, The Netherlands

¹¹ Artie McFerrin Department of Chemical Engineering, Texas A&M University, College Station, TX 78743

¹² University of Southern California, Department of Stem Cell Biology and Regenerative Medicine, Los Angeles CA 90033, USA

¹³ University of Minnesota, School of Physics and Astronomy, Minneapolis, MN 55455, USA

¹⁴ University of Texas – Austin, Department of Biomedical Engineering, Austin TX 78712, USA

¹⁵ Johns Hopkins University, Department of Chemical and Biomolecular Engineering, Baltimore, MD 21218, USA

¹⁶ Stanford University, Department of Mechanical Engineering, Stanford, CA 94305, USA

¹⁷ University of California - Santa Barbara, Department of Mechanical Engineering, Santa Barbara, CA 93106, USA

¹⁸ Rowan University, Department of Biomedical Engineering, Glassboro, NJ 08028, USA

¹⁹ University of Freiburg, Faculty of Biology and Signalling Research Centres BIOSS and CIBSS, Freiburg, Germany

²⁰ Arizona State University, School of Molecular Sciences, Tempe, AZ 85287, USA

* A.P.L, N.S., and O.C. are co-corresponding authors. Address correspondence to <u>allenliu@umich.edu</u>, <u>nstepha1@asu.edu</u>, and <u>chaudhuri@stanford.edu</u>

Abstract

Recent far-reaching advances in synthetic biology have yielded exciting tools for the creation of new materials. Conversely, advances in the fundamental understanding of soft-condensed matter, polymers, and biomaterials offer new avenues to extend the reach of synthetic biology. The broad and exciting range of possible applications have significant implications to address grand challenges in health, biotechnology, and sustainability. Despite the potentially transformative impact that lies at the interface of synthetic biology and biomaterials, the two fields have to date progressed mostly separately. This perspective article provides a review of recent key advances in these two fields, and a roadmap for collaboration at the interface between these two communities. We highlight the near-term applications of this interface to the development of hierarchically structured biomaterials, from bioinspired building blocks to "living" materials that sense and respond based on the reciprocal interactions between materials and embedded cells.

The field of biomaterials research has expanded significantly in the past few decades, with a growing interest in the concept of 'programmability'. A key promise of programmable biomaterials is in their use as a synthetic extracellular matrix (ECM) to direct the fate of cells through tunable properties, encoded using chemical approaches. However, in contrast to natural biomaterials like the native ECM—which is highly dynamic, and continuously modified through reciprocal feedback between cells—many current generation biomaterials are static, and transfer information in one direction (i.e. from the material to the cell).

Synthetic biology has long championed programmability as a central feature. Tunability is achieved by introducing genetic modifications in a systematic way via promoters, inducers, and nucleic acid-modifying enzymes. Recent years have also seen an expansion of synthetic biology tools from their origin in bacterial cells to increasing applications in mammalian cell programming. Meanwhile, a new growing front of research is focusing on the construction of artificial / synthetic cells, which can be programmed to have defined input-output relationships, much like a natural cell. Here, we review the state of the field for both synthetic biology and biomaterials, discuss recent work at this interface, and close with an outlook on future opportunities at the intersection of these areas. The idea for this Perspective was triggered by the stimulating discussions held during the two-day National Science Foundation Square-Table workshop in Alexandria, Virginia in October 2019. Nonetheless, there are numerous distinct research groups, clusters, and centers across the world working at the interface between synthetic biology and engineered materials, and this piece aims to provide a broad overview of the recent achievements in this exciting filed.

New frontiers in synthetic biology

Synthetic biology turned 20 years old in 2020 and represents a research topic with immense potential for transforming life in the 21st century¹. Remarkable achievements in DNA synthesis and assembly, along with standardization of genetic components and automated sequence designs, have helped biology evolve into an engineering science. From the early days of building genetic circuits in bacteria, synthetic biology approaches have now extended to mammalian systems, and even to engineering completely synthetic cells from the bottom up.

Advances in cell-base technology

Synthetic biology has revolutionized our capability to program how cells perceive, process, and react to external information (**Fig. 1a**), and cells can now be engineered to perceive specific inputs, such as the concentration or the kinetics of biological, chemical, or physical stimuli². Of particular interest are optogenetic tools as they allow optical control of molecular reactions with

exquisite spatiotemporal precision, dose-dependency, reversibility, and orthogonality³. The information perceived by such input stimuli can subsequently be processed by synthetic networks inspired by fundamental computational operations and algorithms. First pioneered with computing gene networks⁴, this information-processing capability has recently been transferred to networks of enzymes composed of kinases or proteases^{2,5}. The capability for information processing ranges from Boolean algebra to CRISPR-based networks that record and genetically store sequential information input⁶, or control-theory-inspired integral feedback controllers that maintain specific biological setpoints in fluctuating environments⁷. The concept of cellular information processing can be extended to multicellular systems interconnected by 'chemical wires' to increase, in a modular fashion, the complexity of computational operations⁸. Finally, the outcome of such information processing can be translated into desired cellular outputs like cell growth and differentiation, or the synthesis of a product ranging from simple proteins to complex, hitherto unavailable small molecule drugs⁹.

Cell-free biology and synthetic cells

Cell-free expression (CFE) has emerged as an expedient technology to build *de novo* synthetic cells from the ground up¹⁰. By recapitulating transcription and translation outside living cells, CFE makes it possible to embed selected pathways and functions into cell-sized compartments, to produce desired molecules *in situ* or to engineer life-like processes or properties (**Fig. 1b**). First, CFE provides a minimal environment to rapidly characterize and optimize integrated genetic parts producing biological materials, whose properties may depend on a multitude of interdependent parameters inside living cells^{11,12}. The development of synthetic organelles by spontaneous phase separation in CFE is within reach, with synthetic nucleic acids playing a crucial role^{13,14}. Second, CFE makes it easier to engineer the kinetics of reactions and facilitates the control of biomaterial production across many physical scales¹⁰, enabling the synthesis of adaptive, self-regulating materials¹⁵. Third, CFE is amenable to being embedded into synthetic or composite substrates and biotic-abiotic interfaces¹⁶. Altogether, these advantages are spurring the development of CFE-based synthetic cells with the capability for chemical communication that may one day lead to the development of multifunctional biofilms, proto-tissues, and organoids¹⁷.

Synthetic receptors with novel functionalities

Biological systems, comprised of cells and materials that interact with each other, give rise to a smart, complex system. Cells read, or sense, their surroundings using membrane-spanning receptors and can modulate their behavior in response to changing environments. Recently, synthetic receptors have been developed for programming this crucial behavior^{18,19}. Both the external information sensed (the 'input') and the corresponding change in behavior (the 'output') can be user-defined (**Fig. 1c**). For example, the extracellular domain can be a single-chain antibody that recognizes an exogenous protein, while the intracellular, effector domain can be a synthetic transcription factor. Many of these receptors have been successfully used for improving cell therapy for cancer²⁰ and for programming developmental transitions both *in vitro* and *in vivo*^{8,21}. Intriguingly, some of these receptors have proven to be responsive to ligands presented by materials²², paving the way for programmed interaction between cells and materials.

Application of machine learning and lab automation to synthetic biology

Synthetic biology is on the verge of realizing enormous gains in productivity as each step in the "design, build, test, and learn" engineering cycle benefits from automation (Fig. 1d). Abstraction tools, used in electronic circuit design, have also facilitated designs in synthetic biology. For example, Cello was created as a design environment that translates genetic circuits into DNA sequences²³. In the lab, mobile robotic workstations can be integrated into workflows with existing equipment. More importantly, in contrast to the overworked (and overcaffeinated) PhD student, the robot can work flawlessly around the clock with perfect reproducibility, freeing the student to focus on more creative work²⁴. Microfluidic approaches (aided by design tools like 3DµF²⁵) provide further gains in efficiency by scaling down the required reagents and experimental footprint. Furthermore, the automated workflow can be harnessed by machine learning algorithms for discovery and optimization, e.g. enabling genotype-to-phenotype predictions for optimizing tryptophan production in Saccharomyces cerevisiae²⁶. While the application of machine learning algorithms and artificial intelligence might sound 'non-living', we emphasize the critical role that human/researcher intelligence plays in setting up the parameter space and protocols for such workflows and drawing broader and deeper insights from the results. Lastly, information standards, such graphical notation used in maps of biological processes and machine-readable representations of biochemical models²⁷, facilitate communication between humans and machines, or between independent labs.

New frontiers in biomaterials

Biomaterials include diverse materials, ranging from titanium or silicone implants, metal stents, and plastic drug delivery devices all the way to hydrogels, which are water-swollen polymer networks. In this section, we focus on hydrogels due to their conduciveness to cell-biomaterial interactions in 3D, and the recent explosion of interest in the field. Hydrogels can be formed from fully synthetic polymers, biopolymers derived from natural sources, ECM proteins or other

polypeptides, and even nucleic acids. Key traditional applications of biomaterials include drug delivery, tissue engineering and regenerative medicine, and cell culture models that better recapitulate tissue microenvironments. Tremendous effort has gone into designing biomaterials to better capture the complex features of natural ECMs and to produce desired outcomes in these applications, such as: spatiotemporally controlled drug release kinetics, stem cell differentiation, enhanced wound healing, and immune system reprogramming to target cancer²⁸.

Control over intrinsic biomaterial characteristics

At the molecular level, peptide engineering and new chemistries such as bio-orthogonal or photochemical reactions have been applied to achieve more precise control of mechanical and biochemical properties of biomaterials²⁹ (**Fig. 2a**). Natural ECMs and living tissues are not simple linearly elastic materials, but rather nonlinear viscoelastic systems that undergo irreversible or plastic deformations, motivating the recent development of systems that recapitulate these more complex mechanical behaviours³⁰. For example, viscoelasticity can be achieved by incorporating dynamic or reversible crosslinking interactions such as metal-ligand coordination, host-guest bonds, hydrogen bonds, electrostatic or hydrophobic interactions, and dynamic covalent bonds, in contrast to static covalent bonds that typically result in elastic materials³⁰. These crosslinking methods can be combined with a double network approach to provide additional control of the mechanical behavior of hydrogels, or the signals presented in cellular and biomedical applications³¹. Changes in stiffness and viscoelasticity impact fundamental cell processes, including cell proliferation, apoptosis, migration, and differentiation, making these parameters critical to control in any application involving cells³⁰.

Material engineering at the micrometer (or greater) length scale can also impart mechanical properties critical to cell and tissue functions. Micron-scale elastic fibers comprising, or incorporated into, the biomaterial give rise to nonlinear stress-strain responses under tension due to reorientation and alignment³². Complementarily, void spaces in the biomaterial formed through bioprinting, foaming, or microgel annealing provide control over porosity and bulk mechanical properties, which for example might be important for structural stability of the construct, without altering the nanoscale elasticity that cells sense through focal adhesions³³. The available palette of tools to control both intrinsic mechanical properties and architectural features will continue to expand with advances in bioprinting multi-component, soft material composites³⁴.

Stimulus-responsive dynamic biomaterials

Over the last decade the state of the art in soft biomaterials transformed from mostly passive, static substrates into dynamic and stimulus-responsive materials that undergo large

changes in stiffness, swelling behavior, and 3D structure at the experimentalist's command, which in turn can elicit specific on-demand responses from embedded cells (**Fig. 2b**). These systems usually involve the removal or conformational change of crosslinks inside the material, and stimuli include externally applied cues—either physical (e.g. light, temperature, magnetic fields), or chemical (DNA, proteins, small biomolecules)—to provide (spatio)temporal control over the biomaterial properties³⁵. Recent developments highlight hydrogels that show large on-demand stiffness changes^{36,37} and strategies for the sequential removal and restoration of covalent crosslinks³⁸. Challenges ahead include optimizing response sensitivity (i.e. imparting large changes with small, biologically relevant cues) and the introduction of multiple orthogonal (and reversible) cue responses inside a single biomaterial³⁹. These developments will lay the foundation for incorporating control circuits– used extensively in synthetic biology, but less so in biomaterials– to regulate biomaterial functionality.

Multi-phase biomaterials and their characterization

The human body is full of multi-phase biomaterials, with nearly every tissue containing, at a minimum, cells and a fibrous protein and biopolymer matrix. Such multi-phase biomaterial composites present a challenging target because of their multifunctional and dynamical nature. A recent development toward such systems is the condensation of proteins and other molecules via liquid-liquid phase separation (LLPS)⁴⁰. LLPS, which has been primarily studied in the context of understanding fundamental cellular organization, has emerged as an intriguing paradigm for creating multi-phase protein biomaterials by tuning their amino acid sequence⁴¹. LLPS materials are non-crystalline and stabilized by numerous heterogeneous non-covalent interactions. which can give rise to multiscale structure to the materials. The ability to design protein/polymer systems that selectively co-localize multiple components in a biomolecular condensate is a promising avenue for not only tissue engineering and drug delivery⁴², but also synthetic biology.

Next generation biomaterials

Further advancement of biomaterials will require both the synthesis of new material compositions and advances in manufacturing, processing, printing, and assembly of material architecture⁴³. 3D bioprinting, multistep lithography and stereolithography, or post-processing for functionalization can each be used to create architectures of responsive or functional material domains. The resulting anisotropic responses of such hierarchical materials will control where the responses occur and can direct complex new responses like shape changes⁴⁴. A key challenge with some materials, particularly engineered protein-based or DNA-based biomaterials, will be on

producing materials at the kilogram scale⁴⁵, where they can be used broadly. Advances in biomimetic polymers, i.e. protein-like self-assembly⁴⁶, are a promising alternative for the commercial scale synthesis of shelf-stable materials.

Given the overall complexity of biological systems and the multifarious properties of any given material, the integration of "-omics"-based approaches (e.g. the "materials genome") with machine learning will help generate materials for diverse applications of interest (**Fig. 2c**). For example, the combination of high-throughput screening of novel biomaterials with broad yet detailed outputs and machine learning algorithms have enabled the development of antibiofouling biomaterials⁴⁷ or polymers for pharmaceutical applications⁴⁸. These approaches can identify materials with unprecedented properties and enable better characterization or engineering of biological systems.

Using synthetic biology to fabricate tailored biomaterials

Although synthetic biology and biomaterials have largely evolved as independent fields, the former has tremendous promise to provide natural or artificially designed modules for the latter. Programmed cells can (1) produce novel building blocks, and/or furnish simplified motifs that can be repurposed by materials scientists to create new biomaterials, or (2) synthesize the material of interest directly.

Innovative bioinspired building blocks can create materials with unique chemical and physical structures to achieve desired functions and properties. For example, coiled-coil peptides, aided by computational design, can be assembled into nanofibers with controllable physical characteristics using click chemistry⁴⁹. Similarly, DNA nanotechnology has pushed the boundaries of molecularly programmable shape and functionality⁵⁰. Hybrid systems that combine synthetic DNA with proteins and other biopolymers are enabling building bottom-up hierarchical structures with readily tailored functionalities⁵¹.

In a direct application of synthetic biology-based approaches to biomaterials, bacteria can be used to produce engineered variants of natural proteins as components of materials, for applications ranging from engineering of cellular microenvironments to nanowires and self-healing materials⁵². For example, the SpyTag-SpyCatcher reactive protein partners⁵³ can be incorporated into the bacterial amyloid curli system, leading to tunable functions of the resulting biofilm⁵⁴. Alternatively, this system can be applied to functionalize the 2D surface-layer proteins of *Caulobacter crescentus*⁵⁵, or to engineer the *Salmonella* microcompartment protein EutM to

organize enzymes in a cascade for efficient substrate tunneling⁵⁶. Engineered proteins can also control biomaterial properties using light-responsive proteins, often directly borrowed from the optogenetic toolbox³⁶, to release tethered enzymes or growth factors from polymeric hydrogels using light⁵⁷.

The production of useful *inorganic* biomaterials using synthetic biology is in its infancy but represents an area with great potential, particularly in the context of sustainable building materials. The development of bio-concrete—by precipitating calcium carbonate through a biomineralization process that involves the urease enzyme—is a prime example of using programmed cells to produce a material *in situ*. Microorganisms genetically engineered for slightly lower urease activity produced larger calcite crystals with high moduli⁵⁸, demonstrating that the morphology and material properties of biogenic calcite can be tailored by using a synthetic biology approach. Again by employing microbially induced CaCO₃ precipitation, it was shown that photosynthetic cyanobacteria could biomineralize inert sand-gelatin scaffolds and significantly toughen the hydrogel matrix⁵⁹; regulating the cell metabolic activity that in turn allowed for multiple 'regenerations' using temperature and humidity switches. Mirroring the rewiring of bacterial metabolic networks for enhanced production of foods, fuels, and other chemicals⁶⁰, synthetic biology-based rewiring of cellular circuits can be harnessed to further amplify natural biomineralization processes, or potentially generate novel, bio-orthogonal pathways for inorganic materials production.

Many natural biological materials result from the cooperation of multiple cell types. Secreted glucan chains, which are produced by various species of Gram-negative acetic acid bacteria, can become bundled into cellulose fibrils and form a floating mat around the embedded cells. Recently, an approach to fabricate functional bacterial cellulose using a stable co-culture of yeast and cellulose-producing bacteria was demonstrated⁶¹. Enzymes secreted by yeast can modify bacterial cellulose, generating autonomously grown catalytic materials with enzymatic functions, illustrating that complex biofabricated materials can be achieved by programming multi-cellular consortia.

The studies highlighted above represent the beginning of the convergence between synthetic biology and biomaterials research. Computational approaches are enabling the design of new building blocks inspired by (but not directly derived from) nature. Engineered peptides and proteins are essential building blocks for modular, bottom-up design of innovative materials with programmable and dynamic functions and structures. Controlling the stability, size, and spatial display of chemical functionalities on these molecules will enable well-defined hierarchical structures that span the nano- and micro- to macro-scales. Applying the concepts of parametric

chemistry and tunable hierarchical structuring, a multi-material, 3D printing platform for additive manufacturing of bio-cement has been developed⁶².

Beyond proteins, polysaccharides or hybrid systems that combine proteins. polysaccharides, DNA, and engineered cells enable the bottom-up assembly of multi-component and hierarchically structured materials systems (Fig 3a). For example, plant-derived starches are already used for the production of biodegradable bioplastics, yet their widespread adoption is limited by a combination of high costs and narrow range of material properties. Since metabolic pathways for many polysaccharides are well characterized, the prospect of controlling features like molecular weight distribution, sugar composition, and branching could lead to bioderivedmaterials with a wider range of material properties and replace existing petrochemical-derived materials. Polysaccharides can also be produced in much higher yields (>10 g/L) relative to proteins and nucleic acids, by using engineered cells or enzymes. Structural RNA self-assembly has been investigated as a scaffold for intracellular enzyme display⁶³, raising intriguing guestions about whether oligonucleotide nanomaterials could be secreted extracellularly by cellular systems. Finally, the growing interest in engineering non-living artificial cells (i.e. protocells) from the bottom-up provides yet another opportunity for creating novel biomaterials and/or providing unique sense-response capabilities.

Towards 'living' materials

A second exciting direction for merging synthetic biology and biomaterials is to create 'living' materials that not only instruct the cells but can in turn be modulated *by* the cells, to together provide functionality to the material. For example, all living organisms move in some way; in mammals, myocytes within muscle convert chemical energy into reversible mechanical work, in response to electrical and chemical stimuli. Designing living actuators driven by myocyte activity requires materials that define cell shape and orientation, cytoskeletal assembly and organization, and enable communication with excitatory inputs. Beyond repairing or replacing injured muscle tissue, such materials could create soft, biologic robotics. For example, recent work patterned genetically engineered rat cardiomyocytes that contract in response to light to create a biohybrid laser-guided stingray⁶⁴.

Living organisms also sense and respond to physiological cues, and cells programmed with stimulatory transmembrane receptors can induce numerous responses upon activation, including differentiation and immunomodulation. For example, researchers engineered synthetic Notch receptors that cause cells to undergo an epithelial to mesenchymal transition upon contact from specific sender cell ligands⁶⁵. In addition, T cells can be engineered with transmembrane

receptors that dimerize in the presence of vascular endothelial growth factor and respond by producing a cytokine (IL-2)⁶⁶. In these systems, ligands only induce a positive response, but cells programmed with stimulatory *and* inhibitory transmembrane receptors would provide biological feedback useful for engineering living materials. For example, cells embedded in a temporary scaffold with a stimulatory ligand could secrete a matrix that replaces the scaffold with nascent material containing an inhibitory ligand to regulate production (**Fig 3b**). It was recently shown that cells secrete matrix in biomaterials that they then respond to^{67,68}, and can remodel certain biomaterials through protease activity and mechanical force³⁰, but control over these dynamic interactions is missing. This paradigm has wide applicability ranging from creating regenerative tissues that otherwise have limited healing capacity, to adding lifelike properties to inanimate materials, like bricks that repair themselves (**Fig 3c**).

Another hallmark of living systems is the processing of raw materials into useful products. Cells within exocrine systems like the pancreas act as factories that assemble individual proteins into large complexes that are excreted from the cell. This natural process could be adapted for bioremediation, where dispersed pollutants are converted into environmentally inert materials. Recently, an implantable bioactive material was developed that degrades into succinate, which can enter the tricarboxylic acid cycle in mitochondria to accelerate bone regeneration⁶⁹. In the future, cells programmed with engineered metabolic pathways could serve as central processors that support homeostasis and growth of living materials.

An alternate conception of 'living materials' is systems that can evolve *like* a living system, with successive rounds of selection, 'mutation' (of material properties), and amplification. Such efforts require high-throughput synthesis and characterization/screening, with machine learning approaches to identify the best candidates for subsequent rounds of 'evolution.' Advances in robotic-enabled chemical synthesis and separation provide unprecedented opportunities to rapidly generate polymeric materials. These solution chemistry approaches are complemented by emerging advances in bioprinting (which provide spatial control of material composition⁷⁰) as well as efforts in bio-templated synthesis⁷¹ and engineered evolution, selection, and amplification⁷². However, the development of property libraries linked to formulation and composition remains a significant challenge⁷³, due to the vast property space of biomaterials, which includes (time-dependent) physical, structural, and biochemical parameters. There is also a need for clever screening/selection schemes (devised by talented, though probably still caffeinated, PhD students freshly liberated by automatic workflows to pursue creative work) that can maintain physical connections between functional material properties and specific genetic variants. There is often a

tradeoff between accuracy and speed, requiring a detailed formulation-property analysis on a select subset of systems.

Despite these challenges, several promising platforms have emerged for high throughput analysis, using volumes in the pico- to nanoliter range and measuring changes optically. Polymer microarrays, with the content of each reaction spot encoded by its spatial coordinates, enable screening hundreds or thousands of substrates for supporting the growth of different cell types, such as stem cells or islet cells, or for those minimizing cell adhesion for anti-fouling applications⁷⁴. Alternatively, cells and other materials can be encapsulated in aqueous droplets suspended in an immiscible oil using microfluidics, where each droplet serves as an independent reactor. Coupling continuous flow capabilities to a high droplet generation rate (>10 kHz) enables, for example, the directed evolution of enzymes from a library of >10⁷ enzyme variants within 10 hours⁷⁵. Finally, for materials synthesized directly by cells, flow cytometers or fluorescence-activated cell or droplet sorting⁷⁶ provide powerful interrogation methods. Extending these approaches to measure a wider range of material types and properties is a priority for biomaterials discovery.

High throughput synthesis and emerging advances in characterization provide important inputs to establishing design, build, test, and learn cycles, but advances in data science and highperformance computing infrastructure are also critically important. Standardization of data collection and management are needed, including the wide adoption of uniform naming formats for polymers and composite formulations, and standardizing experimental devices and procedures. Standardized formats are particularly critical for machine learning and algorithmic searching of data sets across numerous laboratories. These are concepts at the heart of the Materials Genome Initiative, launched across multiple federal agencies in the US aimed at "discovering, manufacturing, and deploying advanced materials twice as fast and at a fraction of the cost compared to traditional methods"⁷⁷. It is also likely that granting and regulatory agencies will play a key role in defining and enforcing these metrics within the field. We further highlight the emergence of various interdisciplinary consortiums and research clusters throughout the world that are interested in similar questions. International cooperation can likely make these developments more rapid and efficient.

Outlook

Synthetic biology and biomaterials research will each continue to flourish as independent fields, but the intersection between the two is poised to become a major focus of research efforts over the near- to mid-term, with tremendous potential for pressing materials challenges. Bridging the gap between laboratory demonstrations and fabrication techniques that can be implemented

on larger scales will be increasingly important. Some of the concepts of synthetic biopolymeric materials and cell-based fabrication have already penetrated the commercial arena⁷⁸, including the production of textiles from microbially-produced recombinant spider silk proteins, tissue-engineered leather and meat, colorless polyimides for electronics, and rigid materials based on fungal mycelium. Continued innovation in this area will enable the large-scale production of synthetic biological materials that rival or surpass existing materials, from plastics to concrete.

We have identified three major areas where we envision that this 'living interface' can have the greatest societal impact: medicine, biotechnology, and sustainability. In medicine, nearterm goals include the ability for designer cells to serve as drug delivery vehicles with closed-loop control; long-term goals include moonshots like building artificial organs by hierarchical assembly of engineered cells. In biotechnology, we anticipate an impact in accelerated vaccine production and delivery. An even longer-term vision is the non-biologic evolution of materials, especially in conjunction with dynamic, stimulus-responsive, and 'computational' biomaterials. Finally, in sustainability, we envision many possibilities ranging from cell-based or synthetic cell-based decomposable materials to completely living building materials that can sense, respond, and regenerate autonomously. Given that nature has an ecosystem with balanced biotic and abiotic factors, the living interface of synthetic biology and biomaterials may accelerate how nature comes up with new biomaterials. We recognize that engineering control/feedback of 'living' systems is inherently difficult and will require continuous innovation. We have no doubt that by imparting life-like properties into materials—both in conjunction with cells or mimicking their key properties-scientists will come with many advances that we cannot foresee today, but that will fundamentally transform both basic and applied research across many fields of science.

Acknowledgements

The authors wish to thank all the participants of the second Square Table workshop where the ideas in this perspective article originated from. The workshop was funded by the National Science Foundation (NSF) grant BMAT-1939310. We especially acknowledge Germano lannacchione for his stewardship of the Square Table workshops. The authors also acknowledge support from the National Institutes of Health grants R01 EB030031 (A.P.L), R35 GM138256 (L.M.), R21 CA232244 (K.A.H.), NSF grants CMMI 1846367 (O.C.), DMR-BMAT CAREER 1753387 (N.S.), EF-1934496 (V.N.), DMR-2004875 (N.S.J.), DMR-2004937 (M.V.), CBET-2033654 (B.M.B), DMR-2037055 (S.L.V.), MCB-2033387 (S.T.), DMR-2004796 (J.M.), DMR-2011824 (A.M.K.), the Human Frontiers in Science Program RGP0045/2018 (S.P.), Department of Energy grants DOE BES DE-SC-0010595 (E.F. and R.S.), DGF grant DFG-EXC-2189

(W.W.), Dutch Ministry of Education, Culture and Science- Gravitation 024.001.035 (P.H.J.K.), and the DARPA Engineered Living Materials Program (P.D.A.).

Competing interests

The authors declare no competing interests

Figures and captions

Figure 1| Programmability of synthetic biology. a, The heart of synthetic biology rests on our capability to program cells as information processing units for sensing a variety of inputs and produce discrete actionable outputs. Advances in information processing have been inspired by computational operations and algorithms and more recently propelled by the use of CRISPR-based networks for information recording. b, Artificial / Synthetic cells made from basic biological building blocks and incorporating transcription (TX) and translation (TL) machineries can recapitulate salient features of living cells such as basic chemical signaling, transcriptional dynamics, intracellular organization, and cytoskeleton organization. **c**, Recent advances in mammalian synthetic biology are focused on engineering receptors to allow customized sensing and response behaviors. This leverages the coupling of a desired input to a transcription factor to alter gene expression. **d**, Opportunities for automation in the design-build-test-learn cycle of synthetic biology. To date, partial autonomy exists for portions of the cycle while bridging gaps in the connections and curation of machine-interpretable information are emerging. Supplying goals and interpreting the results based on domain knowledge is an important function of the researcher and will be last to be automated. Figure d is adapted from Ref ⁷⁹.

Figure 2| **The design space for biomaterials. a**, Current design paradigm involves selection of biomaterials, crosslinking type, cell-adhesion ligand type and density, specification of void space, inclusion of elastic fibers, and multi-phase materials in order to control mechanical properties (stiffness, viscoelasticity, nonlinear elasticity, plasticity), biological signaling, and micro-scale architecture of the resulting biomaterial. b, Recent advances in biomaterials where stimuli such as light, temperature, magnetic fields, and biomechanical signals can induce changes in the properties of the biomaterial. c, The combination of high-throughput biomaterials production and omics-type measurements of biomaterial properties with the application of machine learning to identify design rules, may pave the way towards next generation biomaterials. Heatmap in Figure 2c is from Ref^{80.}

Figure 3| Using synthetic biology to fabricate biomaterials with tailored properties toward 'living' materials systems. a, Biomaterials inspired, derived, or produced by natural systems, including proteins, polysaccharides, and DNA, are being used in a variety of applications. In particular, cellular systems are being engineered for the production of materials. An exciting direction for merging synthetic biology and biomaterials is to create 'living' materials that not only instruct the cells, but where the cells in turn modulate the material properties. b, Schematic of cells programmed with biological circuitry. Stimulatory molecules (green circles) induce stimulatory receptor dimerization which causes the cell to perform a specific task (e.g., fluoresce green and secrete material with an inhibitory molecule). Similarly, inhibitory molecules (synthesized material with a red circle) cause inhibitory receptors to dimerize, signaling the cell to stop fluorescing and producing material. c, Schematic of SynBricks. Cells programmed with stimulatory and inhibitory biological feedback can be encapsulated in sacrificial hydrogel scaffolds with stimulatory molecules, causing resident cells to glow green and replace the hydrogel with calcium carbonate (raw material of bricks) tagged with inhibitory molecules. Once the inhibitory molecule concentration surpasses a threshold, cells will stop fluorescing and producing SynBrick material.

References

- 1. Meng, F. & Ellis, T. The second decade of synthetic biology: 2010–2020. *Nat. Commun.* **11**, 5174 (2020).
- 2. SedImayer, F., Aubel, D. & Fussenegger, M. Synthetic gene circuits for the detection, elimination and prevention of disease. *Nat. Biomed. Eng.* **2**, 399–415 (2018).
- 3. Kolar, K., Knobloch, C., Stork, H., Žnidarič, M. & Weber, W. OptoBase: A Web Platform for Molecular Optogenetics. *ACS Synth. Biol.* **7**, 1825–1828 (2018).
- 4. Moon, T. S., Lou, C., Tamsir, A., Stanton, B. C. & Voigt, C. A. Genetic programs constructed from layered logic gates in single cells. *Nature* **491**, 249–253 (2012).
- 5. Gao, X. J., Chong, L. S., Kim, M. S. & Elowitz, M. B. Programmable protein circuits in living cells. *Science (80-.).* **361**, 1252–1258 (2018).
- 6. Tang, W. & Liu, D. R. Rewritable multi-event analog recording in bacterial and mammalian cells. *Science (80-.).* **360**, eaap8992 (2018).
- Aoki, S. K., Lillacci, G., Gupta, A., Baumschlager, A., Schweingruber, D. & Khammash, M. A universal biomolecular integral feedback controller for robust perfect adaptation. *Nature* 570, 533–537 (2019).
- Toda, S., Blauch, L. R., Tang, S. K. Y., Morsut, L. & Lim, W. A. Programming selforganizing multicellular structures with synthetic cell-cell signaling. *Science (80-.).* (2018) doi:10.1126/science.aat0271.
- Smanski, M. J., Zhou, H., Claesen, J., Shen, B., Fischbach, M. A. & Voigt, C. A. Synthetic biology to access and expand nature's chemical diversity. *Nat. Rev. Microbiol.* 14, 135–149 (2016).
- Noireaux, V. & Liu, A. P. The New Age of Cell-Free Biology. Annu. Rev. Biomed. Eng. 22, 51–77 (2020).
- Godino, E., López, J. N., Zarguit, I., Doerr, A., Jimenez, M., Rivas, G. & Danelon, C. Cellfree biogenesis of bacterial division proto-rings that can constrict liposomes. *Commun. Biol.* 3, 539 (2020).
- Garenne, D., Libchaber, A. & Noireaux, V. Membrane molecular crowding enhances MreB polymerization to shape synthetic cells from spheres to rods. *Proc. Natl. Acad. Sci. U. S. A.* **117**, 1902–1909 (2020).
- Saleh, O. A., Jeon, B. J. & Liedl, T. Enzymatic degradation of liquid droplets of DNA is modulated near the phase boundary. *Proc. Natl. Acad. Sci. U. S. A.* (2020) doi:10.1073/pnas.2001654117.
- 14. Sokolova, E., Spruijt, E., Hansen, M. M. K., Dubuc, E., Groen, J., Chokkalingam, V.,

Piruska, A., Heus, H. A. & Huck, W. T. S. Enhanced transcription rates in membrane-free protocells formed by coacervation of cell lysate. *Proc. Natl. Acad. Sci. U. S. A.* **110**, 11692–11697 (2013).

- Green, L. N., Subramanian, H. K. K., Mardanlou, V., Kim, J., Hariadi, R. F. & Franco, E. Autonomous dynamic control of DNA nanostructure self-assembly. *Nat. Chem.* **11**, 510– 520 (2019).
- Efrat, Y., Tayar, A. M., Daube, S. S., Levy, M. & Bar-Ziv, R. H. Electric-Field Manipulation of a Compartmentalized Cell-Free Gene Expression Reaction. ACS Synth. Biol. 7, 1829–1833 (2018).
- Dupin, A. & Simmel, F. C. Signalling and differentiation in emulsion-based multicompartmentalized in vitro gene circuits. *Nat. Chem.* (2019) doi:10.1038/s41557-018-0174-9.
- Santorelli, M., Lam, C. & Morsut, L. Synthetic development: building mammalian multicellular structures with artificial genetic programs. *Curr. Opin. Biotechnol.* 59, 130– 140 (2019).
- Scheller, L., Strittmatter, T., Fuchs, D., Bojar, D. & Fussenegger, M. Generalized extracellular molecule sensor platform for programming cellular behavior article. *Nat. Chem. Biol.* 14, 723–729 (2018).
- 20. Rivière, I. & Sadelain, M. Chimeric Antigen Receptors: A Cell and Gene Therapy Perspective. *Mol. Ther.* **25**, 1117–1124 (2017).
- Stapornwongkul, K. S., de Gennes, M., Cocconi, L., Salbreux, G. & Vincent, J. P. Patterning and growth control in vivo by an engineered GFP gradient. *Science (80-.*). 370, 321–327 (2020).
- Huang, X., Williams, J. Z., Chang, R., Li, Z., Burnett, C. E., Hernandez-Lopez, R., Setiady, I., Gai, E., Patterson, D. M., Yu, W., Roybal, K. T., Lim, W. A. & Desai, T. A. DNA scaffolds enable efficient and tunable functionalization of biomaterials for immune cell modulation. *Nat. Nanotechnol.* Online ahead of print (2020) doi:10.1038/s41565-020-00813-z.
- Nielsen, A. A. K., Der, B. S., Shin, J., Vaidyanathan, P., Paralanov, V., Strychalski, E. A., Ross, D., Densmore, D. & Voigt, C. A. Genetic circuit design automation. *Science (80-.).* 352, aac7341 (2016).
- Burger, B., Maffettone, P. M., Gusev, V. V., Aitchison, C. M., Bai, Y., Wang, X., Li, X., Alston, B. M., Li, B., Clowes, R., Rankin, N., Harris, B., Sprick, R. S. & Cooper, A. I. A mobile robotic chemist. *Nature* 583, 237–241 (2020).

- Sanka, R., Lippai, J., Samarasekera, D., Nemsick, S. & Densmore, D. 3DµF Interactive Design Environment for Continuous Flow Microfluidic Devices. *Sci. Rep.* 9, 9166 (2019).
- Zhang, J., Petersen, S. D., Radivojevic, T., Ramirez, A., Pérez-Manríquez, A., Abeliuk, E., Sánchez, B. J., Costello, Z., Chen, Y., Fero, M. J., Martin, H. G., Nielsen, J., Keasling, J. D. & Jensen, M. K. Combining mechanistic and machine learning models for predictive engineering and optimization of tryptophan metabolism. *Nat. Commun.* **11**, 4880 (2020).
- Waltemath, D., Golebiewski, M., Blinov, M. L., Gleeson, P., Hermjakob, H., Hucka, M., Inau, E. T., Keating, S. M., König, M., Krebs, O., Malik-Sheriff, R. S., Nickerson, D., Oberortner, E., Sauro, H. M., Schreiber, F., Smith, L., Stefan, M. I., Wittig, U. & Myers, C. J. The first 10 years of the international coordination network for standards in systems and synthetic biology (COMBINE). *J. Integr. Bioinform.* **17**, 20200005 (2020).
- 28. Huebsch, N. & Mooney, D. J. Inspiration and application in the evolution of biomaterials. *Nature* **462**, (2009).
- 29. Dimarco, R. L. & Heilshorn, S. C. Multifunctional materials through modular protein engineering. *Adv. Mater.* **24**, 3923–3940 (2012).
- 30. Chaudhuri, O., Cooper-White, J., Janmey, P. A., Mooney, D. J. & Shenoy, V. B. Effects of extracellular matrix viscoelasticity on cellular behaviour. *Nature* **584**, 535–546 (2020).
- Rodell, C. B., MacArthur, J. W., Dorsey, S. M., Wade, R. J., Wang, L. L., Woo, Y. J. & Burdick, J. A. Shear-thinning supramolecular hydrogels with secondary autonomous covalent crosslinking to modulate viscoelastic properties in vivo. *Adv. Funct. Mater.* 25, 636–644 (2015).
- Baker, B. M., Trappmann, B., Wang, W. Y., Sakar, M. S., Kim, I. L., Shenoy, V. B., Burdick, J. A. & Chen, C. S. Cell-mediated fibre recruitment drives extracellular matrix mechanosensing in engineered fibrillar microenvironments. *Nat. Mater.* 14, 1262–1268 (2015).
- Griffin, D. R., Weaver, W. M., Scumpia, P. O., Di Carlo, D. & Segura, T. Accelerated wound healing by injectable microporous gel scaffolds assembled from annealed building blocks. *Nat. Mater.* 14, 737–744 (2015).
- 34. Skylar-Scott, M. A., Mueller, J., Visser, C. W. & Lewis, J. A. Voxelated soft matter via multimaterial multinozzle 3D printing. *Nature* **575**, 330–335 (2019).
- 35. Guo, Z., Liu, H., Dai, W. & Lei, Y. Responsive principles and applications of smart materials in biosensing. *Smart Mater. Med.* **1**, 54–65 (2020).
- 36. Hörner, M., Raute, K., Hummel, B., Madl, J., Creusen, G., Thomas, O. S., Christen, E. H., Hotz, N., Gübeli, R. J., Engesser, R., Rebmann, B., Lauer, J., Rolauffs, B., Timmer, J.,

Schamel, W. W. A., Pruszak, J., Römer, W., Zurbriggen, M. D., Friedrich, C., Walther, A., Minguet, S., Sawarkar, R. & Weber, W. Phytochrome-Based Extracellular Matrix with Reversibly Tunable Mechanical Properties. *Adv. Mater.* **31**, e1806727 (2019).

- de Almeida, P., Jaspers, M., Vaessen, S., Tagit, O., Portale, G., Rowan, A. E. & Kouwer,
 P. H. J. Cytoskeletal stiffening in synthetic hydrogels. *Nat. Commun.* 10, 609 (2019).
- Rosales, A. M., Vega, S. L., DelRio, F. W., Burdick, J. A. & Anseth, K. S. Hydrogels with Reversible Mechanics to Probe Dynamic Cell Microenvironments. *Angew. Chemie* - *Int. Ed.* 56, 12132–12136 (2017).
- Badeau, B. A. & Deforest, C. A. Programming Stimuli-Responsive Behavior into Biomaterials. *Annu. Rev. Biomed. Eng.* 21, 241–265 (2019).
- 40. Chao, Y. & Shum, H. C. Emerging aqueous two-phase systems: From fundamentals of interfaces to biomedical applications. *Chem. Soc. Rev.* **49**, 114–142 (2020).
- Schuster, B. S., Dignon, G. L., Tang, W. S., Kelley, F. M., Ranganath, A. K., Jahnke, C. N., Simpkins, A. G., Regy, R. M., Hammer, D. A., Good, M. C. & Mittal, J. Identifying sequence perturbations to an intrinsically disordered protein that determine its phase-separation behavior. *Proc. Natl. Acad. Sci. U. S. A.* **117**, 11421–11431 (2020).
- Klein, I. A., Boija, A., Afeyan, L. K., Hawken, S. W., Fan, M., Dall'Agnese, A., Oksuz, O., Henninger, J. E., Shrinivas, K., Sabari, B. R., Sagi, I., Clark, V. E., Platt, J. M., Kar, M., McCall, P. M., Zamudio, A. V., Manteiga, J. C., Coffey, E. L., Li, C. H., Hannett, N. M., Guo, Y. E., Decker, T. M., Lee, T. I., Zhang, T., Weng, J. K., Taatjes, D. J., Chakraborty, A., Sharp, P. A., Chang, Y. T., Hyman, A. A., Gray, N. S. & Young, R. A. Partitioning of cancer therapeutics in nuclear condensates. *Science (80-.).* 368, 1386–1392 (2020).
- 43. Champeau, M., Heinze, D. A., Viana, T. N., de Souza, E. R., Chinellato, A. C. & Titotto,
 S. 4D Printing of Hydrogels: A Review. *Adv. Funct. Mater.* **30**, 1910606 (2020).
- Cangialosi, A., Yoon, C. K., Liu, J., Huang, Q., Guo, J., Nguyen, T. D., Gracias, D. H. & Schulman, R. DNA sequence–directed shape change of photopatterned hydrogels via high-degree swelling. *Science (80-.).* 357, 1126–1130 (2017).
- 45. Praetorius, F., Kick, B., Behler, K. L., Honemann, M. N., Weuster-Botz, D. & Dietz, H. Biotechnological mass production of DNA origami. *Nature* **552**, (2017).
- 46. Barbee, M. H., Wright, Z. M., Allen, B. P., Taylor, H. F., Patteson, E. F. & Knight, A. S. Protein-Mimetic Self-Assembly with Synthetic Macromolecules. *Macromolecules* **54**,

(2021).

- Chan, D., CHien, J.-C., Axpe, E., Blankemeier, L., Baker, S. W., Swaminathan, S.,
 Piunova, V. A., Zubarev, D. Y., Soh, H. T. & Appel, E. A. Combinatorial Polyacrylamide
 Hydrogels for Preventing Biofouling on Implantable Biosensors. *bioRxiv* (2020)
 doi:10.1101/2020.05.25.115675.
- 48. Upadhya, R., Kosuri, S., Tamasi, M., Meyer, T. A., Atta, S., Webb, M. A. & Gormley, A. J. Automation and data-driven design of polymer therapeutics. *Adv. Drug Deliv. Rev.* 171, 1–28 (2021).
- Wu, D., Sinha, N., Lee, J., Sutherland, B. P., Halaszynski, N. I., Tian, Y., Caplan, J., Zhang, H. V., Saven, J. G., Kloxin, C. J. & Pochan, D. J. Polymers with controlled assembly and rigidity made with click-functional peptide bundles. *Nature* **574**, 658–662 (2019).
- 50. Seeman, N. C. & Sleiman, H. F. DNA nanotechnology. Nat. Rev. Mater. 3, 17068 (2017).
- Buchberger, A., Simmons, C. R., Fahmi, N. E., Freeman, R. & Stephanopoulos, N. Hierarchical Assembly of Nucleic Acid/Coiled-Coil Peptide Nanostructures. *J. Am. Chem.* Soc. 142, 1406–1416 (2020).
- An, B., Wang, Y., Jiang, X., Ma, C., Mimee, M., Moser, F., Li, K., Wang, X., Tang, T. C., Huang, Y., Liu, Y., Lu, T. K. & Zhong, C. Programming Living Glue Systems to Perform Autonomous Mechanical Repairs. *Matter* 3, 2080–2092 (2020).
- 53. Keeble, A. H. & Howarth, M. Power to the protein: Enhancing and combining activities using the Spy toolbox. *Chem. Sci.* **11**, 7281–7291 (2020).
- 54. Nguyen, P. Q., Botyanszki, Z., Tay, P. K. R. & Joshi, N. S. Programmable biofilm-based materials from engineered curli nanofibres. *Nat. Commun.* **5**, 4945 (2014).
- Charrier, M., Li, D., Mann, V. R., Yun, L., Jani, S., Rad, B., Cohen, B. E., Ashby, P. D., Ryan, K. R. & Ajo-Franklin, C. M. Engineering the S-Layer of Caulobacter crescentus as a Foundation for Stable, High-Density, 2D Living Materials. *ACS Synth. Biol.* 8, 181–190 (2019).
- Zhang, G., Johnston, T., Quin, M. B. & Schmidt-Dannert, C. Developing a Protein Scaffolding System for Rapid Enzyme Immobilization and Optimization of Enzyme Functions for Biocatalysis. ACS Synth. Biol. 8, 1867–1876 (2019).
- Shadish, J. A., Strange, A. C. & Deforest, C. A. Genetically Encoded Photocleavable Linkers for Patterned Protein Release from Biomaterials. *J. Am. Chem. Soc.* 141, 15619– 15625 (2019).
- 58. Heveran, C. M., Liang, L., Nagarajan, A., Hubler, M. H., Gill, R., Cameron, J. C., Cook, S.

M. & Srubar, W. V. Engineered Ureolytic Microorganisms Can Tailor the Morphology and Nanomechanical Properties of Microbial-Precipitated Calcium Carbonate. *Sci. Rep.* **9**, 14721 (2019).

- Heveran, C. M., Williams, S. L., Qiu, J., Artier, J., Hubler, M. H., Cook, S. M., Cameron, J. C. & Srubar, W. V. Biomineralization and Successive Regeneration of Engineered Living Building Materials. *Matter* 2, 481–494 (2020).
- Nielsen, J. & Keasling, J. D. Engineering Cellular Metabolism. *Cell* 164, 1185– 1197 (2016).
- Gilbert, C., Tang, T. C., Ott, W., Dorr, B. A., Shaw, W. M., Sun, G. L., Lu, T. K. & Ellis, T. Living materials with programmable functionalities grown from engineered microbial co-cultures. *Nat. Mater.* (2021) doi:10.1038/s41563-020-00857-5.
- Duro-Royo, J., Van Zak, J., Tai, Y. J., Ling, A. S. & Oxman, N. Parametric chemistry reverse engineering biomaterial composites for additive manufacturing of bio-cement structures across scales. in *Challenges for Technology Innovation: An Agenda for the Future* (CRC Press, 2017). doi:10.1201/9781315198101-39.
- Sachdeva, G., Garg, A., Godding, D., Way, J. C. & Silver, P. A. In vivo co-localization of enzymes on RNA scaffolds increases metabolic production in a geometrically dependent manner. *Nucleic Acids Res.* 42, 9493–9503 (2014).
- Park, S. J., Gazzola, M., Park, K. S., Park, S., Di Santo, V., Blevins, E. L., Lind, J. U., Campbell, P. H., Dauth, S., Capulli, A. K., Pasqualini, F. S., Ahn, S., Cho, A., Yuan, H., Maoz, B. M., Vijaykumar, R., Choi, J. W., Deisseroth, K., Lauder, G. V., Mahadevan, L. & Parker, K. K. Phototactic guidance of a tissue-engineered soft-robotic ray. *Science (80-.).* 353, 158–162 (2016).
- Morsut, L., Roybal, K. T., Xiong, X., Gordley, R. M., Coyle, S. M., Thomson, M. & Lim, W.
 A. Engineering Customized Cell Sensing and Response Behaviors Using Synthetic Notch Receptors. *Cell* 164, 780–791 (2016).
- 66. Schwarz, K. A., Daringer, N. M., Dolberg, T. B. & Leonard, J. N. Rewiring human cellular input-output using modular extracellular sensors. *Nat. Chem. Biol.* **13**, 202–209 (2017).
- 67. Loebel, C., Mauck, R. L. & Burdick, J. A. Local nascent protein deposition and remodelling guide mesenchymal stromal cell mechanosensing and fate in three-dimensional hydrogels. *Nat. Mater.* **18**, (2019).
- Ferreira, S. A., Motwani, M. S., Faull, P. A., Seymour, A. J., Yu, T. T. L., Enayati, M., Taheem, D. K., Salzlechner, C., Haghighi, T., Kania, E. M., Oommen, O. P., Ahmed, T., Loaiza, S., Parzych, K., Dazzi, F., Varghese, O. P., Festy, F., Grigoriadis, A. E., Auner, H.

| 1 | | W., Snijders, A. P., Bozec, L. & Gentleman, E. Bi-directional cell-pericellular |
|----|-----|--|
| 2 | | matrix interactions direct stem cell fate. Nat. Commun. 9, (2018). |
| 3 | 69. | Liu, H., Du, Y., St-Pierre, J. P., Bergholt, M. S., Autefage, H., Wang, J., Cai, M., |
| 4 | | Yang, G., Stevens, M. M. & Zhang, S. Bioenergetic-active materials enhance |
| 5 | | tissue regeneration by modulating cellular metabolic state. Sci. Adv. 6, 32232154 |
| 6 | | (2020). |
| 7 | 70. | Li, Y. C., Zhang, Y. S., Akpek, A., Shin, S. R. & Khademhosseini, A. 4D bioprinting: The |
| 8 | | next-generation technology for biofabrication enabled by stimuli-responsive materials. |
| 9 | | Biofabrication 9 , 012001 (2017). |
| 10 | 71. | Nam, K. T., Kim, D. W., Yoo, P. J., Chiang, C. Y., Meethong, N., Hammond, P. T., |
| 11 | | Chiang, Y. M. & Belcher, A. M. Virus-enabled synthesis and assembly of nanowires for |
| 12 | | lithium ion battery electrodes. Science (80). 312, 885–888 (2006). |
| 13 | 72. | Kan, A. & Joshi, N. S. Towards the directed evolution of protein materials. MRS |
| 14 | | <i>Commun.</i> 9 , 441–455 (2019). |
| 15 | 73. | Green, M. L., Choi, C. L., Hattrick-Simpers, J. R., Joshi, A. M., Takeuchi, I., Barron, S. C., |
| 16 | | Campo, E., Chiang, T., Empedocles, S., Gregoire, J. M., Kusne, A. G., Martin, J., Mehta, |
| 17 | | A., Persson, K., Trautt, Z., Van Duren, J. & Zakutayev, A. Fulfilling the promise of the |
| 18 | | materials genome initiative with high-throughput experimental methodologies. Appl. Phys. |
| 19 | | <i>Rev.</i> 4 , 011105 (2017). |
| 20 | 74. | Algahtani, M. S., Scurr, D. J., Hook, A. L., Anderson, D. G., Langer, R. S., Burley, J. C., |
| 21 | | Alexander, M. R. & Davies, M. C. High throughput screening for biomaterials discovery. J. |
| 22 | | Control. Release 190 , 115–126 (2014). |
| 23 | 75. | Agresti, J. J., Antipov, E., Abate, A. R., Ahn, K., Rowat, A. C., Baret, J. C., Marquez, M., |
| 24 | | Klibanov, A. M., Griffiths, A. D. & Weitz, D. A. Ultrahigh-throughput screening in drop- |
| 25 | | based microfluidics for directed evolution. Proc. Natl. Acad. Sci. U. S. A. 107, 4004–4009 |
| 26 | | (2010). |
| 27 | 76. | Ma, F., Chung, M. T., Yao, Y., Nidetz, R., Lee, L. M., Liu, A. P., Feng, Y., Kurabayashi, K. |
| 28 | | & Yang, G. Y. Efficient molecular evolution to generate enantioselective enzymes using a |
| 29 | | dual-channel microfluidic droplet screening platform. Nat. Commun. 9, 1030 (2018). |
| 30 | 77. | Liu, Y., Niu, C., Wang, Z., Gan, Y., Zhu, Y., Sun, S. & Shen, T. Machine learning in |
| 31 | | materials genome initiative: A review. J. Mater. Sci. Technol. 57, 113–122 (2020). |
| 32 | 78. | Voigt, C. A. Synthetic biology 2020–2030: six commercially-available products that |
| 33 | | are changing our world. Nat. Commun. 11, 6379 (2020). |
| 34 | 79. | Beal, J. & Rogers, M. Levels of autonomy in synthetic biology engineering. Mol. Syst. |
| 35 | | <i>Biol.</i> 16 , e10019 (2020). |
| | | |

| 36 | | |
|----|-----|---|
| 37 | | |
| 38 | 80. | Stowers, R. S., Shcherbina, A., Israeli, J., Gruber, J. J., Chang, J., Nam, S., Rabiee, A., |
| 39 | | Teruel, M. N., Snyder, M. P., Kundaje, A. & Chaudhuri, O. Matrix stiffness induces a |
| 40 | | tumorigenic phenotype in mammary epithelium through changes in chromatin |
| 41 | | accessibility. Nat. Biomed. Eng. 3, 1009–1019 (2019). |

а

C

Output

Sensing Action __ Inputs - Replication **Biological cues** Differentiation Cell-cell-contacts Signalling Information - Biosynthesis Circuits Chemical cues processing - Bioconversion Small molecules - Biodegradation - Movement Macromolecules & assemblies - Forces Concentrations & binding kinetics - Stiffening Outputs 🌄 Boolean algebra - Softening Information storage - Permeability 00 Death • Physical cues Integration Shape Light Oscillation control Temperature Recording Voltage Perfect adaptation ... d Goals Examples Soluble ligand, material-bound ligand Domain Input knowledge Native receptor domain or Sensor Desigr antibody-based ligand binding domain Learn Actuator **Transcription factors** Analytical tool APIs Build Test

Regulated gene expression

b

Harvesting

.

Component

specifications

Protocol

Executable

protocols

Protocol

specifications

Executable

protocols

specifications

Signaling



