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Publication Date
2016-07-14

DOI
10.3354/meps11774

Peer reviewed
Quantifying the top-down effects of grazers on a rocky shore: selective grazing and the potential for competition

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Abstract

Grazers affect the structure of primary producer assemblages, and the details of these interactions have been well described for terrestrial habitats. By contrast, the effect of grazers on the diversity, distribution, and composition of their principal food source has rarely been described for the high intertidal zone of rocky shores, a model system for studying the potential effects of climate change. Along rocky, wave-swept shores in central California, the microphytobenthos (MPB) supports diverse assemblages of limpets and littorine snails, which, at current benign temperatures, could potentially partition food resources in a complementary fashion, thereby enhancing secondary
productivity. Two limpet species in particular, *Lottia scabra* and *L. austrodigitalis*, may partition components of the MPB, and are likely to affect the composition of the MPB on which they graze. In this study, we describe the composition, nutritional value (C:N ratio), and fluorescence (an index of chlorophyll density) of ungrazed, *L. scabra*-grazed and *L. austrodigitalis*-grazed MPB assemblages, each as function of temperature. Fluorescence decreased with increased average daily maximum temperature for ungrazed MPB, but temperature had no discernible effects on either fluorescence or the composition of the MPB of grazed assemblages. *L. austrodigitalis* and *L. scabra* did not partition the MPB, and did not exhibit complementarity. Both species exhibited an ordered grazing scheme, in which limpets grazed down certain components of the MPB before others, and grazing increased the C:N ratio of the MPB, decreasing its nutritional value. Taken together, these results suggest that *L. austrodigitalis* and *L. scabra* may experience increased competition as warming temperatures reduce the available MPB.

**Keywords:** Selective grazing; intertidal zone; biofilm; microalgae; microphytobenthos; *Lottia*; limpets

**Introduction**

The distribution and diversity of primary producers play a major role in structuring communities. In terrestrial environments, plant assemblages and the biological and physical factors that determine their structure have been extensively studied (e.g. Harper 1977, Tilman 1988). The diversity, distribution and abundance of plant species affect a number of ecosystem processes that include productivity (Naeem et al. 1994, Tilman et al. 1996, Hector et al. 1999, Cardinale et al. 2007), functional stability (McNaughton 1977), the distribution and abundance of consumers (MacArthur &
Numerous studies have examined the effects of intertidal grazers on the structure of macroalgal assemblages in the wave-swept, rocky intertidal zone, a model system for experimental ecology (see Underwood 1979, Coleman et al. 2006 and Poore et al. 2012 for relevant syntheses; Dayton 1975, Littler & Littler 1984, Gee & Warwick 1994, Williams et al. 2013, Best et al. 2014 for specific examples). However, relatively few have investigated the role of grazing on the epilithic microphytobenthos (MPB) that dominates the upper intertidal environment (but see Castenholz 1961, Nicotri 1977, Underwood 1984b and Underwood & Murphy 2008), and the composition and distribution of the MPB in the upper intertidal environment has yet to be described at scales relevant to intertidal grazers.

The intertidal MPB is composed of a diverse assemblage of organisms, including cyanobacteria, diatoms, macroalgal spores, germlings and juveniles and invertebrate larvae, all embedded in an extracellular matrix (Wahl 1989, Underwood 1984, Davey & O'Toole 2000). The MPB coats all hard surfaces submerged in water (Sekar et al. 2004), and extends to the top of the splash zone in the intertidal environment (Ricketts et al. 1968). In the upper intertidal zone, where the MPB is the only source of benthic primary productivity, it supports a diverse assemblage of limpets and littorine snails (Ricketts et al. 1968). However, the composition and distribution of components within the MPB, the potential for partitioning of these components by grazers, and the feedbacks between top-down consumption by grazers and bottom-up availability of nutrients in the MPB remain largely unknown.

In diverse terrestrial plant assemblages, niche partitioning can reduce interspecific competition, resulting in enhanced primary productivity relative to the performance of the most productive single-species assemblage (Tilman 2000, Naeem 2002, Hooper et al. 2005, Cardinale et al. 2011). This idea,
called complementarity, is generalizable across trophic levels: increased predator diversity is also associated with higher rates of prey capture due to resource partitioning (Ives et al. 2005, Byrnes et al. 2006; Griffin et al. 2008). Although early studies linking biodiversity to ecosystem functioning highlighted the role of complementarity in temperate grassland assemblages, it also underpins relationships between diversity and productivity in intertidal systems (Griffin et al. 2008, Byrnes & Stachowitz 2009, Cardinale et al. 2011, Bracken & Williams 2013). For example, Griffin et al. (2008) used a factorial mesocosm experiment to explore how the species richness of prey assemblages affects the rate of prey consumption by three species of intertidal crabs. They found that, due to complementarity, all species of crabs ingested more food per individual when their prey was in polyculture than in any single monoculture. Through complementarity, diverse assemblages may also show increased resistance to the effects of climate change (Loreau et al. 2001): if resources become more scarce (as a result of increasing temperature, for instance), organisms that partition resources are more likely to survive and thrive. If upper intertidal grazers are complementary in their consumption of components of the MPB, it may have implications for how those species coexist now, and how they will be influenced by future climate shifts.

Nicotri (1977) measured the effects of foraging by the intertidal limpets *Lottia pelta* Rathke, *Lottia strigatella* Carpenter and *Lottia scutum* Rathke and the littorine snail *Littorina scutulata* Gould on MPB and observed that niche partitioning did not occur. She used transmitted light and scanning electron microscopy to determine that these four species grazed statistically indistinguishable quantities of the same species of cyanobacteria and diatoms from the MPB. However, her study organisms typically exist in different microhabitats, and thus may not always have been in direct competition for food resources. In addition, all four gastropod species supplement their diets with macroalgae, and so may not have experienced evolutionary pressure to specialize on the MPB. Similarly, Hawkins et al. (1989) examined three European gastropods, *Gibbula umbilicalis* da Costa, *Littorina littorea* Linnaeus...
and *Patella vulgata* Linné, and found that their diets were indistinguishable despite differences in radular structure and feeding movements. But again, these species overlap only in a subset of locations across wide geographic ranges (Hawkins et al. 1989), and *G. umbilicalis* and *L. littorea* supplement their diets with detritus and macroalgae, respectively. Organisms that specialize entirely on the MPB and share the same microhabitat may be more likely to exhibit niche partitioning, and thus complementarity.

Recently, strong evidence has emerged that three intertidal gastropod species native to Italy may exhibit complementarity through resource partitioning (Sanz-Lázaro et al. 2015). Sanz-Lázaro et al. (2015) manipulated the density and richness of two limpet species (*Patella ulyssiponensis* and *P. rustica*) and one topshell (*Porchus turbinatus*) and observed the effects on the photosynthetic efficiency and photosynthetic biomass of the MPB. These gastropods share the same microhabitat in the intertidal environment, and they specialize on microalgae as their primary food resource. When two of these species grazed in concert, the photosynthetic efficiency of the MPB significantly increased while photosynthetic biomass did not, suggesting complementarity via resource partitioning. However, these effects were not additive: all three species grazing together reduced photosynthetic efficiency back to levels typical of individual grazers.

Along the central coast of California, two limpet species, *Lottia scabra* Gould and *Lottia austrodigitalis* Murphy, share the upper intertidal habitat, living above the *Mytilus californianus* mussel zone and often beyond the upper extent of the macroalga *Endocladia muricata* (Wolcott 1973). They share this habitat with small barnacles (*Chthamalus* spp.) and a variety of littorine snails (*Littorina* spp.) (Ricketts et al. 1968). Both *L. scabra* and *L. austrodigitalis* live on vertical rock surfaces, although *L. scabra* can also be found on horizontal surfaces (Collins 1976, Hahn & Denny 1989). Both species forage for microalgae twice per day during high tides, and exhibit a general dietary preference for diatoms (Nicotri 1977).
Previous research suggests that *L. austrodigitalis* and *L. scabra* are in direct competition for at least some food resources, and this competition could be driven by several factors. Haven (1973) observed that when *L. austrodigitalis* were excluded from a plot containing *L. scabra* in the intertidal zone at Hopkins Marine Station (HMS) in Pacific Grove, California, the density of the MPB increased, as did *L. scabra* growth rates. This result provides evidence that at least some portion of the MPB is a shared resource between the two species. However, it does not exclude the possibility that there are other components of the MPB that may be partitioned between *L. austrodigitalis* and *L. scabra*.

Food quality might also drive competition between *L. austrodigitalis* and *L. scabra*. Gastropod grazers, including limpets, are frequently selective in their preferences for intertidal algae (Della Santina et al. 1993, Bracken & Low 2012). Often, the preferred species are those with higher internal nitrogen concentrations or lower carbon-to-nitrogen (C:N) ratios (Pedersen & Borum 1996, Bracken et al. 2011, Bracken et al. 2014). This preference for higher nitrogen food sources is consistent with limitation of gastropod abundance and growth by nitrogen availability (Nielsen 2001, Sommer 2001). Differences between the C:N ratios of MPB grazed by *L. austrodigitalis* and *L. scabra* would suggest that they are targeting different food resources.

Lastly, temperature may influence the intensity of competition between these two *Lottia* species by reducing the availability, composition, or quality of an already limited food resource. Dark-adapted fluorescence of intertidal MPB can be used as a proxy for chlorophyll *a* density and is known to decrease as temperature increases (Miller et al. 2015). This suggests that less food is available to epilithic grazers like limpets when rock surface temperatures are high. Additionally, temperature may affect the composition and quality of the MPB. For example, Jackson et al. (2010) found that intertidal MPB on an Australian subtropical beach contained fewer cyanobacteria than that on a temperate beach. Diatoms may also be less plentiful at higher temperatures; they tend to be more abundant lower on shore (Castenholz 1961) and decrease with increasing insolation (Thompson et al. 2004).
Diatoms are a nitrogen-rich food source for intertidal grazers, an increase in temperature may lead to a reduction in food quality, as well as food availability.

In this study, we investigated the effects of limpet grazing on the composition and quality of the upper intertidal MPB. Specifically, we asked:

1) How does temperature influence the density and quality of the MPB?
2) How do *L. austrodigitalis* and *L. scabra* affect the density and composition of the MPB?
3) Do *L. scabra* and *L. austrodigitalis* exhibit complementarity by partitioning components of the MPB during grazing?
4) How does grazing by *L. austrodigitalis* and *L. scabra* affect the nutritional quality of the MPB?

To answer these questions, we examined the dark-adapted fluorescence (a proxy for chlorophyll density) and C:N ratio (an index of food quality) of MPB as a function of average maximum temperature of the substratum. We also compared the dark-adapted fluorescence and composition of MPB that had not been grazed with MPB that had been grazed by *L. scabra*, MPB that had been grazed by *L. austrodigitalis*, and MPB that had been grazed by the natural suite of upper intertidal gastropods. Lastly, we quantified the percent cover of morphotypes in the MPB from each grazing treatment, and compared the results between treatments. We expected that increased temperatures and limpet grazing would decrease dark-adapted fluorescence values, change the composition of the MPB, and increase the C:N ratio. Additionally, if resource partitioning was occurring at the microscopic level, we expected that the composition of the *L. scabra*-grazed MPB would differ from that of the *L. austrodigitalis*-grazed MPB.
Methods

Field experiment

We selected six vertically-oriented field sites in the rocky, wave-swept intertidal zone adjacent to HMS (36.6217° N, 121.9043° W). Sites were between 3 m and 35 m apart. At each site, we deployed four circular aluminum plates (10 cm in diameter, 12 mm thick) in horizontal transects at 1.7 m above mean lower low water (MLLW). The average diurnal tidal range at this site is 1.6 m. We covered each plate with a thin layer of light gray rubber grip tape (Safety Walk Tape, 3M) to encourage microalgal growth. Each plate was attached to the granite substratum with a central bolt, and a cement pad was constructed beneath each plate to ensure good thermal contact with the underlying rock. At each site, one plate held four *L. scabra* and one held four *L. austrodigitalis*. This density (500 limpets m⁻²) corresponded to that observed in limpet-rich areas of the intertidal zone at HMS (Morelissen and Harley 2007). Individuals of about 1 cm in shell length were selected for the experiment, and were corralled using 20 mm tall, stainless steel mesh fences (5.5 mm square mesh size). These fences deterred most limpets from leaving the plates, but an average of 0.11 ± 0.15 *L. scabra* and 1.31 ± 0.94 *L. austrodigitalis* were still lost per month. A third plate at each site had an identical fence and served as a grazer exclusion control, while the fourth plate lacked a fence altogether and allowed access by grazers living on the neighboring natural rock surface. We deployed the plates in June of 2013, and they remained in the field through December of that year. Fences were cleared of any macroalgal growth in the late fall.

It is important to note that in Monterey Bay, *L. austrodigitalis* overlaps with a cryptic congener, *L. digitalis* Rathke. The two species can be reliably identified only through genetic methods. However, prior research at our field site suggests that *L. austrodigitalis* are most commonly found on
vertical rocky surfaces, while *L. digitalis* is associated with the micro-environment provided by the gooseneck barnacle, *Pollicipes polymerus* Sowerby (Crummett & Eernisse 2007). More recent work has also shown that *L. austrodigitalis* makes up 88-89% of the cryptic species pair in high shore habitats at HMS (Dong et al. 2008, Dong & Somero 2009), where our limpets were collected. We refer to *L. austrodigitalis* throughout the remainder of this study, acknowledging that a small percentage of our organisms may in fact be *L. digitalis*.

**Temperature**

Temperature measurements were obtained for each plate using iButton temperature dataloggers (0.5°C resolution, Maxim Integrated, San Jose, CA, USA). We machined a circular pocket in the back of each plate to hold the iButton. The iButtons recorded temperature every 12 minutes, and we downloaded the data every 2 weeks.

At the conclusion of the experiment, the average maximum daily temperature for December 2013 was calculated for each plate. We checked that these temperatures were consistent across sites and grazing treatments using 2-factor ANOVA and Tukey’s HSD test for multiple comparisons. Visual checks were performed to ensure that model assumptions of normality and equal variances were met. Analyses were carried out in MATLAB 2014a.

**Dark-adapted fluorescence**

We surveyed plates on July 10, August 6, September 6, October 6, November 6 and December 1, 2013 during favorable tides and wave conditions. During each survey, we replaced any limpets lost during the previous month and measured the density of the MPB. The density of the MPB was approximated using a PAM fluorometer (Diving-PAM, Walz, Effeltrich, Germany). We took six dark-adapted fluorescence measurements haphazardly on each plate during night time low tides. The dark-
adapted fluorescence value \((F_0)\) from the fluorometer serves as a proxy for the chlorophyll \(a\) density of
the MPB (Barranguet & Kromkamp 2000, Honeywill et al. 2002, Serodio et al. 2008). The tip of the
fiber-optic measuring head of the fluorometer was fitted with a 10 mm spacer to maintain a fixed
distance from the experimental plate, and the opening covered an area of 53 mm\(^2\). The tip was held in
place at each measurement site until the \(F_0\) value stabilized (typically 3-5 seconds) before recording a
value, as recommended by the manufacturer. Because changes in surface moisture can affect \(F_0\) values
(Maggi et al. 2013), we restricted sampling to periods when the plates were moist but not actively
splashed or submerged by the tide.

2-factor ANOVA and Tukey’s HSD test for multiple comparisons were used to compare dark-
adapted fluorescence for December 2013 across sites and grazing treatments. Temperature and dark-
adapted fluorescence are known to be negatively correlated (Miller et al. 2015); analysis of covariance
was used to determine whether the relationship between temperature and dark-adapted fluorescence
was influenced by grazing treatment. Visual checks were performed to ensure that model assumptions
of normality and equal variances were met. Analyses were carried out in MATLAB 2014a.

Scanning Electron Microscopy

After plates were removed from the field, we haphazardly selected and cut three 1 cm\(^2\) samples
of rubber tape from each plate. The samples were soaked in a 2.5% gluteraldehyde solution for 15-45
minutes to preserve cell shape, washed in phosphate-buffered solution (pH 7.9) for 15 minutes, rinsed
in de-ionized water and air dried. Samples were then gold-coated using a SPI-MODULE sputter coater
(Structure Probe Inc., West Chester, PA, USA), and were viewed under a scanning electron microscope
(SEM; FEI Quanta 200, hv = 25kV, spot size = 4 nm) at a magnification of 300x. This magnification
resulted in a 0.159 mm\(^2\) field of view. Nine fields were chosen haphazardly from each sample and
photographed, for a total of 648 images (162 images per treatment).
Identification and Quantification of Morphotypes

We processed each image using the photoQuad image analysis program ([http://www.mar.aegean.gr/sonarlab/photoquad/index.php](http://www.mar.aegean.gr/sonarlab/photoquad/index.php)). A spatially stratified, random field of 50 points was overlaid on the entire image, from which percent cover of the components of the MPB was calculated. Points that fell in areas that were not in focus were excluded from the analysis. We chose the magnification of 300x to visualize diatoms and cyanobacteria, which are the components of the MPB thought to be the preferred food sources for *Lottia* spp. (Castenholz 1961, Haven 1973, Nicotri 1977, Connor 1984). This resolution did not allow us to identify each organism at the species level. Instead, components of the MPB were classified by morphotypes, which took into account general shape, size and other features observable at 300x (Figure 1). These morphotypes were based on the organisms described, identified and pictured by Nagarkar and Williams (1997) and Hawkins et al. (1989). Morphotypes 1-5 were long, narrow organisms and were classified as filamentous cyanobacteria. Morphotypes 6-12 were smaller, boxy or rod-like organisms and were classified as diatoms. Organisms described as morphotype 13 formed encrusting mats with individual components that were impossible to distinguish at 300x. Morphotype 14 were also small encrusting organisms, but they grew in a characteristic diamond pattern. This pattern did not correspond in scale to the topographical features of the rubber grip tape or the radulae of *L. scabra* or *L. austrodigitalis*. Morphotype 15 were small, round organisms, and were classified as coccoidal cyanobacteria. Lastly, morphotype 16 described any other organism observed, while morphotype 17 indicated empty rubber grip tape.

From the random field of 50 points, percent cover of each morphotype was calculated as the number of points covering a particular morphotype divided by the total number of points (usually 50, minus any points not scored due to focus issues). Percent cover of organism type (filamentous...
cyanobacteria, diatoms, encrusting organisms or coccoidal cyanobacteria) was also calculated. We averaged percent cover of morphotypes and organism types within each plate \( (n = 27) \) and we calculated average percent cover of morphotypes and organism types for each grazing treatment across sites \( (n = 6) \). To determine a) how grazing by \( L. \) scabra and \( L. \) austrodigitalis affected MPB composition, and b) whether the two species were partitioning the MPB, we used PRIMER v6 software \( ($http://www.primer-e.com/) \) to generate MDS plots to visualize organism- and morphotype-level variation in assemblage structure among grazing treatments. Analysis of similarity (ANOSIM) was used to assess the relative size of these differences (Clarke & Warwick 2001). Because untransformed data place heavy emphasis on similarities in abundant organisms between assemblages (Clarke & Warwick 2001), square-root and fourth-root transformations of the raw data were also performed. These transformations put increasing emphasis on similarities in rare organisms between assemblages. Finally, the top contributors to differences in MPB composition were identified using similarity percentage tests (SIMPER; Clarke & Warwick 2001).

**Carbon and Nitrogen Elemental Analysis**

A second, larger field experiment was deployed from June 2014 through January of 2015. It was assembled as described above, except six replicate plates of seven grazing treatments were placed at each field site: an ungrazed treatment, three \( L. \) scabra-grazed treatments with four, eight or twelve limpets, and three \( L. \) austrodigitalis-grazed treatments with four, eight or twelve limpets. A subset of these plates with visually apparent MPB were used to evaluate the effects of limpet grazing on the nutritional content of the MPB.

Scrapings of the MPB were obtained from 34 ungrazed, 10 \( L. \) scabra-grazed and 9 \( L. \) austrodigitalis-grazed plates with sufficient MPB to permit sampling. To determine internal carbon and nitrogen concentrations, scrapings were dried to constant mass \( (60 \, ^\circ C \text{ for 72 hours}) \), ground to a
fine powder in stainless steel microvials (Retsch Mixer Mill MM 400, Verder Scientific, Newtown, Pennsylvania, USA) and analyzed on a Flash 2000 Elemental Analyzer configured for analyses of C and N in organic samples (Thermo Fisher, Cambridge, UK). Carbon-to-nitrogen (C:N) ratios were then calculated for each plate.

We tested a set of linear models using likelihood ratio tests to determine whether \( F_0 \), grazing treatment, average daily maximum temperature, or an interaction between these factors helped determine C:N ratios. Due to the relatively small number of limpet-grazed plates with visually apparent MPB growth for the C:N analysis, the number of limpets per plate for limpet-grazed treatments could not be included in our model. ANOVA and Tukey’s HSD tests were used to determine differences in log-transformed C:N ratios among grazing treatments. Visual checks were performed to ensure that model assumptions of normality and equal variances were met. Analyses were carried out in R 3.1.1.

**Effects of Limpet Density**

Although each limpet-grazed plate was initially seeded with four limpets in the 2013 experiment, some animals either escaped from or fell off the plates each month. This prompted us to explore the effects of limpet loss on the composition of the MPB. We examined the relationships between the number of limpets remaining on a plate the month prior to plate recovery from the field (December 2013) and the percent cover of morphotypes at recovery using ANOVA and Tukey’s HSD multiple comparisons test. Visual checks were performed to ensure that model assumptions of normality and equal variances were met. Analyses were carried out in MATLAB 2014a.
Results

Temperature

Grazing treatments in the 2013 experiment experienced average daily maximum temperatures that were statistically indistinguishable from each other (mean ± SE = 14.68 ± 0.42ºC; $F_{3,15} = 0.71$, $p = 0.561$). However, the average daily maximum temperature at site six (18.73 ± 0.51ºC) was significantly higher than that at the five other sites (13.87 ± 0.03ºC; $F_{5,15} = 20.62$, $p < 0.001$).

To determine whether temperature might be influencing the composition of the MPB, we divided our ungrazed plates into cool samples (average maximum daily temperature of less than 14ºC) and warm samples (average maximum daily temperature of 14ºC or greater). An analysis of similarity was performed. We found no evidence of a difference between the composition of the MPB on warmer and cooler ungrazed plates ($R = -0.179$, $p = 0.667$), although our sample size for this test was small ($n = 4$ cool plates, $n = 2$ warm plates). Additionally, the average difference in daily maximum temperature between warm and cool plates was only 3ºC.

Dark-adapted fluorescence

$F_0$, our proxy for microalgal density, was significantly higher for the ungrazed treatment than for the natural-grazed, $L. scabra$-grazed and $L. austrodigitalis$-grazed treatments ($F_{3,15} = 5.94$, $p = 0.005$) (Figure 2). Site was not a significant factor in this analysis ($F_{5,15} = 2.62$, $p = 0.068$). $F_0$ was affected by an interaction between grazing treatment and average maximum daily temperature ($F_{1,16} = 9.47$, $p < 0.001$); $F_0$ was inversely proportional to temperature for the ungrazed treatment (slope of linear model = -27.23 ºC$^{-1}$), but the two variables were uncorrelated for the three grazed treatments (Figure 3).
Organism-Level Assemblage Data

Analysis of similarity on untransformed data indicated that the MPB of the ungrazed treatment was significantly different from that of either the *L. scabra*-grazed treatment ($R = 0.284$, $p = 0.035$) or the *L. austrodigitalis*-grazed treatment ($R = 0.247$, $p = 0.048$) (Figure 4). Additionally, the MPB of the natural-grazed treatment was different from that of the *L. scabra*-grazed treatment ($R = 0.185$, $p = 0.041$), but not from the other treatments. No significant difference was observed between the *L. scabra* and *L. austrodigitalis*-grazed treatments ($R = 0.117$, $p = 0.123$). As increasing emphasis was placed on the presence or absence of rare organisms by square-root and fourth-root transforming the data, the ungrazed MPB differed more from the three grazed MPBs (Table 1).

Similarity percentage tests indicated that differences between the ungrazed MPB and the *L. scabra*-grazed MPB were driven by percent cover of filamentous cyanobacteria, diatoms and encrusting organisms. On average, the ungrazed MPB exhibited a higher percent cover of filamentous cyanobacteria and diatoms, and a lower percent cover of encrusting organisms (Table 2). Similar differences were identified between the ungrazed MPB and the *L. austrodigitalis*-grazed MPB, except these differences were also driven by percent cover of coccoidal cyanobacteria, which was more abundant in the *L. austrodigitalis*-grazed MPB. Differences between the natural-grazed MPB and the *L. scabra*-grazed MPB were driven by filamentous and coccoidal cyanobacteria. The former assemblage exhibited higher percent cover of both organisms relative to the latter.

Morphotype-Level Assemblage Data

Similar trends were exhibited at the morphotype level as those at the organism-level. The ungrazed treatment was significantly different from both the *L. scabra*-grazed treatment ($R = 0.559$, $p = 0.002$) and the *L. austrodigitalis*-grazed treatment ($R = 0.274$, $p = 0.006$) when analyses were done on untransformed data (Figure 5). Once again, the natural-grazed treatment and the *L. scabra*-grazed
treatment also differed ($R = 0.293, p = 0.039$), although this difference became non-significant after transforming the data (Table 1). The specific morphotypes driving these differences in composition were identified. Limpet-grazed treatments exhibited lower percent cover in morphotypes 2 and 5 (cyanobacteria), 8, 9 and 11 (diatoms) and 13 (encrusting organisms), but higher percent cover in morphotype 14 (encrusting organisms) (Table 2).

**Evidence for Resource Partitioning**

Both the organism-level and morphotype-level data indicate no significant differences in composition between the MPB grazed by *L. scabra* and that grazed by *L. austrodigitalis* (Table 1; $R = 0.117, p = 0.123$ and $R = -0.069, p = 0.652$ respectively). Emphasizing rare organisms by transforming the MPB composition data makes these communities appear more similar (Table 2). An MDS plot was generated using only *L. scabra*-grazed and *L. austrodigitalis*-grazed treatments that had the full quota of four limpets during the month prior to sample collection ($n = 8$ out of 12 limpet-grazed plates); after controlling for limpet density, there was no significant difference between *L. scabra* and *L. austrodigitalis*-grazed plates ($R = -0.031, p = 0.393$).

**Nutrient Concentrations and Ratios**

The best linear model describing trends in the log-transformed C:N ratio data included no significant interactions between $F_0$, average maximum daily temperature and the grazing treatment. However, C:N ratios were related independently to both $F_0$ and the grazing treatment ($F_{1,49} = 4.236, p = 0.045$ and $F_{2,49} = 12.977, p < 0.001$ respectively). C:N ratios were found to be negatively related to increased $F_0$, while *L. scabra*-grazed plates were found to be significantly depleted in nitrogen (C:N = $2.832 \pm 0.201$) relative to ungrazed plates (C:N = $2.180 \pm 0.029$), resulting in higher C:N ratios ($p < 0.001$) (Figure 6). *L. austrodigitalis*-grazed plates (C:N = $2.501 \pm 0.138$), however, were not
significantly different from either the *L. scabra*-grazed plates or the ungrazed plates (*p* = 0.0634, *p* = 0.127 respectively).

**Effects of Limpet Density**

Although each limpet-grazed plate was initially seeded with four limpets in the 2013 experiment, some animals either escaped from or fell off the plates each month. These animals were replaced during monthly surveys. The month prior to SEM sample collection (December 2013), no *L. scabra* were lost, but the *L. austrodigitalis* plates lost between 0 and 2 individuals, resulting in an average retention of $3.167 \pm 0.753$ limpets per plate (approximately 400 limpets m$^{-2}$). Significant correlations were found between the number of limpets on the plates at the end of December 2013 and the percent cover of coccoidal cyanobacteria and encrusting organisms (Figure 7). Percent cover of coccoidal cyanobacteria was highest at the intermediate densities of 2 or 3 limpets per plate. Percent cover of encrusting organisms was positively related to limpet density ($F_{10,12} = 5.200$, *p* = 0.046); however, at the morphotype level, morphotype 13 decreased with limpet density while morphotype 14 increased. Finally, during both the 2013 and 2014 experiments, significant negative correlations were found between $F_0$ and the number of limpets remaining on the plates during the final month of each experiment ($F_{1,10} = 5.57$, *p* = 0.04 and $F_{1,17} = 6.9$, *p* = 0.012 respectively). In 2013, increasing grazer density from approximately 250 limpets m$^{-2}$ (2 limpets per plate) to 400 limpets m$^{-2}$ (4 limpets per plate) resulted in a decrease in $F_0$ from 77.167 (n = 1) to 48.083 ± 1.977 (n = 8). In 2014, increasing grazer density from approximately 125 limpets m$^{-2}$ (1 limpet per plate) to 1000 limpets m$^{-2}$ (8 limpets per plate) resulted in a decrease in $F_0$ from 82.667 (n = 1) to 17.833 (n = 1).
Discussion

On the ungrazed plates, high average maximum daily temperatures were correlated with low $F_0$ values. MPB is reduced in biomass and therefore less fluorescent when exposed to warm conditions (Jackson et al. 2010, Miller et al. 2015). However, limpet grazing reduced $F_0$ values to those typical of warm, ungrazed plates, regardless of temperature, suggesting that in intertidal regions where grazers are very sparse, $F_0$ values will be low if the region is warm, and high if the region is cool. The presence of grazers masks this signal, corroborating prior observations that limpet grazing suppresses natural variation in the density of the MPB along intertidal shores (Miller et al. 2015). Our results also indicate that a low $F_0$ value on a warm, ungrazed plate does not necessarily signify a change in assemblage structure at the microscopic level. However, a more extensive study designed to observe the composition of the MPB across a larger range of temperatures and with higher replication would better address this question.

Dark-adapted fluorescence ($F_0$), our proxy for chlorophyll $a$ density within the MPB, was significantly higher on ungrazed plates than on grazed plates, indicating that intertidal grazers reduce the abundance of photosynthesizing organisms in the MPB. However, the average grazed $F_0$ (mean ± SE = 55.8 ± 18.22) was much greater than the zero value obtained from new clean safety walk grip tape, suggesting that photosynthesizing organisms are still plentiful even under constant grazing pressure. Small organisms that limpets are unable to graze may be the primary contributors to this $F_0$ value, and it may represent a baseline signifying the limit to which grazers can deplete food resources. Herbivorous intertidal sea stars, in addition to a variety of gastropod grazers, have been found to have limited ability to deplete epilithic MPB, possibly because certain components are inaccessible, toxic or
indigestible (Nicotri 1977, Underwood 1984, Jackson et al. 2009). However, higher densities of *L. scabra* and *L. austrodigitalis* than our experimental density of about 500 limpets m$^{-2}$ might graze down photosynthesizing organisms even further. Our data from 2014 indicated that densities of 1000 limpets m$^{-2}$ should deplete $F_0$ to about half the level observed at 500 limpets m$^{-2}$. However, this observation was based on data from a single plate. Excluding this sample, the negative correlation between limpet density and $F_0$ was not statistically significant; limpet densities between 125 and 500 m$^{-2}$ grazed the MPB down to an average $F_0$ value of 44.5 ± 0.9. It is worth noting that all of these limpet densities have been observed at HMS. Although Morelissen and Harley (2007) observed 515 *L. scabra* m$^{-2}$ in limpet-rich regions of the intertidal zone at HMS, both *L. scabra* and *L. austrodigitalis* have clumpy distributions that can sometimes result in even higher local densities (D. LaScala-Gruenewald, personal observation).

Effects of Grazers and Grazer Density

The composition of grazed MPB was significantly different from that of ungrazed MPB. The limpet-grazed MPB contained fewer diatoms and filamentous cyanobacteria in particular, which may be the preferred food sources of *Lottia* spp. (Nicotri 1977, Connor 1984, Haven 1973, Hahn & Denny 1989). On plates where fewer than four limpets grazed, there were higher percentages of coccoidal cyanobacteria, and lower percentages of encrusting organisms. This suggests an ordered grazing scheme, where filamentous cyanobacteria and diatoms are consumed first, followed by coccoidal cyanobacteria. This pattern may be due to animal preference (Schmitt 1996), but it is more likely due to topographical constraints. Lateral teeth in *L. scabra* and *L. austrodigitalis* are typically 100 μm long and 50 μm wide at the widest point, while tooth tips are typically separated by distances of 100 μm in the cross-radula direction and 200 μm in the along-radula direction (D. LaScala-Gruenewald, unpublished data). In contrast, coccoidal cyanobacteria and some other organisms common on grazed
plates (morphotypes 5 and 9 for example) are under 30 μm in maximum dimension, and are possibly
difficult to consume, especially if the substratum is rugose at the μm-scale.

Plates open to the natural suite of upper intertidal grazers had compositions that were
intermediate between ungrazed and limpet-grazed plates. Although the number and species of grazers
on the natural-grazed plates were not monitored throughout the experiment, both Littorina spp. and
Lottia spp. were capable of ascending the 12 mm plate edge (D. LaScala-Gruenewald, personal
observation), and the former were occasionally observed foraging on the plates during monthly
surveys. Because no limpets were confined on the natural-grazed plates, they probably experienced
reduced grazing pressure, resulting in higher percent cover of filamentous cyanobacteria and diatoms
relative to the limpet-grazed plates. These results indicate that, at HMS, we should expect to see
assemblages of primarily encrusted organisms where limpets are very dense, and assemblages of
primarily filamentous cyanobacteria and diatoms in regions where limpets and other grazers are very
sparse.

Evidence for Resource Partitioning

No significant differences were observed between the composition of the MPB grazed by L.
scabra and that grazed by L. austrodigitalis. This result is consistent with previous findings (Haven
1973) indicating that these two species are in direct competition for food resources, and fails to support
our hypothesis of complementarity. However, microscopic-level analyses of the MPB may be
important in determining resource partitioning in other scenarios. L. scabra and L. austrodigitalis have
very similar morphology; in fact, their radulae show only very subtle differences in lateral tooth shape,
and in the length of their ribbon segments (Lindberg 1981, D. LaScala-Gruenewald personal
observation). Complementarity might still be observed if a wider array of grazers were considered. In
particular, the abundant upper intertidal periwinkle, Littorina keenae Phillippi, which was not included
in this experiment, has distinctly different radula morphology, radula hardness, and feeding behavior, and may be able to access and consume components of the MPB that are unavailable to limpets (Goodwin & Fish 1977, Hawkins et al. 1989).

Effects of Grazing on Nutrient Ratios

Limpet grazing also affected the C:N ratio of the MPB. Cyanobacteria have been hypothesized to be nutritionally inferior to diatoms (Hargrave 1970, Nicotri 1977), and we found that the ratio of the percent cover of diatoms to the percent cover of cyanobacteria was significantly reduced in MPB grazed by *L. scabra* and *L. austrodigitalis* (ANOVA; $F_{3,20} = 4.150$, $p = 0.019$). *L. scabra*-grazed samples also had significantly increased C:N ratios relative to ungrazed samples, indicating preferential consumption of high-nitrogen portions of the MPB. However, *L. austrodigitalis*-grazed samples had C:N ratios that were intermediate between *L. scabra*-grazed and ungrazed samples. Due to the relatively low number of limpet-grazed plates that had sufficient coverage of MPB to be used for the C:N analysis, we grouped plates foraged by four, eight and twelve limpets, which raises the possibility that differences in C:N ratios on limpet-grazed plates were driven by limpet density. The *L. austrodigitalis*-grazed plates suitable for C:N analysis were initially seeded with an average of 8.4 limpets while the *L. scabra*-grazed plates were seeded only with 4.4 limpets, but by the end of our study, the number of limpets on *L. austrodigitalis* and *L. scabra*-grazed plates were statistically indistinguishable (3.53 ± 0.35). This was likely due to the fact that *L. austrodigitalis* have a higher propensity for escaping our field enclosures than *L. scabra* (Miller et al. 2015). Additionally, no relationship was found between the number of limpets remaining on the plates at the end of the study and C:N ratio.

Prior research does not indicate that cyanobacteria have increased C:N ratios relative to diatoms; both groups of organisms tend to have C:N ratios between 3 and 30 (Brzezinski 1985, Fukuda
et al. 1998, Geider & La Roche 2002, Fu et al. 2007). However, C:N ratios are species- and location-
specific, and vary with nutrient availability and physical parameters such as temperature (Geider & La
Roche 2002). More specific data on C:N ratios of diatoms and cyanobacteria native to the upper
intertidal zone at HMS would be necessary to better evaluate our hypothesis of depleted nutritional
quality.

Relevance and Implications

Our results corroborate previous work investigating potential partitioning of the MPB by
intertidal gastropods. Nicotri (1977) showed that the limpets L. pelta, L. strigatella and L. scutum, in
addition to the littorine snail L. scutulata, did not partition food resources at the microscopic scale.
Similarly, Hawkins et al. (1989) examined three European gastropods, G. umbilicalis, L. littoreia and P.
vulgata, and found that their diets were indistinguishable despite differences in radula structure and
feeding movements. The species in Nicotris (1977) study occupy different microenvironments, and
both studies considered some species that supplement the MPB in their diets with macroalgae.
However, it appears that their results are generalizable to species that compete in the field for the
entirety of their food resources. This is somewhat surprising: if food is limiting in the upper intertidal
environment (Haven 1973, Branch & Branch 1981, Underwood 1979, Underwood 1984a, b, c), there
should be strong selective pressure favoring individuals who can partition the MPB. Nicotri suggests
that the MPB may be too difficult to partition successfully, and the relative sizes of lateral teeth, ventral
plates and maximum dimension of the components of the MPB considered here are consistent with that
conclusion. However, Lottia spp. may still be partitioning resources with the littorine snails Littorina
keenae, L. scutulata and L. plena, which graze on the MPB in the upper intertidal environment at HMS
but have different radula structure. It would be interesting to observe whether the overlaps in diet
reported by Nicotri (1977) and Hawkins (1989) are also true for L. austrodigitalis, L. scabra and
In general, we conclude that *L. scabra* and *L. austrodigitalis* are in direct competition for resources, and that their density affects the composition of the MPB. However, this conclusion has a number of caveats. First, although a large number of SEM images were taken for each treatment, the percent cover of each plate that was imaged was very small (<1%). It is possible that the lack of significant differences between grazing treatments reported in our results was affected by low sampling area, although we doubt that our general conclusions would change given wider sampling. Additionally, we were unable to identify organisms to the species level. Again, doing so would be unlikely to change our general results, but more information might be provided by performing genomic analyses to determine the species involved. Finally, all of our experiments took place on rubber grip tape rather than the limpets natural rock habitat. While this difference in substratum likely influenced the composition of both the ungrazed and grazed MPB, it allowed for consistency across sites and treatments. The native substratum at HMS is granite, an igneous rock containing feldspar, quartz and an array of other rock types that vary based on where and when the rock was formed. Each of these rock types has unique chemical and physical properties, which may affect patterns of growth in the MPB (Cattaneo-Vietti et al. 2002, Faimali et al. 2004, Perotti 2008). The granite at HMS is also topographically variable on scales ranging from meters to millimeters, and topography has been shown to influence the settlement and growth of MPB (Johnson 1994, Schmitt 1996, Keough et al. 1997, Hutchinson et al. 2006). Using rubber grip tape as the substratum for this experiment eliminated potential variability due to rock type and topography, which might otherwise have masked differences in the composition of the MPB due to grazing. Rubber grip tape as an experimental substratum has some drawbacks. Although it is highly rugose across small scales, it does not mimic the larger (5-10 mm) topographic features typical of the native granite substratum at HMS. Additionally, granite is much harder than rubber tape. Prior
research has found that it takes less force to remove crustose algae from a harder surface (Padilla 1985), suggesting that limpets might feed more efficiently on granite. However, granite cannot be deformed by a radula, and limpets on granite may not be able to completely remove the MPB from small-scale topographical features. It is likely that both the rugosity and flexibility of rubber tape influenced the MPB composition observed on both grazed and ungrazed plates.

The deposition of pedal mucus by limpet grazers may also have had an effect on the compositions of the observed MPB. Limpet locomotion leaves behind a thin mucus layer that, in some species, has been shown to persist in the field and attract components of the MPB relative to adjacent mucus-free substratum (Davies & Hawkins 1998). Connor and Quinn (1984) and Connor (1986) examined the capacity for mucus from *L. scabra* and *L. austrodigitalis* to act as a persistent adhesive trap for components of the MPB. They found that mucus from both species do recruit components of the MPB at +1.7 m above MLLW in the intertidal zone adjacent to Bodega Bay Marine Laboratory in Sonoma County, California, but mucus from *L. scabra* was more effective in doing so. Additionally, they found that the median persistence time for trails from both species in the field was 7 days.

Research on another limpet species, *Patella vulgata*, indicated that its mucus was especially effective in trapping diatoms relative to other components of the MPB, and especially effective during the first 24 hours after deposition (Davies et al. 1992). It is highly likely that our limpet-grazed plates were coated in pedal mucus throughout the duration of our field study, and that this mucus recruited additional MPB to grazed areas. However, the effects of this mucus were not sufficient to counter the effects of grazing. *L. scabra* and *L. austrodigitalis*-grazed plates showed statistically indistinguishable percentages of diatoms and cyanobacteria, and dark-adapted fluorescence. Additionally, *L. scabra* exhibited a significantly lower growth rate than *L. austrodigitalis* throughout the study, which fails to reflect a relative increase in food consumed (D. LaScala-Gruenewald, unpublished data). Differences in mucus properties between our study species were likely masked by grazing intensity. Both limpets
cover an average area of about 50 cm² during a foraging bout (D. LaScala-Gruenewald, unpublished data); the plate area (78.54 cm²) was too small to accommodate four limpets without regrazing. The fact that *L. scabra* and *L. austrodigitalis* appear not to partition resources may have implications in the face of future climate change. Prior research indicates that moderate increases in temperature may create a temporary benefit for ectothermic organisms such as limpets (Miller et al. 2015). Increases in limpet growth rate have been observed at higher temperatures, and it has been hypothesized that either temperature effects on physiological processes or a shift towards consuming higher value food items may be mechanistically responsible (Miller et al. 2015). Our data indicate that the former is more likely, since the composition of MPB does not appear to vary across a small temperature range (13-16°C). Provided that sufficient food resources are available, moderate increases in temperature during aerial exposure should result in increases in limpet metabolism and growth (Sanford 2002, Miller et al. 2015). However, our study also suggests that it may be possible for moderate densities of limpets (200 – 500 m⁻²) to reduce the MPB to a level where little or no food is available. Furthermore, the MPB that remains is of lower quality (i.e., higher C:N). Limpets under future climate change conditions may not be able to obtain sufficient resources at current spatial densities, and we may observe declines in densities of upper-intertidal limpets. We might also expect to see more success from organisms that do partition resources; complementarity has been documented between a number of intertidal organisms (e.g. Byrnes & Stachowicz 2009, Griffin et al. 2008, Sanz-Lázaro et al. 2015) and is a leading explanation for how assemblages may cope with a changing climate (Loreau et al. 2001). Additionally, because increased temperatures correspond to decreases in $F₀$, less food may be available as the climate warms, particularly in the summer. The density and composition of the MPB vary seasonally (Underwood 1984, Jackson et al. 2010), and our samples for this study were collected in December, when the MPB biomass was relatively high. As the MPB decreases over the summer months, there will be a corresponding decrease in food availability.
(Thompson 2000). Individuals with limited access to resources are at greater risk of temperature stress, as they are less able to mitigate potential costs through physiological mechanisms such as the heat shock response (Fitzgerald-DeHoog et al. 2012).

Although *L. scabra* and *L. austrodigitalis* do not appear to partition resources at the microscopic level, the density of grazers does have an effect on the composition of the MPB. Both species consume large filamentous cyanobacteria and diatoms before eating the smaller filamentous and coccoidal cyanobacteria. Therefore, we should expect to see MPB relatively rich in filamentous cyanobacteria and diatoms in regions where limpets are sparse. It is also likely that small filamentous and coccoidal cyanobacteria form some baseline density of the MPB that is unavailable to upper intertidal grazers. Prior research suggests that differences in feeding behavior and radula morphology between limpet species may result in differing abilities to graze on certain species of macroalgae (Padilla 1985). Similarly, *L. scabra* and *L. austrodigitalis* may graze on certain morphotypes in the MPB first because of preference, or because of radula morphology and function. The microtopography of the substratum may also play a role. Hutchinson et al. (2006) examined the effects of substratum roughness on the composition of the MPB and molluscan grazing patterns using SEM. Diatoms were more abundant on rougher substrata, and both diatoms and filamentous cyanobacteria were more abundant around surface features. Finally, different species of molluscan grazers varied in their abilities to remove certain diatoms and cyanobacteria. It is likely that an interaction between grazing behavior, radula morphology and physical properties of the substratum (such as roughness and hardness) are responsible for our observed grazing patterns.

In summary, although the MPB contributes strongly to the primary productivity that fuels the upper intertidal ecosystem (Nicotri 1977, Underwood 1984, Thompson et al. 2000), relatively few studies have considered the composition of intertidal MPB at the microscopic level (Nagarkar & Williams 1997, Hill & Hawkins 1990, Hutchinson et al. 2006), and even fewer have examined the
impacts of environmental factors on the composition of the MPB (but see Nicotri 1977, Underwood 1984, Thompson et al. 2004). Here we found that two limpet species, *L. scabra* and *L. austrodigitalis*, appear to compete directly for food resources even at the microscopic scale, and that grazer density influences the composition of the MPB. C:N ratios on grazed and ungrazed plates suggest that limpet grazing may reduce the nutritional content of the MPB via selective consumption of high-N organisms. Additionally, we found that moderate increases in temperature decreased the density of the MPB, but did not change its composition. As the climate continues to warm, we expect to see decreases in the density of the MPB and consequent decreases in grazer density, especially among species that do not partition resources at the microscopic level.

### Acknowledgments

We thank G. Bernatchez for assistance with elemental analyses and E. Duncan, F. King, J. McNamara, B. Pagan and C. Prince for assistance in setting up and maintaining the experiment in the field. This work was funded by NSF grants OCE-1131038 and 1130095 to B. J. Allen and M. W. Denny.

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Table 1. Transforming MPB composition data results in larger differences between grazed and ungrazed assemblages. P-values are displayed from analysis of similarity tests for the treatments being compared when the data underwent no transformation, a square-root transformation, and a fourth-root transformation. These transformations place increasing emphasis on the presence of rare species in assemblages. Data from both the organism-level analysis and morphotype-level analysis are shown. Significant p-values are in bold type; associated R-values are in parentheses.

<table>
<thead>
<tr>
<th>Treatments being compared</th>
<th>Organism-level</th>
<th>Morphotype-level</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No transformation</td>
<td>Square-root</td>
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<tr>
<td>Ungrazed and Natural</td>
<td>0.093 (0.127)</td>
<td>0.087 (0.159)</td>
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<tr>
<td>Ungrazed and L. scabra</td>
<td>0.035 (0.247)</td>
<td>0.028 (0.285)</td>
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<tr>
<td>Ungrazed and L. austrodigitalis</td>
<td>0.048 (0.185)</td>
<td>0.069 (0.191)</td>
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<tr>
<td>Natural and L. scabra</td>
<td>0.766 (-0.068)</td>
<td>0.814 (-0.076)</td>
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<tr>
<td>Natural and L. austrodigitalis</td>
<td>0.123 (0.117)</td>
<td>0.249 (0.069)</td>
</tr>
</tbody>
</table>
Table 2. Comparison of percent cover of organisms and morphotypes across grazing treatments.

In general, grazed treatments showed fewer filamentous cyanobacteria (morphotypes 2 and 5) and diatoms (morphotypes 8, 9 and 11) than the ungrazed treatment. Percent cover of encrusting organisms were also higher on average, due to the larger percentage of morphotype 14. Coccoidal cyanobacteria (morphotype 15) were higher in the natural-grazed and *L. austrodigitalis*-grazed treatments.

Organisms that were relatively abundant in grazed treatments are in bold type, while those that were relatively scarce in grazed treatments are italicized.

<table>
<thead>
<tr>
<th>Organism-level</th>
<th>Ungrazed</th>
<th>Natural</th>
<th><em>L. scabra</em></th>
<th><em>L. austrodigitalis</em></th>
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<td>Filamentous cyanobacteria</td>
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<td>16.00</td>
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<td>Encrusting organisms</td>
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<td>71.83</td>
<td>84.17</td>
<td>76.50</td>
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<td>Coccoidal cyanobacteria</td>
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<td>9.17</td>
<td>4.67</td>
<td>9.50</td>
</tr>
<tr>
<td>Diatoms</td>
<td>10.00</td>
<td>3.00</td>
<td>--</td>
<td>1.00</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Morphotype-level</th>
<th>Ungrazed</th>
<th>Natural</th>
<th><em>L. scabra</em></th>
<th><em>L. austrodigitalis</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>2</td>
<td>1.60</td>
<td>0.02</td>
<td>0.05</td>
<td>0.02</td>
</tr>
<tr>
<td>5</td>
<td>17.11</td>
<td>15.15</td>
<td>8.92</td>
<td>11.56</td>
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<tr>
<td>8</td>
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<tr>
<td>9</td>
<td>6.18</td>
<td>1.11</td>
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<tr>
<td>11</td>
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<td>13</td>
<td>60.53</td>
<td>58.76</td>
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<td>14</td>
<td>3.09</td>
<td>12.83</td>
<td>39.05</td>
<td>32.21</td>
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</table>
Figure 1. Descriptions and images of morphotypes observed in the MPB. Each morphotype is identified by a number (1-17), and pictured above at 300x unless otherwise indicated. Scale bars are
10µm unless otherwise indicated, circles highlight the specific organism being described and all measurements provided are approximate. Arrows point to features referred to in the right-hand panel. The bottom panel includes each morphotype number, in addition to the organisms broader classification (filamentous cyanobacteria, diatom, encrusting organism or coccoidal cyanobacteria) and a summary of the shape and size details used to identify the organism.
Figure 2. The effect of grazing treatment on $F_0$. The *L. scabra*-grazed, *L. austrodigitalis*-grazed and natural-grazed treatments exhibited decreased chlorophyll density ($F_0 = 55.2 \pm 5.2$) relative to the ungrazed treatment ($F_0 = 117.5 \pm 22.2$) during the month prior to SEM sample collection. Black lines indicate the mean values for each treatment, grey rectangles represent one standard deviation and white rectangles represent 95% confidence intervals (n = 6 for all treatments). Letters indicate significantly different $F_0$ values, as determined by Tukey’s HSD test for multiple comparisons.
Figure 3. Correlations between average maximum daily temperature and $F_0$ differed between treatments. Average maximum daily temperature and $F_0$ were significantly more negatively correlated on ungrazed plates (n = 6) than on grazed plates (n = 18, $F_{3,16} = 9.47$, p < 0.001). Temperatures above 16ºC reduced $F_0$ values on ungrazed plates to those typical of grazed plates. Grazing masked this effect of temperature, keeping $F_0$ values at about half of those on ungrazed plates even when temperatures were less than 16ºC.
Figure 4. Ungrazed MPB was significantly different from grazed MPB at the organism level. The assemblage structure of the ungrazed treatment was significantly different from that of the *L. scabra*-grazed and *L. austrodigitalis*-grazed treatments when considering untransformed data ($R = 0.284$, $p = 0.035$ and $R = 0.247$, $p = 0.048$ respectively); the natural-grazed treatment was significantly different from the *L. scabra*-grazed but not the *L. austrodigitalis*-grazed treatment ($R = 0.185$, $p = 0.041$ and $R = -0.068$, $p = 0.766$ respectively); and no significant difference was observed between *L. scabra* and *L. austrodigitalis*-grazed assemblages ($R = 0.117$, $p = 0.123$).
Figure 5. Ungrazed MPB was significantly different from grazed MPB at the morphotype level. The assemblage structure of the ungrazed treatment was significantly different from that of the *L. scabra*-grazed and *L. austrodigitalis*-grazed treatments when considering untransformed data (R = 0.559, p = 0.002 and R = 0.274, p = 0.006 respectively); the natural-grazed treatment was significantly different from the *L. scabra*-grazed but not the *L. austrodigitalis*-grazed treatments (R = 0.293, p = 0.039 and R = 0.078, p = 0.229 respectively); and no significant difference was observed between *L. scabra* and *L. austrodigitalis*-grazed assemblages (R = -0.069, p = 0.652).
**Figure 6. Effects of grazing treatment on C:N ratio.** Grazing by *L. scabra* significantly increased the log(C:N) ratio of the MPB ($F_{2,45} = 12.98$, $p < 0.001$). However, C:N ratios of *L. austrodigitalis*-grazed samples were intermediate between ungrazed and *L. scabra*-grazed samples, and were not significantly different from either sample. Site was not a significant factor in this analysis ($F_{5,45} = 1.38$, $p = 0.248$), and Tukey’s HSD multiple comparisons test was used to determine differences between grazing treatments. Black lines indicate the mean values for each treatment, grey rectangles represent one standard deviation and white rectangles represent 95% confidence intervals ($n_{\text{ungrazed}} = 34$, $n_{\text{L. scabra-grazed}} = 10$, $n_{\text{L. austrodigitalis-grazed}} = 9$). Letters indicate significantly different C:N ratios.
Figure 7. Effects of number of grazers on percent cover of selected morphotypes. On average, more limpet grazers resulted in lower percent cover of encrusting organisms. This was primarily due to increases in morphotype 14; morphotype 13 decreased as the number of grazers increased. Diatoms (morphotypes 8 and 9) were present in very low percentages if any grazers were present. The percent cover of filamentous cyanobacteria (morphotype 5) was, on average, reduced on grazed plates, but exhibited no correlation with number of grazers. Finally, percent cover of coccoidal cyanobacteria (morphotype 15) was highest at intermediate numbers of grazers. Tukey’s HSC multiple comparisons test was used to determine differences between limpet densities. Black lines indicate the mean values for each treatment, grey rectangles represent one standard deviation and white rectangles represent...
942 95% confidence intervals ($n_0 = 6, n_2 = 1, n_3 = 3, n_4 = 8$). Letters indicate significantly different percent covers of morphotypes.
## Supplementary Materials

### Table S1. ANOVA table for Figure 4.

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### Table S2. ANOVA table for Figure 5.

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### Table S3. ANOVA table for Figure 7.

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