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# Association of a *MET* genetic variant with autism-associated maternal autoantibodies to fetal brain proteins and cytokine expression

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The contribution of peripheral immunity to autism spectrum disorders (ASDs) risk is debated and poorly understood. Some mothers of children with ASD have autoantibodies that react to fetal brain proteins, raising the possibility that a subset of ASD cases may be associated with a maternal antibody response during gestation. The mechanism by which the maternal immune system breaks tolerance has not been addressed. We hypothesized that the mechanism may involve decreased expression of the MET receptor tyrosine kinase, an ASD risk gene that also serves as a key negative regulator of immune responsiveness. In a sample of 365 mothers, including 202 mothers of children with ASD, the functional *MET* promoter variant rs1858830 C allele was strongly associated with the presence of an ASD-specific 37 + 73-kDa band pattern of maternal autoantibodies to fetal brain proteins (P = 0.003). To determine the mechanism of this genetic association, we measured MET protein and cytokine production in freshly prepared peripheral blood mononuclear cells from 76 mothers of ASD and typically developing children. The *MET* rs1858830 C allele was significantly associated with MET protein expression (P = 0.025). Moreover, decreased expression of the regulatory cytokine IL-10 was associated with both the *MET* gene C allele (P = 0.001) and reduced MET protein levels (P = 0.002). These results indicate genetic distinction among mothers who produce ASD-associated antibodies to fetal brain proteins, and suggest a potential mechanism for how a genetically determined decrease in MET protein production may lead to a reduction in immune regulation.

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# Introduction

Autism spectrum disorders (ASDs) are a complex, behaviorally defined group of developmental disorders characterized by social deficits, language impairments and repetitive behaviors with restricted interests. Previous studies of twins and sibling recurrence risk have suggested that ASD is among the most heritable of neuropsychiatric disorders, encouraging extensive research into the genetic basis of ASD risk. 1-3 However, a more recent twin study reported that susceptibility to ASD has a much lower genetic heritability component, while the more substantial component of risk was explained by environmental factors in a shared twin environment.4 This would indicate that the gestational environment, susceptible to genetic regulation by both the fetus and the mother, might be a major factor in determining ASD risk. The genetic risk may lie in (a) the direct impact of genetic regulation on brain development, (b) altering sensitivities to environmental factors and (c) disruption of autonomic and peripheral organ development and function that may influence brain development. In this regard, there is a growing body of evidence that, among risk factors, immune system dysregulation may be a feature in this neurodevelopmental disorder.5-7

Growing evidence indicates that maternal immune dysfunction during gestation may contribute to ASD risk. In animal models, maternal immune activation during gestation leads to behavioral changes in subsequent offspring.8 In humans, epidemiologic analyses reported that a family history of autoimmune disorders was more common in families of children with ASD, 9,10 and that immune-mediated disorders (psoriasis, maternal asthma and allergy) occurred more often around the time of pregnancy in the mothers of children with ASD compared with the mothers with typically developing (TD) children. 11 One early report demonstrated that serum from a mother of a child with ASD showed IgG antibody reactivity to rodent Purkinje cells; this same serum was then injected into gestating mice and shown to elicit behavioral deficits in the offspring. 12 In another study, 11 mothers of children with ASD and their affected children were found to have consistent patterns of antibody reactivity against rat prenatal brain proteins. 13 Further studies have highlighted that this reactivity is to fetal and not adult brain proteins. 14 Our recent studies have demonstrated reactivity against fetal brain proteins at  $\sim 37$  and  $73\,\mathrm{kDa}$  in plasma from 7 of 61 (11.5%) mothers of children with ASD but not in 102 mothers of TD children. 15 Injection of rhesus monkeys with human IgG

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from mothers of children with ASD (who were positive for fetal brain antibodies) during gestation resulted in increased stereotypical behavior and hyperactivity in the offspring when compared with the offspring of rhesus monkeys injected with IgG from mothers of TD children. Evidence of a potential role for these antibodies with respect to changes in neurodevelopment was also noted in a recent murine model. To

Although the preponderance of evidence points to an immune-mediated etiology in some cases of ASD, the biological mechanism of the maternal immune dysfunction remains unclear. In an independent line of study, we described robust genetic association with ASD of a functional variant in the 5' promoter of the gene encoding the MET receptor tyrosine kinase. 18 The MET promoter variant, rs1858830, is a common G-to-C single-nucleotide polymorphism (SNP); the 'C' allele is inherited by individuals with ASD more often than predicted by chance and is more common in individuals with ASD than in a sample of the general population. 18-20 Stratifying ASD subpopulations demonstrated that association of the MET 'C' allele is enriched in individuals with co-occurring ASD and gastrointestinal conditions,21 and in the social and communication domains of ASD.<sup>22</sup> Independent evidence indicates decreased expression of the ligand for the MET receptor, hepatocyte growth factor (HGF), in serum of individuals with co-occurring ASD and gastrointestinal conditions.<sup>23</sup> In addition to its roles in brain development and gastrointestinal repair, the MET receptor tyrosine kinase is a key negative regulator of immune responsiveness. Ligation of the MET receptor by its HGF ligand prevents lupus nephritis in chronic graft-versus-host disease,24 ameliorates the progression of experimental autoimmune myocarditis<sup>25</sup> and suppresses collagen-induced arthritis in mice. 26 MET also induces morphogenic changes in antigen-presenting cells. Importantly, MET signaling induces a tolerogenic phenotype in antigen-presenting cells through the induction of IL-10, without affecting their antigen-presenting capabilities.<sup>27,28</sup> This tolerogenic phenotype induces T regulatory cells in response to antigen that would otherwise have broken tolerance and potentially lead to autoimmunity.<sup>28</sup> MET protein expression is decreased in postmortem brain of individuals with the MET 'C' allele.29 If the same genotypeexpression correlation is observed in the immune system, then one prediction is that disrupted MET signaling may confer susceptibility to immune dysregulation. We hypothesized that MET disruption would be associated with markers of immune dysregulation in mothers of children with ASD. To test this hypothesis, we examined the MET promoter variant rs1858830 'C' allele in the context of ASD-specific maternal antibodies to fetal brain proteins, as well as the cytokine profile of peripheral blood cells following immune challenge.

# Materials and methods

**Subjects.** All mothers, (n=365) including those of children with an ASD (n=202) and those of TD control children (n=163), participated in the Childhood Autism Risks from Genetics and the Environment (CHARGE) study.<sup>30</sup> Those children enrolled in the study met the following criteria: (a)

they were between the ages of 24 and 60 months at the time of enrollment, (b) lived with at least one biologic parent, (c) had a parent who spoke either English or Spanish, (d) were born in the state of California and (e) resided in the catchment areas of a specified list of regional centers in Northern California. The CHARGE study is an ongoing population-based case—control study. This study protocol followed the ethical guidelines of the most recent Declaration of Helsinki, and was approved by the institutional review boards at the University of California, Davis, The State of California Department of Developmental Services, and the University of Southern California. Informed consent was obtained before participation.

To confirm the initial diagnosis, all children enrolled in the study were assessed at the UC Davis M.I.N.D. Institute. The diagnosis of ASD was confirmed in all subjects using the standardized Autism Diagnostic Interview-Revised (ADI-R)31 and the Autism Diagnostic Observation Schedule (ADOS) modules 1 or 2.32-35 The ADI-R provides a standardized, semi-structured interview and a diagnostic algorithm for the DSM-IV<sup>36</sup> and ICD-10 definitions of autism.<sup>37</sup> All of the CHARGE study clinical assessment personnel have attained research reliability on the ADI-R and the ADOS. The Social Communication Questionnaire was used to screen for behavioral and developmental characteristics of ASD among TD controls: children who scored above the screening cutoff score of 15 were fully assessed using the ADI-R and ADOS. Those controls that met criteria for autistic disorder were classified as ASD. Controls who did not meet criteria for ASD were classified as TD.

**Autoantibodies.** Western blot data collected and analyzed in a previous study was used for the association analyses described herein. Western blots were performed as previously described using fetal monkey brain extract. 15,38

Genotyping the ASD-associated MET promoter variant rs1858830. As the rs1858830 SNP falls within a highly GCrich region, indirect genotyping methods fail when using genomic DNA. A 652-bp fragment containing the rs1858830 SNP was amplified from 15 ng genomic DNA with primers 5'-GATTTCCCTCTGGGTGGTG-3' (Forward) and 5'-CAAGC CCCATTCTAGTTTCG-3' (Reverse). Each 20-µl reaction included 15 ng genomic DNA, 1 µM each primer, 5% dimethyl sulfoxide, 15 mm MgCl<sub>2</sub>, 0.25 mm each dNTP (dATP, dCTP, dGTP and dTTP), 2.5 U AmpliTag Gold DNA Polymerase (ABI, Carlsbad, CA, USA) and 0.2 U Turbo pfu DNA polymerase (Stratagene, Santa Clara, CA, USA). Cycling conditions were: 95°C for 5 min followed by 35 cycles of 95 °C for 30 s, 61 °C for 30 s and 72 °C for 1 min. Specific amplification of the 652-bp product was confirmed by agarose gel electrophoresis. Each PCR product was subjected to direct re-sequencing using an ABI 3730xl using Big Dye Terminator chemistry. Genotype at the MET rs1858830 locus was determined from the sequencing result using Sequencher software (Gene Codes, Ann Arbor, MI, USA).

Cell culture and stimulation. CHARGE is an ongoing study for which we have collected and banked both plasma



and DNA that allowed us to retroactively analyze subjects for both autoantibody production and genotype at the MET rs1858830 locus. However, cytokine production in response to immune challenge must be done on freshly isolated peripheral blood mononuclear cells (PBMCs). Therefore, a random subset of 76 mothers (22 C/C, 33 C/G and 21 G/G), prospective to study initiation, was used to assess the functional cytokine response in relation to MET rs1858830 genotype. Whole blood was collected through venipuncture into yellow top citrate tubes (BD, Franklin Lakes, NJ, USA) according to study protocol, and centrifuged at 900 a for 10 min to pellet cells. Plasma was collected and immediately frozen in 0.5 ml aliquots at -80 °C until assayed by western blot for the presence of anti-fetal brain antibodies. The cell pellet was then adjusted to appropriate density with HBSS and layered over Histopaque density gradient (Sigma, St Louis, MO, USA) followed by centrifugation at 600 g for 30 min. The PBMC layer was removed and washed two additional times at 900 g for 10 min at which point cells were counted with a hemocytometer, and 300 000 cells per well were plated into 96-well round bottom plates. Cells were then allowed to rest 24h before administration of 5 µg ml<sup>-1</sup> lipopolysaccharide for 48 h to induce the production of an innate immune response. After stimulation, cell culture supernatants and cell pellets were frozen at -80°C until assayed for cytokine levels and MET protein levels, respectively.

Determination of cytokine levels. As a measure of innate immune reactivity following lipopolysaccharide stimulation, cell culture supernatants were analyzed for seven different cytokine and chemokines, including IL-6, GM-CSF, IL-1, IL-12p40, TNF-α, MIP-1β and IL-10. Levels were determined using a commercially available multiplex bead-based kit and run according to manufacturer's instructions (Millipore, Billerica, MA, USA). Briefly, 25 µl of cell culture supernatant was incubated with anti-cytokine-conjugated beads in a 96well filter-bottom plate on a plate shaker overnight at 4 °C. The beads were then washed using a vacuum manifold, and biotin-conjugated detection antibodies were added for a 1-h incubation, followed by the subsequent addition of streptavidin-PE for 30 min. The plates were then read on a Bio-Plex 100 (BioRad, Hercules, CA, USA), and analyzed using Bio-Plex Manager software using a 5-point standard curve. Reference samples were run on each plate to determine assay consistency.

Determination of MET protein levels. After supernatant was extracted, the cultured cells were used to determine the expression of the membrane-bound MET protein. The cell pellet was resuspended in 60-μl Novagen Widescreen protein extraction reagent. Protein assays were performed with the Luminex xMAP technology with a beadbased MET (HGF-Receptor) ELISA kit (Novagen, Rockland, MA, USA), by manufacturer's protocol. Briefly, the isolated protein was mixed with capture beads containing primary antibody directed to MET (HGF-Receptor; Novagen), followed by overnight shaking incubation at 4 °C. Protein was measured on the Bio-Plex Suspension Array System (Bio-Rad, Hercules, CA, USA) at the Beckman Center Core at the USC Norris Cancer Center. Data acquisition and analysis were performed using the Bio-Plex Manager software (Bio-Rad).

**Statistical analyses.** The  $\chi^2$  was used to analyze the association of MET rs1858830 genotype with the presence of maternal antibody. Allelic  $\chi^2$  results are presented; genotypic  $\chi^2$  results are similar. Genotype-dependent differences in MET protein and cytokine production were determined by analysis of variance followed by *post-hoc* Tukey–Kramer tests. Linear regression was used to determine correlation between MET protein expression and cytokine levels. Bonferroni correction for multiple comparisons was used to account for the analysis of multiple cytokines.

## Results

Maternal antibody patterns associated with ASD risk. In a previous report with the CHARGE sample, 30 association with ASD risk was demonstrated for a 37+73-kDa band pattern of maternal antibodies reactive to fetal brain proteins in 61 mothers of children with ASD but not 102 mothers of TD children. 15 Data for the current study were derived from an expanded study to include a total of 202 mothers of children with an ASD and 163 mothers of TD children. 38 Specificity of the combined 37 + 73-kDa band pattern with ASD risk was maintained in this larger sample. The 37+73-kDa band pattern (as illustrated in conjunction with Table 1) was present in 19 of 202 (9%) mothers of children with an ASD, and none of the 163 mothers of TD children (Table 1).

Association of MET rs1858830 C allele with ASDassociated maternal antibody band patterns. We next stratified the sample by maternal antibody patterns that have been shown to be associated with ASD risk in the offspring, and performed genetic association tests with the MET rs1858830 genotype. Among the 19 mothers who were positive for the 37 + 73-kDa band pattern, 11 (58%) were homozygous for the ASD-associated MET rs1858830 C allele. 7 (37%) were C/G heterozygotes and one (5%) was homozygous for the G allele (Table 2). Among the 183 mothers of children with ASD who were negative for the 37 + 73-kDa band pattern, 51 (28%) were C/C homozygous, 79 (43%) were C/G heterozygous and 53 (29%) were G/G homozygous (Table 2). This MET C allele frequency of 76% in the 19 mothers with the 37 + 73-kDa band pattern was significantly higher than the 49% C allele frequency observed

Table 1 Incidence of antibody reactivity to fetal brain proteins in mothers of children with an ASD compared with mothers of TD children

Diagnosis of offspring	N	Positive for 37/73-kDa band pattern (%)	
ASD	202	19 (9)*	
TD	163	0 (0)	
ASD + TD	365	19 (5)	

Abbreviations: ASD, autism spectrum disorder; TD, typically developing. Figure (B) illustrates the 37/73-kDa band pattern by western blot where lane 1 represents an antibody-positive mother of a child with an ASD, and lane 2 represents the mother of a TD control. \*P < 0.0001 ASD vs TD.



Table 2 Association of MET rs1858830 C allele with the ASD-associated 37+73-kDa band pattern

37/73-kDa bands	N	MET C/C (%)	MET C/G (%)	MET G/G (%)	<b>Allelic</b> χ² Ρ
ASD positive	19	11 (58)	7 (37)	1 (5)	Reference
ASD negative	183	51 (28)	79 (43)	53 (29)	0.002
ASD + TD negative	346	101 (29)	154 (45)	91 (26)	0.003

Abbreviations: ASD, autism spectrum disorder; TD, typically developing.

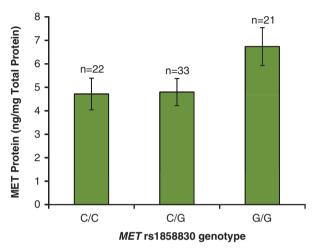
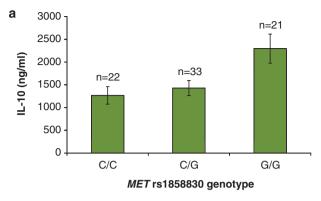


Figure 1 Association of MET rs1858830 genotype with MET protein expression in peripheral blood mononuclear cells (PBMCs). MET protein expression was reduced 1.4-fold in mothers of genotype C/C  $(4.7\pm0.7\,\mathrm{ng\,mg^{-1}}$  total protein) compared with mothers of genotype G/G  $(6.7\pm0.8\,\mathrm{ng\,mg^{-1}}$  total protein) (P = 0.030). MET protein expression was also decreased in mothers of genotype C/G compared with mothers of genotype G/G (P = 0.025). There was no significant difference in MET protein expression between mothers of genotype C/C and C/G (P = 0.463).

in the 183 mothers of children with ASD who did not exhibit the 37 + 73-kDa band pattern (P = 0.002; Table 2). Among 346 mothers of both ASD and TD children who were negative for the 37 + 73-kDa band pattern, 101 (29%) were C/C, 154 (45%) were C/G and 91 (26%) were G/G (Table 2). The 76% frequency of the MET C allele in mothers with the 37 + 73kDa band pattern was also significantly higher than the 51% C allele frequency observed in all 346 mothers lacking the band pattern (P = 0.003; Table 2).

Association of MET rs1858830 genotype with MET protein levels in PBMCs of nonpregnant mothers. We previously reported an association of the MET C allele with decreased MET protein levels in postmortem brain samples from individuals with ASD.<sup>29</sup> To determine if MET rs1858830 genotype also predicts MET protein levels in peripheral blood, we measured MET protein levels in stimulated PBMCs isolated from 76 mothers of ASD and TD children, as MET is only expressed on immune cells in response to activation. As anticipated, results were similar to those observed in postmortem brain where the expression of MET protein was significantly reduced. In mothers with the MET rs1858830 C/C genotype, MET protein expression was reduced 1.4-fold in PBMCs compared with mothers with



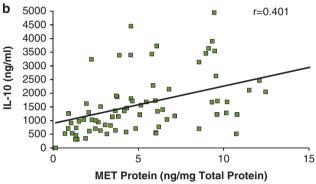


Figure 2 Correlation of both MET rs1858830 genotype (a) and MET protein levels (b) with IL-10 expression in peripheral blood mononuclear cells (PBMCs). (a) IL-10 protein is increased 1.8-fold in G/G genotype mothers (2295  $\pm$  321 ng mg<sup>-1</sup>) compared with C/C genotype mothers (1267  $\pm$  192 ng mg<sup>-1</sup>) (P = 0.001). IL-10 protein is increased 1.6-fold in G/G genotype mothers compared with C/G genotype mothers (1429  $\pm$  165 ng mg $^{-1}$ ) (P=0.005). (**b**) Linear regression indicates highly significant correlation (r = 0.401; P = 0.003) between expression of MET protein and expression of IL-10 protein.

the G/G genotype (P = 0.030; Figure 1). MET protein was also significantly decreased in mothers with the C/G genotype compared with the G/G genotype (P=0.025; Figure 1).

Correlation of both MET C allele and reduced MET protein levels with decreased cytokine IL-10 production in mothers. In the same PBMCs used for cellular MET protein level analysis, we also measured the production of seven cytokines (IL-1, IL-6, IL-10, IL-12(p40), GM-CSF, MIP- $1\beta$  and TNF- $\alpha$ ) following stimulation with lipopolysaccharide of PBMC cultures. Production of the regulatory cytokine IL-10 was significantly associated with MET rs1858830 genotype and MET protein levels. The production of IL-10 was decreased 1.8-fold in mothers of genotype C/C compared with mothers of genotype G/G (P = 0.001; Figure 2a). As expected by the association of MET genotype to MET protein levels, the production of IL-10 was also positively correlated with the amount of MET protein (P = 0.002; Figure 2b). It is important to note that the significance of both of these results survive Bonferroni correction for analysis of seven analytes. The effect of MET genotype was specific to IL-10 as there was no correlation to expression of the other six cytokines

## **Discussion**

The results of this study suggest that the functional *MET* promoter variant rs1858830 C allele is associated with the presence of ASD-associated maternal antibodies to fetal brain proteins. The convergence of these two previously distinct associations with ASD risk (that of maternal antibodies to fetal brain proteins and of the *MET* C allele) provides the first link between an ASD susceptibility gene and a functional immunerelated outcome to that gene.

MET is a receptor tyrosine kinase, serving different functions in distinct cell types, including proliferation, motogenesis, survival and differentiation. MET is best understood as an oncogene involved in the metastasis of several cancers: somatic gain-of-function mutations lead to dramatically increased signaling via the MET receptor, and uncontrolled cell proliferation and migration. MET is also involved in mediating HGF signaling in a number of developmental, physiological and repair processes, including liver regeneration, a gastrointestinal repair, and in serving as a part of a negative feedback loop in immune responses. The MET C allele decreases transcription and results in reduced MET receptor tyrosine kinase protein in postmortem brain samples. 18,29

In the current study, we demonstrate that the METC allele is also associated with decreased expression of MET protein in the immune system. As MET functions as a key negative regulator of immune function, the association of a functional genetic variant and ASD suggests a possible mechanism through which ASD-specific maternal antibodies to fetal brain proteins are generated. In addition to its role in autoimmunity, the MET receptor ligand HGF has also been shown to attenuate inflammatory conditions such as acute hyper responsiveness and airway remodeling in a murine model of asthma, 46 as well as the suppression of acute and chronic cardiac transplant rejection in the mouse. 47 While animal models lend support to an immunosuppressive role, the exact mechanism by which HGF exerts its effects on the immune system has not been fully elucidated. The MET receptor has been detected on monocytes/macrophages, dendritic cells and B cells, but not on T cells, even though HGF has been implicated in the adhesion and migration of memory T cells. 27,44,48-51 Interestingly, MET protein expression is low to undetectable in inactive cells but increases dramatically in response to cellular activation. 44,51 In this study, we provide evidence that subjects harboring the MET 'C' allele are deficient in a crucial immunoregulatory molecule, IL-10, that may leave them more susceptible to prolonged inflammatory conditions.

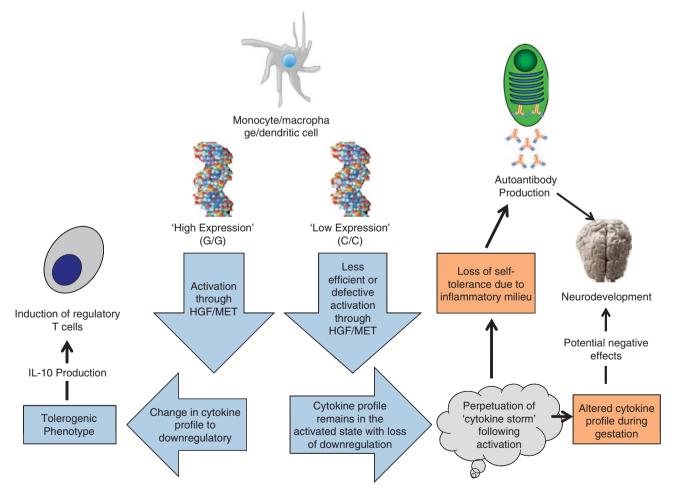


Figure 3 Proposed model of the mechanism by which maternal MET genotype predisposes, via immune dysregulation, toward neurodevelopmental alterations characteristic of ASD.



Interleukin-10 is a well-described immunosuppressive cytokine that has been shown to be upregulated in animal models, in which HGF is important in the prevention of autoimmunity. <sup>25,26,52</sup> In addition, *in vitro* studies on the effects of HGF on both human and mouse antigen-presenting cells implicate IL-10 as a major contributor to the reported anti-inflammatory effects. <sup>28,53</sup> IL-10 has been reported to be decreased in children with ASD compared with controls, <sup>54–56</sup> but only in the context of innate cell activation. In the context of adaptive immune activation or at baseline, IL-10 levels in children with ASD are unchanged. <sup>57–61</sup> However, there have not been any studies investigating the potential role of IL-10 dysregulation in the mothers of children with ASD.

In this study, we describe a decrease in levels of IL-10 that is significantly correlated with levels of MET protein expression. We had anticipated an increase in inflammatory cytokines as MET signaling has been shown to directly modulate the inflammatory response through phosphorylation of GSK3B; however, unlike other studies we did not observe a significant increase in pro-inflammatory cytokine production.<sup>53</sup> This is likely attributed to both the cellular composition of the stimulated cultures (the use of whole PBMC in our study vs the use of bone marrow-derived macrophages in animal studies) as well as the timing of cytokine measurements (48 vs 24 h). IL-10 is most often attributed with the resolution of an inflammatory response. It may be that signaling through the MET receptor functions to shorten the duration of the inflammatory response and not necessarily the magnitude, thereby allowing sustained inflammation in mothers with the MET'C' allele. Such sustained inflammation would provide an environment conducive to loss of tolerance, thereby increasing the risk of autoantibody production, or potentiate a change in the cytokine profile during pregnancy that could negatively affect proper neurodevelopment (Figure 3).

A limitation of this study, and of our predicted model, is the lack of data indicating the factor responsible for initiation of the inflammatory response. While the *MET* 'C' allele may provide susceptibility toward reduced immune regulation, an environmental trigger such as infection, xenobiotic exposure or other stressor is still required to initiate an immune response. This may partially explain why the 'C' allele is evenly distributed among the population but is highly prevalent in mothers who exhibit markers of immune dysregulation, that is, autoantibody production. Future studies in a prospective model that will allow for tracking of environmental insults are needed to explore this mechanism in further detail.

Several studies have now addressed the potential role for maternal antibodies to fetal brain proteins in ASD. 12-17,38 While each of these studies demonstrated both antibody specificity as well as a potential pathologic significance of the maternal autoantibodies, the current study is the first to present evidence suggesting a genetic polymorphism associated with the production of these antibodies. We fully acknowledge that no single gene or environmental factor will account for the entirety of ASD etiologies, and thus our focus on one ASD candidate gene, the MET receptor tyrosine kinase, was based on its bi-functional role in brain and immune development. Our observations that *MET* rs1858830 genotype predicts MET protein expression in the immune system, and that lower MET protein levels are correlated with

lower regulatory cytokine levels, provides mechanistic support for this focus. The studies described herein represent the beginning of our investigation into the potential mechanism(s) by which the maternal immune system becomes susceptible to the generation of antibodies to fetal brain tissue. Identification of a predisposing genetic factor such as MET will contribute greatly toward the understanding of the mechanisms involved in a subset of ASD cases.

## Conflict of interest

The authors declare no conflicts of interest.

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