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The PMA phorbol ester tumor promoter increases canonical Wnt signaling via macropinocytosis

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Abstract Activation of the Wnt pathway lies at the core of many human cancers. Wnt and macropinocytosis are often active in the same processes, and understanding how Wnt signaling and membrane trafficking cooperate should improve our understanding of embryonic development and cancer. Here, we show that a macropinocytosis activator, the tumor promoter phorbol 12-myristate 13-acetate (PMA), enhances Wnt signaling. Experiments using the *Xenopus* embryo as an in vivo model showed marked cooperation between the PMA phorbol ester and Wnt signaling, which was blocked by inhibitors of macropinocytosis, Rac1 activity, and lysosome acidification. Human colorectal cancer tissue arrays and xenografts in mice showed a correlation of cancer progression with increased macropinocytosis/multivesicular body/lysosome markers and decreased GSK3 levels. The crosstalk between canonical Wnt, focal adhesions, lysosomes, and macropinocytosis suggests possible therapeutic targets for cancer progression in Wnt-driven cancers.

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eLife assessment

Altogether, the strength of this **important** study is that it provides **compelling** evidence in several biological models, including *Xenopus* embryos, that Wnt3a increases macropinocytosis and that PMA increases this cellular response. This novel link between Wnt, focal adhesions, lysosomes, and macropinocytosis will be very interesting for cell and tumor biologists. In future work, it will be important to identify the underlying mechanism, i.e., the molecular node whereby this interaction occurs.

Introduction

Cancer progression is a process during which mutations in various signaling pathways accumulate in the cell leading to uncontrolled growth (*Kinzler and Vogelstein, 1996; Hanahan and Weinberg, 2011*). Cancer progression is also influenced by tumor promoters, substances that do not mutate the DNA by themselves but nevertheless facilitate cancer development. In 1941, Berenblum and Rous found that inflammatory agents such as croton oil or turpentine could induce skin cancer when applied repeatedly to mouse or rabbit skin, but only when an initiator mutagen such as benzopyrene or anthracene had been previously painted on that patch of skin (*Berenblum, 1941; Rous and Kidd, 1941*). The active agent in croton oil was later purified and found to be the phorbol ester phorbol 12-myristate 13-acetate (PMA), also known as 12-O-tetradecanoylphorbol-13-acetate (TPA).

The molecular mechanism by which PMA promotes cancer is incompletely understood but involves the activation of protein kinase C (PKC) in the plasma membrane, where PMA mimics the endogenous second messenger diacylglycerol (DAG) (Nishizuka, 1984). Tumor promoters cause chronic inflammation and significantly contribute to tumorigenesis (Weinberg and Weinberg, 2007).

The canonical Wnt/ β -catenin pathway is a major driver of cancer and plays a crucial role in embryonic development (MacDonald et al., 2009; Nusse and Clevers, 2017). In the case of colon cancer, the great majority of tumors are initiated by mutation of the tumor suppressor Adenomatous Polyposis Coli (APC), which is a component of the destruction complex that normally degrades the transcriptional regulator β -catenin, leading to its accumulation in the nucleus and benign polyp formation (Kinzler and Vogelstein, 1996; Segditsas and Tomlinson, 2006). Activation of the Wnt/ β -catenin pathway is also the driving force in many other cancers (Galluzzi et al., 2019). A key regulator of Wnt signaling is Glycogen Synthetase 3 (GSK3), which phosphorylates β -catenin, leading to the formation of a phosphodegron that triggers its degradation in the proteasome. Upon activation of the Wnt receptors, GSK3, together with the destruction complex, is sequestered into multivesicular bodies (MVBs), also known as the late endosomes (Taelman et al., 2010; Vinyoles et al., 2014). The translocation of GSK3 from the cytosol into MVBs/lysosomes leads to the stabilization of many other cellular proteins in addition to β -catenin, in a process designated Wnt-STOP (Wnt-Stabilization of Proteins) (Acebron et al., 2014; Albrecht et al., 2021).

The many GSK3 phosphorylation targets stabilized by Wnt signaling include regulators of macropinocytosis such as Ras, Rac1, and Pak1 (Taelman et al., 2010; Albrecht et al., 2020; Jeong et al., 2012). A recent advance was the realization that Wnt signaling requires macropinocytosis (i.e., cell drinking) in order to traffic activated Wnt receptors and GSK3 to MVBs and lysosomes (Redelman-Sidi et al., 2018; Tejada-Muñoz et al., 2019). Wnt-induced macropinocytosis also causes the internalization of focal adhesions (FAs) and integrins from the cell surface into MVBs (Tejada-Muñoz et al., 2022c). The requirement of macropinocytosis for Wnt signaling is revealed, for example, by the decrease in nuclear β -catenin when colorectal cancer (CRC) cells mutant for APC were treated with macropinocytosis inhibitors such as the derivative of the diuretic Amiloride known as 5-(N-Ethyl-N-isopropyl) Amiloride (EIPA) (Tejada-Muñoz et al., 2019; Tejada-Muñoz and De Robertis, 2022a; Tejada-Muñoz and De Robertis, 2022b).

Macropinocytosis is emerging in the field as a promising target for cancer treatment (Lambies and Commisso, 2022). In addition, it has been found that the macropinocytosis induced by Wnt signaling causes significant lysosome acidification and activation (Albrecht et al., 2020). There is also accumulating evidence showing that the size and number of lysosomes increase during carcinogenesis (Cardone et al., 2005; Kroemer and Jäättelä, 2005; Zhitomirsky and Assaraf, 2016). Lysosomes, previously thought to be the garbage disposal of the cell, have emerged as central organelles in cancer (Kirkegaard and Jäättelä, 2009; Kirkegaard and Jäättelä, 2009).

From the considerations above, investigating the intersection of Wnt signaling, macropinocytosis, lysosomes, FAs, and membrane trafficking, is of great interest. While in most cells macropinocytosis requires stimulation of receptor tyrosine kinase (RTK) (Haigler et al., 1979; Yoshida et al., 2018), Ras (Bar-Sagi and Feramisco, 1986; Commisso et al., 2013), or Wnt (Redelman-Sidi et al., 2018; Tejada-Muñoz et al., 2019), some cells, such as amoebae and macrophages, have constitutive macropinocytosis. Importantly, Swanson discovered that the phorbol ester PMA stimulated macropinocytosis even in macrophages that were thought to have fully activated constitutive macropinocytosis (Swanson, 1989).

In the present study, we asked whether phorbol ester could interact with canonical Wnt signaling. Using the *Xenopus* embryo, which provides a premier model system to analyze the Wnt pathway, we found that PMA, which lacked any effects on its own, greatly sensitized the embryo to dorsalization by Wnt signaling, leading to the formation of radial head structures lacking trunks. The dorsalizing effect of PMA on embryos was blocked by the macropinocytosis inhibitor EIPA, the lysosomal V-AT-Pase inhibitor Bafilomycin A (Baf), or the Rac1 inhibitor EHT1864. Rac1 is an upstream regulator of the p21-activated kinase 1 (Pak1) required for actin-mediated macropinocytosis. Lysosomal activity was specifically required at the 32-cell stage for the activity of the Wnt-mimicking GSK3 inhibitor Lithium Chloride (LiCl). Ectopic axes induced by dominant-negative GSK3 (DN-GSK3- β) or β -catenin mRNAs were blocked by EIPA or dominant-negative Rab7, indicating a widespread requirement of membrane trafficking for Wnt signaling.

The involvement of membrane trafficking in Wnt-driven cancer was supported by immunostaining results of arrayed human CRC histological sections, in which we found that the grade of malignancy increased Pak1, CD63 (an MVB marker), and V0a3 (a V-ATPase subunit that marks lysosomes) levels, while decreasing GSK3. In *Xenopus* embryos, twinning induced by Xwnt8 mRNA microinjection was blocked by inhibition of Rac1. In cultured CRC SW480 cells reconstituted with APC, the FA marker Tes was cytoplasmic, while the same cells mutant for APC (in which the Wnt pathway is activated) Tes was nuclear. The results suggest that macropinocytosis and membrane trafficking play an important role in embryogenesis and cancer progression.

Results

The phorbol ester PMA potentiates Wnt-like signals in *Xenopus* embryos via macropinocytosis and lysosomal acidification

Wnt is one of the earliest signals in the embryo and it induces the formation of the primary dorsal-ventral and head-to-tail axes (Loh *et al.*, 2016; Niehrs, 2012). It is known that β -catenin is required for Wnt signaling activation on the dorsal side during early cleavage. Recently, we reported that inhibition of macropinocytosis with EIPA, or of lysosomal acidification with Baf, ventralized endogenous early dorsal axis formation, leading in particular to reduced head and brain structures (Tejeda-Muñoz and De Robertis, 2022a). Activation of lysosomes on the dorsal side of the embryo was detectable as early as the 64-cell stage (Tejeda-Muñoz and De Robertis, 2022a).

The tumor promoter PMA is known to increase macropinocytosis in macrophages (Swanson, 1989). To test its activity during early development, we used a sensitized system in which a small amount of the GSK3 inhibitor LiCl (4 nM at 300 mM in a single ventral injection at 4-cell) causes only a small dorsalization with slightly enlarged heads (Kao *et al.*, 1986; Tejeda-Muñoz and De Robertis, 2022a). The dorsalizing effects of PMA were most striking when phenotypes were analyzed at the early tailbud stage (Figure 1A–H). LiCl alone expanded head structures only slightly and PMA alone was without effect (Figure 1A, B, E). However, the combination of PMA and LiCl lead to radially dorsalized embryos consisting of head structures lacking trunks (compare Figure 1B–F). This striking phenotype was eliminated by co-injection of EIPA or Baf, suggesting that macropinocytosis and lysosomal acidification are required for the effect of PMA (Figure 1G, H). Injection of Baf alone decreased head and neural development, supporting a requirement for lysosomes for endogenous dorsal axis formation (Figure 1D). Target genes of the early Wnt signal can be detected at blastula (stage 9.5, just after the start of zygotic transcription) by measuring transcripts of *Siamois* and *Xnr3* by qRT-PCR (Tejeda-Muñoz and De Robertis, 2022a). As shown in Figure 1I, PMA alone (4 nM at 500 nM) was without any effect, LiCl increased the Wnt target genes, and the combination of PMA and LiCl synergized significantly elevated *Siamois* and *Xnr3* transcript levels, while EIPA and Baf blocked the increase in *Siamois* and *Xnr3* caused by PMA/LiCl (Figure 1I).

Using the BAR (β -catenin activity reporter) assay system (Biechele and Moon, 2008) in a cultured HEK293 permanent BAR/Renilla cell line, PMA synergized with LiCl when assayed by transcriptional BAR luciferase activity (Figure 1J). PMA on its own had no detectable effect on BAR reporter activity (Figure 1—figure supplement 1J). In this cell line, PMA cooperated with LiCl to increase β -catenin levels (Figure 1—figure supplement 1A–H). PMA alone caused a slight increase in β -catenin protein staining, but this lacked statistical significance (Figure 1—figure supplement 1C, I), presumably due to a small amount of endogenous Wnt secreted by this cell line (Colozza *et al.*, 2020). Importantly, the effect of PMA on the activation of the Wnt pathway by GSK3 inhibition was abrogated by the inhibition of V-ATPase with Baf (Figure 1—figure supplement 1H, I).

To test the effect of PMA on Wnt-induced macropinocytosis we used an uptake assay in which 3T3 cells were treated with dextran for 1 hr (Figure 1K–P). The macropinocytosis marker TMR-dextran 70 kDa has a hydrated diameter of more than 200 nM and is diagnostic of large endocytic cups, in particular when sensitive to the macropinocytosis inhibitor EIPA (Commisso *et al.*, 2013; Commisso *et al.*, 2014). In the presence of Wnt3a, but not in the absence, PMA strongly induced dextran uptake and this effect was blocked by EIPA (Figure 1K–P, quantitated in Figure 1Q). PMA also increased dextran uptake in the SW480 CRC cell line that has constitutive Wnt signaling, even in the absence of added Wnt (Figure 1—figure supplement 1J, K). In time-lapse video microscopy of SW480 cells with the plasma membrane tagged by myristoylated-GFP (mGFP), PMA addition triggered the abundant

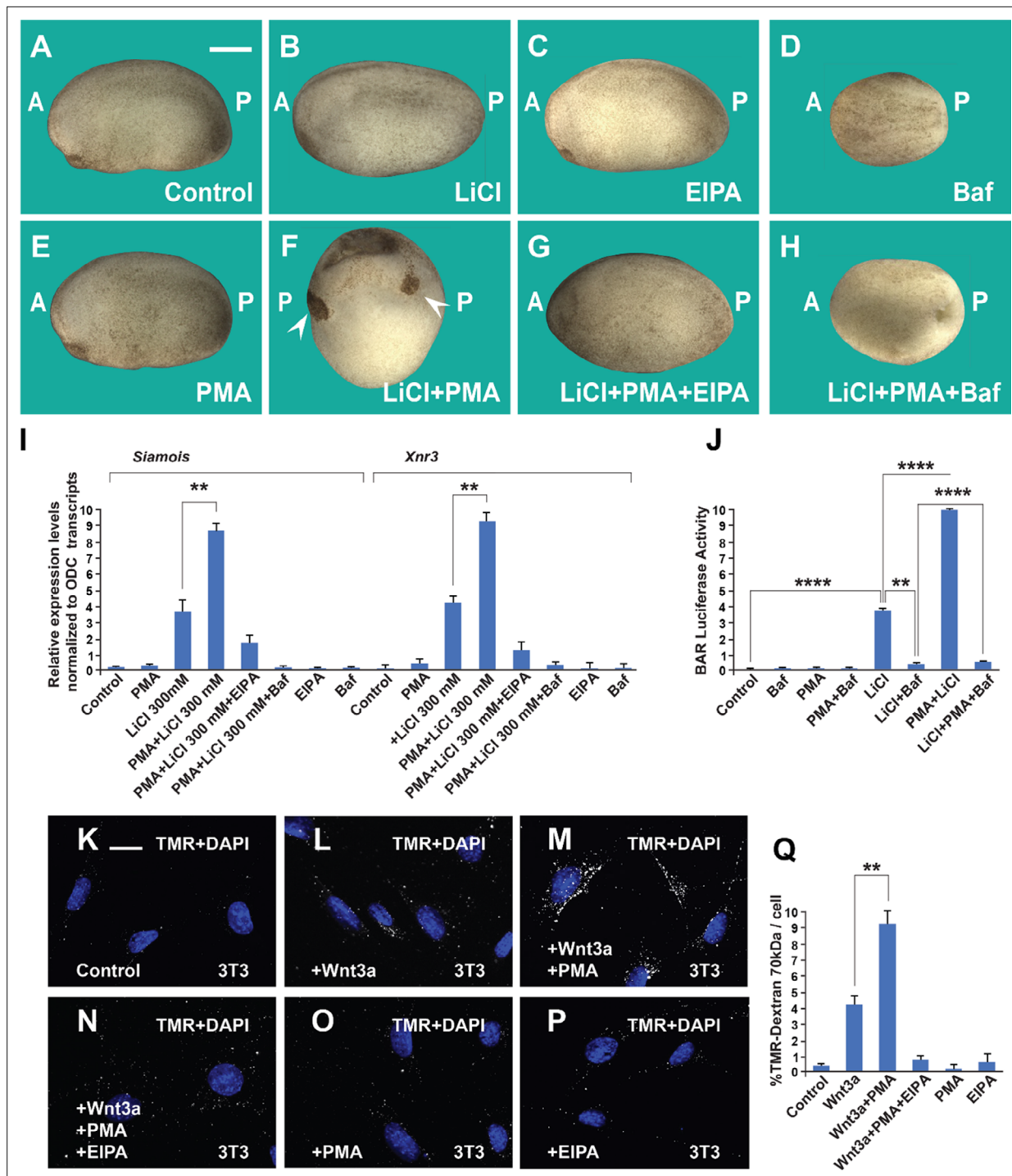


Figure 1. The phorbol ester phorbol 12-myristate 13-acetate (PMA) synergizes with GSK3 inhibition and Wnt3a and this cooperation requires macropinocytosis and lysosome acidification. (A) Uninjected control embryo at stage 24. (B) Lithium Chloride (LiCl) (4 nl 300 mM, 1x ventral) dorsalized the embryo only slightly. (C) 5-(N-Ethyl-N-isopropyl) Amiloride (EIPA) (1 mM, 1x ventral) alone did not produce a distinctive phenotype at this concentration. (D) The vacuolar ATPase (V-ATPase) inhibitor Bafilomycin A1 (Baf) (incubation with 5 μ M for 7 min at 32 cells) ventralized embryos. (E) A single injection of PMA (500 nM) did not show any phenotypic effect. (F) PMA and LiCl strongly synergized, resulting in embryos with radial heads. (G) Co-injection of the EIPA macropinocytosis inhibitor blocked dorsalization caused by LiCl plus PMA. (H) Incubation with Baf lysosomal acidification inhibitor blocked dorsalization caused by LiCl plus PMA. Number of embryos was as follows: A = 70, 100%; B = 80, 98%; C = 75, 94%; D = 82, 92%; E = 76, 100%; F = 112, 98%; G = 75, 93%; H = 85, 98%. Scale bar, 500 μ m. (I) PMA increases the effects of LiCl in Wnt signaling in early embryos. qRT-PCR analysis at blastula stage 9.5 of the Wnt target genes *Siamois* and *Xnr3* normalized for Ornithine decarboxylase (ODC) transcripts. Embryos received a single injection of 4 nl of LiCl 300 mM with or without PMA (500 nM). Error bars denote standard error of the mean (SEM) ($n \geq 3$) (** $p < 0.01$).

Figure 1 continued on next page

Figure 1 continued

(J) Luciferase assay in HEK293BR cells showing cooperation between PMA and LiCl, and its inhibition by Baf. (K–P) PMA and Wnt3a cooperated in stimulating macropinocytosis, which was sensitive to inhibition by EIPA, in 3T3 cells. TMR-dextran 70 kDa was added for 1 hr. (Q) Quantification of the Wnt3a + PMA results. Experiments with cultured cells were biological triplicates. Error bars denote standard deviation (**** $p < 0.0001$ and ** $p < 0.01$).

The online version of this article includes the following figure supplement(s) for figure 1:

Figure supplement 1. In cultured cells, the phorbol ester phorbol 12-myristate 13-acetate (PMA) cooperates with GSK3 inhibition in β -catenin signaling and increases dextran macropinocytosis.

formation of macropinosome cups within minutes of addition when compared to DMSO (Dimethylsulfoxide) controls (**Video 1**).

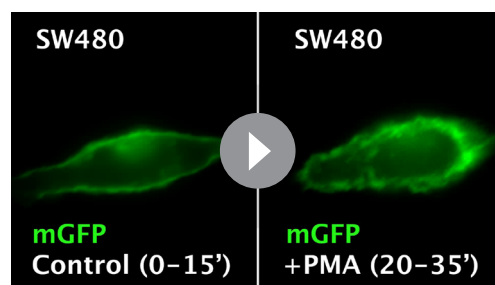
These results in embryos and cultured cells indicate that the tumor promoter PMA potentiates Wnt signaling and that this cooperation requires macropinocytosis and lysosomal acidification.

Lysosomal activity is required at the 32-cell stage for early Wnt activation

In previous work, we had shown that immersion of embryos in the V-ATPase inhibitor Baf at 5 μ M for only 7 min at the 32-cell stage resulted in ventralized microcephalic embryos lacking cement glands (**Tejeda-Muñoz and De Robertis, 2022a**). To test whether this ventralization is caused by inhibition of the endogenous Wnt pathway signal, we now carried out order-of-addition experiments. When an initial Baf incubation of 7 min was followed by a second brief incubation in LiCl while still at the 32-cell stage, the ventralization by Baf was rescued, and the resulting tailbud embryos had a LiCl-like dorsalized phenotype (**Figure 2A–D**). The initial effect of GSK3 by LiCl is to trigger macropinocytosis (**Albrecht et al., 2020**). The phenotypes are supported by the activation of Wnt signaling at the blastula stage, as indicated by qRT-PCR of direct Wnt target genes (**Figure 2E**). In the converse experiment, when embryos were first briefly treated with LiCl, washed, and subsequently incubated for 7 min in Baf, the effect of lysosomal inhibition was dominant, resulting in ventralized tailbud embryos (**Figure 2F–J**). These order-of-addition experiments strongly support the proposal that membrane trafficking and endogenous lysosomal activity are required to generate the initial endogenous Wnt signal that takes place at the 32-cell stage of development (**Tejeda-Muñoz and De Robertis, 2022a**).

Wnt pathway signaling in microinjected embryos or APC mutant cells requires macropinocytosis and membrane trafficking

Next, we tested the requirement for macropinocytosis of different components of the Wnt pathway using the axis duplication assay (**McMahon and Moon, 1989**). Microinjection of Wnt-mimic dominant-



Video 1. An SW480 colorectal cancer (CRC) cell before and after treatment with phorbol 12-myristate 13-acetate (PMA) (0.3 μ M); the plasma membrane was marked by transfection of myristoylated-GFP (mGFP) and filmed for 15 min. Note that PMA greatly enhanced plasma membrane vesicular activity typical of macropinocytosis. Control cell was treated with DMSO alone and shows low but detectable macropinocytosis cup formation. The arrowheads indicate two prominent macropinocytosis vesicles.

<https://elifesciences.org/articles/89141/figures#video1>

negative GSK3 (DN-GSK3- β) mRNA, a catalytically inactive form of GSK3, induced complete axis duplications, which were blocked by the co-injection of the macropinocytosis inhibitor EIPA (1 mM, 4 nl) (**Figure 3A–D**). Secondary axis induction by β -catenin mRNA was also abrogated by EIPA, as well as the membrane trafficking inhibitor dominant-negative Rab7 (DN-Rab7) (**Figure 3E–H**). β -Catenin is a transcriptional activator, and one might have not expected a requirement for membrane trafficking. However, axis formation is a multistep process and appears to require macropinocytosis and Rab7 activity in addition to transcriptional activation of Wnt target genes.

Macropinocytosis is an actin-driven process that is orchestrated by Pak1 kinase, which in turn is activated by Rac1. Rac1 activity is essential for the formation of lamellipodia and macropinocytosis cups (**Hall, 1998**). EHT1864 (EHT) is a specific

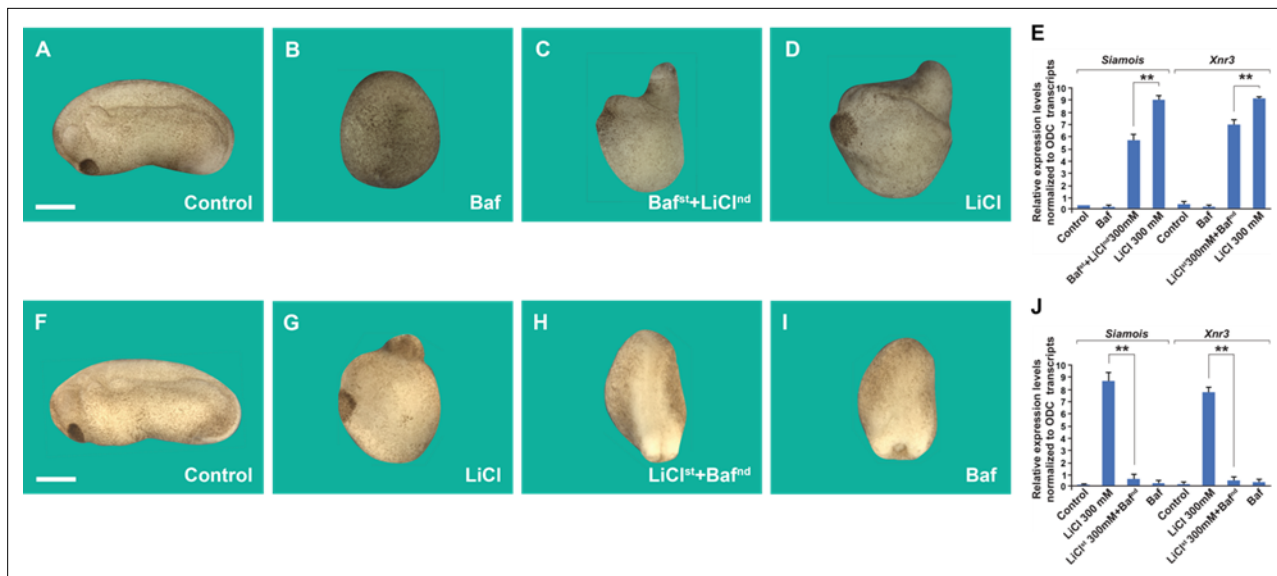


Figure 2. Order-of-addition experiment showing that lysosomal acidification at 32-cell stage is required for early Wnt/ β -catenin activation. (A) Untreated embryo. (B) Baf treatment (5 μ M for 7 min) inhibited endogenous axis formation resulting in ventralized embryos lacking heads at early tailbud. (C) Embryos treated with Baf, washed, and then immersed in 300 mM Lithium Chloride (LiCl) an additional 7 min formed heads, resulting in dorsalized embryos with enlarged heads and short trunks. Note that all treatments were done at 32-cell stage, which is critical for the early Wnt signal. (D) LiCl treatment alone dorsalized embryos. (E) Quantitative RT-PCR (qPCR) for Wnt target genes *Siamois* and *Xnr3* at blastula, confirming that the phenotypic effects are due to early activation of the Wnt pathway. (F) Untreated embryo. (G) Embryos incubated with LiCl at 32-cell were dorsalized. (H) Embryos first treated with LiCl and then subsequently with Baf resulted in ventralized embryos lacking heads. (I) Baf treatment caused ventralization and small heads. (J) Quantitative RT-PCR for Wnt target genes *Siamois* and *Xnr3* showing that inhibiting V-ATPase at the 32-cell stage blocks the effect of earlier LiCl treatment. The numbers of embryos analyzed were as follows: A = 115, 100%; B = 124, 98% with phenotype; C = 132, 97%; D = 99%; F = 132, 100%; G = 135, 99%; H = 126, 97%; I = 129, 98% (scale bars, 500 μ m.). Error bars denote standard error of the mean (SEM) ($n \geq 3$) (** $p < 0.01$).

inhibitor of Rac1 that has the practical advantage of being water soluble (Hampsch et al., 2017). When embryos were immersed in EHT at the critical 32-cell stage for 7 min (10 mM), head structures were greatly reduced, and ventral posterior structures expanded (Figure 4A, B). This was accompanied by a significant increase of transcripts of the ventral markers *Sizzled* and *Vent1* at the blastula stage (Figure 4C).

Microinjection of a small amount of EHT (1 mM, 4 nl, 1x ventral at 4-cell) was without effect on its own but was able to block the axis duplication effect of microinjecting *Wnt8* mRNA (Figure 4D–G). Quantitative RT-PCR analyses at the blastula stage confirmed that inhibiting Rac1 strongly reduced the induction of the Wnt target genes *Siamois* and *Xnr3* caused by *xWnt8* mRNA microinjection at the late blastula stage (Figure 4H).

In SW480 CRC cells, which have constitutively activated Wnt due to mutation of APC (Leibovitz et al., 1976; Faux et al., 2004), EHT inhibited the size of cell spheroids in hanging drop cultures (Figure 4—figure supplement 1A–C) and, importantly, significantly inhibited macropinocytosis uptake of TMR-dextran in these cancer cells (Figure 4—figure supplement 1D–F).

Rac1 itself was stabilized in 3T3 cells by Wnt signaling, as indicated by experiments showing (1) that LiCl treatment increased levels of Rac1 protein (Figure 4I–J’), (2) that cells transfected with constitutively active β -catenin-GFP had higher levels of Rac1 than control untransfected cells (Figure 4K–K’), and (3) that Rac1 immunostaining was stabilized in cells transfected with CA-Lrp6-GFP when compared to untransfected cells (Figure 4L–L’).

The results show that signaling by the Wnt/ β -catenin pathway requires macropinocytosis, Rac1, and membrane trafficking in *Xenopus* embryos and cancer cells.

Tumor progression in human CRCs correlates with a role for membrane trafficking in Wnt/ β -catenin signaling

CRC is particularly favorable for the analysis of tumor progression because it gradually accumulates mutations, and in over 85% of the cases the initial driver mutation is in the APC tumor suppressor

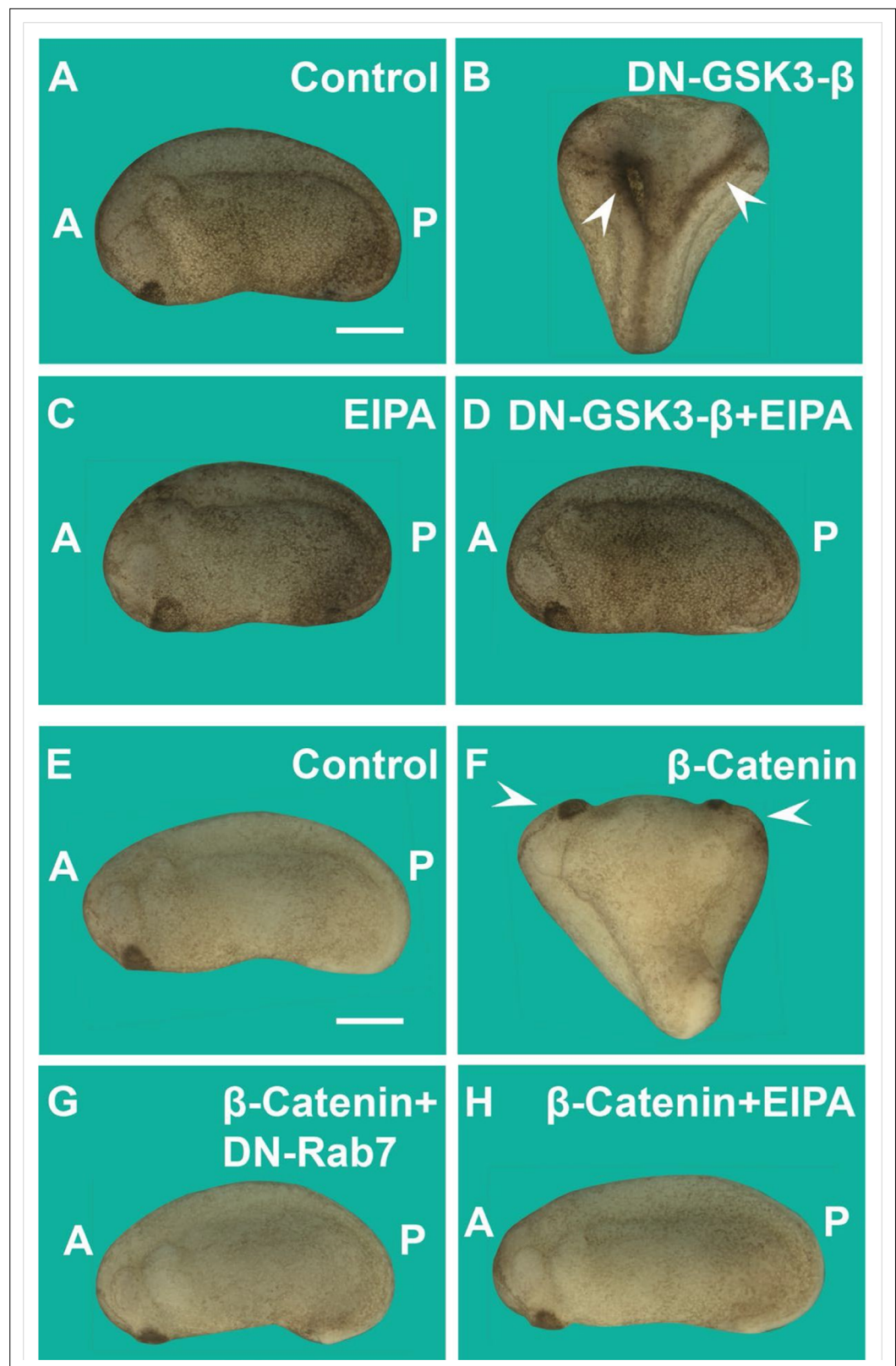


Figure 3. Induction of secondary axes by the GSK3/β-catenin pathway requires membrane trafficking. **(A)** Control embryo at early tailbud. **(B)** Embryo injected with DN-GSK3 (dominant-negative GSK3-β, 150 pg 1x ventral) mRNA, showing double axes (arrowheads). **(C)** Injection of 5-(N-Ethyl-N-isopropyl) Amiloride (EIPA) 1x ventral alone showed no phenotypic effect (4 nl, 1 mM). **(D)** Co-injection of DN-GSK3 and EIPA blocked double axis formation.

Figure 3 continued on next page

Figure 3 continued

(E) Uninjected control embryo. (F) Activation of Wnt signaling via injection of β -catenin mRNA (80 pg) induced complete twinned axes (arrowheads). (G) Embryo co-injected with β -catenin mRNA and DN-Rab7 (500 pg) showing that membrane trafficking is required for secondary axis formation. (H) Co-injection of β -catenin mRNA and the macropinocytosis inhibitor EIPA blocks axial duplication. The numbers of embryos analyzed were as follows: A = 62, 100%; B = 75, 94% with double axes; C = 76, 100%; D = 70, 98%; E = 70, 100%; F = 73, 98%; G = 69, 75%; H = 82, 97%; four independent experiments (scale bars, 500 μ m).

(Kinzler and Vogelstein, 1996; Weinberg and Weinberg, 2007). This offered an opportunity to test our proposal that membrane trafficking and its molecular components play an important role in Wnt-driven cancers (Albrecht et al., 2021). Commercially available arrays of paraffin-sectioned histological sections containing 90 cases of adenocarcinoma of various grades I–IV and 90 samples of corresponding adjacent normal tissues (from TissueArray) were used for immunostaining. Arrays were double stained for β -catenin and antibodies against proteins involved in the macropinocytosis/MVB/lysosome/GSK3 pathway. Each individual section was evaluated, and normal colon, adenocarcinoma I, and adenocarcinoma IV were compared to assess the effects of cancer progression.

As shown in Figure 5, β -catenin levels consistently correlated with grade IV malignancy when compared to normal colon or grade I cancers. Pak1, a kinase required for macropinocytosis and Wnt signaling (Redelman-Sidi et al., 2018; Albrecht et al., 2020) was significantly elevated with increased malignancy (Figure 5A–C). CD63, a tetraspan protein that marks MVB intraluminal vesicles (Escola et al., 1998), reached high levels in adenocarcinoma IV (Figure 5D–F), particularly in cells with the highest β -catenin expression (compare Figure 5F–F'). This increase in MVBs is in agreement with the GSK3 sequestration model of Wnt signaling (Taelman et al., 2010). The V-ATPase subunit V0a3, a marker for acidic lysosomes (Ramirez et al., 2019), also increased strongly with the CRC malignancy (compare Figure 5G–I). Importantly, the cellular levels of total GSK3 decreased with malignancy (Figure 5J–L, compare Figure 5J–L), as predicted by the sequestration model (Taelman et al., 2010). During Wnt signaling, GSK3 is sequestered into MVBs which are trafficked into lysosomes and a decrease was to be expected after sustained Wnt activation. Previously, we had been unable to demonstrate a decrease in total cellular GSK3 levels during acute Wnt signaling experiments (Taelman et al., 2010). The co-localization of β -catenin with Pak1, CD63, V0a3, or GSK3 in human colorectal adenocarcinomas was quantified in Figure 5—figure supplement 1A–D.

These results were confirmed by examining immunostaining levels of Pak1, V0a3, and GSK3 compared to β -catenin protein in mouse xenografts in a CD1 NU/NU nude mouse model injected with SW480 human CRC cells (in which Wnt activation is known to be caused by APC mutation, Faux et al., 2004). In this case, tumor cells were compared to normal colon mouse tissue sections. The results showed that CRC cells had high levels of β -catenin, Pak1, and lysosomes (Figure 5—figure supplement 1E–J), while GSK3 levels were reduced (compare Figure 5—figure supplement 1K–L) in agreement with the results in human tissue arrays.

These in vivo observations in human tumors are consistent with the view that macropinocytosis components, MVBs, and lysosomes increase with CRC malignancy. Further, the decrease in total GSK3 levels suggests a role in the trafficking and degradation of this key enzyme in cancer.

Wnt affects the subcellular localization of FA components

We have previously reported a crosstalk between the Wnt and FA signaling pathways in which Wnt3a treatment rapidly led to the endocytosis of Integrin β 1 and of multiple FA proteins into MVBs (Tejeda-Muñoz et al., 2022c). FAs link the actin cytoskeleton with the extracellular matrix (Figure 6A), and we now investigated whether FA activity is affected by Wnt signaling, PMA treatment, or CRC progression. FA components containing LIM domains can shuttle between the nucleus and cytoplasm (Nix and Beckerle, 1997; Kadrmaz and Beckerle, 2004; Anderson et al., 2021). We examined whether constitutive Wnt signaling regulates FA trafficking using the SW480 and SW480APC system, in which the stable reconstitution with full-length APC restores these CRC cells to a non-malignant phenotype (Faux et al., 2004), which is accompanied by a decrease in β -catenin levels (Figure 6B). As shown in Figure 6C–D", the PET-LIM domain FA protein Tes (also known as Testin) (Garvalov et al., 2003) was cytoplasmic in SW480APC, but was nuclear in SW480 cells in which Wnt is activated by APC mutation. The nuclear localization of Tes in SW480 cells required

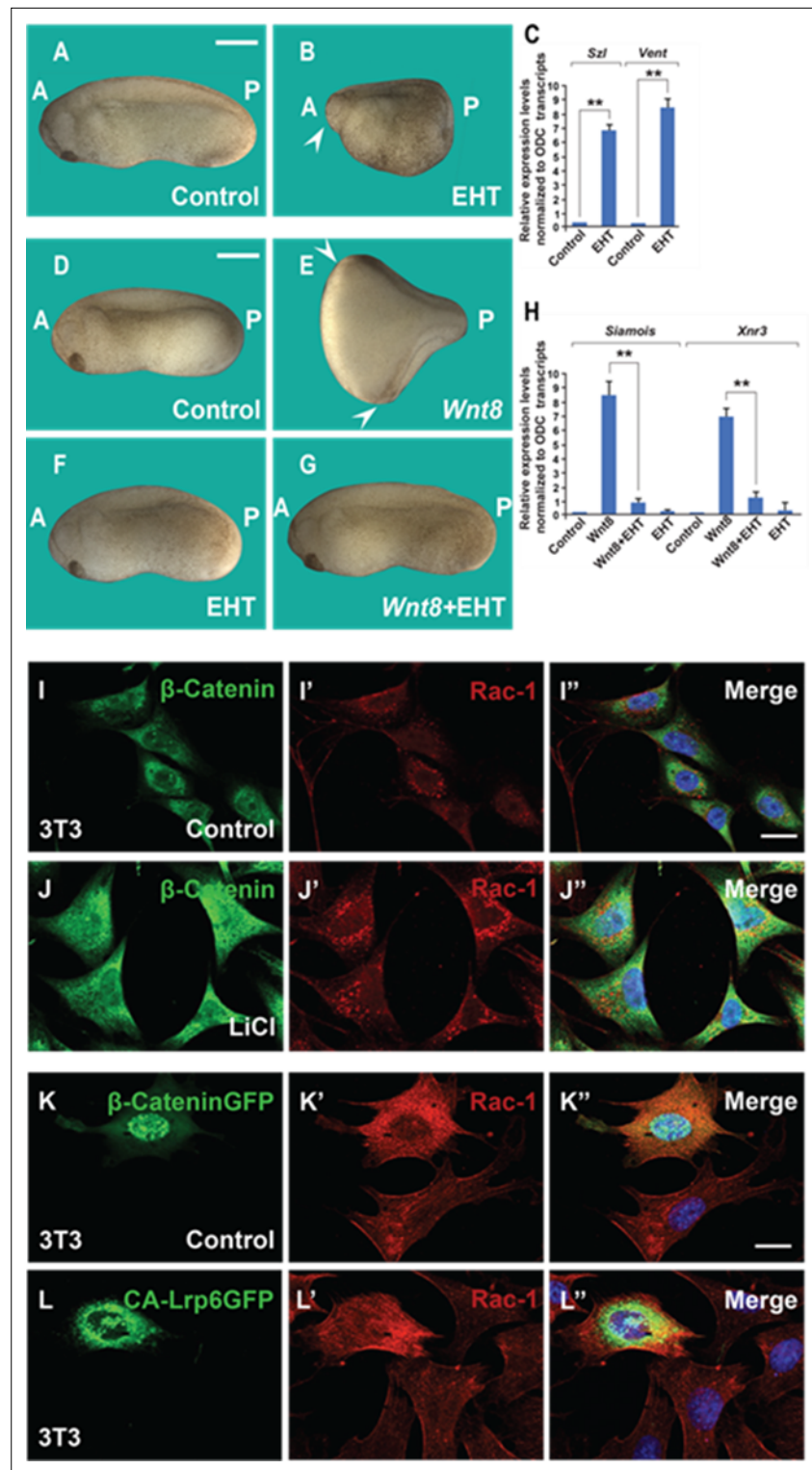


Figure 4. The Rac1 inhibitor EHT1864 blocks Wnt signaling in *Xenopus* embryos, and Rac1 levels are stabilized by treatments that increase Wnt/ β -catenin signaling. (A) Uninjected control embryo. (B) Incubation of the *Xenopus* embryos with the Rac1 inhibitor EHT at 32-cell stage (7 min, 10 mM) resulted in a ventralized phenotype with a small head in the anterior (A, arrowhead) and expanded ventral structures in the posterior (P). Rac1 activity is required for macropinocytosis, see **Figure 4—figure supplement 1**. (C) qRT-PCR of gastrula stage embryos showing increased ventral markers Szl and Vent1 after Rac1 inhibition. (D) Control embryo. (E) Injection of Wnt8 mRNA (2 μ g) induces complete duplicated axes (arrows). (F) Injected embryos with EHT (1 mM, 4 nl 1x ventral)

Figure 4 continued on next page

Figure 4 continued

alone showed no phenotypic effect at this concentration. (G) EHT co-injected with Wnt8 mRNA blocked double axis formation. (H) qRT-PCR of Wnt target genes *Siamois* and *Xnr3* at blastula confirming that Rac1 is required for early Wnt signaling. (I–J) Treatment of 3T3 cells with 40 mM Lithium Chloride (LiCl) increases β -catenin and Rac1 levels. (K–L) Transfection of 3T3 cells with stabilized constitutively active forms of β -cateninGFP or Lrp6GFP increased levels of Rac1 protein in transfected cells compared to untransfected ones. The numbers of embryos analyzed were as follows: A = 52, 100%; B = 47, 95% with ventralized small head phenotype; D = 58, 100%; E = 67; 97%; F = 64, 96%; G = 62, 97%, four independent experiments. Scale bars for embryos 500 μ m; scale bars for immunofluorescence, 10 μ m. qRT-PCR experiments represent biological replicates. Error bars denote standard error of the mean (SEM) ($n \geq 3$) (** $p < 0.01$).

The online version of this article includes the following figure supplement(s) for figure 4:

Figure supplement 1. Macropinocytosis is inhibited in SW480 colorectal cancer (CRC) spheroids by treatment with the Rac1 inhibitor EHT.

macropinocytosis, as suggested by treatment with EIPA which resulted in its re-localization to the cytoplasm (Figure 6E–F). In the nude mouse xenograft model, Tes was abundant in the cytoplasm of normal colon epithelium, while in transplanted SW480 cells it colocalized with β -catenin in the nucleus of those cells with the highest β -catenin levels (Figure 6G–I). The function of TES is poorly understood, but it has been proposed to act as a tumor suppressor (Tatarelli et al., 2000; Tobias et al., 2001).

Focal adhesion kinase (FAK) is a key regulator of FA maintenance and signaling, known to interact with the Wnt pathway in various ways (Chuang et al., 2022). In HEK293 BAR/Renilla cells, treatment with the FAK inhibitor PF-00562271 strongly decreased β -catenin signaling by LiCl inhibition of GSK3 (Figure 6J; see also Figure 6—figure supplement 1A–B). In the context on the *Xenopus* embryo, treatment with the FAK inhibitor (100 μ M, 7 min at 32-cell stage) resulted in ventralized embryos with reduced dorsal-anterior structures such as central nervous system, in particular brain structures stained by the pan-neural marker Sox2 (Figure 6K–L). The addition of the tumor promoter PMA to SW480 cells (in which the Wnt pathway is constitutively activated) strongly increased protein levels of β -catenin and FAK protein (Figure 6—figure supplement 1C–D). The effect of PMA on β -catenin and FAK levels was abrogated by EIPA or Baf treatment (Figure 6—figure supplement 1C–H), suggesting that it requires macropinocytosis and lysosome activity.

Taken together, the results suggest a possible involvement of FA components and FAK in Wnt signaling, in particular during the endogenous early Wnt signal in *Xenopus* embryos. The results also suggest that nuclear Tes could serve as a Wnt-regulated marker in CRC. The regulated shuttling of FA components between nucleus and cytoplasm is probably just the tip of the iceberg of the role of membrane trafficking in Wnt-driven cancers.

Discussion

The present work was inspired by the discovery that PMA increases macropinocytosis (Swanson, 1989). PMA plays an important role in our understanding of tumor biology because it is the archetypal tumor promoter, able to promote tumor growth without further mutation of the DNA (Berenblum, 1941; Weinberg and Weinberg, 2007). Our experiments support the role of membrane trafficking in canonical Wnt signaling in embryos and cancer cells and are summarized in the model in Figure 7. The main finding was a synergistic positive effect between the tumor promoter phorbol ester (PMA) and activation of the Wnt/ β -catenin pathway that was blocked by EIPA or Baf (Figure 1). EIPA is a macropinocytosis inhibitor of the Na⁺/H⁺ exchanger at the plasma membrane; it causes acidification of the submembranous cytoplasm and prevents actin polymerization (Koivusalo et al., 2010). Bafilomycin A (Baf) is an inhibitor of the V-ATPase that drives membrane trafficking and lysosomal acidification. PMA activates the PKC pathway (Takai et al., 1979; Nishizuka, 1984), and there is previous literature on the crosstalk between the PKC and Wnt pathways (Schwarz et al., 2013; Goode et al., 1992). What is novel in the present paper is the crosstalk between macropinocytosis, membrane trafficking, lysosomes, FAs, PMA, and Wnt signaling.

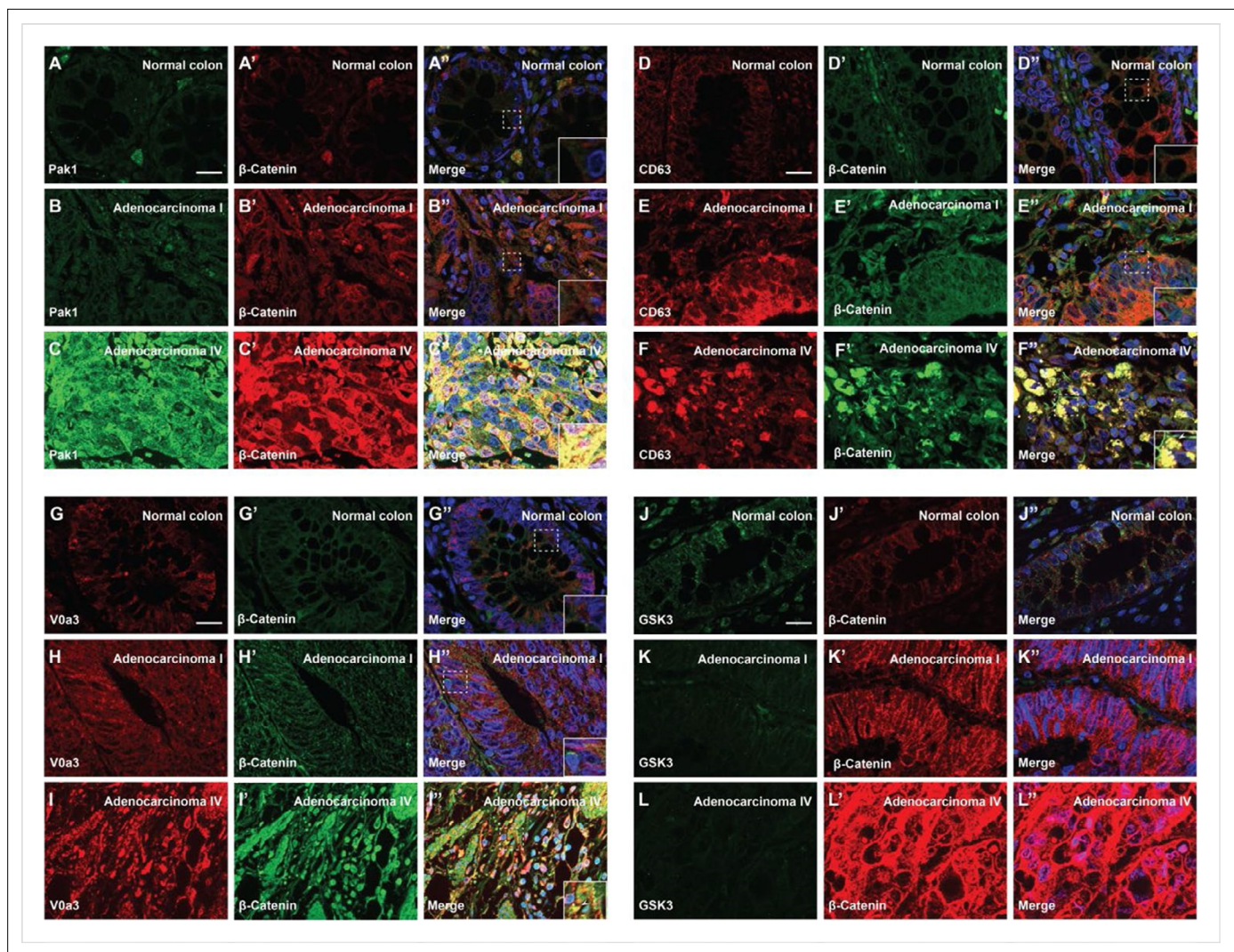


Figure 5. Degree of human colorectal cancer (CRC) malignancy is associated with increased macropinocytosis/multivesicular body (MVB)/lysosome markers and decreased GSK3 levels. (A–A'') Normal human colon paraffin section stained with the macropinocytosis marker Pak1 and β -catenin, respectively. (A'') Merged image with DAPI (4',6-diamidino-2-phenylindole), a few cells colocalize both markers. (B–B'') Pak1 and β -catenin levels are moderately increased at an early stage I CRC adenocarcinoma. (C–C'') Strong colocalization between Pak1 and β -catenin was observed in advanced stage IV CRC (inset). (D–D'') Normal human colon section stained with the MVB marker CD63 and β -catenin; colocalization was not observed. (E–E'') CD63 and β -catenin were stabilized in adenocarcinoma I, and moderate colocalization was found between CD63 and β -catenin. (F–F'') CD63 and β -catenin were strongly stabilized and colocalized in adenocarcinoma stage IV CRC. (F'') Merge; note the striking colocalization between the MVB marker CD63 and β -catenin in advanced stages of cancer (inset). (G–G'') Normal colon stained for V0a3 (a subunit of V-ATPase that marks lysosomes) and β -catenin. (H–H'') Stage I adenocarcinoma with moderately increased levels of lysosomes and β -catenin. (I–I'') Strong co-localization of lysosomes and β -catenin in stage IV CRC (see inset). (J–J'') Human colon array stained with the GSK3 and β -catenin. (K–K'') GSK3 decreases, and β -catenin increases, at early stages of carcinogenesis. (L–L'') GSK3 levels are very low in advanced CRC compared to normal human colon, while β -catenin levels are very high. Scale bars, 10 μ m. Also see **Figure 5—figure supplement 1** for quantifications and mouse xenografts of CRC cell.

The online version of this article includes the following figure supplement(s) for figure 5:

Figure supplement 1. Malignancy of colorectal cancer positively correlates with macropinocytosis, multivesicular body (MVB), and lysosome markers, and inversely correlates with GSK3 levels.

Macropinocytosis, Rac1, and Wnt signaling

Wnt signaling triggers macropinocytosis and lysosomal acidification (Albrecht et al., 2020). In cultured cells, Wnt3a-induced macropinocytosis was increased by PMA and blocked by EIPA. In embryo micro-injections, Baf blocked the synergy between PMA and the Wnt-mimic LiCl. Baf also blocked development of the endogenous axis, particularly the head region when embryos were briefly immersed in Baf

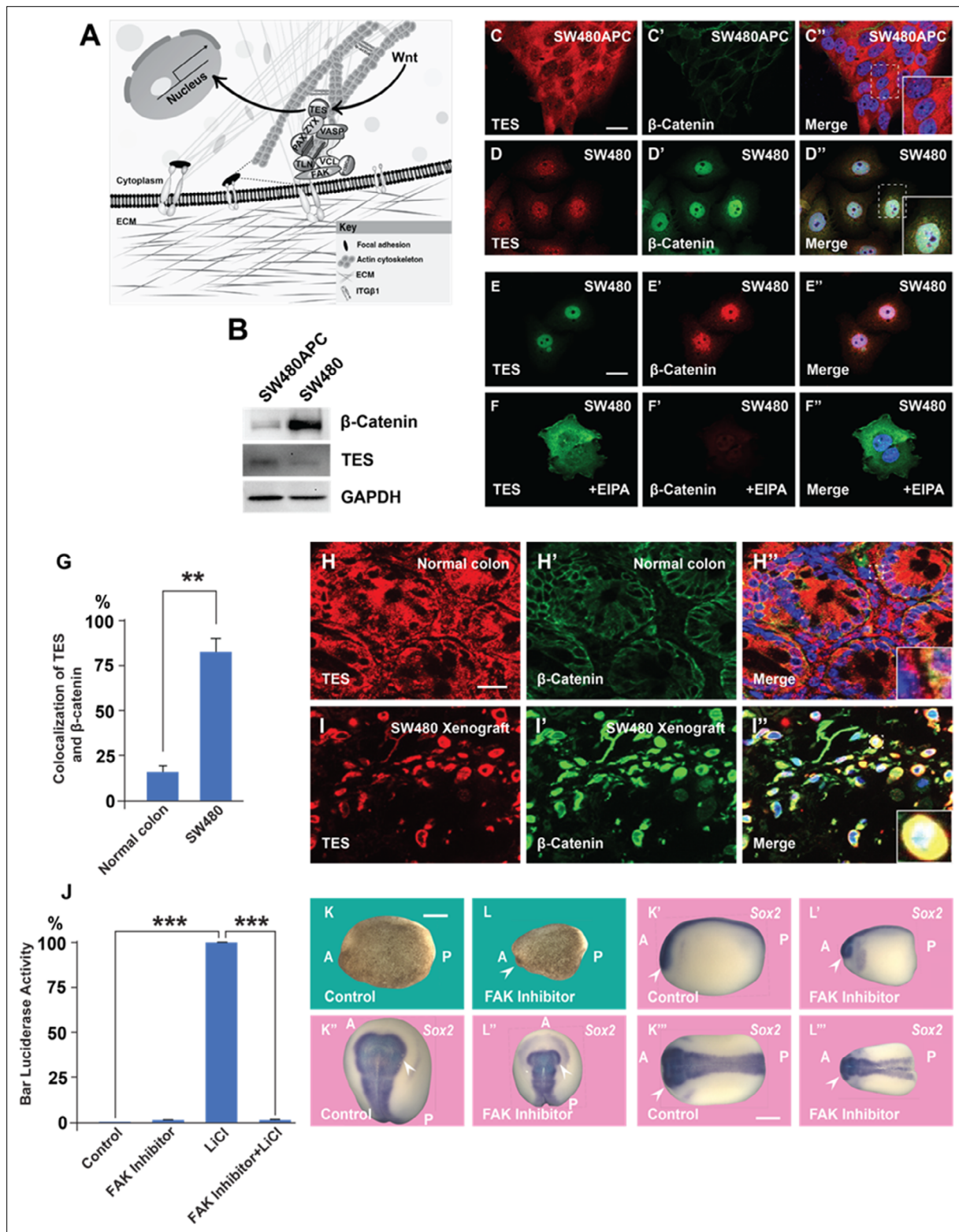


Figure 6. A focal adhesion protein changes its nucleocytoplasmic distribution after Wnt signaling activation, and focal adhesion kinase inhibition affects Wnt/ β signaling. **(A)** Diagram showing how focal adhesions connect the actin cytoskeleton to the extracellular matrix via integrins; Wnt signaling changes the distribution of the focal adhesion protein Tes from focal adhesion sites to the nucleus. **(B)** Western blot of SW480 cells with and without stable expression of Adenomatous Polyposis Coli (APC); note the reduction in Tes expression and increase in β -catenin levels in SW480 cells lacking full-length APC. GAPDH was used as loading control. **(C-C')** Colon cancer cell line SW480 stably restored with full-length APC stained with Tes and β -Catenin. Scale bars are shown in the bottom left of each panel.

Figure 6 continued on next page

Figure 6 continued

β -catenin antibodies. Note that in SW480APC cells TES is absent from the nucleus (inset). (D–D'') In SW480 colorectal cancer (CRC) cells both Tes and β -catenin are located the nucleus (inset). (E–E'') Colon cancer cell line SW480 stained with Tes (in green) and β -catenin (in red) antibodies as a control. (F–F'') Inhibiting macropinocytosis with 5-(N-Ethyl-N-isopropyl) Amiloride (EIPA) (40 μ M) treatment restored the cytoplasmic localization of the focal adhesion protein Tes in SW480 cells and strongly inhibited β -catenin levels. (G) Quantification of colocalization between Tes and β -catenin in normal mouse colon and SW480 xenografts. Error bars denote standard error of the mean (SEM) ($n \geq 3$) (** $p < 0.01$) (H–H'') Immunohistochemistry of the CD1 NU/NU normal mouse colon showing Tes and β -catenin distribution. (H'') Merge showing modest co-localization (inset). (I–I'') Immunohistochemistry images of the CD1 NU/NU mouse xenograft model showing nuclear TES, which is strongly colocalized with β -catenin levels in transplanted human SW480 cancer cells (inset). Scale bars, 10 μ m. (J) Reporter assay in HEK293T cells stably expressing β -catenin activity reporter (BAR) and Renilla, showing strong inhibition of the β -catenin transcriptional activity induced by Lithium Chloride (LiCl) (40 mM) by focal adhesion kinase (FAK) inhibitor PF-00562271 (20 μ M) after overnight treatment. Error bars denote standard error of the mean (SEM) ($n \geq 3$) (** $p < 0.001$). (K–L'') Immersion of *Xenopus* embryos in FAK inhibitor (7 min at 32-cell stage) inhibits head and dorsal development. Bright field and several angles of in situ hybridization with pan-neural marker Sox2 are shown to demonstrate reduction in brain structures indicated by the arrowhead; scale bar 500 μ m.

The online version of this article includes the following source data and figure supplement(s) for figure 6:

Source data 1. Original files with the uncropped western blots used in **Figure 6B**.

Figure supplement 1. The phorbol ester phorbol 12-myristate 13-acetate (PMA) stabilizes focal adhesion kinase (FAK) and enhances β -catenin levels in cells with constitutive Wnt signaling.

at the 32-cell stage, as previously reported (Tejeda-Muñoz and De Robertis, 2022a). Using order-of-addition experiments in which embryos were treated with 7 min pulses of Baf or the GSK3 inhibitor LiCl, we showed that the requirement for lysosome acidification was due to inhibition of the Wnt pathway at the crucial 32-cell stage (Figure 2). Co-injection experiments of DN-GSK3 or β -catenin mRNAs with DN-Rab7 or EIPA supported the view that membrane trafficking and macropinocytosis are essential for the induction of secondary axes in *Xenopus* (Figure 3).

Rac1 is an upstream regulator of the Pak1 kinase required for the actin machinery that drives macropinocytosis (Redelman-Sidi et al., 2018). Rac1 is a member of the family of small Rho GTPases that drives the formation of lamellipodia, membrane ruffles, and macropinocytosis (Hall, 1998; Egami et al., 2014; Fujii et al., 2013). The Rac1 inhibitor EHT1864 (Hampsch et al., 2017) strongly ventralized the endogenous axis of embryos after brief incubation at the critical 32-cell stage (Figure 4). In co-injection experiments EHT blocked the axis-inducing effect of microinjected xWnt8 mRNA. Rac1 is a regulator of multiple signaling cascades and has been reported to increase the nuclear translocation of β -catenin (Wu et al., 2008). Other work has shown that Rac1 binds to β -catenin and promotes the formation of nuclear β -catenin/LEF1 complexes (Jamieson et al., 2015). In the context of the present study, Rac1 protein levels were increased by treatments that enhanced Wnt/ β -catenin signaling. The results suggest that Rac1 may crosstalk with Wnt through macropinocytosis in early development.

Lysosomes, cancer and FAs

The lysosome is considered a promising target for the treatment of cancer (Fehrenbacher and Jäättelä, 2005; Lawrence and Zoncu, 2019). We used human CRC tissue arrays to analyze whether lysosomes and membrane trafficking correlate with cancer progression. Most colorectal tumors are initiated by mutation of APC, which drives Wnt signaling (Kinzler and Vogelstein, 1996; Segditsas and Tomlinson, 2006). It was found that advanced grades of colon carcinoma correlated with increased immunostaining for Pak1 (a driver of macropinocytosis), CD63 (a MVB/late endosome marker), and V0a3 (a lysosomal V-ATPase marker) (Figure 5). Importantly, cancer malignancy correlated with low levels of GSK3 staining, both in human samples and nude mouse SW480 xenografts. Cancer cells require lysosome function and have changes in lysosomal volume and subcellular localization during oncogenic transformation (Kirkegaard and Jäättelä, 2009; Kirkegaard and Jäättelä, 2009; Gocheva and Joyce, 2007; Kolwijck et al., 2010). V-ATPase has been associated with acidification of the extracellular milieu in tumors (Capecchi and Forgac, 2013; Hinton et al., 2009; Perona and Serrano, 1988). Extracellular tumor acidification is probably due to increased numbers of lysosomes which are exocytosed, since V0a3 was located within the cytoplasm in advanced cancer or xenografts in mice (Figure 5I and Figure 5—figure supplement 1I). The results reported here support the view (Taelman et al., 2010; Tejeda-Muñoz and De Robertis, 2022a) that Wnt-driven lysosomal acidification and GSK3 sequestration may play a role in cancer. In addition, MVB/lysosome components could serve as markers for advanced Wnt-driven cancers.

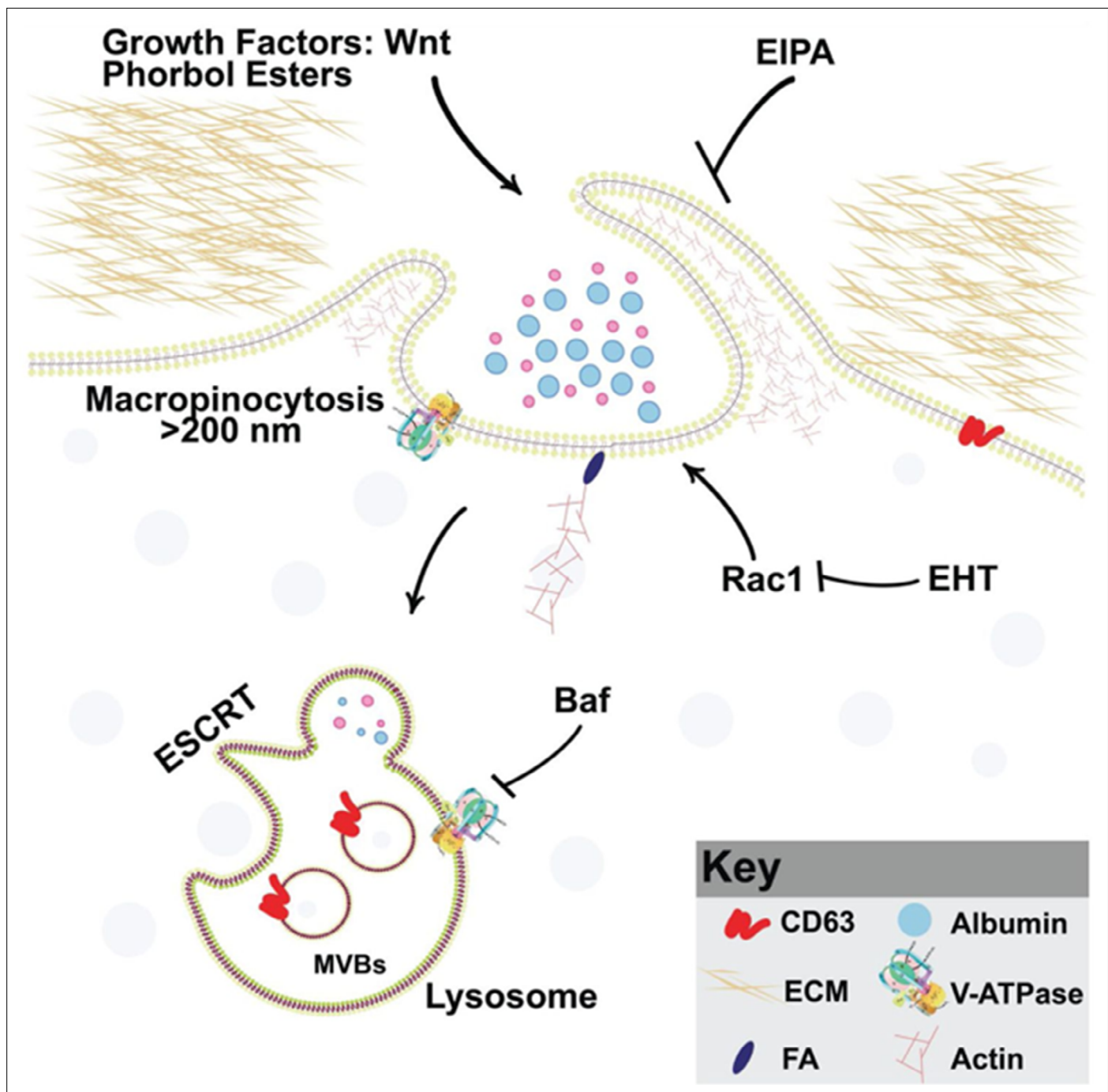


Figure 7. Model of phorbol 12-myristate 13-acetate (PMA) synergy with Wnt signaling through macropinocytosis and membrane trafficking. Wnt and PMA are activators of macropinocytosis, which is a cell drinking process driven by an actin meshwork that requires the small GTPase Rac1. Membrane trafficking into multivesicular bodies (MVBs)/lysosomes (marked by CD63) requires acidification by V-ATPase, which is inhibited by Bafilomycin A (Baf). Phorbol esters such as PMA stimulate macropinocytosis and synergize with Wnt signaling. Inhibiting macropinocytosis with the membrane trafficking inhibitors 5-(N-Ethyl-N-isopropyl) Amiloride (EIPA), Baf, or the Rac1 inhibitor EHT1864 block Wnt signaling and its cooperation with the tumor promoter PMA.

The cytoskeleton interacts with proteins of the extracellular matrix through FAs (Geiger *et al.*, 2001). We previously reported that Wnt causes the endocytosis of FA proteins and depletion of Integrin $\beta 1$ from the cell surface (Tejeda-Muñoz *et al.*, 2022c). We now report that Tes, a member of the PET-LIM family of FA proteins, changed subcellular localization after Wnt activation in SW480 cells. Other FA proteins shuttle between the nucleus, cytoplasm, and FA sites, and may play important roles in cancer

(Chaturvedi et al., 2012; Nix and Beckerle, 1997). FAK is a key signaling kinase that regulates FA signaling. In SW480 cells (which have constitutive Wnt signaling), FAK protein levels were increased by PMA, and this was inhibited by EIPA or Baf. The FAK inhibitor PF-00562271 inhibited β -catenin signaling in Luciferase assays, and reduced dorsal-anterior structures such as CNS in embryos after incubation for 7 min at 32-cell stage that is critical for the early Wnt signaling. These results suggest a possible crosstalk between FAs, Wnt signaling and tumor promoters in cancer progression and embryonic development.

Cooperation between PMA and Wnt

PMA activates PKC signaling by mimicking its physiological second messenger DAG, an activator of PKC. There is previous literature implicating PKC isozymes in the regulation of the Wnt signaling pathway (Fang et al., 2002; Goode et al., 1992) and that different PKC isoforms can have positive or negative effects on Wnt/ β -catenin signaling (Ohno and Nishizuka, 2002). However, the molecular role of PKC isoforms in the Wnt signaling pathway remains poorly understood (Bhatia and Spiegelman, 2005; Schwarz et al., 2013). It has been reported that PMA can stabilize CK1 ϵ (Casein Kinase 1 epsilon), enhance its kinase activity, and induce LRP6 phosphorylation at Thr1479 and Ser1490, leading to the activation of the Wnt/ β -catenin pathway (Su et al., 2018). In the context of the present study, it is relevant that Wnt signaling rapidly triggers macropinocytosis and trafficking into lysosomes (Tejeda-Muñoz et al., 2019). Macropinocytosis requires the activation of phosphoinositide 3 kinase (PI3K) which leads to the formation of patches of PIP₃ in the inner plasma membrane leaflet that initiate the formation of macropinocytotic cups (Egami et al., 2014; Yoshida et al., 2018). During macropinosome formation, the PIP₃ lipid is then converted into DAG by phospholipase C gamma 1 (PLC γ 1), and DAG activates PKC signaling (Yoshida et al., 2018). It seems likely that PMA may facilitate Wnt/ β -catenin signaling by mimicking the DAG lipid in macropinosomes. We have tested this hypothesis by direct addition of DAG to cells and found cooperation with Wnt signaling (Azbazdar et al., 2023).

In the future, it will be interesting to study the role of chronic inflammation on macropinocytosis and tumor promotion. Active PKC phosphorylates many targets, including I κ B kinase (IKK), which becomes activated and phosphorylates inhibitor of NF- κ B (I κ B), targeting it for degradation and triggering inflammation (Weinberg and Weinberg, 2007). Tumor progression is promoted by many chronic inflammatory agents such as turpentine, distilled alcoholic beverages, hepatitis viruses, and *Helicobacter pylori* that do not cause mutations by themselves but regulate the expansion of previously mutant clones (Rous and Kidd, 1941; Weinberg and Weinberg, 2007). Taken together, the results presented here suggest that macropinocytosis, lysosomal activity, and membrane trafficking are possible therapeutic targets for tumor progression in Wnt-driven cancers.

Materials and methods

Key resources table

Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
Cell line (<i>Homo sapiens</i>)	HeLa (human cervical adenocarcinoma)	ATCC	RRID: CVCL_0030	
Cell line (<i>Homo sapiens</i>)	SW480 (human colorectal)	ATCC	RRID: CVCL_0546	
Biological sample (<i>Xenopus laevis</i> male)	Sperm	<i>Xenopus</i> I	5215 Albino <i>Xenopus laevis</i> Sexually Mature Male -/-/M	Used for fertilization
Biological sample (<i>Xenopus laevis</i> female)	Egg	<i>Xenopus</i> I	4280 Sexually Mature Female 10.5–11 cm -/-/F	Used for fertilization
Biological sample (CD1 NU/NU nude mouse female)		Charles River Laboratories		Xenograft
Antibody	Pak1 antibody rabbit polyclonal	Abcam	Cat# ab131522, RRID: AB_11156726	IF (1:100)
Antibody	FAK antibody rabbit polyclonal	Cell Signaling	Cat# 3285, RRID: AB_2269034	IF (1:100)

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Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
Antibody	Rac-1 antibody rabbit polyclonal	Thermo Fisher	Cat# PA1-091	IF (1:100)
Antibody	CD63 antibody mouse monoclonal	Abcam	Cat# ab59479 RRID:AB_940915	IF (1:100)
Antibody	TES antibody rabbit polyclonal	ATLAS	Cat# HPA018123 RRID:AB_1857900	IF (1:100)
Antibody	GSK3 antibody mouse monoclonal	Abcam	Cat# ab93926, RRID:AB_10563643	IF (1:100)
Antibody	Transferrin receptor (TfR) antibody mouse monoclonal	Thermo Fisher	Cat# 13-6800	IF (1:100)
Antibody	ATP6V0a3 antibody rabbit polyclonal	Novus	Cat# nbp1-89333 RRID:AB_11016312	IF (1:100)
Antibody	GAPDH antibody rabbit monoclonal	Cell Signaling	mAb #2118	WB (1:1000)
Antibody	β -Catenin antibody rabbit polyclonal	Thermo Fisher Scientific	Cat# 71-2700; RRID:AB_2533982	IF (1:100)
Antibody	β -Catenin antibody mouse monoclonal	Thermo Fisher Scientific	Cat# MA1-2001; RRID:AB_326078	IF (1:100)
Antibody	IgG, HRP-linked antibody (horse anti-mouse polyclonal)	Cell Signaling	Cat# 7076; RRID:AB_330924	WB (1:2500)
Antibody	IgG, HRP-linked antibody (goat anti-mouse polyclonal)	Cell Signaling	Cat# 7074; RRID:AB_2099233	WB (1:2500)
Antibody	IgG H&L (Alexa Fluor 594) preadsorbed (goat anti-mouse polyclonal)	Abcam	Cat# ab150120; RRID:AB_2631447	IF (1:200)
Antibody	IgG H&L (Alexa Fluor 594) preadsorbed (goat anti-rabbit polyclonal)	Abcam	Cat# ab150084; RRID:AB_2734147	IF (1:200)
Antibody	IgG H&L (Alexa Fluor 488) preadsorbed (goat anti-mouse polyclonal)	Abcam	Cat# ab150117; RRID:AB_2734147	IF (1:200)
Antibody	IgG H&L (Alexa Fluor 488) preadsorbed (ab150081) (goat anti-rabbit polyclonal)	Abcam	Cat# ab150081; RRID:AB_2734747	IF (1:200)
Antibody	IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor 488 (goat anti-mouse polyclonal)	Invitrogen	Cat# A-11001; RRID:AB_2534069	IF (1:200)
Antibody	IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor 568 (goat anti-rabbit polyclonal)	Invitrogen	Cat# A-11011; RRID:AB_143157	IF (1:200)
Recombinant DNA reagent	Dominant-negative (DN)-GSK3-GFP (<i>Xenopus laevis</i>) (plasmid)	Addgene	RRID:Addgene_29681	K85R and K86R
Recombinant DNA reagent	DN-Rab7 (<i>Homo sapiens</i>) (plasmid)	Addgene	RRID:Addgene_12660	Dominant-negative plasmid
Recombinant DNA reagent	pCS2-mGFP (plasmid)	Addgene	RRID:Addgene_14757	Membrane bound form of EGFP
Recombinant DNA reagent	CD63-RFP (<i>Homo sapiens</i>) (plasmid)	Addgene	RRID:Addgene_62964	RFP tag
Recombinant DNA reagent	xWnt8myc (wnt8a.L Frog) (plasmid)	Addgene	RRID:Addgene_16863	
Recombinant DNA reagent	β -Catenin-activated reporter (BAR) (plasmid)	Addgene	RRID:Addgene_12456	Beta-catenin reporter. TCF/LEF sites upstream of a luciferase reporter

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Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
Recombinant DNA reagent	Renilla reporter (plasmid)	Addgene	RRID: Addgene_62186	MuLE (Multiple Lentiviral Expression) Entry vector containing a CMV promoter and renilla luciferase module
Recombinant DNA reagent	Lipofectamine 3000	Thermo Fisher	Cat# L3000001	
Peptide, recombinant protein	Wnt3a	Peprtech	Cat# 315-20	
Commercial assay or kit	mMESSAGE mMACHINE SP6 Transcription Kit	Thermo Fisher	Cat# AM1340	
Commercial assay or kit	Dual-Luciferase Reporter Assay System	Promega	Cat# E1500	
Chemical compound, drug	Fibronectin	Thermo Fisher	Cat# 33016015	
Chemical compound, drug	Dextran Tetramethylrhodamine (TMR-Dx) 70,000	Thermo Fisher	Cat# D1818	
Chemical compound, drug	5-(N-Ethyl-N-isopropyl) amiloride (EIPA)	Sigma	Cat# A3085	
Chemical compound, drug	Bafilomycin A1	Selleckchem	Cat# S1413	
Chemical compound, drug	Digitonin	Sigma	Cat# 300410	
Chemical compound, drug	Phalloidin	Abcam	Cat# ab176759	
Chemical compound, drug	Lithium chloride (LiCl)	Sigma	Cat# L4408	
Chemical compound, drug	PF-00562271 (FAK inhibitor)	Selleckchem	Cat# S2672	
Chemical compound, drug	Phorbol 12-myristate 13-acetate (PMA)	TOCRIS	Cat# 1201	
Chemical compound, drug	5-(N-Ethyl-N-isopropyl) amiloride (EIPA)	Sigma	Cat# A3085	
Chemical compound, drug	EHT 1864	Selleckchem	Catalog# S7482	
Chemical compound, drug	Triton X-100	Thermo Fisher	Cat# HFH10	
Chemical compound, drug	Paraformaldehyde	Sigma	Cat# P6148	
Chemical compound, drug	Fibronectin	Sigma	Cat# F4759	
Chemical compound, drug	Protease inhibitors	Roche	Cat# 04693132001	
Chemical compound, drug	Phosphatase inhibitors	Calbiochem	Cat# 524629	
Software, algorithm	ImageJ	NIH	http://imagej.nih.gov/ij/	
Software, algorithm	AxioVision 4.8	Zeiss	http://Zeiss.com	
Software, algorithm	Zen 2.3 imaging software	Zeiss	http://Zeiss.com	
Software, algorithm	R	R Core Team	https://cran.r-project.org	

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Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
Other	10 cm dish	Thermo Fisher	Cat# 174903	
Other	8-Well glass-bottom chamber slides	ibidi	Cat# 80827	
Other	Circular coverslips	ibidi	Cat# 10815	
Other	12-Well dish	Thermo Fisher	Cat# 150628	
Other	DMEM (culture medium)	Thermo Fisher	Cat# 11965092	
Other	L-15 (culture medium)	Thermo Fisher	Cat# 11415064	
Other	Glutamine	Thermo Fisher	Cat# 25030081	
Other	Fetal bovine serum (FBS)	Thermo Fisher	Cat# 16000044	
Other	Bovine serum albumin (BSA)	Thermo Fisher	Cat# 9048468	
Other	Pen-Strep antibiotics	Thermo Fisher	Cat# 15140122	
Other	PBS	Gibco	Cat# 10-010-023	
Other	PBS	Fisher Scientific	Cat# BP3994	
Other	Fluoroshield Mounting Medium with DAPI	Abcam	Cat# ab104139	
Other	IM 300 microinjection pump	Narishige International USA, Inc	N/A	
Other	Axio Observer Z1 Inverted Microscope with Apotome	Zeiss	N/A	

Materials availability statement

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Edward M. De Robertis.

No new materials were generated in this study. Requests for reagents used should be directed and will be fulfilled by the Lead Contact. No custom code, software, or algorithm central to supporting the main claims of the paper were generated in this manuscript.

Experimental model and subject details

Tissue culture and transfection

HeLa (ATCC, CRL-2648) and HEK293T cells stably expressing BAR and Renilla reporters were cultured in DMEM (Dulbecco's modified Eagle medium), supplemented with 10% fetal bovine serum (FBS), 1% glutamine, and penicillin/streptomycin. SW480 cells and SW480APC cells (*Faux et al., 2004*) were cultured in DMEM/F12 (DMEM:Nutrient Mixture F-12), supplemented with 5% FBS, 1% glutamine, and penicillin/streptomycin. The cells were seeded at a cell density of 20–30%, and experiments were performed when cells reached between 70 and 80% confluency. Cells were cultured for 6–8 hr in a medium containing 2% FBS before all treatments. All cell lines tested negative for mycoplasma contamination. ATCC cell line authentication services were performed to confirm identity of cultured cell types.

Xenopus embryo microinjection

This study was performed in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All of the animals were handled according to approved Institutional Animal Care and Use Committee (IACUC) protocols (D16-00124) of the University of California, Los Angeles, Medical School. The protocol was approved by the Committee on the Ethics of Animal Experiments of the University of California, Los Angeles (Permit Number: ARC-1995-129). *Xenopus laevis* embryos were fertilized in vitro using excised testis and staged as described (*Colozza and De Robertis, 2014; Tejada-Muñoz and De Robertis, 2022a*). In

in vitro synthesized mRNAs were introduced into embryos by microinjection using an IM 300 Microinjector (Narishige International USA, Inc) of 4 nl into the marginal zone of a ventral blastomere at the 4-cell stage. pCS2-DN-GSK3 β , β -catenin, and DN-Rab7 were linearized with NotI and transcribed with SP6 RNA polymerase using the Ambion mMessage mMACHINE kit. Embryos were injected in 1 \times MMR (Marc's Minimal Ringers) and cultured in 0.1 \times MMR.

Human colon cancer tissue array and immunochemistry

Three sets of colon cancer tissue arrays containing 90 cases of adenocarcinoma and 90 adjacent normal colon tissue (180 tissue cores total) were obtained from [TissueArray.com](https://www.tissuearray.com). The company stated that each specimen collected from any clinic was consented to by both the hospital and the individual, and that discrete legal consent forms were obtained and the rights to hold research uses for any purpose or further commercialized uses were waived. Double stained immunohistochemistry was performed on paraffin-embedded tissue samples which were deparaffinized in xylene and rehydrated using graded alcohols. For antigen retrieval, slides were incubated at 95°C for 40 min in citrate buffer (10 mM, 0.05% Tween 20, pH 6.0). Tissue sections were then fixed with 4% paraformaldehyde (Sigma #P6148) for 15 min, treated with 0.2% Triton X-100 in phosphate-buffered saline (PBS; Gibco) for 10 min, and blocked with 5% bovine serum albumin (BSA) in PBS overnight. Primary and secondary antibodies were added overnight at 4°C. The samples were washed three times with PBS after each treatment, and coverslips were mounted with Fluoroshield Mounting Medium with DAPI (ab104139). Immunofluorescence was analyzed and photographed using a Zeiss Imager Z.1 microscope with Apotome.

Xenograft tumor model

For the CD1 NU/NU nude mouse model, SW480 cells were collected by trypsinization. Then, 7.5 \times 10⁵ cells per injection site were resuspended in high-concentration Matrigel (BD Biosciences, Franklin Lakes, NY), diluted in PBS to 50% final concentration, and subcutaneously injected into the flanks of an 8-week-old mouse. After day 7, when the tumor was established, the tumor was measured three times a week until completion. After 3 weeks, tissue was obtained from animals euthanized by CO₂ inhalation. Tumor and normal colon were fixed, paraffin-embedded, and 5 μ m histological sections were mounted on slides. Mouse experiments were approved by the UCLA Animal Research Committee.

Antibodies and reagents

Total β -catenin antibody (1:1000) was purchased from Invitrogen (712700), glyceraldehyde-3-phosphate dehydrogenase antibody (1:1000) and FAK antibody (1:1000, 3285) were obtained from Cell Signaling Technologies, anti-ATP6V0a3 antibody (1:500) was obtained from Novus (nbp1-89333, 1:1000). CD63 antibody was obtained from Abcam.

Antibodies against Pak1 (ab131522), Ras (ab52939), GSK3 (ab93926, 1:4000), and secondary antibodies for immunostaining for cells (ab150120, ab150084, ab150117, ab150081) (1:300) were obtained from Abcam. Antibody against the FA protein Tes (HPA018123, 1:100) was obtained from Atlas antibodies. Secondary antibodies for immunostaining arrays (A-11001, A-11011 were obtained from Invitrogen). HRP-linked secondary antibodies 7076, 7074 at 1:5000 (Cell Signaling) were used for western blots and analyzed with an iBright Imaging system. EIPA (A3085), and LiCl (L4408), were obtained from Sigma. Baf (S1413) and PF-00562271 (FAK inhibitor, S2672) were purchased from Selleckchem. TMR-dextran 70 kDa was obtained from Thermo Fisher (D1818). PMA (1201) was purchased from TOCRIS.

Immunostainings

HeLa, HEK293T, SW480, and SW480APC cells were plated on glass coverslips and transferred to 2% FBS 6–12 hr before overnight experimental treatments. Coverslips were acid washed and treated with Fibronectin (10 μ g/ml for 30 min at 37°C, Sigma F4759) to facilitate cell spreading and adhesion. Cells were fixed with 4% paraformaldehyde (Sigma #P6148) for 15 min, permeabilized with 0.2% Triton X-100 in PBS Gibco for 10 min, and blocked with 5% BSA in PBS for 1 hr.

Primary antibodies were added overnight at 4°C. Cells were washed three times with PBS, and secondary antibodies were applied for 1 hr at room temperature. After three additional washes with PBS, the coverslips were mounted with Fluoroshield Mounting Medium with DAPI (ab104139).

Immunofluorescence was analyzed and photographed using a Zeiss Imager Z.1 microscope with Apotome.

Transfections

Transfections of β -cateninGFP (0.8 mg) and CA-Lrp6GFP (0.8 mg) were performed on 3T3 cells plated on coverslips for 24 hr. Cells were stained following the immunofluorescence protocol described above and stained with Rac-1 antibody.

Western blots

Cell lysates were prepared using RIPA (Radioimmunoprecipitation Assay) buffer (0.1% NP40, 20 mM Tris/HCl pH 7.5), 10% glycerol, together with protease (Roche #04693132001) and phosphatase inhibitors (Calbiochem #524629), and processed as described (Tejeda-Muñoz *et al.*, 2019).

Luciferase assay

Experiments were performed with stably transfected HEK293T cells stably expressing BAR and Renilla reporters (Albrecht *et al.*, 2020), treated with LiCl and with or without Bafilomycin for 8 hr, and Luciferase activity measured with the Dual-Luciferase Reporter Assay System (Promega) according to the manufacturer's instructions, using the Glomax Luminometer (Promega). Luciferase values of each sample were normalized for Renilla activity.

3D spheroid cell culture

SW480 cells were cultured in a Petri dish using DMEM:F-12 medium with 5% FBS. The top cover was removed from 60 mm tissue culture dishes and 3 ml of PBS placed in the bottom of the dish to act as a hydration chamber. Cells were counted and 500 cells were added as 25 μ l drops deposited onto the Petri dish cover, and immediately inverted over the humid chamber. At least 6 drops per condition were plated, keeping enough distance between each other. The inverted drop cultures were incubated at 37°C in 5% CO₂/95% humidity.

The drops were monitored daily; after 4 days, aggregates had been formed and EHT treatment was added to the spheroids. After 4 days, spheroids were incubated with TMR-dextran 70 kDa (1 mg/ml) for 1 hr, and each spheroid was photographed with an Axio Zoom.V16 Stereo Zoom Zeiss microscope with apotome function.

Time-lapse imaging

SW480 cells were seeded on glass bottom culture dishes (MatTek Corp). After 4 hr, the cells were transfected with mGFP with Lipofectamine 3000 (Albrecht *et al.*, 2020). The video was taken after 3 days in culture. Control cells had DMSO added for 15 min. Then, cells were treated with PMA (0.3 μ M) for 15 min. Filming was with a Zeiss Observer.Z1 microscope and AxioVision 4.8.2 SP3 software was used for imaging.

TMR-dextran assays

3T3 or SW480 cells were seeded on cover glass coverslips for 12–18 hr as described in Tejeda-Muñoz *et al.*, 2022c. The cells were incubated with Wnt3a overnight in the presence or absence of PMA and EIPA. After incubation, the cells were treated with 1 mg/ml TMR-dextran 70 kDa for 1 hr, then washed, fixed, and blocked for 1 hr with 5% BSA in PBS to reduce background from nonspecific binding. The coverslips were mounted using Fluoroshield Mounting Medium with DAPI (Abcam, ab104139). Immunofluorescence images were performed using a Zeiss Imager Z.1 microscope with Apotome.

Bafilomycin and FAK inhibitor treatments, microinjection, and in situ hybridization in *Xenopus* embryos

We determined experimentally that optimal ventralization with loss of head structures of *Xenopus* embryos was obtained after incubation with Baf (5 μ M) or FAK inhibitor PF-00562271 (100 μ M) for 7 min at the 32-cell stage in 0.1 \times MMR solution. After incubation, embryos were washed two times with 0.1 \times MMR solution and cultured overnight until early tailbud tadpole stage. Embryos were microinjected once ventrally at the 4-cell stage with any of the following reagents: LiCl (300 mM), PMA (500 nM), EIPA (1 mM), EHT (1 mM), or mRNAs such as DN-GSK3- β (150 pg), DN-Rab-7 (500 pg), and

xWnt8 (2 pg), either alone or co-injected with the other reagents. In some experiments, microinjected embryos were incubated in 0.1× MMR solution until the 32-cell stage and transferred to 0.1× MMR solution with or without Baf (5 μM) and incubated for 7 min. Embryos were then washed twice with 0.1× MMR solution and cultured until tadpole stage. In situ hybridization was performed as described at <http://www.hhmi.ucla.edu/derobertis>.

LiCl and Bafilomycin sequential treatments

Whole embryos at the 32-cell stage were incubated with LiCl (300 mM) alone for 7 min. After this treatment, the embryos were washed two times with 0.1× MMR solution and cultured until the tadpole stage, or incubated again for 7 min in 0.1 MMR solution containing 5 μM Baf, and washed as described before. In the converse experiment, embryos were first treated with Baf and then with LiCl. All treatments were done at the 32-cell stage, which plays a key role in the early Wnt signal (*Kao et al., 1986; Tejeda-Muñoz et al., 2022c*).

qRT-PCR

Quantitative RT-PCR experiments using *Xenopus* embryos were performed as previously described (*Colozza and De Robertis, 2014*). Primer sequences for qRT-PCR were as follows:

Gene	Forward	Reverse	Use
ODC	CAGCTAGCTGTGGTGTGG	CAACATGGAACTCACACC	qRT-PCR
Siamois	AAGATAACTGGCATTCTGAGC	GGTAGGGCTGTGTATTGAAGG	qRT-PCR
Xnr3	CGAGTGCAAGAAGGTGGACA	ATCTTCATGGGGACACGGA	qRT-PCR
Sizzled	GTCTTCCTGCTCTCTGC	AACAGGGAGCACAGGAAG	qRT-PCR
Vent1	GGCACCTGAACGGAAGAA	GATTTTGAACCCAGGTTTTGAC	

Quantification and statistical analysis

Data were expressed as means and standard error of the means (SEMs). Statistical analysis of the data was performed using the Student t-test; a p-value of <0.01** was considered statistically significant for differences between means. Fluorescence was quantified in control versus treated cells using ImageJ software analyses with $n > 30$ cells or $n > 30$ images from arrays from humans or mouse sample per condition. Fluorescence intensity was normalized in images compared in each condition and results from three or more independent experiments were presented as the mean ± SEM.

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Author contributions

Nydia Tejeda-Munoz, Conceptualization, Data curation, Writing – original draft, Writing – review and editing; Yagmur Azbazar, Data curation, Formal analysis, Investigation, Writing – review and editing; Julia Monka, Grace Binder, Alex Dayrit, Raul Ayala, Investigation; Neil O'Brien, Funding acquisition, Investigation; Edward M De Robertis, Conceptualization, Data curation, Funding acquisition, Investigation, Writing – original draft, Project administration, Writing – review and editing

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Ethics

This study was performed in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All of the animals were handled according to approved Institutional Animal Care and Use Committee (IACUC) protocols (D16-00124) of the University of California, Los Angeles, Medical School. The protocol was approved by the Committee on the Ethics of Animal Experiments of the University of California, Los Angeles (Permit Number: ARC-1995-129).

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Additional files

Supplementary files

- MDAR checklist

Data availability

Data have been deposited in the Cell Image Library (CIL): <http://cellimagelibrary.org/groups/55765>.

The following dataset was generated:

Author(s)	Year	Dataset title	Dataset URL	Database and Identifier
Tejeda-Muñoz N, Azbazar Y, Monka J, Binder G, Dayrit A, Ayala R, O'Brien N, De Robertis EM	2023	The PMA phorbol ester tumor promoter increases canonical Wnt signaling via macropinocytosis	http://cellimagelibrary.org/groups/55765	Cell Image Library, 55765

References

- Acebron SP**, Karaulanov E, Berger BS, Huang YL, Niehrs C. 2014. Mitotic wnt signaling promotes protein stabilization and regulates cell size. *Molecular Cell* **54**:663–674. DOI: <https://doi.org/10.1016/j.molcel.2014.04.014>, PMID: 24837680
- Albrecht LV**, Tejeda-Muñoz N, Bui MH, Cicchetto AC, Di Biagio D, Colozza G, Schmid E, Piccolo S, Christofk HR, De Robertis EM. 2020. GSK3 Inhibits Macropinocytosis and Lysosomal activity through the Wnt destruction Complex Machinery. *Cell Reports* **32**:107973. DOI: <https://doi.org/10.1016/j.celrep.2020.107973>, PMID: 32726636
- Albrecht LV**, Tejeda-Muñoz N, De Robertis EM. 2021. Cell biology of canonical wnt signaling. *Annual Review of Cell and Developmental Biology* **37**:369–389. DOI: <https://doi.org/10.1146/annurev-cellbio-120319-023657>, PMID: 34196570

- Anderson CA**, Kovar DR, Gardel ML, Winkelman JD. 2021. LIM domain proteins in cell mechanobiology. *Cytoskeleton* **78**:303–311. DOI: <https://doi.org/10.1002/cm.21677>, PMID: 34028199
- Azbazdar Y**, Tejada-Munoz N, Monka JC, Dayrit A, Binder G, Ozhan G, De Robertis EM. 2023. Addition of exogenous diacylglycerol enhances Wnt/ β -catenin signaling through stimulation of macropinocytosis. *iScience* **26**:108075. DOI: <https://doi.org/10.1016/j.isci.2023.108075>
- Bar-Sagi D**, Feramisco JR. 1986. Induction of membrane ruffling and fluid-phase pinocytosis in quiescent fibroblasts by ras proteins. *Science* **233**:1061–1068. DOI: <https://doi.org/10.1126/science.3090687>, PMID: 3090687
- Berenblum I**. 1941. The cocarcinogenic action of croton resin 1. *Cancer Research* **1**:44–48.
- Bhatia N**, Spiegelman VS. 2005. Activation of Wnt/beta-catenin/TCF signaling in mouse skin carcinogenesis. *Molecular Carcinogenesis* **42**:213–221. DOI: <https://doi.org/10.1002/mc.20077>, PMID: 15765534
- Biechle TL**, Moon RT. 2008. Assaying beta-catenin/TCF transcription with beta-catenin/TCF transcription-based reporter constructs. *Methods in Molecular Biology* **468**:99–110. DOI: https://doi.org/10.1007/978-1-59745-249-6_8, PMID: 19099249
- Capecchi J**, Forgac M. 2013. The Function of Vacuolar ATPase (V-ATPase) a subunit isoforms in invasiveness of MCF10a and MCF10CA1a Human Breast Cancer Cells. *Journal of Biological Chemistry* **288**:32731–32741. DOI: <https://doi.org/10.1074/jbc.M113.503771>
- Cardone RA**, Casavola V, Reshkin SJ. 2005. The role of disturbed pH dynamics and the Na⁺/H⁺ exchanger in metastasis. *Nature Reviews. Cancer* **5**:786–795. DOI: <https://doi.org/10.1038/nrc1713>, PMID: 16175178
- Chaturvedi A**, Hoffman LM, Welm AL, Lessnick SL, Beckerle MC. 2012. The EWS/FLI Oncogene drives changes in Cellular Morphology, Adhesion, and Migration in Ewing Sarcoma. *Genes & Cancer* **3**:102–116. DOI: <https://doi.org/10.1177/1947601912457024>, PMID: 23050043
- Chuang HH**, Zhen YY, Tsai YC, Chuang CH, Hsiao M, Huang MS, Yang CJ. 2022. FAK in Cancer: from Mechanisms to Therapeutic Strategies. *International Journal of Molecular Sciences* **23**:1726. DOI: <https://doi.org/10.3390/ijms23031726>, PMID: 35163650
- Colozza G**, De Robertis EM. 2014. Maternal syntabulin is required for dorsal axis formation and is a germ plasm component in *Xenopus*. *Differentiation; Research in Biological Diversity* **88**:17–26. DOI: <https://doi.org/10.1016/j.diff.2014.03.002>, PMID: 24798204
- Colozza G**, Jami-Alahmadi Y, Dsouza A, Tejada-Muñoz N, Albrecht LV, Sosa EA, Wohlschlegel JA, De Robertis EM. 2020. Wnt-inducible LRP6-APEX2 interacting proteins identify ESCRT machinery and Trk-fused gene as components of the Wnt signaling pathway. *Scientific Reports* **10**:21555. DOI: <https://doi.org/10.1038/s41598-020-78019-5>, PMID: 33299006
- Commisso C**, Davidson SM, Soydaner-Azeloglu RG, Parker SJ, Kamphorst JJ, Hackett S, Grabocka E, Nofal M, Drebin JA, Thompson CB, Rabinowitz JD, Metallo CM, Vander Heiden MG, Bar-Sagi D. 2013. Macropinocytosis of protein is an amino acid supply route in Ras-transformed cells. *Nature* **497**:633–637. DOI: <https://doi.org/10.1038/nature12138>, PMID: 23665962
- Commisso C**, Flinn RJ, Bar-Sagi D. 2014. Determining the macropinocytic index of cells through a quantitative image-based assay. *Nature Protocols* **9**:182–192. DOI: <https://doi.org/10.1038/nprot.2014.004>
- Egami Y**, Taguchi T, Maekawa M, Arai H, Araki N. 2014. Small GTPases and phosphoinositides in the regulatory mechanisms of macropinosome formation and maturation. *Frontiers in Physiology* **5**:374. DOI: <https://doi.org/10.3389/fphys.2014.00374>, PMID: 25324782
- Escola J-M**, Kleijmeer MJ, Stoorvogel W, Griffith JM, Yoshie O, Geuze HJ. 1998. Selective Enrichment of Tetraspan Proteins on the internal vesicles of Multivesicular Endosomes and on Exosomes Secreted by Human B-lymphocytes. *Journal of Biological Chemistry* **273**:20121–20127. DOI: <https://doi.org/10.1074/jbc.273.32.20121>
- Fang X**, Yu S, Tanyi JL, Lu Y, Woodgett JR, Mills GB. 2002. Convergence of Multiple Signaling Cascades at Glycogen Synthase Kinase 3: EDG Receptor-Mediated Phosphorylation and Inactivation by Lysophosphatidic Acid through a Protein Kinase C-Dependent Intracellular Pathway. *Molecular and Cellular Biology* **22**:2099–2110. DOI: <https://doi.org/10.1128/MCB.22.7.2099-2110.2002>
- Faux MC**, Ross JL, Meeker C, Johns T, Ji H, Simpson RJ, Layton MJ, Burgess AW. 2004. Restoration of full-length adenomatous polyposis coli (APC) protein in a colon cancer cell line enhances cell adhesion. *Journal of Cell Science* **117**:427–439. DOI: <https://doi.org/10.1242/jcs.00862>
- Fehrenbacher N**, Jäättelä M. 2005. Lysosomes as targets for cancer therapy. *Cancer Research* **65**:2993–2995. DOI: <https://doi.org/10.1158/0008-5472.CAN-05-0476>, PMID: 15833821
- Fujii M**, Kawai K, Egami Y, Araki N. 2013. Dissecting the roles of RAC1 activation and deactivation in macropinocytosis using microscopic photo-manipulation. *Scientific Reports* **3**:2385. DOI: <https://doi.org/10.1038/srep02385>, PMID: 23924974
- Galluzzi L**, Spranger S, Fuchs E, López-Soto A. 2019. WNT Signaling in Cancer Immunosurveillance. *Trends in Cell Biology* **29**:44–65. DOI: <https://doi.org/10.1016/j.tcb.2018.08.005>, PMID: 30220580
- Garvalov BK**, Higgins TE, Sutherland JD, Zettl M, Scaplehorn N, Köcher T, Piddini E, Griffiths G, Way M. 2003. The conformational state of TES regulates its zyxin-dependent recruitment to focal adhesions. *The Journal of Cell Biology* **161**:33–39. DOI: <https://doi.org/10.1083/jcb.200211015>
- Geiger B**, Bershadsky A, Pankov R, Yamada KM. 2001. Transmembrane crosstalk between the extracellular matrix and the cytoskeleton. *Nature Reviews Molecular Cell Biology* **2**:793–805. DOI: <https://doi.org/10.1038/35099066>
- Gocheva V**, Joyce JA. 2007. Cysteine cathepsins and the cutting edge of cancer invasion. *Cell Cycle* **6**:60–64. DOI: <https://doi.org/10.4161/cc.6.1.3669>, PMID: 17245112

- Goode N**, Hughes K, Woodgett JR, Parker PJ. 1992. Differential regulation of glycogen synthase kinase-3 beta by protein kinase C isotypes. *The Journal of Biological Chemistry* **267**:16878–16882. DOI: [https://doi.org/10.1016/S0021-9258\(18\)41866-2](https://doi.org/10.1016/S0021-9258(18)41866-2), PMID: 1324914
- Haigler HT**, McKanna JA, Cohen S. 1979. Rapid stimulation of pinocytosis in human carcinoma cells A-431 by epidermal growth factor. *The Journal of Cell Biology* **83**:82–90. DOI: <https://doi.org/10.1083/jcb.83.1.82>, PMID: 315944
- Hall A**. 1998. Rho GTPases and the actin cytoskeleton. *Science* **279**:509–514. DOI: <https://doi.org/10.1126/science.279.5350.509>, PMID: 9438836
- Hampsch RA**, Shee K, Bates D, Lewis LD, Désiré L, Leblond B, Demidenko E, Stefan K, Huang YH, Miller TW. 2017. Therapeutic sensitivity to Rac GTPase inhibition requires consequential suppression of mTORC1, AKT, and MEK signaling in breast cancer. *Oncotarget* **8**:21806–21817. DOI: <https://doi.org/10.18632/oncotarget.15586>, PMID: 28423521
- Hanahan D**, Weinberg RA. 2011. Hallmarks of cancer: the next generation. *Cell* **144**:646–674. DOI: <https://doi.org/10.1016/j.cell.2011.02.013>, PMID: 21376230
- Hinton A**, Sennoune SR, Bond S, Fang M, Reuveni M, Sahagian GG, Jay D, Martinez-Zaguilan R, Forgac M. 2009. Function of a subunit isoforms of the V-ATPase in pH homeostasis and in vitro invasion of MDA-MB231 human breast cancer cells. *The Journal of Biological Chemistry* **284**:16400–16408. DOI: <https://doi.org/10.1074/jbc.M901201200>, PMID: 19366680
- Jamieson C**, Lui C, Brocardo MG, Martino-Echarri E, Henderson BR. 2015. RAC1 augments Wnt signaling by stimulating β -catenin-lymphoid enhancer factor-1 complex assembly independent of β -catenin nuclear import. *Journal of Cell Science* **128**:3933–3946. DOI: <https://doi.org/10.1242/jcs.167742>, PMID: 26403202
- Jeong WJ**, Yoon J, Park JC, Lee SH, Lee SH, Kaduwal S, Kim H, Yoon JB, Choi KY. 2012. Ras stabilization through aberrant activation of Wnt/ β -catenin signaling promotes intestinal tumorigenesis. *Science Signaling* **5**:ra30. DOI: <https://doi.org/10.1126/scisignal.2002242>, PMID: 22494971
- Kadmas JL**, Beckerle MC. 2004. The LIM domain: from the cytoskeleton to the nucleus. *Nature Reviews. Molecular Cell Biology* **5**:920–931. DOI: <https://doi.org/10.1038/nrm1499>, PMID: 15520811
- Kao KR**, Masui Y, Elinson RP. 1986. Lithium-induced respecification of pattern in *Xenopus laevis* embryos. *Nature* **322**:371–373. DOI: <https://doi.org/10.1038/322371a0>, PMID: 19140288
- Kinzler KW**, Vogelstein B. 1996. Lessons from hereditary colorectal cancer. *Cell* **87**:159–170. DOI: [https://doi.org/10.1016/s0092-8674\(00\)81333-1](https://doi.org/10.1016/s0092-8674(00)81333-1), PMID: 8861899
- Kirkegaard T**, Jäättelä M. 2009. Lysosomal involvement in cell death and cancer. *Biochimica et Biophysica Acta* **1793**:746–754. DOI: <https://doi.org/10.1016/j.bbamcr.2008.09.008>, PMID: 18948147
- Koivusalo M**, Welch C, Hayashi H, Scott CC, Kim M, Alexander T, Touret N, Hahn KM, Grinstein S. 2010. Amiloride inhibits macropinocytosis by lowering submembranous pH and preventing RAC1 and Cdc42 signaling. *The Journal of Cell Biology* **188**:547–563. DOI: <https://doi.org/10.1083/jcb.200908086>, PMID: 20156964
- Kolwijck E**, Massuger LFAG, Thomas CMG, Span PN, Krasovec M, Kos J, Sweep FCGJ. 2010. Cathepsins B, L and cystatin C in cyst fluid of ovarian tumors. *Journal of Cancer Research and Clinical Oncology* **136**:771–778. DOI: <https://doi.org/10.1007/s00432-009-0716-z>, PMID: 19915865
- Kroemer G**, Jäättelä M. 2005. Lysosomes and autophagy in cell death control. *Nature Reviews. Cancer* **5**:886–897. DOI: <https://doi.org/10.1038/nrc1738>, PMID: 16239905
- Lambies G**, Commisso C. 2022. Macropinocytosis and cancer: From tumor stress to signaling pathways. *Sub-Cellular Biochemistry* **98**:15–40. DOI: https://doi.org/10.1007/978-3-030-94004-1_2, PMID: 35378701
- Lawrence RE**, Zoncu R. 2019. The lysosome as a cellular centre for signalling, metabolism and quality control. *Nature Cell Biology* **21**:133–142. DOI: <https://doi.org/10.1038/s41556-018-0244-7>, PMID: 30602725
- Leibovitz A**, Stinson JC, McCombs WB, McCoy CE. 1976. Classification of human colorectal adenocarcinoma cell lines. *Cancer Research* **36**:4562–4569 PMID: 1000501.
- Loh KM**, van Amerongen R, Nusse R. 2016. Generating cellular diversity and spatial form: Wnt signaling and the evolution of multicellular animals. *Developmental Cell* **38**:643–655. DOI: <https://doi.org/10.1016/j.devcel.2016.08.011>, PMID: 27676437
- MacDonald BT**, Tamai K, He X. 2009. Wnt/beta-catenin signaling: components, mechanisms, and diseases. *Developmental Cell* **17**:9–26. DOI: <https://doi.org/10.1016/j.devcel.2009.06.016>, PMID: 19619488
- McMahon AP**, Moon RT. 1989. Ectopic expression of the proto-oncogene int-1 in *Xenopus* embryos leads to duplication of the embryonic axis. *Cell* **58**:1075–1084. DOI: [https://doi.org/10.1016/0092-8674\(89\)90506-0](https://doi.org/10.1016/0092-8674(89)90506-0), PMID: 2673541
- Niehrs C**. 2012. The complex world of WNT receptor signalling. *Nature Reviews. Molecular Cell Biology* **13**:767–779. DOI: <https://doi.org/10.1038/nrm3470>, PMID: 23151663
- Nishizuka Y**. 1984. The role of protein kinase C in cell surface signal transduction and tumour promotion. *Nature* **308**:693–698. DOI: <https://doi.org/10.1038/308693a0>, PMID: 6232463
- Nix DA**, Beckerle MC. 1997. Nuclear-cytoplasmic shuttling of the focal contact protein, zyxin: A potential mechanism for communication between sites of cell adhesion and the nucleus. *The Journal of Cell Biology* **138**:1139–1147. DOI: <https://doi.org/10.1083/jcb.138.5.1139>, PMID: 9281590
- Nusse R**, Clevers H. 2017. Wnt/ β -Catenin Signaling, Disease, and Emerging Therapeutic Modalities. *Cell* **169**:985–999. DOI: <https://doi.org/10.1016/j.cell.2017.05.016>, PMID: 28575679
- Ohno S**, Nishizuka Y. 2002. Protein kinase C isotypes and their specific functions: prologue. *Journal of Biochemistry* **132**:509–511. DOI: <https://doi.org/10.1093/oxfordjournals.jbchem.a003249>, PMID: 12359062

- Perona R**, Serrano R. 1988. Increased pH and tumorigenicity of fibroblasts expressing a yeast proton pump. *Nature* **334**:438–440. DOI: <https://doi.org/10.1038/334438a0>, PMID: 2900469
- Ramirez C**, Hauser AD, Vucic EA, Bar-Sagi D. 2019. Plasma membrane V-ATPase controls oncogenic RAS-induced macropinocytosis. *Nature* **576**:477–481. DOI: <https://doi.org/10.1038/s41586-019-1831-x>, PMID: 31827278
- Redelman-Sidi G**, Binyamin A, Gaeta I, Palm W, Thompson CB, Romesser PB, Lowe SW, Bagul M, Doench JG, Root DE, Glickman MS. 2018. The canonical wnt pathway drives macropinocytosis in cancer. *Cancer Research* **78**:4658–4670. DOI: <https://doi.org/10.1158/0008-5472.CAN-17-3199>, PMID: 29871936
- Rous P**, Kidd JG. 1941. Conditional neoplasms and subthreshold neoplastic states: A study of the tar tumors of rabbits. *The Journal of Experimental Medicine* **73**:365–390. DOI: <https://doi.org/10.1084/jem.73.3.365>, PMID: 19871084
- Schwarz M**, Münzel PA, Braeuning A. 2013. Non-melanoma skin cancer in mouse and man. *Archives of Toxicology* **87**:783–798. DOI: <https://doi.org/10.1007/s00204-012-0998-9>
- Segditsas S**, Tomlinson I. 2006. Colorectal cancer and genetic alterations in the Wnt pathway. *Oncogene* **25**:7531–7537. DOI: <https://doi.org/10.1038/sj.onc.1210059>, PMID: 17143297
- Su Z**, Song J, Wang Z, Zhou L, Xia Y, Yu S, Sun Q, Liu SS, Zhao L, Li S, Wei L, Carson DA, Lu D. 2018. Tumor promoter TPA activates Wnt/ β -catenin signaling in a casein kinase 1-dependent manner. *PNAS* **115**:E7522–E7531. DOI: <https://doi.org/10.1073/pnas.1802422115>, PMID: 30038030
- Swanson JA**. 1989. Phorbol esters stimulate macropinocytosis and solute flow through macrophages. *Journal of Cell Science* **94 (Pt 1)**:135–142. DOI: <https://doi.org/10.1242/jcs.94.1.135>, PMID: 2613767
- Taelman VF**, Dobrowolski R, Plouhinec J-L, Fuentealba LC, Vorwald PP, Gumper I, Sabatini DD, De Robertis EM. 2010. Wnt signaling requires sequestration of glycogen synthase kinase 3 inside multivesicular endosomes. *Cell* **143**:1136–1148. DOI: <https://doi.org/10.1016/j.cell.2010.11.034>, PMID: 21183076
- Takai Y**, Kishimoto A, Iwasa Y, Kawahara Y, Mori T, Nishizuka Y. 1979. Calcium-dependent activation of a multifunctional protein kinase by membrane phospholipids. *The Journal of Biological Chemistry* **254**:3692–3695. DOI: [https://doi.org/10.1016/S0021-9258\(18\)50638-4](https://doi.org/10.1016/S0021-9258(18)50638-4), PMID: 438153
- Tatarelli C**, Linnenbach A, Mimori K, Croce CM. 2000. Characterization of the human TESTIN gene localized in the FRA7G region at 7q31.2. *Genomics* **68**:1–12. DOI: <https://doi.org/10.1006/geno.2000.6272>, PMID: 10950921
- Tejeda-Muñoz N**, Albrecht LV, Bui MH, De Robertis EM. 2019. Wnt canonical pathway activates macropinocytosis and lysosomal degradation of extracellular proteins. *PNAS* **116**:10402–10411. DOI: <https://doi.org/10.1073/pnas.1903506116>, PMID: 31061124
- Tejeda-Muñoz N**, De Robertis EM. 2022a. Lysosomes are required for early dorsal signaling in the *Xenopus* embryo. *PNAS* **119**:e2201008119. DOI: <https://doi.org/10.1073/pnas.2201008119>, PMID: 35446621
- Tejeda-Muñoz N**, De Robertis EM. 2022b. Wnt, GSK3, and Macropinocytosis. *Sub-Cellular Biochemistry* **98**:169–187. DOI: https://doi.org/10.1007/978-3-030-94004-1_9, PMID: 35378708
- Tejeda-Muñoz N**, Morselli M, Moriyama Y, Sheladiya P, Pellegrini M, De Robertis EM. 2022c. Canonical Wnt signaling induces focal adhesion and Integrin beta-1 endocytosis. *iScience* **25**:104123. DOI: <https://doi.org/10.1016/j.isci.2022.104123>, PMID: 35402867
- Tobias ES**, Hurlstone AF, MacKenzie E, McFarlane R, Black DM. 2001. The TES gene at 7q31.1 is methylated in tumours and encodes a novel growth-suppressing LIM domain protein. *Oncogene* **20**:2844–2853. DOI: <https://doi.org/10.1038/sj.onc.1204433>, PMID: 11420696
- Vinyoles M**, Del Valle-Pérez B, Curto J, Viñas-Castells R, Alba-Castellón L, García de Herreros A, Duñach M. 2014. Multivesicular GSK3 Sequestration upon Wnt Signaling Is Controlled by p120-Catenin/Cadherin Interaction with LRP5/6. *Molecular Cell* **53**:444–457. DOI: <https://doi.org/10.1016/j.molcel.2013.12.010>
- Weinberg RA**, Weinberg RA. 2007. *The Biology of Cancer* Taylor & Francis Group. DOI: <https://doi.org/10.1201/9780203852569>
- Wu X**, Tu X, Joeng KS, Hilton MJ, Williams DA, Long F. 2008. Rac1 activation controls nuclear localization of beta-catenin during canonical Wnt signaling. *Cell* **133**:340–353. DOI: <https://doi.org/10.1016/j.cell.2008.01.052>, PMID: 18423204
- Yoshida S**, Pacitto R, Inoki K, Swanson J. 2018. Macropinocytosis, mTORC1 and cellular growth control. *Cellular and Molecular Life Sciences* **75**:1227–1239. DOI: <https://doi.org/10.1007/s00018-017-2710-y>, PMID: 29119228
- Zhitomirsky B**, Assaraf YG. 2016. Lysosomes as mediators of drug resistance in cancer. *Drug Resistance Updates* **24**:23–33. DOI: <https://doi.org/10.1016/j.drug.2015.11.004>, PMID: 26830313