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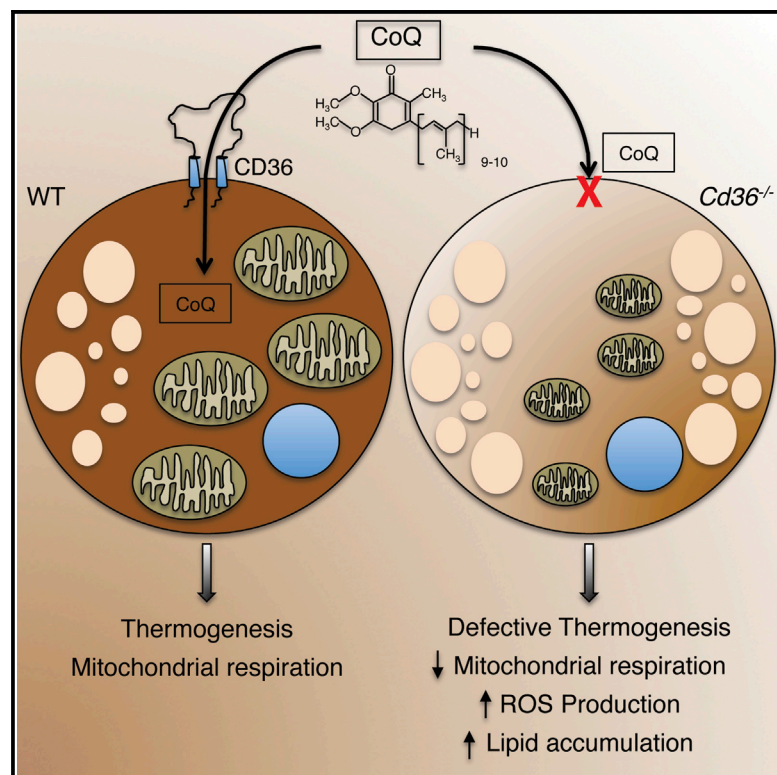
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Dependence of Brown Adipose Tissue Function on CD36-Mediated Coenzyme Q Uptake

Graphical Abstract



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In Brief

Coenzyme Q (CoQ) is an essential component of the mitochondrial electron transport chain, and while exogenous CoQ supplementation has been successful as a therapy for deficient patients, which tissues depend on exogenous CoQ and the mechanism of CoQ uptake is not well understood. Anderson et al. report that the scavenger receptor CD36 drives CoQ uptake by brown adipose tissue and is required for normal BAT function.

Highlights

- Uptake of CoQ by brown adipocytes requires CD36
- BAT lacking CD36 has a deficiency in CoQ levels
- *Cd36*^{-/-} BAT mitochondria display bioenergetic consequences due to CoQ deficiency
- CD36 is required for nonshivering thermogenesis and normal BAT morphology

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Dependence of Brown Adipose Tissue Function on CD36-Mediated Coenzyme Q Uptake

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SUMMARY

Brown adipose tissue (BAT) possesses the inherent ability to dissipate metabolic energy as heat through uncoupled mitochondrial respiration. An essential component of the mitochondrial electron transport chain is coenzyme Q (CoQ). While cells synthesize CoQ mostly endogenously, exogenous supplementation with CoQ has been successful as a therapy for patients with CoQ deficiency. However, which tissues depend on exogenous CoQ uptake as well as the mechanism by which CoQ is taken up by cells and the role of this process in BAT function are not well understood. Here, we report that the scavenger receptor CD36 drives the uptake of CoQ by BAT and is required for normal BAT function. BAT from mice lacking CD36 displays CoQ deficiency, impaired CoQ uptake, hypertrophy, altered lipid metabolism, mitochondrial dysfunction, and defective nonshivering thermogenesis. Together, these data reveal an important new role for the systemic transport of CoQ to BAT and its function in thermogenesis.

INTRODUCTION

Classical brown adipose tissue (BAT) is a unique type of adipose tissue that is composed of adipocytes with multilocular lipid droplets and a large amount of mitochondria, making it a highly metabolically active organ that is responsible for nonshivering thermogenesis both in neonate and adult humans (Aherne and Hull, 1966; Cypess et al., 2009; Heaton, 1972; van Marken Lichtenbelt et al., 2009; Virtanen et al., 2009). The unabated growth of the obesity epidemic and associated metabolic diseases such as type 2 diabetes reflects our current lack of efficient strategies for intervention and treatment of metabolic diseases (Zimmet et al., 2001). Because BAT possesses the inherent ability to dissipate metabolic energy as heat through uncoupled mitochondrial

respiration, expanding BAT or enhancing its respiratory activity could be a strategy for therapeutic intervention.

In order to generate a high metabolic rate for heat production, brown adipocytes contain a large amount of mitochondria densely packed with cristae (Sell et al., 2004). These cells possess an unparalleled ability to generate heat due to the dissociation of electron transport chain respiration from ATP production through uncoupling protein 1 (UCP1) (Argyropoulos and Harper, 2002). Located in the inner mitochondrial membrane, UCP1 provides an alternative route of entry for protons, allowing them to bypass ATP synthase and in turn dissipate energy to generate heat (Argyropoulos and Harper, 2002). Brown adipocyte mitochondria also display an elevated capacity for substrate utilization, relying predominantly on the β oxidation of long-chain fatty acids (LCFAs) (Sell et al., 2004). LCFAs can be taken up from the circulation via fatty-acid transport proteins (FATPs) or can be generated from endocytosed lipoproteins through a process mediated in part by the scavenger receptor (SR) CD36 (Anderson and Stahl, 2013; Coburn et al., 2001; Kuniyasu et al., 2002; Zeng et al., 2003).

The SR family of transmembrane glycoproteins mediates the binding and uptake of a broad range of ligands in a variety of tissues (Greaves and Gordon, 2009). The SR family is comprised of several classes, SR-A through SR-H (Plüddemann et al., 2007). SR class B receptors are unique from other classes of SRs in that they have two transmembrane domains, an extracellular ligand binding loop, and two short intracellular tails (Plüddemann et al., 2007). A defining member of the SR class B receptors is CD36, an 88 kDa SR with multiple binding pockets and hydrophobic segments (Su and Abumrad, 2009). CD36 has been shown to be required in a variety of tissues for the uptake of several hydrophobic molecules, including LCFA and the carotenoid lycopene (Harmon and Abumrad, 1993; Moussa et al., 2011). Originally identified as fatty acid translocase (FAT), CD36 is also an established receptor for compounds of different chemical nature, such as oxidized lipoproteins, thrombospondin, and collagen (Nergiz-Unal et al., 2011). CD36 is structurally most similar to SR-BI, which has been shown to mediate the selective uptake of cholesteryl esters from high-density lipoprotein

(HDL) (Acton et al., 1996; Calvo et al., 1995; Calvo et al., 1998). More recently, it was demonstrated that CD36 is required for the endocytosis of lipoproteins by both macrophages and BAT (Bartelt et al., 2011; Febbraio et al., 2000).

The lipid coenzyme Q (CoQ; also known as ubiquinone) is an essential component of the mitochondrial electron transport chain, functioning as a transporter of electrons from several electron carriers (including complex I, complex II, glycerol-3-phosphate [G3P] dehydrogenase [GPDH], and the electron transferring flavoprotein of fatty acid β oxidation) to complex III as well as an antioxidant (Crane, 2001). CoQ consists of a quinone ring and an isoprenoid side chain of varying length that is derived from the same mevalonate pathway as cholesterol (Bentinger et al., 2010). The longer the isoprenoid side chain, the more hydrophobic the CoQ molecule is (Bentinger et al., 2010). Mice have predominantly CoQ with 9 isoprenoid units (CoQ₉), whereas humans have mostly CoQ₁₀ (Sohet and Delzenne, 2012). A large portion of CoQ is found in the inner mitochondrial membrane, but a smaller portion can also be found in the membranes of other organelles and the cytosol, suggesting that it may have other functions in addition to transporting electrons in the mitochondria (Bhagavan and Chopra, 2006).

Low CoQ levels are associated with cardiomyopathies, aging, and statin-induced myopathies (Bentinger et al., 2010). There are also several genetic mutations directly affecting proteins involved in the synthesis of CoQ, resulting in primary CoQ deficiencies (Quinzii et al., 2008). Therefore, increasing CoQ levels could be therapeutically beneficial for a variety of metabolic diseases. Many tissues endogenously synthesize CoQ, and the contribution of dietary CoQ is thought to be comparatively small (Bhagavan and Chopra, 2006). However, exogenous uptake of CoQ from the circulation can be significant in situations where CoQ levels are already low, such as in genetic deficiencies, myopathies, and aging (Artuch et al., 2009). Exactly how tissues take up CoQ and which tissues primarily depend on exogenous CoQ have not been well studied (Padiilla-López et al., 2009), posing a considerable challenge to the therapeutic use of CoQ in metabolic diseases, aging, and mitochondrial function.

We speculated that, given the ability of CD36 and related receptors to mediate the uptake of hydrophobic molecules in a variety of tissues, CD36 may be required in BAT for the uptake of similar molecules, such as CoQ. Here we report that CD36 is required for CoQ uptake in BAT and the maintenance of normal CoQ levels, and thus nonshivering thermogenesis and BAT function. The ability of CD36 deficient BAT (*Cd36*^{-/-}) to take up CoQ was greatly impaired, and analysis of mitochondrial CoQ levels showed that loss of CD36 caused a BAT-specific decrease in CoQ₉ and CoQ₁₀. While BAT fatty acid uptake rates were unchanged, the ability of *Cd36*^{-/-} BAT to take up another lipid, CoQ, was greatly impaired. This CD36-dependent CoQ deficiency impaired processes linked to classical CoQ function, such as oxidative damage and reduced respiration. Together, these data demonstrate a unique role for exogenous CoQ provided by CD36 in BAT mitochondrial function and reveal an unexpected dependence of BAT homeostasis on the cellular levels of CoQ.

RESULTS

CD36 Drives CoQ Uptake by BAT

CD36 has been shown to enhance uptake of LCFAs in the heart, skeletal muscle, and white adipose tissue (WAT) (Abumrad et al., 1993; Coburn et al., 2000; Van Nieuwenhoven et al., 1995). Besides LCFAs, CD36 and related receptors have been shown to play a role in the cellular uptake of a variety of other hydrophobic molecules, including the selective uptake of cholesterol esters from HDL by SR-B1 (Acton et al., 1996) and carotenoids such as lycopenes by CD36 (Moussa et al., 2011). Since both lycopenes and CoQ are isoprenoids, we wanted to determine whether CD36 can enhance cellular uptake of CoQ, particularly in the context of a mitochondrial-rich tissue. CoQ was administered to cells and animals via Intralipid, which is an emulsion of soy bean oil, phospholipids, and glycerin (Taskinen et al., 1983). We compared the uptake of solvent- and detergent-solubilized CoQ to Intralipid and found that receptor-mediated uptake was only observed with Intralipid (data not shown). When we treated HEK293 cells overexpressing CD36 with CoQ, the CD36-expressing cells took up 57% more CoQ₉ than control cells (Figure 1A). Conversely, mature brown adipocytes from *Cd36*^{-/-} BAT took up 42% less CoQ₁₀ than WT cells, as quantified by high-performance liquid chromatography (HPLC) (Figures 1B and S1A). It is important to note that CoQ₁₀ was used as a substrate for uptake, as endogenous levels are low in murine cells and differed only slightly between WT and *Cd36*^{-/-} mature brown adipocytes (Figure 1B). However, total CoQ levels, which are dominated by CoQ₉, were significantly suppressed in *Cd36*^{-/-} mature brown adipocytes (WT 1.55 ± 0.18 pmol total CoQ/μg protein versus *Cd36*^{-/-} 0.96 ± 0.11 pmol total CoQ/μg protein, *p = 0.015). Taken together, these results suggest a role for CD36 in CoQ uptake, particularly by BAT.

To determine whether CD36 enhances CoQ uptake in vivo, we administered a CoQ-containing Intralipid mix to WT and *Cd36*^{-/-} mice by intraperitoneal injection and then measured CoQ levels by HPLC. At 24-hours postinjection, *Cd36*^{-/-} BAT had taken up significantly less CoQ₁₀ than WT mice (WT 0.159 ± 0.034 pmol CoQ₁₀/μg protein versus *Cd36*^{-/-} 0.018 ± 0.006 pmol CoQ₁₀/μg protein) (Figure 1Ci). Conversely, uptake of exogenous CoQ₁₀ was unaffected in the liver, which took up significant amounts of injected CoQ₁₀, and skeletal muscle, which did not take up appreciable amounts of exogenous CoQ₁₀ (Figures 1Cii and 1Ciii). Interestingly, while there was no difference in serum CoQ₁₀ levels between WT and *Cd36*^{-/-} mice at baseline, *Cd36*^{-/-} mice had significantly more CoQ₁₀ in their serum following injection of CoQ₁₀ (WT 0.009 ± 0.002 pmol CoQ₁₀/μl serum versus *Cd36*^{-/-} 0.026 ± 0.005 pmol CoQ₁₀/μl serum) (Figure 1Civ).

To determine whether CoQ levels are perturbed in other tissues from *Cd36*^{-/-} mice at baseline, we measured CoQ₉ and CoQ₁₀ in BAT, liver, heart, soleus, gastrocnemius, quadriceps, adrenal gland, brain, and serum. While CoQ₁₀ is the predominant species in humans, the mouse CoQ pool is composed of approximately 90% CoQ₉ and 10% CoQ₁₀ (Lass et al., 1997). Interestingly, both CoQ₉ and CoQ₁₀ levels in *Cd36*^{-/-} BAT were significantly decreased by approximately 40%–50% (Figure 1D). This finding was confirmed by mass spectrometry, which

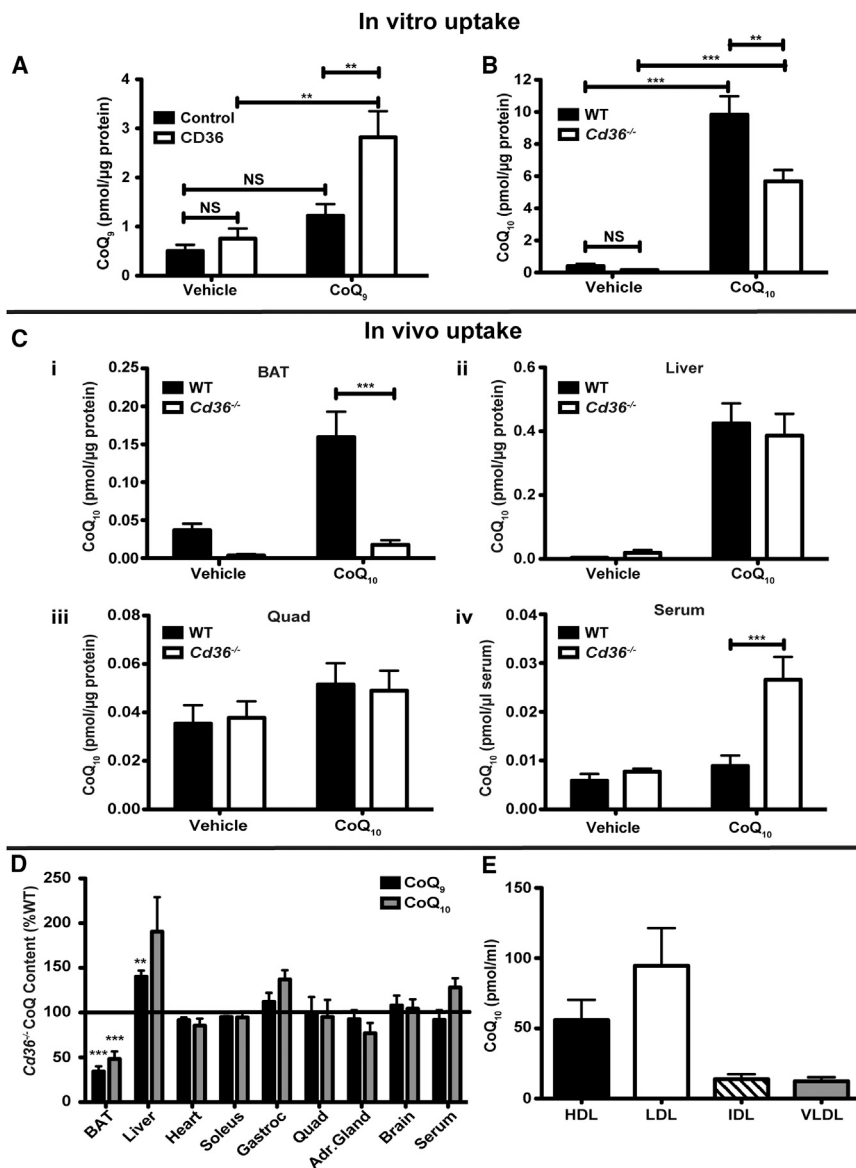


Figure 1. CD36 Is Required for CoQ Transport

(A) HEK293 cells stably expressing CD36 or parent vector (control) were treated with CoQ₉ for 24 hr (n = 5–6 per group). CoQ levels were measured by HPLC and normalized to micrograms of protein.

(B) Primary mature brown adipocytes were isolated from WT and *Cd36*^{-/-} BAT and treated with either vehicle or CoQ₁₀ for 3 hr (n = 5).

(C) (i–iv) WT and *Cd36*^{-/-} mice were injected intraperitoneally with either vehicle (Intralipid) or CoQ₁₀. After 24 hr, tissues were isolated, and CoQ₁₀ levels were measured by HPLC and normalized to micrograms of protein or microliters of serum (n = 8–9).

(D) BAT, liver, heart, soleus, gastrocnemius, quadriceps, adrenal gland, brain, and serum CoQ₉ and CoQ₁₀ levels in *Cd36*^{-/-} relative to WT (100%) were measured by HPLC and normalized to micrograms of protein (n = 3–6). A complete table with the absolute CoQ values is included in Table S1.

(E) CoQ₁₀ levels were measured in lipoprotein fractions from human serum by HPLC and normalized to milliliters of fraction (n = 6).

*p < 0.05; **p < 0.005; ***p < 0.0005. Error bars represent SEM. See also Figure S1 and Table S1.

Alterations of Lipid Metabolism in *Cd36*^{-/-} BAT

CD36 is most abundant in adipose tissue, heart, lung, and muscle, with less expression in spleen and liver (Figures 2A, S2A, and S2B). In addition, there is no effect of browning in WAT by CL-316,243 on the expression of *Cd36* in subcutaneous and visceral WAT depots (Figure S2C). To elucidate the role of CD36 in BAT, we took advantage of mice lacking CD36 (*Cd36*^{-/-}) (Febbraio et al., 1999). The absence of CD36 in the BAT of these mice was confirmed by Western blot (Figures 2A and S2A). To identify specifically where CD36 is expressed in BAT, we

showed significantly decreased levels of both CoQ₉ and CoQ₁₀ in *Cd36*^{-/-} BAT (64% and 80% of WT, respectively) (Figures S1B and S1C). In contrast, CoQ levels in the heart, skeletal muscle, adrenal glands, brain, and serum from *Cd36*^{-/-} mice were normal, while levels in the liver were slightly increased (Figure 1D; Table S1).

These loss-of-function and gain-of-function studies in vitro and in vivo suggest that CD36 facilitates direct cellular uptake of CoQ in BAT. Given that CD36 can facilitate uptake of lipoprotein particles in BAT (Bartelt et al., 2011), we wanted to see in which lipoprotein fraction CoQ is present. CoQ is most abundant in the low-density lipoprotein (LDL) fraction (Figure 1E), but absent from the nonlipoprotein fraction (data not shown), suggesting that it may be transported through the circulation and presented to the cell as a lipoprotein particle component (Tomasetti et al., 1999).

performed an immunolocalization study for CD36 and UCP1. Interestingly, CD36 was found to be located exclusively on the plasma membrane in BAT, supporting a role for CD36 in the uptake of hydrophobic molecules from the circulation (Figure 2B).

Given that CD36 is robustly expressed in BAT and that *Cd36*^{-/-} mice have decreased CoQ levels in BAT, we wanted to determine whether mice lacking CD36 had normal BAT morphology and function. An appreciable difference in the morphology of *Cd36*^{-/-} BAT compared with WT BAT was immediately noticed by direct observation (Figure 2C). BAT lacking CD36 was significantly larger and paler in color compared with BAT from WT mice at room temperature (Figure 2C). Following cold exposure, the characteristic browning of BAT was absent in *Cd36*^{-/-} mice, while the weight difference remained (Figure 2C). 3D reconstruction of confocal microscopy scans of BAT sections stained for neutral lipids with BODIPY confirmed that the marked

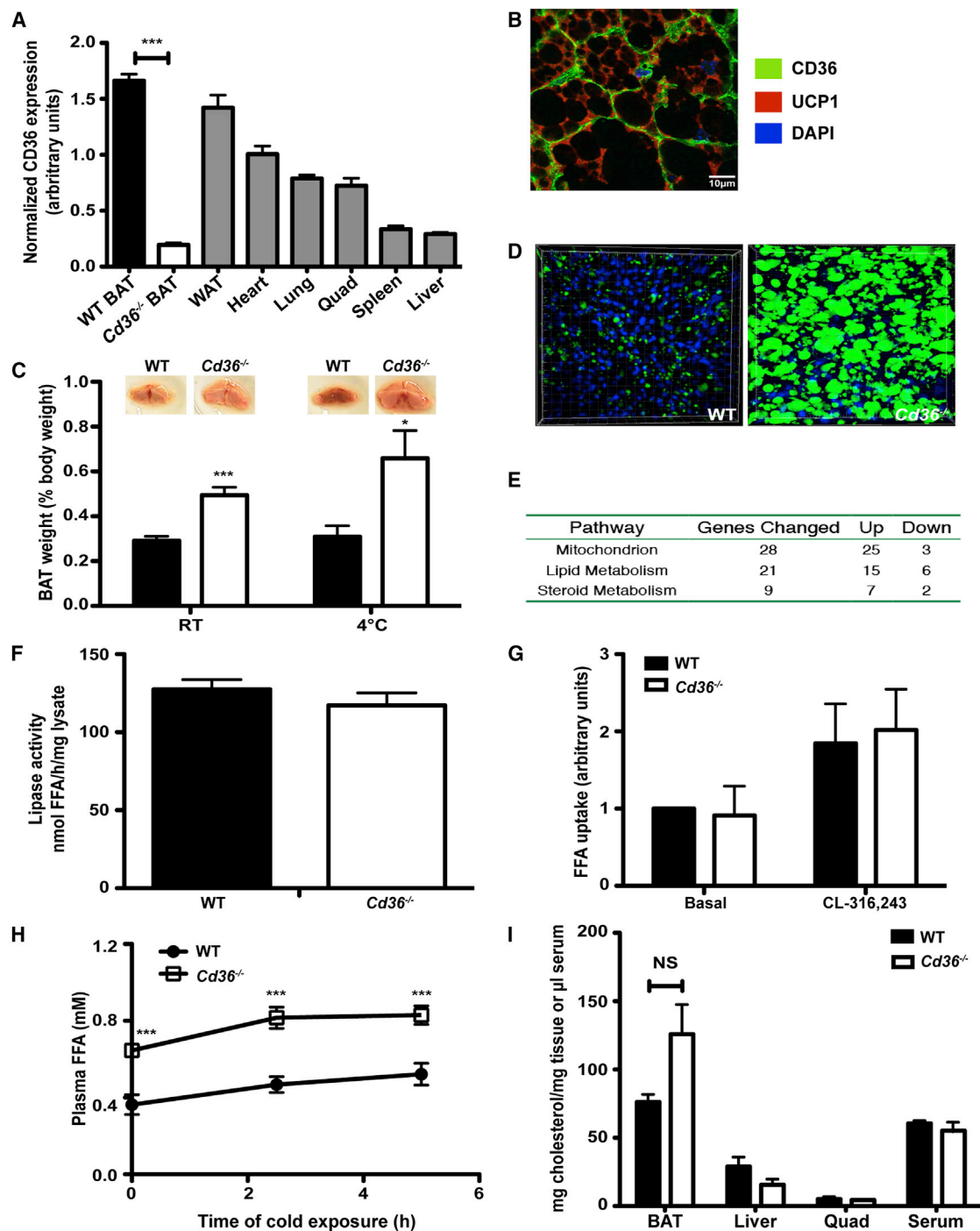


Figure 2. Pathologic TAG Storage in *Cd36*^{-/-} BAT

(A) Western blot for CD36 and β tubulin was performed on lysates from WT and *Cd36*^{-/-} BAT, WAT, heart, lung, quadriceps muscle, spleen, and liver (n = 5). Data are presented in arbitrary units as a ratio of CD36 normalized to tubulin.

(B) Colocalization of CD36 (green), UCP1 (red), and DAPI (blue) in a WT BAT cryosection. Scale bar represents 10 μ m.

(C) Images of BAT from WT and *Cd36*^{-/-} mice and their weights before and after cold exposure (n = 3–6).

(D) Representative images of 3D reconstructions from cold-exposed mouse BAT sections stained with the fluorescently labeled TAG probe BODIPY 493/503 (green) and DAPI (blue).

(E) Genome-wide expression analysis was performed on RNA from WT and *Cd36*^{-/-} BAT (n = 3 per group). A summary table of misregulated genes identified using a $p \leq 0.05$ threshold is presented. A complete list of misregulated genes is included in [Tables S2](#) and [S3](#).

(F) Lipase activity (expressed as nmol of FFA released/hour/mg protein of tissue lysate) from WT and *Cd36*^{-/-} BAT chunks (n = 4).

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hypertrophy in *Cd36*^{-/-} BAT was mainly due to increased lipid droplet accumulation (Figure 2D) and lipid droplet volume (median volume: 18.5 μm^3 WT versus 205 μm^3 *Cd36*^{-/-}). These results show that BAT lacking CD36 displays whitening and increased lipid droplet size.

To determine whether the accumulation of lipids in *Cd36*^{-/-} BAT was due to defective lipolysis, we first measured lipase activity in mature brown adipocytes from WT and *Cd36*^{-/-} mice. No significant difference in lipase activity was observed between WT and *Cd36*^{-/-} BAT (Figure 2F). The lipolysis rate in WAT was unchanged (Figure S3A), indicating that the release of fatty acids from WAT into the circulation was unaffected in *Cd36*^{-/-} mice. However, the lipolysis rate in both the basal state and after stimulation by forskolin/IBMX was significantly reduced in *Cd36*^{-/-} BAT (Figure S3B).

We speculated that the defects observed in *Cd36*^{-/-} BAT morphology could arise from alterations in lipid trafficking. We found that LCFA uptake rates of isolated primary mature brown adipocytes from *Cd36*^{-/-} mice were comparable to WT and responded normally to stimulation by the β 3-adrenergic receptor agonist CL-316,243 (WT 1.84- \pm 0.51-fold change over untreated versus *Cd36*^{-/-} 2.02- \pm 0.53-fold change over untreated) (Figure 2G), while *Lpl* expression in *Cd36*^{-/-} BAT was elevated by 1.48-fold compared with WT BAT (* p = 0.007) (Table S3), arguing against defective free fatty acid (FFA) uptake or production. To determine whether there was a defect in circulating FFA, we measured FFA levels in the serum of WT and *Cd36*^{-/-} mice. Consistent with previous reports (Coburn et al., 2000; Febbraio et al., 1999), circulating FFA levels were actually elevated 2-fold in *Cd36*^{-/-} mice both before and after cold exposure (Figure 2H). Cholesterol levels were unchanged between WT and *Cd36*^{-/-} mice in BAT, liver, quad, and serum (Figure 2I).

Impaired Mitochondrial Substrate Utilization in *Cd36*^{-/-} BAT

To determine whether there are bioenergetic consequences as a result of BAT CoQ deficiency, we assessed LCFA β oxidation in intact mature brown adipocytes using [¹⁴C]-palmitic acid. Despite normal fatty acid uptake potential (Figure 2G), production of ¹⁴CO₂ by *Cd36*^{-/-} brown adipocytes was reduced by approximately 50% compared with WT brown adipocytes, demonstrating that substrate utilization is decreased in *Cd36*^{-/-} BAT and suggesting a potential mitochondrial defect (Figure 3A).

A series of Clark electrode respiratory tests with isolated BAT mitochondria using a variety of substrates was performed. In isolated mitochondria, electrons from pyruvate and malate enter complex I, while electrons from succinate and G3P bypass complex I and are transferred directly to CoQ by complex II or GPDH, respectively (Crane, 2001). The basal rates of respiration were decreased in BAT mitochondria from *Cd36*^{-/-} mice (although not statistically significant for succinate and pyruvate/malate) (Figure 3B). Furthermore, the defect in respiration with G3P

could be rescued with the addition of CoQ₂, suggesting that the cause of impaired respiration is due to the decreased levels of CoQ (Figure 3B). Electron microscopy revealed significantly smaller mitochondria in *Cd36*^{-/-} BAT compared with WT (Figure 3C). Mitochondria in *Cd36*^{-/-} BAT were 53% smaller in area compared with WT (Figure 3D). Furthermore, the mitochondrial volume density in *Cd36*^{-/-} BAT was 51% that of WT (Figure 3E). Taken together, these results demonstrate impaired mitochondrial function in *Cd36*^{-/-} BAT due to decreased CoQ levels.

Genome-wide expression analysis in BAT from WT and *Cd36*^{-/-} mice showed changes in the expression of several functional families, including mitochondrial related genes, but no misregulation of genes encoding electron transport chain proteins (Figure 2E; Tables S2 and S3). To determine whether cytochrome levels within the mitochondrial electron transport chain were affected, we measured the content of cytochromes *c*, *c1*, *b*, and *a*, but saw no differences in their levels in *Cd36*^{-/-} BAT mitochondria (Figure 3F).

Low CoQ levels are associated with increased reactive oxygen species (ROS) production and oxidative damage (Murphy, 2009). To determine whether *Cd36*^{-/-} BAT mitochondria are undergoing oxidative stress, we measured H₂O₂ production. *Cd36*^{-/-} BAT mitochondria had a significantly higher rate of H₂O₂ production (Figure 3G), indicating that there is more ROS production and possibly oxidative damage in the mitochondria of *Cd36*^{-/-} BAT.

Cd36^{-/-} Mice Are Cold Intolerant in spite of Increased Shivering

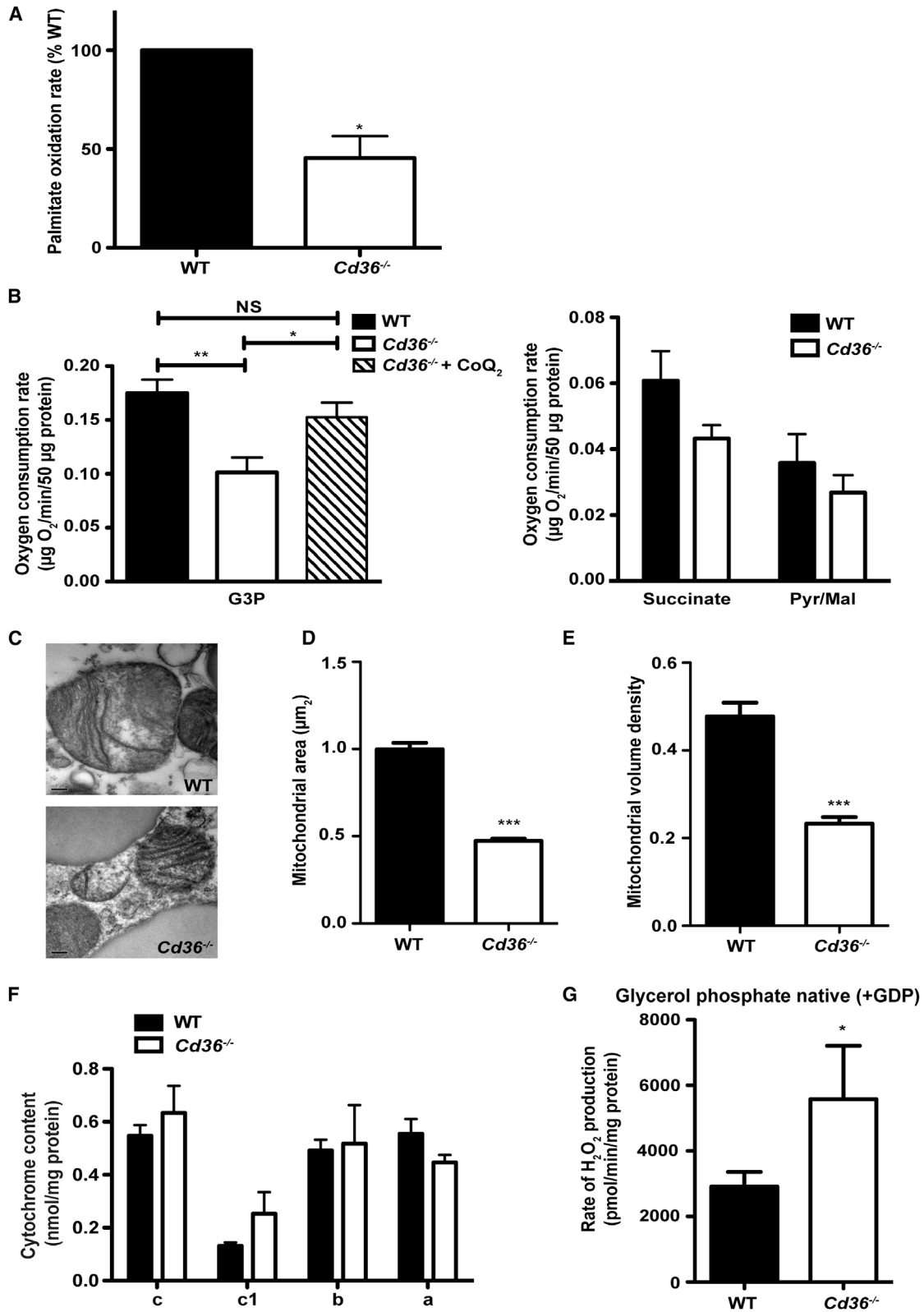
To determine whether the observed altered lipid metabolism and decrease in CoQ levels affected BAT function, we exposed WT and *Cd36*^{-/-} mice to 4°C for 5 hours and then measured their core body temperature. While there was no difference in body temperature under normal conditions, we did detect a prominent thermogenic defect in *Cd36*^{-/-} mice compared with WT mice housed at 4°C (Figure 4A). Since CD36 is also expressed in muscle, we wanted to exclude a contribution of defects from shivering thermogenesis (Bastie et al., 2004; Van Nieuwenhoven et al., 1995). Therefore, we assessed shivering thermogenesis utilizing an innovative wireless accelerometer (Figure 4B) that can be used to detect both animal movement as well as shivering (Figure 4C). We found that neither overall activity (Figure 4D) nor shivering amplitude (Figure 4F) was impaired, yet shivering duration was increased (Figure 4E). These results indicate that *Cd36*^{-/-} mice display a defect in nonshivering thermogenesis and a compensatory but insufficient upregulation of shivering thermogenesis in response to cold exposure. This hypothesis was further supported by the finding that, while resting metabolic rates were comparable between WT and *Cd36*^{-/-} mice, the characteristic increase in VO₂, VCO₂, and heat production in response to a single injection of the β 3-adrenergic receptor

(G) Maximal BODIPY 510/512-labeled FFA uptake capacity of untreated and CL-316,243-treated primary mature brown adipocytes from WT and *Cd36*^{-/-} mice (n = 3).

(H) WT and *Cd36*^{-/-} fasting plasma FFA levels at 0, 2.5, and 5 hr of cold exposure (n = 4).

(I) Cholesterol was extracted from BAT, liver, quadriceps, and serum and normalized to either milligrams of tissue or microliters of serum (n = 3).

* p < 0.05; ** p < 0.005; *** p < 0.0005. Error bars, SEM. See also Figures S2 and S3 and Tables S2 and S3.



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agonist CL-316,243 was absent in *Cd36*^{-/-} mice (Figure 4G). This suggests that altered lipid metabolism and CoQ deficiency in *Cd36*^{-/-} BAT has functional consequences.

DISCUSSION

The unique ability of mitochondria-rich BAT to generate heat and regulate nonshivering thermogenesis, coupled with its recent identification in adult humans, has driven research into the therapeutic use of this tissue for metabolic diseases. The presence of CD36 in BAT and its known role as a transporter of lipids in other tissues prompted us to examine whether CD36 could function as a transporter of the highly hydrophobic mitochondrial electron carrier CoQ in BAT. Here we show that CD36 drives the uptake of CoQ by BAT and is required for normal BAT function. The findings presented in this study demonstrate a unique relationship between CD36 and CoQ in brown fat and identify CD36 and CoQ as new factors that may limit the metabolic activity of BAT.

Efficient import of CoQ specifically into BAT by CD36 is an interesting finding. A number of studies have indicated the importance of CD36 and other class B SRs in the uptake of hydrophobic isoprenoids, such as lycopene, lutein, and β carotene (Borel et al., 2011, 2013; Harmon and Abumrad, 1993; Kiefer et al., 2002; Moussa et al., 2011), whose transport in aqueous environments is reliant on specific protein carriers such as CD36 (Kuzuyama and Seto, 2012; Reboul and Borel, 2011). In addition, genetic variants within *CD36* are associated with varying blood levels of carotenoids (Borel et al., 2011, 2013). Carotenoids are synthesized from isoprenoids, which are highly hydrophobic precursors whose transport in aqueous environments is reliant on specific protein carriers such as CD36 (Kuzuyama and Seto, 2012; Reboul and Borel, 2011). CoQ is another molecule with a hydrophobic isoprenoid side chain whose cellular uptake may be facilitated by transport proteins (Bentinger et al., 2010; Padilla-López et al., 2009). Our finding that *Cd36*^{-/-} BAT had a smaller CoQ pool than WT BAT suggested that CD36 may be required for efficient CoQ uptake or catabolism by BAT.

Loss-of-function and gain-of-function studies in vitro and in vivo demonstrate that, indeed, CD36 facilitates direct cellular uptake of CoQ, and show a clear BAT-autonomous CoQ uptake defect. While the precise mechanism of CoQ uptake by CD36 in BAT remains unclear at this point, uptake of lipoprotein particles by CD36, long believed to be restricted to liver and macrophages, was elegantly shown recently to also occur in BAT (Bartelt et al., 2011). In other tissues, CD36 has been shown to take up cholesteryl esters from HDL, as well as oxidized LDL, through

endocytosis (Acton et al., 1996; Calvo et al., 1998). Furthermore, CoQ is most abundant in the LDL fraction (Figure 1E), suggesting that it may be transported through the circulation and presented to the cell as a lipoprotein particle component (Tomasetti et al., 1999). Therefore, it is possible that CD36 mediates the uptake of lipoproteins containing CoQ through endocytosis. Alternatively, CD36 alone or in conjunction with other proteins may mediate the selective uptake of CoQ from bound lipoproteins.

It is important to note that CoQ deficiency and impaired uptake were found only in BAT lacking CD36 and not in several other tissues tested. In both WT and *Cd36*^{-/-} mice the liver, which has very little CD36, was still able to take up CoQ, while the quadriceps muscle, which has moderate levels of CD36, did not take up exogenous CoQ (Figures 1Cii and 1Ciii). This pattern of tissue CoQ uptake is very similar to the pattern of CD36-dependent lipoprotein uptake by *Cd36*^{-/-} tissues that, like our CoQ uptake, falls into three categories: CD36-dependent uptake (i.e., BAT), CD36-independent uptake (i.e., liver), and low overall uptake, as exemplified by skeletal muscle (Bartelt et al., 2011). These findings demonstrate a tissue-specific role for CD36 not only in lipoprotein uptake but also CoQ uptake.

Overall, BAT is a major sink for exogenous CoQ. In spite of its size, BAT had the highest CoQ uptake rate of all the tissues examined, removing five times as much circulating CoQ as the 16 times larger liver. Moreover, the quadriceps muscle takes up 250 times less CoQ than BAT. Remarkably, *Cd36*^{-/-} mice had three times more CoQ₁₀ in their serum compared with WT mice, yet they had eight times less CoQ₁₀ in their BAT, suggesting that CoQ accumulated in the circulation instead of being taken up by BAT in *Cd36*^{-/-} mice. These data suggest that, of the tissues examined, BAT is a major destination for circulating CoQ and that CD36 plays a role in the clearance of CoQ from the serum.

While we cannot exclude a contribution of CoQ de novo synthesis to the CD36-dependent CoQ deficiency, it is most likely that CoQ synthesis is weakly, if at all, affected. Gene expression analysis by microarray identified a multitude of genes involved in pathways leading to CoQ synthesis that were upregulated rather than downregulated in *Cd36*^{-/-} BAT (Tables S2 and S3). This could be interpreted as a possible compensatory response to the decrease in the pools of CoQ in *Cd36*^{-/-} BAT. For example, HMG-CoA synthase, which catalyzes an early step in the mevalonate synthesis pathway, was upregulated (Tables S2 and S3). This pathway generates both cholesterol and CoQ, and reduction of HMG-CoA synthase, a common precursor in the synthesis of these two molecules, suggested deficiencies in the levels of mevalonate or its derivatives cholesterol and CoQ (Bentinger et al., 2010). However, the level of cholesterol was unchanged between

Figure 3. Impaired Substrate Utilization and Mitochondrial Dysfunction in *Cd36*^{-/-} BAT

- (A) Production of ¹⁴CO₂ from ¹⁴C-[Palmitic acid] in *Cd36*^{-/-} primary mature brown adipocytes relative to WT (100%) and normalized for mg of protein (n = 3).
 (B) WT and *Cd36*^{-/-} BAT mitochondrial respiration rates using G3P with and without CoQ2, succinate, and pyruvate/malate (Pyr/Mal). Oxygen consumption rates (OCRs) were measured and presented as $\mu\text{g O}_2/\text{min}/50 \mu\text{g protein}$ (n = 6–9).
 (C) Representative electron microscopy images from WT and *Cd36*^{-/-} BAT; $\times 43,000$ magnification. Scale bar represents 0.2 μm .
 (D) Quantification of the mitochondrial area from WT and *Cd36*^{-/-} BAT EM images (n = 4).
 (E) Quantification of the mitochondrial volume density from WT and *Cd36*^{-/-} BAT EM images (n = 4).
 (F) Cytochrome content was measured in isolated mitochondria from WT and *Cd36*^{-/-} BAT and normalized to mg of mitochondrial protein (n = 3).
 (G) The rate of H₂O₂ production was measured in WT and *Cd36*^{-/-} BAT mitochondria and presented as pmol/min/mg protein (n = 4).

*p < 0.05; **p < 0.005; ***p < 0.0005. Error bars represent SEM.

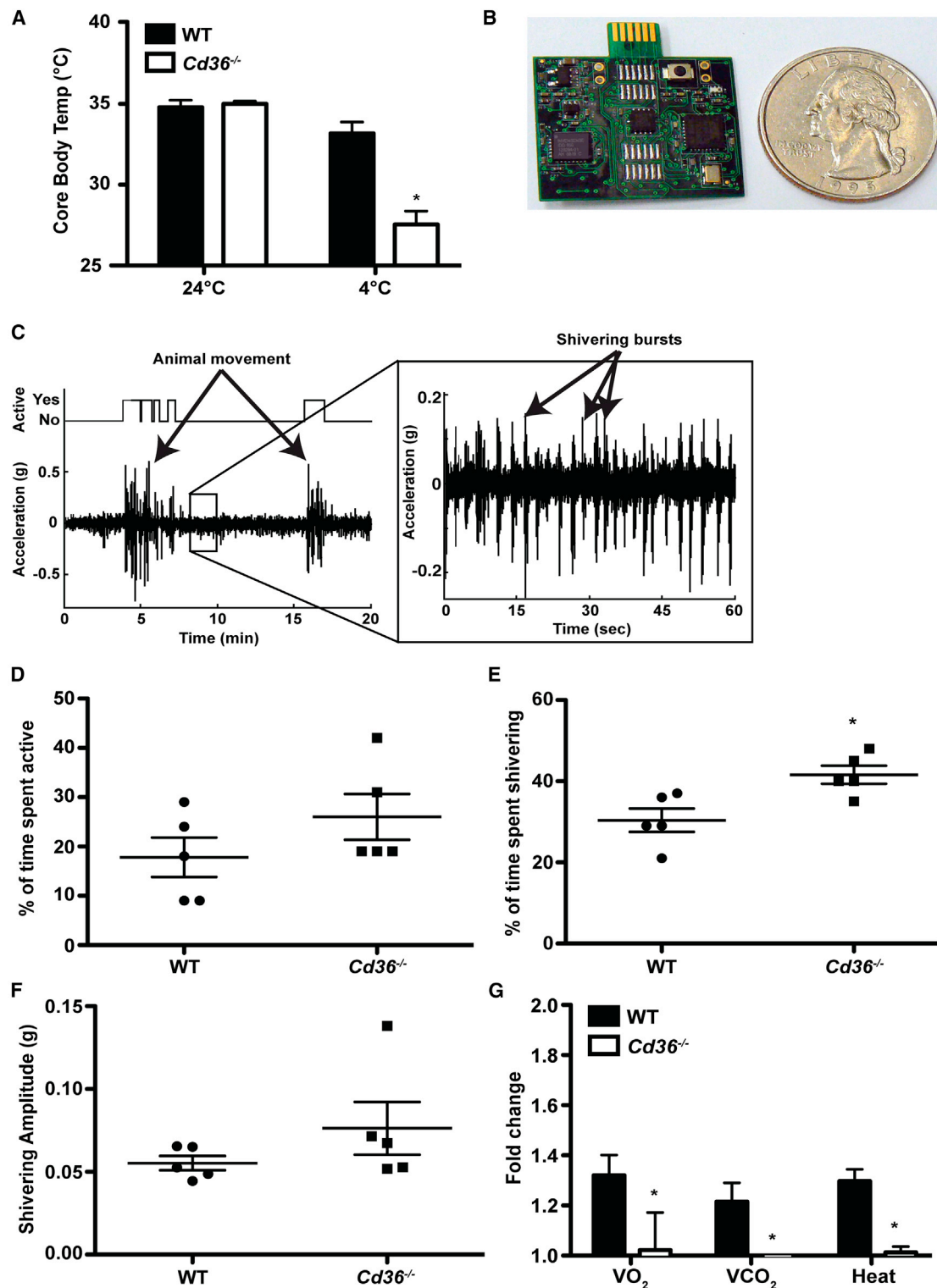


Figure 4. Defective Nonshivering Thermogenesis in *Cd36*^{-/-} BAT

(A) Mouse core body temperature measured rectally before and after 5 hr of cold exposure (n = 6).

(B) The wireless accelerometer used in this study was built at UC Berkeley and is approximately the size of a coin (a U.S. quarter). Shivering and movement were monitored by attaching the device shown to the mouse back.

(C) Accelerograph showing millisecond-resolution monitoring of movement and shivering in a WT mouse. The large bursts of acceleration indicate animal movement. A zoomed-in accelerograph displays regular shivering spikes in a cold-exposed WT mouse.

(legend continued on next page)

WT and *Cd36*^{-/-} BAT (Figure 2I), indicating that the early components of the CoQ synthesis pathway were not affected in *Cd36*^{-/-} BAT. The unchanged cholesterol levels could also hint at a facilitative rather than endocytic uptake mechanism, although further studies are required to address the point. Furthermore, while all tissues synthesize CoQ endogenously at varying rates (Thelin et al., 1992), the biosynthesis of CoQ in BAT is very low, yet its levels are quite high, suggesting that CoQ must be taken up from the circulation (Sekhar et al., 1990). The low CoQ synthesis rate in BAT coupled with our findings that cholesterol levels are unaffected in *Cd36*^{-/-} BAT suggests that loss of CD36 most likely does not affect CoQ synthesis.

Mice lacking CD36 also displayed defective nonshivering thermogenesis. Unfortunately, we were unable to test whether exogenous CoQ would rescue the thermogenic defect in *Cd36*^{-/-} mice, as CoQ₁₀ supplementation by a single intraperitoneal injection failed to raise total CoQ levels in *Cd36*^{-/-} BAT to that of baseline WT levels (Figure S4). This is most likely due to the fact that CD36 is required for the uptake of sufficient CoQ by BAT.

The data presented in this study identify a mechanism for the regulation of BAT function and thermogenesis via CD36-mediated CoQ uptake. We have shown a clear requirement for CD36 in BAT function and CoQ homeostasis. Identifying the precise mechanism of CoQ uptake by CD36 will aid in the enhancement of CoQ supplementation therapies for patients with CoQ deficiencies as well as a way to stimulate brown fat activity for the treatment of obesity and associated metabolic disorders.

EXPERIMENTAL PROCEDURES

Animal Experiments

All animal procedures were approved by the University of California Berkeley Animal Care and Use Committee. *Cd36*^{-/-} mice on a C57BL/6J background were kindly provided by Dr. Maria Febbraio (Febbraio et al., 1999). Control C57BL/6J mice (referred to as WT in this paper) were purchased from the Jackson Laboratory. All animal experiments were conducted in 8- to 20-week-old male mice housed under standard conditions. They were given free access to water and rodent chow (Harlan Teklad #2018). For cold-exposure studies, singly caged mice were housed at 4°C for 5 hr. Core body temperature was measured rectally using a lubricated thermistor probe (Yellow Springs Instruments) at 30 min intervals. Shivering was determined by attaching a wireless accelerometer (weight = 1.5 g) to the mouse back and quantified with millisecond resolution in 3D for 2 hr. For serum analysis, retro-orbital blood was collected from anesthetized mice prior to and following 2.5 and 5 hr cold exposure. Serum samples were analyzed for nonesterified fatty acids and glucose using colorimetric kits according to the manufacturer's protocol (Wako Chemicals).

CoQ Uptake Assays

For cellular uptake of CoQ, cells were incubated with 10 μM CoQ₉ or CoQ₁₀. A 1 mM CoQ stock solution was generated by dissolving CoQ in 100% ethanol and heating for 5 min to 65°C. Ten μl of 1 mM CoQ was then added to 90 μl Intralipid (Sigma #I141) and heated to 65°C for 5 min. This solution was then mixed with 1 ml prewarmed complete media and immediately added to cells.

Cells were incubated at 37°C with 10 μM CoQ for up to 6 hr, at which time the media were removed and cells were washed three times with PBS and harvested for CoQ extraction. For in vivo uptake of CoQ, mice were injected intraperitoneally with 200 μl of either a commercial CoQ₁₀ solution (LiQSORB, Tishcon) or a CoQ₁₀/Intralipid solution. To make this solution, a 29 mM CoQ₁₀ stock solution was generated by dissolving CoQ₁₀ (Sigma #C9538) in 100% ethanol and heating for 15 min to 65°C. Twenty μl of 29 mM CoQ₁₀ was then added to 180 μl Intralipid (Sigma #I141) and heated to 65°C for 15 min. Two hundred μl of this 1.45 mM CoQ₁₀/Intralipid were injected intraperitoneally per mouse. Serum and tissues were collected 24 hr following injection for CoQ extraction.

CoQ Extraction and Measurement by HPLC

CoQ levels were determined as previously described (Podda et al., 1999). In brief, whole tissue, isolated mitochondria, or cells were homogenized in 1 ml PBS containing 0.5 mg/ml of the antioxidant butylated hydroxytoluene. One ml of 0.1 M SDS was added to samples, followed by vortexing and sonication. An aliquot (50 μl) was removed for determining protein concentration by bicinchoninic acid (BCA) assay, and CoQ4 was then added as an internal standard. Two ml of 100% ethanol were added, followed by vortexing and sonication. Two ml of hexane were then added, and the samples were vortexed for 90 s and centrifuged at 1,000 g for 3 min. The upper hexane layer was dried under gentle nitrogen stream in a 37°C water bath. The dried remnants were then resuspended in 160 μl of 100% ethanol and briefly heated at 65°C. Quantification by HPLC was determined using the Waters HPLC-UV detection system. One hundred μl of the resuspension were injected and run on a C18-ODS Hypersil reverse-phase column (Thermo Scientific #30105-254630). A gradient was used consisting of solution A (80% methanol/20% water) and solution B (100% ethanol) with a flow rate of 1 ml/min for 40 min: min 0–16, 39% A and 61% B; over min 16–18 change to 100% B; min 18–28 100% B; over min 28–30 change to 39% A and 61% B; min 30–40, 39% A and 61% B. CoQ₉ peak appears around 24 min, and CoQ₁₀ peak appears around 25 min. A standard curve was generated using commercial CoQ₉ and CoQ₁₀ (Sigma) to determine absolute amounts.

Lipolysis and Lipase Activity

Lipase activity was performed as previously described (Ahmadian et al., 2009). For lipolysis rates, 5 mg of BAT tissue chunks was incubated in serum-free Dulbecco's modified Eagle's medium with and without 200 μM 3-isobutyl-1-methylxanthine (IBMX) and 10 μM forskolin (Sigma). Lipolysis rate was determined at 15 min intervals as the amount of glycerol released using the free glycerol reagent from Sigma (#F6428) according to the manufacturer's recommendations.

LCFA Uptake

Fatty acid uptake assays were performed as previously described (Wu et al., 2006) using primary mature brown adipocytes at 37°C in 1 × Hank's balanced salt solution containing 0.1% fatty acid-free bovine serum albumin (BSA) and BODIPY 500/510 C1, C12-labeled fatty acid (Invitrogen #D3823). The reaction was stopped at 0.5, 1, and 5 min by the addition of ice-cold PBS containing 0.1% BSA and 500 μM Phloretin (Sigma). Single-cell fluorescence was determined at 510 nm by fluorescence-activated cell sorting using the FACScalibur instrument (BD Biosciences).

Palmitic Acid Oxidation

Palmitic acid oxidation assays were performed as previously described (Ahmadian et al., 2009). Primary mature brown adipocytes were incubated at 37°C for 1 hr in sealed vials containing modified Krebs buffer plus 1% BSA, 5 mM glucose, and 5 μM [¹⁴C]-Palmitate (50 mCi/mmol, PerkinElmer). The reaction was stopped with 5N H₂SO₄, and the ¹⁴CO₂ produced was trapped in filter papers soaked in 200 μl 2-phenethylamine (Sigma). The filter papers

(D–F) Motor activity (D), time spent shivering (E), and shivering amplitude (F) of fasted mice exposed to 4°C for 2 hr (n = 5).

(G) Oxygen consumption and carbon dioxide and heat production measurements 3 hr after intraperitoneal administration of the β₃-adrenergic receptor agonist CL-316,243 (n = 3). Data are presented as fold change compared with before CL injection.

*p < 0.05. Error bars represent SEM.

were transferred to scintillation vials containing 5 ml scintillation liquid (Ecoscint XR, National Diagnostics) and the radioactivity was measured in a LS 6000 scintillation counter (Beckman Coulter).

Mitochondrial Isolation, Respiration, and Cytochrome Content

Mitochondria were purified as previously described (Parker et al., 2009). All steps were carried out at 4°C. Tissues were chopped with scissors in sodium chloride-Tris-EDTA (STE) buffer (250 mM sucrose, 5 mM Tris, and 2 mM EGTA [pH 7.4]) plus 1% BSA, homogenized manually (3 strokes with a loose-fit and 10 strokes with a tight-fit dounce homogenizer) and centrifuged at 8,500 *g* for 10 min. For BAT, the top fat layer was removed, and the tube walls were cleaned of fat. The pellet was resuspended in STE with 1% BSA and centrifuged at 400 *g* for 10 min. The supernatant was placed in a new tube and centrifuged at 8,500 *g*. The mitochondrial pellet was washed twice in BSA-free STE and resuspended in BSA-free STE. Protein concentration was determined by the BCA method. Mitochondrial respiration rates were recorded in a Clark-type electrode (Strathkelvin Instruments) as previously described (Oelkrug et al., 2010). Fifty μ g of mitochondria were incubated at 37°C under agitation in a final volume of 100 μ l in buffer containing 50 mM KCl, 10 mM TES, 1 mM KH₂PO₄, and 2 mM MgCl₂, 1 mM EGTA, 0.46 mM CaCl₂, 0.4% BSA, and 1 μ g/ml oligomycin (Sigma) (pH 7.4). Ten mM G3P, 5 mM pyruvate, 5 mM malate, or 4 mM succinate were used as substrates. Twenty μ M CoQ2 were used in rescue experiments. Cytochrome difference spectra were determined by dithionite reduced minus air oxidized scans on a DW-2 Olis dual-beam spectrophotometer. Isolated mitochondria (~0.5–1.0 mg/ml) were incubated in sodium chloride-HEPES-EDTA medium. Concentrations of cytochromes *c*, *c₁*, *b*, and *a* were calculated by simultaneous equation solving according to previously published studies (Schneider et al., 1980; Williams, 1964). The wavelength pairs and extinction coefficients for each cytochrome were cytochrome *c* 550–535 nm, 25.1 mM⁻¹ cm⁻¹; cytochrome *c₁* 554–540 nm, 24.1 mM⁻¹ cm⁻¹; cytochrome *b* 563–577 nm, 23.2 mM⁻¹ cm⁻¹; cytochrome *a* 605–630 nm, 13.1 mM⁻¹ cm⁻¹.

Statistical Analysis

Differences between two groups were determined using unpaired *t* test. Differences between three or more groups were analyzed by one-way ANOVA followed by Tukey's post hoc test for multiple comparisons. Power calculations were performed using online software (<http://statisticalsolutions.net>) with a desired sample size of ten on data sets with an *n* = 3. Data are presented as the mean value, and error bars represent SEM. Asterisks indicate significant comparisons, and specific *p* values are listed in each figure legend.

ACCESSION NUMBERS

The GEO accession number for the microarray data reported in this paper is GSE64060.

SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, four figures, and three tables and can be found with this article online at <http://dx.doi.org/10.1016/j.celrep.2014.12.048>.

AUTHOR CONTRIBUTIONS

C.M.A., M.K., J.W., S.V., R.L.S.G., C.L.Q., R.N., M.J., D.I.B., B.N., C.H., A.-A.N.V., M.J.P., D.Y., K.C., A.Y., and P.V. designed experiments and collected, analyzed, and interpreted data. A.S. conceived and designed the study and analyzed data. M.F. provided the *Cd36*^{-/-} mice. D.K.N., J.L.N., and M.D.B. designed experiments and analyzed data. C.M.A., M.K., and A.S. were involved in drafting or critically revising the manuscript. All authors approved the final version of the manuscript.

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REFERENCES

- Abumrad, N.A., el-Maghrabi, M.R., Amri, E.Z., Lopez, E., and Grimaldi, P.A. (1993). Cloning of a rat adipocyte membrane protein implicated in binding or transport of long-chain fatty acids that is induced during preadipocyte differentiation. Homology with human CD36. *J. Biol. Chem.* *268*, 17665–17668.
- Acton, S., Rigotti, A., Landschulz, K.T., Xu, S., Hobbs, H.H., and Krieger, M. (1996). Identification of scavenger receptor SR-BI as a high density lipoprotein receptor. *Science* *271*, 518–520.
- Aherne, W., and Hull, D. (1966). Brown adipose tissue and heat production in the newborn infant. *J. Pathol. Bacteriol.* *91*, 223–234.
- Ahmadian, M., Duncan, R.E., Varady, K.A., Frasson, D., Hellerstein, M.K., Birkenfeld, A.L., Samuel, V.T., Shulman, G.I., Wang, Y., Kang, C., and Sul, H.S. (2009). Adipose overexpression of desnutrin promotes fatty acid use and attenuates diet-induced obesity. *Diabetes* *58*, 855–866.
- Anderson, C.M., and Stahl, A. (2013). SLC27 fatty acid transport proteins. *Mol. Aspects Med.* *34*, 516–528.
- Argyropoulos, G., and Harper, M.E. (2002). Uncoupling proteins and thermoregulation. *J. Appl. Physiol.* *92*, 2187–2198.
- Artuch, R., Salviati, L., Jackson, S., Hirano, M., and Navas, P. (2009). Coenzyme Q₁₀ deficiencies in neuromuscular diseases. *Adv. Exp. Med. Biol.* *652*, 117–128.
- Bartelt, A., Bruns, O.T., Reimer, R., Hohenberg, H., Ittrich, H., Peldschus, K., Kaul, M.G., Tromsdorf, U.I., Weller, H., Waurisch, C., et al. (2011). Brown adipose tissue activity controls triglyceride clearance. *Nat. Med.* *17*, 200–205.
- Bastie, C.C., Hajri, T., Drover, V.A., Grimaldi, P.A., and Abumrad, N.A. (2004). CD36 in myocytes channels fatty acids to a lipase-accessible triglyceride pool that is related to cell lipid and insulin responsiveness. *Diabetes* *53*, 2209–2216.
- Bentinger, M., Tekle, M., and Dallner, G. (2010). Coenzyme Q—biosynthesis and functions. *Biochem. Biophys. Res. Commun.* *396*, 74–79.
- Bhagavan, H.N., and Chopra, R.K. (2006). Coenzyme Q₁₀: absorption, tissue uptake, metabolism and pharmacokinetics. *Free Radic. Res.* *40*, 445–453.
- Borel, P., de Edelenyi, F.S., Vincent-Baudry, S., Malezet-Desmoulin, C., Margotat, A., Lyan, B., Gorrard, J.M., Meunier, N., Drouault-Holowacz, S., and Bieuevet, S. (2011). Genetic variants in BCMO1 and CD36 are associated with plasma lutein concentrations and macular pigment optical density in humans. *Ann. Med.* *43*, 47–59.
- Borel, P., Lietz, G., Goncalves, A., Szabo de Edelenyi, F., Lecompte, S., Curtis, P., Goumidi, L., Caslake, M.J., Miles, E.A., Packard, C., et al. (2013). CD36 and SR-BI are involved in cellular uptake of provitamin A carotenoids by Caco-2 and HEK cells, and some of their genetic variants are associated with plasma concentrations of these micronutrients in humans. *J. Nutr.* *143*, 448–456.
- Calvo, D., Dopazo, J., and Vega, M.A. (1995). The CD36, CLA-1 (CD36L1), and LIMPII (CD36L2) gene family: cellular distribution, chromosomal location, and genetic evolution. *Genomics* *25*, 100–106.
- Calvo, D., Gómez-Coronado, D., Suárez, Y., Lasunción, M.A., and Vega, M.A. (1998). Human CD36 is a high affinity receptor for the native lipoproteins HDL, LDL, and VLDL. *J. Lipid Res.* *39*, 777–788.
- Coburn, C.T., Knapp, F.F., Jr., Febbraio, M., Beets, A.L., Silverstein, R.L., and Abumrad, N.A. (2000). Defective uptake and utilization of long chain fatty acids in muscle and adipose tissues of CD36 knockout mice. *J. Biol. Chem.* *275*, 32523–32529.

- Coburn, C.T., Hajri, T., Ibrahim, A., and Abumrad, N.A. (2001). Role of CD36 in membrane transport and utilization of long-chain fatty acids by different tissues. *J. Mol. Neurosci.* 16, 117–121, discussion 151–117.
- Crane, F.L. (2001). Biochemical functions of coenzyme Q₁₀. *J. Am. Coll. Nutr.* 20, 591–598.
- Cypess, A.M., Lehman, S., Williams, G., Tal, I., Rodman, D., Goldfine, A.B., Kuo, F.C., Palmer, E.L., Tseng, Y.H., Doria, A., et al. (2009). Identification and importance of brown adipose tissue in adult humans. *N. Engl. J. Med.* 360, 1509–1517.
- Febbraio, M., Abumrad, N.A., Hajjar, D.P., Sharma, K., Cheng, W., Pearce, S.F., and Silverstein, R.L. (1999). A null mutation in murine CD36 reveals an important role in fatty acid and lipoprotein metabolism. *J. Biol. Chem.* 274, 19055–19062.
- Febbraio, M., Podrez, E.A., Smith, J.D., Hajjar, D.P., Hazen, S.L., Hoff, H.F., Sharma, K., and Silverstein, R.L. (2000). Targeted disruption of the class B scavenger receptor CD36 protects against atherosclerotic lesion development in mice. *J. Clin. Invest.* 105, 1049–1056.
- Greaves, D.R., and Gordon, S. (2009). The macrophage scavenger receptor at 30 years of age: current knowledge and future challenges. *J. Lipid Res.* 50 (Suppl), S282–S286.
- Harmon, C.M., and Abumrad, N.A. (1993). Binding of sulfosuccinimidyl fatty acids to adipocyte membrane proteins: isolation and amino-terminal sequence of an 88-kD protein implicated in transport of long-chain fatty acids. *J. Membr. Biol.* 133, 43–49.
- Heaton, J.M. (1972). The distribution of brown adipose tissue in the human. *J. Anat.* 112, 35–39.
- Kiefer, C., Sumser, E., Wernet, M.F., and Von Lintig, J. (2002). A class B scavenger receptor mediates the cellular uptake of carotenoids in *Drosophila*. *Proc. Natl. Acad. Sci. USA* 99, 10581–10586.
- Kuniyasu, A., Hayashi, S., and Nakayama, H. (2002). Adipocytes recognize and degrade oxidized low density lipoprotein through CD36. *Biochem. Biophys. Res. Commun.* 295, 319–323.
- Kuzuyama, T., and Seto, H. (2012). Two distinct pathways for essential metabolic precursors for isoprenoid biosynthesis. *Proc. Jpn. Acad., Ser. B, Phys. Biol. Sci.* 88, 41–52.
- Lass, A., Agarwal, S., and Sohal, R.S. (1997). Mitochondrial ubiquinone homologues, superoxide radical generation, and longevity in different mammalian species. *J. Biol. Chem.* 272, 19199–19204.
- Moussa, M., Gouranton, E., Gleize, B., Yazidi, C.E., Niot, I., Besnard, P., Borel, P., and Landrier, J.F. (2011). CD36 is involved in lycopene and lutein uptake by adipocytes and adipose tissue cultures. *Mol. Nutr. Food Res.* 55, 578–584.
- Murphy, M.P. (2009). How mitochondria produce reactive oxygen species. *Biochem. J.* 417, 1–13.
- Nergiz-Unal, R., Rademakers, T., Cosemans, J.M., and Heemskerk, J.W. (2011). CD36 as a multiple-ligand signaling receptor in atherothrombosis. *Cardiovasc. Hematol. Agents Med. Chem.* 9, 42–55.
- Oelkrug, R., Kutschke, M., Meyer, C.W., Heldmaier, G., and Jastroch, M. (2010). Uncoupling protein 1 decreases superoxide production in brown adipose tissue mitochondria. *J. Biol. Chem.* 285, 21961–21968.
- Padilla-López, S., Jiménez-Hidalgo, M., Martín-Montalvo, A., Clarke, C.F., Navas, P., and Santos-Ocaña, C. (2009). Genetic evidence for the requirement of the endocytic pathway in the uptake of coenzyme Q6 in *Saccharomyces cerevisiae*. *Biochim. Biophys. Acta* 1788, 1238–1248.
- Parker, N., Crichton, P.G., Vidal-Puig, A.J., and Brand, M.D. (2009). Uncoupling protein-1 (UCP1) contributes to the basal proton conductance of brown adipose tissue mitochondria. *J. Bioenerg. Biomembr.* 41, 335–342.
- Plüddemann, A., Neyen, C., and Gordon, S. (2007). Macrophage scavenger receptors and host-derived ligands. *Methods* 43, 207–217.
- Podda, M., Weber, C., Traber, M.G., Milbradt, R., and Packer, L. (1999). Sensitive high-performance liquid chromatography techniques for simultaneous determination of tocopherols, tocotrienols, ubiquinol, and ubiquinone in biological samples. *Methods Enzymol.* 299, 330–341.
- Quinzii, C.M., López, L.C., Naini, A., DiMauro, S., and Hirano, M. (2008). Human CoQ₁₀ deficiencies. *Biofactors* 32, 113–118.
- Reboul, E., and Borel, P. (2011). Proteins involved in uptake, intracellular transport and basolateral secretion of fat-soluble vitamins and carotenoids by mammalian enterocytes. *Prog. Lipid Res.* 50, 388–402.
- Schneider, H., Lemasters, J.J., Höchli, M., and Hackenbrock, C.R. (1980). Fusion of liposomes with mitochondrial inner membranes. *Proc. Natl. Acad. Sci. USA* 77, 442–446.
- Sekhar, B.S., Kurup, C.K., and Ramasarma, T. (1990). Microsomal redox systems in brown adipose tissue: high lipid peroxidation, low cholesterol biosynthesis and no detectable cytochrome P-450. *Mol. Cell. Biochem.* 92, 147–157.
- Sell, H., Deshaies, Y., and Richard, D. (2004). The brown adipocyte: update on its metabolic role. *Int. J. Biochem. Cell Biol.* 36, 2098–2104.
- Sohet, F.M., and Delzenne, N.M. (2012). Is there a place for coenzyme Q in the management of metabolic disorders associated with obesity? *Nutr. Rev.* 70, 631–641.
- Su, X., and Abumrad, N.A. (2009). Cellular fatty acid uptake: a pathway under construction. *Trends Endocrinol. Metab.* 20, 72–77.
- Taskinen, M.R., Nikkilä, E.A., Kuusi, T., and Tulikoura, I. (1983). Changes of high density lipoprotein subfraction concentration and composition by intralipid in vivo and by lipolysis of intralipid in vitro. *Arteriosclerosis* 3, 607–615.
- Thelin, A., Schedin, S., and Dallner, G. (1992). Half-life of ubiquinone-9 in rat tissues. *FEBS Lett.* 313, 118–120.
- Tomasetti, M., Alleva, R., Solenghi, M.D., and Littarru, G.P. (1999). Distribution of antioxidants among blood components and lipoproteins: significance of lipids/CoQ₁₀ ratio as a possible marker of increased risk for atherosclerosis. *Biofactors* 9, 231–240.
- van Marken Lichtenbelt, W.D., Vanhommerig, J.W., Smulders, N.M., Drosaeerts, J.M., Kemerink, G.J., Bouvy, N.D., Schrauwen, P., and Teule, G.J. (2009). Cold-activated brown adipose tissue in healthy men. *N. Engl. J. Med.* 360, 1500–1508.
- Van Nieuwenhoven, F.A., Verstijnen, C.P., Abumrad, N.A., Willemsen, P.H., Van Eys, G.J., Van der Vusse, G.J., and Glatz, J.F. (1995). Putative membrane fatty acid translocase and cytoplasmic fatty acid-binding protein are co-expressed in rat heart and skeletal muscles. *Biochem. Biophys. Res. Commun.* 207, 747–752.
- Virtanen, K.A., Lidell, M.E., Orava, J., Heglind, M., Westergren, R., Niemi, T., Taittonen, M., Laine, J., Savisto, N.J., Enerbäck, S., and Nuutila, P. (2009). Functional brown adipose tissue in healthy adults. *N. Engl. J. Med.* 360, 1518–1525.
- Williams, J.N., Jr. (1964). A method for the simultaneous quantitative estimation of cytochromes a, b, c1, and c in mitochondria. *Arch. Biochem. Biophys.* 107, 537–543.
- Wu, Q., Kazantzis, M., Doege, H., Ortegon, A.M., Tsang, B., Falcon, A., and Stahl, A. (2006). Fatty acid transport protein 1 is required for nonshivering thermogenesis in brown adipose tissue. *Diabetes* 55, 3229–3237.
- Zeng, Y., Tao, N., Chung, K.N., Heuser, J.E., and Lublin, D.M. (2003). Endocytosis of oxidized low density lipoprotein through scavenger receptor CD36 utilizes a lipid raft pathway that does not require caveolin-1. *J. Biol. Chem.* 278, 45931–45936.
- Zimmet, P., Alberti, K.G., and Shaw, J. (2001). Global and societal implications of the diabetes epidemic. *Nature* 414, 782–787.