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### Authors

Richards, Emilie J  
Servedio, Maria R  
Martin, Christopher H

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# Searching for Sympatric Speciation in the Genomic Era

Emilie J. Richards, Maria R. Servedio, and Christopher H. Martin\*

**Sympatric speciation illustrates how natural and sexual selection may create new species in isolation without geographic barriers. However, recent genomic reanalyses of classic examples of sympatric speciation reveal complex histories of secondary gene flow from outgroups into the radiation. In contrast, the rich theoretical literature on this process distinguishes among a diverse range of models based on simple genetic histories and different types of reproductive isolating barriers. Thus, there is a need to revisit how to connect theoretical models of sympatric speciation and their predictions to empirical case studies in the face of widespread gene flow. Here, theoretical differences among different types of sympatric speciation and speciation-with-gene-flow models are reviewed and genomic analyses for distinguishing which models apply to case studies based on the timing and function of adaptive introgression are proposed. Investigating whether secondary gene flow contributed to reproductive isolation is necessary to test whether predictions of theory are ultimately borne out in nature.**

panmictic gene flow and no initial divergence at the start of speciation.<sup>[2–6]</sup> In the context of theoretical speciation models, this type of speciation process is the most difficult because the starting conditions involve no pre-existing divergence among loci involved in reproductive isolation. Instead, linkage disequilibrium (LD) must build up through time within a population through the action of disruptive natural selection and strong assortative mating by ecotype, despite the countervailing eroding force of recombination.<sup>[7–10]</sup>

Recently, the definition of sympatric speciation has been expanded to focus more on biogeographical context in line with Mayr's original definition,<sup>[11]</sup> in which the speciation process is defined as sympatric a) as long as diverging populations are within "cruising range" of each other and b) regardless of whether secondary gene flow provided alleles con-

## 1. Introduction: What Is Sympatric Speciation?

As Mayr famously quipped, sympatric speciation is like the Lernean Hydra: "which grew two new heads whenever one of its heads was cut off" (p. 451<sup>[1]</sup>). The latest incarnation of this phenomenon has occurred over the past decade: sympatric speciation now means two different things to different research groups. We stress that our goal here is not to offer a new definition of sympatric speciation nor grow a new head on the hydra, but only to clarify existing usage so that we can focus on reconciling diverse theoretical models with existing empirical examples of this process.


Subsequent to Mayr's classic definition based on geography (1947), sympatric speciation was redefined over the past two decades in a population genetic context as the most extreme endpoint on the continuum of divergence with gene flow:

contributing to reproductive isolating barriers in sympatry. Cruising range provides a practical empirical definition of gene flow between diverging sympatric populations, allowing for some geographic or microallopatric population structure.<sup>[11,12]</sup> Allowing for the secondary gene flow of alleles contributing some or all of the reproductive isolation between sympatric populations also expands the definition of sympatric speciation to include both "hard" and "easy" processes under one umbrella. Theoretical models show that speciation is much easier from starting conditions that involve some level of initial divergence and/or restricted gene flow,<sup>[13]</sup> for example, if the alleles necessary for reproductive isolation first become physically linked in allopatry (Box 1).

These two definitions, the population genetic and biogeographic, reflect different perspectives on the value of studying sympatric speciation. The biogeographic definition (for clarity, we will refer to this process as "easy sympatric speciation"), with its broader range of starting conditions that are easier to verify in nature, increases, perhaps vastly, the number of empirical speciation events that could be categorized as examples of sympatric speciation. This definition values the frequency of sympatric diverging populations in nature compared to allopatric speciation as an estimate of the overall importance of "sympatry" in contributing to biodiversity. The population genetic definition (for clarity, we will refer to this more restrictive process as "hard sympatric speciation"), with its narrow set of starting conditions that remain challenging to verify in nature, finds value in studying both the easy and hard processes of sympatric speciation defined in theoretical models.

E. J. Richards, Dr. M. R. Servedio, Dr. C. H. Martin  
 Department of Biology  
 University of North Carolina at Chapel Hill  
 Chapel Hill, NC 27599, USA  
 E-mail: chmartin@unc.edu

E. J. Richards, Dr. C. H. Martin  
 Department of Integrative Biology and Museum of Vertebrate Zoology  
 University of California  
 Berkeley, CA 94720, USA

 The ORCID identification number(s) for the author(s) of this article can be found under <https://doi.org/10.1002/bies.201900047>.

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Namely, it values the theoretical possibility of creating new species solely through the power of divergent selection alone, regardless of whether this process is common in nature. Here we focus on the types of questions that genomic data now allow us to ask to improve the search for examples of both the easy and hard processes of sympatric speciation, and investigate the range of speciation mechanisms found in nature from among those shown to be plausible in theory.

Under the biogeographical definition of sympatric speciation, there is little difference in terms of the speciation mechanisms said to be involved in scenarios that start with initial panmixia (i.e., hard sympatric speciation) versus those that start with some geographic or microallopatric population structure (i.e., easy sympatric speciation<sup>[11,12]</sup>). However, this contrasts with the theoretical literature, which differentiates models of hard sympatric speciation from other models of speciation with gene flow. Indeed, theory teaches us that the hard process of sympatric speciation (without the aid of secondary gene flow contributing to reproductive isolation) is uniquely and notoriously difficult,<sup>[25]</sup> in part because quite specific conditions of resource availability,<sup>[8,20]</sup> mating traits and preferences,<sup>[14,26]</sup> and search costs<sup>[27]</sup> must be met for it to occur. Some argue that the effort to discern the exact geographic scenario and initial conditions of speciation would be better spent on finding loci involved in reproductive isolation (i.e., “barrier loci”<sup>[28,29]</sup>). This is an important first step and we can glean something about the process of speciation from gene annotations of barrier loci and linkage architecture. However, understanding whether any one particular locus or potential mechanism was necessary for speciation often requires placing genomic discoveries in the context of speciation models that explicitly compare the importance of such factors and mechanisms in driving divergence, models whose outcomes are highly dependent on the initial conditions before sympatric divergence.

## 2. Different Mechanistic Processes Underlie Divergence in Sympatry

Regardless of definition, it is necessary to distinguish among different sympatric divergence processes to understand which classes of speciation models and predictions apply to specific case studies. We here distinguish different scenarios (**Figure 1**) that will result in two sister species in sympatry based on whether secondary gene flow aided in population divergence: 1) hard sympatric speciation without gene flow; 2) hard sympatric speciation in the presence of a) neutral secondary gene flow or b) after differential sorting of an ancestral hybrid swarm. In the latter case, we also distinguish whether the ancestral hybrid swarm population achieved panmixia before later divergence (i.e., hard sympatric speciation); otherwise, differential sorting of haplotypes within the hybrid swarm is better described by secondary contact speciation-with-gene-flow models rather than sympatric speciation models. 3) Easy sympatric speciation may be aided by secondary gene flow that a) triggers initial sympatric divergence or b) increases divergence after initial divergence in sympatry becomes stalled, an outcome of many sympatric speciation models without sufficiently strong disruptive selection.<sup>[21,31–33]</sup> Finally, 4) secondary contact after a period of allopatry between two populations can result in

coexistence or reinforcement if there is no collapse into a single admixed population<sup>[13,22,34,35]</sup> or extinction of one or both populations.

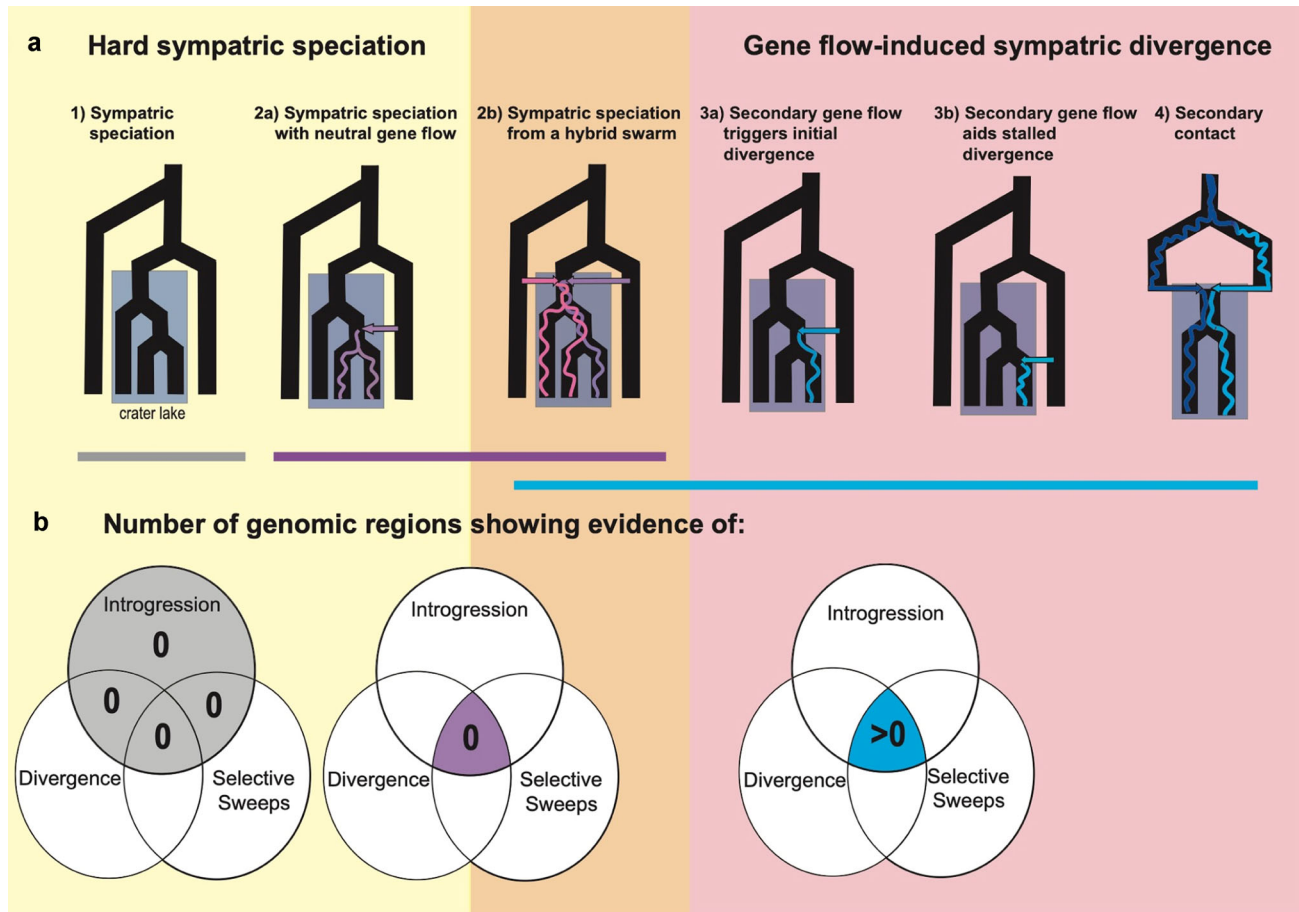
### 2.1. Some Scenarios of Sympatric Divergence Are Easier Than Others

We consider scenarios 1 and 2a to be examples of hard sympatric speciation, whereas scenarios 3 and 4 would be examples of speciation aided by secondary gene flow, a much easier process in theory. Interestingly, hybrid swarm scenarios (2b) exist in a gray area since substantial initial gene flow from multiple sources may increase ecological or preference variation within a population that is sufficient to trigger later sympatric divergence, even without segregating inversions or genetic incompatibilities.<sup>[15,36,37]</sup> So far, we know of no examples of scenario 1 within any case study of sympatric sister species examined using genomic tools; even long diverged species show some evidence of introgression from outgroups in their past.<sup>[38]</sup> In contrast, sympatric speciation with neutral gene flow (scenario 2a, and conditionally scenario 2b) and speciation aided by gene flow (scenarios 3 and 4) frequently appear to operate concurrently even within a single sympatric adaptive radiation.<sup>[39–41]</sup>

### 2.2. Why Should We Distinguish between Different Sympatric Divergence Scenarios?

It is important to distinguish these scenarios because theoretical models predict that sympatric divergence unaided by any form of secondary gene flow is substantially more difficult than other speciation with gene flow scenarios (Box 1). Gene flow throughout the speciation process allows recombination to break down LD among alleles associated with ecological divergence and assortative mating. There are also three different types of reproductive isolating traits to consider within sympatric speciation models: the most difficult process involves independently segregating loci for ecotype, female preferences, and male traits within the population, whereas sympatric divergence is much easier if any of these three types of traits are combined (i.e., cannot become disassociated by gene flow and recombination), such as assortative mating based on phenotype matching instead of separate loci for preference and traits<sup>[42,43]</sup> or “magic” traits (such as assortative mating based on microhabitat preference<sup>[8,9,44]</sup>). Sympatric speciation by sexual selection alone is also theoretically possible (albeit considered highly unlikely) if there is substantial preference variation either initially within the population or through secondary gene flow.<sup>[14,15]</sup>

Any form of LD among ecological and mate choice loci formed in allopatry, whether due to physical linkage, selection, or drift, can thus tend to shift the initial starting conditions of panmixia in favor of sympatric divergence.<sup>[22]</sup> However, LD without physical linkage subsides within a relatively small number of generations after secondary sympatry and thus may not allow sufficient time for the evolution of assortative mating



**Figure 1.** Genomic signatures of sympatric speciation and speciation with gene flow. Speciation scenarios are grouped into hard sympatric speciation scenarios (yellow box; harder process in theoretical models) and other divergence scenarios that can also occur in sympatry (red box; easier processes, which we refer to broadly as sympatric divergence here). Speciation from a hybrid swarm (orange box) can fall under either class of scenarios and additional information is necessary to determine what category of speciation models best describes this process. A) The timing of gene flow relative to divergence can be used to distinguish between speciation scenarios. The colored arrows represent gene flow events and the colored lines within the tree are simplified representations of a signature of introgression from that gene flow event into the sympatric species. B) Venn diagrams illustrating the number of genomic windows across the entire genome expected to have overlapping signatures of introgression (e.g.,  $f_d$  outliers), genetic divergence (e.g.,  $F_{ST}$  and  $D_{xy}$  outliers), and selective sweeps (e.g., SweeD) for each speciation scenario.<sup>[30]</sup> The highlighted sections of the Venn diagrams indicate the key signature that can be used to distinguish between the scenarios. The scenarios that are expected to leave very similar signatures of overlap are grouped by the bars colored with their respective Venn diagram.

within the population. In contrast, pre-existing physical linkage among ecological loci has been shown to increase the probability of divergence, especially when it captures already divergent alleles, as is more likely after a period of divergence in allopatry before secondary contact.<sup>[18,45]</sup> Similarly, physical linkage can cause preference and trait alleles to mimic phenotype matching, although even tight linkage can break down over long timescales (shown in a model with population structure<sup>[17]</sup>). Segregating inversions in the ancestral population are now well-known empirical examples of physical linkage promoting divergence in sympatry.<sup>[18,46]</sup> Sympatric divergence is also limited by many other restrictive conditions including the costs of female choosiness and strengths of disruptive selection and assortative mating.

Despite extensive searches for examples of sympatric speciation in the wild, there are few convincing case studies due to the

difficulty of ruling out historical allopatric scenarios (see below) and ruling out a role of introgression in speciation. Furthermore, the role of magic traits or matching vs preference/trait mechanisms is not fully understood in any existing case study. Thus, we still have very limited empirical tests of an extensive theoretical literature and diverse competing models of the notoriously difficult process of sympatric speciation.<sup>[42,43,47,48]</sup>

### 3. The Classic Criteria for Sympatric Speciation Do Not Distinguish between Different Sympatric Divergence Scenarios

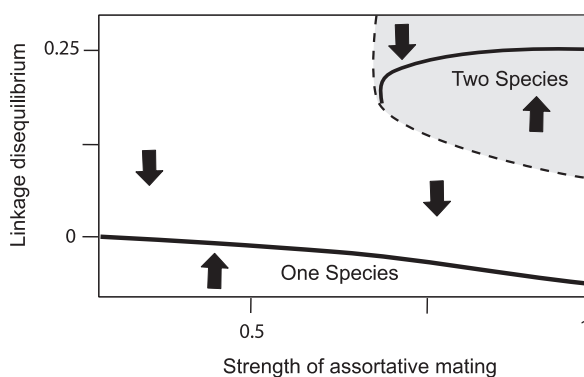
There are four traditional criteria for demonstrating hard sympatric speciation without secondary gene flow (e.g., scenario 1 in Figure 1A): 1) sister species have to be reproductively

**Box 1**  
**Why do we care whether speciation is sympatric?**

Inferences from theoretical models predict that, under a scenario of speciation with gene flow (scenario 3), introgression can make the process of speciation much easier in three ways. First, by introducing additional variation in ecological traits into the population, introgression could potentially facilitate a branching process due to competition for resources (although we are not aware of a model that assesses this precise situation, it can be inferred from the dynamics of Dieckmann and Doebeli<sup>[8]</sup>). Second, introgression of novel alleles for mating preferences may provide a boost in preference variation that could be an important trigger to aid the evolution of assortative mating under a preference/trait mechanism, which requires preference variation to be large.<sup>[14,15]</sup> For example, we found evidence of secondary gene flow of olfactory alleles shortly before the rapid divergence of a Cameroon cichlid radiation in Lake Ejagham, which may have boosted preference variation.<sup>[16]</sup> Third, secondary gene flow after a period of allopatric isolation may lead to increased LD between assortative mating and ecological loci or among ecological loci. It seems logical that this might facilitate sympatric speciation as this metric is often described as progress along the speciation continuum. However, initial LD has been shown not to matter much in at least some scenarios<sup>[7]</sup> because without physical linkage, LD breaks down quickly. However, physical linkage may enable these alleles to remain in association for a sufficient time for assortative mating to evolve within the population.<sup>[17]</sup> Initial LD may also increase the probability of allelic capture by an inversion or for selection for new mutations within an inversion that may

affect both ecology and assortment.<sup>[18]</sup> Finally, higher LD among ecological loci may in some cases increase the probability of sympatric divergence, but this is in effect similar to varying effect sizes of alleles at ecological loci (e.g., many small-effect alleles within a region resemble a large-effect locus<sup>[19]</sup>). These predictions could also apply to sympatric radiations. For example, some classic sympatric speciation models<sup>[8]</sup> can yield many more than just two species if left to run for more generations.<sup>[20,21]</sup>

The fundamental difference between sympatric speciation and speciation with gene flow, including secondary contact scenarios, lies in the fact that very often multiple equilibrium states exist in speciation models, such that loss of divergence and maintenance of divergence in the presence of gene flow are both possible outcomes, depending on the starting conditions of a population (this is nicely illustrated for one measure of divergence by Kirkpatrick and Ravnigé,<sup>[22]</sup> **Figure 2**). In such cases, speciation is much more easily reached from starting conditions that match those of two populations that have diverged largely in allopatry due to the large amount of allelic variation or pre-existing phenotypic bimodality and assortative mating.<sup>[13]</sup> Even for scenarios of speciation with gene flow that are much easier, such as geographic separation between two incipient species that are undergoing gene flow or involving some form of magic trait, differentiation is much more difficult to reach or maintain from an initially homogeneous population than from an initially differentiated one.<sup>[23,24]</sup>



**Figure 2.** Two equilibrium cases exist for the LD, a proxy for differentiation into two distinct “species” in this proof-of-concept model, that can be maintained between two loci that are under disruptive selection and determine assortative mating. With little initial LD, the one-species equilibrium is likely to be reached even when the intensity of assortment is high. When LD in the traits is initially large, as can be the case if there is initially divergence in allopatry, the two-species equilibrium can be reached instead. Adapted with permission.<sup>[22]</sup> Copyright 2002, The University of Chicago Press.<sup>[22]</sup>

isolated, 2) form a monophyletic group, 3) largely overlap in ranges, and 4) have biogeographic and evolutionary histories that make periods of allopatric divergence highly unlikely.<sup>[6]</sup> Very few case studies have been able to meet these rigorous criteria despite intensive searches.<sup>[2,6]</sup> This has led to the prominent status of crater lake cichlids as some of the best examples of sympatric speciation in the wild due to the uniform shape of isolated volcanic lakes, which convincingly rule out phases of allopatry due to water-level changes (Box 2).<sup>[61]</sup>

The monophyly criterion assumes that monophyly arises only when a single ancestral population underlies the present-day daughter species. This is typically met by inferring a single phylogeny from one or more loci. This single point-estimate view of evolutionary history is problematic because it obscures the presence of non-bifurcating relationships among organisms (e.g., sister species that derived ancestry from multiple source populations due to extensive gene flow or hybrid speciation) and the real variation in evolutionary histories among genes across the genome itself.<sup>[62]</sup> Few regions of the genome may initially contribute to reproductive isolation resulting in a heterogeneous genomic landscape of differentiation among incipient species,<sup>[63]</sup> a pattern now extensively supported across case studies.<sup>[64–66]</sup> Therefore, monophyletic relationships are

**Box 2**

**Evidence for sympatric speciation from crater lake cichlid radiations.**

There are relatively few volcanic chains of crater lakes containing fishes in the tropics, notably found in Cameroon, Nicaragua, Tanzania, Uganda, Madagascar, and Papua New Guinea.<sup>[49–51]</sup> Although sympatric radiations of endemic fishes are known from other isolated saline, alkali, postglacial, and ancient lakes, only four lineages of cichlids have radiated in the world's crater lakes (**Figure 3**). The most diverse radiation is Barombi Mbo, Cameroon, with eleven endemic cichlid species, followed by Lake Bermin, Cameroon, with nine.<sup>[52]</sup> Nicaraguan crater lakes reach up to five species,<sup>[53]</sup> the East African craters never exceed two sympatric species,<sup>[49,54]</sup> and Madagascar's crater lakes contain a single endemic cichlid.<sup>[51]</sup> It remains unknown why regional and lineage diversity varies so greatly because there appears to be no relationship between the occurrence of endemic cichlid radiations and crater lake size or age (up to approximately 5 km diameter and 2 million years old).<sup>[55]</sup>

The evidence for secondary gene flow is remarkably similar across all crater lake cichlid radiations examined with genomic data so far. Admixture proportions with outgroups are frequently detected within the range of 1–4%: 0.6% in Lake Barombi Mbo *Sarotherodon* (percentage of polyphyletic trees in *Saguaro*<sup>[56]</sup>), 1.1% in Lake Massoko *Astatotilapia* (Patterson's  $D$ <sup>[49]</sup>), 4.3% in Lake Apoyo *Amphilophus* (demographic model<sup>[53]</sup>), and 4.4% in Lake Ejagham *Coptodon* (1138  $f_d$  outliers<sup>[16]</sup>), although notably these studies all used different metrics of introgression. No case studies have yet found evidence of substantial divergence in allopatry followed by secondary contact (but see Lake Xiloá *Amphilophus*<sup>[53]</sup>). Instead, nearly all studies have concluded sympatric divergence with periodic or continuous gene flow, potentially from an initial hybrid swarm population (i.e., introgression from multiple outgroup populations).

Secondary gene flow may have triggered sympatric divergence in a radiation of three *Coptodon* cichlids in Lake Ejagham: demographic analyses of whole genomes suggest that this population did not diversify for 8000 years despite frequent gene flow until an influx of olfactory receptor alleles 1000

years ago, coinciding with the first sympatric divergence in the lake.<sup>[16]</sup> In Lake Victoria, segregating opsin alleles in riverine cichlid populations were differentially sorted among Lake Victorian cichlids and may have triggered their diversification.<sup>[40]</sup>

Evidence for hard sympatric speciation in crater lake cichlids without the presence of secondary gene flow remains elusive, but some studies have suggested that introgressed variation may not have played a role in speciation between sympatric species. Malinsky et al.<sup>[49]</sup> showed that 1.1% introgression occurred long before the major divergence of a shallow/deep-water sister species pair of cichlids in Lake Massoko, Tanzania. These timing estimates and the observation of a weak correlation between regions of elevated  $F_{st}$  and candidate introgressed regions were used to argue that secondary gene flow may not have played a causal role in divergence in this system. However, this initial introgression may still have aided later sympatric divergence (which admittedly is very difficult to rule out) and nothing is yet known about selective sweeps of introgressed variation.

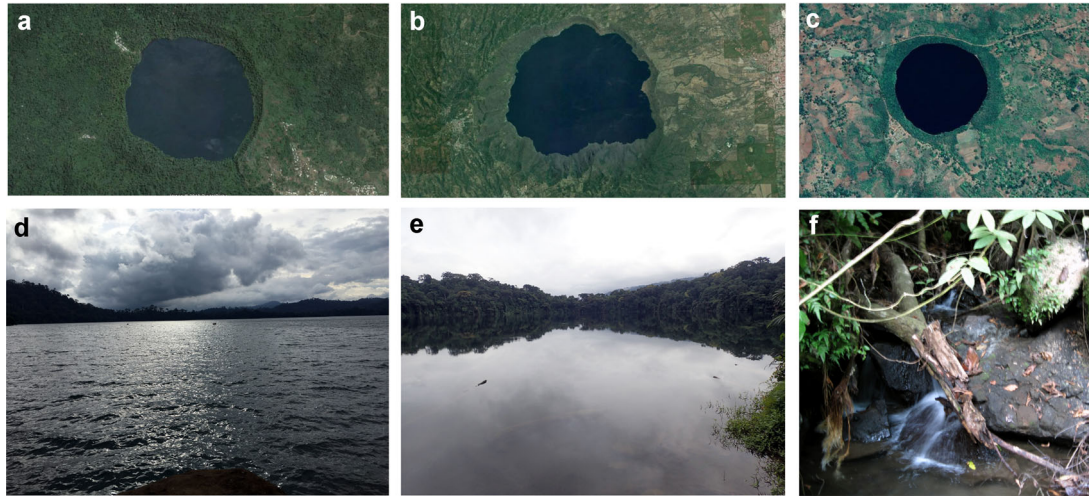
Very recent sympatric divergence in some crater lakes or the proliferation of many species from a few colonization events may also suggest that divergence occurred in sympatry without the aid of gene flow;<sup>[53,57]</sup> however, in the former case it remains unclear if incipient divergence will continue to complete reproductive isolation or become stalled as appears to be the case in some species complexes of Cameroon crater lake cichlids.<sup>[39,58]</sup> Very rare secondary gene flow into the Barombi Mbo cichlid radiation (0.6% introgression) without a clear functional role provides weak evidence of sympatric divergence, but more functional characterization and timing of introgression are needed.<sup>[56]</sup> The recent advent of transgenic reporters, clustered regularly interspaced short palindromic repeats (CRISPR)–CRISPR-associated protein 9 (Cas9), and in situ hybridization genetic tools within Nicaraguan crater lake cichlids provides much promise for future investigations of the role of introgression in sympatric divergence.<sup>[59,60]</sup>

consistent with, but not exclusive to, a scenario of sympatric speciation. Examining heterogeneous evolutionary histories across regions relevant to speciation is thus crucial for understanding the processes and conditions under which sympatric divergence can occur.

#### **4. The “New” Problem of Sympatric Speciation Is to Establish or Reject a Functional Role for Secondary Gene Flow**

While genomics has increased our ability to resolve evolutionary relationships among organisms, it has also revealed more

complex evolutionary histories of multiple colonizations and extensive secondary gene flow in nearly all examples of sympatric speciation that have been examined with genomic data so far<sup>[16,49,53,54,56,67–71]</sup> (e.g., to our knowledge Lord Howe Island palms and indigobirds have not yet been directly examined for secondary gene flow with an outgroup). Indeed, only a handful of genes may directly contribute to the speciation process whereas the rest of the genome is porous to gene flow while reproductive isolation is incomplete.<sup>[63,72]</sup> Examples of sympatric speciation without secondary gene flow (scenario 1) are now even rarer after applying modern genomic tools to search for introgression. Instead, it is still possible that even the hard process of sympatric speciation may occur in the face of secondary gene flow in nearly



**Figure 3.** Examples of volcanic crater lakes containing endemic cichlid radiations around the globe: a,d,f) Barombi Mbo, Cameroon, and its only outlet stream; b) Lake Apoyo, Nicaragua, c) Lake Massoko, Tanzania, e) Lake Bermin, Cameroon. Satellite images (a–b) from Google, DigitalGlobe 2019 and (c) Google, CNES/Airbus 2019; (d–f) by Christopher H. Martin.

all these examples (scenario 2a<sup>[56]</sup>). Importantly, most evidence of secondary gene flow impacting putative examples of sympatric speciation comes from genome-wide tests of introgression from outgroup lineages that do not look at how that secondary gene flow has impacted reproductive isolating barriers between diverging populations in sympatry.<sup>[53,69]</sup> In case studies of sympatric speciation that involve radiations of species, secondary gene flow may also impact only some of the diverging populations such that some species within a radiation may better represent sympatric speciation scenarios than others. Therefore, introgression detected at the genome-wide level from lineages outside the speciation event tells us that secondary gene flow has occurred, but little about the divergence process among incipient sympatric species and how that gene flow shaped the process of speciation.

The challenge of understanding the hard process of sympatric speciation in the genomic era is establishing or rejecting a functional role for the secondary gene flow commonly present during the speciation process, in effect ruling out scenarios 3 and 4 in favor of scenario 2 (Figure 1). Even if signatures of secondary gene flow are detected, speciation could still have occurred solely via mechanisms of hard sympatric speciation if that secondary gene flow did not play a causal role in divergence (scenario 2a and possibly 2b). In contrast, secondary gene flow could play a causal role if it introduced novel genetic variation or physically linked alleles (e.g., a segregating inversion) that promoted divergence through mechanisms such as inflating variance through the creation of a hybrid swarm (scenario 2b),<sup>[36,37,40,73]</sup> adaptive introgression (scenario 3),<sup>[30,74–76]</sup> transgressive segregation (scenarios 2 and 3),<sup>[77,78]</sup> or hybrid speciation.<sup>[79]</sup> Beyond examples of sympatric speciation, genetic variation brought in through gene flow with divergent lineages has been found in many empirical examples of rapid speciation and recognized as a potential mechanism for rapid speciation and adaptive radiation.<sup>[80]</sup> Here we propose and discuss genomic analyses that may help to establish or reject a functional role of

secondary gene flow in the speciation process (Figure 1). This is necessary to identify putative cases of hard sympatric speciation when gene flow appears to be nearly universal in the wild, particularly among sympatric diverging populations.

## 5. Genomic Analyses Can Aid in Distinguishing between Different Scenarios of Sympatric Speciation

Although genome-wide analyses of introgression provide a starting point, ultimately consideration of the time of arrival and functional role of each introgressed region within extant sympatric sister species pairs will be necessary to distinguish between hard sympatric speciation in which incidental gene flow does not contribute to reproductive isolating barriers (scenario 2a) versus easy sympatric speciation in which divergence is aided by secondary gene flow (scenario 3; e.g., segregating inversions<sup>[81,82]</sup> or balancing selection on regions containing multiple barrier loci<sup>[83,84]</sup>). We suggest four major types of genomic analyses to address questions about the role of secondary gene flow and identify sympatric speciation with gene flow: analyses to 1) estimate the timing of introgression into sympatric sister species relative to their divergence time, 2) infer the presence and timing of selective sweeps within sympatric sister species, 3) annotate candidate adaptive introgression regions for functional elements or trait associations that may be relevant to speciation, and 4) if closely related nonspeciating outgroups are available, confirm the lack of selective sweeps of these regions in outgroups. Some of these analyses are already being applied to examples of sympatric speciation in the wild, particularly crater lake cichlid systems, taking on the difficult task of distinguishing between sympatric speciation scenarios. These analyses are by no means trivial as evidence for either “hard” or “easy” sympatric speciation scenarios in the wild remains sparse, but recently developed methods have made it possible to start addressing such challenging questions.

**Box 3**

**Tools for detecting and timing adaptive introgression.**

- 1) Detecting and timing introgression. Although there are a variety of tests to detect gene flow on a local scale or within sliding genomic windows, currently three major types of demographic coalescent modeling approaches can infer the timing of introgression based on different genomic information: 1) the distribution of allele frequencies from genotype data (site frequency spectrum<sup>[85,86]</sup>), 2) the distribution of haplotype block lengths from phased genomes,<sup>[87,88]</sup> and 3) variation in coalescent patterns among gene trees.<sup>[89]</sup> Distinguishing introgressed variation from variation that stems from the incomplete sorting of ancestral polymorphisms between diverging species (which does not rule out a scenario of hard sympatric speciation) is still challenging because these two processes can often lead to very similar patterns in the genome. However, new population genetic simulation methods enable comparisons of genetic patterns simulated under arbitrarily complex demographic scenarios.<sup>[90]</sup> For example, we may now be able to simulate genomes evolving under complex histories of gene flow and introgression compared to simulated genomes evolving under complex histories of ancestral population structure and biased sorting of ancestral genetic variation, and then use subtle differences between the two simulated datasets to detect introgressed variation in empirical genomes. Improvements in machine-learning methods to distinguish between demographic scenarios that can produce similar genetic patterns are also advancing rapidly.<sup>[91–94]</sup>
- 2) Timing of selective sweeps. Recent methods for estimating the age of a selective sweep exploit different aspects about the pattern of variation surrounding the allele on its haplotype background. These include heuristic approaches that use point estimates of mean haplotype length or the number of derived mutations within a chosen distance of the site,<sup>[95,96]</sup> model-based approaches that use demographic information and summary statistics of allele frequencies and LD to model a distribution of ages that fit the observed data,<sup>[97–99]</sup> and full sequence approaches that leverage the length of ancestral haplotypes surrounding the beneficial allele and the accumulation of derived mutations.<sup>[100,101]</sup>
- 3) Functional analyses of introgressed variants. Functional annotation of introgressed regions minimally involves searching an annotated reference genome for genes with relevant functions known from model organisms. Intergenic regions can be searched for evidence of strong sequence conservation across taxa<sup>[102]</sup> or potential regulatory elements.<sup>[103]</sup> Additionally, genome-wide association studies (GWAS) can identify variants in introgressed regions correlated with reproductive isolating barriers. Functional validation of gene and regulatory element variants through genome-editing experiments is also becoming increasingly tractable for non-model organisms.<sup>[59]</sup>

**5.1. Is the Observed Secondary Gene Flow Concurrent with Divergence Times?**

Estimating the duration of gene flow and the timing of introgression into a sister species from an outgroup relative to the timing of divergence between sympatric sister species will help distinguish between scenarios of sympatric speciation, speciation with gene flow, and secondary contact. If populations diverged in sympatry independent of any concurrent secondary gene flow (scenario 2a), we might expect to see weak concordance of the timing of gene flow with divergence times among species; for example, discrete gene flow events that date well before or after divergence times among species. In the case of both discrete gene flow events surrounding divergence time estimates or continuous gene flow from the time of colonization to the present, more information about function and selection on regions introgressed near the time of speciation will be needed. Increasingly sophisticated approaches for detecting fine-scale patterns of introgression and inferring the timing and duration of gene flow from genomic data are becoming available (Box 3).

**5.2. Did Any of the Introgressed Regions Experience Selective Sweeps and Did the Timing of These Sweeps Align with Species Divergence Time?**

We can use information about selective sweeps of introgressed variation to further characterize the role of secondary gene flow in sympatric divergence. When an allele is selectively favored in a population, positive selection may cause it to increase in frequency and form a localized selective sweep of reduced genetic variation surrounding the adaptive variant.<sup>[104]</sup> Such regions of high differentiation in recently diverged species are often targeted as candidates for speciation genes, although other processes not directly associated with speciation can lead to similar patterns of high heterogeneity in differentiation across a genome.<sup>[29,105,106]</sup> If speciation was recent or ongoing, there may be strong signatures of a selective sweep for particular haplotypes in at least one of the sister species for regions involved in the divergence process (e.g., regions containing selective sweeps overlap regions of strong divergence; Figure 1b). If secondary gene flow was neutral with respect to speciation, we may find no signatures of selective



sweeps in those introgressed regions. However, care should be taken with any significance thresholds used for calling regions candidates for adaptive introgression, ideally thresholds resulting from genetic simulations. For example, false positives overlapping among all three categories are possible depending on the frequency of regions that are strongly differentiated, experienced a selective sweep, or introgressed, even if secondary gene flow was neutral.

Importantly, a sweep of the same introgressed region in both sympatric sister species may be interpreted as adaptation to the same new environment, which may not contribute to reproductive isolation between the pair (dependent on their respective genetic backgrounds<sup>[107,108]</sup>). However, this pattern is also consistent with the sweep of a region contributing to a “one-allele” mechanism of mate choice,<sup>[7,42,43]</sup> such as increased female choosiness in both sympatric sister species, which would contribute to reproductive isolation.<sup>[109]</sup> Thus, selective sweeps of an introgressed region in both sympatric sister species do not rule out its role in aiding the speciation process.

Alternatively, if selective sweeps are detected, the timing of selective sweeps can give indirect evidence about their role in speciation. If the timing of introgression predates the timing of the selective sweep, it is challenging to infer the importance of an introgressed region for speciation because LD among loci relevant to speciation may take time to build up. However, the absence of selective sweeps or occurrence of introgression long after species divergence would suggest that introgression was not relevant to speciation.

### 5.3. Is There Support for a Causal Role of Secondary Gene Flow Based on Functional Genetic Analyses of Variants in the Region?

Another potential source of evidence for the functional importance of gene flow can come from GWAS between variants in introgressed regions and traits involved in ecological or sexual isolation between sister species. The conservation of sequences within introgressed regions across taxa may also provide strong evidence of a functional role (e.g., PhastCons<sup>[102]</sup>). However, many complex traits are driven by a large number of variants of small effect and ruling out a functional role for gene flow from gene annotations is difficult (e.g., see the omnigenic model<sup>[110]</sup>). Finally, and most powerfully, genome editing and gene expression reporter systems are increasingly tractable in nonmodel systems.<sup>[59,111]</sup> This is ultimately an asymmetric problem: finding evidence that an introgressed region may have contributed to reproductive isolation is easier than demonstrating that no introgressed regions contributed to reproductive isolation in any way.<sup>[56]</sup>

### 5.4. Are There Similar Patterns of Selection or Divergence in the Introgressed Regions in Closely Related Outgroup Populations?

A thorough investigation of these same regions in outgroups to the sympatric species gives added power to distinguish whether secondary gene flow aided sympatric divergence. If nondiversifying,

closely related species exist in similar environments and have not diversified in a similar manner but share signatures of selective sweeps in the same regions, then the observed introgression may have been neutral relative to speciation, e.g., due to adaptations to shared changes in climate, pathogens, or shared regions of reduced recombination or increased background selection. Similarly, several studies comparing genomic landscapes of differentiation across closely related taxa have found that high differentiation observed in the same genomic regions across taxa reflects the action of linked selection across low-recombination regions rather than selection against gene flow at barrier loci.<sup>[112–115]</sup>

## 6. Conclusions and Outlook

Sympatric speciation remains among the most controversial evolutionary processes, beloved by theorists and long sought after by empiricists. While evidence of divergence under the biogeographic definition of sympatry is mounting using traditional genetic criteria of monophyly,<sup>[2]</sup> genomic data have now revealed the pervasiveness of secondary gene flow and introgression in many of these examples. Future fine-scale investigations of introgression will likely continue to paint a complex picture of the role of secondary gene flow in speciation. Establishing or ruling out a role for secondary gene flow in speciation and discerning which putative cases studies evolved through an “easy” or a “hard” process of sympatric speciation in the wild will be a formidable task, yet a worthwhile one in its revelation of the sheer power of divergent selection to create species in nature.

Nearly all existing case studies of sympatric speciation involve some form of automatic magic trait, such as assortative mating by habitat,<sup>[81,116,117]</sup> along a depth gradient,<sup>[49]</sup> or environment-induced phenology shifts.<sup>[118]</sup> We think that an outstanding remaining question is whether the hard process of sympatric speciation occurs in nature without the aid of some form of magic trait, as originally demonstrated to be possible in theory.<sup>[8]</sup> The highly polygenic and multidimensional nature of adaptation and mate choice suggests that an “all-of-the-above” speciation scenario containing a mix of preference/trait, magic trait, and phenotype matching (in which each trait is affected by a wide distribution of allelic effect sizes with varying times of arrival) will be the norm in nature. In contrast, although numerous and diverse, most speciation models continue to address these mechanisms in a piecemeal fashion with an assumption of large-effect alleles. It remains unclear how different mechanisms, effect sizes, and times of arrival will interact and compete within a single model.

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## Conflict of Interest

The authors declare no conflict of interest.

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- [1] E. Mayr, *Animal Species and Evolution*, Belknap, Cambridge, MA **1963**.
- [2] D. I. Bolnick, B. M. Fitzpatrick, *Annu. Rev. Ecol. Evol. Syst.* **2007**, *38*, 459.
- [3] B. M. Fitzpatrick, J. A. Fordyce, S. Gavrilets, *J. Evol. Biol.* **2008**, *21*, 1452.
- [4] B. M. Fitzpatrick, J. A. Fordyce, S. Gavrilets, *J. Evol. Biol.* **2009**, *22*, 2342.
- [5] S. Via, *Trends Ecol. Evol.* **2001**, *16*, 381.
- [6] J. A. Coyne, A. Orr, *Speciation*, Sutherland, MA **2004**.
- [7] J. Felsenstein, *Evolution* **1981**, *35*, 124.
- [8] U. Dieckmann, M. Doebeli, *Nature* **1999**, *400*, 354.
- [9] S. Gavrilets, *Fitness Landscape and Origin of Species*, Princeton University Press, Princeton, NJ **2004**.
- [10] M. Doebeli, U. Dieckmann, J. A. Metz, D. Tautz, *Evolution* **2005**, *59*, 691.
- [11] J. Mallet, A. Meyer, P. Nosil, J. L. Feder, *J. Evol. Biol.* **2009**, *22*, 2332.
- [12] A. D. Foote, *Trends Ecol. Evol.* **2018**, *33*, 85.
- [13] S. P. Otto, M. R. Servedio, S. L. Nuismer, *Genetics* **2008**, *179*, 2091.
- [14] F. J. Weissing, P. Edelaar, G. S. vanDoorn, *Behav. Ecol. Sociobiol.* **2011**, *65*, 461.
- [15] G. S. vanDoorn, F. J. Weissing, *Selection* **2002**, *2*, 17.
- [16] J. W. Poelstra, E. J. Richards, C. H. Martin, *Mol. Ecol.* **2018**, *27*, 4270.
- [17] M. Servedio, R. Bürger, *Genes* **2018**, *9*, 217. <https://doi.org/10.3390/genes9040217>
- [18] M. Kirkpatrick, N. Barton, *Genetics* **2006**, *173*, 419.
- [19] S. Yeaman, S. Aeschbacher, R. Bürger, *Mol. Ecol.* **2016**, *25*, 2542.
- [20] J. Polechová, N. H. Barton, S. Gavrilets, *Evolution* **2005**, *59*, 1194.
- [21] D. I. Bolnick, *J. Theor. Biol.* **2006**, *241*, 734.
- [22] M. Kirkpatrick, V. Ravnigne, *Am. Nat.* **2002**, *159*, S22.
- [23] O. Cotto, M. R. Servedio, *Am. Nat.* **2017**, *190*, 680.
- [24] H. Sachdeva, N. H. Barton, *Evolution* **2017**, *71*, 1478.
- [25] J. A. Coyne, A. H. Orr, *Evolution* **1989**, *43*, 362.
- [26] P. Norvaišas, E. Kisd, *Am. Nat.* **2012**, *180*, 388.
- [27] M. Kopp, J. Hermisson, *J. Evol. Biol.* **2008**, *21*, 1005.
- [28] P. Nosil, D. Schluter, *Trends Ecol. Evol.* **2011**, *26*, 160.
- [29] M. Ravinet, R. Faria, R. K. Butlin, J. Galindo, N. Bierne, M. Rafajlović, M. A. F. Noor, B. Mehlig, A. M. Westram, *J. Evol. Biol.* **2017**, *30*, 1450.
- [30] E. J. Richards, C. H. Martin, *PLoS Genet.* **2017**, *13*, e1006919.
- [31] C. Matessi, A. Gimelfarb, S. Gavrilets, *Selection* **2002**, *2*, 41.
- [32] D. I. Bolnick, M. Doebeli, *Evolution* **2003**, *57*, 2433.
- [33] R. Bürger, K. A. Schneider, *Am. Nat.* **2006**, *167*, 190.
- [34] M. Turelli, N. H. Barton, J. A. Coyne, *Trends Ecol. Evol.* **2001**, *16*, 330.
- [35] A. G. Garner, B. E. Goulet, M. C. Farnitano, Y. F. Molina-Henao, R. Hopkins, *Genes* **2018**, *9*. <https://doi.org/10.3390/genes9040191>
- [36] O. Seehausen, *Trends Ecol. Evol.* **2004**, *19*, 198.
- [37] O. Seehausen, *J. Evol. Biol.* **2013**, *26*, 279.
- [38] D. A. Turissini, D. R. Matute, *PLoS Genet.* **2017**, *13*, e1006971.
- [39] C. H. Martin, *Am. Nat.* **2012**, *180*, E90.
- [40] J. I. Meier, D. A. Marques, S. Mwaiko, C. E. Wagner, L. Excoffier, O. Seehausen, *Nat. Commun.* **2017**, *8*, 14363.
- [41] C. E. Wagner, A. R. McCune, *Evolution* **2009**, *63*, 1312.
- [42] M. Kopp, M. R. Servedio, T. C. Mendelson, R. J. Safran, R. L. Rodríguez, M. E. Hauber, E. C. Scordato, L. B. Symes, C. N. Balakrishnan, D. M. Zonana, G. S. vanDoorn, *Am. Nat.* **2018**, *191*, 1.
- [43] M. R. Servedio, J. W. Boughman, *Annu. Rev. Ecol. Evol. Syst.* **2017**, *48*, 85.
- [44] M. R. Servedio, G. S. V. Doorn, M. Kopp, A. M. Frame, P. Nosil, *Trends Ecol. Evol.* **2011**, *26*, 389.
- [45] J. L. Feder, P. Nosil, *Evolution* **2009**, *63*, 3061.
- [46] Z. L. Fuller, C. J. Leonard, R. E. Young, S. W. Schaeffer, N. Phadnis, *PLoS Genet.* **2018**, *14*, e1007526.
- [47] S. Gavrilets, *J. Hered.* **2014**, *105*, 743.
- [48] S. Gavrilets, *Evolution* **2003**, *57*, 2197.
- [49] M. Malinsky, R. J. Challis, A. M. Tyers, S. Schiffels, Y. Terai, B. P. Ngatunga, E. A. Miska, R. Durbin, M. J. Genner, G. F. Turner, *Science* **2015**, *350*, 1493.
- [50] O. Seehausen, *Proc. R. Soc. Biol. Sci.* **2006**, *273*, 1987.
- [51] J. S. Sparks, *Mol. Phylogenet. Evol.* **2004**, *30*, 599.
- [52] U. K. Schliwen, D. Tautz, S. Pääbo, *Nature* **1994**, *368*, 629.
- [53] A. F. Kautt, G. Machado-Schiaffino, A. Meyer, *PLoS Genet.* **2016**, *12*, e1006157.
- [54] G. Machado-Schiaffino, A. F. Kautt, H. Kusche, A. Meyer, *BMC Evol. Biol.* **2015**, *15*, 15. <https://doi.org/10.1186/s12862-015-0287-3>
- [55] C. E. Wagner, L. J. Harmon, O. Seehausen, *Ecol. Lett.* **2014**, *17*, 583.
- [56] E. J. Richards, J. W. Poelstra, C. H. Martin, *Evol. Lett.* **2018**, *2*, 524.
- [57] K. R. Elmer, T. K. Lehtonen, A. F. Kautt, C. Harrod, A. Meyer, *BMC Biol.* **2010**, *8*, 8. <https://doi.org/10.1186/1741-7007-8-60>
- [58] C. H. Martin, *Evolution* **2013**, *67*, 2114.
- [59] C. F. Kratochwil, M. M. Sefton, Y. Liang, A. Meyer, *BMC Dev. Biol.* **2017**, *17*, 15.
- [60] C. F. Kratochwil, M. M. Sefton, A. Meyer, *BMC Dev. Biol.* **2015**, *15*, 12.
- [61] M. Barluenga, K. N. Stölting, W. Salzburger, M. Muschick, A. Meyer, *Nature* **2006**, *439*, 719.
- [62] M. W. Hahn, L. Nakhleh, *Evolution* **2016**, *70*, 7.
- [63] C. I. Wu, *J. Evol. Biol.* **2001**, *14*, 851.
- [64] J. A. McGirr, C. H. Martin, *Mol. Biol. Evol.* **2016**, *34*, 873.
- [65] M. C. Fontaine, J. B. Pease, A. Steele, R. M. Waterhouse, D. E. Neafsey, I. V. Sharakhov, X. Jiang, A. B. Hall, F. Catteruccia, E. Kakani, S. N. Mitchell, Y. C. Wu, H. A. Smith, R. R. Love, M. K. Lawniczak, M. A. Slotman, S. J. Emrich, M. W. Hahn, N. J. Besansky, *Science* **2015**, *347*, 1258522.
- [66] C. R. Campbell, J. W. Poelstra, A. D. Yoder, *Biol. J. Linn. Soc.* **2018**, *124*, 561.
- [67] M. F. Geiger, J. K. McCrary, U. K. Schliwen, *PLoS One* **2013**, *8*, e74901.
- [68] J. Igea, D. Bogarín, A. S. T. Papadopoulos, V. Savolainen, *Evolution* **2015**, *69*, 482.
- [69] C. H. Martin, J. S. Cutler, J. P. Friel, C. Denning Touokong, G. Coop, P. C. Wainwright, *Evolution* **2015**, *69*, 1406.
- [70] K. R. Elmer, S. Fan, H. Kusche, M. Luise Spreitzer, A. F. Kautt, P. Franchini, A. Meyer, *Nat. Commun.* **2014**, *5*, 5168.
- [71] M. Barluenga, A. Meyer, *BMC Evol. Biol.* **2010**, *10*, 326.
- [72] C.-I. Wu, C.-T. Ting, *Nat. Rev. Genet.* **2004**, *5*, 114.
- [73] C. H. Martin, *Evolution* **2016**, *70*, 2504.
- [74] P. W. Hedrick, *Mol. Ecol.* **2013**, *22*, 4606.
- [75] E. Huerta-Sánchez, X. Jin, Asan, Z. Bianba, B. M. Peter, N. Vinckenbosch, Y. Liang, X. Yi, M. He, M. Somel, P. Ni, B. Wang, X. Ou, Huasang, J. Luosang, Z. X. P. Cuo, K. Li, G. Gao, Y. Yin, W. Wang, X. Zhang, X. Xu, H. Yang, Y. Li, J. Wang, J. Wang, R. Nielsen, *Nature* **2014**, *512*, 194.

- [76] S. Stankowski, M. A. Streisfeld, *Proc. R. Soc. London, Ser. B* **2015**, 282, 20151666.
- [77] L. H. Rieseberg, M. A. Archer, R. K. Wayne, *Heredity* **1999**, 83, 363.
- [78] K. Kagawa, G. Takimoto, *Ecol. Lett.* **2017**, 21, 264.
- [79] M. Schumer, G. G. Rosenthal, P. Andolfatto, *Evolution* **2014**, 68, 1553.
- [80] D. A. Marques, J. I. Meier, O. Seehausen, *Trends Ecol. Evol.* **2019**, <https://doi.org/10.1016/j.tree.2019.02.008>
- [81] J. L. Feder, S. H. Berlocher, J. B. Roethele, H. Dambroski, J. J. Smith, W. L. Perry, V. Gavrilovic, K. E. Filchak, J. Rull, M. Aluja, *Proc. Natl. Acad. Sci. U. S. A.* **2003**, 100, 10314.
- [82] L. Fishman, A. Stathos, P. M. Beardsley, C. F. Williams, J. P. Hill, *Evolution* **2013**, 67, 2547.
- [83] R. F. Guerrero, M. W. Hahn, *Mol. Ecol.* **2017**, 26, 5362.
- [84] T. C. Nelson, W. A. Cresko, *Evol. Lett.* **2018**, 2, 9.
- [85] L. Excoffier, I. Dupanloup, E. Huerta-Sánchez, V. C. Sousa, M. Foll, *PLoS Genet.* **2013**, 9, e1003905.
- [86] R. N. Gutenkunst, R. D. Hernandez, S. H. Williamson, C. D. Bustamante, *PLoS Genet.* **2009**, 5, e1000695.
- [87] P.-R. Loh, M. Lipson, N. Patterson, P. Moorjani, J. K. Pickrell, D. Reich, B. Berger, *Genetics* **2013**, 193, 1233.
- [88] B. Vernot, J. M. Akey, *Science* **2014**, 343, 1017.
- [89] I. Gronau, M. J. Hubisz, B. Gulko, C. G. Danko, A. Siepel, *Nat. Genet.* **2011**, 43, 1031.
- [90] B. C. Haller, P. W. Messer, *Mol. Biol. Evol.* **2019**, 36, 632.
- [91] A. D. Kern, D. R. Schrider, *G3: Genes, Genomes, Genet.* **2018**, 8, 1959.
- [92] D. R. Schrider, F. K. Mendes, M. W. Hahn, A. D. Kern, *Genetics* **2015**, 200, 267.
- [93] D. R. Schrider, J. Ayroles, D. R. Matute, A. D. Kern, *PLoS Genet.* **2018**, 14, e1007341.
- [94] D. R. Schrider, A. D. Kern, *PLoS Genet.* **2016**, 12, e1005928.
- [95] R. R. Hudson, *J. Mol. Evol.* **2007**, 64, 702.
- [96] G. Coop, K. Bullaughey, F. Luca, M. Przeworski, *Mol. Biol. Evol.* **2008**, 25, 1257.
- [97] S. Belezza, A. M. Santos, B. McEvoy, I. Alves, C. Martinho, E. Cameron, M. D. Shriver, E. J. Parra, J. Rocha, *Mol. Biol. Evol.* **2013**, 30, 24.
- [98] S. Nakagome, G. Alkorta-Aranburu, R. Amato, B. Howie, B. M. Peter, R. R. Hudson, A. Di Rienzo, *Mol. Biol. Evol.* **2016**, 33, 657.
- [99] L. Ormond, M. Foll, G. B. Ewing, S. P. Pfeifer, J. D. Jensen, *Mol. Ecol.* **2016**, 25, 157.
- [100] H. Chen, J. Hey, M. Slatkin, *Theor. Popul. Biol.* **2015**, 99, 18.
- [101] J. Smith, G. Coop, M. Stephens, J. Novembre, *Mol. Biol. Evol.* **2018**, 35, 1003.
- [102] A. Siepel, G. Bejerano, J. S. Pedersen, A. S. Hinrichs, M. Hou, K. Rosenbloom, H. Clawson, J. Spieth, L. D. W. Hillier, S. Richards, G. M. Weinstock, R. K. Wilson, R. A. Gibbs, W. J. Kent, W. Miller, D. Haussler, *Genome Res.* **2005**, 15, 1034.
- [103] S. Chatterjee, N. Ahituv, *Annu. Rev. Genomics Hum. Genet.* **2017**, 18, 45.
- [104] J. M. Smith, J. Haigh, *Genet. Res.* **1974**, 23, 23.
- [105] M. W. Nachman, B. A. Payseur, *Philos. Trans. R. Soc., B* **2012**, 367, 409.
- [106] T. E. Cruickshank, M. W. Hahn, *Mol. Ecol.* **2014**, 23, 3133.
- [107] K. N. Stölting, M. Paris, C. Meier, B. Heinze, S. Castiglione, D. Bartha, C. Lexer, *New Phytol.* **2015**, 207, 723.
- [108] C. S. Clarkson, D. Weetman, J. Essandoh, A. E. Yawson, G. Maslen, M. Manske, S. G. Field, M. Webster, T. Antão, B. MacInnis, D. Kwiatkowski, M. J. Donnelly, *Nat. Commun.* **2014**, 5, 4248.
- [109] D. Ortíz-Barrientos, M. A. F. Noor, *Science* **2005**, 310, 1467.
- [110] E. A. Boyle, Y. I. Li, J. K. Pritchard, *Cell* **2017**, 169, 1177.
- [111] P. A. Cleves, J. C. Hart, R. M. Agoglia, M. T. Jimenez, P. A. Erickson, L. Gai, C. T. Miller, *PLoS Genet.* **2018**, 14, e1007449.
- [112] N. Vijay, M. Weissensteiner, R. Burri, T. Kawakami, H. Ellegren, J. B. W. Wolf, *Mol. Ecol.* **2017**, 26, 4284.
- [113] B. M. VanDoren, L. Campagna, B. Helm, J. C. Illera, I. J. Lovette, M. Liedvogel, *Mol. Ecol.* **2017**, 26, 3982.
- [114] T. Ma, K. Wang, Q. Hu, Z. Xi, D. Wan, Q. Wang, J. Feng, D. Jiang, H. Ahani, R. J. Abbott, M. Lascoux, E. Nevo, J. Liu, *Proc. Natl. Acad. Sci. U. S. A.* **2018**, 115, E236.
- [115] K. E. Delmore, J. S. Lugo Ramos, B. M. VanDoren, M. Lundberg, S. Bensch, D. E. Irwin, M. Liedvogel, *Evol. Lett.* **2018**, 2, 76.
- [116] G. L. Bush, *Annu. Rev. Ecol. Syst.* **1975**, 6, 339.
- [117] M. D. Sorenson, K. M. Sefc, R. B. Payne, *Nature* **2003**, 424, 928.
- [118] V. Savolainen, M.-C. Anstett, C. Lexer, I. Hutton, J. J. Clarkson, M. V. Norup, M. P. Powell, D. Springate, N. Salamin, W. J. Baker, *Nature* **2006**, 441, 210.