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Effects of hyperoxia and hypoxia on the physiological traits responsible for obstructive sleep apnoea

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Key points
- Changes in the level of inspired oxygen have dramatic effects on the pathophysiology of obstructive sleep apnoea (OSA): hyperoxia reduces the severity of OSA in some but not all patients, whereas hypoxia transforms obstructive events into central events. Given that OSA is likely to result from the interaction of key pathophysiological traits, including a compromised pharyngeal anatomy, inadequate upper airway muscle function, a large ventilatory response to a disturbance in ventilation (high loop gain) and a low arousal threshold, we examined how changes in oxygen levels alter these traits.
- Our study demonstrates that the beneficial effect of hyperoxia on OSA severity is solely based on its ability to attenuate loop gain, whereas hypoxia increases loop gain and the arousal threshold in addition to improving pharyngeal collapsibility.
- Such effects help to explain why oxygen therapy may not work in every patient with OSA and explain the disappearance of OSA and the emergence of central events during hypoxic conditions.

Abstract Oxygen therapy is known to reduce loop gain (LG) in patients with obstructive sleep apnoea (OSA), yet its effects on the other traits responsible for OSA remain unknown. Therefore, we assessed how hyperoxia and hypoxia alter four physiological traits in OSA patients. Eleven OSA subjects underwent a night of polysomnography during which the physiological traits were measured using multiple 3-min ‘drops’ from therapeutic continuous positive airway pressure (CPAP) levels. LG was defined as the ratio of the ventilatory overshoot to the preceding reduction in ventilation. Pharyngeal collapsibility was quantified as the ventilation at CPAP of 0 cmH₂O. Upper airway responsiveness was defined as the ratio of the increase in ventilation to the increase in ventilatory drive across the drop. Arousal threshold was estimated as the level of ventilatory drive associated with arousal. On separate nights, subjects were submitted to hyperoxia (n = 9; F₁O₂ ~0.5) or hypoxia (n = 10; F₁O₂ ~0.15) and the four traits were reassessed. Hyperoxia lowered LG from a median of 3.4 (interquartile range (IQR): 2.6–4.1) to 2.1 (IQR: 1.3–2.5; P < 0.01), but did not alter the remaining traits. By contrast, hypoxia increased LG [median: 3.3 (IQR: 2.3–4.0) vs. 6.4 (IQR: 4.5–9.7); P < 0.005]. Hypoxia additionally increased the arousal threshold (mean ± s.d. 10.9 ± 2.1 l min⁻¹ vs. 13.3 ± 4.3 l min⁻¹; P < 0.05) and improved pharyngeal collapsibility (mean ± s.d. 3.4 ± 1.4 l min⁻¹ vs. 4.9 ± 1.3 l min⁻¹; P < 0.05), but did not alter upper airway responsiveness (P = 0.7). This study demonstrates that the beneficial effect of hyperoxia on the severity of OSA is primarily based on its ability to reduce LG. The effects of hypoxia described above may explain the disappearance of OSA and the emergence of central sleep apnoea in conditions such as high altitude.
Introduction

The pathophysiology of obstructive sleep apnoea (OSA) is multi-factorial. Several key factors, known as physiological ‘traits’, have been shown to combine to cause OSA. These include: (i) poor upper airway anatomy that predisposes the airway to collapse; (ii) poor ability of the upper airway muscles to respond to a respiratory challenge and stiffen or dilate the airway; (iii) a low respiratory arousal threshold that causes an individual to arouse from sleep for very small increases in respiratory drive, and (iv) a hypersensitive ventilatory control system often referred to as a system with a high loop gain (LG) (Gold et al. 1985; Wellman et al. 2011).

Over the years, many investigators have examined the use of supplemental oxygen therapy as a therapy for OSA. However, the effects of supplemental oxygen on the severity of OSA and its consequences are highly variable (Wellman et al. 2008; Mehta et al. 2013; Xie et al. 2013). Small physiological studies indicate that oxygen therapy significantly improves the apnoea–hypopnoea index (AHI) in 36–50% of individuals, whereas OSA severity remains unchanged or worsens in other patients. For those patients in whom supplemental oxygen is beneficial, it is likely that it improves OSA by reducing the sensitivity of the ventilatory control system (i.e. by decreasing LG) (Wellman et al. 2008; Xie et al. 2013). However, like any drug, oxygen may have other important physiological effects. Although oxygen may be able to reduce the sensitivity of the ventilatory control system, the reduction in ventilatory drive may have the unwanted effect of reducing the respiratory output to the upper airway muscles (Alekando, 2004), which could potentially increase upper airway collapsibility and reduce pharyngeal dilator muscle responsiveness. Such a worsening of these traits may explain why a proportion of OSA patients do not improve or actually worsen.

By contrast, exposure to hypoxaemia, such as that which may occur at altitude or in heart failure, has been clinically observed to change OSA to central sleep apnoea (CSA) (Warner et al. 1987; Burgess et al. 2004, 2006; Patz et al. 2006; Nussbaumer-Ochsner et al. 2010), which suggests that hypoxaemia may improve the upper airway anatomy or responsiveness in addition to elevating LG. It is well documented that hypoxia will raise LG (Khoo et al. 1982; Solin et al. 2000; Sands et al. 2011; Andrews et al. 2012) and that a high LG amplifies small disturbances in ventilation, yielding cyclic oscillations in ventilatory drive, as seen in CSA. However, in addition to raising LG, the conversion of OSA to CSA suggests that hypoxia might also improve the pharyngeal anatomy or responsiveness through an increased drive to the upper airway muscles (Jordan et al. 2010). However, to date there has been no systematic investigation of how either hyperoxia or hypoxia alter the underlying physiology in patients with OSA. Accordingly, the aim of this study was to assess how changes in oxygen levels alter the physiological traits responsible for OSA. The preliminary results of this analysis have been published in abstract form (Edwards et al. 2013a).

Methods

Participants

Eleven patients (five male, six female) with documented OSA defined as an AHI of >10 events h⁻¹ (mean ± S.D. 49.9 ± 22.9 events h⁻¹) were recruited from the sleep clinic at the Brigham and Women’s Hospital. All subjects were currently treated with continuous positive airway pressure (CPAP) and had documented adherence of usage of >5 h night⁻¹ during the month prior to enrolment. Subjects were excluded if they had any of the following conditions: concurrent sleep disorders; renal insufficiency; neuromuscular disease; uncontrolled diabetes mellitus; CSA; heart failure; uncontrolled hypertension, or a thyroid disorder. Subjects were also screened to ensure they were not taking any medications that might alter sleep or are known to affect respiration or pharyngeal muscle control. Written informed consent was obtained before subjects were enrolled in the study, which was approved by the Partners’ Human Research Committee and conformed to the standards set by the Declaration of Helsinki.

Experimental design and protocol

All subjects underwent two or three overnight studies in our laboratory. During the initial overnight study, a baseline assessment of the four physiological traits (described below) was conducted. During the following visits, the traits were reassessed while subjects breathed 15% O₂ balance N₂ (hypoxic condition) or 50% O₂ balance N₂ (hyperoxic condition). The order in which
subjects were subjected to hyperoxia and hypoxia nights was randomized and at least 1 week was allowed to elapse between visits.

At each visit, subjects were instrumented with standard polysomnography apparatus that included equipment for electroencephalography (EEG), electro-oculography, sub-mental electromyography, electrocardiography and arterial oxygen saturation monitored at the finger. Subjects were also fitted with a nasal mask (Gel Mask; Respironics, Inc., Murrysville, PA, USA) through which measurements of mask pressure (Validyne Engineering Corp., Northridge, CA, USA), ventilatory flow (pneumotachometer model 3700A; Hans-Rudolph, Inc., Shawnee, KS, USA), tidal volume and end-tidal CO₂ (Vacumed, Vacumetrics, Inc., Ventura, CA, USA) were obtained. The mask was connected to a positive/negative pressure source (Respironics, Inc.) to enable rapid switching between CPAP levels. During the hypoxia and hyperoxia studies, the air inlet for the pressure delivery device was connected to a large respiratory balloon that was constantly filled with the appropriate gas mixture to ensure a continuous delivery of the desired gas. All signals were sampled at 125 Hz and displayed using Nihon Kohden (Tokyo, Japan) and Spike 2 (Cambridge Electronic Design Ltd, Cambridge, UK) software.

Once all the equipment was in place, subjects were asked to sleep in the supine position. Following sleep onset, the level of CPAP was titrated to eliminate all sleep-disordered breathing. When patients were asleep in stable non-rapid eye movement (nREM) sleep, the four traits [pharyngeal anatomy/collapsibility, LG, upper airway muscle responsiveness (gain) and arousal threshold] were assessed (Fig. 1). The method for measuring these traits has been described in detail previously (Wellman et al. 2011; Edwards et al. 2012, 2014) and involves 2–3 min CPAP ‘drops’ to sub-therapeutic levels during stable nREM sleep that are repeatedly performed throughout the night. Briefly, when CPAP is dropped to a sub-therapeutic level, a reduction in ventilation is caused by the partial obstruction of the airway. The ‘passive’ anatomy (i.e. pharyngeal collapsibility) is determined by plotting mask pressure versus ventilation for the second and third breaths of all the pressure drops throughout the entire night. The data are fit with a linear regression line and ventilation at zero mask pressure (\(V_0\)) is used to measure pharyngeal anatomy/collapsibility. After the initial reduction in ventilation, ventilatory drive will begin to rise as a result of the accumulation of CO₂ and this stimulus may activate the upper airway muscles in an attempt to reopen the airway and recover lost ventilation. Despite this partial recovery, ventilation often remains depressed or reduced below the eupnoeic level despite the increased levels of ventilatory drive. The level to which ventilatory drive has risen over the course of the drop can then be determined by abruptly returning CPAP to the therapeutic level and measuring the overshoot in ventilation. The steady-state LG is then measured as the ratio of this ventilatory overshoot (or response) to the net reduction or disturbance in ventilation from baseline. In order to be utilized in the calculation of LG, ventilation during the last 60 s of the drop must be significantly lower than eupnoeic ventilation on optimum CPAP and no arousals can occur during this interval. The components of LG, controller gain (ventilatory sensitivity to CO₂) and plant gain (change in end-tidal CO₂ for a corresponding change in ventilation) were also measured. Plant gain was defined as the reciprocal of the slope of the metabolic hyperbola during sleep, and controller gain as (LG)/(plant gain). The responsiveness of the upper airway muscles, which we refer to as the ‘upper airway gain’ (UAG), is measured by first calculating the difference between ventilation at the start and end of the drop, which represents how much ventilation has been recovered over the course of the drop. The ratio of this difference to the amount by which ventilation overshoots (i.e. the increase in ventilatory drive over the course of the drop) when mask pressure is returned to the holding pressure represents the ability of the airway to stiffen or dilate in response to an increase in ventilatory drive. All LG and UAG measurements were calculated from CPAP drops that did not end in arousal, and all measurements were averaged to determine a mean value for each subject.

In addition to its use in the calculation of LG and UAG, the time course of ventilation following the return to the therapeutic pressure allows a delay and a time constant to be derived (Wellman et al. 2011). Importantly, once the LG, delay and time constant are known, the time course of the rise in ventilatory drive during each drop can be determined using a dynamic model of the ventilatory control system. Briefly, the observed changes in ventilation that occur during each CPAP drop were input into the transfer function model with the known steady-state LG, time constant and delay, which computationally transformed the changes in ventilation into a ventilatory drive signal. Once ventilatory drive is calculated, the arousal threshold can be quantified from any CPAP drop during which an arousal occurred (defined as an increase of > 3 s in EEG frequency). Specifically, the arousal threshold was calculated as the level of ventilatory drive immediately preceding the arousal.

Given the importance of arousals in promoting a ventilatory overshoot (Khoo & Berry, 1996; Khoo et al. 1996) and ventilatory instability, we also examined the effects of hyperoxia and hypoxia on the magnitude and damping characteristics of the ventilatory response to spontaneous arousal (VRA). In order for a spontaneous arousal to be included in our analysis, it had to occur while the subject was on therapeutic CPAP, last 3–15 s, occur during stage 2–4 nREM sleep and be preceded and followed by ≥1 min of stable nREM sleep following...
pre-established guidelines (Jordan et al. 2004; Edwards et al. 2013b). Arousalswere discarded if a mask leak, a change in the level of CPAP or mouth expiration occurred within 60 s before or after the arousal. Breath-by-breath measurements of inspired minute ventilation ($V_I$) and end-tidal CO$_2$ ($P_{CO_2}$) were interpolated at 0.25 s intervals for 60 s prior to and 60 s following each arousal (start arousal = time zero), designated as time = 0. Ventilation was then normalized to the mean ventilation using the 60 s prior to the arousal. We calculated the size of the average ventilatory overshoot (defined as the peak ventilation within 15 s of time = 0), the size of the secondary undershoot (defined as the nadir ventilation within 45 s of time = 0) and the ratio of these two values (Fig. 1C) to provide another measure of the stability of the ventilatory control system. A large ratio indicates a more unstable system, whereas a low value indicates a more stable system.

**Statistical analysis**

In order to maximize our sample size because several participants did not complete all three conditions, the effects of hyperoxia and hypoxia on OSA traits were assessed independently using either paired $t$ tests or the signed rank test depending on whether the data were normally distributed, with Bonferroni correction for multiple comparisons (i.e. hyperoxic and hypoxic conditions). All statistical analyses were performed using SigmaPlot Version 11.0 (Systat Software, Inc., San Jose, CA, USA). A $P$-value of $\leq 0.05$ was considered to indicate statistical significance. Values are presented as means ± S.E.M. or medians [interquartile range (IQR)] as appropriate.

**Results**

The mean ± S.D. age and body mass index of our patients were 50.4 ± 5.5 years and 36.6 ± 5.7 kg m$^{-2}$, respectively. Of the 11 subjects who completed the baseline study, 10 patients provided trait measurements during hypoxia and nine provided trait measurements during hyperoxia. The effects of hyperoxia and hypoxia on resting ventilatory parameters, the therapeutic CPAP level used during the study and the numbers of CPAP drops performed to assess the traits are shown in Table 1. Compared with baseline values, hyperoxia raised mean overnight oxygen saturation and hypoxia lowered it. Minute ventilation and end-tidal CO$_2$ remained unaltered by the level of oxygen, although transient changes were observed when the patients were initially switched into hyperoxia or hypoxia. During the hypoxia night, the majority of patients ($n = 8$) developed short epochs of cyclic central apnoeas/hypopnoeas most commonly following either an arousal or the ventilatory overshoot consequent to the return of CPAP to therapeutic levels. When the traits were assessed under the different oxygen conditions, no differences emerged in the therapeutic CPAP level used, the number of CPAP drops performed on each night, or the number of CPAP drops used to obtain LG/upper airway gain measurements.

**Effects of hyperoxia on OSA traits**

Figure 2 demonstrates that hyperoxia lowered LG from a median of 3.4 (IQR: 2.6–4.1) to 2.1 (IQR: 1.3–2.5) ($P < 0.01$) as a result of a reduction in controller gain [0.47 l min$^{-1}$ mmHg$^{-1}$ (IQR: 0.30–0.60 l min$^{-1}$ mmHg$^{-1}$) vs. 0.25 l min$^{-1}$ mmHg$^{-1}$ (IQR: 0.19–0.34 l min$^{-1}$ mmHg$^{-1}$); $P < 0.01$] as plant gain remained unchanged (7.5 ± 0.5 mmHg l$^{-1}$ min$^{-1}$ vs. 7.4 ± 0.4 mmHg l$^{-1}$ min$^{-1}$; $P = $ NS). There was a trend for hyperoxia to increase the circulatory delay (6.1 ± 1.1 s vs. 11.1 ± 1.6 s; $P = 0.12$), although this difference failed to reach statistical significance. However, hyperoxia did not alter the time constant of the ventilatory overshoot (53.6 ± 8.4 s vs. 79.3 ± 17.9 s; $P = 0.6$), and nor did it alter the upper airway anatomy/collapsibility, arousal threshold or UAG (Fig. 3).

**Effects of hypoxia on OSA traits**

Sustained overnight hypoxia increased LG [3.3 (IQR: 2.3–4.0) vs. 6.4 (IQR: 4.5–9.7); $P < 0.005$] via increases in controller gain [0.42 (IQR: 0.27–0.59) vs. 0.76 (IQR: 0.60–1.41); $P < 0.005$. It also decreased the circulatory delay (6.2 ± 1.0 s vs. 2.5 ± 0.4 s; $P < 0.005$). Exposure to sustained hypoxia additionally increased the arousal threshold (10.9 ± 0.7 l min$^{-1}$ vs. 13.3 ± 1.4 l min$^{-1}$; $P < 0.05$) and improved pharyngeal collapsibility (3.4 ± 0.4 l min$^{-1}$ vs. 4.9 ± 0.4 l min$^{-1}$; $P < 0.05$), but did not alter UAG (Fig. 4).

**Effects of oxygen on VRA**

The VRA could be assessed in seven of the nine patients who completed the hyperoxia nights and in all patients who completed the hypoxia nights. Compared with baseline levels, the level of oxygen did not alter the number or duration of arousals included in the analysis (Table 1). The effects of hypoxia and hyperoxia on VRA are depicted in Fig. 5. There was no difference in the magnitude of VRA with either hypoxia or hyperoxia in comparison with baseline conditions, although there was a trend for the overshoot to decrease with hyperoxia ($P = 0.06$). Compared with baseline, hypoxia significantly increased the magnitude of the ventilatory undershoot, whereas hyperoxia reduced it. These changes resulted in hypoxia significantly increasing the ventilatory undershoot.
Effects of oxygen level on the four physiological traits

Effects of hyperoxia. In the present study, hyperoxia consistently lowered the steady-state LG as predicted by theory (Khoo et al. 1982) and has been previously demonstrated experimentally (Gautier et al. 1986; Chowdhuri et al. 2010a). Furthermore, the magnitude of the decrease in LG was driven solely by reductions in controller gain and is strikingly similar to the reductions in controller gain observed with the administration of sustained hyperoxia during sleep in healthy volunteers (Chowdhuri et al. 2010a). At first, our results seem inconsistent with those of our previous study, in which we reported that the ‘dynamic’ LG was lowered only in those individuals who had a high LG at baseline (Wellman et al. 2008). Although the steady-state and dynamic LGs are not directly comparable, if we estimate the ‘dynamic’ LG using our CPAP dial-down technique [see Wellman et al. (2011) and Edwards et al. (2012) for details], we see that the majority of subjects in the current study also had a somewhat high LG at baseline [median LG: 0.71 (IQR: 0.34–0.84)].

Although it is likely that the present study was statistically underpowered to detect a significant increase in the circulatory delay, we did observe a strong trend for this to increase with hyperoxia. An increase in the delay might occur because: (i) hyperoxia is able to blunt the fast responsive peripheral chemoreceptors and the changes in ventilation subsequently observed reflect the response of the more ‘sluggish’ central chemoreceptors, or (ii) hyperoxia has depressive effects on cardiac function: it has been shown to reduce cardiac output in patients with congestive heart failure in a dose-dependent manner.

**Table 1. Effects of oxygen therapy on resting ventilatory and sleep parameters, continuous positive airway pressure (CPAP) drops performed and number of arousals included in the ventilatory response to spontaneous arousal (VRA) analysis**

<table>
<thead>
<tr>
<th></th>
<th>Baseline (n = 11)</th>
<th>Hyperoxia (n = 9)</th>
<th>Hypoxia (n = 10)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Resting ventilatory parameters</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Minute ventilation (l min(^{-1}))</td>
<td>7.6 ± 1.1</td>
<td>7.5 ± 0.9</td>
<td>7.6 ± 0.7</td>
</tr>
<tr>
<td>End-tidal CO(_2) (mmHg)</td>
<td>39.4 ± 2.4</td>
<td>38.2 ± 1.7</td>
<td>40.0 ± 2.9</td>
</tr>
<tr>
<td>Mean overnight O(_2) saturation (%)</td>
<td>95.0 ± 1.4</td>
<td>97.3 ± 0.9(^*)</td>
<td>84.3 ± 1.8(^*)</td>
</tr>
<tr>
<td><strong>Sleep parameters</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total recording duration (min)</td>
<td>364.9 ± 59.0</td>
<td>347.9 ± 48.0</td>
<td>337.9 ± 48.0</td>
</tr>
<tr>
<td>Total sleep duration (min)</td>
<td>265.1 ± 31.5</td>
<td>229.4 ± 33.6</td>
<td>266.2 ± 57.1</td>
</tr>
<tr>
<td>nREM duration (min)</td>
<td>240.0 ± 31.2</td>
<td>176.5 ± 32.1</td>
<td>230.3 ± 58.3</td>
</tr>
<tr>
<td>Stage 1</td>
<td>65 ± 38.9</td>
<td>49.1 ± 23.2</td>
<td>50.7 ± 24.5</td>
</tr>
<tr>
<td>Stage 2</td>
<td>172.6 ± 35.1</td>
<td>176.5 ± 32.1</td>
<td>176.3 ± 39.2</td>
</tr>
<tr>
<td>Stage 3–4</td>
<td>0 (0–1.4)</td>
<td>0.5 (0–5.5)</td>
<td>0.3 (0–0.5)</td>
</tr>
<tr>
<td>REM duration (min)</td>
<td>25.1 ± 16.1</td>
<td>25.9 ± 14.4</td>
<td>36.0 ± 11.5</td>
</tr>
<tr>
<td>Sleep efficiency (%)</td>
<td>73.9 ± 11.0</td>
<td>74.8 ± 14.1</td>
<td>79.1 ± 13.5</td>
</tr>
<tr>
<td><strong>CPAP used and drops performed</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Therapeutic pressure (cmH(_2)O)</td>
<td>11.4 ± 1.9</td>
<td>10.6 ± 2.6</td>
<td>12.0 ± 2.4</td>
</tr>
<tr>
<td>Total CPAP drops (n)</td>
<td>27.6 ± 7.8</td>
<td>21.9 ± 3.6</td>
<td>16.3 ± 7.6</td>
</tr>
<tr>
<td>CPAP drops to assess LG/UAG (n)</td>
<td>4.7 ± 2.9</td>
<td>7.4 ± 3.6</td>
<td>3.9 ± 2.1</td>
</tr>
<tr>
<td><strong>VRA analysis</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Arousal number (n)</td>
<td>4.8 ± 1.6</td>
<td>4.7 ± 2.6</td>
<td>6.6 ± 2.8</td>
</tr>
<tr>
<td>Arousal duration (s)</td>
<td>6.9 ± 1.4</td>
<td>7.4 ± 1.6</td>
<td>8.3 ± 1.7</td>
</tr>
</tbody>
</table>

Values are means ± s.d. Abbreviations: LG, loop gain; nREM, non-rapid eye movement; REM, rapid eye movement; UAG, upper airway gain. *P < 0.05 compared with data for the baseline night.
Figure 1. Techniques for measuring the physiological traits in obstructive sleep apnoea and assessing the ventilatory response to spontaneous arousal

A, a schematic of the ventilatory response to a continuous positive airway pressure (CPAP) drop demonstrates how all changes in ventilation were used to assess the physiological traits. Determining pharyngeal collapsibility, loop gain and upper airway gain: the drop in CPAP causes an immediate reduction in resting ventilation ($\dot{V}_{\text{eupnoea}}$) as a result of airway narrowing. The breaths (2–3) following the reduction in CPAP were used to calculate the pharyngeal collapsibility or $\dot{V}_0$.

The inset shows how the breaths from the current drop (circled) are placed on a graph of ventilation versus mask pressure in order to calculate $\dot{V}_0$. This initial reduction in ventilation leads to an increase in respiratory drive over the course of the drop. We measure how much ventilatory drive accumulates by rapidly restoring CPAP therapy and measuring the overshoot in ventilation ($x$). The ratio of this ventilatory response or overshoot ($x$) to the net reduction in ventilation during the drop period ($y$) provides a measure of loop gain ($x/y$). A delay ($\delta$) and time constant ($\tau$) are then estimated from the dynamics of the ventilatory overshoot.

In response to the increase in drive ($x$), the subject activates the upper airway muscles and partially reopens the airway, allowing ventilation to recover slightly ($z$). The ratio of the compensatory increase in ventilation ($z$) to the increase in ventilatory drive ($x$) across the drop provides a measure of neuromuscular compensation ($z/x$), to which we refer as the upper airway gain. B, determining the arousal threshold: now that we know the LG, $\delta$ and $\tau$, a ventilatory drive signal (red line) can be calculated for each CPAP drop. In CPAP drops that cause an arousal, we quantify the arousal threshold as the level of ventilatory drive immediately preceding the arousal. C, to assess the effect of hypoxia and hyperoxia on the ventilatory response to spontaneous arousal, we calculated the ratio of the reduction in ventilation following the initial overshoot ($y$) and the magnitude of this overshoot ($x$). The solid and dashed grey lines demonstrate how a minimally and a highly underdamped system respond respectively for the same ventilatory overshoot.
(Haque et al. 1996), as well as to impair cardiac relaxation and increased left ventricle filling pressures (Mak et al. 2001). Nonetheless, an increase in circulatory delay may be a contributing factor to the longer respiratory events often observed in OSA patients receiving supplemental oxygen (Wellman et al. 2008; Mehta et al. 2013).

Importantly, our finding that hyperoxia did not alter any of the remaining traits suggests that the ability of oxygen therapy to improve OSA severity is driven primarily by its ability to reduce LG in normoxic individuals, specifically through reductions in the sensitivity of the carotid bodies (i.e. controller gain). Such a finding is consistent with results in animal studies that have shown that denervation of the carotid body either prevents the apnoea and periodic breathing consequent to transient ventilatory overshoots (Nakayama et al. 2003) or the unstable breathing caused in heart failure models (Marcus et al. 2014). The ubiquitous finding that oxygen therapy improves OSA severity in a proportion of individuals, whereas the remaining patients gain little or no benefit (Martin et al. 1982; Smith et al. 1984; Gold et al. 1985, 1986; Pokorski & Jernajczyk, 2000; Landsberg et al. 2001; Kumagai et al. 2008; Mehta et al. 2013), highlights the importance of understanding that OSA is caused by both anatomical and non-anatomical factors (Wellman et al. 2011; Eckert et al. 2013). If a patient has a highly collapsible airway, as recent data indicate that 23% of patients do (Eckert et al. 2013), then he or she will have OSA regardless of whether there are abnormalities in any of the other physiological traits (i.e. LG). In such patients, we expect that reducing LG with therapies such as oxygen or acetazolamide will be of little benefit in terms of reducing the AHI. However, if a patient’s anatomy is of the vulnerable type found in the overwhelming majority of OSA subjects (Eckert et al. 2013), then whether or not he or she has a high LG (or defects in the other non-anatomical traits) will play a large role in whether the individual will develop OSA (i.e. LG is an effect modifier), as well as how that person will respond to treatment with oxygen. Considering an elevated LG as an effect modifier helps to explain why treatments that are intended to reduce LG often improve OSA in some but not all patients, even if they do universally lower LG as observed in the current study. Firstly, the fact that OSA is not completely resolved in most patients by such therapies suggests that an elevated LG is not the only factor causing OSA. Secondly, the reason why such therapies do not work in everyone is that these previous studies were conducted in unselected patients. If we could lower LG in patients with a mild vulnerability to upper airway collapse, who represent patients in whom an elevated LG is a large contributor to the sleep disorder (Eckert et al. 2013; Wellman et al. 2004), we predict that these patients would show dramatic improvements in the severity of their OSA. Although our hypotheses need to be tested rigorously in well-designed clinical trials, we hope that these concepts will allow clinicians to move beyond the ‘one size fits all’ treatment approach of CPAP and to begin to tailor alternative therapies to the needs of individuals based on their underlying physiology (Jordan et al. 2014; Malhotra, 2014).

![Figure 2. Effects of hyperoxia and hypoxia on ventilatory control characteristics](image)
Effects of hypoxia. By contrast with hyperoxia, exposure to sustained overnight hypoxia had an interesting effect on OSA traits. As expected, hypoxia raised LG via an increase in controller gain, the magnitude of which was increased by ~80% from its baseline value. Notably, this increase is remarkably similar to the increase in controller gain (83%) observed after short periods of episodic hypoxia in healthy volunteers (Chowdhuri et al. 2010b). The improvement in pharyngeal collapsibility with hypoxia is likely to be attributable to an increase in respiratory output to the upper airway muscles providing a ‘stiffer’ and less collapsible airway. Similar improvements in upper airway collapsibility have been documented in response to sustained CO₂ exposure (Jordan et al. 2010) in OSA patients. Despite the improvement in the collapsibility of the upper airway, hypoxia did not alter the responsiveness of the upper airway muscles (i.e. upper airway gain), a finding which is consistent with those of the study by Eckert et al. (2008), which demonstrated that the activation of the genioglossus muscle (a major upper airway dilator muscle) in response to brief negative pressure pulses applied in both wake and sleep was unaltered by hypoxia. Lastly, hypoxia also raised the arousal threshold by 22% in the current study. This finding is consistent with that of a previous study in healthy participants demonstrating that hypoxia increases

Figure 3. Effects of hyperoxia on anatomy, arousal threshold and upper airway gain
Hyperoxia did not alter the passive anatomy (A), the arousal threshold (B) or the upper airway gain (C).

Figure 4. Effects of hypoxia on anatomy, arousal threshold and upper airway gain
Hypoxia significantly improved the passive anatomy (A) and increased the arousal threshold (B), but did not statistically alter the upper airway gain (C).
the respiratory arousal threshold by ~25% and the time to arousal following either resistive loading or airway occlusion (Hlavac et al. 2006). The mechanism(s) by which acute hypoxia increases the arousal threshold are unclear, but it has been proposed that hypoxia is an important neuro-inhibitory modulator that can depress respiratory afferent transmission. Taken together, these findings may help to explain the clinical observation in patients with OSA that sustained hypoxia transforms obstructive events into predominantly central events and reduces the proportion of events with arousals.

Acute sustained hypoxia during sleep not only occurs at altitude but is a key feature of many medical disorders, including congestive heart failure, chronic obstructive pulmonary disease and obesity hypoventilation syndrome, as well as moderate–severe OSA. However, the ramifications of the role of intermittent hypoxia in the pathogenesis of OSA have not been fully elucidated. Certainly, an elevated controller gain (and thus LG) in untreated OSA patients can be reversed with CPAP treatment, suggesting that an increased LG can be a consequence of OSA (Loewen et al. 2009; Salloum et al. 2009). Such a finding is consistent with the various animal studies in which exposure to intermittent hypoxia (and hypercapnia) has been shown to increase the sensitivity of the peripheral chemoreceptors. Animal studies have also shown that intermittent hypoxia may attenuate the responsiveness/recruitment of the genioglossus muscle (Edge et al. 2012), although this may be counteracted by long-term facilitation of the muscle (Tadjalli et al. 2010). Lastly, Sforza et al. (1999) reported that in OSA patients, the arousal threshold increased shortly after sleep onset, peaked between the second and third hours of the night and remained at this level for the duration of the night. Studies in sleeping neonatal animals suggest that increases in the arousal threshold can be induced by

![Figure 5. Effects of oxygen on the ventilatory response to arousal](image_url)
intermittent hypoxia (Johnston et al. 1998; Durand et al. 2004; Waters & Tinworth, 2005). However, whether or not such changes are driven by the sleep fragmentation associated with repetitive arousals from sleep or intermittent hypoxia per se in patients with OSA remains unclear.

**Effect of oxygen level on VRA**

The mechanisms that determine the magnitude of the VRA have been attributed to a combination of: (i) the sudden removal of the sleep-induced increase in upper airway resistance; (ii) a reflex ‘startle-like’ mechanism that is independent of ventilatory sensitivity during wakefulness, and (iii) the restoration of the waking chemical drive at the increased $P_{CO_2}$ level which occurs during sleep (Phillipson, 1978; Khoo et al. 1998; Horner et al. 2001). The observation that the magnitude of the VRA is similar whether chemical drive is elevated with hypoxia or depressed with hyperoxia suggests that the overshoot in ventilation following a spontaneous arousal is chemoreceptor-independent, an observation congruent with studies suggesting its magnitude is in part similar to a ‘startle-like’ response (Horner et al. 2001; Trinder et al. 2006).

The role of arousals in the pathogenesis of OSA has been widely debated in the literature. The immediate impact of arousal is to restore pharyngeal patency and waking muscle tone in an attempt to prevent large falls in oxygen level. Furthermore, in some patients frequent recurrent arousals can act to recruit upper airway muscle activity progressively, which can lead to improved airway patency and periods of stable breathing (Jordan et al. 2011). By contrast, a large ventilatory response to arousal can also promote dynamic ventilatory instability (Khoo et al. 1996), which may contribute to OSA severity in certain individuals. To date, the effects of hyperoxia and hypoxia on VRA have not been assessed. Although there were no differences in the magnitude of the initial overshoot, we did find that the magnitude of the ventilatory undershoot following arousal was greater in hypoxia and smaller in hyperoxia, which is consistent with what we would expect physiologically. The magnitude of the ventilatory undershoot following arousal is dependent not only upon how much the $P_{aCO_2}$ changes, but how close eupnoeic $P_{aCO_2}$ is from the apnoea threshold, which is a function of controller gain (Dempsey, 2004). As the increase in ventilation following spontaneous arousal remained constant among the three conditions (and thus reduction in $P_{aCO_2}$), the changes in the undershoot with oxygen reflect the differences in controller gain. Such findings suggest that the level of oxygen may be an important contributor to whether arousals promote dynamic ventilatory instability and contribute to OSA severity.

**Methodological considerations**

Although our estimates of LG using our published technique often produce values similar to those obtained with other methods (Salloum et al. 2009), we have never formally validated this measurement. One way to validate our methodology for the determination of LG is to assess whether it is sensitive to the known effects of varying oxygen levels on LG: hypoxia increases LG by increasing the slope of the hypercapnic ventilatory response (i.e. controller gain), whereas hyperoxia does the opposite (Nielsen & Smith, 1951; Khoo et al. 1982; Dahan et al. 1990; Mohan & Duffin, 1997; Xie et al. 2006; Chowdhuri et al. 2010a). The observation that our measurements of LG show the expected directional changes with different levels of oxygen gives us confidence that the method of measuring LG from disturbances in ventilation provided by CPAP drops is a robust technique (Edwards et al. 2011, 2012).

Many of the previous investigations or randomized controlled trials assessing the effects of oxygen in the treatment of OSA have delivered 100% oxygen at flow rates of 2–4 l min$^{-1}$, which corresponds to an $F_{O_2}$ of roughly 27–33% and may be appreciably lower than the 40% administered in the current study. We elected to create hyperoxic levels which would ensure the inhibition of the peripheral chemoreceptors (i.e. controller gain) similar to those we and others have used previously (Wellman et al. 2008; Chowdhuri et al. 2010a; Xie et al. 2013). Thus there is a possibility that the reduction in LG with hyperoxia may have been greater than expected during clinical studies, although we believe that this situation is unlikely to be the case for two reasons: (i) the mean overnight oxygen saturation level in the current study is similar to that seen in previous clinical investigations, and (ii) physiological studies in OSA patients who have used 40–50% oxygen (Xie et al. 2013) have shown a reduction in AHI similar to those in the numerous clinical studies (Mehta et al. 2013). By contrast with the level of hyperoxia, the level of hypoxia administered in the current study is similar to the oxygen level at an altitude of ~2700 m. This level was chosen in order to elucidate the physiology responsible for the switching of the OSA phenotype that has been previously reported to occur at this altitude (Burgess et al. 2004, 2006; Nussbaumer-Ochsner et al. 2010).

It was initially surprising that sustained hyperoxia and hypoxia seemingly had no effect on resting ventilation and end-tidal $CO_2$: The finding that we did not observe a systematic change in either ventilatory characteristic may reflect the fact that the actual changes that occur in these patients are small and, because of the large individual variability, are not captured by our small sample size (i.e. the study was insufficiently powered to detect differences in resting ventilation). However, the lack of change may actually be a real phenomenon as other small studies have
reported these ventilatory variables to remain unchanged during sustained hypoxic (Hlavac et al. 2006; Eckert et al. 2008) or hyperoxic (Xie et al. 2013) conditions.

We chose to study patients with OSA rather than assessing the effect that different levels of oxygen would have on the physiology of healthy participants (i.e. without OSA) for two reasons. Firstly, several previous investigations have already directly or indirectly assessed the effects of oxygen levels on several of the physiological traits measured in this study (using a variety of different techniques) in healthy participants and have been discussed above. Secondly, our main aim was to understand the mechanisms responsible for the hyperoxia-induced reduction in OSA severity, as well as the hypoxia-induced obstructive–central switch in patients with OSA. Therefore, we needed to study the relevant population (i.e. subjects with OSA). Our current work is limited by the fact that the complex nature of our study design did not allow us to assess how the changes in OSA traits during hyperoxia and hypoxia translate into alterations in the severity and pattern of sleep-disordered breathing. Nonetheless, the findings of the current study provide valuable information that helps to explain many of the clinically observed effects of different oxygen levels.

Conclusions

In summary, the major findings of our study highlight key alterations in the pathophysiology causing OSA in response to sustained exposure to both hyperoxia and hypoxia. Our study demonstrates that the beneficial effect of hyperoxia on OSA severity is based solely on its ability to attenuate LG, whereas hypoxia increased LG and the arousal threshold, in addition to improving pharyngeal collapsibility. Such effects help to explain why oxygen therapy may not work in all patients with OSA and account for the disappearance of OSA and the emergence of central events during hypoxic conditions.

References


### Additional information

**Competing interests**

R.L.O. consults for Respironics, Inc. A.W. is a consultant for Philips Respironics, Inc., SOVA Pharmaceuticals, Inc. and Apnex Medical, Inc. A.W.'s interests were reviewed and are managed by the Brigham and Women's Hospital and Partners HealthCare in accordance with their conflict of interest policies. A.M. was a consultant for Philips Respironics, Sleep HealthCenters, Sleep Group Solutions, Apnex Medical, Inc., Pfizer, Inc. and Apnicure, Inc., but has relinquished all outside personal income since May 2012. D.P.W. was the chief medical officer for Philips Respironics until 31 December 2012 and remains a consultant for that company. He is now the chief scientific officer for Apnicure, Inc. None of the other authors have any financial relationship with a commercial entity that has an interest in the subject of this manuscript and have no other conflicts of interest.

**Author contributions**

B.A.E., A.W., A.M., S.A.S. and D.P.W. contributed to the conception and design of the experiments. B.A.E., S.A.S., R.L.O., P.R.G., J.P.B., D.P.W., A.M. and A.W. were involved in the collection, analysis and interpretation of data. All authors contributed to the drafting or critical revision of the article for important intellectual content. All authors approved the final version of this article.

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