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SYNTHESIS & INTEGRATION

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Ecological restoration using intertidal foundation species: Considerations and potential for rockweed restoration

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Abstract

Foundation species, such as trees, corals, grasses, oysters, and rockweeds, must be common and abundant to effectively modify the physical environment and increase biodiversity by buffering environmental stress. Yet many of these important species have been declining due to disease, climate change, and other factors. A prime example is the precipitous population decline of marine rockweeds, which is attributed to increased urbanization and its accompanying impacts. Rockweeds provide three-dimensional habitat in harsh rocky intertidal environments and regulate ecosystem functioning, essential roles that no substitute species are capable of filling. Recovery of impacted rockweed populations is typically slow and unpredictable due to their limited dispersal capacity. These issues have motivated efforts to conserve remaining populations of rockweeds and reestablish or enhance depleted ones. Successfully doing so requires a robust understanding of factors that affect survival of the species and the processes that influence ecosystem structure, along with rigorous scientific testing of restoration methods and the factors that affect restoration success. In this comprehensive review, we summarize the current knowledge of rockweed ecology, highlight studies that could inform restoration practices, and recommend ways to improve our ability to implement scalable restoration of rockweeds and accompanying ecosystem-wide benefits.

KEYWORDS

climate change, foundation species, fucoids, population decline, reestablish, rocky intertidal ecosystems

INTRODUCTION

Ecosystems are often characterized by conspicuous organisms referred to as foundation species (sensu Dayton, 1972) that support biodiversity, enhance ecosystem function and

stability, and are often culturally valuable. In terrestrial ecosystems, foundation species range in scale from large canopy-forming trees to diminutive grasses. Similarly, marine habitats support a wide spectrum of species that structure communities, including kelps, mangroves,

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corals, seagrasses, mussels, and oysters. Fundamentally, the influence foundation species have on ecosystems is commensurate with their overall abundance; declines in abundance of foundation species can have cascading detrimental effects at population and community levels (Ellison et al., 2005, 2019). For example, the precipitous decline of shade-tolerant eastern hemlock trees in North America resulted in significant reductions in aquatic invertebrates as well as declines in sediment retention and productivity in streams (Degrassi et al., 2015; Ellison et al., 2005).

Rocky intertidal foundation species, such as mussels, barnacles, and seaweeds, are common in temperate regions and provide three-dimensional habitat required by other species to survive. Rockweeds, a group of conspicuous brown seaweeds (Phaeophyceae) in the order Fucales (Fucoids), are often the dominant macroalgae in temperate intertidal and shallow subtidal rocky habitats (Figure 1). Ecologically, the importance of these seaweeds in regulating ecosystem functioning and facilitating high biodiversity is well recognized, and rockweeds are increasingly being used as indicator species to assess the status of rocky intertidal ecosystems for management purposes (e.g., Maki, 1991; Murray et al., 2016; Orlando-Bonaca et al., 2008). Economically, rockweeds are valuable as commercially harvested species (Lotze et al., 2019), and they provide important ecological services supporting commercially harvested taxa, including refuge for lobsters (Schmidt et al., 2011) and nursery habitat for fish (Gullo, 2002; Vercaemer et al., 2018).

Rockweed populations, like many other species considered foundational to ecosystems, have sustained extensive degeneration and range contractions worldwide (e.g., Nicastro et al., 2013; Wernberg et al., 2011). For example, the southern range edge for the rockweed *Fucus vesiculosus* has shifted northward 1250 km from Morocco to Portugal (Nicastro et al., 2013), and the species has declined in distribution by more than 95% in the Baltic Sea (Torn et al., 2006). Globally, the causes of population declines are not well known, can result from complex interactions of multiple sources of stressors, and vary greatly among geographic locations and species. Rockweeds are known to be sensitive to both natural and anthropogenic perturbations, including severe storms (Underwood, 1998), ice scour (McCook & Chapman, 1991, 1997), pollution (Bellgrove et al., 1997), trampling (Bertocci et al., 2011), and climate change (Álvarez-Losada et al., 2020).

The decline of rockweed populations, as well as the positive effects that rockweeds have on the biological communities they inhabit, have motivated efforts to conserve remaining populations and to reestablish or enhance depleted ones (e.g., Coleman & Wernberg, 2017; Stekoll & Deysher, 1996; Whitaker, 2009; Whitaker et al., 2010). However, successful reestablishment of depleted populations requires understanding the factors that affect survival of the species along with rigorous scientific testing of outplanting methods. An understanding of community structure, including biotic interactions, is also helpful to ensure restoration is effective (Underwood, 1996). Here, we will (1) explain why rockweeds are prime subjects for restoration, (2) summarize published evidence of changes in rockweed populations, (3) identify and discuss the factors limiting population recovery, and (4) review past restoration activities and provide recommendations for future ecological restoration efforts.



FIGURE 1 Rockweeds are often the dominant seaweed in temperate rocky intertidal ecosystems. (Left) The rockweed *Silvetia compressa* in the upper intertidal zone on the Channel Islands, California (photo credit: S. Whitaker 2021). (Right) *Fucus distichus* covers most of the mid and upper intertidal gradient on Calvert Island in British Columbia, Canada (photo credit: L. Anderson 2012).

ROCKWEEDS AS FOUNDATION SPECIES

Rockweeds play a strong role in the structuring of communities via provision of habitat and modifications to the environment (Bellgrove et al., 2017). The three-dimensional rockweed canopy provides a complex matrix of microhabitats, increasing niche availability for a high diversity of taxa (e.g., Bertness et al., 1999; Fredriksen et al., 2005; Hily & Jean, 1997; Johnson & Scheibling, 1987; Schiel & Lilley, 2011). Different sets of species can be found living in the understory of the rockweed canopy, as epiphytes attached to the rockweeds, along the edge of canopy limits, and as mobile organisms freely moving through a rockweed bed (Figure 2). Rockweed canopies modify both abiotic and biotic conditions, either directly by providing shelter or reducing environmental stress or indirectly by altering biotic interactions (e.g., Bertness et al., 1999; Beermann et al., 2013; Scrosati, 2016; Scrosati & Ellrich, 2018; Watt & Scrosati, 2013a, 2013b). Alterations to the environment can have both positive and negative effects on associated species, but, generally, rockweed facilitates the presence of long-lived taxa over opportunistic species, thereby increasing community stability, resiliency, and diversity (e.g., Eriksson, 2007; Jenkins et al., 1999; Lilley & Schiel, 2006).

Modifications to the physical environment by rockweed canopies include a reduction in the rate of evaporation and temperature change during emersion periods

(Scrosati, 2016; Scrosati et al., 2021; Scrosati & Ellrich, 2018; Watt & Scrosati, 2013a, 2013b), light attenuation, and slowing of water flow. For instance, during warm periods at low tide, air temperature can be 5-16°C lower under the rockweed canopy compared to exposed rock surfaces (Beermann et al., 2013; Bertness et al., 1999; Brawley & Johnson, 1991). Similarly, evaporation rates underneath the canopy during low tides can be significantly lower than rates on the open rock (Brawley & Johnson, 1991, 1993; Bertness et al., 1999; Beermann et al., 2013). For example, in the high intertidal zone, Bertness et al. (1999) found that water loss during emersion was nearly 60% outside the canopy, while only \sim 5% underneath the canopy. The refuge from thermal stress and desiccation provided by rockweeds is particularly important since these are important factors affecting the ability of organisms to survive in harsh intertidal ecosystems.

Because of the canopy structure and modifications to the environment that rockweed beds provide, they harbor a biologically diverse community of epifaunal and understory species (Bertness et al., 1999; Colman, 1940; Eriksson et al., 2006; Fredriksen et al., 2005; Gunnill, 1985; Hawkins & Hartnoll, 1983; Hily & Jean, 1997; Jenkins et al., 2004; Johnson & Scheibling, 1987; Lilley & Schiel, 2006; Lubchenco, 1983; Sapper & Murray, 2003; Schiel & Lilley, 2011; Thompson et al., 1996). In England, for example, Colman (1940) identified 177 invertebrate species under the canopies provided by three species of *Fucus*.



FIGURE 2 The canopy provided by rockweed supports a diverse assemblage of rocky intertidal taxa in the California Channel Islands (photo credit: M. Ready 2022).

Silvetia beds in southern California, USA, had 47 species of algae, 20 sessile invertebrate species, and 44 mobile invertebrate species for a total of 107 taxa living under the frond canopy (Sapper & Murray, 2003). In comparison, communities associated with another well-documented ecosystem engineer, the mussel Mytilus californianus on the eastern north Pacific coast, harbor a similar level of diversity with a range of 80-120 species found within a mussel bed within a site (Kanter, 1977, 1978, 1979; Straughan & Kanter, 1977). Rockweeds can also be important for many other taxa, including macroinvertebrates and fish that use the habitat for shorter periods of time, and birds that feed on the invertebrates found within the canopy (Hamilton, 2001; Hamilton & Nudds, 2003). For example, crabs, lobsters (Phillippi et al., 2014; Schmidt et al., 2011; Vercaemer et al., 2018), and many fishes, including some of commercial value (Gagnon et al., 2019; Gullo, 2002; Mattsson, 2019; Rangeley & Kramer, 1995; Vercaemer et al., 2018), utilize rockweed beds as feeding areas or nursery habitat.

In addition to providing habitat for the organisms that live in the understory, rockweed canopies often facilitate the recruitment of a diversity of taxa (Bertness et al., 1999; Jenkins et al., 2004), including conspecifics (Bertness & Leonard, 1997; Brawley & Johnson, 1991; Moeller, 2002; Philbrick, 2004; Vadas et al., 1990; van Tamelen et al., 1997). Recruitment, a process vital to population maintenance and long-term persistence, is typically enhanced in the presence of rockweed canopy cover because desiccation and thermal stress are reduced during periods of emersion compared to bare rock surfaces (Beermann et al., 2013; Vadas et al., 2004). During periods of immersion, the frond canopy formed by rockweeds slows water flow, which allows for gametes or larvae to settle (Dayton, 1985).

Rockweeds are also large contributors to intertidal and coastal food webs. Productivity of fucoids is considerably higher than that of many other primary producers, especially in the intertidal zone (Bordeyne et al., 2015; Golléty et al., 2008; Tait et al., 2014; Tait & Schiel, 2010). For example, in France, Bordeyne et al. (2015) found that Fucus beds accounted for 77%-97% of carbon dioxide flux, with gross primary productivity being 7-9 times higher than that measured in eelgrass (Zostera noltii) communities (Ouisse et al., 2010) and up to 44 times higher than soft bottom habitats (Migné et al., 2004) or sandy beach communities (Hubas et al., 2006). This fucoid production forms the base of many food webs (Bertness & Leonard, 1997; Jenkins et al., 2004; Lubchenco, 1983; Moore, 1977), providing food for numerous grazers, including amphipods (Moore, 1977), isopods, decapods, littorine snails (Bertness & Leonard, 1997; Lubchenco, 1983), turban snails (Steinberg, 1985), and limpets (Hawkins et al., 2008;

Jenkins et al., 2004). Feeding studies confirm that grazers consume rockweeds, including *Fucus* and *Silvetia* (Kubanek et al., 2004; Steinberg, 1985). Steinberg (1985) observed that gastropods, including turban snails and periwinkle snails that commonly seek refuge in rockweed canopies, consistently preferred feeding on brown algae, including *Silvetia*, over red algal species.

Given their high productivity rates and relatively high biomass turnover, 40%–70% annually (Schmidt et al., 2011; Vadas et al., 2004), rockweeds are also major contributors to detrital pools (Bishop et al., 2010; Golléty et al., 2010; Renaud et al., 2015), providing carbon for detrital consumers. Rockweed detrital contributions are not only important within the rockweed habitat, but cross-boundary transport can export production into nearby habitats as well. For example, up to 82% of macrophyte (primarily rockweed) production was exported outside of the intertidal zone in a Canadian Bay (Vadas et al., 2004).

ROCKWEED DECLINES

Fucoids are declining locally and regionally on temperate rocky shores in the northern hemisphere (Davies et al., 2007; Strain et al., 2014; Wahl et al., 2015). There is also evidence of range contractions (Duarte et al., 2013; Nicastro et al., 2013) and localized extinctions (Duarte et al., 2013; Fales & Smith, 2022; Fernández, 2016; Lamela-Silvarrey et al., 2012; Martínez et al., 2012; Viejo et al., 2011) of rockweeds in multiple regions. Large declines in subtidal fucoids, such as *Sargassaceae*, have also been documented (reviewed by Falace et al., 2010).

In a thorough literature search on changes in populations of true rockweeds in the family Fucaceae, we found many examples of local extinctions, range shifts, and population declines (Table 1). We did multiple searches across several databases, including Web of Science, ProQuest, and Google Scholar, to find literature pertaining to long-term population declines in the family Fucaceae. Search terms included "rockweed, fucoid, Ascophyllum, Fucus, Pelvetia, Pelvetiopsis, Silvetia" and "change, declines, and population decline," among other key terms. Though we also performed targeted searches for studies reporting stability or increases in fucoid populations, fewer examples were found, which in part may be due to publication bias (Appendix S1: Table S1). Studies that fit the inclusion criteria are included in Table 1, categorized by species, region, and the hypothesized driver of change, if described. Most of the studies did not experimentally or quantitatively evaluate the cause of declines or local extinctions but rather qualitatively discussed the likely drivers or causes of change. Nevertheless, these insights may help focus future analyses and experiments.

TABLE 1 Studies observing population changes of multiple species of rockweed from the family Fucaceae from four regions and the discussed causes of change.

		Discussed causes of change				change		
Species	Type of change	I	II	III	IV	v	VI	Studies
Northeast Pacific								
Silvetia compressa	Decline		Х	Х				Barry et al. (1995), Gerrard (2005), Goodson (2003), Graham et al. (2018), Sagarin et al. (1999), Whitaker et al. (unpublished manuscript), and Widdowson (1971)
Pelvetiopsis californica	Decline; Local extinctions		Х				Х	Fales and Smith (2022) and Thom and Widdowson (1978)
Fucus distichus	Decline		Х	Х				Barry et al. (1995), Driskell et al. (2001), and Weitzman et al. (2021)
Northeast Atlantic								
Ascophyllum nodosum	Decline	Х	Х					Davies et al. (2007)
Fucus guiryi	Decline; Local extinctions; Range change		Х					Riera et al. (2015)
Fucus serratus	Decline; Range change	Х	Х					Álvarez-Losada et al. (2020), Duarte et al. (2013), Fernández (2016), Reichert and Buchholz (2006), and Fernández and Anadón (2008)
Fucus spiralis	Decline; Local extinctions	х	Х					Lamela-Silvarrey et al. (2012)
Fucus vesiculosus	Decline	Х	Х					Fernández (2016), Lamela-Silvarrey et al. (2012), and Piñeiro-Corbeira et al. (2016)
Pelvetia canaliculata	Decline; Local extinctions	Х	Х					Lamela-Silvarrey et al. (2012)
Baltic Sea								
Fucus vesiculosus	Decline			Х				Kukk and Martin (1992) and Mäkinen et al. (1984)
Mediterranean								
Fucus virsoides	Decline	х		Х	Х	Х		Battelli (2016), Munda (1982), Munda (1993), and Rindi et al. (2020)

Note: The discussed causes of change are indicated by roman numerals: I, herbivory; II, climate change; III, water quality, pollution, oil spill, sedimentation; IV, habitat loss; V, introduced species; VI, other impacts.

The causes of declines in rockweed populations varied across regions and time (Table 1). Eutrophication or decreased water quality in the Baltic Sea played a role in many subtidal rockweed declines documented in the 1980s, while climate change or extreme climate events and stressors from multiple sources were more frequently associated with declines in the 1990s and onward, particularly in the northeast Pacific and Atlantic. Most of the discussed causes of rockweed declines were anthropogenic in origin, including climate change, water quality, habitat loss, and introduced species, with the exception of increased herbivory documented in the Mediterranean, northeast Atlantic, and the Baltic Sea (Table 1). Unfortunately, very few studies have evaluated the mechanisms driving these declines, so further work is needed to disentangle the complex causes of global declines.

FACTORS AFFECTING POPULATIONS AND LIMITING RECOVERY

Fucoid functioning and fitness are influenced by numerous natural and anthropogenic stressors (e.g., Schiel & Foster, 2006; Wahl et al., 2015 and references therein). Natural abiotic stressors include low (Pearson & Davison, 1993, 1994) and high (Bell, 1993; Kübler & Davison, 1993) air temperature, decreased humidity (Brawley & Johnson, 1991), variations in nutrient supply (Korpinen et al., 2010), irradiance (Martínez et al., 2012; Schonbeck & Norton, 1980) and osmotic shock, and extremes in sea temperature, wave energy, and nutrients (Pielou, 1981; Vadas et al., 1990). Human-caused stressors include urbanization (Coleman & Wernberg, 2017), eutrophication (Bellgrove et al., 1997), trampling (Araújo et al., 2009; Bertocci et al., 2011; Denis, 2003; Irvine, 2005; Keough & Quinn, 1998; Schiel & Taylor, 1999), coastal sedimentation (Schiel et al., 2006), petroleum spills (Crowe et al., 2000), harvesting (Boaden & Dring, 1980), invasive species (Sutherland et al., 2014), and climate change (Keser et al., 2005; Sagarin et al., 1999). Individually, each of these abiotic drivers imposes serious impacts on fucoids. Collectively, the effects are likely to be synergistically detrimental (Hurd et al., 2014).

Rockweeds lack the ability to reliably disperse over long distances, which affects the ability of their populations to recover following declines. Dispersal of rockweeds is mostly limited to the movement of early life stages, including propagules and zygotes (Schiel & Foster, 2006), and is generally low, since the propagules are relatively large, released near the substratum (Dudgeon et al., 2001; Havs, 2006; Johnson & Brawley, 1998), and adhere to the substratum with a polyphenolic adhesive shortly after fertilization (Vreeland et al., 1993). For some species, the distance spores are capable of traveling is further reduced by the thick mucilage that surrounds them and slows their movement (Pearson & Brawley, 1996). Their propensity to time gamete release to periods of calm environmental conditions, such as low tide or low water motion, putatively sensed via carbon acquisition (Pearson et al., 1998), also limits dispersal in fucoids (Pearson & Brawley, 1996). Dispersal distance can vary among different rockweed taxa but is generally less than 30 m (Chapman, 1995) and more often less than 10 m (Serrão et al., 1996; Williams & Di Fiori, 1996). For example, propagules of the rockweed Ascophyllum nodosum can disperse up to 6 m from the parent source (Dudgeon et al., 2001), while Silvetia compressa recruits typically within <3 m, but mostly less than 1 m from the parent (Williams & Di Fiori, 1996). Longer distance dispersal is possible through drifting of detached reproductive fronds and is common for species in the family Sargassaceae (Fucales) that are neutrally or positively buoyant due to floating pneumatocysts (Chapman, 1995; Deysher & Norton, 1981; Hawes et al., 2017; Norton & Mathieson, 1983; Schiel, 1985). The importance of drifting as a dispersal method for other taxa, including rockweeds, has been hypothesized, but strong evidence is lacking (McKenzie & Bellgrove, 2009).

Rockweed recovery can also be inhibited or slowed due to temporal and spatial limitations in settlement and recruitment. Rockweeds typically are reproductive during relatively short periods during specific seasons, and, although they tend to be highly reproductive with very high fertilization rates (>95%; Brawley, 1992; Pearson & Brawley, 1996), early postsettlement (EPS) life stages have extremely high mortality rates (e.g., Brawley & Johnson, 1991; Chapman & Johnson, 1990; Gunnill, 1980; Johnson & Brawley, 1998; McLachlan, 1974; Moeller, 2002; Vadas et al., 1992). For example, Moeller (2002) found that EPS mortality during peak egg production was more than 99.99%. Lamote and Johnson (2008) found the density of recruits from three rockweed species to be exponentially higher under the canopy during all three years of their study, while Moeller (2002) found peak egg production for Silvetia to be markedly higher underneath the canopy than areas outside the rockweed bed. EPS mortality due to desiccation is particularly high when the rockweed canopy is limited in extent or absent (Johnson & Brawley, 1998; Lamote & Johson, 2008; Moeller, 2002). At the same time, EPS survival is reduced under the rockweed canopy due to mechanical dislodgement caused by the sweeping motion of fronds (Johnson & Brawley, 1998). For example, Johnson and Brawley (1998) observed settlement of EPS Silvetia recruits to be 1-2 orders of magnitude greater under the adult canopy, yet juveniles were most abundant outside of the canopy.

As a result of short-distance dispersal and temporal and spatial constraints on successful recruitment, recovery of fucoids is highly variable following disturbances (Bellgrove et al., 2017; Jenkins et al., 1999, 2004; Underwood, 1998, 1999). Recovery time can also be affected by numerous other factors, including the intensity and frequency of perturbations, the size of the area disturbed or proportion of the population removed, and the timing and frequency of disturbance (Bertocci et al., 2005; Farrell, 1989; Foster et al., 2003; Kim & DeWreede, 1996; Paine & Levin, 1981; Sousa, 1979, 1984). Frequent disturbances can inhibit fucoid recovery because a large population with a contiguous canopy can never form as individuals are being repetitively removed over time. The timing of the disturbance can also be important, depending on, for example, whether recovery of a disturbed area occurs during optimal environmental conditions or when fucoids are in peak reproductive periods (Kim et al., 2017; Kim & DeWreede, 1996). In the Gulf of Maine, disturbance timing has been proposed to determine whether a fucoid-dominated system can recover or shift to an alternative stable state dominated by mussels and barnacles (Petraitis & Dudgeon, 1999, but see Bertness et al., 2002). An alternative stable state can also be induced by the severity of the disturbance, such as a stable Ascophyllum-dominated

community shifting to a stable *Fucus*-dominated community when a large area of *Ascophyllum* has been manually removed (Menge et al., 2017).

Recovery times for fucoids are further complicated by alterations in biotic interactions as a consequence of the disturbance itself or due to indirect, cascading effects from the removal of the canopy-forming rockweed (e.g., Kim, 1997). For example, canopy removal and resulting decreased whiplash can result in an increase in barnacle recruitment and space occupancy, inhibiting subsequent recruitment of fucoids (Kordas & Dudgeon, 2009). In another study, removal of an A. nodosum canopy resulted in a cascade of events that inhibited the rockweed from fully recovering more than 12 years after removal (Jenkins et al., 1999, 2004). Without the protection of the Ascophyllum frond canopy from desiccation stress, understory turf-forming seaweeds disappeared, which resulted in an increase in limpets that grazed down subsequent rockweed recruits and other seaweeds within which rockweeds settle (Jenkins et al., 1999).

RESTORATION

Restoration ecology is a relatively new discipline, which may be why it is inherently expensive and associated with somewhat high uncertainty of success and feasibility. This is particularly true for marine restoration, which, in comparison to terrestrial systems, is still in its infancy (Bayraktarov et al., 2016; De Groot et al., 2013). Comparably few studies involve marine restoration (Benayas et al., 2009; Blignaut et al., 2013), even though these systems have often been heavily urbanized and degraded (Halpern et al., 2008). For example, in a sample of 224 ecological restoration studies published from 2004 to 2013, marine habitats were the least represented (~15%) relative to terrestrial and freshwater environments (Kollmann et al., 2016). Although restoration can be expensive, the cost of restoration is not necessarily correlated with success. Bayraktarov et al. (2016) found the most successful restoration efforts to be highly contingent upon ecosystem type, site selection, and techniques applied rather than cost. Despite costs and variable levels of success, restoration ecology has strong potential to be a promising tool for repairing degraded ecosystems, particularly as ecological theory evolves and is used more consistently to inform restoration practices (Peterson et al., 2003). Opportunities for practicing restoration will undoubtedly continue through legally mandated responses to impacts on natural resources (e.g., US Congress, 1980, 1990).

To date, a substantial amount of marine restoration studies has focused on a few select taxa, such as seagrasses, corals, oysters, and kelp, all of which are considered to be ecosystem engineers. Notably rare, however, are studies on restoration of other ecologically important canopy-forming macroalgal engineers, such as rockweeds. Certainly, the marked global decline of fucoid algae (Piñeiro-Corbeira et al., 2016; Vogt & Schramm, 1991; Wahl et al., 2015) and their influence on community structure have spurred an increased urgency in developing strategies to halt and reverse their loss. Thus far, fucoid restoration has predominantly focused on nonrockweed, subtidal taxa, such as Sargassum and Cystoseira (Campbell et al., 2014; La Fuente et al., 2019; Lardi et al., 2022; Perkol-Finkel et al., 2012; Yoon et al., 2014); only a handful of studies have focused on the restoration of intertidal rockweeds in the family Fucaceae (e.g., Gao et al., 2017; Jonsson et al., 2006; Kautsky et al., 2019; Stekoll & Deysher, 1996; Tronske, 2020; Whitaker et al., 2010).

To reverse losses of fucoids, a wide range of human-mediated actions have been applied with variable levels of success, including passive restoration approaches that involve removing the source of the impact. However, the causes of declines are often unclear or multifactorial (Piñeiro-Corbeira et al., 2016). In cases where the causes of fucoid losses are known, there is some support for the efficacy of passive restoration. For example, in the Baltic Sea, where large declines in subtidal rockweeds were caused by eutrophication (Kautsky et al., 1986; Torn et al., 2006; Vogt & Schramm, 1991), substantial recovery has occurred following reductions in nutrient loads (Eriksson et al., 1998; Nilsson et al., 2004). However, in other regions where water quality has improved, recovery of fucoid populations has failed to occur (Coleman et al., 2008; Díez et al., 2009; Soltan et al., 2001), potentially due to low standing stock or other discussed processes limiting rockweed resilience. In harvested rockweed populations, the implementation of sustainable management strategies, including equipment regulations, harvest limits, maximum cutting heights, and longer periods between harvests, has resulted in reduced impacts (Gendron et al., 2018; Ugarte & Sharp, 2001).

In cases where rockweeds have failed to recover naturally, active restoration techniques have been applied, consisting primarily of transplantation of individuals from the wild, cultivating outplants, constructing artificial habitat or substrate, and seeding with fertile branches or propagules (Campbell et al., 2014; Kautsky et al., 2019; La Fuente et al., 2019). Transplantation of whole individuals, or thalli, in particular, has been relatively successful (Figure 3). For example, Gao et al. (2017) attributed the recruitment of ~1000 rockweed individuals to a transplant method they applied involving the attachment of fertile rockweed thalli to polyethylene rope, which, in turn, was affixed to the substratum.



FIGURE 3 Rockweed transplant being attached via marine epoxy (photo credit: M. Ready 2022).

Experimental transplantation of S. compressa by chipping off rock pieces with rockweeds attached from donor sites and affixing the rock to the substrate at restoration sites using marine epoxy resulted in survival rates as high as ~60% (Tronske, 2020; Whitaker et al., 2010). Success was particularly high when transplants were placed on sloped surfaces protected from direct sunlight and trampling by visitors. More than a decade later, at a site where rockweed had become locally extinct for well over two decades, despite only <70 rockweed thalli being transplanted (Whitaker et al., 2010), the small population has expanded in 2022 to include well over a thousand individuals (Figure 4). More recent efforts by the authors to outplant Silvetia at several locations have shown early success with high survivorship (>75% after ~4 months) and subsequent recruitment of new individuals.

Some rockweed restoration studies have utilized seeding, construction of artificial substrates, or a combination of these techniques to stimulate or enhance recruitment of rockweeds. Whitaker (2009) collected fertile receptacles of *Silvetia* and affixed them to the substrate in mesh containers, while Stekoll and Deysher (1996) attached entire fertile *Fucus gardneri* individuals to erosion control netting and inoculated the plots with a solution of zygotes. More recently, the authors have preliminarily tested attaching fertile branches to the substrate, stimulating gamete release in the field, and outplanting of discs with germlings from laboratory gamete release. Though these studies found that seeding techniques were unsuccessful, similar approaches have been

found to be successful for other Fucales that are predominantly subtidal, such as *Cystoseira* (Verdura et al., 2018) and *Sargassum* (Yu et al., 2012). Although Stekoll and Deysher (1996) were unable to enhance recruitment via seeding erosion control netting, they observed that moisture retention facilitated by the netting may have enhanced recruitment of zygotes from the surrounding *Fucus* population. In a factorial field experiment, Whitaker et al. (2010) found that the presence of artificial canopy significantly enhanced survival of *Silvetia* transplants.

The deployment of artificial substrates or structures that mimic the positive effect of a rockweed canopy by reducing desiccation and facilitating natural recruitment warrants further exploration. Studies employing the use of artificial substrate for restoration of fucoids have primarily been conducted on subtidal species, but the techniques utilized may also be applicable to rockweed restoration. These studies have largely focused on cultivating germlings on artificial substrates in vitro, then outplanting the substrate into the wild. For instance, Chai et al. (2014) cultured Sargassum germlings in vitro on "breeding boards" that were then mounted on concrete blocks and placed at the restoration site. The germlings survived and, after one year, had grown to an average length of ~15 cm. Growth of Gongolaria germlings in laboratory conditions also showed promise for use in restoration efforts (Lardi et al., 2022), though outplanting of germlings has not been tested. A similar technique could potentially be utilized in rockweed restoration by cultivating rockweed germlings in vitro onto artificial substrates

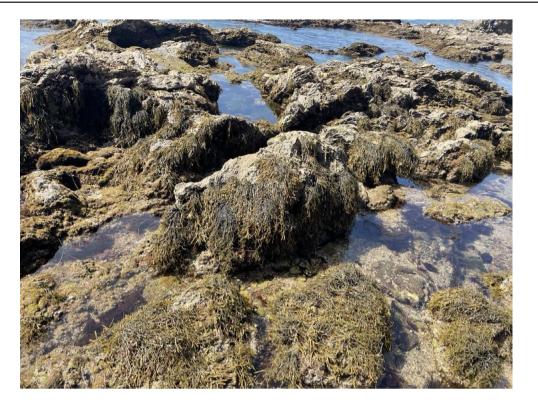


FIGURE 4 *Silvetia compressa* at Little Corona del Mar in Newport Beach, California, 14 years after restoration (photo credit: J. Smith 2021).

such as ceramic tiles or epoxy discs, which may then be affixed to the reef with bolts or marine epoxy. Early efforts by the authors show that *Silvetia* can be stimulated to release gametes with germlings settling on discs in the laboratory, but germlings on outplanted discs have not survived. Several additional studies have utilized in vitro propagation of fucoids onto artificial substrates with varying levels of success (Choi et al., 2003; Falace et al., 2006; Kautsky et al., 2019; La Fuente et al., 2019; Leung et al., 2014; Terawaki et al., 2003; Verdura et al., 2018; Yoon et al., 2014; Yatsuya, 2010, and references therein).

Continued exploration of restoration techniques for rockweeds is needed, particularly with the use of seeding and lab cultivations. Though, as previously discussed, transplanting entire thalli has been one of the more successful techniques implemented in rockweed restoration, it also has a potential impact on the donor population. This is a particularly important consideration for those species that have experienced steep population declines and have limited distribution. Seeding and outplanting techniques limit the amount of material harvested from the donor population, as fertile receptacles (Whitaker, 2009), branches (Verdura et al., 2018), or even drift material (Yatsuya, 2010) can be collected and used to propagate individuals in vitro or be placed at a restoration site to establish a source of propagules that may facilitate natural recruitment.

CONCLUSIONS AND RECOMMENDATIONS

Fucoid declines are predicted to increase in frequency and intensity in the future, with rising temperatures accompanying climate change (Kay et al., 2016; Keser et al., 2005; Rugiu et al., 2018; Takolander et al., 2017; Wilson et al., 2015). Recovery of damaged rockweed populations is slow and unpredictable (Hill, 1980; Vesco & Gillard, 1980) since dispersal is limited and early life stages experience extreme mortality (Moeller, 2002). Recovery is particularly limited when perturbations result in low abundances or local extirpations because of insufficient adult sources for recruitment and reestablishment. Assisted recovery can facilitate a return to stable populations, particularly for those populations with low abundances, whose inhibited recovery may otherwise result in continued declines and eventual extirpation. Because they are foundational species, rockweed recovery can increase the biodiversity of the entire rocky intertidal ecosystem that depends on them. For these reasons, rockweeds are optimal candidates for restoration. Although restoration in rocky intertidal habitats is in its infancy, with very few examples of tested approaches, some methods for successfully restoring rockweeds have been identified through rigorous empirical testing (Gao et al., 2017; Whitaker et al., 2010).

Scaling up rockweed restoration remains a central challenge. Although some work has been done in support of this goal (e.g., seeding and cultivation), these studies were largely unsuccessful (e.g., Stekoll & Deysher, 1996; Whitaker et al., 2010). Cultivation techniques for rockweeds have been developed (e.g., Pearson et al., 1998), but extremely high levels of mortality for early life stages of rockweed species require vast numbers of propagules or cultivation to larger sizes to enable successful restoration. More research and development are needed to effectively cultivate rockweed to stages that are resistant to mortality in large enough numbers to be used in restoration projects.

In the face of climate change, relying on historical reference states as a target for restoration may be ineffective if the changing climate is a major factor contributing to habitat degradation and species decline. Current regulatory frameworks understandably emphasize conserving essential species within their indigenous range, and the concept of assisted migration, also known as managed relocation (Richardson et al., 2009) or assisted colonization (Hällfors et al., 2014), is controversial. For restoration to be successful in a rapidly changing climate, nevertheless, the specific environmental requirements of individual species and the possible impacts of such management actions should be seriously considered (Harris et al., 2006).

Rocky intertidal ecosystems have tremendous value through the provision of significant economic benefits for tourism, fisheries, and real estate, vital ecosystem services, such as shoreline stabilization and protection, and a rich environment for education, research, and recreation. As a result, rocky shores have long had great cultural and historical significance, including for tribal nations. However, rocky intertidal ecosystems are vulnerable to impacts of human activities, such as urbanization (Vogt & Schramm, 1991), overexploitation (Harley & Rogers-Bennett, 2004), pollution (Eriksson et al., 1998, 2002; Kangas et al., 1982; Kautsky et al., 1986; Kukk & Martin, 1992; Mäkinen et al., 1984; Middelboe & Sand-Jensen, 2000; Nilsson et al., 2004; Torn et al., 2006), and climate change (e.g., Barry et al., 1995; Sagarin et al., 1999). Many rocky shore ecosystems, particularly near urban centers, are already degraded, and that trend will continue as coastal development progresses. Thus, effective and quantifiable restoration is needed to maintain habitat health and resilience as well as the valuable services that rocky intertidal ecosystems provide. Restoring key foundation species, such as rockweeds, is an efficient way to achieve broader benefits to ecosystem integrity. It is critically necessary to identify effective methods for restoring rocky intertidal communities at meaningful spatial scales to counter future large- and small-scale impacts on this ecologically important and valuable marine

ecosystem. Restoring foundational rockweeds and their associated communities is one of them and will help buffer against the impacts of climate change and other catastrophic events in the future, as well as prevent larger scale losses of intertidal communities.

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DATA AVAILABILITY STATEMENT

No data were collected for this study.

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REFERENCES

- Álvarez-Losada, Ó., J. Arrontes, B. Martínez, C. Fernández, and R. M. Viejo. 2020. "A Regime Shift in Intertidal Assemblages Triggered by Loss of Algal Canopies: A Multidecadal Survey." *Marine Environmental Research* 160: 104981. https://doi.org/ 10.1016/j.marenvres.2020.104981.
- Araújo, R., S. Vaselli, M. Almeida, E. Serrão, and I. Sousa-Pinto. 2009. "Effects of Disturbance on Marginal Populations: Human Trampling on Ascophyllum nodosum Assemblages at its Southern Distribution Limit." Marine Ecology Progress Series 378: 81–92. https://doi.org/10.3354/meps07814.
- Barry, J. P., C. H. Baxter, R. D. Sagarin, and S. E. Gilman. 1995. "Climate-Related, Long-Term Faunal Changes in a California Rocky Intertidal Community." *Science* 267(5198): 672–5. https://doi.org/10.1126/science.267.5198.672.
- Battelli, C. 2016. "Disappearance of *Fucus virsoides* J. Agardh from the Slovenian Coast (Gulf of Trieste, Northern Adriatic)." *Annales: Series Historia Naturalis* 26(1): 1–12. https://doi.org/ 10.19233/ASHN.2016.1.
- Bayraktarov, E., M. I. Saunders, S. Abdullah, M. Mills, J. Beher, H. P. Possingham, P. J. Mumby, and C. E. Lovelock. 2016.
 "The Cost and Feasibility of Marine Coastal Restoration." *Ecological Applications* 26(4): 1055–74. https://doi.org/10. 1890/15-1077.
- Beermann, A. J., J. A. Ellrich, M. Molis, and R. A. Scrosati. 2013. "Effects of Seaweed Canopies and Adult Barnacles on Barnacle Recruitment: The Interplay of Positive and Negative Influences." *Journal of Experimental Marine Biology and Ecology* 448: 162–70. https://doi.org/10.1016/j.jembe.2013. 07.001.
- Bell, E'Lise C. 1993. "Photosynthetic Response to Temperature and Desiccation of the Intertidal Alga Mastocarpus papillatus." Marine Biology 117(2): 337–46. https://doi.org/10.1007/ BF00345679.

- Bellgrove, A., M. N. Clayton, and G. P. Quinn. 1997. "Effects of Secondarily Treated Sewage Effluent on Intertidal Macroalgal Recruitment Processes." *Marine and Freshwater Research* 48(2): 137–46. https://doi.org/10.1071/mf96011.
- Bellgrove, A., P. F. McKenzie, H. Cameron, and J. B. Pocklington. 2017. "Restoring Rocky Intertidal Communities: Lessons from a Benthic Macroalgal Ecosystem Engineer." *Marine Pollution Bulletin* 117(1-2): 17–27. https://doi.org/10.1016/j.marpolbul. 2017.02.012.
- Benayas, J. M. R., A. C. Newton, A. Diaz, and J. M. Bullock. 2009. "Enhancement of Biodiversity and Ecosystem Services by Ecological Restoration: A Meta-Analysis." *Science* 325(5944): 1121–4. https://doi.org/10.1126/science.1172460.
- Bertness, M. D., and G. H. Leonard. 1997. "The Role of Positive Interactions in Communities: Lessons from Intertidal Habitats." *Ecology* 78(7): 1976–89. https://doi.org/10.1890/ 0012-9658(1997)078[1976:TROPII]2.0.CO;2.
- Bertness, M. D., G. H. Leonard, J. M. Levine, P. R. Schmidt, and A. O. Ingraham. 1999. "Testing the Relative Contribution of Positive and Negative Interactions in Rocky Intertidal Communities." *Ecology* 80(8): 2711–26. https://doi.org/10.1890/ 0012-9658(1999)080[2711:TTRCOP]2.0.CO;2.
- Bertness, M. D., G. C. Trussell, P. J. Ewanchuk, and B. R. Silliman. 2002. "Do Alternate Stable Community States Exist in the Gulf of Maine Rocky Intertidal Zone?" *Ecology* 83(12): 3434–48. https://doi.org/10.1890/0012-9658(2002)083[3434:DASCSE]2.0. CO;2.
- Bertocci, I., R. Araújo, S. Vaselli, and I. Sousa-Pinto. 2011. "Marginal Populations under Pressure: Spatial and Temporal Heterogeneity of Ascophyllum nodosum and Associated Assemblages Affected by Human Trampling in Portugal." Marine Ecology Progress Series 439: 73–82. https://doi.org/ 10.3354/meps09328.
- Bertocci, I., E. Maggi, S. Vaselli, and L. Benedetti-Cecchi. 2005. "Contrasting Effects of Mean Intensity and Temporal Variation of Disturbance on a Rocky Seashore." *Ecology* 86(8): 2061–7. https://doi.org/10.1890/04-1698.
- Bishop, M. J., M. A. Coleman, and B. P. Kelaher. 2010. "Cross-Habitat Impacts of Species Decline: Response of Estuarine Sediment Communities to Changing Detrital Resources." *Oecologia* 163(2): 517–25. https://doi.org/10.1007/s00442-009-1555-y.
- Blignaut, J., K. J. Esler, M. P. de Wit, D. Le Maitre, S. J. Milton, and J. Aronson. 2013. "Establishing the Links between Economic Development and the Restoration of Natural Capital." *Current Opinion in Environmental Sustainability* 5(1): 94–101. https:// doi.org/10.1016/j.cosust.2012.12.003.
- Boaden, P. J. S., and M. T. Dring. 1980. "A Quantitative Evaluation of the Effects of Ascophyllum Harvesting on the Littoral Ecosystem." *Helgoländer Meeresuntersuchungen* 33(1): 700–10. https://doi.org/10.1007/BF02414790.
- Bordeyne, F., A. Migné, and D. Davoult. 2015. "Metabolic Activity of Intertidal *Fucus* spp. Communities: Evidence for High Aerial Carbon Fluxes Displaying Seasonal Variability." *Marine Biology* 162(10): 2119–29. https://doi.org/10.1007/s00227-015-2741-6.
- Brawley, S. H. 1992. "Fertilization in Natural Populations of the Dioecious Brown Alga *Fucus ceranoides* and the Importance of the Polyspermy Block." *Marine Biology* 113(1): 145–57. https:// doi.org/10.1007/BF00367648.

- Brawley, S. H., and L. E. Johnson. 1991. "Survival of Fucoid Embryos in the Intertidal Zone Depends upon Developmental Stage and Microhabitat." *Journal of Phycology* 27(2): 179–86. https://doi.org/10.1111/j.0022-3646.1991.00179.x.
- Brawley, S. H., and L. E. Johnson. 1993. "Predicting Desiccation Stress in Microscopic Organisms the Use of Agarose Beads to Determine Evaporation within and between Intertidal Microhabitats." *Journal of Phycology* 29(4): 528–35. https://doi. org/10.1111/j.1529-8817.1993.tb00154.x.
- Campbell, A. H., E. M. Marzinelli, A. Vergés, M. A. Coleman, and P. D. Steinberg. 2014. "Towards Restoration of Missing Underwater Forests." *PLoS One* 9(1): e84106. https://doi.org/ 10.1371/journal.pone.0084106.
- Chai, Z., Y. Huo, Q. He, X. Huang, X. Jiang, and P. He. 2014. "Studies on Breeding of Sargassum vachellianum on Artificial Reefs in Gouqi Island, China." Aquaculture 424–425: 189–93. https://doi.org/10.1016/j.aquaculture.2013.12.038.
- Chapman, A. R. O. 1995. "Functional Ecology of Fucoid Algae: Twenty-Three Years of Progress." *Phycologia* 34(1): 1–32. https://doi.org/10.2216/i0031-8884-34-1-1.1.
- Chapman, A. R. O., and C. R. Johnson. 1990. "Disturbance and Organization of Macroalgal Assemblages in the Northwest Atlantic." *Hydrobiologia* 192(1): 77–121. https://doi.org/10.1007/ BF00006228.
- Choi, C. G., H. G. Kim, and C. H. Sohn. 2003. "Transplantation of Young Fronds of Sargassum horneri for Construction of Seaweed Beds." Korean Journal of Fisheries and Aquatic Sciences 36(5): 469–73.
- Coleman, M. A., B. P. Kelaher, P. D. Steinberg, and A. J. K. Millar. 2008. "Absence of a Large Brown Macroalga on Urbanized Rocky Reefs Around Sydney, Australia, and Evidence for Historical Decline." *Journal of Phycology* 44(4): 897–901. https://doi.org/10.1111/j.1529-8817.2008.00541.x.
- Coleman, M. A., and T. Wernberg. 2017. "Forgotten Underwater Forests: The Key Role of Fucoids on Australian Temperate Reefs." *Ecology and Evolution* 7(20): 8406–18. https://doi.org/ 10.1002/ece3.3279.
- Colman, J. 1940. "On the Faunas Inhabiting Intertidal Seaweeds." Journal of the Marine Biological Association of the United Kingdom 24(1): 129–83. https://doi.org/10.1017/ S0025315400054503.
- Crowe, T. P., R. C. Thompson, S. Bray, and S. J. Hawkins. 2000. "Impacts of Anthropogenic Stress on Rocky Intertidal Communities." *Journal of Aquatic Ecosystem Stress and Recovery* 7(4): 273–97. https://doi.org/10.1023/A:1009911928100.
- Davies, A. J., M. P. Johnson, and C. A. Maggs. 2007. "Limpet Grazing and Loss of Ascophyllum nodosum Canopies on Decadal Time Scales." Marine Ecology Progress Series 339: 131–41. https://doi.org/10.3354/meps339131.
- Dayton, P. K. 1972. "Toward an Understanding of Community Resilience and the Potential Effects of Enrichments to the Benthos at McMurdo Sound, Antarctica." In Proceedings of the Colloquium on Conservation Problems in Antarctica 81–96. Lawrence, KS: Allen Press.
- Dayton, P. K. 1985. "Ecology of Kelp Communities." Annual Review of Ecology and Systematics 16: 215–45.
- Degrassi, A., S. T. Brantley, C. R. Levine, R. Miller, J. Mohan, S. Record, and A. M. Ellison. 2015. "The Loss of Foundation

Species Revisited." *PeerJ PrePrints* 3: e1409v1. https://doi.org/ 10.7287/peerj.preprints.1409v1.

- Denis, T. G. 2003. "Effects of Human Foot Traffic on the Standing Stocks, Size Structures, and Reproduction of Southern California Populations of the Intertidal Rockweed Silvetia compressa (O. Fucales)." MS Thesis, California State University.
- Deysher, L., and T. A. Norton. 1981. "Dispersal and Colonization in Sargassum muticum (Yendo) Fensholt." Journal of Experimental Marine Biology and Ecology 56(2): 179–95. https://doi.org/10.1016/0022-0981(81)90188-X.
- Díez, I., A. Santolaria, A. Secilla, and J. M. Gorostiaga. 2009. "Recovery Stages over Long-Term Monitoring of the Intertidal Vegetation in the 'Abra de Bilbao' Area and on the Adjacent Coast (N. Spain)." *European Journal of Phycology* 44(1): 1–14. https://doi.org/10.1080/09670260802158642.
- Driskell, W. B., J. L. Ruesink, D. C. Lees, J. P. Houghton, and S. C. Lindstrom. 2001. "Long-Term Signal of Disturbance: *Fucus* gardneri After the Exxon Valdez Oil Spill." *Ecological* Applications 11(3): 815–27. https://doi.org/10.1890/1051-0761 (2001)011[0815:LTSODF]2.0.CO;2.
- Duarte, L., R. M. Viejo, B. Martínez, M. deCastro, M. Gómez-Gesteira, and T. Gallardo. 2013. "Recent and Historical Range Shifts of Two Canopy-Forming Seaweeds in North Spain and the Link with Trends in Sea Surface Temperature." Acta Oecologica 51: 1–10. https://doi.org/10.1016/j.actao.2013.05.002.
- Dudgeon, S., J. E. Kübler, W. A. Wright, R. L. Vadas, Sr., and P. S. Petraitis. 2001. "Natural Variability in Zygote Dispersal of Ascophyllum nodosum at Small Spatial Scales." Functional Ecology 15(5): 595–604. https://doi.org/10.1046/j.0269-8463.2001.00559.x.
- Ellison, A., H. Buckley, B. Case, D. Cardenas, A. Duque, J. Lutz, J. Myers, D. Orwig, and J. Zimmerman. 2019. "Species Diversity Associated with Foundation Species in Temperate and Tropical Forests." *Forests* 10(2): 128. https://doi.org/10.3390/f10020128.
- Ellison, A. M., M. S. Bank, B. D. Clinton, E. A. Colburn, K. Elliott, C. R. Ford, D. R. Foster, et al. 2005. "Loss of Foundation Species: Consequences for the Structure and Dynamics of Forested Ecosystems." *Frontiers in Ecology and the Environment* 3(9): 479–86. https://doi.org/10.1890/1540-9295 (2005)003[0479:LOFSCF]2.0.CO;2.
- Eriksson, B. K. 2007. "Dominance by a Canopy Forming Seaweed Modifies Resource and Consumer Control of Bloom-Forming Macroalgae." *Oikos* 116(7): 1211–9. https://doi.org/10.1111/j. 0030-1299.2007.15666.x.
- Eriksson, B. K., G. Johansson, and P. Snoeijs. 1998. "Long-Term Changes in the Sublittoral Zonation of Brown Algae in the Southern Bothnian Sea." *European Journal of Phycology* 33(3): 241–9. https://doi.org/10.1080/09670269810001736743.
- Eriksson, B. K., G. Johansson, and P. Snoeijs. 2002. "Long-Term Changes in the Macroalgal Vegetation of the Inner Gullmar Fjord, Swedish Skagerrak Coast." *Journal of Phycology* 38(2): 284–96. https://doi.org/10.1046/j.1529-8817.2002.00170.x.
- Eriksson, B. K., A. Rubach, and H. Hillebrand. 2006. "Biotic Habitat Complexity Controls Species Diversity and Nutrient Effects on Net Biomass Production." *Ecology* 87(1): 246–54. https://doi.org/10.1890/05-0090.
- Falace, A., G. Alongi, M. Cormaci, G. Furnari, D. Curiel, E. Cecere, and A. Petrocelli. 2010. "Changes in the Benthic Algae along the Adriatic Sea in the Last Three Decades." *Chemistry*

and Ecology 26(sup1): 77–90. https://doi.org/10.1080/ 02757541003689837.

- Falace, A., E. Zanelli, and G. Bressan. 2006. "Algal Transplantation as a Potential Tool for Artificial Reef Management and Environmental Mitigation." *Bulletin of Marine Science* 78(1): 161–6.
- Fales, R., and J. R. Smith. 2022. "Long-Term Change in a High-Intertidal Rockweed (*Pelvetiopsis californica*) and Community-Level Consequences." *Marine Biology* 169(3): 34. https://doi.org/10.1007/s00227-022-04022-1.
- Farrell, T. M. 1989. "Succession in a Rocky Intertidal Community: The Importance of Disturbance Size and Position within a Disturbed Patch." *Journal of Experimental Marine Biology and Ecology* 128(1): 57–73. https://doi.org/10.1016/0022-0981(89) 90092-0.
- Fernández, C. 2016. "Current Status and Multidecadal Biogeographical Changes in Rocky Intertidal Algal Assemblages: The Northern Spanish Coast." *Estuarine, Coastal and Shelf Science* 171: 35–40. https://doi.org/10.1016/j.ecss.2016.01.026.
- Fernández, C., and R. Anadón. 2008. "The Cantabrian Coast: A Scenario of Changes in the Distribution of Intertidal Communities." Algae 39: 30–2.
- Foster, M. S., E. W. Nigg, L. M. Kiguchi, D. D. Hardin, and J. S. Pearse. 2003. "Temporal Variation and Succession in an Algal-Dominated High Intertidal Assemblage." *Journal of Experimental Marine Biology and Ecology* 289(1): 15–39. https://doi.org/10.1016/S0022-0981(03)00035-2.
- Fredriksen, S., H. Christie, and B. A. Sæthre. 2005. "Species Richness in Macroalgae and Macrofauna Assemblages on *Fucus serratus* L. (Phaeophyceae) and *Zostera marina* L. (Angiospermae) in Skagerrak, Norway." *Marine Biology Research* 1: 2–19. https://doi.org/10.1080/17451000510018953.
- Gagnon, K., M. Gräfnings, and C. Boström. 2019. "Trophic Role of the Mesopredatory Three-Spined Stickleback in Habitats of Varying Complexity." *Journal of Experimental Marine Biology and Ecology* 510: 46–53. https://doi.org/10.1016/j.jembe.2018.10.003.
- Gao, X., H. G. Choi, S. K. Park, J. R. Lee, J. H. Kim, H. Zi-Min, and K. W. Nam. 2017. "Growth, Reproduction and Recruitment of *Silvetia siliquosa* (Fucales, Phaeophyceae) Transplants Using Polyethylene Rope and Natural Rock Methods." *Algae* 32(4): 337–47. https://doi.org/10.4490/algae.2017.32.12.6.
- Gendron, L., A. Merzouk, P. Bergeron, and L. E. Johnson. 2018. "Managing Disturbance: The Response of a Dominant Intertidal Seaweed Ascophyllum nodosum (L.) Le Jolis to Different Frequencies and Intensities of Harvesting." Journal of Applied Phycology 30(3): 1877–92. https://doi.org/10.1007/ s10811-017-1346-5.
- Gerrard, A. L. 2005. "Changes in the Rocky Intertidal Floras along the Palos Verdes Peninsula (Los Angeles County) since E.Y. Dawson's Surveys in the Late 1950s." MS Thesis, California State University.
- Golléty, C., A. Migné, and D. Davoult. 2008. "Benthic Metabolism on a Sheltered Rocky Shore: Role of the Canopy in the Carbon Budget." *Journal of Phycology* 44(5): 1146–53. https://doi.org/ 10.1111/j.1529-8817.2008.00569.x.
- Golléty, C., P. Riera, and D. Davoult. 2010. "Complexity of the Food Web Structure of the Ascophyllum nodosum Zone Evidenced by a Δ13C and Δ15N Study." Journal of Sea Research 64(3): 304–12. https://doi.org/10.1016/j.seares.2010.04.003.

- Goodson, J. 2003. "Long-Term Changes in Rocky Intertidal Populations and Communities at Little Corona Del Mar, California: A Synthesis Using Traditional and Non-Traditional Data." MS Thesis, California State University.
- Graham, S., B. Hong, S. Mutschler, B. Saunders, and J. Bredvik. 2018. "Changes in Abundance of *Silvetia compressa* at San Clemente Island before and during the 2015–2016 El Niño." *Western North American Naturalist* 78(4): 605–16. https://doi. org/10.3398/064.078.0405.
- Groot, D., S. Rudolf, J. Blignaut, S. van der Ploeg, J. Aronson, T. Elmqvist, and J. Farley. 2013. "Benefits of Investing in Ecosystem Restoration." *Conservation Biology* 27(6): 1286–93. https://doi.org/10.1111/cobi.12158.
- Gullo, A. M. 2002. "The Value of Rockweed (Ascophyllum nodosum) as Habitat for Tidepool Fishes." MS Thesis, University of Maine.
- Gunnill, F. C. 1980. "Demography of the Intertidal Brown Alga Pelvetia fastigiata in Southern California, USA." Marine Biology 59(3): 169–79. https://doi.org/10.1007/BF00396865.
- Gunnill, F. C. 1985. "Growth, Morphology, and Microherbivore Faunas of *Pelvetia fastigiata* (Phaeophyta, Fucaceae) at La Jolla, CA, USA." *Botanica Marina* 28(5): 187–99. https://doi. org/10.1515/botm.1985.28.5.187.
- Hällfors, M. H., E. M. Vaara, M. Hyvärinen, M. Oksanen, L. E. Schulman, H. Siipi, and S. Lehvävirta. 2014. "Coming to Terms with the Concept of Moving Species Threatened by Climate Change – A Systematic Review of the Terminology and Definitions." *PLoS One* 9(7): e102979. https://doi.org/10. 1371/journal.pone.0102979.
- Halpern, B. S., S. Walbridge, K. A. Selkoe, C. V. Kappel, F. Micheli, C. D'Agrosa, J. F. Bruno, et al. 2008. "A Global Map of Human Impact on Marine Ecosystems." *Science* 319(5865): 948–52. https://doi.org/10.1126/science.1149345.
- Hamilton, D. J. 2001. "Feeding Behavior of Common Eider Ducklings in Relation to Availability of Rockweed Habitat and Duckling Age." Waterbirds: The International Journal of Waterbird Biology 24(2): 233–41. https://doi.org/10.2307/1522035.
- Hamilton, D. J., and T. D. Nudds. 2003. "Effects of Predation by Common Eiders (*Somateria mollissima*) in an Intertidal Rockweed Bed Relative to an Adjacent Mussel Bed." *Marine Biology* 142(1): 1–12. https://doi.org/10.1007/s00227-002-0935-1.
- Harley, C. D. G., and L. Rogers-Bennett. 2004. "The Potential Synergistic Effects of Climate Change and Fishing Pressure on Exploited Invertebrates on Rocky Intertidal Shores." *CalCOFI Report* 45: 98–110.
- Harris, J. A., R. J. Hobbs, E. Higgs, and J. Aronson. 2006. "Ecological Restoration and Global Climate Change." *Restoration Ecology* 14(2): 170–6. https://doi.org/10.1111/j. 1526-100X.2006.00136.x.
- Hawes, N. A., D. I. Taylor, and D. R. Schiel. 2017. "Transport of Drifting Fucoid Algae: Nearshore Transport and Potential for Long Distance Dispersal." *Journal of Experimental Marine Biology and Ecology* 490: 34–41. https://doi.org/10.1016/j. jembe.2017.02.001.
- Hawkins, S. J., and R. G. Hartnoll. 1983. "Changes in a Rocky Shore Community: An Evaluation of Monitoring." *Marine Environmental Research* 9(3): 131–81. https://doi.org/10.1016/ 0141-1136(83)90051-X.

- Hawkins, S. J., P. J. Moore, M. T. Burrows, E. Poloczanska, N. Mieszkowska, R. J. H. Herbert, S. R. Jenkins, R. C. Thompson, M. J. Genner, and A. J. Southward. 2008.
 "Complex Interactions in a Rapidly Changing World: Responses of Rocky Shore Communities to Recent Climate Change." *Climate Research* 37(2–3): 123–33. https://doi.org/10. 3354/cr00768.
- Hays, C. G. 2006. "Ecological Consequences of Dispersal and Gene Flow in an Intertidal Alga." PhD diss., University of California. https://search.proquest.com/openview/7b21bf85fd 97f3062f17b1cbc0c22305/1?pq-origsite=gscholar&cbl=18750& diss=y.
- Hill, M. L. 1980. "Structure, Organization, and Persistence of the *Pelvetia fastigiata* (Phaeophyceae, Fucales) Community on a Rocky Intertidal Shoreline at Dana Point, Orange County, California." MA Thesis, California State University.
- Hily, C., and F. Jean. 1997. "Macrobenthic Biodiversity in Intertidal Habitats of the Iroise Biosphere Reserve (Brittany, France)." *Journal of the Marine Biological Association of the UK* 77(2): 311–23. https://doi.org/10.1017/S0025315400071691.
- Hubas, C., D. Davoult, T. Cariou, and L. F. Artigas. 2006. "Factors Controlling Benthic Metabolism during Low Tide along a Granulometric Gradient in an Intertidal Bay (Roscoff Aber Bay, France)." *Marine Ecology Progress Series* 316: 53–68. https://doi.org/10.3354/meps316053.
- Hurd, C. L., P. J. Harrison, K. Bischof, and C. S. Lobban. 2014. Seaweed Ecology and Physiology. Cambridge: Cambridge University Press.
- Irvine, K. 2005. "Influence of Trampling Intensity versus Hydration State on Loss of Biomass from the Intertidal Rockweed, *Fucus* gardneri." Coastal Management 33(4): 471–81. https://doi.org/ 10.1080/08920750500218003.
- Jenkins, S. R., S. J. Hawkins, and T. A. Norton. 1999. "Interaction between a Fucoid Canopy and Limpet Grazing in Structuring a Low Shore Intertidal Community." *Journal of Experimental Marine Biology and Ecology* 233(1): 41–63. https://doi.org/10. 1016/S0022-0981(98)00128-2.
- Jenkins, S. R., T. A. Norton, and S. J. Hawkins. 2004. "Long Term Effects of Ascophyllum nodosum Canopy Removal on Mid Shore Community Structure." Journal of the Marine Biological Association of the United Kingdom 84(2): 327–9. https://doi. org/10.1017/S0025315404009221h.
- Johnson, L. E., and S. H. Brawley. 1998. "Dispersal and Recruitment of a Canopy-Forming Intertidal Alga: The Relative Roles of Propagule Availability and Post-Settlement Processes." *Oecologia* 117(4): 517–26. https://doi.org/10.1007/s004420050688.
- Johnson, S. C., and R. E. Scheibling. 1987. "Structure and Dynamics of Epifaunal Assemblages on Intertidal Macroalgae Ascophyllum nodosum and Fucus vesiculosus in Nova Scotia, Canada." Marine Ecology Progress Series 37(2/3): 209–27 https://www.jstor.org/stable/24824696.
- Jonsson, P. R., L. Granhag, P. S. Moschella, P. Åberg, S. J. Hawkins, and R. C. Thompson. 2006. "Interactions between Wave Action and Grazing Control the Distribution of Intertidal Macroalgae." *Ecology* 87(5): 1169–78. https://doi.org/10.1890/ 0012-9658(2006)87[1169:IBWAAG]2.0.CO;2.
- Kangas, P., H. Autio, G. Hällfors, H. Luther, Å. Nieme, and H. Salemaa. 1982. "A General Model of the Decline of *Fucus*

vesiculosus at Tvärminne, South Coast of Finland in 1977-81." Acta Botanica Fennica 118: 1–27.

- Kanter, R. G. 1977. "Structure and Diversity in *Mytilus californianus* (Mollusca: Bivalvia) Communities." PhD diss., University of Southern California.
- Kanter, R. G. 1978. Intertidal Study of the Southern California Bight.1.2: Mussel Communities, Vol III. Los Angeles, CA: U.S. Bureau of Land Management, Department of the Interior.
- Kanter, R. G. 1979. Intertidal Study of the Southern California Bight.7.0: Mussel Community Studies, Vol II. Los Angeles, CA: U.S.Bureau of Land Management, Department of the Interior.
- Kautsky, N., H. Kautsky, U. Kautsky, and M. Waern. 1986. "Decreased Depth Penetration of *Fucus vesiculosus* (L.) since the 1940's Indicates Eutrophication of the Baltic Sea." *Marine Ecology Progress Series* 28: 1–8.
- Kautsky, L., S. Qvarfordt, and E. Schagerström. 2019. "Fucus vesiculosus Adapted to a Life in the Baltic Sea: Impacts on Recruitment, Growth, Re-Establishment and Restoration." Botanica Marina 62(1): 17–30. https://doi.org/10.1515/bot-2018-0026.
- Kay, L. M., A. L. Schmidt, K. L. Wilson, and H. K. Lotze. 2016. "Interactive Effects of Increasing Temperature and Nutrient Loading on the Habitat-Forming Rockweed Ascophyllum nodosum." Aquatic Botany 133: 70–8. https://doi.org/10.1016/j. aquabot.2016.06.002.
- Keough, M. J., and G. P. Quinn. 1998. "Effects of Periodic Disturbances from Trampling on Rocky Intertidal Algal Beds." *Ecological Applications* 8(1): 141–61. https://doi.org/10.1890/ 1051-0761(1998)008[0141:EOPDFT]2.0.CO;2.
- Keser, M., J. T. Swenarton, and J. F. Foertch. 2005. "Effects of Thermal Input and Climate Change on Growth of Ascophyllum nodosum (Fucales, Phaeophyceae) in Eastern Long Island Sound (USA)." Journal of Sea Research 54(3): 211–20. https://doi.org/10.1016/j.seares.2005.05.001.
- Kim, J. H. 1997. "The Role of Herbivory, and Direct and Indirect Interactions, in Algal Succession." *Journal of Experimental Marine Biology and Ecology* 217(1): 119–35. https://doi.org/10. 1016/S0022-0981(97)00054-3.
- Kim, J. H., and R. E. DeWreede. 1996. "Effects of Size and Season of Disturbance on Algal Patch Recovery in a Rocky Intertidal Community." *Marine Ecology Progress Series* 133(1/3): 217–28. https://doi.org/10.3354/meps133217.
- Kim, H. H., Y. W. Ko, K. M. Yang, G. Sung, and J. H. Kim. 2017.
 "Effects of Disturbance Timing on Community Recovery in an Intertidal Habitat of a Korean Rocky Shore." *Algae* 32(4): 325–36. https://doi.org/10.4490/algae.2017.32.12.7.
- Kollmann, J., S. T. Meyer, R. Bateman, T. Conradi, M. M. Gossner, M. de Souza, J. Mendonça, et al. 2016. "Integrating Ecosystem Functions into Restoration Ecology—Recent Advances and Future Directions." *Restoration Ecology* 24(6): 722–30. https:// doi.org/10.1111/rec.12422.
- Kordas, R. L., and S. Dudgeon. 2009. "Modeling Variation in Interaction Strength between Barnacles and Fucoids." *Oecologia* 158(4): 717–31. https://doi.org/10.1007/s00442-008-1183-y.
- Korpinen, S., V. Jormalainen, and E. Pettay. 2010. "Nutrient Availability Modifies Species Abundance and Community Structure of *Fucus*-Associated Littoral Benthic Fauna."

Marine Environmental Research 70(3-4): 283-92. https://doi. org/10.1016/j.marenvres.2010.05.010.

- Kubanek, J., S. E. Lester, W. Fenical, and M. E. Hay. 2004. "Ambiguous Role of Phlorotannins as Chemical Defenses in the Brown Alga *Fucus vesiculosus.*" *Marine Ecology Progress Series* 277: 79–93. https://doi.org/10.3354/meps277079.
- Kübler, J. E., and I. R. Davison. 1993. "High-Temperature Tolerance of Photosynthesis in the Red Alga *Chondrus crispus.*" *Marine Biology* 117(2): 327–35. https://doi.org/10.1007/BF00345678.
- Kukk, H., and G. Martin. 1992. "Long-Term Dynamics of the Phytobenthos in Paernu Bay, the Baltic Sea." *Proceedings of* the Estonian Academy of Sciences, Ecology 2(3): 110–8.
- La Fuente, D., M. C. Gina, V. Asnaghi, S. Kaleb, and A. Falace. 2019. "First Ex Situ Outplanting of the Habitat-Forming Seaweed *Cystoseira amentacea Var. stricta* from a Restoration Perspective." *PeerJ* 7: e7290. https://doi.org/10.7717/peerj.7290.
- Lamela-Silvarrey, C., C. Fernández, R. Anadón, and J. Arrontes. 2012. "Fucoid Assemblages on the North Coast of Spain: Past and Present (1977 - 2007)." *Botanica Marina* 55(3): 199–207. https://doi.org/10.1515/bot-2011-0081.
- Lamote, M., and L. E. Johnson. 2008. "Temporal and Spatial Variation in the Early Recruitment of Fucoid Algae: The Role of Microhabitats and Temporal Scales." *Marine Ecology Progress Series* 368: 93–102. https://doi.org/10.3354/ meps07592.
- Lardi, P. I., I. Varkitzi, K. Tsiamis, S. Orfanidis, D. Koutsoubas, A. Falace, and M. Salomidi. 2022. "Early Development of Gongolaria Montagnei (Fucales, Phaeophyta) Germlings under Laboratory Conditions, with a View to Enhancing Restoration Potential in the Eastern Mediterranean." *Botanica Marina* 65(4): 279–87. https://doi.org/10.1515/bot-2021-0105.
- Leung, Y. H., C. W. Yeung, and P. O. Ang. 2014. "Assessing the Potential for Recovery of a Sargassum siliquastrum Community in Hong Kong." Journal of Applied Phycology 26(2): 1097–106. https://doi.org/10.1007/s10811-013-0097-1.
- Lilley, S. A., and D. R. Schiel. 2006. "Community Effects Following the Deletion of a Habitat-Forming Alga from Rocky Marine Shores." *Oecologia* 148(4): 672–81. https://doi.org/10.1007/ s00442-006-0411-6.
- Lotze, H. K., I. Milewski, J. Fast, L. Kay, and B. Worm. 2019. "Ecosystem-Based Management of Seaweed Harvesting." *Botanica Marina* 62(5): 395–409. https://doi.org/10.1515/bot-2019-0027.
- Lubchenco, J. 1983. "Littornia and Fucus: Effects of Herbivores, Substratum Heterogeneity, and Plant Escapes During Succession." Ecology 64(5): 1116–23. https://doi.org/10.2307/1937822.
- Maki, A. W. 1991. "The Exxon Valdez Oil Spill: Initial Environmental Impact Assessment. Part 2." Environmental Science and Technology 25(1): 24–9. https://doi.org/10.1021/es00013a001.
- Mäkinen, A., I. Haahtela, H. Ilvessalo, J. Lehto, and O. Rönnberg. 1984. "Changes in the Littoral Rocky Shore Vegetation of the Seili Area, SW Archipelago of Finland." *Ophelia* 3: 157–66.
- Martínez, B., F. Arenas, S. Marcos Rubal, R. E. Burgués, J. I. García-Plazaola, F. L. Figueroa, R. Pereira, L. Saldaña, I. Sousa-Pinto, and A. Trilla. 2012. "Physical Factors Driving Intertidal Macroalgae Distribution: Physiological Stress of a Dominant Fucoid at Its Southern Limit." *Oecologia* 170(2): 341–53. https://doi.org/10.1007/s00442-012-2324-x.

- Mattsson, E. 2019. "Importance of *Fucus vesiculosus* (Bladderwrack) for Coastal Fish Communities in the Baltic Sea." MS Thesis, Stockholm University.
- McCook, L. J., and A. R. O. Chapman. 1991. "Community Succession Following Massive Ice-Scour on an Exposed Rocky Shore: Effects of Fucus Canopy Algae and of Mussels during Late Succession." *Journal of Experimental Marine Biology and Ecology* 154(2): 137–69. https://doi.org/10.1016/0022-0981(91) 90161-O.
- McCook, L. J., and A. R. O. Chapman. 1997. "Patterns and Variations in Natural Succession Following Massive Ice-Scour of a Rocky Intertidal Seashore." *Journal of Experimental Marine Biology and Ecology* 214(1): 121–47. https://doi.org/10. 1016/S0022-0981(96)02751-7.
- McKenzie, P. F., and A. Bellgrove. 2009. "Dislodgment and Attachment Strength of the Intertidal Macroalga Hormosira banksii (Fucales, Phaeophyceae)." Phycologia 48(5): 335–43. https://doi.org/10.2216/08-96.1.
- McLachlan, J. 1974. "Effects of Temperature and Light on Growth and Development of Embryos of *Fucus edentatus* and *F. distichus* ssp. *distichus.*" *Canadian Journal of Botany* 52(5): 943–51. https://doi.org/10.1139/b74-121.
- Menge, B. A., M. E. S. Bracken, J. Lubchenco, and H. M. Leslie. 2017. "Alternative State? Experimentally Induced Fucus Canopy Persists 38 Yr in an Ascophyllum-Dominated Community." Ecosphere 8(3): e01725. https://doi.org/10.1002/ ecs2.1725.
- Middelboe, A. L., and K. Sand-Jensen. 2000. "Long-Term Changes in Macroalgal Communities in a Danish Estuary." *Phycologia* 39(3): 245–57. https://doi.org/10.2216/i0031-8884-39-3-245.1.
- Migné, A., N. Spilmont, and D. Davoult. 2004. "In Situ Measurements of Benthic Primary Production During Emersion: Seasonal Variations and Annual Production in the Bay of Somme (Eastern English Channel, France)." Continental Shelf Research 24(13): 1437–49. https://doi.org/10.1016/j.csr.2004.06.002.
- Moeller, J. M. 2002. "Seasonal and Spatial Patterns of Reproduction and Recruitment in a Population of the Intertidal Rockweed *Silvetia compressa* (Fucales)." MS Thesis, California State University.
- Moore, P. G. 1977. "Organization in Simple Communities: Observations on the Natural History of Hyale nilssoni (Amphipoda) in High Littoral Seaweeds." In Biology of Benthic Organisms, edited by B. F. Keegan, P. O. Ceidigh, and P. J. S. Boaden, 443–51. Oxford: Pergamon Press. https://doi.org/10. 1016/B978-0-08-021378-1.50050-6.
- Munda, I. M. 1982. "The Effect of Organic Pollution on the Distribution of Fucoid Algae from the Istrian Coast (Vicinity of Rovinj)." Acta Adriatica 23(1/2): 329–37.
- Munda, I. M. 1993. "Changes and Degradation of Seaweed Stands in the Northern Adriatic." *Hydrobiologia* 260/261(1): 239–53. https://doi.org/10.1007/BF00049025.
- Murray, S. N., S. B. Weisberg, P. T. Raimondi, R. F. Ambrose, C. A. Bell, C. A. Blanchette, J. L. Burnaford, et al. 2016. "Evaluating Ecological States of Rocky Intertidal Communities: A Best Professional Judgment Exercise." *Ecological Indicators* 60: 802–14. https://doi.org/10.1016/j.ecolind.2015.08.017.
- Nicastro, K. R., G. I. Zardi, S. Teixeira, J. Neiva, E. A. Serrão, and G. A. Pearson. 2013. "Shift Happens: Trailing Edge Contraction Associated with Recent Warming Trends

Threatens a Distinct Genetic Lineage in the Marine Macroalga *Fucus vesiculosus.*" *BMC Biology* 11(1): 6. https://doi.org/10.1186/1741-7007-11-6.

- Nilsson, J., R. Engkvist, and L.-E. Persson. 2004. "Long-Term Decline and Recent Recovery of Fucus Populations along the Rocky Shores of Southeast Sweden, Baltic Sea." Aquatic Ecology 38: 587–98. https://doi.org/10.1007/s10452-004-5665-7.
- Norton, T. A., and A. C. Mathieson. 1983. "The Biology of Unattached Seaweeds." In *Progress in Phycological Research*, edited by F. Round and D. Chapman, 333–86. Amsterdam: Elsevier. http://oceanrep.geomar.de/44479/.
- Orlando-Bonaca, M., L. Lipej, and S. Orfanidis. 2008. "Benthic Macrophytes as a Tool for Delineating, Monitoring and Assessing Ecological Status: The Case of Slovenian Coastal Waters." *Marine Pollution Bulletin* 56(4): 666–76. https://doi. org/10.1016/j.marpolbul.2007.12.018.
- Ouisse, V., A. Migné, and D. Davoult. 2010. "Seasonal Variations of Community Production, Respiration and Biomass of Different Primary Producers in an Intertidal Zostera noltii Bed (Western English Channel, France)." Hydrobiologia 649(1): 3–11. https://doi.org/10.1007/s10750-010-0254-3.
- Paine, R. T., and S. A. Levin. 1981. "Intertidal Landscapes: Disturbance and the Dynamics of Pattern." *Ecological Monographs* 51(2): 145–78. https://doi.org/10.2307/2937261.
- Pearson, G. A., and S. H. Brawley. 1996. "Reproductive Ecology of Fucus distichus (Phaeophyceae): An Intertidal Alga with Successful External Fertilization." Marine Ecology Progress Series 143: 211–23. https://doi.org/10.3354/meps143211.
- Pearson, G. A., and I. R. Davison. 1993. "Freezing Rate and Duration Determine the Physiological Response of Intertidal Fucoids to Freezing." *Marine Biology* 115(3): 353–62. https:// doi.org/10.1007/BF00349832.
- Pearson, G. A., and I. R. Davison. 1994. "Freezing Stress and Osmotic Dehydration in *Fucus distichus* (Phaeophyta): Evidence for Physiological Similarity." *Journal of Phycology* 30(2): 257–67. https://doi.org/10.1111/j.0022-3646.1994.00257.x.
- Pearson, G. A., E. A. Serrão, and S. H. Brawley. 1998. "Control of Gamete Release in Fucoid Algae: Sensing Hydrodynamic Conditions Via Carbon Acquisition." *Ecology* 79(5): 1725–39. https://doi.org/10.1890/0012-9658(1998)079[1725:COGRIF]2.0. CO;2.
- Perkol-Finkel, S., F. Ferrario, V. Nicotera, and L. Airoldi. 2012. "Conservation Challenges in Urban Seascapes: Promoting the Growth of Threatened Species on Coastal Infrastructures." *Journal of Applied Ecology* 49(6): 1457–66. https://doi.org/10. 1111/j.1365-2664.2012.02204.x.
- Peterson, C. H., J. H. Grabowski, and S. P. Powers. 2003. "Estimated Enhancement of Fish Production Resulting from Restoring Oyster Reef Habitat: Quantitative Valuation." *Marine Ecology Progress Series* 264: 249–64. https://doi.org/10. 3354/meps264249.
- Petraitis, P. S., and S. R. Dudgeon. 1999. "Experimental Evidence for the Origin of Alternative Communities on Rocky Intertidal Shores." *Oikos* 84(2): 239–45. https://doi.org/10.2307/3546718.
- Philbrick, P. K. B. 2004. "Factors Restricting Recruitment of Ascophyllum nodosum L (Le Jolis)." PhD diss., University of New Hampshire.
- Phillippi, A., K. Tran, and A. Perna. 2014. "Does Intertidal Canopy Removal of *Ascophyllum nodosum* Alter the Community

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Structure Beneath?" Journal of Experimental Marine Biology and Ecology 461: 53–60. https://doi.org/10.1016/j.jembe.2014.07.018.

- Pielou, E. C. 1981. "Rapid Estimation of the Standing Crop of Intertidal Fucoids on an Exposed Shore." Journal of Environmental Management 13(1): 82–94.
- Piñeiro-Corbeira, C., R. Barreiro, and J. Cremades. 2016. "Decadal Changes in the Distribution of Common Intertidal Seaweeds in Galicia (NW Iberia)." *Marine Environmental Research* 113: 106–15. https://doi.org/10.1016/j.marenvres.2015.11.012.
- Rangeley, R. W., and D. L. Kramer. 1995. "Use of Rocky Intertidal Habitats by Juvenile Pollock Pollachius virens." Marine Ecology Progress Series 126: 9–17. https://doi.org/10.3354/meps126009.
- Reichert, K., and F. Buchholz. 2006. "Changes in the Macrozoobenthos of the Intertidal Zone at Helgoland (German Bight, North Sea): A Survey of 1984 Repeated in 2002." *Helgoland Marine Research* 60(3): 213–23. https://doi. org/10.1007/s10152-006-0037-4.
- Renaud, P. E., T. S. Løkken, L. L. Jørgensen, J. Berge, and B. J. Johnson. 2015. "Macroalgal Detritus and Food-Web Subsidies along an Arctic Fjord Depth-Gradient." *Frontiers in Marine Science* 2: 31. https://doi.org/10.3389/fmars.2015.00031.
- Richardson, D. M., J. J. Hellmann, J. S. McLachlan, D. F. Sax, M. W. Schwartz, E. Patrick Gonzalez, J. Brennan, et al. 2009. "Multidimensional Evaluation of Managed Relocation." Proceedings of the National Academy of Sciences of the United States of America 106(24): 9721–4. https://doi.org/10.1073/ pnas.0902327106.
- Riera, R., C. Sangil, and M. Sansón. 2015. "Long-Term Herbarium Data Reveal the Decline of a Temperate-Water Algae at Its Southern Range." *Estuarine, Coastal and Shelf Science* 165: 159–65. https://doi.org/10.1016/j.ecss.2015.05.008.
- Rindi, F., B. Gavio, P. Díaz-Tapia, C. G. Di Camillo, and T. Romagnoli. 2020. "Long-Term Changes in the Benthic Macroalgal Flora of a Coastal Area Affected by Urban Impacts (Conero Riviera, Mediterranean Sea)." *Biodiversity and Conservation* 29: 2275–95. https://doi.org/10.1007/s10531-020-01973-z.
- Rugiu, L., I. Manninen, E. Rothäusler, and V. Jormalainen. 2018. "Tolerance and Potential for Adaptation of a Baltic Sea Rockweed under Predicted Climate Change Conditions." *Marine Environmental Research* 134: 76–84. https://doi.org/10. 1016/j.marenvres.2017.12.016.
- Sagarin, R. D., J. P. Barry, S. E. Gilman, and C. H. Baxter. 1999.
 "Climate-Related Change in an Intertidal Community over Short and Long Time Scales." *Ecological Monographs* 69(4): 465–90. https://doi.org/10.1890/0012-9615(1999)069[0465: CRCIAI]2.0.CO;2.
- Sapper, S. A., and S. N. Murray. 2003. "Variation in Structure of the Subcanopy Assemblage Associated with Southern California Populations of the Intertidal Rockweed Silvetia Compressa (Fucales)." *Pacific Science* 57(4): 433–62. https://doi.org/10.1353/ psc.2003.0035.
- Schiel, D. R. 1985. "A Short-Term Demographic Study of Cystoseira osmundacea (Fucales: Cystoseiraceae) in Central California." Journal of Phycology 21(1): 99–106. https://doi.org/10.1111/j. 0022-3646.1985.00099.x.
- Schiel, D. R., and M. S. Foster. 2006. "The Population Biology of Large Brown Seaweeds: Ecological Consequences of Multiphase Life Histories in Dynamic Coastal Environments." *Annual Review of*

Ecology, Evolution, and Systematics 37(1): 343-72. https://doi. org/10.1146/annurev.ecolsys.37.091305.110251.

- Schiel, D. R., and S. A. Lilley. 2011. "Impacts and Negative Feedbacks in Community Recovery over Eight Years Following Removal of Habitat-Forming Macroalgae." *Journal* of Experimental Marine Biology and Ecology 407(1): 108–15. https://doi.org/10.1016/j.jembe.2011.07.004.
- Schiel, D. R., and D. I. Taylor. 1999. "Effects of Trampling on a Rocky Intertidal Algal Assemblage in Southern New Zealand." Journal of Experimental Marine Biology and Ecology 235(2): 213–35. https://doi.org/10.1016/S0022-0981(98)00170-1.
- Schiel, D. R., S. A. Wood, R. A. Dunmore, and D. I. Taylor. 2006. "Sediment on Rocky Intertidal Reefs: Effects on Early Post-Settlement Stages of Habitat-Forming Seaweeds." *Journal* of Experimental Marine Biology and Ecology 331(2): 158–72. https://doi.org/10.1016/j.jembe.2005.10.015.
- Schmidt, A. L., M. Coll, T. N. Romanuk, and H. K. Lotze. 2011. "Ecosystem Structure and Services in Eelgrass Zostera marina and Rockweed Ascophyllum nodosum Habitats." Marine Ecology Progress Series 437: 51–68. https://doi.org/10.3354/ meps09276.
- Schonbeck, M. W., and T. A. Norton. 1980. "Factors Controlling the Lower Limits of Fucoid Algae on the Shore." Journal of Experimental Marine Biology and Ecology 43(2): 131–50. https://doi.org/10.1016/0022-0981(80)90021-0.
- Scrosati, R. A. 2016. "Community-Level Facilitation by Macroalgal Foundation Species Peaks at an Intermediate Level of Environmental Stress." *bioRxiv*: 091777. https://doi.org/10. 1101/091777.
- Scrosati, R. A., A. M. Catalán, and N. Valdivia. 2021. "Macroalgal Canopies Reduce Beta Diversity in Intertidal Communities." *Botanica Marina* 64(5): 419–25. https://doi.org/10.1515/bot-2021-0023.
- Scrosati, R. A., and J. A. Ellrich. 2018. "Thermal Moderation of the Intertidal Zone by Seaweed Canopies in Winter." *Marine Biology* 165(7): 115.
- Serrão, E. A., G. Pearson, L. Kautsky, and S. H. Brawley. 1996. "Successful External Fertilization in Turbulent Environments." Proceedings of the National Academy of Sciences of the United States of America 93(11): 5286–90. https://doi.org/ 10.1073/pnas.93.11.5286.
- Soltan, D., M. Verlaque, C. F. Boudouresque, and P. Francour. 2001. "Changes in Macroalgal Communities in the Vicinity of a Mediterranean Sewage Outfall After the Setting Up of a Treatment Plant." *Marine Pollution Bulletin* 42(1): 59–70. https:// doi.org/10.1016/S0025-326X(00)00116-8.
- Sousa, W. P. 1979. "Experimental Investigations of Disturbance and Ecological Succession in a Rocky Intertidal Algal Community." *Ecological Monographs* 49(3): 227–54.
- Sousa, W. P. 1984. "Intertidal Mosaics: Patch Size, Propagule Availability, and Spatially Variable Patterns of Succession." *Ecology* 65(6): 1918–35. https://doi.org/10.2307/1937789.
- Steinberg, P. D. 1985. "Feeding Preferences of *Tegula funebralis* and Chemical Defenses of Marine Brown Algae." *Ecological Monographs* 55(3): 333–49. https://doi.org/10.2307/1942581.
- Stekoll, M. H., and L. Deysher. 1996. "Recolonization and Restoration of Upper Intertidal *Fucus gardneri* (Fucales, Phaeophyta)

Following the Exxon Valdez Oil Spill." *Hydrobiologia* 326/327(1): 311–6. https://doi.org/10.1007/BF00047824.

- Strain, E. M. A., R. J. Thomson, F. Micheli, F. P. Mancuso, and L. Airoldi. 2014. "Identifying the Interacting Roles of Stressors in Driving the Global Loss of Canopy-Forming to Mat-Forming Algae in Marine Ecosystems." *Global Change Biology* 20(11): 3300–12. https://doi.org/10.1111/gcb.12619.
- Straughan, D., and R. G. Kanter. 1977. Southern California Baseline Study: Mussel Community Study, Vol III. Report 2.2. Los Angeles, CA: Bureau of Land Management, Department of the Interior.
- Sutherland, W. J., R. Aveling, T. M. Brooks, M. Clout, L. V. Dicks, L. Fellman, E. Fleishman, et al. 2014. "A Horizon Scan of Global Conservation Issues for 2014." *Trends in Ecology & Evolution* 29(1): 15–22. https://doi.org/10.1016/j.tree.2013.11.004.
- Tait, L. W., I. Hawes, and D. R. Schiel. 2014. "Shining Light on Benthic Macroalgae: Mechanisms of Complementarity in Layered Macroalgal Assemblages." *PLoS One* 9(12): e114146. https://doi.org/10.1371/journal.pone.0114146.
- Tait, L. W., and D. R. Schiel. 2010. "Primary Productivity of Intertidal Macroalgal Assemblages: Comparison of Laboratory and In Situ Photorespirometry." *Marine Ecology Progress Series* 416: 115–25. https://doi.org/10.3354/meps08781.
- Takolander, A., E. Leskinen, and M. Cabeza. 2017. "Synergistic Effects of Extreme Temperature and Low Salinity on Foundational Macroalga *Fucus vesiculosus* in the Northern Baltic Sea." *Journal of Experimental Marine Biology and Ecology* 495: 110–8. https://doi.org/10.1016/j.jembe.2017. 07.001.
- Terawaki, T., K. Yoshikawa, G. Yoshida, M. Uchimura, and K. Iseki. 2003. "Ecology and Restoration Techniques for Sargassum Beds in the Seto Inland Sea, Japan." *Marine Pollution Bulletin* 47(1–6): 198–201. https://doi.org/10.1016/ S0025-326X(03)00054-7.
- Thom, R. M., and T. B. Widdowson. 1978. "A Resurvey of E. Yale Dawson's 42 Intertidal Algal Transects on the Southern California Mainland after 15 Years." Bulletin of the Southern California Academy of Sciences 77(1): 1–13.
- Thompson, R. C., B. J. Wilson, M. L. Tobin, A. S. Hill, and S. J. Hawkins. 1996. "Biologically Generated Habitat Provision and Diversity of Rocky Shore Organisms at a Hierarchy of Spatial Scales." *Journal of Experimental Marine Biology and Ecology* 202(1): 73–84. https://doi.org/10.1016/0022-0981(96)00032-9.
- Torn, K., D. Krause-Jensen, and G. Martin. 2006. "Present and Past Depth Distribution of Bladderwrack (*Fucus vesiculosus*) in the Baltic Sea." *Aquatic Botany* 84(1): 53–62. https://doi. org/10.1016/j.aquabot.2005.07.011.
- Tronske, N. 2020. "Experimental Restoration of a Canopy-Forming Foundational Rockweed Species in the Rocky Intertidal Zone: Effects of Silvetia compressa Transplant Density on Thalli Survival and Growth and Subcanopy Recruitment and Community Composition." MS Thesis, California State Polytechnic University.
- Ugarte, R. A., and G. Sharp. 2001. "A New Approach to Seaweed Management in Eastern Canada: The Case of Ascophyllum nodosum." Cahiers de Biologie Marine 42(1/2): 63–70.
- Underwood, A. J. 1996. "Detection, Interpretation, Prediction and Management of Environmental Disturbances: Some Roles for Experimental Marine Ecology." *Journal of Experimental*

Marine Biology and Ecology 200(1-2): 1-27. https://doi.org/10. 1016/S0022-0981(96)02637-8.

- Underwood, A. J. 1998. "Grazing and Disturbance: An Experimental Analysis of Patchiness in Recovery from a Severe Storm by the Intertidal Alga Hormosira banksii on Rocky Shores in New South Wales." Journal of Experimental Marine Biology and Ecology 231(2): 291–306. https://doi.org/ 10.1016/S0022-0981(98)00091-4.
- Underwood, A. J. 1999. "Physical Disturbances and Their Direct Effect on an Indirect Effect: Responses of an Intertidal Assemblage to a Severe Storm." *Journal of Experimental Marine Biology and Ecology* 232(1): 125–40. https://doi.org/10. 1016/S0022-0981(98)00105-1.
- US Congress. 1980. "Comprehensive Environmental Response, Compensation, and Liability Act of 1980." 42 U.S.C. § 9601 et seq. Pub.L. 96–510.
- US Congress. 1990. "Oil Pollution Act. 'Bill Summary & Status 101st Congress (1989–1990) – H.R.1465'." 33 U.S.C. §2701 et seq. http:// thomas.loc.gov/.
- Vadas, S., L. Robert, S. Johnson, and T. A. Norton. 1992. "Recruitment and Mortality of Early Post-Settlement Stages of Benthic Algae." *British Phycological Journal* 27(3): 331–51. https://doi.org/10.1080/00071619200650291.
- Vadas, S., L. Robert, W. A. Wright, and B. F. Beal. 2004. "Biomass and Productivity of Intertidal Rockweeds (Ascophyllum nodosum LeJolis) in Cobscook Bay." Northeastern Naturalist 11(sp2): 123–42. https://doi.org/10.1656/1092-6194(2004)11 [123:BAPOIR]2.0.CO;2.
- Vadas, R. L., W. A. Wright, and S. L. Miller. 1990. "Recruitment of Ascophyllum nodosum: Wave Action as a Source of Mortality." Marine Ecology Progress Series 61(3): 263–72. https://doi.org/ 10.3354/meps061263.
- van Tamelen, P. G., M. S. Stekoll, and L. Deysher. 1997. "Recovery Processes of the Brown Alga *Fucus gardneri* Following the 'Exxon Valdez' Oil Spill: Settlement and Recruitment." *Marine Ecology Progress Series* 160: 265–77. https://doi.org/10. 3354/meps160265.
- Vercaemer, B., M. C. Wong, and M. A. Bravo. 2018. "Fish Assemblages in Rockweed (Ascophyllum nodosum (L.) Le Jolis) Beds on the Atlantic Coast of Nova Scotia, Canada." Canadian Technical Report of Fisheries and Aquatic Sciences 3249. 34 pp.
- Verdura, J., M. Sales, E. Ballesteros, M. E. Cefalì, and E. Cebrian. 2018. "Restoration of a Canopy-Forming Alga Based on Recruitment Enhancement: Methods and Long-Term Success Assessment." *Frontiers in Plant Science* 9: 1832. https://doi. org/10.3389/fpls.2018.01832.
- Vesco, L. L., and R. M. Gillard. 1980. "Recovery of Benthic Marine Populations along the Pacific Coast of the United States Following Natural and Man-Made Disturbances Including Pertinent Life History Information." POCS Reference Paper No. 53–4. Pacific Outer Continental Shelf Office: Bureau of Land Management, Department of the Interior.
- Viejo, R. M., B. Martínez, J. Arrontes, C. Astudillo, and L. Hernández. 2011. "Reproductive Patterns in Central and Marginal Populations of a Large Brown Seaweed: Drastic Changes at the Southern Range Limit." *Ecography* 34(1): 75–84. https://doi.org/ 10.1111/j.1600-0587.2010.06365.x.

- Vogt, H., and W. Schramm. 1991. "Conspicuous Decline of *Fucus* in Kiel Bay (Western Baltic): What Are the Causes?" *Marine Ecology Progress Series* 69: 189–94. https://doi.org/10.3354/ meps069189.
- Vreeland, V., E. Grotkopp, S. Espinosa, D. Quiroz, W. M. Laetsch, and J. West. 1993. "The Pattern of Cell Wall Adhesive Formation by *Fucus* Zygotes." *Hydrobiologia* 260(1): 485–91. https://doi.org/10.1007/BF00049060.
- Wahl, M., M. Molis, A. J. Hobday, S. Dudgeon, R. Neumann, P. Steinberg, A. H. Campbell, E. Marzinelli, and S. Connell. 2015. "The Responses of Brown Macroalgae to Environmental Change from Local to Global Scales: Direct versus Ecologically Mediated Effects." *Perspectives in Phycology* 2(1): 11–29. https://doi.org/10.1127/pip/2015/0019.
- Watt, C. A., and R. A. Scrosati. 2013a. "Bioengineer Effects on Understory Species Richness, Diversity, and Composition Change Along an Environmental Stress Gradient: Experimental and Mensurative Evidence." *Estuarine, Coastal* and Shelf Science 123: 10–8. https://doi.org/10.1016/j.ecss. 2013.02.006.
- Watt, C. A., and R. A. Scrosati. 2013b. "Regional Consistency of Intertidal Elevation as a Mediator of Seaweed Canopy Effects on Benthic Species Richness, Diversity, and Composition." *Marine Ecology Progress Series* 491: 91–9. https://doi.org/10. 3354/meps10521.
- Weitzman, B., B. Konar, K. Iken, H. Coletti, D. Monson, R. Suryan, T. Dean, D. Hondolero, and M. Lindeberg. 2021. "Changes in Rocky Intertidal Community Structure During a Marine Heatwave in the Northern Gulf of Alaska." *Frontiers in Marine Science* 8: 556820. https://doi.org/10.3389/fmars.2021.556820.
- Wernberg, T., B. D. Russell, M. S. Thomsen, C. F. Gurgel, C. J. Bradshaw, E. S. Poloczanska, and S. D. Connell. 2011. "Seaweed Communities in Retreat from Ocean Warming." *Current Biology* 21(21): 1828–32. https://doi.org/10.1016/j.cub. 2011.09.028.
- Whitaker, S. G. 2009. "Experimental Re-establishment of the Rocky Intertidal Brown Alga *Silvetia compressa* at Little Corona del Mar." M.S. Thesis, California State University.
- Whitaker, S. G., J. R. Smith, and S. N. Murray. 2010. "Reestablishment of the Southern California Rocky Intertidal Brown Alga, *Silvetia compressa*: An Experimental Investigation of Techniques and Abiotic and Biotic Factors That Affect Restoration Success." *Restoration Ecology* 18(S1): 18–26. https://doi.org/10.1111/j.1526-100X.2010.00717.x.

- Widdowson, T. B. 1971. "Changes in the Intertidal Algal Flora of the Los Angeles Area Since the Survey by E. Yale Dawson in 1956-1959." Bulletin of the Southern California Academy of Sciences 70(1): 2–16.
- Williams, S. L., and R. E. Di Fiori. 1996. "Genetic Diversity and Structure in *Pelvetia fastigiata* (Phaeophyta: Fucales): Does a Small Effective Neighborhood Size Explain Fine-Scale Genetic Structure?" *Marine Biology* 126(3): 371–82. https:// doi.org/10.1007/BF00354619.
- Wilson, K. L., L. M. Kay, A. L. Schmidt, and H. K. Lotze. 2015. "Effects of Increasing Water Temperatures on Survival and Growth of Ecologically and Economically Important Seaweeds in Atlantic Canada: Implications for Climate Change." *Marine Biology* 162(12): 2431–44. https://doi.org/10.1007/s00227-015-2769-7.
- Yatsuya, K. 2010. "Techniques for the Restoration of *Sargassum* Beds on Barren Grounds." *Bulletin of the Fisheries Research Agency* 32: 69–73.
- Yoon, J. T., S. M. Sun, and G. Chung. 2014. "Sargassum Bed Restoration by Transplantation of Germlings Grown under Protective Mesh Cage." Journal of Applied Phycology 26(1): 505–9. https://doi.org/10.1007/s10811-013-0058-8.
- Yu, Y. Q., Q. S. Zhang, Y. Z. Tang, S. B. Zhang, L. Zhi Cheng, S. H. Chu, and X. X. Tang. 2012. "Establishment of Intertidal Seaweed Beds of *Sargassum thunbergii* through Habitat Creation and Germling Seeding." *Ecological Engineering* 44: 10–7. https://doi.org/10.1016/j.ecoleng.2012.03.016.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

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