

UCLA

UCLA Previously Published Works

Title

Systematic Proteomic Identification of the Heat Shock Proteins (Hsp) that Interact with Estrogen Receptor Alpha (ER α) and Biochemical Characterization of the ER α -Hsp70 Interaction

Permalink

<https://escholarship.org/uc/item/1v20v8s1>

Journal

PLOS ONE, 11(8)

ISSN

1932-6203

Authors

Dhamad, Ahmed E
Zhou, Zhenqi
Zhou, Jianhong
[et al.](#)

Publication Date

2016

DOI

10.1371/journal.pone.0160312

Peer reviewed

RESEARCH ARTICLE

Systematic Proteomic Identification of the Heat Shock Proteins (Hsp) that Interact with Estrogen Receptor Alpha (ER α) and Biochemical Characterization of the ER α -Hsp70 Interaction

Ahmed E. Dhamad, Zhenqi Zhou[‡], Jianhong Zhou, Yuchun Du*

Department of Biological Sciences, University of Arkansas, Fayetteville, Arkansas, United States of America

[‡] Current address: David Geffen School of Medicine, Department of Medicine, Division of Endocrinology, Diabetes and Hypertension, UCLA, Los Angeles, California, United States of America

* ydu@uark.edu



CrossMark
click for updates

OPEN ACCESS

Citation: Dhamad AE, Zhou Z, Zhou J, Du Y (2016) Systematic Proteomic Identification of the Heat Shock Proteins (Hsp) that Interact with Estrogen Receptor Alpha (ER α) and Biochemical Characterization of the ER α -Hsp70 Interaction. PLoS ONE 11(8): e0160312. doi:10.1371/journal.pone.0160312

Editor: Didier Picard, University of Geneva, SWITZERLAND

Received: December 7, 2015

Accepted: July 18, 2016

Published: August 2, 2016

Copyright: © 2016 Dhamad et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper.

Funding: This work was supported by National Institutes of Health (NIH) grants P30 GM103450 and R03 CA169692, and a grant from the Arkansas Biosciences Institute (YD). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Abstract

Heat shock proteins (Hsps) are known to associate with estrogen receptors (ER) and regulate ER-mediated cell proliferation. Historically, the studies in this area have focused on Hsp90. However, some critical aspects of the Hsp-ER α interactions remain unclear. For example, we do not know which Hsps are the major or minor ER α interactants and whether or not different Hsp isoforms associate equally with ER α . In the present study, through a quantitative proteomic method we found that 21 Hsps and 3 Hsp cochaperones were associated with ER α in human 293T cells that were cultured in a medium containing necessary elements for cell proliferation. Four Hsp70s (Hsp70-1, Hsc70, Grp75, and Grp78) were the most abundant Hsps identified to associate with ER α , followed by two Hsp90s (Hsp90 α and Hsp90 β) and three Hsp110s (Hsp105, HspA4, and HspA4L). Hsp90 α was found to be 2–3 times more abundant than Hsp90 β in the ER α -containing complexes. Among the reported Hsp cochaperones, we detected prostaglandin E synthase 3 (p23), peptidyl-prolyl cis-trans isomerase FKBP5 (FKBP51), and E3 ubiquitin-protein ligase CHIP (CHIP). Studies with the two most abundant ER α -associated Hsps, Hsp70-1 and Hsc70, using human breast cancer MCF7 cells demonstrate that the two Hsps interacted with ER α in both the cytoplasm and nucleus when the cells were cultured in a medium supplemented with fetal bovine serum and phenol red. Interestingly, the ER α -Hsp70-1/Hsc70 interactions were detected only in the cytoplasm but not in the nucleus under hormone starvation conditions, and stimulation of the starved cells with 17 β -estradiol (E2) did not change this. In addition, E2-treatment weakened the ER α -Hsc70 interaction but had no effect on the ER α -Hsp70-1 interaction. Further studies showed that significant portions of Hsp70-1 and Hsc70 were associated with transcriptionally active chromatin and inactive chromatin, and the two Hsps interacted with ER α in both forms of the chromatins in MCF7 cells.

Introduction

Estrogen receptor alpha (ER α) is a nuclear transcription factor that controls the expression of estrogen responsive genes. Like other members of steroid receptor (SR) superfamily including androgen receptor, progesterone receptor, glucocorticoid receptor and mineralocorticoid receptor, the responsiveness of ER α to its ligands such as 17 β -estradiol (E2) is regulated by heat shock proteins (Hsps) and their cochaperones [1–3]. In the absence of estrogenic ligands, ER α is assembled into an Hsp90-based chaperone protein complex, which keeps ER α in a ligand-binding competent but inactive state and prevents it from binding to estrogen-response elements [4–7]. Unliganded ER α is a short-lived protein with a half-life of 4–5 h and is constantly degraded [8]. The degradation is mediated by E3 ubiquitin-protein ligase CHIP (CHIP) and through the ubiquitin-proteasome pathway [9–11]. Upon binding of its ligands, ER α dissociates from Hsp90, dimerizes, binds to the estrogen-response elements, and induces transcription of its target genes through recruiting co-activators [12, 13]. Hsp90 is essential for ER α hormone binding [6], dimer formation [12], and binding to the estrogen-response elements [14].

The Hsps are highly conserved chaperones and play important roles in protein folding, assembly, trafficking and disposition, and stress responses [15, 16]. Human Hsps are classified into six families, Hsp110 (HspH), Hsp90 (HspC), Hsp70 (HspA), Hsp40 (DNAJ), small Hsps (HspB), and chaperonin (HspD/E and CCT) [17, 18]. Hsps vary substantially from one to another with regards to function, expression, and subcellular localization. Some Hsps are constitutively expressed such as Hsc70 and Hsp90 β , whereas others are induced by stresses such as Hsp70-1 and Hsp90 α [19, 20]. While some Hsps are localized in specific cellular compartments, such as Grp75 in the mitochondria and Grp78 in the endoplasmic reticulum, most Hsps are localized in the cytoplasm and the nucleus [21, 22]. Hsp70 and Hsp90 are among the most abundant cellular proteins, with each family accounting for 1–2% of total cellular protein under normal conditions and 2–4% under stress conditions [23–26]. Despite the fact that Hsp70 and Hsp90 are among the main conserved protective systems in cells [27], they are substantially overexpressed in cancer cells, and the upregulations correlate with poor prognosis [28, 29]. Because of the important roles of Hsp70 and Hsp90 in regulating SRs, and the “addiction” of cancer cells to higher levels of Hsps, inhibitors of Hsp70 and Hsp90 are actively being pursued for treating cancers [23, 24, 28, 30–32].

The extensive studies on the interactions of Hsps with SRs including ER α over the past five decades have established the fundamental roles of Hsps, Hsp90 in particular, in regulating SRs [33]. However, some details are missing and in some cases results are controversial. For examples, because Hsp90 α and Hsp90 β share 86% sequence [34], it is expected that the two isoforms have similar functions in cells. Probably because of this reason, many publications on studying the roles of Hsp90 in regulating SRs even did not mention which isoforms they used. However, while Hsp90 α -knockout mice are viable, Hsp90 β -knockout mice are lethal [35, 36]. As myoblasts differentiate into myotubes, Hsp90 α disappears and only Hsp90 β remains, and the isoform switch is essential for the differentiation [37]. These results suggest that there are critical differences between the two isoforms. Through a quantitative proteomic approach, we have comprehensively identified cellular proteins that are associated with ER α in human 293T cells that were grown in a “complete” culture medium [a medium that was supplemented with growth stimulating factors including phenol red and fetal bovine serum (FBS)]. Here we present the results revealing the interactions between ER α and Hsps/cochaperones at the proteome level. Our proteomic data demonstrate that four Hsp70 family members, Hsp70-1, Hsc70, Grp75 and Grp78, were the predominant Hsps that were associated with ER α in 293T cells, followed by two Hsp90 family members, Hsp90 α and Hsp90 β , and three Hsp110 family

members, Hsp105, HspA4 and HspA4L. In addition, three Hsp cochaperones, prostaglandin E synthase 3 (p23), peptidyl-prolyl cis-trans isomerase FKBP5 (FKBP51) and CHIP, were also identified to associate with ER α . Studies with the two most abundant ER α -associated Hsps, Hsp70-1 and Hsc70, suggest that these two Hsps interact with ER α in the cytoplasm and the nucleus when human breast cancer MCF7 cells were cultured in the conventional laboratory conditions. However, under hormone starvation, the ER α -Hsp70-1/Hsc70 interactions were observed only in the cytosol, and E2 stimulation did not change the pattern. The E2-treatment weakened the ER α -Hsc70 interaction but had no effect on the ER α -Hsp70-1 interaction. Different from Hsp90 α , significant portions of Hsp70-1 and Hsc70 were found to be associated with transcriptionally active chromatin and inactive chromatin, and the two Hsps interacted with ER α in both forms of the chromatins in MCF7 cells.

Materials and Methods

Cell culture, proteome labeling, and affinity purification

We used the SILAC/AACT (stable isotope labeling with amino acids in cell culture/amino acid-coded tagging) approach to label the proteome of cells [38, 39]. A population of human embryonic kidney 293T cells (American Type Culture Collection, Manassas, VA) were cultured in labeled (Arg-¹³C₆ and Lys-¹³C₆ ¹⁵N₂) Dulbecco's Modified Eagle's Medium (DMEM, Thermo Fisher Scientific, Waltham, MA) with 10% dialyzed FBS and 1% penicillin and streptomycin for two weeks and then transiently transfected with a plasmid expressing Flag tag alone. A second population of 293T cells were cultured in unlabeled DMEM with 10% FBS and 1% penicillin and streptomycin, and transiently transfected with a plasmid expressing Flag-ER α . The two population of cells were harvested 48 h after transfection, washed with cold PBS, and incubated in 5 packed cell pellet volumes of lysis buffer I [20 mM Tris-HCl, pH 7.5, 0.5% NP-40, 1 mM EDTA, 10 nM E2, protease inhibitors (Roche, Indianapolis, IN), and phosphatase inhibitors (1 mM Na₃VO₄, 10 mM NaF, and 10 mM β -glycerophosphate)] on ice for 30 min. The cells were then lysed by douncing with a 15-mL glass dounce homogenizer with a tight-fitting type B pestle (Kontes Glass Co., Vineland, NJ). After adding NaCl and glycerol to final concentrations of 125 mM and 10%, respectively, the extracts were centrifuged at 20,000 g for 15 min at 4°C. The resulting pellets were resuspended in lysis buffer I supplemented with 125 mM NaCl and 10% glycerol and extracted again with sonication (Branson Digital Sonifier 450, Branson Ultrasonics Co., CT) [40]. Protein concentration of the combined and cleared supernatant was determined, and equal amounts of the labeled and unlabeled cell extracts were separately incubated with pre-washed Flag M2 resin (Sigma-Aldrich, St. Louis, MO) for 5 h at 4°C with end-to-end rotation. The beads were then washed extensively with lysis buffer I supplemented with 125 mM NaCl and 10% glycerol. The bound proteins were eluted with elution buffer (10 mM Tris-HCl, pH 7.5, 350 mM NaCl, 1 mM EDTA, 250 mM 3X Flag peptides, and protease inhibitors). The eluates of the two affinity purifications were mixed and fractionated with a 12% SDS-PAGE gel for liquid chromatography-tandem mass spectrometry (LC-MS/MS) analysis. Human breast cancer MCF7 cells (American Type Culture Collection, Manassas, VA) were maintained in Minimum Essential Medium α (MEM α ; Thermo Fisher Scientific, Waltham, MA) with 5% FBS and 1% penicillin and streptomycin.

LC-MS/MS, database search, and data analysis

In-gel digestion, LC-MS/MS analysis, and protein identification/quantification with the Maxquant (version 1.0.13.13) and Mascot (version 2.2; Matrix Science, Boston, MA) by searching against a composite target-decoy International Protein Index (IPI) human protein database (version 3.52) were performed as described previously [41]. In this SILAC/AACT approach,

because the Flag-ER α expressing cells and the Flag expressing cells were cultured in the unlabeled medium and stable-isotope-labeled medium, respectively, and the eluates from the two affinity purifications of equal amounts of the unlabeled cell extract and labeled cell extract were mixed and analyzed by LC-MS/MS, the relative intensities of the paired isotopic peaks of peptides (i.e., light/heavy ratios: L/H ratios) reflect the binding profile of the protein to ER α . Whereas the L/H ratios for the nonspecific binding proteins were around 1, the ratios for the proteins that specifically bind to ER α were significantly larger than 1 due to affinity enrichment of the proteins [42, 43]. Search results were further processed by Scaffold software (version 4.4.7; Proteome Software Inc., Portland, OR) for viewing protein and peptide identification information. In the Scaffold analysis, protein identification probability with at least two peptides was set to 99% and the peptide identification probability was set to 95%. The normalized spectral abundance factors (NcSAFs) were calculated as described [44, 45]. The normalization was applied only to the identified Hsps and cochaperones to estimate the relative level of each protein within the identified Hsps and cochaperones that were associated with ER α [44, 45]. Spectral counts for peptides shared among the identified Hsps were counted only once, and distributed based on the number of unique spectral counts to each isoform [46].

The E2 treatment and subcellular fractionation

The MCF7 cells were cultured in the phenol-red free MEM α supplemented with 5% charcoal-treated FBS (Hyclone, Logan, UT) for 3–4 days and then treated with either 100 nM E2 or ethanol (control) for 24 h. The cells were then harvested, washed twice with cold PBS, resuspended in 5 packed cell pellet volumes of hypotonic buffer (10 mM Tris-HCl, pH 8.0, 5 mM KCl, and 1.5 mM MgCl₂, and protease inhibitors) supplemented with 100 nM E2 for the E2-treated cells or ethanol for the control cells. The cells were incubated on ice for 20 min. After adding phosphatase inhibitors (1 mM Na₃VO₄, 10 mM NaF, and 10 mM glycerophosphate) to the cell suspension, the cells were lysed by douncing 12 times with a 15-mL glass dounce homogenizer with a tight-fitting type B pestle. After centrifugation at 500 g for 10 min at 4°C, the pellet was saved and the supernatant was cleared by centrifugation at 10,000 g for 15 min at 4°C. The cleared supernatant was supplemented with 15 mM Tris-HCl, pH 8.0, 140 mM NaCl, 1% Triton X-100, 0.1% SDS and 3 mM EDTA, and saved as cytosolic fraction. The pellet from the 500 g centrifugation was resuspended in hypotonic buffer and dounced 5 times. After centrifugation at 500 g for 10 min at 4°C, the pellet was washed twice with hypotonic buffer and saved as nuclei. The isolated nuclei were resuspended in lysis buffer II (25 mM Tris-HCl, pH 8.0, 140 mM NaCl, 1% Triton X-100, 3 mM EDTA, 0.1% SDS, protease inhibitors, and phosphate inhibitors) supplemented with 100 nM E2 for the E2-treated samples or ethanol for the control samples. The nuclei were then sonicated on ice, centrifuged at 10,000 g for 15 min at 4°C, and the resulting supernatant was designated as nuclear fraction.

Cross-linking, immunoprecipitation (IP), and Western blotting

In-cell cross-linking was performed using the cell-permeable cross-linking reagent dithiobis (succinimidylpropionate) (DSP) (Thermo Fisher Scientific, Waltham, MA). The MCF7 cells in plates were washed twice with PBS at room temperature and incubated with 1 mM DSP in DMEM at 37°C for 15 min. After removal of the cross-linker solution, the cells were incubated with quenching solution (100 mM Tris-HCl, pH 8.0 in DMEM) at 37°C for 10 min. Quenching solution was removed, and the cells were washed twice with PBS and lysed for IPs. The IPs and Western blotting were performed as described previously [47, 48]. Antibodies used in this study were purchased from the following commercial sources: Anti-ER α , p300, and NCoR antibodies from Santa Cruz Biotech (Santa Cruz, Dallas, TX; catalog no.: Anti-ER α , sc-8002;

anti-p300, sc-584; anti-NCoR, sc-1609), anti-Hsp70-1 and Hsc70 antibodies from Enzo life science (Farmingdale, NY; catalog no.: anti-Hsp70-1, ADI-SPA-810; anti-Hsc70, ADI-SPA-815), anti-Hsp90 α from Epitomics (Burlingame, CA; catalog no., 3670-1), anti-histone H3 from Cell signaling (Danvers, MA; catalog no., 9715), and anti-tubulin from Sigma-Aldrich (St. Louis, MO; catalog no., T9026). Quantification of protein bands in Western blotting was performed using ImageJ software.

Extraction of chromatin-binding protein, and transcriptionally active chromatin and inactive chromatin

Chromatin-binding protein was extracted with 0.3% SDS and 250 units/mL benzonase as described by Yang *et al.* (2014) [49]. Briefly, after MCF7 cells were resuspended in a radioimmunoprecipitation assay (RIPA) buffer (50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.25% deoxycholic acid, 1% NP-40, and 1 mM EDTA) supplemented with 200 μ M phenylmethylsulfonyl fluoride (PMSF), 1 mM sodium orthovanadate and protease inhibitors, the cells were homogenized by passing through a 22G needle 10 times, followed by an incubation on ice for 20 min. The chromatin was separated from the soluble protein (S) by a centrifugation at 1,000 g, and the isolated chromatin was extracted with 0.3% SDS and 250 units/mL benzonase (EMD Millipore, Billerica, MA) on ice for 10 min. The digested chromatin was centrifuged at 1,000 g, and the resulting supernatant was designated as chromatin-binding protein (CB). Transcriptionally active chromatin and inactive chromatin were extracted with different concentrations of salt according to Henikoff *et al.* (2009) and Yang *et al.* (2014) [49, 50]. Briefly, after MCF7 cells were lysed with a lysis buffer (10 mM Tris-HCl, pH 8.0, 10 mM KCl, 1.5 mM MgCl₂, 340 mM sucrose, 10% glycerol, 1 mM DTT, 0.1% Triton X-100, and protease inhibitors) on ice for 8 min, cytoplasmic protein (C) was separated from the nuclei with a 1,300 g centrifugation. The washed nuclei were digested with 2,000 gel units/mL micrococcal nuclease (New England Biolabs, Ipswich, MA) in the lysis buffer described above plus 1 mM CaCl₂ at 37°C for 10 min, and the reaction was stopped by 2 mM EGTA. After centrifugation at 1,300 g for 10 min at 4°C, the supernatant (nuclear soluble protein: NS) was removed and the digested nuclei were washed and first treated with 150 mM NaCl at 4°C for 2 h for extracting active chromatin (Ch1) and then with 600 mM NaCl at 4°C overnight for extracting inactive chromatin (Ch2).

Statistical analysis

Statistical analysis was performed using one-way ANOVA (PSI-PLOT, Pearl River, NY). A *p*-value of <0.05 was considered significant.

Results

Identification of Hsps and their cochaperones that associate with ER α

We used a SILAC/AACT-based quantitative proteomic method to systematically identify cellular proteins that were associated with ER α [42, 43]. Through this approach, a subset of Hsps and their cochaperones were identified to associate with ER α (Table 1). Most of the Hsps and cochaperones were identified with high confidence with LC-MS/MS (S1 Table), which can be reflected by the very low PEP (posterior error probability) values for the identifications (Table 1).

To examine the abundance of the identified Hsps and cochaperones that were associated with ER α , we calculated NcSAF for each protein [51]. NcSAF is based on spectral counting for each protein in LC-MS/MS analysis, and a larger NcSAF value reflects the higher abundance of the protein in biological samples [44, 46, 51, 52]. The most abundant Hsps that were associated

Table 1. Heat shock proteins and their cochaperones that were identified to associate with ERα in human cells.

Family	Gene names	Protein names (short names)	UniProt ID	Unique peptides	Unique spectra	Sequence coverage (%)*	NcSAF	L/H ratios†	PEP‡	
Hsp70	<i>HSPA1A</i>	Heat shock 70 kDa protein 1A/1B (Hsp70-1)	P08107	28	927	60.5	0.286	8.9	0	
	<i>HSPA2</i>	Heat shock-related 70 kDa protein 2	P54652	12	38	37.6	0.012	8.0	3.2E-198	
	<i>HSPA5</i>	78 kDa glucose-regulated protein (Grp78)	P11021	35	266	51.4	0.080	10.7	0	
	<i>HSPA6</i>	Heat shock 70 kDa protein 6	P17066	6	15	22.9	0.005	8.0	1.1E-121	
	<i>HSPA8</i>	Heat shock cognate 71 kDa protein (Hsc70)	P11142	25	810	60.5	0.248	11.7	0	
	<i>HSPA9</i>	Stress-70 protein, mitochondrial (Grp75)	P38646	33	425	49.9	0.124	12.1	0	
Hsp90	<i>HSP90AA1</i>	Heat shock protein Hsp90-alpha (Hsp90α)	P07900	18	221	45.4	0.057	6.7	3.3E-195	
	<i>HSP90AB1</i>	Heat shock protein Hsp90-beta (Hsp90β)	P08238	19	72	47.1	0.022	5.5	1.7E-144	
Hsp110	<i>HSPH1</i>	Heat shock protein 105 kDa (Hsp105)	Q92598	25	74	34.1	0.016	24.6	3.7E-241	
	<i>HSPA4</i>	Heat shock 70 kDa protein 4 (HspA4)	P34932	38	154	54.0	0.032	4.7	0	
	<i>HSPA4L</i>	Heat shock 70 kDa protein 4L (HspA4L)	O95757	22	60	35.9	0.012	26.2	9.2E-163	
Hsp40	<i>DNAJA2</i>	DnaJ homolog subfamily A member 2	O60884	2	4	6.1	0.002	2.1	9.0E-22	
	<i>DNAJA3</i>	DnaJ homolog subfamily A member 3, mitochondrial	Q96EY1	5	16	16.9	0.005	12.7	1.0E-55	
	<i>DNAJB1</i>	DnaJ homolog subfamily B member 1	P25685	4	6	12.6	0.003	5.6	1.7E-15	
	<i>DNAJB4</i>	DnaJ homolog subfamily B member 4	Q9UDY4	2	4	5.9	0.002	4.7	2.6E-07	
	<i>DNAJB6</i>	DnaJ homolog subfamily B member 6	O75190	3	8	9.2	0.004	2.3	1.2E-12	
	<i>DNAJC7</i>	DnaJ homolog subfamily C member 7	Q99615	4	8	9.9	0.003	2.0	4.9E-25	
	<i>DNAJC9</i>	DnaJ homolog subfamily C member 9	Q8WXX5	10	21	35.0	0.013	14.7	0.068	
	<i>DNAJC10</i>	DnaJ homolog subfamily C member 10	Q8IXB1	2	6	2.8	0.001	5.9	1.2E-07	
	Small Hsps	<i>HSPB8</i>	Heat shock protein beta-8 (Hsp22)	Q9UJY1	2	4	9.7	0.003	12.0	0.0035
	Chaperonin	<i>HSPE1</i>	10 kDa heat shock protein, mitochondrial	P61604	3	10	31.4	0.016	15.6	8.3E-27
Cochaperones	<i>STUB1</i>	E3 ubiquitin-protein ligase CHIP (CHIP)	Q9UNE7	14	57	45.5	0.030	21.6	1.5E-111	
	<i>FKBP5</i>	Peptidyl-prolyl cis-trans isomerase FKBP5 (FKBP51)	Q13451	18	39	38.5	0.014	16.5	8.0E-108	
	<i>PTGES3</i>	Prostaglandin E synthase 3 (p23)	Q15185	4	12	15.6	0.012	171.1	2.0E-08	

* Coverage of all peptide sequences matched to the identified protein sequence (%).

† Ratios of light peptides (derived from Flag-ERα-expressing cells) versus heavy peptides (derived from Flag alone-expressing cells).

‡PEP: posterior error probability.

doi:10.1371/journal.pone.0160312.t001

with ER α were four Hsp70 family members, Hsp70-1, Hsc70, Grp75 and Grp78, with the NcSAF values in the range of 0.08–0.286. The L/H ratios for all the identified Hsp70s varied in a narrow range of from 8 to 12, suggesting they were enriched by affinity purification similarly.

Two Hsp90 family members and three Hsp110 family members were also identified to be abundant in the ER α -containing complexes, though at significantly less levels than the four Hsp70 family members described above (Table 1). Among the 5 reported Hsp90 members [17], Hsp90 α and Hsp90 β , which share 86% sequence homology [34], were identified to associate with ER α . The NcSAF values for Hsp90 α and Hsp90 β were 0.057 and 0.022, respectively, and thus the former was 2.6-fold of that of the latter, suggesting that Hsp90 α is 2–3 times more abundant than Hsp90 β in the ER α -containing protein complexes. It is known that while the expression of Hsp90 α is inducible, Hsp90 β is constitutively expressed [53]. The L/H ratios were similar for Hsp90 α and Hsp90 β (6.7 and 5.5, respectively), suggesting the proportions of those that were specifically associated with ER α to those of non-specific bindings for the two isoforms were similar. The Hsp110 members are known as nucleotide exchanger factors (NEFs) of Hsp70 and interact with Hsc70 [17, 54]. Three Hsp110 members, Hsp105, HspA4 and HspA4L, were identified to abundantly associate with ER α (Table 1). The abundances of the three Hsp110 members were comparable to those of Hsp90 α and Hsp90 β , with the NcSAF values in the range of 0.012–0.032. HspA4 and HspA4L were originally considered as members of Hsp70 [55], but now are classified as members of the Hsp110 family [56]. It is noteworthy that Hsp105 and HspA4L were identified with high L/H ratios, suggesting that they were highly enriched by anti-Flag antibody.

The Hsp40 (DNAJ proteins) constitutes the largest subgroup of the Hsp family, up to 50 members, in human cells. One of the major functions of Hsp40 is to couple with Hsp70 to facilitate folding of Hsp70 client proteins [27]. We identified eight Hsp40 members in this study, and all of them were identified with smaller NcSAF values compared with other identified Hsps except for DNAJC9, which was identified with a NcSAF value comparable to those for the Hsp110 members. These results suggest that the majority of Hsp40 members are not abundant in the ER α -containing complexes. Based on the fact that Hsp40 physically interacts with Hsp70 [27], it is likely that Hsp40 interacts with ER α indirectly and the interactions are mediated by Hsp70.

Multiple Hsp cochaperones, including p23, FKBP51, FKBP52, protein phosphatase 5 (PP5) and cyclophilin 40 (Cyp40), have been reported to couple with Hsp90 to facilitate the function of SRs [1, 33]. Most of these cochaperones contain tetratricopeptide repeat domains, which bind to the EEDV motif of Hsp90/Hsp70 [57], and are typically assembled into SR complexes at the final stages of assembly to form the mature, hormone-competent states of SRs [58, 59]. Among the reported cochaperones, we identified p23, FKBP51, and CHIP but were not able to detect FKBP52, Cyp40, and PP5 (Table 1). Notably, CHIP was identified with a larger NcSAF value (0.03), which was comparable to those for the two Hsp90 family members and the three Hsp110 family members, suggesting that CHIP is also abundantly associated with ER α . CHIP has been shown to interact with ER α via its tetratricopeptide repeat domain and mediates ER α degradation through the ubiquitin-proteasome pathway in the nucleus [10, 11].

Hsp70-1 and Hsc70 interact with ER α in the cytoplasm and the nucleus

The role of Hsp90 in regulating the assembly, trafficking, and transcriptional activity of ER α has been studied extensively [1]. Compared with Hsp90, much less is known about the role of Hsp70 in regulating ER α and some results are controversial [5, 7, 60]. In this study, we found that Hsp70-1 and Hsc70 were the two most abundant Hsps that were associated with ER α (Table 1). As the first step to characterizing these important interactions, we proceeded to

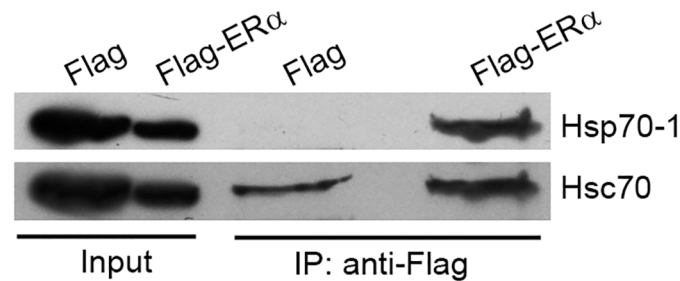


Fig 1. Verification of the interaction between Flag-ER α and endogenous Hsp70-1/Hsc70. The 293T cells were transfected with a plasmid that expresses Flag alone (control) or Flag-ER α . Forty eight hours after transfection, the cells were harvested, lysed, and the resulting total protein was pulled down by immobilized anti-Flag antibody. The bound proteins were analyzed with Western blotting using anti-Hsp70-1 and anti-Hsc70 antibodies.

doi:10.1371/journal.pone.0160312.g001

verify the interaction of ER α with Hsp70-1/Hsc70 using IP and Western blotting. Consistent with our proteomic data, the IP results obtained with the 293T cells ectopically expressing Flag-ER α demonstrate that Flag-ER α interacted with endogenous Hsp70-1 and Hsc70 (Fig 1). To examine if endogenous ER α interacts with endogenous Hsp70-1/Hsc70 and determine the subcellular site where the ER α -Hsp70-1/Hsc70 interactions occur in ER α -positive breast cancer cells, we performed IPs using cytosolic and nuclear proteins of human breast cancer MCF7 cells as starting materials, respectively. The results demonstrate that anti-ER α antibody precipitated significantly more Hsp70-1 than the control IgG precipitated in both the cytosolic fractions and the nuclear fractions (Fig 2A, top row; Fig 2B, left panel). However, the amounts of Hsc70 that were precipitated by anti-ER α antibody and the control IgG were not statistically significant. In addition, we observed large variations on Hsc70 in the IP results among different sample preparations (Fig 2A, middle row; Fig 2B, left panel). The interactions between SRs and Hsps are typically transient and weak by nature [61]. To confirm the interaction of endogenous ER α with Hsc70 and to further validate the specific ER α -Hsp70-1 interaction, we used the cell-permeable cross-linking reagent DSP to treat MCF7 cells and then used whole cell lysate of the DSP-treated cells to perform IPs and Western blotting. The results demonstrate that anti-ER α antibody precipitated significantly more Hsp70-1 and Hsc70 proteins than the IgG precipitated after the cross-linking treatment (Fig 2C). These results suggest that Hsp70-1 and Hsc70 indeed specifically interact with ER α in addition to the nonspecific interactions. We have confirmed the effectiveness of our subcellular fractionation by performing Western blot analysis using markers of the cytoplasm and the nucleus (Fig 2B, right panel).

Hsp70-1 and Hsc70 interact with ER α in transcriptionally active and inactive chromatin

To characterize the interactions of ER α with Hsp70-1/Hsc70, we fractionated MCF7 cell extracts into soluble protein (S), chromatin-binding protein (CB) and the remaining pellet (P), and analyzed those fractions with Western blotting. The results demonstrate that significant portions of Hsp70-1 and Hsc70 were associated with chromatin and the remaining pellets (Fig 3A). In contrast, the amount of Hsp90 α associated with chromatin was neglectable and none was detected in the remaining pellet. As expected, a large portion of ER α , a transcriptional factor, was also associated with chromatin and the pellet. The analysis of a marker of chromatin-binding protein, histone H3, confirmed that the method we used for extracting chromatin-binding protein was effective (Fig 3A). To examine how Hsp70-1 and Hsc70 are associated with chromatin, we fractionated MCF7 cell extracts into cytoplasmic protein (C), nuclear

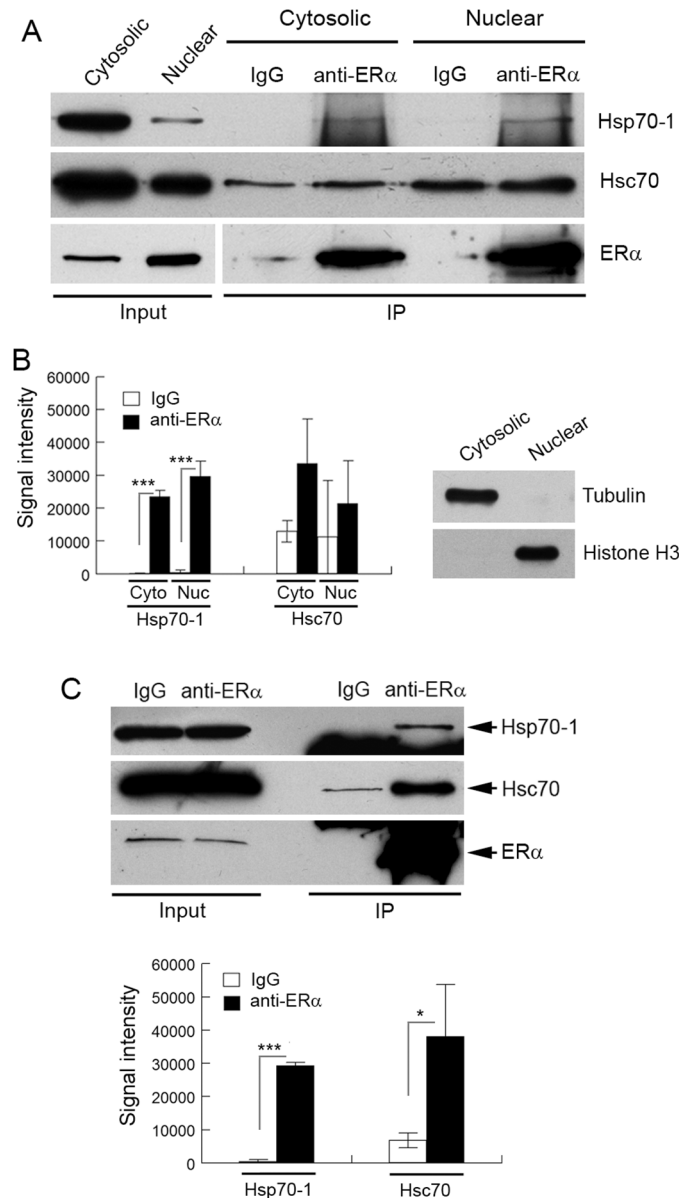


Fig 2. Endogenous ER α interacts with endogenous Hsp70-1 and Hsc70. (A) The cytosolic and nuclear extracts of MCF7 cells were immunoprecipitated by anti-ER α antibody or an isotype-matched, unrelated control IgG, and the immunoprecipitated protein was analyzed by Western blotting with the indicated antibodies. (B) Left panel, quantification of the IP protein bands in Western blots. Signal intensity values were arbitrary numbers obtained by analyzing the protein bands with ImageJ software. Right panel, validation of the cytosolic and nuclear fractionations. Tubulin and histone H3 were used as markers for the cytosolic and nuclear fractions, respectively. (C) The MCF7 cells were treated with the cell-permeable cross-linking reagent DSP and whole cell lysate of the DSP-treated cells was immunoprecipitated by anti-ER α antibody or a control IgG, followed by Western blot analyses with the indicated antibodies. Values in the Western blot quantifications in (B) and (C) were the means \pm S.D. of three separate sample preparations. Cyto, cytosolic; Nuc, nuclear. * and *** denote $p < 0.05$ and $p < 0.001$, respectively.

doi:10.1371/journal.pone.0160312.g002

soluble protein (NS), transcriptionally active chromatin (Ch1), and inactive chromatin (Ch2) [49]. The results demonstrate that significant portions of Hsp70-1 and Hsc70 were associated with active chromatin and inactive chromatin (Fig 3B). In contrast, only a tiny amount of Hsp90 α was associated with active chromatin and none was detected to associate with inactive

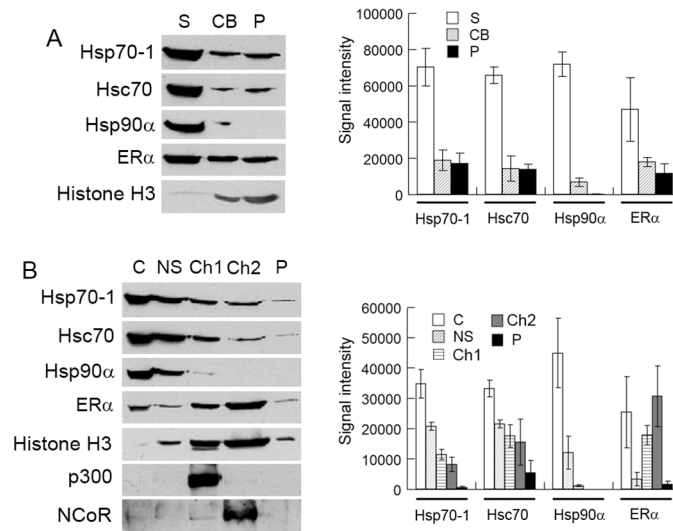


Fig 3. Hsp70-1 and Hsc70 are associated with chromatin. (A) The MCF7 cell extract was fractionated into soluble protein (S), chromatin-binding protein (CB), and the pellet (P), and then analyzed by Western blotting with the indicated antibodies (left panel). Right panel, quantification of Western blots. (B) The MCF7 cell extract was fractionated into cytoplasmic protein (C), nuclear soluble protein (NS), transcriptionally active chromatin (Ch1) and inactive chromatin (Ch2), and analyzed by Western blotting with the indicated antibodies (left panel). Right panel, quantification of Western blots. Histone H3, p300, and NCoR were used as markers of chromatin-binding protein, active chromatin, and inactive chromatin, respectively. Signal intensity values in the Western blot quantifications were arbitrary numbers obtained by analyzing the protein bands with ImageJ software. Values in the Western blot quantifications were the means \pm S.D. of three separate sample preparations.

doi:10.1371/journal.pone.0160312.g003

chromatin. The portions of Hsp70-1, Hsc70, and Hsp90 α that existed as nuclear soluble protein were comparable among the three Hsps (Fig 3B). These results suggest that different from Hsp90 α , which is localized almost exclusively in the cytoplasm and in the nucleus as non-chromatin-binding protein, Hsp70-1 and Hsc70 are also associated with active chromatin and inactive chromatin in addition to being localized in the cytoplasm and in the nucleus as non-chromatin-binding protein. Strikingly, a large portion of ER α was associated with inactive chromatin when the MCF7 cells were cultured in the “complete” medium. We have verified our active/inactive chromatin extraction protocol with a well-established coactivator-p300 and a corepressor-NcoR, which are typically associated with transcriptionally active chromatin and inactive chromatin, respectively [62, 63] (Fig 3B, left panel).

To examine in which subcellular fraction Hsp70-1 and Hsc70 interact with ER α , we performed IPs using fractionated (cytoplasmic, nuclear soluble, active chromatin, and inactive chromatin fractions) proteins from MCF7 as starting materials. The results demonstrate that anti-ER α antibody precipitated significantly more Hsp70-1 and Hsc70 than the control IgG precipitated in all four fractions tested except for Hsc70 in the cytosolic fraction due to large variations among different sample preparations (Fig 4). We have confirmed the presence of ER α in the expected samples by probing the membrane with anti-ER α body (Fig 4, middle panel). It seemed that the precipitated amounts of Hsp70-1 and Hsc70 correlated with the amount of ER α that was precipitated, which in turn seemed to be correlated with the level of ER α in input samples (Fig 4, top and middle panels; S1 Fig). In addition, despite that the majority of Hsp70-1 and Hsc70 were localized in cytoplasm and in the nucleus as soluble protein (Figs 3B and 4, top panel), significant portions of the ER α -Hsp70-1 and ER α -Hsc70 interactions occurred in the active chromatin and inactive chromatin (Fig 4, middle and low

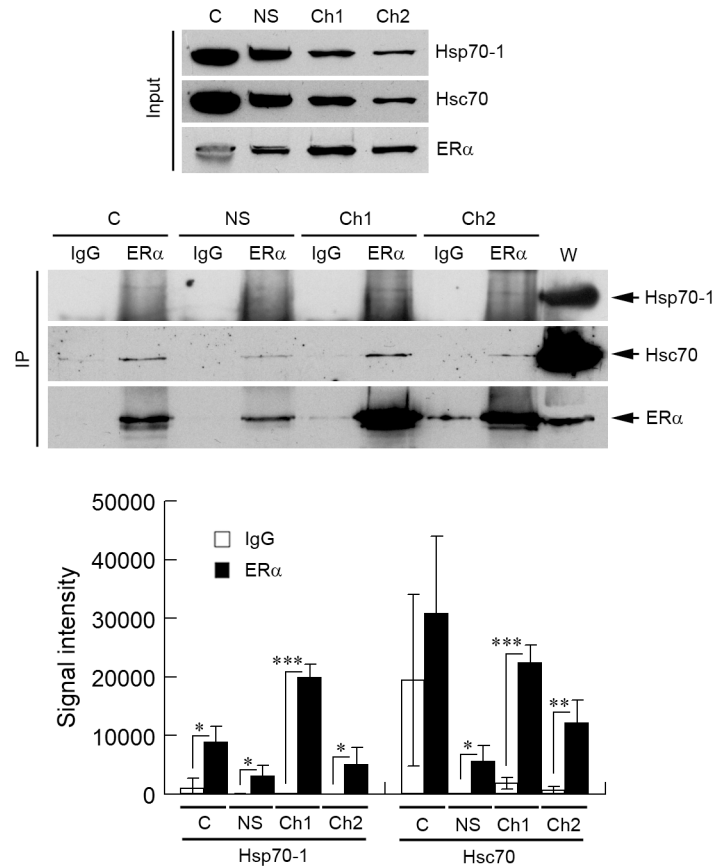


Fig 4. Hsp70-1 and Hsc70 interact with ER α in chromatin. Anti-ER α antibody (ER α) and an isotype-matched, unrelated control IgG were used to immunoprecipitate proteins from cytoplasmic (C), nuclear soluble (NS), transcriptionally active chromatin (Ch1), and inactive chromatin (Ch2) fractions prepared from MCF7 cells. The subcellular proteins were prepared as for Fig 3 except that the inactive chromatin (Ch2) was obtained through sonication instead of elution with 600 mM NaCl. Signal intensity values in the Western blot quantifications were arbitrary numbers obtained by analyzing the protein bands with ImageJ software. Values in the Western blot quantifications were the means \pm S.D. of three separate sample preparations. W, whole cell lysate. *, **, and *** denote $p < 0.05$, $p < 0.01$, and $p < 0.001$, respectively.

doi:10.1371/journal.pone.0160312.g004

panels), suggesting that the levels of Hsp70-1 and Hsc70 do not affect the amounts of the ER α -Hsp70-1 and ER α -Hsc70 interactions. In short, the results in this section demonstrate that Hsp70-1 and Hsc70 interact with ER α in both active chromatin and inactive chromatin.

ER α interacts with Hsp70-1/Hsc70 in the cytoplasm under conditions of hormone starvation/stimulation

To examine the effect of estrogens on the interaction of ER α with Hsp70-1/Hsc70 in ER α -positive cells, we cultured MCF7 cells in the phenol-red free MEM α supplemented with 5% charcoal-treated FBS for 3–4 days, and then treated the cells with either 100 nM E2 or ethanol (control) for 24 h. We then harvested the cells, fractionated the cell extracts into cytosolic and nuclear fractions, and performed IPs using the cytosolic and nuclear fractions, respectively, as starting materials. The results demonstrate that anti-ER α antibody immunoprecipitated more Hsp70-1 and Hsc70 than the IgG precipitated in the cytosolic fractions (Fig 5A, low panel; compare lane 2 with lane 1, and lane 4 with lane 3; Fig 5B, left panel), suggesting that ER α interacts with Hsp70-1 and Hsc70 in the cytoplasm under conditions of hormone

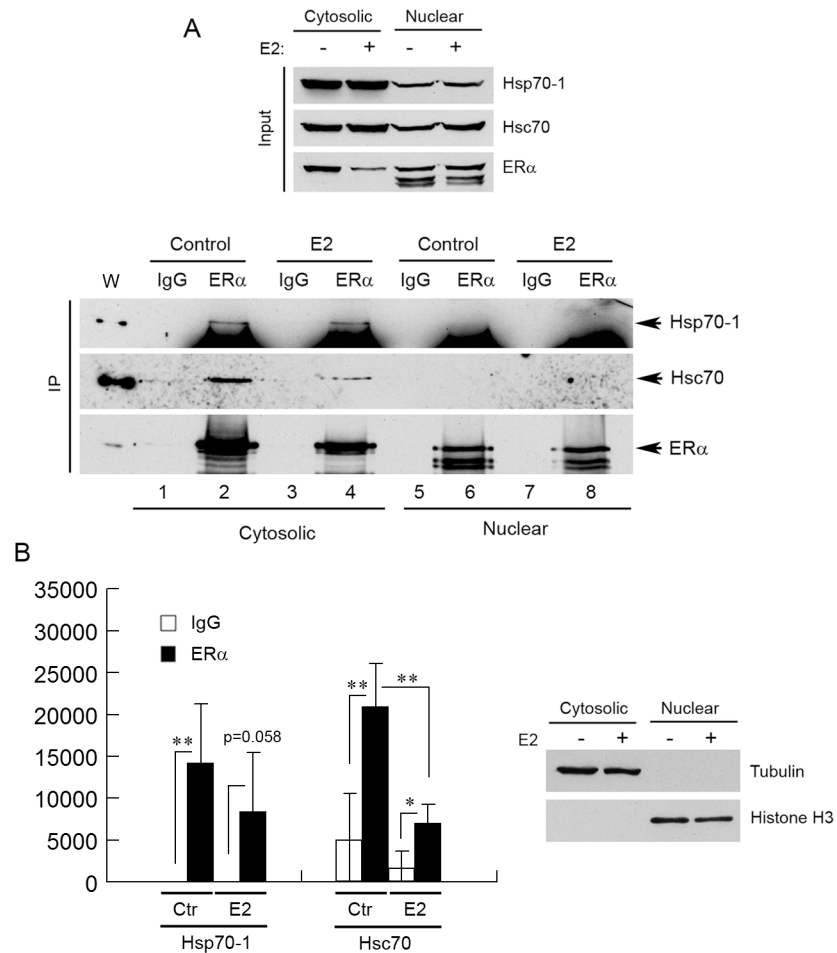


Fig 5. ER α interacts with Hsp70-1 and Hsc70 in the cytoplasm under conditions of hormone starvation/stimulation. (A) The MCF7 cells were cultured under hormone starvation conditions for 3–4 days and then treated with either 100 nM E2 or ethanol (control) for 24 h. The cytosolic and nuclear extracts of the treated cells were then immunoprecipitated by anti-ER α antibody or a control IgG, and the immunoprecipitated protein was analyzed by Western blotting with the indicated antibodies. (B) Left panel, quantification of Western blots. Only the Hsp70-1 and Hsc70 protein bands in the cytosolic fractions were quantified. Signal intensity values in the Western blot quantifications were arbitrary numbers obtained by analyzing the protein bands with ImageJ software. Values in the Western blot quantifications were the means \pm S.D. of four separate sample preparations. Right panel, validation of the cytosolic and nuclear fractionations. Tubulin and histone H3 were used as markers for the cytosolic and nuclear fractions, respectively. W, whole cell lysate. Ctr, control. * and ** denote $p < 0.05$ and $p < 0.01$, respectively.

doi:10.1371/journal.pone.0160312.g005

starvation and the subsequent hormone stimulation. The E2 treatment had no significant effect on the ER α -Hsp70-1 interaction, but significantly weakened the interaction between ER α and Hsc70 in the cytoplasm (Fig 5A, low panel; compare lane 4 with lane 2; Fig 5B, left panel). These results are consistent with the previous observations, which showed that Hsp70 was still associated with progesterone receptors in the presence of progesterone but the levels of the association decreased compared with in the absence of progesterone [64, 65]. Anti-ER α antibody did not precipitate any detectable amount of Hsp70-1 and Hsc70 from the nuclear fractions either in the absence or presence of E2 (Fig 5A, low panel; lanes 5–8). Compared with the results shown in Fig 2, which were obtained with the MCF7 cells cultured under conventional laboratory conditions (i.e., a culture medium supplemented with 5% FBS

and phenol red), the ER α -Hsp70-1/Hsc70 interactions observed under E2 starvation/stimulation conditions appeared to be different: under the former conditions the interactions were observed in both the cytoplasm and the nucleus (Fig 2) and under the latter conditions in the cytoplasm only (Fig 5). These results suggest that certain factors, potentially not just E2, in the culture media dictate whether ER α interacts with Hsp70-1/Hsc70 in the cytoplasm or the nucleus.

To examine how estrogens affect the association of Hsp70-1 and Hsc70 with chromatin, we cultured MCF7 cells under hormone-starvation conditions for 4 days and then treated the cells with either 100 nM E2 or ethanol (control) for 24 h, fractionated the treated cells into cytoplasmic (C), nuclear soluble (NS), transcriptionally active chromatin (Ch1) and inactive chromatin (Ch2) fractions, and analyzed those fractions with Western blotting. The E2 treatment caused significant reduction of ER α as a cytoplasmic protein and as a nuclear soluble protein, suggesting that E2 treatment causes translocation of ER α from the cytoplasm to the nucleus, and eventually the majority of the soluble nuclear ER α to chromatin (Fig 6). In addition, E2 significantly increased the distribution of Hsp90 α in the nucleus as nuclear soluble protein. Compared with the dynamic changes in ER α and Hsp90 α , E2 had no significant effect on the distribution of Hsp70-1 and Hsc70 among the different fractions (Fig 6).

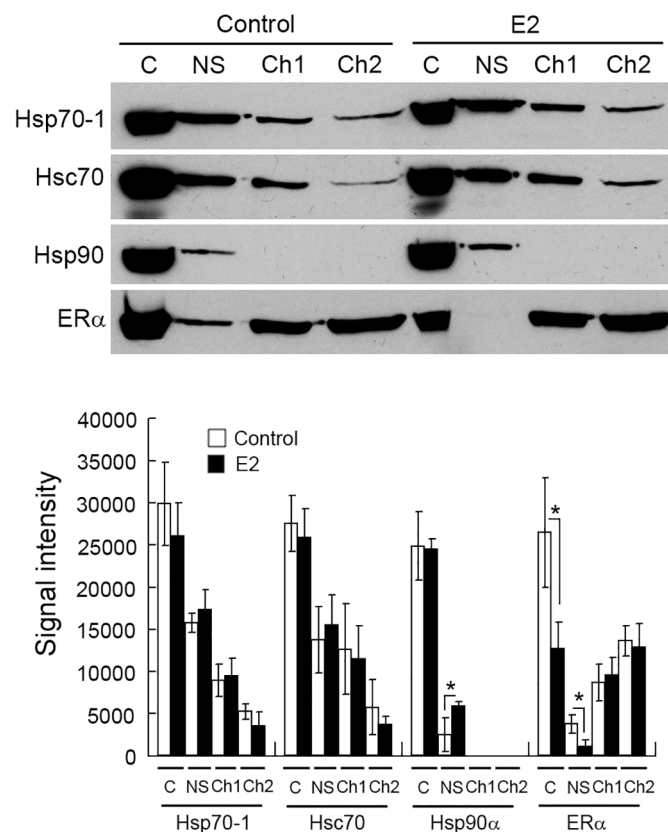


Fig 6. Estradiol does not affect the association of Hsp70-1 and Hsc70 with chromatin. The MCF7 cell extract was fractionated into cytoplasmic protein (C), nuclear soluble protein (NS), transcriptionally active chromatin (Ch1) and inactive chromatin (Ch2), and analyzed by Western blotting with the indicated antibodies (top panel). Signal intensity values in the Western blot quantifications were arbitrary numbers obtained by analyzing the protein bands with ImageJ software. Values in the Western blot quantifications were the means \pm S.D. of three separate sample preparations. * denotes $p < 0.05$.

doi:10.1371/journal.pone.0160312.g006

Discussion

Through a quantitative proteomic approach, we identified 21 Hsps and 3 Hsp cochaperones that associate with ER α . The most abundant Hsps that were identified to associate with ER α were four Hsp70 members, followed by two Hsp90 members and three Hsp110 members when 293T cells were cultured in “complete” medium. Within the Hsp70 family, Hsp70-1 and Hsc70 stood out as the most abundant Hsps that associate with ER α , followed by Grp75 that is localized in the mitochondria, and Grp78 that is localized in the endoplasmic reticulum. The two most common Hsp90 family members, Hsp90 α and Hsp90 β [53], were also identified to abundantly associate with ER α , though at much less abundant levels than the four Hsp70 family members. It is generally believed that ER α interacts with Hsp90 only in the absence of ligands, and dissociates from Hsp90 in the presence of ligands [1, 33, 57]. In this study, although we did not add any exogenous estrogenic ligands (such as E2) to the media for culturing the 293T cells for proteomic identification, we cultured the cells in “complete” medium that contains phenol red, which is known to act as a weak estrogen to stimulate proliferation of ER α -positive cells [66] and FBS, which contains steroid hormones [67]. In addition, we included 10 nM E2 in the lysis buffer for preparing total cellular protein for LC-MS/MS analysis. The identification of Hsp90 α and Hsp90 β as ER α interacting proteins under the present cell culture and affinity purification conditions suggests that Hsp90 could also complex with ER α , at least partially, in the presence of estrogenic ligands. This conclusion is consistent with the notion that the dynamic and transient interaction of steroid-bound SRs with Hsp90 may be required for the cytoplasmic-nuclear trafficking of SRs in cells [61].

Historically, the attention in studying the role of Hsps in regulating the assembly, trafficking, and transcriptional activity of ER α has been focused on Hsp90 [1, 33]. Through conventional liquid chromatography or affinity purification, it has been well established that Hsp90 interacts with ER α in a variety of tissue/cells in the absence of ligands [33]. Because of its role in controlling SRs including ERs, and a separate role in protecting oncoproteins, Hsp90 inhibitors are in clinical trials for treating cancer [23, 24]. Compared with Hsp90, much less is known about Hsp70 in regulating ER α . In this study, we found that Hsp70-1 and Hsc70 were the most abundant Hsps that associate with ER α (Table 1). Interestingly, despite that the majority of Hsp70-1 and Hsc70 were localized in the cytoplasm, comparable amounts of cytoplasmic and nuclear hsp70-1 and Hsc70 were precipitated by anti-ER α antibody (Fig 2). Furthermore, significant portions of Hsp70-1 and Hsc70 were associated with active chromatin and inactive chromatin (Fig 3), and the two Hsps interacted with ER α in both forms of the chromatins (Fig 4). These results are consistent with the observation that the association of Hsp70 with SRs does not affect DNA binding activity of SRs [68]. In contrast, Hsp90 α was almost exclusively localized in the cytoplasm and in the nucleus as non-chromatin-binding protein (Fig 3), which is consistent with the previous observations that SR-Hsp90 complexes are not associated with DNA and that dissociation of Hsp90 from SRs leads to DNA-binding of SRs [69, 70]. Unlike the ER α -Hsp90 association that is normally hormone-dependent [1, 33, 57], Hsp70 is still associated with SRs in the presence of steroid hormones [64, 65, 68, 71], which was also observed in this study (Fig 5). These results suggest that Hsp70 may play a dramatically different role in regulating ER biological activities compared with Hsp90. Perhaps, cells have evolved two distinct Hsp chaperone systems as repressors to keep ER α in the inactive states in transcription—one is “off-site” (not associated with chromatin) and ligand responsive, which is mediated by Hsp90, and one is “on-site” (associated with chromatin) and not/partially ligand responsive, which is mediated by Hsp70. If this is the case, it would be interesting to examine how these two chaperone systems interplay to regulate ER α transcriptional activities in a broad context such as tissue development and homeostasis.

In addition to functioning as nuclear receptors and transcription factors in the nucleus, ERs also act as signaling molecules in the plasma membrane and are localized in the mitochondria and the endoplasmic reticulum [72–74]. In this study, three mitochondrial Hsp members, Grp75, HSPE1, DNAJA3, were identified to associate with ER α . In particular, Grp75 was identified as a major ER α interactant (Table 1). When nuclear-gene-encoded proteins, such as ER α , are transported into mitochondria via posttranslational import, the proteins are imported into mitochondria in the unfolded states and need to be properly folded after the import. It would be interesting to determine whether Grp75, HSPE1, and DNAJA3 are merely responsible for folding imported ER α in the mitochondria or play additional roles in regulating ER α biological activities in the mitochondria. Several lines of evidence suggest that ERs may play important roles in the mitochondria. For example, it is known that a portion of cellular ERs are localized to mitochondria and the relative distribution of ERs into the mitochondrial pool is regulated by estrogens [47, 48, 75–78]. In addition, it has been shown that mitochondrial DNA contains estrogen response elements [79] and that mitochondrial structure and some important functions are influenced by estrogenic ligands. In addition to the mitochondria, ERs are also localized in the plasma membrane and the endoplasmic reticulum in the extra-nuclear compartments [72, 74, 80–82]. Posttranslational palmitoylation of ER can contribute to ER membrane localization [83]. Interestingly, both ER α and ER β were found to localize in the rough endoplasmic reticulum and secretory vesicles [82]. This raises the possibility that ERs could also be transported to the membranes through vesicular transport. However, virtually nothing is known about endoplasmic reticulum ER besides its localization. In this study, we found that Grp78, an Hsp that is localized in the endoplasmic reticulum, was abundantly associated with ER α (Table 1). At present, it is not clear whether the identification of Grp78 as a potential ER α interactant reflects a need of this Hsp in mediating ER α in this organelle.

Supporting Information

S1 Fig. Comparison of extraction of inactive chromatin with 600 mM NaCl and sonication.

There was an inconsistency between Figs 3 and 4 in the main text with regard to the relative content of ER α in inactive chromatin (Ch2). When inactive chromatin was extracted with 600 mM NaCl, which was the case for Fig 3, ER α content in inactive chromatin was the highest among the five fractions examined (Fig 3). However, when inactive chromatin was extracted with sonication, which was the case for Fig 4, ER α content was lower in inactive chromatin than in active chromatin (Fig 4, top panel). To examine whether the inconsistency was caused by different extraction methods, we extracted cytoplasmic (C), nuclear soluble (NS), and active chromatin (Ch1) from two populations of MCF7 cells as described in the main text, followed by extraction of inactive chromatin from the first population of cells with 600 mM NaCl and from the second population of cells with sonication. The results demonstrate that sonication extracted less ER α in inactive chromatin fraction compared to 600 mM NaCl extraction (S1 Fig), suggesting that the lower input ER α content in inactive chromatin fraction shown in the Fig 4 resulted from less efficient extraction of inactive chromatin by sonication compared to 600 mM NaCl extraction.

(TIFF)

S1 Table. The list of peptides identified for the heat shock proteins and cochaperones that associate with estrogen receptor alpha.

(XLS)

Author Contributions

Conceived and designed the experiments: AED ZZ JZ YD.

Performed the experiments: AED ZZ JZ YD.

Analyzed the data: AED JZ YD.

Contributed reagents/materials/analysis tools: AED ZZ JZ YD.

Wrote the paper: AED YD.

References

1. Picard D. Chaperoning steroid hormone action. *Trends Endocrinol Metab.* 2006; 17(6):229–35. PMID: [16806964](#)
2. Cheung J, Smith DF. Molecular chaperone interactions with steroid receptors: an update. *Mol Endocrinol.* 2000; 14(7):939–46. PMID: [10894145](#)
3. Pratt WB, Galigniana MD, Morishima Y, Murphy PJ. Role of molecular chaperones in steroid receptor action. *Essays Biochem.* 2004; 40:41–58. PMID: [15242338](#)
4. Chambraud B, Berry M, Redeuilh G, Chambon P, Baulieu EE. Several regions of human estrogen receptor are involved in the formation of receptor-heat shock protein 90 complexes. *J Biol Chem.* 1990; 265(33):20686–91. PMID: [2243115](#)
5. Segnitz B, Gehring U. Subunit structure of the nonactivated human estrogen receptor. *Proc Natl Acad Sci U S A.* 1995; 92(6):2179–83. PMID: [7892243](#)
6. Fliss AE, Benzeno S, Rao J, Caplan AJ. Control of estrogen receptor ligand binding by Hsp90. *J Steroid Biochem Mol Biol.* 2000; 72(5):223–30. PMID: [10822011](#)
7. Landel CC, Kushner PJ, Greene GL. The interaction of human estrogen receptor with DNA is modulated by receptor-associated proteins. *Mol Endocrinol.* 1994; 8(10):1407–19. PMID: [7854357](#)
8. Eckert RL, Mullick A, Rorke EA, Katzenellenbogen BS. Estrogen receptor synthesis and turnover in MCF-7 breast cancer cells measured by a density shift technique. *Endocrinology.* 1984; 114(2):629–37. PMID: [6690295](#)
9. Reid G, Hubner MR, Metivier R, Brand H, Denger S, Manu D, et al. Cyclic, proteasome-mediated turnover of unliganded and liganded ER α on responsive promoters is an integral feature of estrogen signaling. *Mol Cell.* 2003; 11(3):695–707. PMID: [12667452](#)
10. Fan M, Park A, Nephew KP. CHIP (carboxyl terminus of Hsc70-interacting protein) promotes basal and geldanamycin-induced degradation of estrogen receptor- α . *Mol Endocrinol.* 2005; 19(12):2901–14. PMID: [16037132](#)
11. Tateishi Y, Kawabe Y, Chiba T, Murata S, Ichikawa K, Murayama A, et al. Ligand-dependent switching of ubiquitin-proteasome pathways for estrogen receptor. *EMBO J.* 2004; 23(24):4813–23. PMID: [15538384](#)
12. Powell E, Wang Y, Shapiro DJ, Xu W. Differential requirements of Hsp90 and DNA for the formation of estrogen receptor homodimers and heterodimers. *J Biol Chem.* 2010; 285(21):16125–34. doi: [10.1074/jbc.M110.104356](#) PMID: [20353944](#)
13. Beato M, Klug J. Steroid hormone receptors: an update. *Hum Reprod Update.* 2000; 6(3):225–36. PMID: [10874567](#)
14. Inano K, Curtis SW, Korach KS, Omata S, Horigome T. Heat shock protein 90 strongly stimulates the binding of purified estrogen receptor to its responsive element. *J Biochem.* 1994; 116(4):759–66. PMID: [7883750](#)
15. Lindquist S, Craig EA. The heat-shock proteins. *Annu Rev Genet.* 1988; 22:631–77. PMID: [2853609](#)
16. Hunt C, Morimoto RI. Conserved features of eukaryotic hsp70 genes revealed by comparison with the nucleotide sequence of human hsp70. *Proc Natl Acad Sci U S A.* 1985; 82(19):6455–9. PMID: [3931075](#)
17. Kampinga HH, Hageman J, Vos MJ, Kubota H, Tanguay RM, Bruford EA, et al. Guidelines for the nomenclature of the human heat shock proteins. *Cell Stress Chaperones.* 2009; 14(1):105–11. doi: [10.1007/s12192-008-0068-7](#) PMID: [18663603](#)
18. Schlesinger MJ. Heat shock proteins. *J Biol Chem.* 1990; 265(21):12111–4. PMID: [2197269](#)
19. Dworniczak B, Mirault ME. Structure and expression of a human gene coding for a 71 kd heat shock 'cognate' protein. *Nucleic Acids Res.* 1987; 15(13):5181–97. PMID: [3037489](#)

20. Daugaard M, Rohde M, Jaattela M. The heat shock protein 70 family: Highly homologous proteins with overlapping and distinct functions. *FEBS Lett.* 2007; 581(19):3702–10. PMID: [17544402](#)
21. Terada K, Mori M. Human DnaJ homologs dj2 and dj3, and bag-1 are positive cochaperones of hsc70. *J Biol Chem.* 2000; 275(32):24728–34. PMID: [10816573](#)
22. Hattori H, Kaneda T, Lokeshwar B, Laszlo A, Ohtsuka K. A stress-inducible 40 kDa protein (hsp40): purification by modified two-dimensional gel electrophoresis and co-localization with hsc70(p73) in heat-shocked HeLa cells. *J Cell Sci.* 1993; 104 (Pt 3):629–38. PMID: [8314866](#)
23. Whitesell L, Lindquist SL. HSP90 and the chaperoning of cancer. *Nat Rev Cancer.* 2005; 5(10):761–72. PMID: [16175177](#)
24. Messaoudi S, Peyrat JF, Brion JD, Alami M. Recent advances in Hsp90 inhibitors as antitumor agents. *Anticancer Agents Med Chem.* 2008; 8(7):761–82. PMID: [18855578](#)
25. Sanchez ER. Chaperoning steroidal physiology: lessons from mouse genetic models of Hsp90 and its cochaperones. *Biochim Biophys Acta.* 2012; 1823(3):722–9. doi: [10.1016/j.bbamcr.2011.11.006](#) PMID: [22155719](#)
26. Taipale M, Jarosz DF, Lindquist S. HSP90 at the hub of protein homeostasis: emerging mechanistic insights. *Nat Rev Mol Cell Biol.* 2010; 11(7):515–28. doi: [10.1038/nrm2918](#) PMID: [20531426](#)
27. Kampinga HH, Craig EA. The HSP70 chaperone machinery: J proteins as drivers of functional specificity. *Nat Rev Mol Cell Biol.* 2010; 11(8):579–92. doi: [10.1038/nrm2941](#) PMID: [20651708](#)
28. Murphy ME. The HSP70 family and cancer. *Carcinogenesis.* 2013; 34(6):1181–8. doi: [10.1093/carcin/bgt111](#) PMID: [23563090](#)
29. Pick E, Kluger Y, Giltzane JM, Moeder C, Camp RL, Rimm DL, et al. High HSP90 expression is associated with decreased survival in breast cancer. *Cancer Res.* 2007; 67(7):2932–7. PMID: [17409397](#)
30. Zaarur N, Gabai VL, Porco JA Jr., Calderwood S, Sherman MY. Targeting heat shock response to sensitize cancer cells to proteasome and Hsp90 inhibitors. *Cancer Res.* 2006; 66(3):1783–91. PMID: [16452239](#)
31. Yaglom JA, Gabai VL, Sherman MY. High levels of heat shock protein Hsp72 in cancer cells suppress default senescence pathways. *Cancer Res.* 2007; 67(5):2373–81. PMID: [17332370](#)
32. Howe MK, Bodoor K, Carlson DA, Hughes PF, Alwarawrah Y, Loisel DR, et al. Identification of an allosteric small-molecule inhibitor selective for the inducible form of heat shock protein 70. *Chem Biol.* 2014; 21(12):1648–59. doi: [10.1016/j.chembiol.2014.10.016](#) PMID: [25500222](#)
33. Pratt WB, Toft DO. Steroid receptor interactions with heat shock protein and immunophilin chaperones. *Endocr Rev.* 1997; 18(3):306–60. PMID: [9183567](#)
34. Hickey E, Brandon SE, Smale G, Lloyd D, Weber LA. Sequence and regulation of a gene encoding a human 89-kilodalton heat shock protein. *Mol Cell Biol.* 1989; 9(6):2615–26. PMID: [2527334](#)
35. Grad I, Cederroth CR, Walicki J, Grey C, Barluenga S, Winssinger N, et al. The molecular chaperone Hsp90alpha is required for meiotic progression of spermatocytes beyond pachytene in the mouse. *PLoS One.* 2010; 5(12):e15770. doi: [10.1371/journal.pone.0015770](#) PMID: [21209834](#)
36. Voss AK, Thomas T, Gruss P. Mice lacking HSP90beta fail to develop a placental labyrinth. *Development.* 2000; 127(1):1–11. PMID: [10654595](#)
37. Echeverria PC, Briand PA, Picard D. A remodeled Hsp90 molecular chaperone ensemble with the novel co-chaperone Aarsd1 is required for muscle differentiation. *Mol Cell Biol.* 2016.
38. Ong SE, Blagoev B, Kratchmarova I, Kristensen DB, Steen H, Pandey A, et al. Stable isotope labeling by amino acids in cell culture, SILAC, as a simple and accurate approach to expression proteomics. *Mol Cell Proteomics.* 2002; 1(5):376–86. PMID: [12118079](#)
39. Zhu H, Pan S, Gu S, Bradbury EM, Chen X. Amino acid residue specific stable isotope labeling for quantitative proteomics. *Rapid Commun Mass Spectrom.* 2002; 16(22):2115–23. PMID: [12415544](#)
40. Zhou J, Du Y. Acquisition of resistance of pancreatic cancer cells to 2-methoxyestradiol is associated with the upregulation of manganese superoxide dismutase. *Mol Cancer Res.* 2012; 10(6):768–77. doi: [10.1158/1541-7786.MCR-11-0378](#) PMID: [22547077](#)
41. Liu L, Zhou J, Wang Y, Mason RJ, Funk CJ, Du Y. Proteome alterations in primary human alveolar macrophages in response to influenza A virus infection. *J Proteome Res.* 2012; 11(8):4091–101. doi: [10.1021/pr3001332](#) PMID: [22709384](#)
42. Du Y, Zhou J, Fan J, Shen Z, Chen X. Streamline proteomic approach for characterizing protein-protein interaction network in a RAD52 protein complex. *J Proteome Res.* 2009; 8(5):2211–7. doi: [10.1021/pr800662x](#) PMID: [19338310](#)
43. Du YC, Gu S, Zhou J, Wang T, Cai H, Macinnes MA, et al. The dynamic alterations of H2AX complex during DNA repair detected by a proteomic approach reveal the critical roles of Ca(2+)/calmodulin in

- the ionizing radiation-induced cell cycle arrest. *Mol Cell Proteomics*. 2006; 5(6):1033–44. PMID: [16522924](#)
44. Florens L, Carozza MJ, Swanson SK, Fournier M, Coleman MK, Workman JL, et al. Analyzing chromatin remodeling complexes using shotgun proteomics and normalized spectral abundance factors. *Methods*. 2006; 40(4):303–11. PMID: [17101441](#)
 45. Paoletti AC, Parmely TJ, Tomomori-Sato C, Sato S, Zhu D, Conaway RC, et al. Quantitative proteomic analysis of distinct mammalian Mediator complexes using normalized spectral abundance factors. *Proc Natl Acad Sci U S A*. 2006; 103(50):18928–33. PMID: [17138671](#)
 46. Zhang Y, Wen Z, Washburn MP, Florens L. Refinements to label free proteome quantitation: how to deal with peptides shared by multiple proteins. *Anal Chem*. 2010; 82(6):2272–81. doi: [10.1021/ac9023999](#) PMID: [20166708](#)
 47. Zhou Z, Zhou J, Du Y. Estrogen receptor beta interacts and colocalizes with HADHB in mitochondria. *Biochem Biophys Res Commun*. 2012; 427(2):305–8. doi: [10.1016/j.bbrc.2012.09.047](#) PMID: [23000159](#)
 48. Zhou Z, Zhou J, Du Y. Estrogen receptor alpha interacts with mitochondrial protein HADHB and affects beta-oxidation activity. *Mol Cell Proteomics*. 2012; 11(7):M111 011056.
 49. Yang L, Gal J, Chen J, Zhu H. Self-assembled FUS binds active chromatin and regulates gene transcription. *Proc Natl Acad Sci U S A*. 2014; 111(50):17809–14. doi: [10.1073/pnas.1414004111](#) PMID: [25453086](#)
 50. Henikoff S, Henikoff JG, Sakai A, Loeb GB, Ahmad K. Genome-wide profiling of salt fractions maps physical properties of chromatin. *Genome Res*. 2009; 19(3):460–9. doi: [10.1101/gr.087619.108](#) PMID: [19088306](#)
 51. Zybilov B, Mosley AL, Sardi ME, Coleman MK, Florens L, Washburn MP. Statistical analysis of membrane proteome expression changes in *Saccharomyces cerevisiae*. *J Proteome Res*. 2006; 5(9):2339–47. PMID: [16944946](#)
 52. Zhu W, Smith JW, Huang CM. Mass spectrometry-based label-free quantitative proteomics. *J Biomed Biotechnol*. 2010; 2010:840518. doi: [10.1155/2010/840518](#) PMID: [19911078](#)
 53. Sreedhar AS, Kalmar E, Csermely P, Shen YF. Hsp90 isoforms: functions, expression and clinical importance. *FEBS Lett*. 2004; 562(1–3):11–5. PMID: [15069952](#)
 54. Vos MJ, Hageman J, Carra S, Kampinga HH. Structural and functional diversities between members of the human HSPB, HSPH, HSPA, and DNAJ chaperone families. *Biochemistry*. 2008; 47(27):7001–11. doi: [10.1021/bi800639z](#) PMID: [18557634](#)
 55. Fathallah DM, Cherif D, Dellagi K, Arnaout MA. Molecular cloning of a novel human hsp70 from a B cell line and its assignment to chromosome 5. *J Immunol*. 1993; 151(2):810–3. PMID: [8335910](#)
 56. Kaneko Y, Kimura T, Kishishita M, Noda Y, Fujita J. Cloning of *apg-2* encoding a novel member of heat shock protein 110 family. *Gene*. 1997; 189(1):19–24. PMID: [9161406](#)
 57. Carrello A, Allan RK, Morgan SL, Owen BA, Mok D, Ward BK, et al. Interaction of the Hsp90 cochaperone cyclophilin 40 with Hsc70. *Cell Stress Chaperones*. 2004; 9(2):167–81. PMID: [15497503](#)
 58. Ratajczak T, Carrello A. Cyclophilin 40 (CyP-40), mapping of its hsp90 binding domain and evidence that FKBP52 competes with CyP-40 for hsp90 binding. *J Biol Chem*. 1996; 271(6):2961–5. PMID: [8621687](#)
 59. Storer CL, Dickey CA, Galigniana MD, Rein T, Cox MB. FKBP51 and FKBP52 in signaling and disease. *Trends Endocrinol Metab*. 2011; 22(12):481–90. doi: [10.1016/j.tem.2011.08.001](#) PMID: [21889356](#)
 60. Klinge CM, Brolly CL, Bambara RA, Hilf R. hsp70 is not required for high affinity binding of purified calf uterine estrogen receptor to estrogen response element DNA in vitro. *J Steroid Biochem Mol Biol*. 1997; 63(4–6):283–301. PMID: [9459195](#)
 61. Pratt WB. The role of the hsp90-based chaperone system in signal transduction by nuclear receptors and receptors signaling via MAP kinase. *Annu Rev Pharmacol Toxicol*. 1997; 37:297–326. PMID: [9131255](#)
 62. Perissi V, Rosenfeld MG. Controlling nuclear receptors: the circular logic of cofactor cycles. *Nat Rev Mol Cell Biol*. 2005; 6(7):542–54. PMID: [15957004](#)
 63. Rosenfeld MG, Lunyak VV, Glass CK. Sensors and signals: a coactivator/corepressor/epigenetic code for integrating signal-dependent programs of transcriptional response. *Genes Dev*. 2006; 20(11):1405–28. PMID: [16751179](#)
 64. Kost SL, Smith DF, Sullivan WP, Welch WJ, Toft DO. Binding of heat shock proteins to the avian progesterone receptor. *Mol Cell Biol*. 1989; 9(9):3829–38. PMID: [2779568](#)
 65. Smith DF, Faber LE, Toft DO. Purification of unactivated progesterone receptor and identification of novel receptor-associated proteins. *J Biol Chem*. 1990; 265(7):3996–4003. PMID: [2303491](#)

66. Berthois Y, Katzenellenbogen JA, Katzenellenbogen BS. Phenol red in tissue culture media is a weak estrogen: implications concerning the study of estrogen-responsive cells in culture. *Proc Natl Acad Sci U S A*. 1986; 83(8):2496–500. PMID: [3458212](#)
67. Darbre P, Yates J, Curtis S, King RJ. Effect of estradiol on human breast cancer cells in culture. *Cancer Res*. 1983; 43(1):349–54. PMID: [6847777](#)
68. Srinivasan G, Patel NT, Thompson EB. Heat shock protein is tightly associated with the recombinant human glucocorticoid receptor:glucocorticoid response element complex. *Mol Endocrinol*. 1994; 8(2):189–96. PMID: [8170475](#)
69. Denis M, Poellinger L, Wikstrom AC, Gustafsson JA. Requirement of hormone for thermal conversion of the glucocorticoid receptor to a DNA-binding state. *Nature*. 1988; 333(6174):686–8. PMID: [3374615](#)
70. Sanchez ER, Meshinchi S, Tienrungroj W, Schlesinger MJ, Toft DO, Pratt WB. Relationship of the 90-kDa murine heat shock protein to the untransformed and transformed states of the L cell glucocorticoid receptor. *J Biol Chem*. 1987; 262(15):6986–91. PMID: [3294824](#)
71. Sanchez ER, Hirst M, Scherrer LC, Tang HY, Welsh MJ, Harmon JM, et al. Hormone-free mouse glucocorticoid receptors overexpressed in Chinese hamster ovary cells are localized to the nucleus and are associated with both hsp70 and hsp90. *J Biol Chem*. 1990; 265(33):20123–30. PMID: [2243084](#)
72. Govind AP, Thampan RV. Membrane associated estrogen receptors and related proteins: localization at the plasma membrane and the endoplasmic reticulum. *Mol Cell Biochem*. 2003; 253(1–2):233–40. PMID: [14619974](#)
73. Hammes SR, Levin ER. Extranuclear steroid receptors: nature and actions. *Endocr Rev*. 2007; 28(7):726–41. PMID: [17916740](#)
74. Levin ER. Plasma membrane estrogen receptors. *Trends Endocrinol Metab*. 2009; 20(10):477–82. doi: [10.1016/j.tem.2009.06.009](#) PMID: [19783454](#)
75. Chen JQ, Delannoy M, Cooke C, Yager JD. Mitochondrial localization of ERalpha and ERbeta in human MCF7 cells. *Am J Physiol Endocrinol Metab*. 2004; 286(6):E1011–22. PMID: [14736707](#)
76. Pedram A, Razandi M, Wallace DC, Levin ER. Functional estrogen receptors in the mitochondria of breast cancer cells. *Mol Biol Cell*. 2006; 17(5):2125–37. PMID: [16495339](#)
77. Yager JD, Chen JQ. Mitochondrial estrogen receptors—new insights into specific functions. *Trends Endocrinol Metab*. 2007; 18(3):89–91. PMID: [17324583](#)
78. Yang SH, Liu R, Perez EJ, Wen Y, Stevens SM Jr., Valencia T, et al. Mitochondrial localization of estrogen receptor beta. *Proc Natl Acad Sci U S A*. 2004; 101(12):4130–5. PMID: [15024130](#)
79. Demonacos CV, Karayanni N, Hatzoglou E, Tsiriyiotis C, Spandidos DA, Sekeris CE. Mitochondrial genes as sites of primary action of steroid hormones. *Steroids*. 1996; 61(4):226–32. PMID: [8733006](#)
80. Blaustein JD, Lehman MN, Turcotte JC, Greene G. Estrogen receptors in dendrites and axon terminals in the guinea pig hypothalamus. *Endocrinology*. 1992; 131(1):281–90. PMID: [1612006](#)
81. Peralta Soler A, Aoki A. Immunocytochemical detection of estrogen receptors in a hormone-unresponsive mammary tumor. *Histochemistry*. 1989; 91(4):351–6. PMID: [2732099](#)
82. Gonzalez M, Reyes R, Damas C, Alonso R, Bello AR. Oestrogen receptor alpha and beta in female rat pituitary cells: an immunochemical study. *Gen Comp Endocrinol*. 2008; 155(3):857–68. PMID: [18067893](#)
83. Pedram A, Razandi M, Sainson RC, Kim JK, Hughes CC, Levin ER. A conserved mechanism for steroid receptor translocation to the plasma membrane. *J Biol Chem*. 2007; 282(31):22278–88. PMID: [17535799](#)