

## UC Irvine

### UC Irvine Previously Published Works

**Title**

Spinal NGF Restores Opioid Sensitivity in Neuropathic Rats: Possible Role of NGF as a Regulator of CCK-Induced Anti-Opioid Effects

**Permalink**

<https://escholarship.org/uc/item/1w46p18r>

**Journal**

Pain Research and Management, 5(1)

**ISSN**

1203-6765

**Authors**

Cahill, Catherine M  
Coderre, Terence J

**Publication Date**

2000

**DOI**

10.1155/2000/347212

**Copyright Information**

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

# Spinal NGF restores opioid sensitivity in neuropathic rats: Possible role of NGF as a regulator of CCK-induced anti-opioid effects

Catherine M Cahill PhD<sup>1</sup>, Terence J Coderre PhD<sup>2,3,4</sup>

CM Cahill, TJ Coderre.

**Spinal NGF restores opioid sensitivity in neuropathic rats: Possible role of NGF as a regulator of CCK-induced anti-opioid effects.**

**Pain 2000;5(1):49-57.**

The breadth of peripheral effects produced by nerve growth factor (NGF) in nociceptive processing has been well documented. However, less is known about the functional significance of central NGF in nociceptive transmission. The effect of NGF on the nervous system is dependent on the developmental stage. During the prenatal developmental period, NGF is critical for survival of nociceptors; in the postnatal period it regulates the expression of nociceptor phenotype, and in the adult it contributes to pain following an inflammatory insult. The implications for central NGF in the expression and regulation of spinal neuropeptides that are involved in pain mechanisms are reviewed. Knowledge has been gained by studies using peripheral nerve injury models that cause a deprivation of central NGF. These models also give rise to the development of pain syndromes, which encompass spontaneous pain, hyperalgesia and allodynia, routinely referred to as neuropathic pain. These models provide an approach for examining the contribution of central NGF to nociceptive transmission. Chronic pain emanating from a nerve injury is typically refractory to traditional analgesics such as opioids. Recent evidence suggests that supplementation of spinal NGF restores morphine-induced antinociception in an animal model of neuropathic pain. This effect appears to

be mediated by alterations in spinal levels of cholecystokinin. The authors hypothesize that NGF is critical in maintaining neurochemical homeostasis in the spinal cord of nociceptive neurons, and that supplementation may be beneficial in restoring and/or maintaining opioid analgesia in chronic pain conditions resulting from traumatic nerve injury.

**Key Words:** *Allodynia; Cholecystokinin; Nerve growth factor; Opiate; Pain*

**Le NGF spinal rétablit la sensibilité aux opioïdes chez les rats neuropathiques : rôle possible du NGF comme régulateur des effets anti-opioïdes induits par la cholécystokinine**

**RÉSUMÉ :** L'étendue des effets périphériques produits par le facteur de croissance nerveuse (NGF) dans le processus nociceptif est bien documentée. Cependant, la signification fonctionnelle du NGF central dans la transmission nociceptive est moins connue. L'effet du NGF sur le système nerveux dépend du stade de développement de ce système. Pendant la période de développement prénatale, le NGF est essentiel à la survie des nocicepteurs ; dans la période postnatale, il régule l'expression du phénotype des nocicepteurs, et chez l'adulte, il contribue à la douleur résultant d'une atteinte inflammatoire. Les

*voir page suivante*

<sup>1</sup>Department of Neurology and Neurosurgery, Montreal Neurological Institute, McGill University, <sup>2</sup>Pain Mechanisms Laboratory, Clinical Research Institute of Montreal, <sup>3</sup>Department of Psychology, McGill University, and <sup>4</sup>Centre de recherche en sciences neurologiques et Département de médecine, Université de Montréal, Montréal, Québec

Correspondence and reprints: Catherine M Cahill, Department of Neurology and Neurosurgery, Montreal Neurological Institute, 3801 University Street, Montreal, Quebec H3A 2B4. Telephone 514-398-1913, fax 514-398-5871, e-mail cmcahill@yahoo.com

implications du NGF central dans l'expression et dans la régulation des neuropeptides spinaux qui sont impliqués dans les mécanismes de la douleur sont passées en revue. Les connaissances se sont aussi améliorées en procédant à des études utilisant des modèles de lésions nerveuses périphériques qui entraînent une carence de NGF central. Ces modèles entraînent aussi le développement de syndromes douloureux, qui englobent la douleur spontanée, l'hyperalgie et l'allodynie, communément appelés douleur neuropathique. Ces modèles fournissent une approche pour étudier la contribution du NGF central à la transmission nociceptive. Une douleur chronique provenant d'une lésion nerveuse est typiquement réfractaire aux analgésiques classiques

comme les opioïdes. Des données récentes laissent à penser que l'administration d'un supplément de NGF spinal rétablit l'antinociception induite par la morphine dans un modèle animal de douleur neuropathique. Cet effet semble être médié par des modifications survenant dans les niveaux de cholécystokinine spinale. Les auteurs émettent l'hypothèse que le NGF est essentiel au maintien de l'homéostasie neurochimique des neurones nociceptifs dans la moelle épinière et que l'administration d'un supplément de NGF pourrait permettre de rétablir et/ou de maintenir une analgésie opioïde dans les cas de douleur chronique résultant d'une atteinte nerveuse d'origine traumatique.

**I**njury to the peripheral or central nervous system (CNS) gives rise to the development of pain syndromes that usually include spontaneous pain, hyperalgesia and allodynia (1,2), routinely referred to as neuropathic pain. Following peripheral nerve injury in humans, neuropathic pain develops, including mechanical allodynia, defined as perception of pain in response to a normally innocuous tactile stimulus (3). Several animal models of neuropathic pain have been developed to investigate the mechanisms and pathologies that precipitate chronic pain (4-6). The development of neuropathic pain has been attributed to an increase in excitability, a decrease in inhibition and structural reorganization of neurons within the dorsal spinal cord (7) (Table 1). One of the morphological changes that occurs following damage to a peripheral nerve is the sprouting of myelinated cutaneous A-beta fibres into lamina II of the dorsal spinal cord (8-12). This creates close proximity between fibres that normally mediate nonpainful mechanical information and primary afferents that are involved in nociceptive transmission. In fact, one study reported that the A fibres that sprout within the superficial lamina make synaptic connections (11). It has been hypothesized that this morphological change underlies the expression of mechanical allodynia. One might speculate that the activation of large afferent fibres could cause the depolarization of small diameter afferent nerve terminals, and thus induce the release of transmitters and neuromodulators involved in nociceptive transmission. Therefore, innocuous mechanical stimuli could give rise to a painful response via the new synaptic connections made between large and small diameter afferent terminals within the superficial dorsal horn. It is not known what triggers the morphological change; however, there is a correlation between the reduction in the retrograde supply of peripherally produced trophic factors (including nerve growth factor [NGF]) and nerve injury-induced effects. Certainly, NGF is reduced in sensory neurons following axotomy (13). We review evidence that NGF can reverse some of the adverse effects produced by peripheral nerve injury.

The treatment of neuropathic pain is considered to be highly contentious, but a review evaluated various existing treatments (14). Generally, neuropathic pain states are unresponsive to traditional analgesics such as opioids (15-17). Several animal studies have confirmed clinical observations that opioids have little or no therapeutic benefit in alleviating

**TABLE 1**  
**Neuronal and molecular mechanisms of neuropathic pain**

Afferent nerve terminal	Release of local factors (cytokines, nerve growth factor)
	Nerve sprouting
	Increased sensitivity of neuronal sprouts to mechanical, chemical and thermal stimuli
Dorsal root ganglia	Spontaneous activity
	Increased innervation of A fibres by sympathetic terminals
	Increased evoked activity
Spinal cord	Sprouting of large afferent terminals into 'nociceptive lamina'
	Expansion of receptive fields
	Changes in neuropeptide levels and their receptors
	Central sensitization

neuropathic pain. Thus, in rats that exhibit nerve injury-induced allodynia and hyperalgesia, intrathecal morphine was found to be ineffective at increasing mechanical response thresholds, or withdrawal latencies to radiant heat or cold water (18-22). Various hypotheses have been formulated to explain the lack of opioid effectiveness, including a reduction in the number of opioid receptors, and activation of *N*-methyl-D-aspartate (NMDA) and/or cholécystokinine (CCK) receptors.

#### CCK IN NOCICEPTION

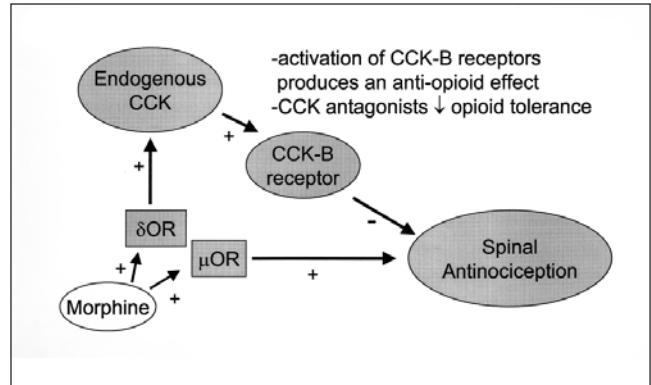
CCK has been demonstrated to be an important modulator within the mammalian nervous system, including having a role in the transmission and modulation of nociceptive information (23,24). The predominant form of CCK in the brain is an eight-amino acid residue peptide (CCK<sub>26-33</sub>) that exists in both sulphated and, to a lesser extent, desulphated forms. Pharmacological studies have identified two major subtypes of CCK receptors based on their ability to recognize sulphated or desulphated CCK (25,26). This classification was confirmed by the recent cloning of both CCK-A and CCK-B receptors (27). Anatomically, CCK and its receptors are present in various areas of the CNS that are involved in nociceptive processing. CCK mRNA (28,29) and peptides (30) have been identified in small and medium sized dorsal root gan-

glion (DRG) cells, although other studies using both *in situ* hybridization (28,29) and immunocytochemical (31) techniques have found very few DRG cells expressing CCK. Others have claimed that CCK is not normally present in normal DRG cells in the rat (32,33). In the rat dorsal horn of the spinal cord, numerous interneurons and descending fibres contain CCK (29,34,35).

CCK differs from most other neuropeptides that modulate nociceptive transmission in that it appears to act indirectly by interaction with the opioid system. The hypothesis that CCK may be an anti-opioid peptide is derived from studies demonstrating that exogenous application of CCK attenuates the analgesic effect of morphine and beta-endorphin (36,37). This observation has since been confirmed by many reports that used both behavioural and electrophysiological nociceptive techniques. Under normal conditions, CCK causes a marked inhibition of the antinociceptive effects of morphine and selective mu opioid receptor agonists (38,39), and attenuates morphine-induced inhibition of C fibre-evoked discharges of dorsal horn nociceptive neurons (37). Endogenous CCK tonically inhibits opioid-induced antinociception, and selective CCK antagonists potentiate morphine-induced antinociception (23,24,40-45). Furthermore, an antisense oligonucleotide directed against the CCK-B receptor mRNA was shown to enhance morphine-induced antinociception (46), suggesting that CCK produces a tonic inhibition of morphine-induced analgesic effects through an action at CCK-B receptors. The results of a recent clinical study underscore the contribution of CCK and its anti-opioid effects in chronic pain states because the CCK receptor antagonist proglumide increased the analgesic effect of morphine in some patients with chronic benign pain (47).

Many studies have demonstrated that opioids enhance the release of CCK through activation of opioid receptors on CCK-containing neurons (Figure 1). Recently, morphine was shown to evoke the release of CCK from cortical regions via activation of a delta-opioid receptor (48). High concentrations of morphine or a delta-opioid receptor agonist, (D-Ser8)-leucine enkephalin-Thr, were shown to enhance a calcium-dependent release of CCK from the rat substantia nigra (49), and [D-Ala<sup>2</sup>]deltorphin augmented CCK release from dorsal horn lumbar spinal cord slices (50). This excitatory action of opioids via activation of the delta-opioid receptor subtype has also been implicated in the development of opioid tolerance.

CCK does not elicit hyperalgesic effects, nor do CCK antagonists produce antinociception, indicating that endogenous CCK has no tonic inhibition on endogenous opioids (51). The mechanism by which CCK produces its anti-opioid effect remains elusive, although many hypotheses have been formulated. Within the CNS, the distribution of CCK parallels that of the endogenous opioids within the pain processing areas (52-55), providing anatomical evidence that a functional relationship may exist between these two transmitter systems. One of the possible mechanisms of CCK's anti-opioid activity was proposed to be consequential to CCK producing an attenuation of the binding affinity of mu



**Figure 1** Schematic representation of the interaction between cholecystokinin (CCK) and the opioid system at the level of the spinal cord. Chronic administration of morphine has been shown to increase endogenous CCK via activation of the delta opioid receptor (OR). In turn, CCK decreases the levels of endogenous enkephalins and activates CCK-B receptors to elicit an anti-opioid effect, resulting in the attenuation of morphine-induced antinociception. Blockade of CCK-B receptors with selective antagonists has been shown to attenuate the development of tolerance to opioid-induced antinociception

opioid receptors for its ligands (56). It has also been proposed that CCK's anti-opioid effects involve interactions between intracellular signalling cascades following activation of both CCK and mu opioid receptors (57). Although anatomical studies have yet to verify the coexistence of CCK-B and mu opioid receptors on the same nerve terminal, CCK-B receptor binding sites are present on capsaicin-sensitive small diameter primary afferent neurons (58). Moreover, CCK reverses the effects of mu opioid agonists in a whole cell patch clamp recording of acutely dissociated DRG neurons (59). Together, these studies strongly suggest the colocalization of CCK-B and opioid receptors, and the occurrence of CCK-opioid interactions within the same primary afferent neuron.

It has been established that CCK receptor signalling occurs predominantly via the phospholipase C-1,4,5-inositol triphosphate/1,2-diacylglycerol (IP<sub>3</sub>/DAG)-protein kinase C (PKC) pathway (27). CCK receptors activate pertussis toxin-sensitive G proteins that are coupled to membrane-bound phospholipase C, which in turn causes an increase in IP<sub>3</sub> and the subsequent release of intracellular calcium. DAG activates PKC, resulting in its translocation to the membrane, where it can phosphorylate various proteins. It has been shown that stimulation of PKC can result in the phosphorylation and desensitization of opioid receptors (60-62). It is highly possible that this signalling cascade may underlie CCK's anti-opioid effects because it has recently been demonstrated that local pretreatment with a PKC inhibitor, calphostin C, abolished the inhibitory effects of CCK on opioid-induced antinociception (63).

#### CCK INCREASES FOLLOWING NERVE INJURY

It was hypothesized that one of the possible reasons for the relative refractory analgesic efficacy of opioids in neuropathic pain may be an increase in CCK in the spinal cord. Thus, the lack of morphine effectiveness in alleviating neuropathic nociception may partially result from an increase in

this anti-opioid peptide following peripheral nerve injury. Indeed, intrathecal co-administration of a CCK antagonist and morphine has been found to produce an antiallodynic effect in nerve-injured rats, while intrathecal morphine alone was without effect (64). Moreover, it was shown that CCK antagonists could potentiate the antihyperalgesic effect of morphine in nerve-injured rats, although this effect varied among various animal models of neuropathic pain (65). Others have demonstrated that systemic administration of a CCK-B antagonist prevents the development of opioid insensitivity in an animal model of peripheral neuropathy (66). In keeping with the CCK hypothesis, in the flexor-reflex model of spinalized rats, systemic morphine exhibited a reduced antinociceptive potency in axotomized rats compared with normal rats, and administration of a CCK-B antagonist was found to potentiate the effect of morphine in this model (21). It was suggested that the anti-opioid effect elicited by CCK may be due to tonic inhibition of enkephalin release because the enhanced effect of morphine with a CCK-B antagonist was prevented by pretreatment with a selective delta-opioid receptor antagonist (45,64). Supporting evidence was provided by another study that showed that the systemic administration of a CCK-B receptor antagonist produced antiallodynic effects that were naloxone reversible in a model of chronic pain induced by a spinal cord lesion (67).

Changes in CCK levels have been shown to occur following peripheral nerve injury, with a dramatic upregulation of CCK-like material and CCK mRNA in the rat occurring after sciatic nerve transection (29,68). In situ hybridization studies have also shown a dramatic increase in the expression of CCK-B receptor mRNA in rat DRG neurons after peripheral nerve section (69) and in primary afferents after unilateral section of the sciatic nerve (20). One of the mechanisms for the increase in CCK following nerve injury may be the enhanced activity at NMDA receptors, because the activation of NMDA receptors was found to elicit the release of CCK-like immunoreactivity from rat cerebral cortex *in vivo* (70), from cortical slices (71) and, more recently, from synaptosomes (72). These studies provide a rationale for the hypothesis that the increase in the synthesis and release of CCK from excitatory interneurons in the dorsal spinal cord following peripheral nerve injury may impede opioid-induced antinociception (73,74).

### NGF IN NOCICEPTION

NGF has an established role in the survival, differentiation, development and maintenance of phenotype for small diameter, primary afferent neurons involved in nociceptive transmission (75). Nociceptive neurons are generally thought to express the NGF-selective trkA receptor. In the absence of NGF, C and A-delta fibres are absent in sensory ganglia, and their corresponding nerve endings are missing both in the periphery and spinal cord (76,77). Overexpression of NGF produces the opposite effect, resulting in the hypertrophy of small diameter, primary afferent neurons innervating the skin (77-79), a condition that produces thermal hyperalgesia in the affected animal (80). The scope of NGF's action

in establishing and maintaining nociceptors for normal pre- and postnatal development is well documented (81).

The contribution of NGF to inflammatory hyperalgesia was suggested to be mediated by its activation of high affinity trkA receptors on mast cells and primary sensory neurons (82). Peripherally produced NGF normally maintains the sensitivity of nociceptive sensory neurons, but in some inflammatory states, an increase in NGF occurs in the skin (83). The augmentation of NGF appears to be partly responsible for the hyperalgesia and allodynia that typically accompany such an insult. Thus, neutralization of NGF during inflammation with the use of either antibodies directed at NGF (84,85) or trkA-immunoglobulin G immunoadhesion molecules for NGF (86) attenuates hyperalgesia. Moreover, NGF is sufficient to induce hyperalgesia because administration either locally or systemically induces a decrease in thermal nociceptive thresholds (85,87). Similarly, it was demonstrated that NGF was necessary for the mechanical hyperalgesia that ensues following an inflammatory insult (88). The peripheral and central mechanisms implicated in NGF-induced hyperalgesia have been extensively reviewed (81).

NGF plays a role in the dynamic control of neuropeptide levels in adult sensory neurons (89-91) and, therefore, may contribute to inflammatory hyperalgesia by altering the release of peptides from sensory neurons. The role of peripheral NGF in nociception is demonstrated in studies showing that anti-NGF or trkA fusion molecules attenuate Freund's adjuvant-induced increase in preprotachykinin A mRNA (92), substance P and calcitonin gene-related peptide (CGRP) (86,91).

### ALTERATIONS IN NGF FOLLOWING NERVE INJURY

Although NGF is not necessary for neuronal survival in adults, it does appear to influence neuronal growth following denervation induced by peripheral nerve damage. In sensory systems, NGF is specifically taken up along peripheral and central processes of sensory neurons and retrogradely transported to the cell body (93). The depletion of NGF in the spinal dorsal horn following sciatic nerve axotomy (10) or chronic constriction injury (12) is thought to precipitate collateral sprouting of large diameter afferent fibres of laminae III to V into lamina II of the spinal cord. In support of this hypothesis, it was discovered that supplementation of spinal NGF by continuous intrathecal infusion suppressed the sprouting of these axon terminals into neighbouring denervated lamina (94). Sciatic nerve transection was shown to decrease the density of NGF binding sites on DRG neurons, and intrathecal NGF infusion partially reversed this reduction but did not influence NGF binding to neurons with intact axons (95).

Studies have demonstrated that disruption of NGF transport is correlated with changes in neuropeptide levels in the spinal cord and DRG that occur following peripheral nerve axotomy or constriction injury (95,96). Moreover, continuous infusion of NGF to the proximal stump of a transected sciatic nerve mitigates some of the morphological, biochemical and electrophysiological alterations in axotomized DRG



TABLE 2

Percentage maximum possible effect (MPE) produced by intrathecal morphine (20 µg) obtained on day 14 following sciatic nerve constriction of nerve-injured animals

Stimulus	% MPE of intrathecal morphine (20 µg) in neuropathic rats (day 16 after injury)	
	Chronic spinal IgG	Chronic spinal NGF
Thermal plantar test	12.5±2.80	35.2±4.50*
Cold water response frequency	20.6±1.59	74.7±10.1**
Cold water response duration	21.9±4.80	87.2±15.3**
50% Von Frey threshold	26.1±3.86	59.4±10.3*

Values are means ± SEM (n = five to nine per group). Statistical analysis using a paired t test revealed that mechanical response thresholds, and withdrawal latencies to either cold or heat following intrathecal morphine in nerve growth factor (NGF)-treated rats were significantly increased compared with pre-morphine values. \*P=0.05; \*\*P=0.01. IgG Immunoglobulin G

perikarya (97-102). Several studies have provided evidence that NGF is involved in neural, anatomical and molecular plasticity of primary afferents (103-106).

Although the occurrence of collateral sprouting and the decrease in spinal NGF are accepted, the clinical relevance of these events elicited in animal models of neuropathic pain is unclear. However, a functional correlate is suggested in that NGF supplementation attenuates injury-induced allodynia and hyperalgesia (107,108). There is also evidence that NGF may be neuroprotective to DRG neurons against drug- and diabetes-induced neuropathies (109). Beneficial effects of NGF supplementation have also been reported in alleviating neuropathic pain, in that infusion of NGF directly on the sciatic nerve prevents the development of thermal hyperalgesia and partially blocks mechanical allodynia in a sciatic nerve constriction model (110,111). However, local administration of anti-NGF decreased the severity of autotomy and blocked collateral sprouting, suggesting that neutralization of peripheral NGF may also be advantageous in blocking neuropathic pain (111).

### NGF REVERSES CCK-INDUCED OPIOID INSENSITIVITY

As discussed above, many studies have demonstrated a decrease in NGF in the sciatic nerve and central terminals in animal models of neuropathic pain (112). However, Verge and colleagues (105) demonstrated that delayed chronic intrathecal administration of NGF counteracted sciatic nerve constriction-induced changes in neuropeptide content, including the injury-induced increase in CCK. The phenotypic changes displayed within sensory neurons of the dorsal horn can be pre-empted by supplementation with NGF (95,113-115). Thus, nerve injury-induced increases in CCK may partially mediate the lack of morphine effectiveness. We propose that supplementation of NGF may help to restore CCK levels to preinjury levels, thus allowing morphine to elicit its full antinociceptive actions. Moreover, other studies have demonstrated that NGF can regulate CCK levels. Intraventricular administration of NGF has been shown to decrease

TABLE 3

Percentage change in morphine-induced antinociception by cholecystokinin (CCK)-B receptor antagonist

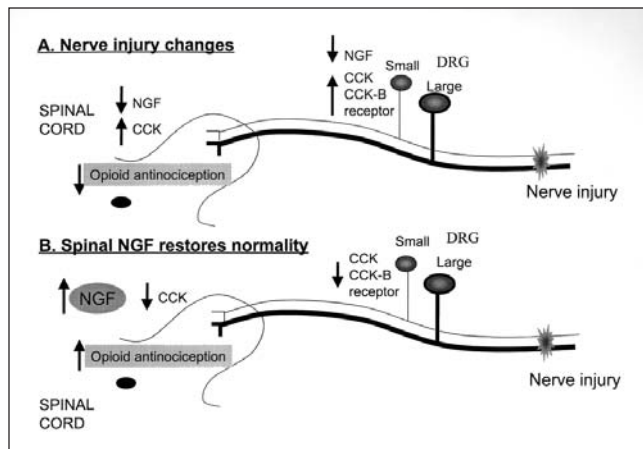
Immunoglobulin G	Antinerve growth factor
8.3±0.72	84.8±10.9*

Percentage change in morphine (5 µg)-induced antinociception following pre-treatment of intrathecal vehicle or CCK receptor antagonist (LY225910, 10 nmol) in rats chronically infused for seven days with intrathecal immunoglobulin G or antinerve growth factor (NGF) via osmotic minipumps. Statistical analysis using a paired t test revealed that the percentage increase in antinociception elicited by intrathecal morphine following LY225910 in anti-NGF treated rats was significantly increased compared with that in IgG-treated rats. \*P=0.05

CCK levels within the hypothalamus (115). This may explain the observation in mice overexpressing NGF that there is an enhanced efficacy of morphine following induction of thermal hyperalgesia.

We have confirmed the observation that intrathecal morphine is ineffective in increasing mechanical response thresholds, latencies to radiant heat or sensitivity to cold water in rats exhibiting nerve injury-induced allodynia and hyperalgesia (18-22). Nevertheless, delayed intrathecal NGF infusion was beneficial in restoring morphine-induced anti-hyperalgesic and antiallodynic effects indicative of neuropathic pain (unpublished data). In the study by Cahill andCoderre (unpublished data), delayed chronic intrathecal infusion of NGF could not alleviate decreased nociceptive thresholds in neuropathic rats, it only influenced morphine sensitivity (Table 2). While the mechanism by which NGF may regulate morphine antinociception in this model remains elusive, we have implicated the potential relevance of CCK as an important inhibitory modulator of opioid antinociception in neuropathic pain states.

To validate further the hypothesis that endogenous spinal NGF is important in maintaining opioid-induced antinociception, we attempted to mimic the decreased effectiveness of opioid antinociception that occurs in neuropathic pain models by neutralizing endogenous levels of NGF with chronic intrathecal infusion of antibodies directed against purified 2.5S NGF. A previous study (116) demonstrated that changes in density and distribution of calcitonin gene-related peptide could be manipulated by changes in endogenous levels of NGF. Thus, chronic intrathecal anti-NGF treatment resulted in altered expression of CGRP in the dorsal horn of rat spinal cord (117). We recently discovered (unpublished data) that treating rats with intrathecal anti-NGF had no significant effect on morphine-induced antinociception compared with that seen in control rats infused with immunoglobulin G (IgG). However, whereas morphine-induced antinociception was potentiated by pretreatment with an intrathecal CCK-B receptor antagonist in anti-NGF-treated rats, there was no significant effect in IgG-treated controls (Table 3). These results draw a positive correlation between a decrease in NGF and CCK receptor antagonism restoring opioid antinociception. It is tempting to suggest that endogenous NGF appears to be important in maintaining the neuropeptide homeostasis,



**Figure 2)** Schematic representation of the changes that occur following peripheral nerve injury. These changes include an increase in the levels of cholecystokinin (CCK) and CCK-B receptors in dorsal root ganglia cells and spinal cord. Alterations in the spinal levels of nerve growth factor (NGF), whereby nerve injury disrupts retrograde transport to the cell bodies and the spinal cord, cause a decrease in the availability of spinal NGF. The increase in CCK and its receptor elicits an anti-opioid effect thereby resulting in an insensitivity to opioids used in the treatment of neuropathic pain. The augmentation of spinal CCK and CCK-B receptors may be the result of the attenuation of spinal NGF. Intrathecal infusion of NGF has been shown to attenuate spinal CCK and reverse opioid insensitivity in an animal model of neuropathic pain. DRG Dorsal root ganglion

including CCK, that is required for maintaining opioid effectiveness (Figure 2).

## CONCLUSIONS

Many authors have attempted to understand the mechanisms involved in the development and maintenance of chronic pain. The ultimate goal of these authors is to facilitate the development of new therapies for optimal pain treatment. Pain is generated by a number of processes that are qualitatively different, requiring multiple modalities for treatment. Accordingly, opioids are often very effective for the management of pain with an inflammatory origin, whereas opioids

have low efficacy or no effect on alleviating pain resulting from nerve injuries. The upregulation of CCK that ensues following nerve injury depresses the ability of endogenous opioids to modulate nociceptive transmission, resulting in the appearance of neuropathic pain syndromes and the reduction of the effectiveness of exogenously administered opioids.

NGF may play a role in the central plasticity and sensitization that occur following traumatic nerve injury. The low amount of central NGF, due to the attenuation of its transport to the CNS, forms part of the adaptive response that may inadvertently lead to the development of chronic pain. Changes in NGF may be related to the clinical phenomenon of opioid insensitivity that occurs in chronic pain of neuropathic origin. Nerve injury leads to complex changes in neuropeptide expression within primary afferents that correlate with changes in their phenotype; these events, particularly increases in CCK and its receptor, are thought to elicit neuropathic pain. It must be kept in mind that there are considerable species and strain differences in the reaction to neuronal damage (68). Moreover, variations in peptide levels are evident in the various animal models implemented for the induction of neuropathic pain-like characteristics within a species strain. Thus, clinically useful drugs for treating neuropathic pain may need to be tailored to specific patient populations where the pathology is well defined and symptoms have been properly charted.

Consequently, it has been proposed that NGF may, at the appropriate time, dose, site and mode of administration, provide prophylaxis and treatment in conditions that lead to chronic pain (117). Although intrathecal NGF does not alter neuropathic nociceptive behaviours, it does restore antiallodynic and antihyperalgesic effects of morphine. We propose that by normalizing the spinal levels of CCK, intrathecal administration of NGF may ameliorate the chronic pain associated with traumatic nerve injury by re-establishing the effectiveness of conventional opioid therapies for pain management.

## REFERENCES

- Merskey H, Bogduk N. Classification of Chronic Pain: Descriptions of Chronic Pain Syndromes and Definitions of Pain Terms. Seattle: IASP Press, 1994.
- Wiesenfeld-Hallin Z, Hao J-X, Xu X-J. Mechanisms of central pain. In: Jensen TJ, Turner JA, Wiesenfeld-Hallin Z, eds. Proceedings of the 8th World Congress on Pain, Progress in Pain Research and Management. Seattle: IASP Press, 1997:575-89.
- Coderre TJ, Katz J, Vaccarino AL, Melzack R. Contribution of central neuroplasticity to pathological pain: review of clinical and experimental evidence. *Pain* 1993;52:259-85.
- Bennett GJ, Xie YK. A peripheral mononeuropathy in rat that produces disorders of pain sensation like those seen in man. *Pain* 1988;33:87-107.
- Kim SH, Chung JM. An experimental model for peripheral neuropathy produced by segmental spinal nerve ligation in the rat. *Pain* 1992;50:355-63.
- Seltzer Z, Dubner R, Shir Y. A novel behavioral model of neuropathic pain disorders produced in rats by partial sciatic nerve injury. *Pain* 1990;43:205-18.
- Woolf CJ, Doubell TP. The pathophysiology of chronic pain – increased sensitivity to low threshold A beta-fibre inputs. *Curr Opin Neurobiol* 1994;4:525-34.
- Koerber HR, Mirmics K, Brown PB, Mendel LM. Central sprouting and functional plasticity of regenerated primary afferents. *J Neurosci* 1994;14:3655-71.
- Shortland P, Woolf CJ. Chronic peripheral nerve section results in a rearrangement of the central axonal arborizations of axotomized A beta primary afferent neurons in the rat spinal cord. *J Comp Neurol* 1993;330:65-82.
- Woolf CJ, Shortland P, Coggeshall RE. Peripheral nerve injury triggers central sprouting of myelinated afferents. *Nature* 1992;355:75-8.
- Woolf CJ, Shortland P, Reynolds M, Ridings J, Doubell T, Coggeshall RE. Reorganization of central terminals of myelinated primary afferents in the rat dorsal horn following peripheral axotomy. *J Comp Neurol* 1995;360:121-34.
- Nakamura S, Myers RR. Myelinated afferents sprout into lamina II of L3-5 dorsal horn following chronic constriction nerve injury in rats. *Brain Res* 1999;818:285-90.
- Raivich G, Hellweg R, Kreutzberg GW. NGF receptor-mediated reduction in axonal NGF uptake and retrograde transport following

- sciatic nerve injury and during regeneration. *Neuron* 1991;7:151-64.
14. MacFarlane BV, Wright A, O'Callaghan J, Benson HAE. Chronic neuropathic pain and its control by drugs. *Pharmacol Ther* 1997;75:1-19.
  15. Arnér S, Meyerson BA. Lack of analgesic effect of opioids on neuropathic and idiopathic forms of pain. *Pain* 1988;33:11-23.
  16. Max MB, Schafer SC, Culnane M, Dubner R, Gracely RH. Association of pain relief with drug side effects in postherpetic neuralgia: a single-dose study of clonidine, codeine, ibuprofen, and placebo. *Clin Pharmacol Ther* 1988;43:363-71.
  17. Kupers RC, Konings H, Adriaensen H, Gybels JM. Morphine differentially affects the sensory and affective pain ratings in neurogenic and idiopathic forms of pain. *Pain* 1991;47:5-12.
  18. Xu XJ, Wiesenfeld-Hallin Z. The threshold for the depressive effect of intrathecal morphine on the spinal nociceptive flexor reflex is increased during autotomy after sciatic nerve section in rats. *Pain* 1991;46:223-9.
  19. Mao J, Price DD, Mayer DJ. Experimental mononeuropathy reduces the antinociceptive effects of morphine: implications for common intracellular mechanisms involved in morphine tolerance and neuropathic pain. *Pain* 1995;61:353-64.
  20. Xu XJ, Puke MJ, Verge VM, Wiesenfeld-Hallin Z, Hughes J, Hökfelt T. Up-regulation of cholecystokinin in primary sensory neurons is associated with morphine insensitivity in experimental neuropathic pain in the rat. *Neurosci Lett* 1993;152:129-32.
  21. Xu XJ, Hao JX, Seiger A, Hughes J, Hökfelt T, Wiesenfeld-Hallin Z. Chronic pain-related behaviors in spinally injured rats: evidence for functional alterations of the endogenous cholecystokinin and opioid systems. *Pain* 1994;56:271-7.
  22. Ossipov MH, Lopez Y, Nichols ML, Bian D, Porreca F. Inhibition by spinal morphine of the tail-flick response is attenuated in rats with nerve ligation injury. *Neurosci Lett* 1995;199:83-6.
  23. Baber NS, Dourish CT, Hill DR. The role of CCK, caerulein, and CCK antagonists in nociception. *Pain* 1989;39:307-28.
  24. Stanfa L, Dickenson A, Xu XJ, Wiesenfeld-Hallin Z. Cholecystokinin and morphine analgesia: variations on a theme. *Trends Pharmacol Sci* 1994;15:65-6.
  25. Gardner JD, Jensen RT. Secretagogue receptors on pancreatic acinar cells. In: Johnson LR, ed. *Physiology of the Gastrointestinal Tract*, 2nd edn. New York: Raven Press, 1987:1109-27.
  26. Innis RB, Snyder SH. Distinct cholecystokinin receptors in brain and pancreas. *Proc Natl Acad Sci (USA)* 1980;77:6917-21.
  27. Wank SA. Cholecystokinin receptors. *Am J Physiol* 1995;269:G628-46.
  28. Seroogy KB, Mohapatra NK, Lund PK, Rethelyi M, McGehee DS, Perl ER. Species-specific expression of cholecystokinin messenger RNA in rodent dorsal root ganglia. *Brain Res Mol Brain Res* 1990;7:171-6.
  29. Verge VM, Wiesenfeld-Hallin Z, Hökfelt T. Cholecystokinin in mammalian primary sensory neurons and spinal cord: in situ hybridization studies in rat and monkey. *Eur J Neurosci* 1993;5:240-50.
  30. Dalsgaard CJ, Vincent SR, Hökfelt T, et al. Co-existence of cholecystokinin- and substance P-like peptides in neurons of the dorsal root ganglia of the rat. *Neurosci Lett* 1982;33:159-63.
  31. Williams RG, Dimaline R, Varro A, Isetta AM, Trizio D, Dockray GJ. Cholecystokinin octapeptide in rat central nervous system: immunocytochemical studies using a monoclonal antibody that does not react with CGRP. *Neurochem Int* 1987;11:433-42.
  32. Marley PD, Nagy JI, Emson PC, Rehfeld JF. Cholecystokinin in the rat spinal cord: distribution and lack of effect on neonatal capsaicin treatment and rhizotomy. *Brain Res* 1982;238:494-8.
  33. Ju G, Hökfelt T, Fischer JA, Frey P, Rehfeld JF, Dockray GJ. Does cholecystokinin-like immunoreactivity in rat primary sensory neurons represent calcitonin gene-related peptide? *Neurosci Lett* 1986;68:305-10.
  34. Fuji K, Senba E, Fugii S, et al. Distribution, ontogeny and projections of cholecystokinin-8, vasoactive intestinal peptide and gamma-aminobutyrate-containing neuron systems in the rat spinal cord: an immunohistochemical analysis. *Neuroscience* 1985;14:881-94.
  35. Cortés R, Arvidsson U, Schalling M, Ceccatelli S, Hökfelt T. In situ hybridization studies on mRNAs for cholecystokinin, calcitonin gene-related peptide and choline acetyltransferase in the lower brain stem, spinal cord and dorsal root ganglia of rat and guinea pig with special reference to motoneurons. *J Chem Neuroanat* 1990;3:467-85.
  36. Faris PL, Komisaruk BR, Watkins LR, Mayer DJ. Evidence for the neuropeptide cholecystokinin as an antagonist of opiate analgesia. *Science* 1983;219:310-2.
  37. Kellstein DE, Price DD, Mayer DJ. Cholecystokinin and its antagonist lorglumide respectively attenuate and facilitate morphine-induced inhibition of C-fiber evoked discharges of dorsal horn nociceptive neurons. *Brain Res* 1991;540:302-6.
  38. Magnuson DS, Sullivan AF, Simmonet G, Roques BP, Dickenson AH. Differential interactions of cholecystokinin and FLFQPQRF-NH<sub>2</sub> with mu and delta opioid antinociception in rat spinal cord. *Neuropeptides* 1990;16:213-8.
  39. Watkins LR, Kinscheck IB, Kaufman EF, Miller J, Frenk H, Mayer DJ. Cholecystokinin antagonists selectively potentiate analgesia induced by endogenous opiates. *Brain Res* 1985;327:181-90.
  40. Stanfa LC, Dickenson AH. Cholecystokinin as a factor in the enhanced potency of spinal morphine following carrageenin inflammation. *Br J Pharmacol* 1993;108:967-73.
  41. Noble F, Biommaert A, Fournie-Zaluski MC, Roques BP. A selective CCK-B receptor antagonist potentiates mu-, but not delta-opioid receptor-mediated antinociception in the formalin test. *Eur J Pharmacol* 1995;273:145-51.
  42. Dourish CT, O'Neill MF, Coughlan J, Kitchener SJ, Hawley D, Iversen SD. The selective CCK-B receptor antagonist L-365,260 enhances morphine analgesia and prevents morphine tolerance in the rat. *Eur J Pharmacol* 1990;176:35-44.
  43. Wiesenfeld-Hallin Z, Xu XJ, Hughes J, Horwell DC, Hökfelt T. PD134308, a selective antagonist of cholecystokinin type-B receptor, enhances the analgesic effect of morphine and synergistically interacts with intrathecal galanin to depress spinal nociceptive reflexes. *Proc Natl Acad Sci (USA)* 1990;87:7105-9.
  44. Zhou Y, Sun YH, Zhang ZW, Han JS. Increased release of immunoreactive cholecystokinin octapeptide by morphine and potentiation of mu-opioid analgesia by CCKB receptor antagonist L-365,260 in rat spinal cord. *Eur J Pharmacol* 1993;234:147-54.
  45. Vanderah TW, Bernstein RN, Yamamura HI, Hruby VJ, Porreca F. Enhancement of morphine antinociception by a CCKB antagonist in mice is mediated via opioid delta receptors. *J Pharmacol Exp Ther* 1996;278:212-9.
  46. Vanderah TW, Lai J, Yamamura HI, Porreca F. Antisense oligodeoxynucleotide to the CCKB receptor produces naltrindole- and [Leu5]enkephalin antiserum-sensitive enhancement of morphine antinociception. *Neuroreport* 1994;5:2601-5.
  47. McCleane GJ. The cholecystokinin antagonist proglumide enhances the analgesic efficacy of morphine in human with chronic benign pain. *Anesth Analg* 1998;87:1117-20.
  48. Becker C, Hamon M, Cesselin F, Benoliel JJ. Delta(2)-opioid receptor mediation of morphine-induced CCK release in the frontal cortex of the freely moving rat. *Synapse* 1999;34:47-54.
  49. Benoliel JJ, Mauborgne A, Bourgoin S, Legrand JC, Hamon M, Cesselin F. Opioid control of the in vitro release of cholecystokinin-like material from the rat substantia nigra. *J Neurochem* 1992;58:916-22.
  50. Benoliel JJ, Mauborgne A, Bourgoin S, Hamon M, Cesselin F. Opioid control of CCK release from spinal cord of morphine-tolerant and non-tolerant rats. 1st European Opioid Conference, Surrey, Sept 9-12, 1997. (Abst T17)
  51. Wiesenfeld-Hallin Z, Xu XJ. The role of cholecystokinin in nociception, neuropathic pain and opiate tolerance. *Regul Pept* 1996;65:23-8.
  52. Beinfeld MC, Meyer DK, Eskay RL, Jensen RT, Brownstein MJ. The distribution of cholecystokinin immunoreactivity in the central nervous



- system of the rat as determined by radioimmunoassay. *Brain Res* 1981;212:51-7.
53. Stengaard-Pedersen K, Larsson LI. Localization and opiate receptor binding of enkephalin, CCK and ACTH/beta-endorphin in the rat central nervous system. *Peptides* 1981;2 (Suppl 1):3-19.
  54. Gall C, Lauterborn J, Burks D, Serogy K. Co-localization of enkephalin and cholecystokinin in discrete areas of rat brain. *Brain Res* 1987;403:403-8.
  55. Pohl M, Benoliel JJ, Bourgoin S, et al. Regional distribution of calcitonin gene-related peptide-, substance P-, cholecystokinin-, Met<sup>5</sup>-enkephalin-, and dynorphin A (1-8)-like material in the spinal cord and dorsal root ganglia of adult rats: effects of dorsal rhizotomy and neonatal capsaicin. *J Neurochem* 1990;55:1122-30.
  56. Wang XJ, Han JS. Modification of cholecystokinin octapeptide of the binding of mu-, delta- and kappa-opioid receptors. *J Neurochem* 1990;55:1379-82.
  57. Wang XJ, Ren MF, Han JS. Mobilization of calcium from intracellular stores is one of the mechanisms underlying the antioioid effect of cholecystokinin octapeptide. *Peptides* 1992;13:947-51.
  58. Ghilardi JR, Allen CJ, Vigna SR, McVey DC, Mantyh PW. Trigeminal and dorsal root ganglion neurons express CCK receptor binding sites in the rat, rabbit, and monkey: possible site of opiate-CCK analgesic interactions. *J Neurosci* 1992;12:4854-66.
  59. Liu NJ, Xu T, Xu C, et al. Cholecystokinin octapeptide reverses mu-opioid-receptor-mediated inhibition of calcium current in rat dorsal root ganglion neurons. *J Pharmacol Exp Ther* 1995;275:1293-9.
  60. Mestek A, Hurlley JH, Bye LS, et al. The human mu opioid receptor: modulation of functional desensitization by calcium/calmodulin-dependent protein kinase and protein kinase C. *J Neurosci* 1995;15:2396-406.
  61. Pei G, Kieffer BL, Lefkowitz RJ, Freedman NJ. Agonist-dependent phosphorylation of the mouse delta-opioid receptor: involvement of G protein-coupled receptor kinases but not protein kinase C. *Mol Pharmacol* 1995;48:173-7.
  62. Schafer M, Zhou L, Stein C. Cholecystokinin inhibits peripheral opioid analgesia in inflamed tissue. *Neuroscience* 1998;82:603-11.
  63. Nichols ML, Bian D, Ossipov MH, Lai J, Porecca F. Regulation of morphine antiallodynic efficacy by cholecystokinin in a model of neuropathic pain in rats. *J Pharmacol Exp Ther* 1995;275:1339-45.
  64. Yamamoto T, Sakashita Y. Differential effects of intrathecally administered morphine and its interaction with cholecystokinin-B antagonist on thermal hyperalgesia following two models of experimental mononeuropathy in the rat. *Anesthesiology* 1999;90:1382-91.
  65. Idanpaan-Heikkila JJ, Guilbaud G, Kayser V. Prevention of tolerance to the antinociceptive effects of systemic morphine by a selective cholecystokinin-B receptor antagonist in a rat model of peripheral neuropathy. *J Pharmacol Exp Ther* 1997;282:1366-72.
  66. Xu XJ, Hökfelt T, Hughes J, Wiesenfeld-Hallin Z. The CCK-B antagonist C1988 enhances the reflex-depressive effect of morphine in axotomized rats. *Neuroreport* 1994;5:718-20.
  67. Hökfelt T, Zhang X, Wiesenfeld-Hallin Z. Messenger plasticity in primary sensory neurons following axotomy and its functional implications. *Trends Neurosci* 1994;17:22-30.
  68. Zhang X, Dagerlind A, Elde RP, et al. Marked increase in cholecystokinin B receptor messenger RNA levels in rat dorsal root ganglia after peripheral axotomy. *Neuroscience* 1993;57:227-33.
  69. Ossipov MH, Maln TP, Lai J, Porreca F. Opioid pharmacology of acute and chronic pain. In: Dickenson A, Besson JM, eds. *The Pharmacology of Pain. Handbook of Experimental Pharmacology*, vol 130. Berlin: Springer-Verlag, 1997:305-34.
  70. Yaksh TL, Furui T. Release of cholecystokinin-like immunoreactivity from rat cerebral cortex and spinal cord: modulatory effects of excitatory and inhibitory amino acids. In: Hughes J, Dockray G, Woodruff G, eds. *The Neuropeptide Cholecystokinin (CCK)*. Chichester: Ellis Horwood, 1989:81-90.
  71. Bandopadhyay R, de Belleruche J. Regulation of CCK release in cerebral cortex by *N*-methyl-D-aspartate receptors: sensitivity to APV, MK-801, kynurenate, magnesium and zinc ions. *Neuropeptides* 1991;18:159-63.
  72. Paulice P, Gemignani A, Raiteri M. Evidence for functional native NMDA receptors activated by glycine or D-serine alone in the absence of glutamatergic coagonist. *Eur J Neurosci* 1998;10:2934-44.
  73. Xu X-J, Wiesenfeld-Hallin Z. Novel modulators in nociception. In: Dickenson A, Besson JM, eds. *The Pharmacology of Pain. Handbook of Experimental Pharmacology*, vol 130. Berlin: Springer-Verlag, 1997:201-34.
  74. Ritter AM, Lewin GR, Kremer NE, Mendell LM. Requirement for nerve growth factor in the development of myelinated nociceptors in vivo. *Nature* 1991;350:500-2.
  75. Crowley C, Spencer SD, Nishimura MC, et al. Mice lacking nerve growth factor display perinatal loss of sensory and sympathetic neurons yet develop basal forebrain cholinergic neurons. *Cell* 1994;76:1001-11.
  76. Rice FL, Albers KM, Davis BM, et al. Differential dependency of unmyelinated and A delta epidermal and upper dermal innervation on neurotrophins, trk receptors, and p75LNGFR. *Dev Biol* 1998;198:57-81.
  77. Albers KM, Wright DE, Davis BM. Overexpression of nerve growth factor in epidermis of transgenic mice causes hypertrophy of the peripheral nervous system. *J Neurosci* 1994;14:1422-32.
  78. Davis BM, Fundin BT, Albers KM, Goodness TP, Cronk KM, Rice FL. Overexpression of nerve growth factor in skin causes preferential increases among innervation to specific sensory targets. *J Comp Neurol* 1997;387:489-506.
  79. Davis BM, Lewin GR, Mendell LM, Jones ME, Albers KM. Altered expression of nerve growth factor in the skin of transgenic mice leads to changes in response to mechanical stimuli. *Neuroscience* 1993;56:789-92.
  80. Woolf CJ, Ma QP, Allchorne A, Poole S. Peripheral cell types contributing to the hyperalgesic action of nerve growth factor in inflammation. *J Neurosci* 1996;16:2716-23.
  81. Mendell LM, Albers KM, Davis BM. Neurotrophins, nociceptors and pain. *Microscopy Res Tech* 1999;45:252-61.
  82. Aloe L, Tuveri MA, Levi-Montalcini R. Studies on carrageenan-induced arthritis in adult rats: presence of nerve growth factor and role of sympathetic innervation. *Rheumatol Int* 1992;12:213-6.
  83. Lewin GR, Rueff A, Mendell LM. Peripheral and central mechanisms of NGF-induced hyperalgesia. *Eur J Neurosci* 1994;6:1903-12.
  84. Woolf CJ, Safieh-Garabedian B, Ma QP, Crilly P, Winter J. Nerve growth factor contributes to the generation of inflammatory sensory hypersensitivity. *Neuroscience* 1994;62:327-31.
  85. McMahon SB, Bennett DL, Priestley JV, Shelton DL. The biological effects of endogenous nerve growth factor on adult sensory neurons revealed by a trkA-IgG fusion molecule. *Nat Med* 1995;1:774-80.
  86. Bennett G, al-Rashed S, Hoult JR, Brain SD. Nerve growth factor induced hyperalgesia in the rat hind paw is dependent on circulating neutrophils. *Pain* 1998;77:315-22.
  87. Ma QP, Woolf CJ. The progressive tactile hyperalgesia induced by peripheral inflammation is nerve growth factor dependent. *Neuroreport* 1997;8:807-10.
  88. Otten U, Goedert M, Mayer N, Lembeck F. Requirement of nerve growth factor for the development of substance P-containing sensory neurones. *Nature* 1980;287:158-9.
  89. Lindsay RM, Harmor AJ. Nerve growth factor regulates expression of neuropeptide genes in adult sensory neurons. *Nature* 1989;337:362-4.
  90. Donnerer J, Schuligoi R, Stein C. Increased content and transport of substance P and calcitonin gene-related peptide in sensory nerves innervating inflamed tissue: evidence for a regulatory function of nerve growth factor in vivo. *Neuroscience* 1992;49:693-8.
  91. Leslie TA, Emson PC, Dowd PM, Woolf CJ. Nerve growth factor contributes to the up-regulation of growth-associated protein 43 and preprotachykinin A messenger RNAs in primary sensory neurons following peripheral inflammation. *Neuroscience* 1995;67:753-61.
  92. Richardson PM, Riopelle RJ. Uptake of nerve growth factor along

- peripheral and spinal axons of primary sensory neurons. *J Neurosci* 1984;4:1683-9.
93. Bennett DL, French J, Priestley JV, McMahon SB. NGF but not NT-3 or BDNF prevents the A fiber sprouting into lamina II of the spinal cord that occurs following axotomy. *Mol Cell Neurosci* 1996;8:211-20.
  94. Verge VM, Merlio JP, Grondin J, et al. Colocalization of NGF binding sites, trk mRNA, and low-affinity NGF receptor mRNA in primary sensory neurons: responses to injury and infusion of NGF. *J Neurosci* 1992;12:4011-22.
  95. Csillik B, Schwab ME, Thoenen H. Transganglionic regulation of central terminals of dorsal root ganglion cells by nerve growth factor (NGF). *Brain Res* 1985;331:11-5.
  96. Schwartz JP, Pearson J, Johnson EM. Effect of exposure to anti-NGF on sensory neurons of adult rats and guinea pigs. *Brain Res* 1982;244:378-81.
  97. Fitzgerald M, Wall PD, Goedert M, Emson PC. Nerve growth factor counteracts the neurophysiological and neurochemical effects of chronic sciatic nerve section. *Brain Res* 1985;332:131-41.
  98. Otto D, Unsicker K, Grothe C. Pharmacological effects of nerve growth factor and fibroblast growth factor applied to the transected sciatic nerve on neuron death in adult rat dorsal root ganglia. *Neurosci Lett* 1987;83:156-60.
  99. Rich KM, Luszczynski JR, Osborne PA, Johnson EM Jr. Nerve growth factor protects adult sensory neurons from cell death and atrophy caused by nerve injury. *J Neurocytol* 1987;16:261-8.
  100. Gold BG, Mobley WC, Matheson SF. Regulation of axonal caliber, neurofilament content, and nuclear localization in mature sensory neurons by nerve growth factor. *J Neurosci* 1991;11:943-55.
  101. Verge VM, Riopelle RJ, Richardson PM. Nerve growth factor receptors on normal and injured sensory neurons. *J Neurosci* 1989;9:914-22.
  102. Isaacson LG, Ondris D, Crutcher KA. Plasticity of mature sensory cerebrovascular axons following intracranial infusion of nerve growth factor. *J Comp Neurol* 1995;361:451-60.
  103. Ma W, Ribeiro-da-Silva A, Noel G, Julien JP, Cuervo AC. Ectopic substance P and calcitonin gene-related peptide immunoreactive fibres in the spinal cord of transgenic mice over-expressing nerve growth factor. *Eur J Neurosci* 1995;7:2021-35.
  104. Mulderrry PK. Neuropeptide expression by newborn and adult rat sensory neurons in culture: effects of nerve growth factor and other neurotrophic factors. *Neuroscience* 1994;59:673-88.
  105. Verge VM, Richardson PM, Wiesenfeld-Hallin Z, Hökfelt T. Differential influence of nerve growth factor on neuropeptide expression in vivo: a novel role in peptide suppression in adult sensory neurons. *J Neurosci* 1995;15:2081-96.
  106. Dray A, Urban L. New pharmacological strategies for pain relief. *Annu Rev Pharmacol Toxicol* 1996;36:253-80.
  107. Herzberg U, Eliav E, Dorsey JM, Gracely RH, Kopin IJ. NGF involvement in pain induced by chronic constriction injury of the rat sciatic nerve. *Neuroreport* 1997;8:1613-8.
  108. Rogers BC. Development of recombinant human nerve growth factor (rhNGF) as a treatment for peripheral neuropathic disease. *Neurotoxicology* 1996;17:865-70.
  109. Ren K, Thomas DA, Dubner R. Nerve growth factor alleviates a painful peripheral neuropathy in rats. *Brain Res* 1995;699:286-92.
  110. Ro LS, Chen ST, Tang LM, Jacobs JM. Effect of NGF and anti-NGF on neuropathic pain in rats following chronic constriction injury of the sciatic nerve. *Pain* 1999;79:265-74.
  111. Garrett NE, Kidd BL, Cruwys SC, Tomlinson DR. Effect of streptozotocin-diabetes on knee joint inflammation-induced changes in substance P and nerve growth factor in the rat. *Brain Res Mol Brain Res* 1996;42:272-8.
  112. Delcroix JD, Tomlinson DR, Fernyhough P. Diabetes and axotomy-induced deficits in retrograde axonal transport of nerve growth factor correlate with decreased levels of p75<sup>NTR</sup> protein in lumbar dorsal root ganglia. *Brain Res Mol Brain Res* 1997;51:82-90.
  113. McMahon SB, Bennett DLH. Growth factors and pain. In: Dickenson S, Besson J-M, eds. *The Pharmacology of Pain. Handbook of Experimental Pharmacology*, vol 130. Berlin: Springer-Verlag, 1997:135-66.
  114. Zhou XF, Rush RA, McLachlan EM. Differential expression of the p75 nerve growth factor receptor in glia and neurons of the rat dorsal root ganglia after peripheral nerve transection. *J Neurosci* 1996;16:2901-11.
  115. Lapchak PA, Araujo DM. NGF suppression of weight gain in adult female rats correlates with decreased hypothalamic cholecystokinin levels. *Brain Res* 1994;655:12-6.
  116. Christensen MD, Hulsebosch CE. Spinal cord injury and anti-NGF treatment results in changes in CGRP density and distribution in the dorsal horn in the rat. *Exp Neurol* 1997;147:463-75.
  117. Anand P. Nerve growth factor regulates nociception in human health and disease. *Br J Anaesth* 1995;75:201-8.
-



**Hindawi**  
Submit your manuscripts at  
<http://www.hindawi.com>

