

UCLA

UCLA Previously Published Works

Title

Spatial and Temporal Cortical Variability Track With Age and Affective Experience During Emotion Regulation in Youth

Permalink

<https://escholarship.org/uc/item/20q4p92c>

Journal

Developmental Psychology, 55(9)

ISSN

0012-1649

Authors

Moreira, João F Guassi
McLaughlin, Katie A
Silvers, Jennifer A

Publication Date

2019-09-01

DOI

10.1037/dev0000687

Peer reviewed

Running head: NEURAL VARIABILITY AND EMOTION REGULATION

Spatial and Temporal Cortical Variability Track with Age and Affective Experience During
Emotion Regulation in Youth

João F. Guassi Moreira¹, Katie A. McLaughlin², & Jennifer A. Silvers^{1*}

¹Department of Psychology, University of California, Los Angeles

²Department of Psychology, University of Washington, Seattle

Manuscript Submission for *Developmental Psychology*
2019 Special Issue on Development of Human Emotion

*Corresponding Author
Jennifer A. Silvers
Department of Psychology
1285 Franz Hall
Box 951563
Los Angeles, CA 90095-1563
310-794-4953
silvers@ucla.edu

NEURAL VARIABILITY AND EMOTION REGULATION

Abstract

Variability is a fundamental feature of human brain activity that is particularly pronounced during development. However, developmental neuroimaging research has only recently begun to move beyond characterizing brain function across development exclusively in terms of magnitude of neural activation to incorporating estimates of variability. No prior neuroimaging study has done so in the domain of emotion regulation. We investigated how age and affective experiences influence spatial and temporal variability in neural activity during emotion regulation. In the current study, 70 typically developing youth aged 8-17 years completed a cognitive reappraisal task of emotion regulation while undergoing functional magnetic resonance imaging. Estimates of spatial and temporal variability during emotion regulation were calculated across a network of brain regions, defined *a priori*, and were then related to age and affective experiences. Results showed that increasing age was associated with reduced spatial and temporal variability in a set of frontoparietal regions (e.g., dorsomedial prefrontal cortex, superior parietal lobule) known to be involved in effortful emotion regulation. In addition, youth who reported less negative affect during emotion regulation had less spatial variability in the ventrolateral prefrontal cortex. We interpret age-related reductions in spatial and temporal variability as evidence of neural specialization. These results imply that the development of emotion regulation is undergirded by a process of neural specialization and open up a host of possibilities for incorporating neural variability into the study of emotion regulation development.

Keywords: Emotion; Emotion Regulation; Development; Variability; Neuroimaging

NEURAL VARIABILITY AND EMOTION REGULATION

Spatial and Temporal Cortical Variability Track with Age and Affective Experience During Emotion Regulation in Youth

Advances in *in vivo* functional neuroimaging have granted psychologists profound insights into how the human brain develops and functions. While most research in developmental cognitive neuroscience has focused on comparing the *magnitude* of neural responses at different ages, accumulating evidence suggests that it is equally important to assess neural *variability* – that is, how the neural signal varies across time and space both within and between individuals (e.g., Durston et al., 2006; Heller & Casey, 2016; Nomi, Bolt, Ezie, Uddin, & Heller, 2017). This is because age-related differences in neural variability likely reflect important developmental processes, including the degree of specialization and experience-based plasticity (e.g., pruning) in neural circuits across age (Casey, 2015; Durston et al., 2006). Emotion regulation presents itself as a particularly important skill to be assessed in a developmental neural variability framework because it exhibits protracted maturation and is critical for wellbeing (Cole & Deater-Deckard, 2009; Gross, 2015; McLaughlin, Garrard, & Somerville, 2015). However, virtually all developmental neuroimaging studies of emotion regulation to date have concentrated on age-related differences in the magnitude of brain activity across individuals and have ignored how variability *within* individuals during emotion regulation differs across development. The present study sought to address this knowledge gap by investigating how within-subject variability in neural activity during emotion regulation—both spatial and temporal—is associated with age and affective experience in youth.

Prior Investigations of Emotion Regulation Neurodevelopment Omit Variability.

Inspired by behavioral work demonstrating that emotion regulation abilities improve significantly during childhood and adolescence (Crone & van der Molen, 2004; Kim & Richardson, 2010; Silvers et al., 2012; Somerville, Jones, & Casey, 2010; Thompson &

NEURAL VARIABILITY AND EMOTION REGULATION

Goodman, 2010), a growing number of neuroimaging studies have begun to explore the neural bases of emotion regulation in youth. One popular means for doing so has been to employ functional magnetic resonance imaging (fMRI) in conjunction with cognitive reappraisal—a widely studied and adaptive emotion regulation strategy that involves thinking about a stimulus differently in order to modulate its emotional import (Denny & Ochsner, 2014; Giuliani & Pfeifer, 2015; Gross, 2015; Ochsner, Silvers, & Buhle, 2012). In healthy adults, reappraisal recruits frontoparietal regions commonly implicated in cognitive control, including the ventrolateral, dorsolateral and dorsomedial prefrontal cortex (vlPFC, dlPFC, and dmPFC, respectively) as well as superior parietal cortex (SPL; Buhle et al., 2014; Ochsner et al., 2004, 2012; Ochsner & Gross, 2005). Across development, increasing age is associated with increased recruitment of these frontoparietal regions along with improvements in reappraisal ability, suggesting that emotion regulation success improves as cognitive control abilities become more fine-tuned (McRae et al., 2012; Silvers et al., 2016; Silvers, Shu, Hubbard, Weber, & Ochsner, 2015). While informative, these initial developmental neuroimaging studies of reappraisal have relied heavily on univariate analyses that characterize age-related changes in terms of peak or mean BOLD signal. As such, this existing work has helped advance the field, but has also overlooked the role that neural variability plays in emotion regulation.

Neural variability stands to inform novel hypotheses in developmental affective science in several ways. Studying within-person variability can identify another dimension by which brain regions and networks differ or co-vary. Doing so may help to uncover hidden network states across brain regions implicated in emotion regulation (Seghier & Price, 2018). This could be relevant for further taxonomizing the individual neurocognitive processes that constitute emotion regulation strategies such as cognitive reappraisal. Relatedly, variability may serve as

NEURAL VARIABILITY AND EMOTION REGULATION

another metric for assessing maturation of emotion regulation and related affective phenomena.

The dynamic systems view of development posits that variability within individuals is a hallmark of developmental processes (Smith & Thelen, 2003)—quantifying variability might allow for more precise mapping of individual growth curves for brain development as it relates to emotion regulation skills across age.

Variability is a Key Feature of Neural Function and a Catalyst for Development.

Variability is a fundamental feature of brain activity that is distinct from “randomness” or “noise” (Pinneo, 1966). Neural activity is organized according to structured spatial and temporal profiles (Christophel, Iamshchinina, Yan, Allefeld, & Haynes, 2018; Huth, Heer, Griffiths, Theunissen, & Gallant, 2016; Huth, Nishimoto, Vu, & Gallant, 2012; Luciana, Wahlstrom, Porter, & Collins, 2012). While not always thought of in terms of variability, the fact that spatial and temporal activation patterns within-individuals vary between different psychological processes suggests that variability is a defining feature of brain function (Etzel, Zacks, & Braver, 2013; Patel, Kaplan, & Snyder, 2014). Put differently, neuronal computations vary for different psychological processes, leading neuroimaging data associated with different processes to vary as well (Kriegeskorte, Cusack, & Bandettini, 2010). The mere fact that seeing a face evokes a different pattern of activity than seeing a house, for example, illustrates that configurations of brain activity are spatially variable (Haxby et al., 2001). Examining variability is therefore one meaningful way to characterize the neural substrates of psychological processes.

Though within-individual variability pertaining to the neurodevelopment of emotion regulation has not been examined, prior research in psychology and developmental neuroscience can help scaffold the present investigation. Psychologically, children try a variety of strategies during development (i.e., exhibit high variability) before becoming expert in a narrower set of

NEURAL VARIABILITY AND EMOTION REGULATION

strategies (Roalf et al., 2014; Siegler, 1994, 2007). Brain development follows a similar pattern. Functionally, brain responses may exhibit “focalization” across age – a shift from a more variable spatial signature of activity to one that is more concentrated, and potentially, specialized (Dehaene-Lambertz, Monzalvo, & Dehaene, 2018; Durston et al., 2006; Richardson, Lisandrelli, Riobueno-Naylor, & Saxe, 2018). Importantly, however, evidence for the shift from diffuse to focused activity has come from studies examining average levels of brain activity across individuals of different ages rather than examining within-subject variability. These patterns of functional maturation may be driven, in part, by pruning of initially over-produced synaptic connections to produce increasingly efficient and specialized neural networks that retain only their most essential connections (DuPre & Spreng, 2017; Durston et al., 2006; Foulkes & Blakemore, 2018; Sowell, Thompson, Tessner, & Toga, 2001; Vij, Nomi, Dajani, & Uddin, 2018). These findings suggest that neurodevelopment is characterized by a transition from diffuse and spatially variable patterns of activity towards optimized and constrained functional networks. Investigating neural variability promises to enhance understanding of neurodevelopment and generate novel hypotheses for future research (Poldrack, 2015).

Incorporating Variability into the Study of Emotion Regulation Neurodevelopment.

We used the work summarized above to guide our query into how neural variability supports the neurodevelopment of emotion regulation. We focused specifically on spatial and temporal variability.

Spatial Variability. Spatial variability, or differences in how activation is distributed across a brain region, is an important organizational feature of neural activity. Psychological processes, including emotional experiences, are encoded in richly detailed spatial topographies that blanket the cortical landscape (Chang, Gianaros, Manuck, & Krishnan, 2015; Huth et al.,

NEURAL VARIABILITY AND EMOTION REGULATION

2016, 2012; Kriegeskorte et al., 2008). Prior work in developmental neuroscience suggests that such topographies are diffuse earlier in life and become focalized with age and experience (Dehaene-Lambertz et al., 2018; Durston et al., 2006; Park et al., 2004; Polk et al., 2002). Examining spatial variability during a specific psychological process may be one way to infer how specialized a given brain region is for that process for a given individual. For example, if an individual shows a low degree of spatial variability within ventrolateral prefrontal cortex (vlPFC) during emotion regulation, this would suggest that activity is concentrated to a specific subset of the neurons in that region. In contrast, a high degree of spatial variability in another individual would suggest that the computations occurring in vlPFC that support emotion regulation are being carried out across a larger population of neurons for that person. Examining spatial variability promises to uncover insights about the functional architecture of neurodevelopment that has relevance not only for emotion regulation but for many other psychological processes. In the current study, we implemented a novel method of estimating spatial variability by repurposing an analytic technique from economics—Gini coefficients—to serve as a metric of spatial variability in a given brain region. As will be described later, Gini coefficients are an especially useful tool because they can yield a numerical index of spatial variability, helping quantify complex theoretical concepts such as focalization.

This approach can be particularly informative for affective neuroscience by way of its ability to highlight whether certain brain regions co-vary or differ in their topographic organization by telling us how patterns of activity are arranged across the surface of cortex (i.e., concentrated or diffusely). For instance, vlPFC, dlPFC, and dmPFC may all show similar magnitudes of activity during reappraisal, but vlPFC and dlPFC may have more similar topographies (i.e., both diffuse or both focal), indicating that a different psychological sub-

NEURAL VARIABILITY AND EMOTION REGULATION

process is being implemented by those brain regions compared to the computations in dmPFC (e.g., working memory vs mentalizing). This would help test dominant theories that posit multiple executive functions support complex, cognitively demanding emotion regulation strategies (Ochsner, Silvers, & Buhle, 2012).

Temporal Variability. While spatial variability can describe how brain function is organized topographically, temporal variability can lend insight into how brain activity changes over time to meet the dynamic demands of one's environment. This is especially relevant for psychological phenomena that change over time, such as emotion regulation (Aldao, Sheppes, & Gross, 2015; Heller & Casey, 2016). Studying temporal variability in brain activation across multiple instances of emotion regulation can lend insight into how neural computations adjust according to varying regulatory demands. Situations that require emotion regulation vary markedly – emotion regulation may be required to maintain calm in a traffic jam or to respond to the loss of a loved one. Learning to consistently mount an effective regulatory response to variable emotional events is a significant developmental hurdle. Given that youth tend to experience more intense and labile emotions, and yet have fewer cognitive resources to draw from, we might expect them to display less consistent (i.e., more variable) neural recruitment during self-regulation (Larson, Csikszentmihalyi, & Graef, 1980; Mischel & Mischel, 1983; Silvers et al., 2012; Somerville et al., 2010). As individuals mature and experience with emotion regulation grows, however, neural computations underlying emotion regulation are likely to become more consistent (i.e., less variable; Koolschijn, Schel, de Rooij, Rombouts, & Crone, 2011; Ordaz, Foran, Velanova, & Luna, 2013).

Current Study.

NEURAL VARIABILITY AND EMOTION REGULATION

In the current study, we used functional magnetic resonance imaging (fMRI) to examine spatial and temporal variability of frontoparietal brain responses during emotion regulation in a sample of typically developing youth. We further sought to characterize how spatial and temporal variability related to age and affective experiences during emotion regulation in this sample. By doing so, we were able to observe how two forms of within-individual neural variability related to age and experiences of affect during emotion regulation (Foulkes & Blakemore, 2018). Given the paucity of research on this matter, this research was exploratory and guided by open questions regarding neural variability and emotion regulation rather than formal hypotheses.

Methods

Participants. Participants included 70 youth (34 female) ranging in age from 8.08 to 17.00 years ($M_{age} = 12.70$). These participants were drawn from a larger sample taking part in a longitudinal study aimed at understanding the effects of childhood maltreatment on affective neurodevelopment. Only youth from the non-maltreated community control group were selected for the current analyses. Exclusion criteria for this group included exposure to childhood maltreatment or violence, presence of a developmental disorder (e.g., autism), psychotropic medication use, and $IQ < 75$. To qualify for inclusion in the current analysis, participants had to have: (i) accompanying behavioral data from the in-scan emotion regulation task (described below); (ii) low levels of motion during the scan (see below); and (iii) anatomical images that were free of abnormalities or artifacts. Participants and their families were recruited from a large, metropolitan area in the Pacific Northwest. Parents provided written consent and children provided written assent in accordance with the University of Washington Institutional Review Board. Participants were compensated \$75 for completing the brain scan.

NEURAL VARIABILITY AND EMOTION REGULATION

fMRI Paradigm. During the fMRI scan, participants completed a computerized version of a cognitive reappraisal task adapted from prior developmental studies of emotion regulation (McLaughlin, Peverill, Gold, Alves, & Sheridan, 2015; Silvers et al., 2016). During the task, participants viewed a series of developmentally appropriate aversive and neutral images modeled after the International Affective Picture System (IAPS) stimulus set (Lang, Bradley, & Cuthbert, 2008). Great care was taken to create and validate a stimulus set that would be appropriate to use in children and adolescents. Aversive images all depicted conflict between individuals or personal struggles (e.g., an individual sitting alone in sadness). Moreover, such images were especially relevant for youth because they portrayed youth in aversive scenarios common to their lives (e.g., depiction of bullying/fighting behaviors, adults fighting in front of children). Images were purchased from a stock photography website (<https://www.shutterstock.com>), including 225 negative and 150 neutral images. In a pilot study, 120 children aged 6-16 years (50% female) provided ratings of valence, arousal, and dominance for a randomized selection of 80 images using the standardized assessment mannequins used to norm the IAPS stimuli. The stimuli and normative ratings are available on the lab website of the principal investigator of the original study (www.stressdevelopmentlab.org). Images were selected from the larger stimulus set based on the valence ratings from the pilot study; the images with the most negative valence ratings were selected for the negative condition, and those closest to the mid-point of the valence scale were selected for the neutral condition. This custom image validation procedure is a notable strength, as most developmental imaging investigations of emotion regulation typically rely on stimulus sets that have been developed and validated for use in adults.

During the task, participants were instructed to either passively observe or reappraise the images via psychological distancing. Afterwards, participants provided their ratings of negative

NEURAL VARIABILITY AND EMOTION REGULATION

affect along a four-point Likert scale (see Figure 1). Participants reappraised negative stimuli on 20 trials, passively observed negative stimuli on 20 trials, and passively observed neutral stimuli on 20 trials. Prior to each image, an instructional cue was displayed that informed participants whether they were to passively observe ('Look') or regulate ('Far') the subsequent image. For 'Look' trials, participants were told to passively observe the stimulus and react to it as they normally would. During 'Far' trials, participants were trained to reappraise the image in a way that made it psychologically distant, such as pretending they were physically standing far from the image in each scene or that they were behind a movie camera, recording the events shown in the picture. Since our interests lay in characterizing spatial and temporal variability during active emotion regulation, we focused on 'Far' trials for the current report.

Prior to scanning, participants completed a training session where they received information about the meaning of the instructional cues and how they were to think about stimuli presented after each type of cue. Experimenters then shared several examples of each condition to participants before asking them to complete a series of 5 practice trials for each condition. During the in-scan task instructional cues were presented for 2 seconds and stimuli were jittered such that they were displayed for 6s – 10s. The Likert scale was presented thereafter for 4 seconds followed by a 1.5s – 6.5s ITI. Participants completed four runs, each lasting approximately 220s (runs ranged between 110 and 115 volumes in length). The stimuli used for 'Look' and 'Far' trials did not differ in their normative ratings of valence and arousal. The task was programmed using E-Prime (Psychology Software Tools, Inc., <http://www.pstnet.com>).

fMRI Data Acquisition and Preprocessing.

fMRI Data Acquisition. Prior to image acquisition, participants younger than 12 years old or who exhibited any signs of nervousness about scanning were taken to a mock MRI scanner to

NEURAL VARIABILITY AND EMOTION REGULATION

become familiarized with the scanning environment and trained on how to minimize head motion. These participants watched a film on a back-projector with a head-mounted motion tracker. The film paused if a head movement exceeding 2mm occurred, helping participants quickly learn to keep still while in the mock scanner bore. In addition to this measure, participants were packed into the head coil with an inflated, head-stabilizing pillow to restrict movement.

Image acquisition was conducted on a 3T Phillips Achieva scanner at the University of Washington (Seattle) Integrated Brain Imaging Center. A 32-channel head coil was implemented, along with the use of a parallel image acquisition system. T1-weighted, magnetization-prepared rapid acquisition gradient echo (MPRAGE) volumes were acquired (TR = 2530ms, TE = 1640-7040 μ s, 7° flip angle, 256 mm² FOV, 176 slices, 1 mm³ isotropic voxel size). Blood oxygenation level dependent (BOLD) signal during functional runs was acquired using a gradient-echo T2*-weighted echoplanar imaging (EPI) sequence. Thirty-two 3-mm thick slices were acquired parallel to the AC-PC line (TR = 2000ms, TE = 30ms, 90° flip angle, 256 x 256 FOV, 64 x 64 matrix size).

fMRI Data Pre-Processing. Prior to pre-processing, data were visually inspected for artifacts and anatomical abnormalities. fMRI data were pre-processed and analyzed using the fMRI Expert Analysis Tool (FEAT, version 6.00) of the FMRIB Software Library package (FSL, version 5.0.9; fsl.fmrib.ox.ac.uk). Pre-processing consisted of using the brain extraction tool (BET) to remove non-brain tissue from functional and structural runs, spatial realignment of functional volumes to correct for head motion using MCFLIRT, and hi-pass filtering the data (100s cutoff). The extent of participant head motion was further estimated by running FSL Motion Outliers to record volumes that exceeded a 0.9 mm threshold of framewise displacement

NEURAL VARIABILITY AND EMOTION REGULATION

(FD) (Siegel et al., 2014). Runs were discarded if participants exceeded this threshold for more than 25 volumes (~20% of a single run). Six participants had at least one run discarded due to excessive head motion (Mean number of discarded runs for eligible participants = 2 runs). The average number of volumes exceeding our FD threshold per run, per participant was 2.333 ($SD = 4.04$, range = 0-16.5); prior to discarding runs it was 3.014 ($SD = 6.09$, range = 0 - 30.75). Since a goal of the study was to examine how spatial variability—both between subjects and across time within subjects—related to age and experiences of negative affect, we elected not to spatially smooth our data. Data were pre-whitened to account for autocorrelated residuals across time. In addition to spatial and temporal variability analyses described below, we also ran traditional univariate analyses. A description of preprocessing for said analyses and their results can be found in the Supplement.

ROI Definition. We identified seven brain regions implicated in emotion regulation from a prior meta-analysis (Buhle, Silvers, et al., 2014). For the rest of this report, we refer to these seven regions as “ROIs”. Each ROI from the original meta-analysis contained one global maxima and at least one other local maxima (that is, local within the cluster). Global and local maxima from these ROIs are described as “spheres”.

We defined ROIs based on a multi-kernel density meta-analysis of cognitive reappraisal fMRI studies (Buhle, Silvers, et al., 2014) and created spheres around the maxima in each ROI (global and local). ROIs were identified based on clusters reported in Table 2 of Buhle, Silvers, et al., (2014), resulting in 7 ROIs: right dorsolateral prefrontal cortex (R dlPFC), left dorsolateral prefrontal cortex (L dlPFC), right ventrolateral prefrontal cortex (R vlPFC), right dorsomedial prefrontal cortex (R dmPFC), left middle temporal gyrus (L MTG), left superior parietal lobule (L SPL), and right superior parietal lobule (R SPL). We then drew spherical masks (4mm radius)

NEURAL VARIABILITY AND EMOTION REGULATION

around each maxima (global and local). On the rare occasions that spheres extended beyond the boundaries of the brain, spheres were moved inward. Our choice to use 4mm radius spheres was motivated by the fact that anything smaller would have rendered an insufficient number of voxels with which to calculate Gini coefficients, and that anything larger would have resulted in overlapping spheres. Clusters varied substantially in size and some had multiple local maxima (i.e., subclusters). In total, we created 32 spheres across the seven ROIs (7 L dlPFC; 3 R dlPFC; 4 R vlPFC; 7 R dmPFC; 2 L MTG; 5 L SPL; 4 R SPL). Spatial and temporal variability estimates were computed using their respective GLMs (described below) with this set of spheres. Estimates of variability from each sphere were then submitted to multi-level measurement models for further analysis (described in the ‘Statistical Approach’ section). Figure 2 provides an illustration of our ROI definition. Because our focus was on understanding variability in the regions known to instantiate top-down emotion regulation, we did not evaluate either type of variability in the amygdala.

Spatial Variability Estimation. We first submitted each participant’s data to a fixed effects analysis using a standard general linear model (GLM) in FSL. The reappraisal task was modeled with five task regressors, each convolved with the canonical HRF (double gamma). A regressor for the instructional cue (‘Instruction’), one for each task condition (‘Far’, ‘Look-Negative’, & ‘Look-Neutral’), and a final regressor for the affect rating period (‘Rating’) were modeled. Slice-timing effects were corrected for by including the temporal derivative of each task regressor in the model. Rotation and translation parameters obtained from MCFLIRT, along with their derivatives and the squares of each, were added as nuisance regressors to reduce the effects of head motion on functional data. Volumes exceeding 0.9 mm in FD were censored at this stage of this analysis using output from FSL Motion Outliers. A second level analysis, which

NEURAL VARIABILITY AND EMOTION REGULATION

averaged contrast estimates within subject, was carried out using a fixed effects model by forcing the random effects variance to zero. Registration of functional data to the high resolution structural (MPRAGE) was carried out using FSL's boundary based registration algorithm (Greve & Fischl, 2009). Each participant's MPRAGE was then non-linearly registered to the MNI152 template image (10mm warp resolution), and the transformation matrix was subsequently used to warp the functional images.

We used univariate activation estimates from the voxels within each sphere to calculate Gini coefficients, a simple but powerful way to quantify spatial variability (Guest & Love, 2017; Leech et al., 2014; Pyatt, 1976). The Gini coefficient was originally developed to study income inequality within specified geographic locations (e.g., cities, countries; Pyatt, 1976). Gini coefficients can range in value from 0 to 1. In the context of income inequality, a Gini coefficient of 0 means that everyone in a given location has exactly the same income; a coefficient of 1 means that one person has all the income and no one else has any. In the context of fMRI data, Gini coefficients can be used to measure the inequality of activation (i.e., BOLD response) among voxels within a given ROI, with greater inequality of activation implying that a smaller subset of voxels account for a larger portion of BOLD signal within the ROI during a given psychological process. In other words, a higher Gini coefficient (i.e., value closer to 1) represents greater inequality of neural activation within a sphere and means that fewer voxels are accounting for more of the activity within the ROI. As this occurs, more activation is peaked around said voxels and less activity is distributed across the rest of the sphere. Larger Gini coefficients indicate that relatively more activation is distributed across a relatively smaller space. Therefore, higher coefficients correspond to less spatial variability across a sphere. In other words, a greater Gini coefficient indicates activity is confined to fewer voxels and is less

NEURAL VARIABILITY AND EMOTION REGULATION

free to vary across the sphere (see Figure 3 for a visualization). Gini coefficients are useful because they can yield a single numerical index of spatial variability, thus helping to quantify complex theoretical concepts such as focalization. Our procedure for quantifying spatial variability via Gini coefficients follows.

As stated previously, our primary focus was on identifying variability during emotion regulation. As such, parameter estimates from the GLM were used to create a linear contrast image comparing the regulation condition (Far) to baseline. These voxelwise contrast values, in the form of Zstat images, were extracted from each sphere using the `cosmo_fmri_dataset()` command in the MatLab-based CoSMoMVPA toolbox (Oosterhof, Connolly, & Haxby, 2016). The subsequent vector of parameter estimates from each voxel within each sphere was sorted by magnitude and minimum-centered (i.e., we centered at the minimum by subtracting the lowest value in the vector from all elements). Minimum centering ensured that the Gini coefficients remained bound between 0 and 1. Following this step, we used the ordered, minimum-centered vector of parameter estimates to calculate a Gini coefficient. The equation follows.

$$(1) \quad \frac{\sum_{i=1}^n (2i-n-1)x_i}{n \sum_{i=1}^n x_i}$$

Where i represents the rank of a given voxel's activation, n is the total number of voxels, and x_i is the activation value (i.e., parameter estimate) for the i -th voxel. Overall, all subjects had 32 Gini coefficients, each corresponding to a sphere from our ROI list. Importantly, an additional analysis revealed that Gini coefficients were orthogonal to estimates of activity magnitude

NEURAL VARIABILITY AND EMOTION REGULATION

(details and caveats can be found in the Supplement). Figure 4 displays an overview of Gini coefficient calculation.

Temporal Variability Estimation. Temporal variability was estimated by taking the standard deviation of a beta-series (Rissman, Gazzaley, & D’Esposito, 2004) within each sphere. This was executed in three steps. First, we estimated brain activity for each trial using the Least Squares Single (LSS) approach (Mumford, Davis, & Poldrack, 2014; Mumford, Turner, Ashby, & Poldrack, 2012). A fixed-effects GLM was created for each regulation (Far) trial such that the trial of interest was given its own regressor in its own GLM and other trials for that condition were modeled with a separate nuisance regressor. Trials belonging to other conditions (e.g., ‘Look-Negative’, ‘Look-Neutral’, ‘Instruction’, etc.) and motion parameters (standard + extended + volumes exceeding 0.9mm FD) were modeled as their own regressors but not analyzed further. An LSS approach was selected because it is flexible for various types of analyses and results in less multi-collinearity between activation estimates for each trial compared to the Least Squares All (LSA) approach. The second step entailed using parameter estimates from each trial-specific GLM to create a linear contrast image comparing each regulate (Far) trial to baseline. We extracted average estimates of activation within a given sphere across the beta series from the Zmap of the regulate (Far) > baseline contrast. This resulted in an $n \times p$ matrix for each subject, where n is the number of trials and p is the number of spheres. Each matrix entry represents the mean activity of the p -th sphere at the n -th trial. The standard deviation of each column was then taken as the third and final step, yielding p estimates of temporal variability (where p corresponds to the number of spheres) for each subject. Each participant had 32 estimates of temporal variability, each corresponding to a sphere. Figure 5 depicts an overview of this estimation procedure.

NEURAL VARIABILITY AND EMOTION REGULATION

Statistical Approach.

We used multi-level measurement models to characterize spatial and temporal variability during emotion regulation. These models were further utilized to examine links between variability, age, and affective experience. Below, we describe our motivation for this approach and its benefits before elaborating the modeling procedure in detail. Code and data are publicly available on the open science framework (osf.io/42fkx/).

Motivation. As described above, we chose our ROIs based on clusters identified by a recent meta-analysis (Buhle et al., 2014). Importantly, the seven primary clusters of activation reported in this meta-analysis varied widely in terms of voxel size ($k = 77$ to $k = 517$). Using these whole clusters as ROIs would have been problematic, as some clusters would gain enhanced precision from data pooled across a greater number of voxels while making comparisons between regions difficult to interpret. One way to overcome this problem would be to draw a sphere around the global maxima for each cluster. However, this approach is also flawed because it ignores additional information from the rest of the cluster not included in the sphere. The approach used in the present manuscript represent an effort to incorporate as much information as possible from each ROI while maintaining an equal size for each sphere.

The first step of our approach involved drawing spheres (4mm radius) around *all* the maxima (both global and local) within each cluster defined by Buhle et al. (2014). We then estimated spatial and temporal variability within each maxima sphere and importantly, *nested* each local maxima sphere by its ROI (i.e., global maxima). We treated these measurements as manifest (i.e., observed) variables in a multi-level measurement model to estimate latent values for each ROI that varied between subjects. For example, the Buhle et al. meta-analysis identified a 175-voxel cluster in right dlPFC (labeled as right middle frontal gyrus in the meta-analysis).

NEURAL VARIABILITY AND EMOTION REGULATION

This large cluster's peak was at X=60, Y=24, Z=3 but also contained two local maxima (X=48, Y=24, Z=9 ; X=48, Y=15, Z=6). For this particular cluster, three spheres were constructed (one at each peak) and were then nested under a supra-heading of right dlPFC. This allowed us to accurately estimate indices of variability within each ROI that also appropriately incorporated the size of each ROI.

Our multi-level measurement model approach has several benefits. It makes use of data from many spheres within each ROI while using equal sized spheres between ROIs; it minimizes the possibility of including information that was still present in the original ROI-defining meta-analysis due to shared error variance (i.e., from method variance) across studies; it appropriately acknowledges the multi-level structure of the data (multiple measurements nested within participants); it provides a more analytically stable solution than simply running OLS regressions with many terms from our spheres in a single model and avoids the problem of multiple comparisons from running many smaller, ROI-specific OLS regression models.

Procedure. After obtaining estimates of spatial and temporal variability, we analyzed these metrics in two separate multi-level measurement models. Specifically, we estimated latent indices of spatial and temporal variability for each ROI and investigated their relationship with age and self-reported affect. Our analytic approach used multi-level modeling to take estimates of variability from spheres drawn around the maxima within each ROI and create a more accurate estimate for the ROI that they comprised. Put another way, the estimates of variability from each sphere of an ROI were used to calculate a latent value for the entire ROI. The relationships between each latent variable (one for each ROI) and its manifest variables (i.e., spheres) were allowed to vary by participant. We further modeled the variance of these latent variables for each ROI as a function of between-person variables of interest: age and average

NEURAL VARIABILITY AND EMOTION REGULATION

ratings of negative affect during emotion regulation. Two multi-level measurement models—one to measure latent spatial variability and another to measure latent temporal variability—were estimated using Hierarchical Linear Modeling (HLM for windows, version 6.06; Raudenbush & Byrk, 2002).

Within-Person Spatial Variability Model. The within-person measurement model for estimating spatial variability follows.

$$(2) G_{ip} = \pi_{1i} * \text{ldIPFC} + \pi_{2i} * \text{rdIPFC} + \pi_{3i} * \text{rvIPFC} + \pi_{4i} * \text{rdmPFC} + \pi_{5i} * \text{IMTG} + \pi_{6i} * \text{ISPL} + \pi_{7i} * \text{rSPL} + e_{ip}$$

Here, G_{ip} represents the Gini coefficient, our index of spatial variability, for the i -th individual at the p -th sphere. The slopes in this equation ($\pi_{1i} - \pi_{7i}$) each represent participant-specific, latent Gini coefficients for a given ROI. That is, they are the idealized estimates of Gini coefficients for each ROI, purged of measurement error. Variables labeled by ROI (e.g., ldIPFC, IMTG, etc) reflect overparameterized dummy codes that signify which ROI each G_{ip} belonged to. In other words, Equation 2 creates a latent, idealized Gini coefficient for every ROI using the observed Gini coefficients from each ROI's constituent spheres while discarding measurement error (e_{ij}). Notably, the lack of an intercept in this model is intentional, as it allows information about average Gini coefficients for each ROI to be encoded in the slopes, (i.e., π 's).

Within-Person Temporal Variability Model. The within-person measurement model for estimating temporal variability followed a nearly identical equation as the preceding model.

$$(3) SD_{ip} = \pi_{1i} * \text{ldIPFC} + \pi_{2i} * \text{rdIPFC} + \pi_{3i} * \text{rvIPFC} + \pi_{4i} * \text{rdmPFC} + \pi_{5i} * \text{IMTG} + \pi_{6i} * \text{ISPL} + \pi_{7i} * \text{rSPL} + e_{ip}$$

NEURAL VARIABILITY AND EMOTION REGULATION

SD_{ip} represents the standard deviation of the p -th sphere's beta-series for the i -th individual. Paralleling the previous equation, slopes in this equation represent idealized, participant-specific estimates of temporal variability. The same set-of overparameterized dummy codes were used to indicate to which ROI each sphere's temporal variability estimate "belonged".

Between-Person Model. Between-person equations for the two models were identical in form (but of course differentially represented spatial and temporal variability):

$$(4) \quad \pi_{1i} = \gamma_{10} + \gamma_{11} * \text{Age} + \gamma_{12} * \text{Neg. Aff.} + r_{1i}$$

$$(5) \quad \pi_{2i} = \gamma_{20} + \gamma_{21} * \text{Age} + \gamma_{22} * \text{Neg. Aff.} + r_{2i}$$

$$(6) \quad \pi_{3i} = \gamma_{30} + \gamma_{31} * \text{Age} + \gamma_{32} * \text{Neg. Aff.} + r_{3i}$$

$$(7) \quad \pi_{4i} = \gamma_{40} + \gamma_{41} * \text{Age} + \gamma_{42} * \text{Neg. Aff.} + r_{4i}$$

$$(8) \quad \pi_{5i} = \gamma_{50} + \gamma_{51} * \text{Age} + \gamma_{52} * \text{Neg. Aff.} + r_{5i}$$

$$(9) \quad \pi_{6i} = \gamma_{60} + \gamma_{61} * \text{Age} + \gamma_{62} * \text{Neg. Aff.} + r_{6i}$$

$$(10) \quad \pi_{7i} = \gamma_{70} + \gamma_{71} * \text{Age} + \gamma_{72} * \text{Neg. Aff.} + r_{7i}$$

In the between-person part of the model, participant-specific latent variables (π 's) are modeled as a function of overall fixed effects for the latent variables, age, and mean self-reported negative affect during active regulation (γ 's). The addition of participant specific deviations to the between-person part of the model allowed the within-person latent variables to vary between individuals (r 's). This allows us to test how age and affective experience relate to participants'

NEURAL VARIABILITY AND EMOTION REGULATION

spatial and temporal variability for each ROI. Both age and average ratings of negative affect (Neg. Aff.) were grand mean centered before being entered into the model.

Results

Behavioral Results. The average rating of mean participant negative affect during Far trials was 1.948 (SD = .499), meaning that our participants were on average rating the negative stimuli during reappraisal as inducing low-medium negative affect. The average ratings of negative affect during Look-Negative and Look-Neutral trials were 2.48 (SD = .562) and 1.08 (SD = 0.171), respectively. We conducted a paired samples t-test between average Far and Look-Negative ratings as a manipulation check, revealing significantly different means ($t(69) = -10.37$, $p < .001$). Age was unrelated to mean ratings of negative affect during Far-Negative ($r = -0.092$, $p > .250$), Look-Negative ($r = 0.184$, $p = .128$), and Look-Neutral ($r = 0.117$, $p > .250$) trials. However, the capacity to reappraise—defined as the percent change in ratings of negative affect between the Far and Look-Negative conditions—increased with age ($r = 0.377$, $p < .01$), consistent with prior work (Silvers et al., 2012). As noted earlier, the focus of the study was on emotion regulation so all subsequent analyses focus only on the Far condition.

Gini Coefficient Validation. Given that Gini Coefficients have only recently been introduced to neuroimaging research (Guest & Love, 2017; Leech et al., 2014), it was necessary to validate this novel technique in three ways. We first confirmed that Gini coefficients were not simply capturing information about the magnitude of neural activity. That is, our Gini coefficients were orthogonal to ROI means and peaks and thus represent information about *variability* and not magnitude (page 2 of Supplement). Following this, we next tested whether Gini coefficients reflected greater focalization of activation. Consistent with this hypothesis, we found that greater Gini coefficients reflected the spatial coalescence of an ROI's most active

NEURAL VARIABILITY AND EMOTION REGULATION

voxels. That is, the most active voxels in a given ROI were more likely to be closer together and not distributed across the entire sphere. Lastly, we sought to determine whether Gini coefficients were actually simply a proxy for gray matter tissue composition within our ROIs. Results indicated that Gini coefficients are independent of gray matter tissue composition. Statistical output and technical details about these analyses are reported in full in the Supplement.

Spatial Variability. As reported in Table 1, relatively similar estimates (ranging from .27-.28) of spatial variability (i.e., Gini coefficients) were observed across ROIs. Low Gini coefficients suggest that activity was distributed somewhat evenly over voxels within each ROI. Given the results of our validation analyses described above (and detailed in the supplement), these Gini coefficients are interpreted as potentially reflecting the presence of a centralized hub of activity, as the most active voxels were concentrated in a focal set of adjacent voxels. The fact that different ROIs had similar mean amounts of spatial variability implies a preserved spatial topography across ROIs during emotion regulation.

Across participants, estimates of latent spatial variability in left SPL were marginally related to age ($\gamma = .002$, $SE = .001$, $p = .051$; see Table 1). As age increased, fewer voxels in left SPL accounted for more activation than in younger participants. In terms of affect, lower levels of negative affect during reappraisal (i.e., more successful emotion regulation) were associated with a greater Gini coefficient in vIPFC activation ($\gamma = -.013$, $SE = .007$, $p = .049$). This means that successful emotion regulation in youth was associated with having fewer voxels in right vIPFC account for more of the activation.

Temporal Variability. Results of the temporal variability measurement model are listed in Table 2. In contrast to what was observed with spatial variability, temporal variability varied markedly across the 7 ROIs. Descriptively, temporal variability followed an apparent spatial

NEURAL VARIABILITY AND EMOTION REGULATION

gradient, such that prefrontal regions generally showed relatively less variability in activation across time compared to parietal and temporal regions.

We observed significant, and marginally significant, associations with age in right dmPFC ($\gamma = -.010$, $SE = .004$, $p = .005$), right vIPFC ($\gamma = -.009$, $SE = .005$, $p = .092$), and left MTG ($\gamma = -.007$, $SE = .004$, $p = .079$). Specifically, as age increased temporal variability across these three ROIs decreased. A supplemental analysis showed that estimates of temporal variability were modestly related to gray matter tissue composition (Supplementary Table 4). We thus re-ran our analyses while controlling for gray matter tissue composition in each sphere. Results indicated that the findings with right dmPFC and left MTG remained significant and marginally significant, respectively; the vIPFC trend did not remain marginally significant. Gray matter composition was directly associated with temporal variability in vIPFC and was inversely related to temporal variability in dmPFC; gray matter composition was unrelated to temporal variability for all other ROIs in addition to also having no relationship with age. Detailed statistical output of analyses concerning gray matter composition can be accessed in the Supplement (Supplementary Tables 5 & 6). These results suggest that age is accompanied by more stable, consistent brain activation in fronto-temporal ROIs implicated in emotion regulation, even when controlling for gray matter composition.

Discussion

Variability within-individuals is a critical feature of neural activity that has been previously overlooked in developmental neuroscience studies of emotion regulation. The present study sought to quantify two types of neural variability during emotion regulation—temporal and spatial—and relate them to age and individual differences in affective experience. We found that age was associated with spatial variability in SPL, while affective experience was associated

NEURAL VARIABILITY AND EMOTION REGULATION

with spatial variability in vIPFC. By contrast, age was linked with temporal variability in MTG and dmPFC. These results help characterize how spatial and temporal profiles of brain activity give rise to emotion regulatory abilities in youth and inform future work aimed at unpacking the role of variability in affective neurodevelopment.

Spatial Variability Marks Selective Specialization. We found that less spatial variability in right vIPFC activation was associated with less negative affect during emotion regulation. We further observed that age predicted less spatial variability in left SPL activation during emotion regulation. Follow-up analyses revealed that ROIs with less spatial variability tended to have highly active voxels tightly coalesced around a central focal point and less activation elsewhere. These results imply that the development of emotion regulation is supported by a shift towards decreased variability and increased spatial specialization in the prefrontal and parietal cortices. While the cross-sectional nature of the present study design precludes formal testing of this hypothesis, it may motivate future longitudinal work aimed at interrogating this possibility. Overall, our findings confer several important theoretical ramifications.

The present results enhance our understanding of the roles that vIPFC and SPL play in supporting the development of emotion regulation. Prior work suggests that these two regions are recruited to help meet the working memory and attentional demands of cognitive reappraisal (Buhle et al., 2014; Ochsner et al., 2012). Developmentally, both have also been linked to age-related improvements in cognitive control and emotion regulation (Durstun et al., 2006; Silvers et al., 2016). The present findings tentatively support and extend existing theories that posit that age-related changes in self-regulation hinge, in part, upon the ability to successfully deploy a burgeoning repertoire of executive functions (see Guassi Moreira & Silvers, 2018 and Calkins & Marcovitch, 2010 for overviews). Our results may help to explain prior work demonstrating that

NEURAL VARIABILITY AND EMOTION REGULATION

youth use effortful emotion regulation strategies like reappraisal less frequently and less effectively than older individuals (Mischel & Mischel, 1983; Mischel & Baker, 1975; Silvers et al., 2016). While prior work has tied the magnitude of prefrontal and parietal activation to developmental differences in emotion regulation (McRae et al., 2012; Silvers et al., 2016; Silvers, Shu, Hubbard, Weber, & Ochsner, 2015), here we have done the same with variability of activation in these regions. Although we found initial evidence that decreased spatial variability is associated with more successful emotion regulation (i.e., greater ability), it is important to note that we did not link neural variability to youth's trait-like tendency to engage in emotion regulation. While this possibility is enticing to consider for future research, our data can only fuel speculation on this topic. Regardless, the present findings suggest that prefrontal and parietal specialization might be as important to consider as overall activation when characterizing the neurodevelopment of self-regulatory abilities.

Outside of vIPFC and SPL, most other regions (e.g., dlPFC, dmPFC, etc) displayed diffuse spatial topographies during reappraisal and spatial variability did not differ as a function of age or affective experience. This suggests that age and affective experience co-vary with spatial variability in a selective subset of regions. This could indicate that (in youth, at least) certain cognitive features of regulation rely on diffusely distributed patterns of activity, whereas other features are instead dependent on a selective set of modular, specialized clusters of activity. If such is the case, it is interesting to consider why different patterns were observed in vIPFC and SPL than other brain regions. Given that these two brain regions have been particularly strongly linked with the variant of reappraisal employed here (emotional distancing), this could suggest that spatial variability is a meaningful indicator of age or individual differences only in brain regions most crucial for performing a given cognitive process. This raises the question of

NEURAL VARIABILITY AND EMOTION REGULATION

whether distinct or similar patterns of spatial variability might be observed for other psychological processes that change during childhood and adolescence. For example, if youth were asked to complete a theory of mind task (e.g., Spunt & Adolphs, 2014), we might expect age and task performance to be associated with less spatial variability in cortical midline structures like dmPFC that support social cognition (Blakemore & Mills, 2014; Pfeifer & Peake, 2012). While testing such possibilities is outside of the scope of the present study, the findings presented here may motivate future research aimed at examining how changes in spatial variability contribute to a range of neurodevelopmental processes in social cognitive and affective domains.

Another exciting notion raised by our results, consistent with a dynamic systems view of development (Smith & Thelen, 2003), is that each ROI represents a unique hub in an emotion regulation network, functions optimally at different amounts of within-individual variability, and follows its own developmental trajectory, nested within a broader hierarchical trajectory (e.g., the network's trajectory, the brain's trajectory). This notion follows from the fact that some of our ROIs were more variable than others and not all showed associations with age. Accordingly, the optimal levels of variability in each hub may be age-dependent, such that more variability is necessary at some ages and not during others. If such a 'variability-architecture' were to be confirmed by additional work, it would lend further support to extant neurodevelopmental theories of emotion regulation that posit regulation is comprised of emergent, but ultimately separable, psychological modules (Guassi Moreira & Silvers, 2018). Future research can directly test this by comparing ROI variability between youths and adults to identify the optimal, or normative, level of variability for each ROI, while also examining how such variability changes over the course of development to elucidate whether ROIs are similar or disparate in their

NEURAL VARIABILITY AND EMOTION REGULATION

maturational trajectories. Testing such possibilities, to which our data cannot currently speak to, encapsulates the next step in a line of research examining how spatial variability characterizes the functional architecture of a range of developmental phenotypes.

Temporal Variability Reflects Increased Stability with Age. We found that older individuals, relative to younger ones, showed decreased variability in neural responding over the course of the emotion regulation task. Specifically, age was associated with marginally less variable activation in left MTG, and significantly less variability in dmPFC. Changes in dmPFC variability are especially interesting considering that mentalizing processes are required to track one's self-regulation progress during reappraisal and other effortful emotion regulation strategies (Ochsner et al., 2012). Decreased temporal variability in dmPFC may be linked to developmental differences in maintaining goal states and self-monitoring, both of which are needed to guide future action (Kurby & Zacks, 2008; Northoff & Bermpohl, 2004; Richmond & Zacks, 2017). Greater temporal variability in dmPFC at younger ages could reflect an unstable or fluctuating ability to implement emotion regulation (Pfeifer & Peake, 2012). If this is the case, we might hypothesize that as individuals mature, their internal representations of self-regulatory states stabilize and dmPFC variability decreases. Despite having no way of empirically confirming the plausibility of this notion given our current experimental paradigm, it helps generate exciting new angles for future research.

Another explanation for the temporal variability results has to do with the social nature of the stimuli used in the present study. Specifically, our age-related findings in dmPFC may be partly explained in terms of ongoing maturation of the social brain (Blakemore & Mills, 2014; Pfeifer & Blakemore, 2012; Pfeifer & Peake, 2012). Social cognitive processes change dramatically during the first two decades of life, especially from late childhood through

NEURAL VARIABILITY AND EMOTION REGULATION

adolescence (Choudhury, Blakemore, & Charman, 2006; Eisenberg, Spinrad, & Knafo-Noam, 1998; Richardson et al., 2018). Such behavioral development has been linked to structural and functional maturation of cortical midline structures, including dmPFC (Blakemore & Mills, 2014; Mills, Goddings, Clasen, Giedd, & Blakemore, 2014; Mills, Lalonde, Clasen, Giedd, & Blakemore, 2014). It is possible that our age results in dmPFC were observed because the pruning of dmPFC pathways promotes a more consistent pattern of BOLD activation across repeated instances of regulation and less temporal variability (Blakemore & Mills, 2014; Mills, Lalonde, et al., 2014; Pfeifer & Blakemore, 2012; Somerville et al., 2013; Sowell et al., 2001).

Our findings demonstrating an inverse relationship between age and temporal variability are simultaneously consistent and at odds with the existing literature. On the one hand, some prior research in adult samples suggests that greater temporal variability is an adaptive feature of neural responding because it endows the brain with the ability to easily access different ‘network states’ that may be needed to complete mental tasks (Garrett, Kovacevic, McIntosh, & Grady, 2010; Garrett, Kovacevic, McIntosh, & Grady, 2011; Petroni et al., 2018). Thus, one might predict that age would predict more flexible (i.e., variable) responses. On the other hand, one could argue that older adolescents may simply need to exert less effort while regulating and display reduced temporal variability across repeated instances of regulation as a result. Indeed, our data appear to support this latter account – older individuals showed less temporal variability on a trial-by-trial basis than younger individuals in certain regions, consistent with prior developmental findings in other domains (Koolschijn, Schel, de Rooij, Rombouts, & Crone, 2011; Ordaz, Foran, Velanova, & Luna, 2013). Our findings might also be explained by prior work showing that age-related trajectories of temporal variability in the brain do not follow a single pattern, but instead show regionally-specific increases and decreases in variability (Nomi

NEURAL VARIABILITY AND EMOTION REGULATION

et al., 2017; Petroni et al., 2018). This raises the possibility that development is characterized by a process of ‘variability tuning’, where an intermediate amount of variability is necessary for the execution of psychological tasks, but what is classified as “intermediate” varies across brain regions. Our own findings cannot directly confirm or falsify this possibility, but the notion is supported by recently published work which espouses a ‘goldilocks’ view of variability (insufficient or excessive variability is detrimental) (Dinstein, Heeger, & Behrmann, 2015).

Limitations. This study is not without limitations, especially since it is the first to examine neural variability in the context of emotion regulation development. First and foremost, since we relied on a cross-sectional design, we cannot characterize true neurodevelopmental trajectories without longitudinal data (e.g., Snijders & Bosker, 1998; Singer & Willett, 2003). Relatedly, without a group of younger children and adults, it is hard to make inferences about developmental processes that extend beyond the adolescent and late childhood years (e.g., a developmental switch; Casey, 2015; Gee et al., 2014) Another limitation lies in our use of measurement models, which provide a simple way to summarize our indices of variability across each ROI, but (possibly incorrectly) assume that the microarchitecture of different cortical spheres is similar in nature.

The ROIs used in the present study were drawn from a meta-analysis that was nearly exclusively based on adult studies. This methodological choice was due to the fact that no meta-analyses based exclusively on pediatric studies exist (Guassi Moreira & Silvers, 2018), and that doing so provided us with a way to independently define ROIs (Kriegeskorte et al., 2009). Unfortunately, this means that the ROIs studied here may not be representative of the precise ROIs that youth use when engaging in emotion regulation. Indeed, it is possible that children use different brain regions than adolescents or adults in order to accomplish the same emotion

NEURAL VARIABILITY AND EMOTION REGULATION

regulation goals, although it is worth noting this is unlikely given that no prior data to our knowledge have consistently observed activations in children in brain regions not observed in adults. Future work in larger samples might opt to sidestep this issue by defining ROIs in a subset of their sample and using the rest of the sample for independent analyses or by collecting additional runs of an emotion regulation task specifically devoted to ROI localization and identification.

An additional, infrequently discussed limitation revolves around the notion that the scanner is anxiety-provoking and thus constitutes a unique emotional context (Eatough, Shirtcliff, Hanson, & Pollak, 2009; Galván, Van Leijenhorst, & McGlennen, 2012). If this were the case, it could impact regulatory processes in ways that obfuscate the true, ecological nature of youth's emotion regulation. That said, care was taken in the present study to reduce this possibility by familiarizing participants with the scanning environment through the use of mock scanning.

Given that our study was the first to examine variability in youth's neural responses during emotion regulation, we opted for an fMRI paradigm that was highly consistent with those previously used in youth (McLaughlin, Peverill, Gold, Alves, & Sheridan, 2015; McRae et al., 2012; Silvers et al., 2015). This decision led us to exclusively examine regulation of negative emotion (inclusive of multiple negative emotional states). It is possible that our results might not generalize to regulation of other emotional states and categories. Indeed, other work has suggested that age-related differences in the behavioral and neural correlates of emotion regulation vary across different types of emotional stimuli (Silvers et al., 2012, 2014). Future work ought to examine whether the results reported herein not only generalize across different

NEURAL VARIABILITY AND EMOTION REGULATION

variants of reappraisal and related emotion regulation strategies, but also across different emotional states.

Lastly, our study was exploratory by definition because it was the first of its kind. The present results do not confirm any hypotheses, but rather generate novel ones to be tested in future studies. This limitation, like the others above, must be taken into careful consideration when evaluating the impact of our findings with respect to the broader literature.

Concluding Remarks. Variability is a fundamental feature of neurodevelopment. Our results suggest that variability is also central to the acquisition of effective emotion regulation. This investigation was the first to incorporate neural variability into developmental research on emotion regulation. We showed that age was associated with increasing spatial focalization of activity in some brain regions, whereas other regions exhibited spatial topographies that were age- and experience-invariant. We also showed that age is marked by stability of neural responses over the course of repeated emotion regulation. Excitingly, this work contributes towards identifying the mechanisms that encode the neural bases of emotion regulation in youth.

NEURAL VARIABILITY AND EMOTION REGULATION

Acknowledgements

This research was supported by a grant (#MH103291) from the National Institute of Mental Health awarded to KAM. Preparation of this manuscript was supported by a National Science Foundation Graduate Research Fellowship (Fellow ID: 2016220797) to JFGM. Gratitude is extended towards Drs. Carolyn Parkinson and Jesse Rissman for their feedback about the study concept and towards Emilia Ninova for reflections on the manuscript.

NEURAL VARIABILITY AND EMOTION REGULATION

References

- Aldao, A., Sheppes, G., & Gross, J. J. (2015). Emotion Regulation Flexibility. *Cognitive Therapy and Research*, 39(3), 263–278. <https://doi.org/10.1007/s10608-014-9662-4>
- Blakemore, S.-J., & Mills, K. L. (2014). Is adolescence a sensitive period for sociocultural processing? *Annual Review of Psychology*, 65, 187–207. <https://doi.org/10.1146/annurev-psych-010213-115202>
- Buhle, J. T., Silvers, J. A., Wage, T. D., Lopez, R., Onyemekwu, C., Kober, H., ... Ochsner, K. N. (2014). Cognitive reappraisal of emotion: A meta-analysis of human neuroimaging studies. *Cerebral Cortex*, 24(11), 2981–2990. <https://doi.org/10.1093/cercor/bht154>
- Casey, B. J. (2015). Beyond simple models of self-control to circuit-based accounts of adolescent behavior. *Annual Review of Psychology*, 66, 295–319. <https://doi.org/10.1146/annurev-psych-010814-015156>
- Chang, L. J., Gianaros, P. J., Manuck, S. B., & Krishnan, A. (2015). A sensitive and specific neural signature for picture-induced negative affect. *PLOS Biology*, 13(6), 1–28. <https://doi.org/10.1371/journal.pbio.1002180>
- Choudhury, S., Blakemore, S. J., & Charman, T. (2006). Social cognitive development during adolescence. *Social Cognitive and Affective Neuroscience*, 1(3), 165–174. <https://doi.org/10.1093/scan/nsl024>
- Christophel, T. B., Iamshchinina, P., Yan, C., Allefeld, C., & Haynes, J.-D. (2018). Cortical specialization for attended versus unattended working memory. *Nature Neuroscience*. <https://doi.org/10.1038/s41593-018-0094-4>
- Cole, P. M., & Deater-Deckard, K. (2009). Emotion regulation, risk, and psychopathology. *Journal of Child Psychology and Psychiatry and Allied Disciplines*, 50(11), 1327–1330. <https://doi.org/10.1111/j.1469-7610.2009.02180.x>
- Crone, E. A., & van der Molen, M. W. (2004). Developmental changes in real life decision making: Performance on a gambling task previously shown to depend on the ventromedial prefrontal cortex. *Developmental Neuropsychology*, 25(3), 251–279. <https://doi.org/10.1207/s15326942dn2503>
- Dehaene-Lambertz, G., Monzalvo, K., & Dehaene, S. (2018). The emergence of the visual word form: Longitudinal evolution of category-specific ventral visual areas during reading acquisition. *PLOS Biology*, 1–34. <https://doi.org/10.1371/journal.pbio.2004103>
- Dinstein, I., Heeger, D. J., & Behrmann, M. (2015). Neural variability: Friend or foe? *Trends in Cognitive Sciences*, 19(6), 322–328. <https://doi.org/10.1016/j.tics.2015.04.005>
- DuPre, E., & Spreng, R. N. (2017). Structural covariance networks across the lifespan, from 6-94 years of age. *Network Neuroscience*, 1–38. https://doi.org/10.1162/NETN_a_00016
- Durston, S., Davidson, M. C., Tottenham, N., Galvan, A., Spicer, J., Fossella, J. A., & Casey, B. J. (2006). A shift from diffuse to focal cortical activity with development. *Developmental Science*, 9(1), 1–8. <https://doi.org/10.1111/j.1467-7687.2005.00454.x>
- Eatough, E. M., Shirtcliff, E. A., Hanson, J. L., & Pollak, S. D. (2009). Hormonal reactivity to MRI scanning in adolescents. *Psychoneuroendocrinology*, 34(8), 1242–1246. <https://doi.org/10.1111/j.1743-6109.2008.01122.x>
- Eisenberg, N., Spinrad, T. L., & Knafo-Noam, A. (1998). *Prosocial Development*. John Wiley & Sons. <https://doi.org/10.1002/9781118963418.childpsy315>
- Etzel, J. A., Zacks, J. M., & Braver, T. S. (2013). Searchlight analysis: Promise, pitfalls, and potential. *NeuroImage*, 78, 261–269. <https://doi.org/10.1016/j.actbio.2009.04.013>

NEURAL VARIABILITY AND EMOTION REGULATION

- Foulkes, L., & Blakemore, S. J. (2018). Studying individual differences in human adolescent brain development. *Nature Neuroscience*, 1–9. <https://doi.org/10.1038/s41593-018-0078-4>
- Galván, A., Van Leijenhorst, L., & McGlennen, K. M. (2012). Considerations for imaging the adolescent brain. *Developmental Cognitive Neuroscience*, 2(3), 293–302. <https://doi.org/10.1016/j.dcn.2012.02.002>
- Garrett, D. D., Kovacevic, N., McIntosh, A. R., & Grady, C. L. (2010). Blood oxygen level-dependent signal variability is more than just noise. *The Journal of Neuroscience*, 30(14), 4914–4921. <https://doi.org/10.1523/JNEUROSCI.5166-09.2010>
- Garrett, D. D., Kovacevic, N., McIntosh, A. R., & Grady, C. L. (2011). The importance of being variable. *Journal of Neuroscience*, 31(12), 4496–4503. <https://doi.org/10.1523/JNEUROSCI.5641-10.2011>
- Gross, J. J. (2015). Emotion regulation: Current status and future prospects. *Psychological Inquiry*, 26(1), 1–26. <https://doi.org/10.1080/1047840X.2014.940781>
- Guassi Moreira, J. F., & Silvers, J. A. (2018). In due time: Neurodevelopmental considerations in the study of emotion regulation. In P. M. Cole & T. Hollenstein (Eds.), *Emotion Regulation: A Matter of Time* (pp. 111–134). New York: Routledge.
- Guest, O., & Love, B. C. (2017). What the success of brain imaging implies about the neural code. *ELife*, 6, 1–16. <https://doi.org/10.7554/eLife.21397>
- Haxby, J. V., Gobbini, M. I., Furey, M. L., Ishai, A., Schouten, J. L., & Pietrini, P. (2001). Distributed and overlapping representations of face and objects in ventral temporal cortex. *Science*, 293(5539), 2425–2430. <https://doi.org/10.1126/science.1063736>
- Heller, A. S., & Casey, B. J. (2016). The neurodynamics of emotion: Delineating typical and atypical emotional processes during adolescence. *Developmental Science*, 19(1), 3–18. <https://doi.org/10.1111/desc.12373>
- Huth, A. G., Heer, W. A. De, Griffiths, T. L., Theunissen, F. E., & Gallant, J. L. (2016). Natural speech reveals the semantic maps that tile human cerebral cortex. *Nature*, 532(7600), 453–458. <https://doi.org/10.1038/nature17637>
- Huth, A. G., Nishimoto, S., Vu, A. T., & Gallant, J. L. (2012). A continuous semantic space describes the representation of thousands of object and action categories across the human brain. *Neuron*, 76(6), 1210–1224. <https://doi.org/10.1016/j.neuron.2012.10.014>
- Kim, J. H., & Richardson, R. (2010). New findings on extinction of conditioned fear early in development: Theoretical and clinical implications. *Biological Psychiatry*, 67(4), 297–303. <https://doi.org/10.1016/j.biopsych.2009.09.003>
- Koolschijn, P. C. M. P., Schel, M. A., de Rooij, M., Rombouts, S. A. R. B., & Crone, E. A. (2011). A three-year longitudinal functional magnetic resonance imaging study of performance monitoring and test-retest reliability from childhood to early adulthood. *Journal of Neuroscience*, 31(11), 4204–4212. <https://doi.org/10.1523/JNEUROSCI.6415-10.2011>
- Kriegeskorte, N., Cusack, R., & Bandettini, P. (2010). How does an fMRI voxel sample the neuronal activity pattern: Compact-kernel or complex spatiotemporal filter? *NeuroImage*, 49(3), 1965–1976. <https://doi.org/10.1016/j.neuroimage.2009.09.059>
- Kriegeskorte, N., Mur, M., Ruff, D. A., Kiani, R., Bodurka, J., Esteky, H., ... Bandettini, P. A. (2008). Matching categorical object representations in inferior temporal cortex of man and monkey. *Neuron*, 60(6), 1126–1141. <https://doi.org/10.1016/j.neuron.2008.10.043>
- Kriegeskorte, N., Simmons, W. K., Bellgowan, P. S., & Baker, C. I. (2009). Circular analysis in systems neuroscience: The dangers of double dipping. *Nature Neuroscience*, 12(5), 535–

NEURAL VARIABILITY AND EMOTION REGULATION

540. <https://doi.org/10.1038/nn.2303>
- Kurby, C. A., & Zacks, J. M. (2008). Segmentation in the perception and memory of events. *Trends in Cognitive Sciences*, 12(2), 72–79. <https://doi.org/10.1016/j.tics.2007.11.004>
- Lang, P. J., Bradley, M. M., & Cuthbert, B. N. (2008). *International affective picture system (IAPS): Affective ratings of pictures and instruction manual. Technical Report A-8*. <https://doi.org/10.1016/j.epr.2006.03.016>
- Larson, R., Csikszentmihalyi, M., & Graef, R. (1980). Mood variability and the psycho-social adjustment of adolescents. *Journal of Youth & Adolescence*, 9(6), 469–490. https://doi.org/10.1007/978-94-017-9094-9_15
- Leech, R., Scott, G., Carhart-Harris, R., Turkheimer, F., Taylor-Robinson, S. D., & Sharp, D. J. (2014). Spatial dependencies between large-scale brain networks. *PLoS ONE*, 9(6). <https://doi.org/10.1371/journal.pone.0098500>
- Luciana, M., Wahlstrom, D., Porter, J. N., & Collins, P. F. (2012). Dopaminergic modulation of incentive motivation in adolescence: Age-related changes in signaling, individual differences, and implications for the development of self-regulation. *Developmental Psychology*, 48(3), 844–861. <https://doi.org/10.1037/a0027432>
- McLaughlin, K. A., Garrard, M. C., & Somerville, L. H. (2015). What develops during emotional development? A component process approach to identifying sources of psychopathology risk in adolescence. *Dialogues in Clinical Neuroscience*, 17(4), 403–410.
- McLaughlin, K. A., Peverill, M., Gold, A. L., Alves, S., & Sheridan, M. A. (2015). Child maltreatment and neural systems underlying emotion regulation. *Journal of the American Academy of Child and Adolescent Psychiatry*, 54(9), 753–62. <https://doi.org/10.1016/j.jaac.2015.06.010>
- McRae, K., Gross, J. J., Weber, J., Robertson, E. R., Sokol-Hessner, P., Ray, R. D., ... Ochsner, K. N. (2012). The development of emotion regulation: An fMRI study of cognitive reappraisal in children, adolescents and young adults. *Social Cognitive and Affective Neuroscience*, 7(1), 11–22. <https://doi.org/10.1093/scan/nsr093>
- Mills, K. L., Goddings, A. L., Clasen, L. S., Giedd, J. N., & Blakemore, S. J. (2014). The developmental mismatch in structural brain maturation during adolescence. *Developmental Neuroscience*, 36(3–4), 147–160. <https://doi.org/10.1159/000362328>
- Mills, K. L., Lalonde, F., Clasen, L. S., Giedd, J. N., & Blakemore, S. J. (2014). Developmental changes in the structure of the social brain in late childhood and adolescence. *Social Cognitive and Affective Neuroscience*, 9(1), 123–131. <https://doi.org/10.1093/scan/nss113>
- Mischel, H. N., & Mischel, W. (1983). The development of children's knowledge of self-control strategies. *Child Development*, 54(3), 603. <https://doi.org/10.2307/1130047>
- Mischel, W., & Baker, N. (1975). Cognitive appraisals and transformations in delay behavior. *Journal of Personality and Social Psychology*, 31(2), 254–261. <https://doi.org/10.1037/h0076272>
- Mumford, J. A., Davis, T., & Poldrack, R. A. (2014). The impact of study design on pattern estimation for single-trial multivariate pattern analysis. *NeuroImage*, 103, 130–138. <https://doi.org/10.1016/j.neuroimage.2014.09.026>
- Mumford, J. A., Turner, B. O., Ashby, F. G., & Poldrack, R. A. (2012). Deconvolving BOLD activation in event-related designs for multivoxel pattern classification analyses. *NeuroImage*, 59(3), 2636–2643. <https://doi.org/10.1016/j.neuroimage.2011.08.076>
- Nomi, J. S., Bolt, T. S., Ezie, C. E. C., Uddin, L. Q., & Heller, A. S. (2017). Moment-to-moment BOLD Signal variability reflects regional changes in neural flexibility across the lifespan.

NEURAL VARIABILITY AND EMOTION REGULATION

- The Journal of Neuroscience*, 37(22), 5539–5548.
<https://doi.org/10.1523/JNEUROSCI.3408-16.2017>
- Northoff, G., & Bermpohl, F. (2004). Cortical midline structures and the self. *Trends in Cognitive Sciences*, 8(3), 102–107. <https://doi.org/10.1016/j.tics.2004.01.004>
- Ochsner, K. N., & Gross, J. J. (2005). The cognitive control of emotion. *Trends in Cognitive Sciences*.
- Ochsner, K. N., Ray, R. D., Cooper, J. C., Robertson, E. R., Chopra, S., Gabrieli, J. D. E., & Gross, J. J. (2004). For better or for worse: Neural systems supporting the cognitive down- and up-regulation of negative emotion. *NeuroImage*, 23(2), 483–499.
<https://doi.org/10.1016/j.neuroimage.2004.06.030>
- Ochsner, K. N., Silvers, J. A., & Buhle, J. T. (2012). Review and evolving model of the cognitive control of emotion. *Annals of the New York Academy of Sciences*, (1251), E1–E24. <https://doi.org/10.1111/j.1749-6632.2012.06751.x>.Functional
- Oosterhof, N. N., Connolly, A. C., & Haxby, J. V. (2016). CoSMoMVPA: Multi-modal multivariate pattern analysis of neuroimaging data in Matlab/GNU Octave. *Frontiers in Neuroinformatics*, 10, 1–27. <https://doi.org/10.3389/fninf.2016.00027>
- Ordaz, S. J., Foran, W., Velanova, K., & Luna, B. (2013). Longitudinal growth curves of brain function underlying inhibitory control through adolescence. *The Journal of Neuroscience*, 33(46), 18109–18124. <https://doi.org/10.1523/JNEUROSCI.1741-13.2013>
- Park, D. C., Polk, T. A., Park, R., Minear, M., Savage, A., & Smith, M. R. (2004). Aging reduces neural specialization in ventral visual cortex. *Proceedings of the National Academy of Sciences*, 101(35), 13091–13095. <https://doi.org/10.1073/pnas.0405148101>
- Patel, G. H., Kaplan, D. M., & Snyder, L. H. (2014). Topographic organization in the brain: Searching for general principles. *Trends in Cognitive Sciences*, 18(7), 351–363.
<https://doi.org/10.1016/j.tics.2014.03.008>
- Petroni, A., Cohen, S. S., Ai, L., Langer, N., Henin, S., Vanderwal, T., ... Parra, L. C. (2018). The variability of neural responses to naturalistic videos change with age and sex. *ENEURO*, 5(February), ENEURO.0244-17.2017. <https://doi.org/10.1523/ENEURO.0244-17.2017>
- Pfeifer, J. H., & Blakemore, S. J. (2012). Adolescent social cognitive and affective neuroscience: Past, present, and future. *Social Cognitive and Affective Neuroscience*, 7(1), 1–10.
<https://doi.org/10.1093/scan/nsr099>
- Pfeifer, J. H., & Peake, S. J. (2012). Self-development: Integrating cognitive, socioemotional, and neuroimaging perspectives. *Developmental Cognitive Neuroscience*, 2(1), 55–69.
<https://doi.org/10.1016/j.dcn.2011.07.012>
- Pinneo, L. R. (1966). On noise in the nervous system. *Psychological Review*, 73(3), 242–247.
- Poldrack, R. A. (2015). Is “efficiency” a useful concept in cognitive neuroscience?
Developmental Cognitive Neuroscience, 11, 12–17.
<https://doi.org/10.1016/j.dcn.2014.06.001>
- Polk, T. A., Stallcup, M., Aguirre, G. K., Alsop, D. C., D’Esposito, M., Detre, J. A., & Farah, M. J. (2002). Neural specialization for letter recognition. *Journal of Cognitive Neuroscience*, 14(2), 145–159. <https://doi.org/10.1162/089892902317236803>
- Pyatt, G. (1976). On the interpretation and disaggregation of gini coefficients. *The Economic Journal*, 86(342), 243–255.
- Richardson, H., Lisandrelli, G., Riobueno-Naylor, A., & Saxe, R. (2018). Development of the social brain from age three to twelve years. *Nature Communications*, 9(1), 1027.
<https://doi.org/10.1038/s41467-018-03399-2>

NEURAL VARIABILITY AND EMOTION REGULATION

- Richmond, L. L., & Zacks, J. M. (2017). Constructing Experience: Event Models from Perception to Action. *Trends in Cognitive Sciences*, 21(12), 962–980. <https://doi.org/10.1016/j.tics.2017.08.005>
- Rissman, J., Gazzaley, A., & D’Esposito, M. (2004). Measuring functional connectivity during distinct stages of a cognitive task. *NeuroImage*, 23(2), 752–763. <https://doi.org/10.1016/j.neuroimage.2004.06.035>
- Roalf, D. R., Gur, R. E., Ruparel, K., Calkins, M., Satterthwaite, T., Bilker, W. B., ... Ruben, C. (2014). Within-individual variability in neurocognitive performance: age and sex-related differences in children and youths from ages 8 to 21. *Neuropsychology*, 28(4), 506–518. <https://doi.org/10.1037/neu0000067>. Within-Individual
- Seghier, M. L., & Price, C. J. (2018). Interpreting and utilising intersubject variability in brain function. *Trends in Cognitive Sciences*, 22(6), 517–530. <https://doi.org/10.1016/j.tics.2018.03.003>
- Siegel, J. S., Power, J. D., Dubis, J. W., Vogel, A. C., Church, J. A., Schlaggar, B. L., & Petersen, S. E. (2014). Statistical improvements in functional magnetic resonance imaging analyses produced by censoring high-motion data points. *Human Brain Mapping*, 35(5), 1981–1996. <https://doi.org/10.1002/hbm.22307>. Statistical
- Siegler, R. S. (1994). Cognitive Variability: A Key to Understanding Cognitive Development. *Current Directions in Psychological Science*, 3(1), 1–5.
- Siegler, R. S. (2007). Cognitive variability. *Developmental Science*, 10(1), 104–109. <https://doi.org/10.1111/j.1467-7687.2007.00571.x>
- Silvers, J. A., Insel, C., Powers, A., Franz, P., Helion, C., Martin, R. E., ... Ochsner, K. N. (2016). vIPFC – vmPFC – amygdala interactions underlie age-related differences in cognitive regulation of emotion. *Cerebral Cortex*, ([Epub Ahead of Print]), 1–13. <https://doi.org/10.1093/cercor/bhw073>
- Silvers, J. A., Insel, C., Powers, A., Franz, P., Weber, J., Mischel, W., ... Ochsner, K. N. (2014). Curbing craving: Behavioral and brain evidence that children regulate craving when instructed to do so but have higher baseline craving than adults. *Psychological Science*, 25(10), 1932–1942. <https://doi.org/10.1177/0956797614546001>
- Silvers, J. A., McRae, K., Gabrieli, J. D. E., Gross, J. J., Remy, K. A., & Ochsner, K. N. (2012). Age-related differences in emotional reactivity, regulation, and rejection sensitivity in adolescence. *Emotion*, 12(6), 1235–1247. <https://doi.org/10.1037/a0028297>. Age-Related
- Silvers, J. A., Shu, J., Hubbard, A. D., Weber, J., & Ochsner, K. N. (2015). Concurrent and lasting effects of emotion regulation on amygdala response in adolescence and young adulthood. *Developmental Science*, 18(5), 771–784. <https://doi.org/10.1111/desc.12260>
- Smith, L. B., & Thelen, E. (2003). Development as a dynamic system. *Trends in Cognitive Sciences*, 7(8), 343–348. [https://doi.org/10.1016/S1364-6613\(03\)00156-6](https://doi.org/10.1016/S1364-6613(03)00156-6)
- Somerville, L. H., Jones, R. M., & Casey, B. J. (2010). A time of change: Behavioral and neural correlates of adolescent sensitivity to appetitive and aversive environmental cues. *Brain and Cognition*, 72(1), 124–133. <https://doi.org/10.1016/j.bandc.2009.07.003>
- Somerville, L. H., Jones, R. M., Ruberry, E. J., Dyke, J. P., Glover, G., & Casey, B. J. (2013). The medial prefrontal cortex and the emergence of self-conscious emotion in adolescence. *Psychological Science*, 24(8), 1554–1562. <https://doi.org/10.1177/0956797613475633>
- Sowell, E. R., Thompson, P. M., Tessner, K. D., & Toga, A. W. (2001). Mapping continued brain growth and gray matter density reduction in dorsal frontal cortex: Inverse relationships during postadolescent brain maturation. *The Journal of Neuroscience*, 21(22),

NEURAL VARIABILITY AND EMOTION REGULATION

8819–29. <https://doi.org/21/22/8819> [pii]

Thompson, R., & Goodman, M. (2010). Development of emotion regulation. In A. M. Kring & D. M. Sloan (Eds.), *Emotion Regulation and Psychopathology: A transdiagnostic Approach to Etiology and Treatment* (pp. 38–58). New York, NY: The Guilford Press.

Vij, S. G., Nomi, J. S., Dajani, D. R., & Uddin, L. Q. (2018). Evolution of spatial and temporal features of functional brain networks across the lifespan. *NeuroImage*.
<https://doi.org/10.1016/j.neuroimage.2018.02.066>

NEURAL VARIABILITY AND EMOTION REGULATION

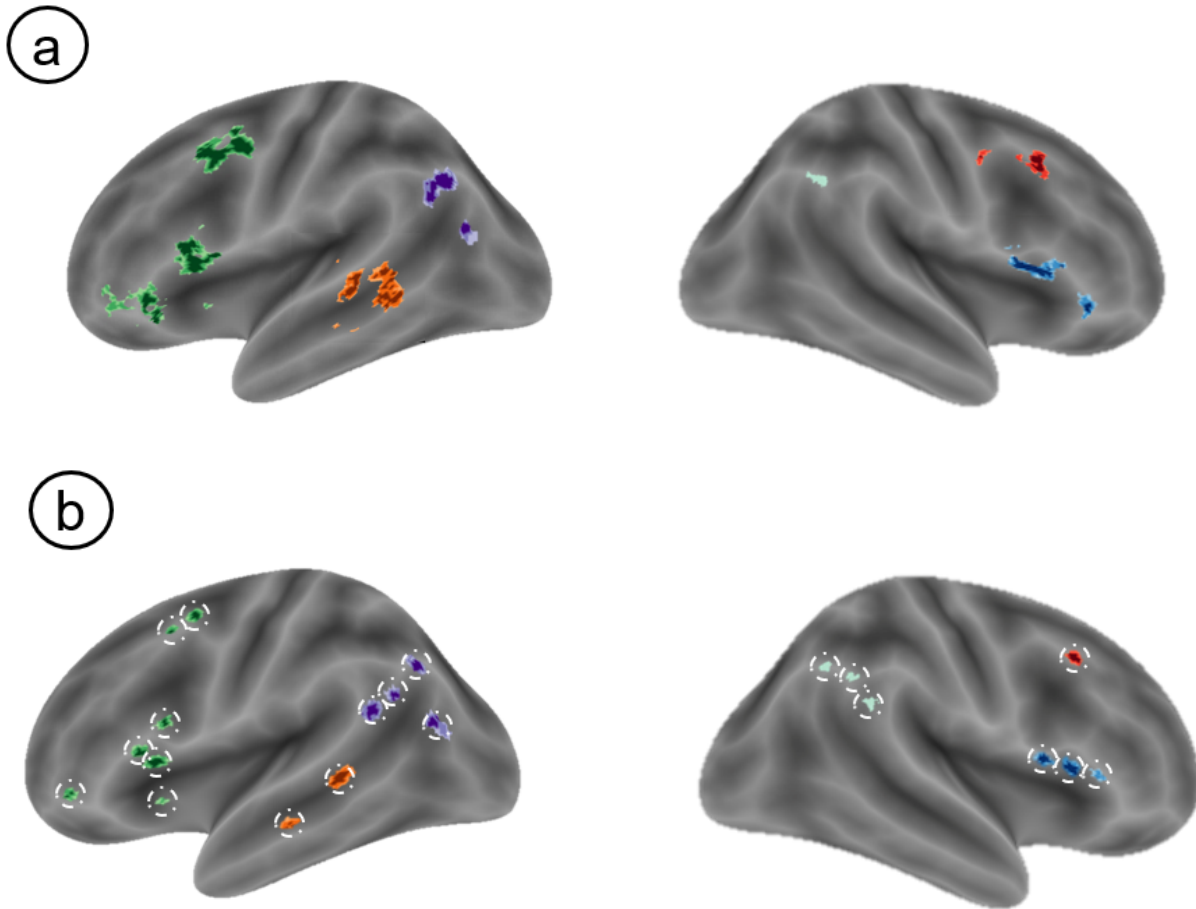
Figure 1. Overview of Emotion Regulation Paradigm



Note. 's' refers to seconds. 'Far' refers to the cue participants received to regulate via distancing; passive viewing cue ('Look') is not pictured. Participants regulated aversive images and passively viewed both aversive and neutral images (not pictured). Each trial was followed by a jittered ITI lasting between 1.5-6.5s.

NEURAL VARIABILITY AND EMOTION REGULATION

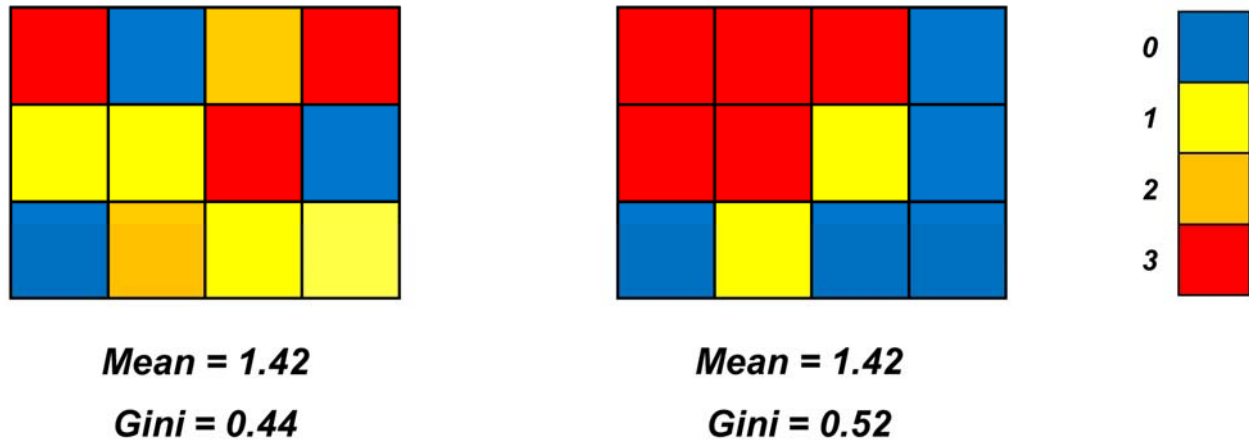
Figure 2. Illustrative Schematic of ROI Definition.



Note. Clusters from a prior meta-analysis (a) were used to define spheres nested within ROIS (b). For clarity, not all significant clusters from Buhle, Silvers (2014) are depicted in the figure. Only left dlPFC (green), left MTG (orange), left SPL (purple), right SPL (light turquoise), right dlPFC (red) and right vlPFC (blue) are depicted and dmPFC has been omitted. Certain ROIs contained spheres (e.g., right dlPFC) are not shown because they were below the lateral surface, could not be surface rendered.

NEURAL VARIABILITY AND EMOTION REGULATION

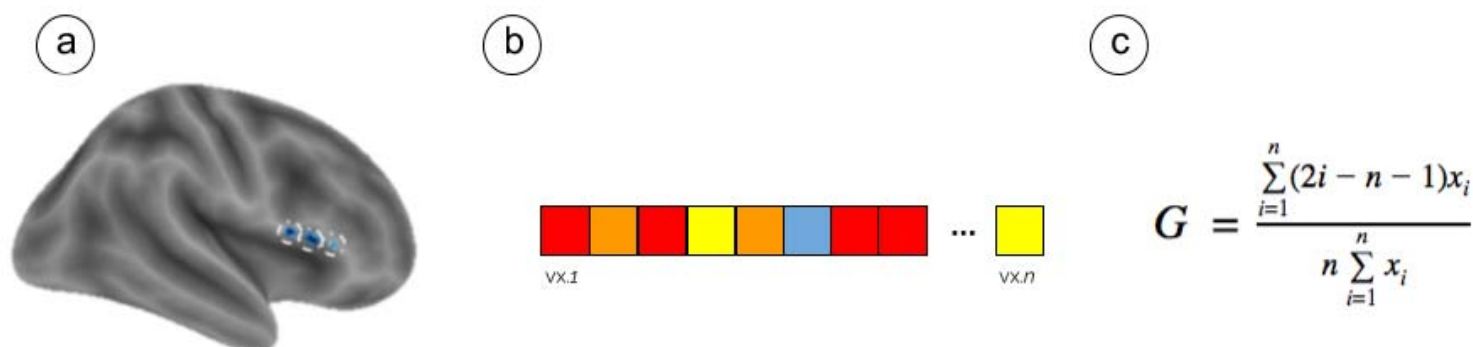
Figure 3. Conceptual Overview of Gini Coefficients and Spatial Variability



Note. The two grids each represent hypothetical voxels from an ROI, with color codes indicating each individual voxel's level of activity. Though the mean level of activity is the same in each voxel, the grid on the right has fewer voxels accounting for more of the ROI's activation, with overall activity less diffusely distributed across the ROI. In this manner, Gini coefficients can measure the spatial variability of an ROI.

NEURAL VARIABILITY AND EMOTION REGULATION

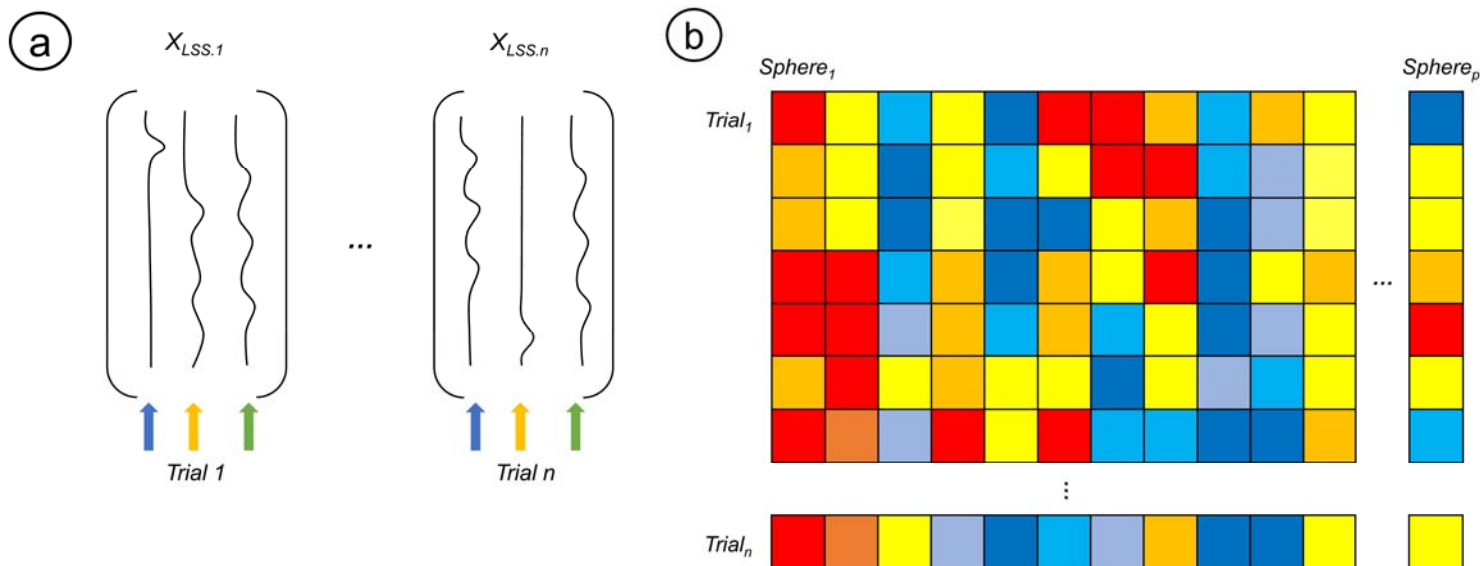
Figure 4. Using Gini Coefficients to Quantify Spatial Variability



Note. Gini coefficients were computed across a series of spheres for neural activity during emotion regulation relative to baseline. Spheres drawn from a set of *a priori* ROIs, such as the exemplars depicted in (a), were used to extract data from activation maps of regulate (Far) > Baseline (fixation) for each participant. The data consisted of a vector of voxels falling within the sphere ($vx.1$ to $vx.n$), each with an estimate of activation for their respective condition (b). This vector was sorted in ascending order, minimum-centered, and submitted to calculation in order to yield a Gini coefficient (c). The vector of activations displayed here is purely for illustrative purposes and does not depict actual data.

NEURAL VARIABILITY AND EMOTION REGULATION

Figure 5. Using Least-Squares Single Trial Analyses to Estimate Temporal Variability.



Note. Least Squares Single (LSS) analyses were used to estimate trial-specific activation for all ‘Far’ trials. In LSS, each trial is estimated in its own GLM where it is the target trial (a). As shown in the schematic, the target trial is given its own regressor (blue arrow), all other trials in that condition are dumped into a nuisance regressor (yellow arrow), and regressors from other conditions of interest are modeled as they normally would otherwise (green arrow). This process is repeated so that one has as many GLMs as target trials. Activations from the resulting beta-series were extracted from our spheres of interest to yield a $n \times p$ matrix, where each entry represents a measure of brain activity for the p -th sphere during the n -th trial (b). Individual subject estimates of temporal variability for each sphere were calculated by taking the standard deviation of each sphere’s respective beta-series. The design matrices and beta-series matrix displayed here are purely for illustrative purposes and do not depict actual data.

NEURAL VARIABILITY AND EMOTION REGULATION

Table 1. Fixed Effects of Latent Gini Coefficients and Moderators from the Measurement Model

<i>ROI</i>	γ	<i>SE</i>	<i>p</i>	<i>SD(r_i)</i>
L dlPFC	0.278	0.002	-	0.007
Age	0.000	0.001	0.617	
Neg. Affect	-0.009	0.005	0.112	
R dlPFC	0.280	0.003	-	0.006
Age	0.001	0.001	0.512	
Neg. Affect	-0.002	0.006	0.782	
R vIPFC	0.277	0.004	-	0.011
Age	0.001	0.001	0.640	
Neg. Affect	-0.013	0.007	0.049*	
R dmPFC	0.284	0.003	-	0.007
Age	0.000	0.001	0.866	
Neg. Affect	0.002	0.004	0.673	
L MTG	0.274	0.005	-	0.015
Age	-0.001	0.002	0.689	
Neg. Affect	0.001	0.010	0.907	
L SPL	0.280	0.003	-	0.011
Age	0.002	0.001	0.051 [□]	
Neg. Affect	-0.002	0.006	0.705	
R SPL	0.279	0.003	-	0.007
Age	-0.002	0.001	0.212	
Neg. Affect	-0.001	0.006	0.884	

Note. Age and Neg. Affect refer to each participant's age and average rating of negative affect during the task (both mean centered). The γ 's next to each ROI label represent fixed effects of latent Gini coefficients for those regions. The γ 's below them represent the respective associations between age and experiences of negative affect and each participant's specific latent Gini coefficient for each ROI. *SD(r_i)* represents the standard deviation of between person residuals in predicting individual specific latent Gini coefficients from fixed effects of latent Gini coefficients, age, and average negative affect. * $p < .05$, [□] $p < .10$.

NEURAL VARIABILITY AND EMOTION REGULATION

Table 2. Fixed Effects of Latent Temporal Variability and Moderators from the Measurement Model

<i>ROI</i>	γ	<i>SE</i>	<i>p</i>	<i>SD(r_i)</i>
L dlPFC	0.435	0.010	-	0.076
Age	-0.002	0.003	0.452	
Neg. Affect	0.031	0.019	0.107	
R dlPFC	0.567	0.015	-	0.110
Age	-0.006	0.006	0.280	
Neg. Affect	-0.010	0.026	0.690	
R vlPFC	0.456	0.013	-	0.088
Age	-0.009	0.005	0.092 [□]	
Neg. Affect	0.041	0.029	0.153	
R dmPFC	0.529	0.011	-	0.080
Age	-0.010	0.004	0.005*	
Neg. Affect	0.032	0.023	0.160	
L MTG	0.439	0.011	-	0.068
Age	-0.007	0.004	0.079 [□]	
Neg. Affect	0.034	0.022	0.132	
L SPL	0.542	0.011	-	0.081
Age	0.001	0.004	0.801	
Neg. Affect	0.028	0.024	0.243	
R SPL	0.534	0.013	-	0.096
Age	-0.006	0.005	0.243	
Neg. Affect	0.015	0.031	0.614	

Note. Age and Neg. Affect respectively refer to each participant's age and average rating of negative affect during the task (both mean centered). The γ 's next to each ROI label represent fixed effects of latent temporal variability for those regions. The γ 's below them represent the respective associations between age and experiences of negative affect and each participant's specific latent temporal variability for each ROI. *SD(r_i)* represents the standard deviation of between person residuals in predicting individual specific latent temporal variability from fixed effects of latent temporal variability, age, and average negative affect. Robust standard errors are reported. * $p < .05$, [□] $p < .10$.