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Integrative genomic analysis in African American children with asthma finds three novel loci associated with lung function

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Abstract

Bronchodilator (BD) drugs are commonly prescribed for treatment and management of obstructive lung function present with diseases such as asthma. Administration of BD medication can partially or fully restore lung function as measured by pulmonary function tests. The genetics of baseline lung function measures taken before BD medication have been extensively studied, and the genetics of the BD response itself have received some attention. However, few studies have focused on the genetics of post-BD lung function. To address this gap, we analyzed lung function

CONFLICT OF INTERESTS

The authors declare that there are no conflict of interests.

DATA AVAILABILITY STATEMENT

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AUTHOR CONTRIBUTIONS

P. C. G., K. L. K., and M. J. W. designed the study. S. S. O., S. S., C. E., and E. G. B. recruited study subjects and generated the data. A. C. Y. M., J. R. E., D. U., and S. H. cleaned and organized the data and provided analytic support. P. C. G., K. L. K., E. Y. L., O. R., M. G. C., and M. J. W. performed the analysis. A. K. L. and L. S. B. provided clinical pharmacological expertise for interpretation of results. E. G. B. and B. E. H. funded the study. E. G. B. supervised all recruitment. All authors contributed to manuscript writing and editing.

The data that support the findings of this study are available in the NCBI dbGaP repository under accession number phs000921.v1.p1. SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

phenotypes in 1103 subjects from the Study of African Americans, Asthma, Genes, and Environment, a pediatric asthma case–control cohort, using an integrative genomic analysis approach that combined genotype, locus-specific genetic ancestry, and functional annotation information. We integrated genome-wide association study (GWAS) results with an admixture mapping scan of three pulmonary function tests (forced expiratory volume in 1 s [FEV₁], forced vital capacity [FVC], and FEV₁/FVC) taken before and after albuterol BD administration on the same subjects, yielding six traits. We identified 18 GWAS loci, and five additional loci from admixture mapping, spanning several known and novel lung function candidate genes. Most loci identified via admixture mapping exhibited wide variation in minor allele frequency across genotyped global populations. Functional fine-mapping revealed an enrichment of epigenetic annotations from peripheral blood mononuclear cells, fetal lung tissue, and lung fibroblasts. Our results point to three novel potential genetic drivers of pre- and post-BD lung function: *ADAMTS1, RAD54B*, and *EGLN3*.

Keywords

admixture; African American; asthma; GWAS; integrative genomic analysis; lung function

1 | INTRODUCTION

Asthma is a disease characterized by episodic obstruction of airways that affects nearly 339 million people worldwide (The Global Asthma Network, 2018) and is the most common chronic disease among children. Asthma constitutes a massive global economic burden, representing \$81.9 billion in medical costs in the United States alone (Nurmagambetov et al., 2018). As a complex disease, asthma results from both environmental and genetic factors, with genetic heritability estimates ranging from 0.35 to 0.90 (Ober & Yao, 2011). The advent of genome-wide association studies (GWAS; Risch & Merikangas, 1996), combined with progressively larger sample sizes in recent years, has enabled researchers to query the genetic basis of asthma at unprecedented scale, with numerous loci identified in autoimmune and inflammatory pathways (Demenais et al., 2018). However, these loci account for a small portion of asthma liability (Demenais et al., 2018).

Pulmonary function tests are recommended to guide the diagnosis of asthma and monitor patient status (Asthma and Allergy Foundation of America, 2019). During these tests, patients breathe through a spirometer that captures key measures of lung function, including the forced expiratory volume in 1 s (FEV₁), which measures initial forced exhalatory capacity; the forced vital capacity (FVC), which measures the maximum total volume of air that a patient can forcibly exhale; and their ratio (FEV₁/FVC). Lung function measures can be population-normalized according to expected lung function values that account for age, sex, height, and ethnicity of the patient (Hankinson et al., 1999). Spirometric measurements can be taken both before bronchodilator (BD) treatment (pre-BD) and after (post-BD) to further understand lung function status. Historically, baseline lung function is measured with pre-BD measures, but among people with asthma, post-BD lung function may best reflect lung health (Brehm et al., 2015).

While the genetic contribution to asthma and lung function has been extensively studied via GWAS, most analyses have relied on subjects of European descent (Demenais et al., 2018; Johansson et al., 2019; Pickrell et al., 2016; Z. Zhu et al., 2018). This overrepresentation of ethnically white subjects in biomedical research has impaired the generalizability of genetic studies of complex disease (Burchard, 2014; Bustamante et al., 2011; Popejoy & Fullerton, 2016). Ethnic differences in lung function, particularly between non-Hispanic African Americans and European Americans, have been reported for over 40 years (Binder et al., 1976; Glindmeyer et al., 1995; Hsi et al., 1983; Rossiter & Weill, 1974; Schwartz et al., 1988). Ethnic disparities in lung function were attributed to population differences in sitting height, as increased height leads to increased lung capacity. However, adjustment for sitting height only explains 42%-50% of ethnic differences in lung function between African Americans and European Americans (Harik-Khan et al., 2004), suggesting that a simplistic reduction to ethnic differences in height cannot account for the observed disparity in lung function. Unequal socioeconomic conditions were also thought to contribute to ethnic differences in lung function (Braun, 2015; Quanjer, 2013, 2015), but socioeconomic factors only account for 7%-10% of unexplained variance (Harik-Khan et al., 2004). Self-identified race or ethnicity are commonly used in the clinic to interpret lung function measures, but these are not ideal variables for understanding genetic differences in lung function between populations. Kumar et al. (2010) observed that the proportion of global African genetic ancestry is inversely correlated with lung function. Spear et al. (2019) later observed population differences among African Americans, Mexican Americans, and Puerto Ricans in BD drug response to albuterol, the short-acting β_2 -adrenergic receptor agonist that is the most commonly prescribed drug for the treatment of acute asthma symptoms. Specifically, Spear et al. performed admixture mapping, a technique designed to identify regions of the genome where locus-specific ancestry drives variation in a disease trait (Shriner, 2013) that has been helpful in studies of complex diseases, including asthma and breast cancer (Féjerman et al., 2012; Pino-Yanes et al., 2015). However, admixture mapping studies comparing baseline and post-BD lung function have not yet been performed in African Americans. In this study, we address this gap in knowledge by evaluating the effect of locusspecific ancestry on both pre- and post-BD lung function measures in a pediatric casecontrol cohort of non-Hispanic African Americans children and adolescents.

2 | METHODS

2.1 | Ethics statement and data availability

Data from the Study of African Americans, Asthma, Genes, and Environments (SAGE) cohort were used for this study. The data that support the findings of this study are available in the NCBI dbGaP repository under ascension number phs000921.v1.p1. Data from SAGE were approved for human subjects research under expedited review by IRB 10–02877 at the University of California, San Francisco with reference #244919. All subjects gave written consent for genotyping, phenotyping, and data usage for general research use.

2.2 | Study population

SAGE is a case–control cross-sectional cohort study of genetics and gene–environment interactions in non-Hispanic African American children and adolescents in the United

States. SAGE includes detailed clinical, social, and environmental data on both asthma and asthma-related conditions. Full details of the SAGE study protocols are described in detail elsewhere (Borrell et al., 2013; Mak et al., 2018; Nishimura et al., 2013; Thakur et al., 2013). Briefly, SAGE was initiated in 2006 and recruited participants with and without asthma through a combination of clinic- and community-based recruitment centers in the San Francisco Bay Area. All participants in SAGE self-identified as African American and self-reported that all four grandparents were non-Hispanic African American. To reduce population heterogeneity resulting from very recent admixture, we only analyzed subjects who affirmatively self-identified as having been born in the United States and whose parents were also born in the United States.

Pulmonary function tests were taken before administration of albuterol BD medication for all individuals, both those with and without asthma. Post-BD spirometry measures were performed only for individuals with asthma. Analyses of pre-BD lung function measures included all 1103 asthma cases and controls with complete covariate information. Post-BD analyses were performed on the 831 asthma cases with post-BD measurements.

2.3 | Genotyping and quality control

DNA was isolated from whole blood collected from SAGE participants at the time of study enrollment as described previously (Borrell et al., 2013). DNA was extracted using the Wizard® Genomic DNA Purification kits (Promega). Samples were genotyped with the Affymetrix Axiom LAT1 array (World Array 4, Affymetrix).

Genotype quality control was performed in PLINK v1.9 (Chang et al., 2015). Of the 772,703 genotyped variants, 111,901 single nucleotide polymorphisms (SNPs) were excluded from analysis due to genotype missingness more than 5% (n = 28,211), minor allele frequency (MAF) less than 1% (n = 80,420) or deviation from Hardy-Weinberg expectations (HWE) at p < 0.001 (n = 3270). The final set included 660,802 genotyped markers (Table S1).

Genotyped SNPs were submitted to the Michigan Imputation Server (Das et al., 2016), phased using EAGLE v2.3 (Loh et al., 2016), and imputed from the 1000 Genomes Project reference panel (The 1000 Genomes Project Consortium, 2015) using Minimac3 (Das et al., 2016). Imputed SNPs with imputation $R^2 < .3$, with deviation from HWE $p < 10^{-4}$, or with MAF < 1% were discarded. Of the 47,101,126 imputed SNPs, a total of 31,146,322 were culled due to either low MAF (n = 31,095,418) or deviation from HWE (n = 50,904). All variants in the imputed set showed a genotype missingness of no more than 5%. The final number of SNPs used in association analyses was 15,954,804 (Table S1).

2.4 | Outcome phenotypes

Pulmonary function testing was performed at the time of recruitment according to the American Thoracic Society/European Respiratory Society standards (Miller et al., 2005; Pellegrino et al., 2005; Wanger et al., 2005) with a KoKo PFT Spirometer (nSpire Health Inc.). Spirometry was performed both before and 15 min after administration of four puffs of albuterol (90 μ g per puff) through a 5-cm plastic mouthpiece from a standard metered-dose inhaler. Patients were assessed for the following spirometric measures before and after BD drug usage (pre-BD and post-BD, respectively): (a) FEV₁, (b) FVC, and (c) FEV₁/FVC. A

total of six phenotypes were assessed for genotype association: pre-BD FEV₁ (pre-FEV₁), pre-BD FVC (pre-FVC), pre-BD FEV₁/FVC (pre- FEV₁/FVC), post-BD FEV₁ (post-FEV₁), post-BD FVC (post-FVC), and post-BD FEV₁/FVC (post-FEV₁/FVC). All phenotype values were normalized based on the expected lung function values calculated from the Hankinson equations (Hankinson et al., 1999), which account for age, sex, height, and self-reported ethnicity. Phenotype distributions were checked for normality and to detect outliers. Outliers were determined using the method of Tukey fences (John Tukey, 1977). For each phenotype, we computed the first quartile value (Q1), the third quartile value (Q3), and the interquartile range (IQR). We declared as outliers all values outside of the range

[Q1 - 3(IQR), Q3 + 3(IQR)].

Individuals with outlier values for a phenotype were removed from association analyses for that phenotype.

2.5 | Covariates

2.5.1 | Age, sex, and body mass index (BMI)—Biometric covariates such as age, sex, BMI, and height were measured directly at time of recruitment. BMI was categorized into underweight, normal, overweight, and obese, according to CDC guidelines for defining childhood obesity (Barlow, 2007; Cote et al., 2013; Whitlock et al., 2005). An overweight status was defined as a BMI at or above the 85th percentile for the general population of children of the same sex and in the same age group. An obese status was defined as a BMI at or above the 95th percentile. Underweight individuals (bottom 5th percentile, n = 9) were excluded from analysis.

2.5.2 | Asthma status—Case status was defined as physician-diagnosed asthma supported by reported asthma medication use and symptoms of coughing, wheezing, or shortness of breath in the 2 years preceding enrollment.

2.5.3 | Maternal educational attainment—Maternal educational attainment was measured at recruitment and included in analyses to control for socioeconomic status. It was coded as total years of education completed from the first grade: for example, a complete K-6 education was 6 years, a complete high school education was 12 years, and any additional years (college or trade school and beyond) were counted as 1 year each.

2.5.4 Genetic ancestry—Previous literature on the genetics of non-Hispanic African Americans has observed global genetic ancestry proportions of 73.2% West African, 24.0% European, and 0.8% Amer-indigenous (Bryc et al., 2015), strongly suggesting that a reference panel of African and European was sufficient for accurate global genetic ancestry estimates in SAGE. Global genetic ancestry was estimated for each individual with the ADMIXTURE software (Alexander et al., 2009) in supervised learning mode assuming one West African and one European ancestral population, with HapMap Phase III YRI and CEU populations as references (The International HapMap 3 Consortium, 2010). Local ancestry estimation was performed with RFMix (Maples et al., 2013; Spear et al., 2019) using the same two-way ancestry reference from HapMap Phase III.

2.5.5 | Estimation of genetic relatedness and genotype principal components

--Genetic relatedness matrices (GRMs) were generated in R using GENESIS (Gogarten et al., 2019), which provides a computational pipeline for handling complex population structure using principal components analysis (PCA). We used PCAir (Conomos et al., 2015) to correct for distant population structure accounting for relatedness, and PC-Relate (Conomos et al., 2016) to adjust for genetic relatedness in recently admixed populations. The resulting principal components provide better correction for population stratification in admixed populations compared to standard PCA on genotypes (Patterson et al., 2006). Additionally, since GRMs produced by GENESIS are corrected for cryptic relatedness, the resulting association test statistics do not suffer inflation resulting from confounding relatedness in our sample.

2.6 | Genetic association analyses

Genotype association testing was performed with the MLMA-LOCO algorithm from GCTA (Yang et al., 2011, 2014) to correct for population structure using GRMs generated with GENESIS. Association testing of outcome phenotypes with allele dosages at 15,954,804 biallelic SNPs was performed with a "leave one chromosome out" model to avoid double-fitting tested variants. Other variables included in models were age, sex, BMI, maternal educational attainment, and three genotype principal components. Models of pre-BD also included asthma status.

The suggestive and significant association thresholds for each outcome phenotype were determined by the effective number of independent statistical tests (M_{eff}) calculated with CODA (Plummer et al., 2006). CODA computes M_{eff} using the autocorrelation of p values from GWAS. This produces population-specific Bonferroni thresholds that account for correlation between statistical tests without increasing the Type I error rate (Sobota et al., 2015). For our analyses, M_{eff} ranged from 488,819 to 507,975 (Table S2). The Bonferroni corrected genome-wide significance threshold was computed as $0.05/M_{eff}$, while the suggestive threshold was computed as $(1/M_{eff})$, yielding a single pair of thresholds for all six outcome phenotypes considered: $p < 1.99 \times 10^{-6}$ for suggestive association, and $p < 9.95 \times 10^{-8}$ for significant association (Table S2).

Admixture mapping analyses were performed using linear regression models in R and local ancestry calls from RFMix for 454,322 genotyped SNPs. Counts of 0, 1, or 2 alleles of African descent were computed for each person at each SNP. Phenotypes were then regressed onto ancestral allele counts for each SNP while including age, sex, height, BMI, maternal educational attainment (as a proxy for socioeconomic status), and global African genetic ancestry proportion as covariates. Analyses with pre-BD outcome measures also included asthma status as a covariate.

2.7 | Fine-mapping genetic associations

Functional fine-mapping with PAINTOR (Kichaev et al., 2014) was used to identify putative causal variants in novel loci deemed statistically significant by admixture mapping. PAINTOR applies a Bayesian probabilistic framework to integrate functional annotations, association summary statistics (*Z*-scores), and linkage disequilibrium information for each

locus to prioritize the most likely causal variants in a given region. Functional annotations were selected per locus as recommended by the authors of PAINTOR (Kichaev, 2017). A subset of lung- and blood-related functional annotations from the Roadmap Epigenomics Project (Roadmap Epigenomics Consortium et al., 2015) and the ENCODE Consortium (ENCODE Project Consortium, 2012) were assessed for their individual improvement to the posterior probability of causality; the top five minimally correlated annotations were selected for each locus.

2.8 | Annotation tools

The NHGRI/EBI GWAS Catalog (Buniello et al., 2019), Ensembl Genome Browser release 98 (Cunningham et al., 2019) and gnomAD browser v3.0 (Karczewski et al., 2020) were used to look up known associations at significant loci according to our analyses. Annotation lookups in the gnomAD browser v3.0 used hg38 coordinates translated from our hg19-aligned genotypes via liftOver (Hinrichs et al., 2006). Data management, statistical analysis, and figure generation made extensive use of GNU parallel (Tange, 2018) and several R packages, including data.table, doParallel, optparse, ggplot2, and the tidyverse bundle (Davis, 2020; Dowle & Srinivasan, 2020; Microsoft Corporation & Weston, 2019; Wickham, 2016; Wickham & Garrett, 2017).

3 | RESULTS

3.1 | Cohort characteristics

Characteristics of all SAGE participants included in analyses are shown in Table 1. Distributions of each lung function measure stratified by case–control status and BD administration (pre-BD vs. post-BD) are shown in Figure S1. FVC showed no significant difference between asthma cases and controls (Kruskal-Wallis p = .073), while stratification by case–control status yielded significantly different distributions for FEV₁ (Kruskal-Wallis $p = 4.8 \times 10^{-7}$) and FEV₁/FVC (Kruskal-Wallis $p = 1.5 \times 10^{-7}$). Among cases, statistically significant differences were observed between distributions of pre-BD and post-BD measures of FEV₁ (Kruskal-Wallis $p = 1.2 \times 10^{-38}$), FVC (Kruskal-Wallis $p = 5.4 \times 10^{-16}$), and FEV₁/FVC (Kruskal-Wallis $p = 4.0 \times 10^{-29}$), illustrating a measurable effect of BD medication on lung function.

The global African genetic ancestry proportion in our sample varied from 30.7% to 100%, with an average proportion of 80.2% (Figure S2), concordant with empirically observed averages (Baharian et al., 2016). Global ancestry contained the same information as the first genotype principal component ($R^2 = 0.984$, Figure S3).

3.2 | Genetic association testing finds novel and known loci

Figure 1 shows results from GWAS performed on pre-BD phenotypes (pre-FEV₁, pre-FVC, and pre-FEV₁/FVC) and post-BD phenotypes (post-FEV₁, post-FVC, and post-FEV₁/FVC) using linear mixed modeling. The association results showed no evidence of genomic inflation, with genetic control λ ranging from 0.98 to 0.99 (Table S2 and Figure S4). Table 2 lists the 18 genome-wide significant associations found, each associated with exactly one of the six lung function measures. An additional 252 variants were suggestively associated with

at least one phenotype (Tables S3–S8). Of the 18 variants, 4 variants on chromosome 13 in a region spanned by the gene *ATP8A2* were associated with pre-FEV₁/FVC (Figure S6). Two variants on chromosome 16 that were associated with pre-FVC flanked the promoter region of *IRX3* (Figure S7). A third variant associated with pre-FVC was located on chromosome 20 near *THBD* (Figure S8), a gene linked to venous thromboembolism in African American and Afro-Caribbean individuals (Hernandez et al., 2016). Two variants associated with Post-FVC were in a gene-rich region on chromosome 19 (Figure S9), with the peak near *TMIGD2* and *SHD*, while eight other variants pointed to a second gene-rich region on chromosome 11 near *CXCR5* and *HYOU1* (Figure S10). Post-FEV₁/FVC was associated with a region on chromosome 15 near the genes *AKAP13* and *ADAMTS7P4* (Figure S11).

Among the suggestive associations, a variant on chromosome 12 associated with post-FEV1 was near *BTBD11* (Figure S12), a gene previously associated with post-FEV₁, post-FEV₁/ FVC, and FEV₁, the change in lung function due to BD administration (Hardin et al., 2016; Lutz et al., 2015), as well as BMI (Kichaev et al., 2019). A suggestive association with pre-FEV₁ on chromosome 12 fell near *SCARB1* (Figure S13), which was previously associated with FEV₁ and FVC (Wyss et al., 2018) and HDL cholesterol levels (Wojcik et al., 2019). Another suggestive association with pre-FEV₁ on chromosome 20 was near the gene *PTPRT* (Figure S14), which was previously associated with thromboembolism susceptibility in 5334 African American individuals (Heit et al., 2017).

3.3 | Admixture mapping identified five novel loci not found by GWAS

Table 3 shows five regions where admixture proportions were statistically significantly associated with one of the six phenotypes. The three pre-BD phenotypes (pre-FEV₁, pre-FVC, pre-FEV₁/FVC) were each associated with one region, while post-FVC was associated with two distinct regions. Post-FEV₁ and post-FEV₁/FVC had no significant associations. None of the regions overlapped with those significant in our GWAS, and none showed large deviations from mean genome-wide African genetic ancestry. A small region on chromosome 21 that was significantly associated with pre-FEV₁ flanked the genes *ADAMTS1* and *ADAMTS5* (Figure S15). The region on chromosome 4 associated with pre-FVC pointed to two candidate genes, *RCHY1* and *THAP6*, that had no prior lung disease associations (Figure S17). A region on chromosome 19 associated with pre-FEV₁/FVC spanned the genes *ZNF557* and *INSR* (Figure S18). Post-FVC was associated with two regions, one on chromosome 8 spanning the genes *ESRP1*, *INTS8*, *TP53INP1*, and *NDUFAF6* (Figure S19), and another on chromosome 14 encompassing *EGLN3* and *SNX6* (Figure S20).

3.4 | Functional fine-mapping found three novel putatively causal loci for lung function phenotypes

Table 4 lists the most probable causal SNP for each of the five admixture mapping loci according to PAINTOR. SNP rs13615 showed the highest probability of causality (0.630) with pre-FEV₁ on locus 1 (Figure 2). This variant falls within the 3'-untranslated region (3'-UTR) of *ADAMTS1*, suggesting that *ADAMTS1* drives the admixture mapping association and not its physical neighbor *ADAMTS5*. The MAF of rs13615 in African and African diaspora populations was lower than every other global population (2.6% AFR vs. 7.0%–

54.5% other populations, gnomAD v3; see Figure S21). The SNP rs10857225 emerged as the most likely causal variant (probability 0.361) for the association of pre-FVC with locus 2 on chromosome 4 (Figure 3). This variant is located within an intron of the gene THAP6, suggesting that THAP6 is more likely the causal gene behind the association with pre-FVC. In contrast to locus 1, the MAF of rs10857225 is highest in global African populations and markedly lower in other global populations (59.1% AFR vs. 28.1%-38.4% other populations, gnomAD v3). Locus 3 on chromosome 19 associated with pre-FEV₁/FVC, and locus 4 on chromosome 8 associated with post-FVC, showed little information gain from functional fine-mapping. The driving variant for locus 3, SNP rs72986681, was located in the 3'-UTR of ZNG557, but showed a low probability of causality (0.168, Figure S22). The most probable marker for locus 4, the SNP rs2470740, which is located in intron 2 of RAD54B, showed an even lower probability of causality (0.109, Figure S23). Functional fine-mapping of locus 5, a region on chromosome 14 associated with post-FVC, yielded the SNP rs1351618 with a moderate probability of causality (.390, Figure 4). rs1351618 is located in an intron of EGLN3. As with locus 2, rs1351618 had a much higher MAF in populations of African ancestry versus other global populations (12.4% AFR vs. <2.2% other populations, gnomAD v3).

4 | DISCUSSION

We analyzed the genetic basis of six lung function phenotypes in 1103 non-Hispanic African American children with and without asthma. The phenotypes consisted of three standard spirometric measures—FEV₁, FVC, and FEV₁/FVC—measured before and after administration of BD medication. Our GWAS identified 18 genome-wide significant loci, while our integrative genetic analysis approach that layered GWAS, admixture mapping, and functional fine-mapping identified another five putatively causal loci that could drive differences between pre- and post-BD lung function.

The four variants on chromosome 13 associated with pre-FEV₁/FVC pointed to ATP8A2 as a candidate gene. ATP8A2 encodes an ATPase involved in phospholipid transport and is highly expressed in brain tissue, testes, and the adrenal glands, and to a lesser degree in the lung (Fagerberg et al., 2014). Mutations in ATP8A2 have been linked to several neurological disorders (Martín-Hernández et al., 2016). Two variants on chromosome 16 that were associated with pre-FVC point to IRX3 as a candidate gene. IRX3 encodes a homeobox protein crucial for neural development, and its promoters previously showed a long-range interaction with the FTO gene. Expression levels of the FTO gene are known to influence BMI and are of great interest in type II diabetes and obesity research (Smemo et al., 2014). Post-FVC showed associations on two chromosomes. Notable genes near the association peak on chromosome 19 included MAP2K2 and ZBTB7A, genes associated with variation in the two genes closest to the association peak, TMIGD2 and SHD, have not been previously associated with any traits. TMIGD2 is involved in T-cell costimulation and the immune response through an interaction with BMI, visceral adiposity, and eosinophil counts (Kichaev et al., 2019; Pulit et al., 2019; Rüeger et al., 2018); and CHAF1A, HDGFL2, PLIN4, ANKRD24, MPND, and SH3GL1, previously associated with corpuscular volume and hemoglobin concentration (Astle et al., 2016; Kichaev et al., 2019; van Rooij et al., 2017). Interestingly, HHLA2, suggesting that it could possibly play an immune or allergic

response role in lung function (Y. Zhu et al., 2013). Among the genes within or near the chromosome 11 peak, two emerge as potentially key loci. The first is CXCR5, which has been linked to increased risk of childhood onset asthma (Ferreira et al., 2019; Johansson et al., 2019; Pividori et al., 2019) and respiratory disease (Kichaev et al., 2019), as well as related allergic and immunological conditions such as eczema, leukocyte count, rheumatoid arthritis, and Sjögren's syndrome (Ferreira et al., 2017; Kichaev et al., 2019; Laufer et al., 2019; Lessard et al., 2013). The second is HYOU1, which has been associated with BMI and post-FEV₁/FVC (Lutz et al., 2015; Pulit et al., 2019). Post-FEV₁/FVC was associated with two genes, AKAP13 and ADAMTS7P4. AKAP13 has been previously associated with numerous conditions, including interstitial lung disease and psoriasis in European individuals (Fingerlin et al., 2013; Tsoi et al., 2015) as well as weight, BMI, and cardiovascular traits such as blood pressure and hemoglobin count in multiple populations (Giri et al., 2019; Kichaev et al., 2019). ADAMTS7P4 was previously associated with red blood cell volume (Kichaev et al., 2019). The statistically suggestive association of post-FEV₁ with *BTBD11* pointed to previous associations with various lung function measures, including post-FEV₁, post-FEV₁/FVC, and FEV₁ (Hardin et al., 2016; Lutz et al., 2015). These previously detected associations were based on much larger sample sizes than what was available to us: the associations with post-FEV₁ and post-FEV₁/FVC found by Lutz et al. were discovered in a population of 10,094 European and 3260 African American smokers with chronic obstructive pulmonary disorder (COPD), while the association with FEV₁ found by Hardin et al. was based on 5766 Europeans and 811 African Americans with COPD, suggesting that our inability to reach genome-wide significance in our sample was due to insufficient statistical power.

Among significant and suggestive GWAS loci, the association of post-FVC with variants in or near *CXCR5* and *HYOU1* is the only one that replicates known lung function loci: *CXCR5* was previously associated with asthma (Ferreira et al., 2019; Johansson et al., 2019; Pividori et al., 2019), and *HYOU1* was previously associated with post-FEV₁/FVC (Lutz et al., 2015). The association with post-FEV₁/FVC comes from an adult COPD cohort ascertained by smoking status; in contrast, SAGE is a pediatric asthma cohort. The mechanism by which *HYOU1* affects lung function in both youth and adults is unclear. Nevertheless, the overlap of post-BD pulmonary function measures at this locus suggests that the region encompassing *CXCR5* and *HYOU1* plays a role in lung disease among people with obstructive lung function.

Admixture mapping identified five genomic regions where variation in genetic ancestry was significantly associated with phenotypic variation. Locus 1 on chromosome 21 spanned the genes *ADAMTS1* and *ADAMTS5*, which encode extracellular proteases within the same protein family but with different consequences for disease. Although both genes have been linked to blood protein levels (Suhre et al., 2017), *ADAMTS1* has been associated with pre-FVC (Kichaev et al., 2019) and is expressed in arterial, adipose, and lung tissue, while *ADAMTS5* is not appreciably expressed in the lung (Figure S16). Further fine-mapping with PAINTOR places the most likely causal SNP (rs13615) within the 3'-UTR of *ADAMTS1*. Although the region is sparsely genotyped, and follow-up with whole genome sequencing data in this region is recommended, these results suggest that *ADAMTS1* may be functionally related to lung function. Interestingly, the association of *ADAMTS1* with pre-

FVC (Kichaev et al., 2019) was discovered in a European sample of substantially larger size than our cohort, highlighting the ability of admixture mapping to detect associations in scenarios with low statistical power. Locus 3, spanning a region on chromosome 19 that was associated with pre-FEV₁/FVC, contains the genes *ZNF557* and *INSR. ZNF557* has not been previously associated with any traits, while *INSR* is the well-known insulin receptor that has been previously associated with childhood onset asthma in our own cohort (White et al., 2016), as well as blood pressure levels, triglyceride levels, HDL cholesterol levels, and hypothyroidism across multiple populations (Bentley et al., 2019; Ehret et al., 2016; Kichaev et al., 2019; Klarin et al., 2018). Post-FVC showed two distinct admixture mapping signals. The first region on chromosome 8, which we call Locus 4, includes the genes *ESRP1*, *INTS8*, *TP53INP1* and *NDUFAF6* was previously associated with type II diabetes and eosinophil counts (Kichaev et al., 2019; Mahajan et al., 2018). The second region, Locus 5, spans *EGLN3* and *SNX6*, both of which show previous associations with blood phenotypes such as blood pressure and hematocrit levels (Astle et al., 2016; Evangelou et al., 2018).

Overall, evaluation of our GWAS and admixture mapping lung function results suggests that genetics of this trait underlie some pleiotropy observed across pulmonary, hematological, cardiovascular, and obesity-related traits. Such pleiotropy has been observed in UK BioBank participants: as lung function decreases, BMI and type II diabetes incidence increases, as well as levels of eosinophils and neutrophils, both of which are common biomarkers for allergic disease (Figure S25; McInnes et al., 2019). The link between obesity and lung function is particularly interesting since obesity is a known asthma comorbidity, and lung function may play a role in obese asthma (Baffi et al., 2015; Gruchała-Niedoszytko et al., 2015). Our findings suggest that genetically based differences in lung function may provide a link between obesity and asthma.

It is curious that the regions identified by admixture mapping and subjected to functional fine-mapping did not overlap with the statistically significant GWAS loci. We attribute this in part to the different types of information used by each approach: GWAS analyzes how allelic variation affects a trait, while admixture mapping analyzes the phenotypic consequences of variation in genetic ancestry. Our integrative approach deprioritized results solely from GWAS, an approach driven by the fact that a supermajority of individual GWAS results fall in noncoding regions of the genome and are consequently notoriously difficult to interpret. By integrating GWAS summary statistics with loci identified via admixture mapping, we found that three of the admixture mapping-based loci-ADAMTS1, THAP6, and EGLN3—had evidence of causal effects. Each of the sentinel SNPs tagging these genes showed a notable difference in ancestral allele frequency: populations of African descent had either the highest or the lowest MAF among all global populations, likely the result of admixture mapping prioritizing loci that varied by genetic ancestry. None of these loci have been previously associated with lung traits, highlighting the strength of our integrative analysis. The association with *EGLN3* is particularly curious since it has been previously associated with a variety of traits, including heart rate response to β -blocker therapy (Shahin et al., 2018). Short-acting β -2 agonists such as albuterol selectively target β -2 receptors in the lungs, while the first-generation β -blockers taken for cardiac conditions bind to both β -1 and β -2 receptors, affecting the heart as well the lungs. Bronchospasm and FEV₁ reduction are clinically significant side effects of first-generation β -1 selective and nonselective β -

blockers for cardiac conditions. Consequently, these nonselective β-blockers must be initiated with caution and close monitoring in patients with asthma (Christiansen & Zuraw, 2019). β-blockers lower blood pressure by reducing heart rate and cardiac contractility and are less effective in people with high levels of African genetic ancestry (Brewster & Seedat, 2013; Whelton et al., 2018). It has been previously observed that African Americans with asthma demonstrate lower BD drug response than European Americans (Blake et al., 2008), suggesting a possible pharmacological interaction between β–2 receptors and African ancestry. Furthermore, *EGLN3* is strongly expressed in cardiac tissue, suggesting that *EGLN3* could possibly influence post-FVC through cardiac phenotypes (Figure S24). Further functional studies are required to elucidate the role of *EGLN3* on lung function and BD drug response.

This study has some important limitations. First, while our data set includes rich phenotyping of pulmonary traits with socioeconomic and biometric measures, it still constitutes a somewhat small sample by modern measures. The tradeoff between rich phenotyping and increased sample size is not trivial, particularly for studies of populations traditionally underrepresented in genetic research, a challenge that still plagues large studies like NHLBI TOPMed and NIH Million Veterans Program. Our approach of layering multiple types of genomic information serves as a partial workaround. However, by layering GWAS with admixture mapping and functional fine-mapping, we restricted our focus to regions where variants showed differential ancestry, which excluded our strongest GWAS hits. While PAINTOR can provide evidence of putatively causal markers that do not necessarily meet strict genome-wide thresholds of significance, it was not designed with admixed populations in mind. We applied suitably rigorous statistical stringency to both our GWAS and our admixture mapping results by thresholding to the effective number of independent tests as estimated by the CODA software, an approach designed to produce population-specific significance thresholds (Sobota et al., 2015), so we are confident that these regions are indeed correlated to the phenotype. Nevertheless, further research is needed to understand how the hierarchical Bayesian model in the PAINTOR inference engine behaves in regions of heterogeneous ancestry. Finally, if the truly causal variant is not genotyped, then PAINTOR and similar fine-mapping frameworks suffer a performance hit that is difficult to rectify without ultra-fine genotyping in the locus of interest (e.g., from whole genome sequencing). We suspect that our relatively low probabilities of causality can be attributed to a confluence of differential ancestry, small sample size, and insufficiently resolved fine-mapping, which could be improved in future studies by using whole-genome sequencing data instead of imputed genotypes.

Our integrative analysis approach leverages available functional annotations and genetic ancestry estimates in the absence of molecular data to yield some promise for discovery of novel loci. Our study is limited to three tiers—genotypes, genetic ancestry, and functional annotations—and makes use of gene expression results from GTEx v8. However, it does not directly incorporate any transcriptomic, metabolomic, proteomic, or methylomic information. As large multiomic data sets from NHLBI TOPMed, UK Biobank, and the NIH Million Veterans Program become available, the need for integrative genomic approaches to studying complex diseases will increase. Future multiomic models of complex diseases, including obstructive lung function disorders, may deliver on the promise of precision

medicine and provide actionable clinical translation of biomedical and pharmacogenomic insights into novel therapies.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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REFERENCES

- Alexander DH, Novembre J, & Lange K (2009). Fast model-based estimation of ancestry in unrelated individuals. Genome Research, 19(9), 1655–1664. 10.1101/gr.094052.109 [PubMed: 19648217]
- Asthma and Allergy Foundation of America. (2019). Asthma diagnosis. https://www.aafa.org/lung-function-tests-diagnose-asthma/
- Astle WJ, Elding H, Jiang T, Allen D, Ruklisa D, Mann AL, Mead D, Bouman H, Riveros-Mckay F, Kostadima MA, Lambourne JJ, Sivapalaratnam S, Downes K, Kundu K, Bomba L, Berentsen K, Bradley JR, Daugherty LC, Delaneau O, ... Soranzo N (2016). The allelic landscape of human

blood cell trait variation and links to common complex disease. Cell, 167(5), 1415–1429e19. 10.1016/j.cell.2016.10.042 [PubMed: 27863252]

- Baffi CW, Winnica DE, & Holguin F (2015). Asthma and obesity: Mechanisms and clinical implications. Asthma Research and Practice, 1, 1 10.1186/s40733-015-0001-7 [PubMed: 27965756]
- Baharian S, Barakatt M, Gignoux CR, Shringarpure S, Errington J, Blot WJ, Bustamante CD, Kenny EE, Williams SM, Aldrich MC, & Gravel S (2016). The great migration and African-American genomic diversity. PLOS Genetics, 12(5), e1006059 10.1371/journal.pgen.1006059 [PubMed: 27232753]
- Barlow SE (2007). Expert committee recommendations regarding the prevention, assessment, and treatment of child and adolescent overweight and obesity: Summary report. Pediatrics, 120(Suppl 4), S164–S192. 10.1542/peds.2007-2329C [PubMed: 18055651]
- Bentley AR, Sung YJ, Brown MR, Winkler TW, Kraja AT, Ntalla I, Schwander K, Chasman DI, Lim E, Deng X, Guo X, Liu J, Lu Y, Cheng C-Y, Sim X, Vojinovic D, Huffman JE, Musani SK, Li C, ... Cupples LA (2019). Multiancestry genome-wide gene-smoking interaction study of 387,272 individuals identifies new loci associated with serum lipids. Nature Genetics, 51(4), 636–648. 10.1038/s41588-019-0378-y [PubMed: 30926973]
- Binder RE, Mitchell CA, Schoenberg JB, & Bouhuys A (1976). Lung Function among Black and White children. American Review of Respiratory Disease, 114(5), 955–959. 10.1164/ arrd.1976.114.5.955
- Blake K, Madabushi R, Derendorf H, & Lima J (2008). Population pharmacodynamic model of bronchodilator response to inhaled albuterol in children and adults with asthma. Chest, 134(5), 981– 989. 10.1378/chest.07-2991 [PubMed: 18583517]
- Borrell LN, Nguyen EA, Roth LA, Oh SS, Tcheurekdjian H, Sen S, Davis A, Farber HJ, Avila PC, Brigino-Buenaventura E, LeNoir MA, Lurmann F, Meade K, Serebrisky D, Rodriguez-Cintron W, Kumar R, Rodriguez-Santana JR, Thyne SM, & Burchard EG (2013). Childhood obesity and asthma control in the GALA II and SAGE II studies. American Journal of Respiratory and Critical Care Medicine, 187(7), 697–702. 10.1164/rccm.201211-2116OC [PubMed: 23392439]
- Braun L (2015). Race, ethnicity and lung function: A brief history. Canadian Journal of Respiratory Therapy, 51(4), 99–101. [PubMed: 26566381]
- Brehm JM, Man Tse S, Croteau-Chonka DC, Forno E, Litonjua AA, Raby BA, Chen W, Yan Q, Boutaoui N, Acosta-Pérez E, Avila L, Weiss ST, Soto-Quiros M, Cloutier MM, Hu D, Pino-Yanes M, Wenzel SE, Spear ML, Kolls JK, ... Celedón JC (2015). A genome-wide association study of post-bronchodilator lung function in children with asthma. American Journal of Respiratory and Critical Care Medicine, 192(5), 634–637. 10.1164/rccm.201501-0047LE [PubMed: 26325155]
- Brewster LM, & Seedat YK (2013). Why do hypertensive patients of African ancestry respond better to calcium blockers and diuretics than to ACE inhibitors and β-adrenergic blockers? A systematic review. BMC Medicine, 11, 141 10.1186/1741-7015-11-141 [PubMed: 23721258]
- Bryc K, Durand EY, Macpherson JM, Reich D, & Mountain JL (2015). The genetic ancestry of African Americans, Latinos, and European Americans across the United States. American Journal of Human Genetics, 96(1), 37–53. 10.1016/j.ajhg.2014.11.010 [PubMed: 25529636]
- Buniello A, MacArthur JAL, Cerezo M, Harris LW, Hayhurst J, Malangone C, McMahon A, Morales J, Mountjoy E, Sollis E, Suveges D, Vrousgou O, Whetzel PL, Amode R, Guillen JA, Riat HS, Trevanion SJ, Hall P, Junkins H, ... Parkinson H (2019). The NHGRI-EBI GWAS catalog of published genome-wide association studies, targeted arrays and summary statistics 2019. Nucleic Acids Research, 47(D1), D1005–D1012. 10.1093/nar/gky1120 [PubMed: 30445434]
- Burchard EG (2014). Medical research: Missing patients. Nature News, 513(7518), 301–302. 10.1038/513301a
- Bustamante CD, Burchard EG, & De la Vega FM (2011). Genomics for the world. Nature, 475(7355), 163–165. 10.1038/475163a [PubMed: 21753830]
- Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, & Lee JJ (2015). Second-generation PLINK: Rising to the challenge of larger and richer datasets. GigaScience, 4(1), 7 10.1186/ s13742-015-0047-8 [PubMed: 25722852]
- Christiansen SC, & Zuraw BL (2019). Treatment of hypertension in patients with asthma. New England Journal of Medicine, 381(11), 1046–1057. 10.1056/NEJMra1800345

- Conomos MP, Miller MB, & Thornton TA (2015). Robust inference of population structure for ancestry prediction and correction of stratification in the presence of relatedness. Genetic Epidemiology, 39(4), 276–293. 10.1002/gepi.21896 [PubMed: 25810074]
- Conomos MP, Reiner AP, Weir BS, & Thornton TA (2016). Model-free estimation of recent genetic relatedness. The American Journal of Human Genetics, 98(1), 127–148. 10.1016/ j.ajhg.2015.11.022 [PubMed: 26748516]
- Cote AT, Harris KC, Panagiotopoulos C, Sandor GGS, & Devlin AM (2013). Childhood obesity and cardiovascular dysfunction. Journal of the American College of Cardiology, 62(15), 1309–1319. 10.1016/j.jacc.2013.07.042 [PubMed: 23954339]
- Cunningham F, Achuthan P, Akanni W, Allen J, Amode MR, Armean IM, Bennett R, Bhai J, Billis K, Boddu S, Cummins C, Davidson C, Dodiya KJ, Gall A, Girón CG, Gil L, Grego T, Haggerty L, Haskell E, ... Flicek P (2019). Ensembl 2019. Nucleic Acids Research, 47(D1), D745–D751. 10.1093/nar/gky1113 [PubMed: 30407521]
- Das S, Forer L, Schönherr S, Sidore C, Locke AE, Kwong A, Vrieze SI, Chew EY, Levy S, McGue M, Schlessinger D, Stambolian D, Loh P-R, Iacono WG, Swaroop A, Scott LJ, Cucca F, Kronenberg F, Boehnke M, ... Fuchsberger C, (2016). Next-generation genotype imputation service and methods. Nature Genetics, 48, 1284–1287. 10.1038/ng.3656 [PubMed: 27571263]
- Davis TL (2020). optparse: Command Line Option Parser. R package (Version 1.6.6) [Computer software]. https://CRAN.R-project.org/package=optparse
- Demenais F, Margaritte-Jeannin P, Barnes KC, Cookson WOC, Altmüller J, Ang W, Barr RG, Beaty TH, Becker AB, Beilby J, Bisgaard H, Bjornsdottir US, Bleecker E, Bønnelykke K, Boomsma DI, Bouzigon E, Brightling CE, Brossard M, Brusselle GG, ... Nicolae DL (2018). Multiancestry association study identifies new asthma risk loci that colocalize with immune-cell enhancer marks. Nature Genetics, 50(1), 42–53. 10.1038/s41588-017-0014-7 [PubMed: 29273806]
- Dowle M, & Srinivasan A (2020). data.table: Extension of data.frame [Computer software]. https:// Rdatatable.gitlab.io/data.table
- Ehret GB, Ferreira T, Chasman DI, Jackson AU, Schmidt EM, Johnson T, Thorleifsson G, Luan J, Donnelly LA, Kanoni S, Petersen A-K, Pihur V, Strawbridge RJ, Shungin D, Hughes MF, Meirelles O, Kaakinen M, Bouatia-Naji N, Kristiansson K, ... Munroe PB (2016). The genetics of blood pressure regulation and its target organs from association studies in 342,415 individuals. Nature Genetics, 48(10), 1171–1184. 10.1038/ng.3667 [PubMed: 27618452]
- ENCODE Project Consortium. (2012). An integrated encyclopedia of DNA elements in the human genome. Nature, 489(7414), 57–74. 10.1038/nature11247 [PubMed: 22955616]
- Evangelou E, Warren HR, Mosen-Ansorena D, Mifsud B, Pazoki R, Gao H, Ntritsos G, Dimou N, Cabrera CP, Karaman I, Ng FL, Evangelou M, Witkowska K, Tzanis E, Hellwege JN, Giri A, Velez Edwards DR, Sun YV, Cho K, ... Million Veteran Program (2018). Genetic analysis of over 1 million people identifies 535 new loci associated with blood pressure traits. Nature Genetics, 50(10), 1412–1425. 10.1038/s41588-018-0205-x [PubMed: 30224653]
- Fagerberg L, Hallström BM, Oksvold P, Kampf C, Djureinovic D, Odeberg J, Habuka M, Tahmasebpoor S, Danielsson A, Edlund K, Asplund A, Sjöstedt E, Lundberg E, Szigyarto CA-K, Skogs M, Takanen JO, Berling H, Tegel H, Mulder J, ... Uhlén M (2014). Analysis of the human tissue-specific expression by genome-wide integration of transcriptomics and antibody-based proteomics. Molecular & Cellular Proteomics, 13(2), 397–406. 10.1074/mcp.M113.035600
- Ferreira MA, Vonk JM, Baurecht H, Marenholz I, Tian C, Hoffman JD, Helmer Q, Tillander A, Ullemar V, van Dongen J, Lu Y, Rüschendorf F, Esparza-Gordillo J, Medway CW, Mountjoy E, Burrows K, Hummel O, Grosche S, Brumpton BM, ... Paternoster L (2017). Shared genetic origin of asthma, hay fever and eczema elucidates allergic disease biology. Nature Genetics, 49(12), 1752–1757. 10.1038/ng.3985 [PubMed: 29083406]
- Ferreira MAR, Mathur R, Vonk JM, Szwajda A, Brumpton B, Granell R, Brew BK, Ullemar V, Lu Y, & Jiang Y, 23 and Me Research Team, eQTLGen Consortium, BIOS Consortium, Magnusson PKE, Karlsson R, Hinds DA, Paternoster L, Koppelman GH, & Almqvist C (2019). Genetic architectures of childhood- and adult-onset asthma are partly distinct. The American Journal of Human Genetics, 104(4), 665–684. 10.1016/j.ajhg.2019.02.022 [PubMed: 30929738]
- Fingerlin TE, Murphy E, Zhang W, Peljto AL, Brown KK, Steele MP, Loyd JE, Cosgrove GP, Lynch D, Groshong S, Collard HR, Wolters PJ, Bradford WZ, Kossen K, Seiwert SD, du Bois RM,

Garcia CK, Devine MS, Gudmundsson G, ... Schwartz DA (2013). Genome-wide association study identifies multiple susceptibility loci for pulmonary fibrosis. Nature Genetics, 45(6), 613–620. 10.1038/ng.2609 [PubMed: 23583980]

- Féjerman L, Chen GK, Eng C, Huntsman S, Hu D, Williams A, Pasaniuc B, John EM, Via M, Gignoux C, Ingles S, Monroe KR, Kolonel LN, Torres-Mejía G, Pérez-Stable EJ, Burchard EG, Henderson BE, Haiman CA, & Ziv E (2012). Admixture mapping identifies a locus on 6q25 associated with breast cancer risk in US Latinas. Human Molecular Genetics, 21(8), 1907–1917. 10.1093/hmg/ddr617 [PubMed: 22228098]
- Giri A, Hellwege JN, Keaton JM, Park J, Qiu C, Warren HR, Torstenson ES, Kovesdy CP, Sun YV, Wilson OD, Robinson-Cohen C, Roumie CL, Chung CP, Birdwell KA, Damrauer SM, DuVall SL, Klarin D, Cho K, Wang Y, ... Edwards TL (2019). Trans-ethnic association study of blood pressure determinants in over 750,000 individuals. Nature Genetics, 51(1), 51–62. 10.1038/ s41588-018-0303-9 [PubMed: 30578418]
- Glindmeyer HW, Lefante JJ, McColloster C, Jones RN, & Weill H (1995). Blue-collar normative spirometric values for Caucasian and African-American men and women aged 18 to 65. American Journal of Respiratory and Critical Care Medicine, 151(2 Pt 1), 412–422. 10.1164/ ajrccm.151.2.7842200 [PubMed: 7842200]
- Gogarten SM, Sofer T, Chen H, Yu C, Brody JA, Thornton TA, Rice KM, & Conomos MP (2019). Genetic association testing using the GENESIS R/Bioconductor package. Bioinformatics, 10.1093/ bioinformatics/btz567
- Gruchała-Niedoszytko M, Niedoszytko M, Sanjabi B, van der Vlies P, Niedoszytko P, Jassem E, & Małgorzewicz S (2015). Analysis of the differences in whole-genome expression related to asthma and obesity. Polskie Archiwum Medycyny Wewnetrznej, 125(10), 722–730. 10.20452/pamw.3109 [PubMed: 26252510]
- Hankinson JL, Odencrantz JR, & Fedan KB (1999). Spirometric reference values from a sample of the general U.S. population. American Journal of Respiratory and Critical Care Medicine, 159(1), 179–187. 10.1164/ajrccm.159.1.9712108 [PubMed: 9872837]
- Hardin M, Cho MH, McDonald M-L, Wan E, Lomas DA, Coxson HO, MacNee W, Vestbo J, Yates JC, Agusti A, Calverley PMA, Celli B, Crim C, Rennard S, Wouters E, Bakke P, Bhatt SP, Kim V, Ramsdell J, ... COPDGene Investigators—clinical centers (2016). A genome-wide analysis of the response to inhaled β2-agonists in chronic obstructive pulmonary disease. The Pharmacogenomics Journal, 16(4), 326–335. 10.1038/tpj.2015.65 [PubMed: 26503814]
- Harik-Khan RI, Muller DC, & Wise RA (2004). Racial difference in lung function in African-American and White children: Effect of anthropometric, socioeconomic, nutritional, and environmental factors. American Journal of Epidemiology, 160(9), 893–900. 10.1093/aje/kwh297 [PubMed: 15496542]
- Heit JA, Armasu SM, McCauley BM, Kullo IJ, Sicotte H, Pathak J, Chute CG, Gottesman O, Bottinger EP, Denny JC, Roden DM, Li R, Ritchie MD, & de Andrade M (2017). Identification of unique venous thromboembolism-susceptibility variants in African-Americans. Thrombosis and Haemostasis, 117(4), 758–768. 10.1160/TH16-08-0652 [PubMed: 28203683]
- Hernandez W, Gamazon ER, Smithberger E, O'Brien TJ, Harralson AF, Tuck M, Barbour A, Kittles RA, Cavallari LH, & Perera MA (2016). Novel genetic predictors of venous thromboembolism risk in African Americans. Blood, 127(15), 1923–1929. 10.1182/blood-2015-09-668525 [PubMed: 26888256]
- Hinrichs AS, Karolchik D, Baertsch R, Barber GP, Bejerano G, Clawson H, Diekhans M, Furey TS, Harte RA, Hsu F, Hillman-Jackson J, Kuhn RM, Pedersen JS, Pohl A, Raney BJ, Rosenbloom KR, Siepel A, Smith KE, Sugnet CW, ... Kent WJ (2006). The UCSC genome browser database: Update 2006. Nucleic Acids Research, 34(Database issue), D590–D598. 10.1093/nar/gkj144 [PubMed: 16381938]
- Hsi BP, Hsu KHK, & Jenkins DE (1983). Ventilatory functions of normal children and young adults: Mexican-American, white, and black. III. Sitting height as a predictor. The Journal of Pediatrics, 102(6), 860–865. 10.1016/S0022-3476(83)80012-2 [PubMed: 6854449]
- Johansson Å, Rask-Andersen M, Karlsson T, & Ek WE (2019). Genome-wide association analysis of 350 000 Caucasians from the UK Biobank identifies novel loci for asthma, hay fever and eczema. Human Molecular Genetics, 28(23), 4022–4041. 10.1093/hmg/ddz175 [PubMed: 31361310]

Tukey John. (1977). Exploratory data analysis. Addison-Wesley.

- Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q, Collins RL, Laricchia KM, Ganna A, Birnbaum DP, Gauthier LD, Brand H, Solomonson M, Watts NA, Rhodes D, Singer-Berk M, England EM, Seaby EG, Kosmicki JA, ... Genome Aggregation Database Consortium. (2020). The mutational constraint spectrum quantified from variation in 141,456 humans. Nature, 581(7809), 434–443. 10.1038/s41586-020-2308-7 [PubMed: 32461654]
- Kichaev G (2017). PAINTOR v3.0. https://github.com/gkichaev/PAINTOR_V3.0
- Kichaev G, Bhatia G, Loh P-R, Gazal S, Burch K, Freund MK, Schoech A, Pasaniuc B, & Price AL (2019). Leveraging polygenic functional enrichment to improve GWAS power. American Journal of Human Genetics, 104(1), 65–75. 10.1016/j.ajhg.2018.11.008 [PubMed: 30595370]
- Kichaev G, Yang W-Y, Lindstrom S, Hormozdiari F, Eskin E, Price AL, Kraft P, & Pasaniuc B (2014). Integrating functional data to prioritize causal variants in statistical fine-mapping studies. PLOS Genetics, 10(10), e1004722 10.1371/journal.pgen.1004722 [PubMed: 25357204]
- Klarin D, Damrauer SM, Cho K, Sun YV, Teslovich TM, Honerlaw J, Gagnon DR, DuVall SL, Li J, Peloso GM, Chaffin M, Small AM, Huang J, Tang H, Lynch JA, Ho Y-L, Liu DJ, Emdin CA, Li AH, ... Assimes TL (2018). Genetics of blood lipids among ~300,000 multi-ethnic participants of the Million Veteran Program. Nature Genetics, 50(11), 1514–1523. 10.1038/s41588-018-0222-9 [PubMed: 30275531]
- Kumar R, Seibold MA, Aldrich MC, Williams LK, Reiner AP, Colangelo L, Galanter J, Gignoux C, Hu D, Sen S, Choudhry S, Peterson EL, Rodriguez-Santana J, Rodriguez-Cintron W, Nalls MA, Leak TS, O'Meara E, Meibohm B, Kritchevsky SB, ... Burchard EG (2010). Genetic ancestry in lung-function predictions. The New England Journal of Medicine, 363(4), 321–330. 10.1056/ NEJMoa0907897 [PubMed: 20647190]
- Laufer VA, Tiwari HK, Reynolds RJ, Danila MI, Wang J, Edberg JC, Kimberly RP, Kottyan LC, Harley JB, Mikuls TR, Gregersen PK, Absher DM, Langefeld CD, Arnett DK, & Bridges SL (2019). Genetic influences on susceptibility to rheumatoid arthritis in African-Americans. Human Molecular Genetics, 28(5), 858–874. 10.1093/hmg/ddy395 [PubMed: 30423114]
- Lessard CJ, Li H, Adrianto I, Ice JA, Rasmussen A, Grundahl KM, Kelly JA, Dozmorov MG, Miceli-Richard C, Bowman S, Lester S, Eriksson P, Eloranta M-L, Brun JG, Gøransson LG, Harboe E, Guthridge JM, Kaufman KM, Kvarnström M, ... Sivils KL (2013). Variants at multiple loci implicated in both innate and adaptive immune responses are associated with Sjögren's syndrome. Nature Genetics, 45(11), 1284–1292. 10.1038/ng.2792 [PubMed: 24097067]
- Loh P-R, Danecek P, Palamara PF, Fuchsberger C, Reshef YA, Finucane HK, Schoenherr S, Forer L, McCarthy S, Abecasis GR, Durbin R, & Price AL (2016). Reference-based phasing using the Haplotype Reference Consortium panel. Nature Genetics, 48(11), 1443–1448. 10.1038/ng.3679 [PubMed: 27694958]
- Lutz SM, Cho MH, Young K, Hersh CP, Castaldi PJ, McDonald M-L, Regan E, Mattheisen M, DeMeo DL, Parker M, Foreman M, Make BJ, Jensen RL, Casaburi R, Lomas DA, Bhatt SP, Bakke P, Gulsvik A, Crapo JD, ... COPDGene Investigators (2015). A genome-wide association study identifies risk loci for spirometric measures among smokers of European and African ancestry. BMC Genetics, 16, 138 10.1186/s12863-015-0299-4 [PubMed: 26634245]
- Mahajan A, Taliun D, Thurner M, Robertson NR, Torres JM, Rayner NW, Payne AJ, Steinthorsdottir V, Scott RA, Grarup N, Cook JP, Schmidt EM, Wuttke M, Sarnowski C, Mägi R, Nano J, Gieger C, Trompet S, Lecoeur C, ... McCarthy MI (2018). Fine-mapping type 2 diabetes loci to singlevariant resolution using high-density imputation and islet-specific epigenome maps. Nature Genetics, 50(11), 1505–1513. 10.1038/s41588-018-0241-6 [PubMed: 30297969]
- Mak ACY, White MJ, Eckalbar WL, Szpiech ZA, Oh SS, Pino-Yanes M, Hu D, Goddard P, Huntsman S, Galanter J, Wu AC, Himes BE, Germer S, Vogel JM, Bunting KL, Eng C, Salazar S, Keys KL, Liberto J, ... NHLBI Trans-Omics for Precision Medicine (TOPMed) Consortium (2018). Whole-genome sequencing of pharmacogenetic drug response in racially diverse children with asthma. American Journal of Respiratory and Critical Care Medicine, 197(12), 1552–1564. 10.1164/ rccm.201712-2529OC [PubMed: 29509491]
- Maples BK, Gravel S, Kenny EE, & Bustamante CD (2013). RFMix: A discriminative modeling approach for rapid and robust local-ancestry inference. American Journal of Human Genetics, 93(2), 278–288. 10.1016/j.ajhg.2013.06.020 [PubMed: 23910464]

- Martín-Hernández E, Rodríguez-García ME, Camacho A, Matilla-Dueñas A, García-Silva MT, Quijada-Fraile P, Corral-Juan M, Tejada-Palacios P, de Las Heras RS, Arenas J, Martín MA, & Martínez-Azorín F (2016). New ATP8A2 gene mutations associated with a novel syndrome: Encephalopathy, intellectual disability, severe hypotonia, chorea and optic atrophy. Neurogenetics, 17(4), 259–263. 10.1007/s10048-016-0496-y [PubMed: 27679995]
- McInnes G, Tanigawa Y, DeBoever C, Lavertu A, Olivieri JE, Aguirre M, & Rivas MA (2019). Global Biobank Engine: Enabling genotype-phenotype browsing for biobank summary statistics. Bioinformatics, 35(14), 2495–2497. 10.1093/bioinformatics/bty999 [PubMed: 30520965]
- Microsoft Corporation & Weston S (2019). doParallel: Foreach Parallel Adaptor for the 'parallel' Package. R package (Version 1.0.15) [Computer Software]. https://CRAN.R-project.org/ package=doParallel
- Miller MR, Crapo R, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, Enright P, van der Grinten CPM, Gustafsson P, Jensen R, Johnson DC, MacIntyre N, McKay R, Navajas D, Pedersen OF, Pellegrino R, Viegi G, Wanger J, & ATS/ERS Task Force (2005). General considerations for lung function testing. The European Respiratory Journal, 26(1), 153–161. 10.1183/09031936.05.00034505 [PubMed: 15994402]
- Nishimura KK, Galanter JM, Roth LA, Oh SS, Thakur N, Nguyen EA, Thyne S, Farber HJ, Serebrisky D, Kumar R, Brigino-Buenaventura E, Davis A, LeNoir MA, Meade K, Rodriguez-Cintron W, Avila PC, Borrell LN, Bibbins-Domingo K, Rodriguez-Santana JR, ... Burchard EG (2013). Early-life air pollution and asthma risk in minority children. The GALA II and SAGE II studies. American Journal of Respiratory and Critical Care Medicine, 188(3), 309–318. 10.1164/ rccm.201302-0264OC [PubMed: 23750510]
- Nurmagambetov T, Kuwahara R, & Garbe P (2018). The economic burden of asthma in the United States, 2008–2013. Annals of the American Thoracic Society, 15(3), 348–356. 10.1513/ AnnalsATS.201703-259OC [PubMed: 29323930]
- Ober C, & Yao T-C (2011). The genetics of asthma and allergic disease: A 21st century perspective. Immunological Reviews, 242(1), 10–30. 10.1111/j.1600-065X.2011.01029.x [PubMed: 21682736]
- Patterson N, Price AL, & Reich D (2006). Population structure and eigenanalysis. PLOS Genetics, 2(12), e190 10.1371/journal.pgen.0020190 [PubMed: 17194218]
- Pellegrino R, Viegi G, Brusasco V, Crapo RO, Burgos F, Casaburi R, Coates A, van der Grinten CPM, Gustafsson P, Hankinson J, Jensen R, Johnson DC, MacIntyre N, McKay R, Miller MR, Navajas D, Pedersen OF, & Wanger J (2005). Interpretative strategies for lung function tests. The European Respiratory Journal, 26(5), 948–968. 10.1183/09031936.05.00035205 [PubMed: 16264058]
- Pickrell JK, Berisa T, Liu JZ, Ségurel L, Tung JY, & Hinds DA (2016). Detection and interpretation of shared genetic influences on 42 human traits. Nature Genetics, 48(7), 709–717. 10.1038/ng.3570 [PubMed: 27182965]
- Pino-Yanes M, Thakur N, Gignoux CR, Galanter JM, Roth LA, Eng C, Nishimura KK, Oh SS, Vora H, Huntsman S, Nguyen EA, Hu D, Drake KA, Conti DV, Moreno-Estrada A, Sandoval K, Winkler CA, Borrell LN, Lurmann F, ... Burchard EG (2015). Genetic ancestry influences asthma susceptibility and lung function among Latinos. The Journal of Allergy and Clinical Immunology, 135(1), 228–235. 10.1016/j.jaci.2014.07.053 [PubMed: 25301036]
- Pividori M, Schoettler N, Nicolae DL, Ober C, & Im HK (2019). Shared and distinct genetic risk factors for childhood-onset and adult-onset asthma: Genome-wide and transcriptome-wide studies. The Lancet. Respiratory Medicine, 7(6), 509–522. 10.1016/S2213-2600(19)30055-4 [PubMed: 31036433]
- Plummer M, Best N, Cowles K, & Vines K (2006). CODA: Convergence diagnosis and output analysis for MCMC. R News, 6(1), 7–11.
- Popejoy AB, & Fullerton SM (2016). Genomics is failing on diversity. Nature, 538(7624), 161–164. 10.1038/538161a [PubMed: 27734877]
- Pulit SL, Stoneman C, Morris AP, Wood AR, Glastonbury CA, Tyrrell J, Yengo L, Ferreira T, Marouli E, Ji Y, Yang J, Jones S, Beaumont R, Croteau-Chonka DC, Winkler TW, Consortium G, Hattersley AT, Loos RJF, Hirschhorn JN, ... Lindgren CM (2019). Meta-analysis of genome-wide association studies for body fat distribution in 694 649 individuals of European ancestry. Human Molecular Genetics, 28(1), 166–174. 10.1093/hmg/ddy327 [PubMed: 30239722]

- Quanjer PH (2013). Lung function, race and ethnicity: A conundrum. European Respiratory Journal, 41(6), 1249–1251. 10.1183/09031936.00053913
- Quanjer PH (2015). Lung function, genetics and socioeconomic conditions. The European Respiratory Journal, 45(6), 1529–1533. 10.1183/09031936.00053115 [PubMed: 26028617]
- Risch N, & Merikangas K (1996). The future of genetic studies of complex human diseases. Science, 273(5281), 1516–1517. 10.1126/science.273.5281.1516 [PubMed: 8801636]
- Ro, admap Epigenomics Consortium, Kundaje A, Meuleman W, Ernst J, Bilenky M, Yen A, Heravi-Moussavi A, Kheradpour P, Zhang Z, Wang J, Ziller MJ, Amin V, Whitaker JW, Schultz MD, Ward LD, Sarkar A, Quon G, Sandstrom RS, Eaton ML, ... Kellis M (2015). Integrative analysis of 111 reference human epigenomes. Nature, 518(7539), 317–330. 10.1038/nature14248 [PubMed: 25693563]
- van Rooij FJA, Qayyum R, Smith AV, Zhou Y, Trompet S, Tanaka T, Keller MF, Chang L-C, Schmidt H, Yang M-L, Chen M-H, Hayes J, Johnson AD, Yanek LR, Mueller C, Lange L, Floyd JS, Ghanbari M, Zonderman AB, ... Ganesh SK (2017). Genome-wide trans-ethnic meta-analysis identifies seven genetic loci influencing erythrocyte traits and a role for RBPMS in erythropoiesis. American Journal of Human Genetics, 100(1), 51–63. 10.1016/j.ajhg.2016.11.016 [PubMed: 28017375]
- Rossiter CE, & Weill H (1974). Ethnic differences in lung function: Evidence for proportional differences. International Journal of Epidemiology, 3(1), 55–61. 10.1093/ije/3.1.55 [PubMed: 4838716]
- Rüeger S, McDaid A, & Kutalik Z (2018). Evaluation and application of summary statistic imputation to discover new height-associated loci. PLOS Genetics, 14(5), e1007371 10.1371/ journal.pgen.1007371 [PubMed: 29782485]
- Schwartz J, Katz SA, Fegley RW, & Tockman MS (1988). Sex and race differences in the development of lung function. American Review of Respiratory Disease, 138(6), 1415–1421. 10.1164/ajrccm/ 138.6.1415
- Shahin MH, Conrado DJ, Gonzalez D, Gong Y, Lobmeyer MT, Beitelshees AL, Boerwinkle E, Gums JG, Chapman A, Turner ST, Cooper-DeHoff RM, & Johnson JA (2018). Genome-wide association approach identified novel genetic predictors of heart rate response to β-blockers. Journal of the American Heart Association, 7(5), 10.1161/JAHA.117.006463
- Shriner D (2013). Overview of admixture mapping. Current Protocols in Human Genetics, 76(1), 1.23.1–1.23.8. 10.1002/0471142905.hg0123s76
- Smemo S, Tena JJ, Kim K-H, Gamazon ER, Sakabe NJ, Gómez-Marín C, Aneas I, Credidio FL, Sobreira DR, Wasserman NF, Lee JH, Puviindran V, Tam D, Shen M, Son JE, Vakili NA, Sung H-K, Naranjo S, Acemel RD, ... Nóbrega MA (2014). Obesity-associated variants within FTO form long-range functional connections with IRX3. Nature, 507(7492), 371–375. 10.1038/nature13138 [PubMed: 24646999]
- Sobota RS, Shriner D, Kodaman N, Goodloe R, Zheng W, Gao Y-T, Edwards TL, Amos CI, & Williams SM (2015). Addressing population-specific multiple testing burdens in genetic association studies, Annals of Human Genetics (79, pp. 136–147. 2 10.1111/ahg.12095 [PubMed: 25644736]
- Spear ML, Hu D, Pino-Yanes M, Huntsman S, Eng C, Levin AM, Ortega VE, White MJ, McGarry ME, Thakur N, Galanter J, Mak ACY, Oh SS, Ampleford E, Peters SP, Davis A, Kumar R, Farber HJ, Meade K, ... Burchard EG (2019). A genome-wide association and admixture mapping study of bronchodilator drug response in African Americans with asthma. The Pharmacogenomics Journal, 19(3), 249–259. 10.1038/s41397-018-0042-4 [PubMed: 30206298]
- Suhre K, Arnold M, Bhagwat AM, Cotton RJ, Engelke R, Raffler J, Sarwath H, Thareja G, Wahl A, DeLisle RK, Gold L, Pezer M, Lauc G, El-Din Selim MA, Mook-Kanamori DO, Al-Dous EK, Mohamoud YA, Malek J, Strauch K, ... Graumann J (2017). Connecting genetic risk to disease end points through the human blood plasma proteome. Nature Communications, 8, 14357 10.1038/ ncomms14357
- Tange O (2018). GNU Parallel 2018. Ole Tange. 10.5281/zenodo.1146014
- Thakur N, Oh SS, Nguyen EA, Martin M, Roth LA, Galanter J, Gignoux CR, Eng C, Davis A, Meade K, LeNoir MA, Avila PC, Farber HJ, Serebrisky D, Brigino-Buenaventura E, Rodriguez-Cintron W, Kumar R, Williams LK, Bibbins-Domingo K, ... Burchard EG (2013). Socioeconomic status

and childhood asthma in urban minority youths. The GALA II and SAGE II studies. American Journal of Respiratory and Critical Care Medicine, 188(10), 1202–1209. 10.1164/ rccm.201306-1016OC [PubMed: 24050698]

The 1000 Genomes Project Consortium. (2015). A global reference for human genetic variation. Nature, 526(7571), 68–74. 10.1038/nature15393 [PubMed: 26432245]

The Global Asthma Network. (2018). The Global Asthma Report 2018 http:// www.globalasthmareport.org/foreword/summaries.php

The International HapMap 3 Consortium. (2010). Integrating common and rare genetic variation in diverse human populations. Nature, 467(7311), 52–58. 10.1038/nature09298 [PubMed: 20811451]

Tsoi LC, Spain SL, Ellinghaus E, Stuart PE, Capon F, Knight J, Tejasvi T, Kang HM, Allen MH, Lambert S, Stoll SW, Weidinger S, Gudjonsson JE, Koks S, Kingo K, Esko T, Das S, Metspalu A, Weichenthal M, ... Elder JT (2015). Enhanced meta-analysis and replication studies identify five new psoriasis susceptibility loci. Nature Communications, 6, 7001 10.1038/ncomms8001

- Wanger J, Clausen JL, Coates A, Pedersen OF, Brusasco V, Burgos F, Casaburi R, Crapo R, Enright P, van der Grinten CPM, Gustafsson P, Hankinson J, Jensen R, Johnson D, Macintyre N, McKay R, Miller MR, Navajas D, Pellegrino R, & Viegi G (2005). Standardisation of the measurement of lung volumes. The European Respiratory Journal, 26(3), 511–522. 10.1183/09031936.05.00035005 [PubMed: 16135736]
- Whelton P.aulK, Carey Robert M, Aronow Wilbert S, Casey Donald E, Collins Karen J, Dennison Himmelfarb Cheryl, DePalma Sondra M, Gidding S.amuel, Jamerson Kenneth A, Jones Daniel W, MacLaughlin E.ricJ., Paul M.untner, Bruce O.vbiagele, Smith Sidney C, Spencer Crystal C, Stafford Randall S, Taler Sandra J, Thomas Randal J, Williams Kim A, ... Wright Jackson T (2018). 2017 ACC/AHA/AAPA/ABC/ACPM/AGS/APhA/ASH/ASPC/NMA/PCNA guideline for the prevention, detection, evaluation, and management of high blood pressure in adults: A report of the american college of cardiology/american heart association task force on clinical practice guidelines. Hypertension, 71(6), e13–e115. 10.1161/HYP.0000000000000065 [PubMed: 29133356]
- White MJ, Risse-Adams O, Goddard P, Contreras MG, Adams J, Hu D, Eng C, Oh SS, Davis A, Meade K, Brigino-Buenaventura E, LeNoir MA, Bibbins-Domingo K, Pino-Yanes M, & Burchard EG (2016). Novel genetic risk factors for asthma in African American children: Precision medicine and the SAGE II Study. Immunogenetics, 68(6–7), 391–400. 10.1007/ s00251-016-0914-1 [PubMed: 27142222]
- Whitlock EP, Williams SB, Gold R, Smith PR, & Shipman SA (2005). Screening and interventions for childhood overweight: A summary of evidence for the US Preventive Services Task Force. Pediatrics, 116(1), e125–e144. 10.1542/peds.2005-0242 [PubMed: 15995013]
- Wickham H (2016). ggplot2: Elegant graphics for data analysis. Springer-Verlag http://ggplot2.org
- Wickham H, & Garrett G (2017). R for Data Science. O'Reilly Media, Inc. https://r4ds.had.co.nz/
- Wojcik GL, Graff M, Nishimura KK, Tao R, Haessler J, Gignoux CR, Highland HM, Patel YM, Sorokin EP, Avery CL, Belbin GM, Bien SA, Cheng I, Cullina S, Hodonsky CJ, Hu Y, Huckins LM, Jeff J, Justice AE, ... Carlson CS (2019). Genetic analyses of diverse populations improves discovery for complex traits. Nature, 570(7762), 514–518. 10.1038/s41586-019-1310-4 [PubMed: 31217584]
- Wyss AB, Sofer T, Lee MK, Terzikhan N, Nguyen JN, Lahousse L, Latourelle JC, Smith AV, Bartz TM, Feitosa MF, Gao W, Ahluwalia TS, Tang W, Oldmeadow C, Duan Q, de Jong K, Wojczynski MK, Wang X-Q, Noordam R, ... London SJ (2018). Multiethnic meta-analysis identifies ancestry-specific and cross-ancestry loci for pulmonary function. Nature Communications, 9(1), 1–15. 10.1038/s41467-018-05369-0
- Yang J, Lee SH, Goddard ME, & Visscher PM (2011). GCTA: A tool for genome-wide complex trait analysis. American Journal of Human Genetics, 88(1), 76–82. 10.1016/j.ajhg.2010.11.011 [PubMed: 21167468]
- Yang J, Zaitlen NA, Goddard ME, Visscher PM, & Price AL (2014). Advantages and pitfalls in the application of mixed-model association methods. Nature Genetics, 46(2), 100–106. 10.1038/ ng.2876 [PubMed: 24473328]

- Zhu Y, Yao S, Iliopoulou BP, Han X, Augustine MM, Xu H, Phennicie RT, Flies SJ, Broadwater M, Ruff W, Taube JM, Zheng L, Luo L, Zhu G, Chen J, & Chen L (2013). B7–H5 costimulates human T cells via CD28H. Nature Communications, 4, 2043 10.1038/ncomms3043
- Zhu Z, Lee PH, Chaffin MD, Chung W, Loh P-R, Lu Q, Christiani DC, & Liang L (2018). A genomewide cross-trait analysis from UK Biobank highlights the shared genetic architecture of asthma and allergic diseases. Nature Genetics, 50(6), 857–864. 10.1038/s41588-018-0121-0 [PubMed: 29785011]



FIGURE 1.

Manhattan plots summarizing GWAS *p* values for all six lung function phenotypes. The solid red line denotes genome-wide significance ($p < 9.95 \times 10^{-8}$), while the dashed blue line marks the suggestive threshold ($p < 1.99 \times 10^{-6}$), per CODA calculations. Variants with a *p* value greater than 0.05 were deemed uninformative and therefore not plotted

Credible Set: 95.0%

LD (r²)

LD (r)

0.8 1.0





FIGURE 2.

A CANVIS plot of results from PAINTOR functional fine-mapping for locus 1, an association with pre-FEV₁ on chromosome 21. The SNP rs13615, which sits in the 3'-UTR of the gene ADAMTS1, attains a posterior probability of causality of 0.630. The panels show, from top to bottom, the posterior probability of causality; the five most informative functional annotations; GWAS p values; and local linkage disequilibrium expressed as a signed Pearson correlation. 3'-UTR, 3'-untranslated region; FEV₁, forced expiratory volume in 1 s; SNP, single nucleotide polymorphism

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FIGURE 3.

PAINTOR results for locus 2, an association on chromosome 4 with pre-FVC. The sentinel SNP, rs10857225, corresponds with a GWAS peak that does not pass Bonferroni correction for statistical significance. The highlighted peak tags the intron of the gene *THAP6*. FVC, forced vital capacity; SNP, single nucleotide polymorphism



FIGURE 4.

PAINTOR fine-mapping results for locus 5, corresponding to a region on chromosome 14 associated with post-FVC. The most likely causal SNP, rs1351618, tags an intron of the gene *EGLN3*. FVC, forced vital capacity; SNP, single nucleotide polymorphism

TABLE 1

Summary statistics of phenotypes and covariates from the SAGE cohort

Characteristics	Cases	Controls	Total
Subjects (n)	831	272	1,103
Age (year)	14.1 (3.66)	16.3 (3.77)	14.7 (3.8)
Female (<i>n</i>)	406	166	572
Height (cm)	158 (14.34)	162.4 (13.26)	159.1 (14.2)
African ancestry (%)	80.4 (0.1)	79.6 (0.1)	80.2 (0.1)
Maternal education (yr)	12.4 (1.47)	12.2 (1.5)	12.3 (1.48)
Obesity status			
Obese (n)	276	74	350
Nonobese (n)	555	198	753
Pre-FEV ₁	103 (13.79)	98.1 (13.02)	99.3 (13.77)
Pre-FVC	103.4 (12.84)	105.1 (13.09)	103.8 (12.92)
Pre-FEV ₁ /FVC	95.1 (9.35)	98.4 (8.2)	95.9 (9.19)
Post-FEV ₁	107 (13.44)	n/a	n/a
Post-FVC	109 (14.42)	n/a	n/a
Post-FEV ₁ /FVC	99 (7.83)	n/a	n/a

Note: Displayed numbers are either counts (n) or averages followed by standard errors in parentheses. Units are listed where appropriate. An "n/a" appears where measurements were taken on cases only

Abbreviations: FEV1, forced expiratory volume in 1 s; FVC, forced vital capacity.

Significant asso	ociatio	on results fro	m G	VAS					
Phenotype	Chr	Position (bp)	A1	A2	MAF	ą	SE	<i>p</i> value	Genes
Post-FEV1/FVC	15	85798401	A	IJ	0.018	-6.7846	1.27	9.05×10^{-8}	ADAMTS74P, AKAP13
Post-FVC	11	118932913	U	H	0.010	18.9094	3.29	$9.47 imes 10^{-9}$	CXCR5, HY0U1
Post-FVC	11	118902275	U	H	0.010	17.9775	3.29	4.83×10^{-8}	CXCR5, HYOUI
Post-FVC	11	118905095	A	IJ	0.010	17.9775	3.29	4.83×10^{-8}	CXCR5, HY0U1
Post-FVC	11	118905316	Н	U	0.010	17.9775	3.29	4.83×10^{-8}	CXCR5, HYOUI
Post-FVC	11	118906065	A	IJ	0.010	17.9775	3.29	4.83×10^{-8}	CXCR5, HY0U1
Post-FVC	11	118906240	Н	U	0.010	17.9775	3.29	4.83×10^{-8}	CXCR5, HY0U1
Post-FVC	11	118906745	U	IJ	0.010	17.9775	3.29	4.83×10^{-8}	CXCR5, HYOUI
Post-FVC	11	118907923	IJ	H	0.010	18.2978	3.41	$7.85 imes 10^{-8}$	CXCR5, HYOUI
Post-FVC	19	4289259	Н	U	0.106	5.1672	0.967	$9.21 imes 10^{-8}$	TMIGD2, SHD
Post-FVC	19	4291817	Н	U	0.111	5.2674	0.963	4.52×10^{-8}	TMIGD2, SHD
Pre-FEV1/FVC	13	26235394	IJ	A	0.010	-10.5334	1.94	$5.80 imes 10^{-8}$	ATP8A2
Pre-FEV1/FVC	13	26247080	IJ	A	0.010	-10.5334	1.94	$5.80 imes 10^{-8}$	ATP8A2
Pre-FEV1/FVC	13	26262378	Н	IJ	0.011	-10.5325	1.94	$5.81 imes 10^{-8}$	ATP8A2
Pre-FEV1/FVC	13	26268604	A	U	0.011	-10.5325	1.94	$5.81 imes 10^{-8}$	ATP8A2
Pre-FVC	20	22900228	IJ	A	0.010	18.0953	2.93	6.77×10^{-10}	THBD
Pre-FVC	16	54327903	IJ	A	0.023	9.8543	1.78	2.83×10^{-8}	IRX3, FTO
Pre-FVC	16	54327610	A	IJ	0.034	8.3689	1.53	4.85×10^{-8}	IRX3, FTO

Abbreviations: A1, major allele; A2, minor allele; B, effect size; FEV1, forced expiratory volume in 1 s; FVC, forced vital capacity; Genes, any genes within proximity of the associated variants; MAF, minor allele frequency; SE, standard error of the estimate of β . Note: The p values for all SNPs listed here met the significance threshold of 9.95×10^{-8} . SNPs were specified by chromosome (Chr) and physical position in base pairs (bp).

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							TABLE 3			
Admix	ture mapping	in SA	GE identifie	ed five regi	ons with stat	tistically sig	gnificant asso	ociation to at	least one j	phenotype
Locus	Phenotype	Chr	Start (bp)	End (bp)	Length (kb)	Threshold	$SNP_{Admix}(n)$	SNP _{GWAS} (n)	AFR (%)	Genes
1	Pre-FEV1	21	28,209,667	28,240,392	30.72	1.03×10^{-4}	4	215	78.7	ADAMTSI
2	Pre-FVC	4	75,555,658	76,873,740	1318.08	9.64×10^{-5}	102	7905	79.8	RCHY1, TI

hg19 coordinates. The total length of the region is given in kilobasepairs (kb). The threshold for statistical significance is given for each region. The column "Genes" lists genes physically within and near Note: The regions are arbitrarily numbered from 1 to 5 and defined by phenotype, chromosome (Chr), physical starting point (in base pairs), and end point (in base pairs). Physical positions are given in EGLN3, SNX6 80.5 1982 95 9.93×10^{-5} 311.50 34,595,061 34,283,561 4 the associated regions. Post-FVC ŝ

ESRP1, INTS8, TP53INP1, NDUFAF6

80.8

2822

 9.93×10^{-5} $1.04 imes 10^{-4}$

432.65 59.09

95,820,594

95,387,941

 ∞

Post-FVC

4

7,127,294

7,068,207

19

Pre-FEV1/FVC

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376

18 45

RCHY1, THAP6 INSR, ZNF557

78.7 79.8 80.4

counts the number (m) of genotyped SNPs from admixture mapping that met the significance threshold; SNPGWAS, the total number of GWAS SNPs (genotyped and/or imputed) in the admixture mapping Abbreviations: AFR, the percentage (%) of local genetic ancestry of African origin; FEV1, forced expiratory volume in 1 s; FVC, forced vital capacity; SNP, single nucleotide polymorphism; SNPAdmix. region.

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TABLE 4

Results from PAINTOR highlighting the most probably causal SNPs for each locus as defined by admixture mapping

Locus	Phenotype	Chr	Position (bp)	SNP	Ref allele	Alt allele	MAF (%)	P-value	Annotation	Pr (causal)
1	$Pre-FEV_1$	21	28,209,667	rs13615	A	IJ	2.55	6.95×10^{-3}	ADAMTSI, 3' -UTR	0.630
2	Pre-FVC	4	76,464,584	rs10857225	A	С	59.07	3.11×10^{-6}	THAP6, Intron Variant	0.361
ю	Pre-FEV ₁ /FVC	19	7,087,789	rs72986681	IJ	A	1.54	7.25×10^{-4}	ZNF557, 3' -UTR	0.168
4	Post-FVC	8	95,399,551	rs2470740	А	Т	23.82	2.04×10^{-3}	RAD54B, Intron Variant	0.109
5	Post-FVC	14	34,531,633	rs1351618	C	Т	12.40	$1.99 imes 10^{-4}$	EGLN3, Intron Variant	0.390
				,				. . i		

Note: As in Table 3, the loci are arbitrarily numbered from 1 to 5 and defined by phenotype and chromosome. The physical position (in basepairs) of the most likely causal SNP is given in hg19 coordinates. MAFs are taken from global populations from the gnomAD server v3. The displayed p values are from our discovery GWAS. Pr(causal) is computed from PAINTOR.

Abbreviations: 3'-UTR, 3'-untranslated region; Chr, chromosome; FEV1, forced expiratory volume in 1 s; FVC, forced vital capacity; MAF, minor allele frequency; Pr (causal), probability of causality; SNP, single nucleotide polymorphism.