

Lawrence Berkeley National Laboratory

Lawrence Berkeley National Laboratory

Title

Aging, tumor suppression and cancer: High-wire act!

Permalink

<https://escholarship.org/uc/item/2332r6q5>

Author

Campisi, Judith

Publication Date

2004-08-15

Peer reviewed



Aging, tumor suppression and cancer: high wire-act!

Judith Campisi^{a,b,*}

^a*Life Sciences Division, Lawrence Berkeley National Laboratory, 1 Cyclotron Road, Berkeley, CA 94720, USA*

^b*Buck Institute for Age Research, 8001 Redwood Blvd., Novato, CA 94545, USA*

Abstract

Evolutionary theory holds that aging is a consequence of the declining force of natural selection with age. We discuss here the evidence that among the causes of aging in complex multicellular organisms, such as mammals, is the antagonistically pleiotropic effects of the cellular responses that protect the organism from cancer. Cancer is relatively rare in young mammals, owing in large measure to the activity of tumor suppressor mechanisms. These mechanisms either protect the genome from damage and/or mutations, or they elicit cellular responses—apoptosis or senescence—that eliminate or prevent the proliferation of somatic cells at risk for neoplastic transformation. We focus here on the senescence response, reviewing its causes, regulation and effects. In addition, we describe recent data that support the idea that both senescence and apoptosis may indeed be the double-edged swords predicted by the evolutionary hypothesis of antagonistic pleiotropy—protecting organisms from cancer early in life, but promoting aging phenotypes, including late life cancer, in older organisms.

© 2004 Published by Elsevier Ireland Ltd.

Keywords: Antagonistic pleiotropy; Apoptosis; Cellular senescence; Gene expression Pattern; p53; p16

1. Introduction

There have been extraordinary advances in the last decade in understanding the evolution of genomes and the genetic basis for aging. The idea that aging is under genetic control may now seem obvious, especially considering the sometimes large differences in life span among organisms with comparatively similar genomes (Williams, 1957). Recent discoveries, however, have explicitly identified evolutionarily conserved genes that are important regulators of life span, as well as early life fitness, among diverse species (Guarente and Kenyon, 2000; Kirkwood and Austad, 2000; Walker et al., 2000; Finch and Ruvkun, 2001). In general, these recent findings support modern evolutionary theories of aging. They have uncovered candidate genes on which evolution likely acted to produce species-specific life spans, and elucidated conserved pathways within organisms that link metabolism, reproduction and life span. At present, we still know remarkably little about the cellular and molecular bases for longevity differences among species.

However, owing in large measure to our recent understanding of the genetic similarities in longevity pathways among species, we are gaining important insights into the mechanisms that control aging within species.

Here, we review the evolutionary theory of antagonistic pleiotropy, and emerging evidence that aging in complex multicellular organisms is caused in part by the antagonistically pleiotropic effects of tumor suppressive mechanisms—mechanisms that evolved to prevent the development of cancer in young organisms.

2. Evolution of life spans and cancer

2.1. Environment and the evolution of genomes

An obvious tenet of any evolutionary theory is that heritable traits, including species-specific life spans, are controlled by genes, and that these genes in turn evolved in response to environmental pressures. In addition, the environments in which genomes evolve are typically fraught with natural hazards—predators, infection, food scarcity, harsh climatic conditions, etc., which generally kill

* Tel.: +1 510 486 4416; fax: +1 510 486 4545.

E-mail address: jcampisi@lbl.gov.

organisms long before they reach “old age”. That is, the environments in which genomes evolve typically eliminate reproductively fit organisms at relatively young ages. Consequently, any gene mutation or multi-gene process that has deleterious albeit delayed effects—delayed with respect to the average age at which the environment eliminates organisms—will be retained. Thus, aging is thought to occur because phenotypes that have escaped the force of natural selection persist (Williams, 1957; Kirkwood and Austad, 2000; Finch and Ruvkun, 2001).

Consider, now, that genomes evolve relatively slowly, and that environments can change relatively rapidly. This is precisely what has happened to modern humans. We have, very rapidly on evolutionary time scales, eliminated many of the extrinsic hazards that dominated the environments in which our genome evolved. Parenthetically, we have also done this for our favorite animal models for biomedical research, at least within the confines of our animal colonies! Hence, aging—those delayed deleterious phenotypes that escaped the force of natural selection during evolution—is pervasive, especially among developed populations, which have been most successful in eliminating extrinsic hazards.

2.2. Cancer and age-related disease

Just as aging is considered by evolutionary biologists to be a consequence of the declining force of natural selection with age, age-related diseases can be considered phenotypes that have escaped the force of natural selection. Age-related diseases are generally considered to be degenerative in nature, a result of the overall decline in tissue structure and function that is a hallmark of aging. There are, however, exceptions. The most notable of these is cancer.

In simple terms, cancer can be considered a gain-of-function disease (discussed further below). That is, cells must acquire functions (e.g., hyperproliferation, resistance to cell death, migratory and invasive properties) in order to give rise to malignant tumors. Of particular interest, cancer is not a universal feature of aging. There are many organisms, including some of our favorite organisms for studying aging (*Drosophila melanogaster*; *Caenorhabditis elegans*), which never develop cancer. Here, we define cancer as an ectopic mass of supernumerary cells that develops postnatally and has the potential to kill the organism. What distinguishes organisms that do and do not develop cancer? One obvious distinction is the presence of renewable somatic tissues—somatic tissues that contain dividing or division-competent cells—in the adults. Thus, cell division in adult somatic tissues appears to put organisms at risk for developing cancer. This is not surprising, given that we now know that DNA replication puts cells at endanger for acquiring and fixing mutations (Kunkel and Bebenek, 2000; van Brabant et al., 2000; Friedberg et al., 2002; Thompson and Schild, 2002), and that somatic mutations are a major cause of cancer (Bishop,

1995; Simpson and Camargo, 1998; Gray and Collins, 2000; Knudson, 2000).

Why would evolution select for organisms with renewable tissues, given the danger of developing cancer? The benefits of renewable tissues—the ability to regenerate or repair tissues damaged by injury or endogenous degenerative processes—may have outweighed their risks, in conjunction with the co-evolution of mechanism to suppress cancer (discussed below). Moreover, we speculate that the evolution of renewable tissues afforded organisms long life spans.

3. Tumor suppressor mechanisms

Because cell division can lead to mutations and hence cancer, organisms with renewable tissues had to evolve strategies to prevent the cancer. Collectively, these strategies are termed tumor suppressor mechanisms. Tumor suppressor mechanisms can, in general, be broadly classified into two major categories—caretakers and gatekeepers (Kinzler and Vogelstein, 1997).

Caretaker tumor suppressors act on the genome, generally by preventing or repairing DNA damage. Thus, caretaker tumor suppressors restrain the development of cancer by suppressing the development of mutations. Since mutations not only cause cancer, but have also been proposed to independently contribute to aging, genes that encode caretaker tumor suppressor functions are straightforward longevity assurance genes (Martin, 1966; Dolle et al., 2002; Vijg and Dolle, 2002; Hasty et al., 2003).

Gatekeeper tumor suppressors, by contrast, act on cells, causing them to die (apoptosis) or permanently arrest proliferation (senescence). Thus, gatekeeper tumor suppressors restrain the development of cancer by eliminating or preventing the growth (used here interchangeably with growth) of potential cancer cells. In contrast to the caretakers, we suggest that genes encoding gatekeeper tumor suppressors have both beneficial (anti-cancer) and deleterious (pro-aging) effects, depending on the age of the organism. That is, gatekeeper tumor suppressors are an example of evolutionary antagonistic pleiotropy, which, as discussed below, is hypothesized to explain at least in part why organisms age.

4. Antagonistic pleiotropy

Because aging is a consequence of the declining force of natural selection with age, traits that benefit young organisms—suppressing cancer, for example—can have unselected deleterious effects—driving aging phenotypes, for example—later in the life span. This is the concept of evolutionary antagonistic pleiotropy (Williams, 1957; Rose, 1991; Kirkwood and Austad, 2000; Finch and Ruvkun, 2001). In simple terms, antagonistic pleiotropy holds that what is good for an organism when it is young can be bad for

164 it when it is old. Antagonistic pleiotropy occurs because
 165 genes with the “good” attributes of optimizing the fitness of
 166 young organisms evolved under the pressure of natural
 167 environments, in which, as discussed above, extrinsic
 168 hazards are high and old organisms are relatively rare. If
 169 the same genes have “bad” attributes, but their manifesta-
 170 tion is delayed—that is, manifest only after most of the
 171 population has been eliminated by extrinsic hazards—they
 172 cannot be eliminated by natural selection. Hence, genes with
 173 delayed deleterious actions (antagonistic pleiotropy) can
 174 persist. The consequences of their deleterious effects, then,
 175 are abundantly evident only in populations in which the
 176 hazardous environmental pressures have eased rapidly
 177 relative to the pace at which genomes evolve—the situation
 178 humans in the developed world (and mice in our animal
 179 colonies) now face.

180 How might the gatekeeper tumor suppressor genes—
 181 those that control apoptosis and cell senescence—be
 182 antagonistically pleiotropic?

183 Apoptosis prevents cancer by virtually eliminating cells
 184 that are damaged or otherwise potentially oncogenic (Reed,
 185 1999; Green and Evan, 2002; Hickman et al., 2002). On the
 186 other hand, apoptosis can eventually deplete tissues of their
 187 constituent cells and/or deplete the stem cell pools that
 188 replenish renewable tissues (Joaquin and Gollapudi, 2001;
 189 Weinstein and Ciszek, 2002; Zhang and Herman, 2002;
 190 Campisi, 2003a, 2003b). Hence, with increasing age,
 191 apoptosis might cause an overall loss of tissue structure and
 192 function, a hallmark of aging. Apoptosis may be an especially
 193 important contributor to the degenerative diseases of aging.

194 Along the same lines of reasoning, cellular senescence
 195 prevents cancer by arresting the growth of potentially
 196 oncogenic cells (Sager, 1991; Bringold and Serrano, 2000;
 197 Lundberg et al., 2000; Reddel, 2000; Campisi, 2001).
 198 However, with increasing age, senescent cells, which are
 199 incapable of regeneration and show marked changes in
 200 function (discussed below), can accumulate. Again, this
 201 accumulation can lead to an overall loss of tissue structure
 202 and function (Campisi, 1996, 2003a, 2003b; Smith and
 203 Pereira-Smith, 1996; Faragher, 2000). Like apoptosis,
 204 cellular senescence may contribute to the degenerative
 205 diseases of aging. In addition, because the secretory
 206 phenotype of senescent cells can alter the local tissue
 207 milieu, senescent cells may also contribute to the
 208 hyperproliferative diseases of aging, including—ironi-
 209 cally—cancer (Campisi, 1997, 2003a, 2003b).

210 At present, very little is known about what determines
 211 whether mammalian cells recover, die or senesce in the face
 212 of damage or stress. Some cell types—for example, T
 213 cells—are more prone to undergo apoptosis than senes-
 214 cence, whereas the opposite is true for other cell types—for
 215 example, fibroblasts. In addition, the level and type of stress
 216 may determine whether cells undergo an apoptotic or
 217 senescence response. Whatever the case, at least in
 218 mammals, both apoptosis and senescence important
 219 defenses against the development of cancer, yet both

processes have the potential to be deleterious with time, and
 hence in older organisms.

Here, we review the evidence that the gatekeeper tumor
 suppressors that control apoptosis and senescence may be
 antagonistically pleiotropic. We focus our discussion
 primarily on the mammalian senescence response, which
 arguably has more complex age-related consequences in vivo
 than the apoptotic response, and for which there is mounting
 evidence for a role in aging. However, it is important to bear in
 mind that parallel arguments may hold for apoptosis.

5. The senescence response

5.1. Causes of senescence

The senescence response was first formally described as
 the process that limits the proliferation of human cells in
 culture (Hayflick, 1965). We now know that this limit is due
 in large measure to the loss of telomeric DNA that occurs
 when cells that do not express telomerase undergo DNA
 replication (Levy et al., 1992; Wright and Shay, 2001).
 Telomeres, the DNA sequence and proteins that cap the ends
 of linear chromosomes, are essential chromosomal ele-
 ments, loss of which causes genomic instability, an
 enormous risk factor for malignant transformation (Artandi
 and DePinho, 2000; Shay and Wright, 2001; Kim et al.,
 2002; Blasco, 2003). Thus, the senescence response to short
 dysfunctional telomeres serves to arrest the growth of cells
 in danger of genomic instability, consistent with its role in
 tumor suppression.

In the last decade, it has become clear that many events and
 stimuli in addition to telomere dysfunction—all of which put
 cells at risk for neoplastic transformation—can induce a
 senescence response. These events include DNA damage
 (DiLeonardo et al., 1994; Chen et al., 1995; Robles and
 Adami, 1998), as well as perturbations to chromatin
 organization (Ogryzko et al., 1996; Jacobs et al., 1999;
 Itahana et al., 2003; Narita et al., 2003). They also include the
 expression of certain oncogenes (Serrano et al., 1997; Zhu et
 al., 1998; Dimri et al., 2000) that deliver supraphysiological
 mitogenic signals to cells, and the overexpression of certain
 tumor suppressor genes (Sugrue et al., 1997; McConnell et
 al., 1998; Dai and Enders, 2000; Dimri et al., 2000;
 Beausejour et al., 2003). The most potent tumor suppressor
 genes that induce senescence when overexpressed are those
 that encode components of the p53 and pRB tumor suppressor
 pathways, both of which are crucial for the senescence
 response (Shay et al., 1991; Bringold and Serrano, 2000;
 Lundberg et al., 2000; Campisi, 2001; Itahana et al., 2001).

5.2. The senescent phenotype

The senescence response is not a simple arrest of cell
 proliferation. Rather, senescent cells adopt a complex
 phenotype that entails many changes in gene expression

(Cristofalo and Pignolo, 1993; Campisi et al., 1996; Faragher, 2000; Krtolica and Campisi, 2002). In addition to imposing a block to cell cycle progression, the senescence response causes changes in cell morphology (generally, adoption of an enlarged flattened shape). It also renders many (although not all) cell types resistant to apoptotic signals. Furthermore, senescent cells acquire cell-type specific functional changes. Thus, senescent cells fail to proliferate, but also become resistant to elimination by apoptosis and do not function normally. We hypothesize that the resistance to apoptosis may explain why senescent cells accumulate, while their altered function may explain how they contribute to aging and age-related disease.

What are the functional changes that accompany the senescence response? These changes have been best characterized in fibroblasts, the cell type that synthesizes and maintains the stroma, the structure that underlies the cells of epithelial tissues and regulates their function (Donjacour and Cunha, 1991). Of particular interest, senescent human fibroblasts develop a secretory phenotype characterized by increased secretion of extracellular matrix remodeling enzymes, inflammatory cytokines and epithelial growth factors (Campisi, 1996; Faragher, 2000; Krtolica and Campisi, 2002). These secreted molecules can, at least in principle, have a field effect—altering the microenvironment of the surrounding tissue with respect to structure, inflammation status and epithelial function.

Because the senescent phenotype entails functional changes that can alter tissue structure and function, senescent cells—as they accumulate—may progressively promote the decline in tissue structure and function that characterizes aging. It is in this way that the senescence response may be antagonistically pleiotropic—protecting organisms from cancer at young ages, but promoting aging phenotypes at old ages. How much evidence is there for this idea?

6. Testing the hypothesis that gatekeeper tumor suppressors, specifically the senescence response, is antagonistically pleiotropic

The hypothesis that gatekeeper tumor suppressors, and particularly the senescence response, is antagonistically pleiotropic makes a number of predictions, not all of which have been tested experimentally. Here, we review the major predictions and, where applicable, the pertinent experimental results.

6.1. Do senescent cells exist and accumulate with age *in vivo*?

A prime prediction of the above hypothesis is that senescent cells exist and accumulate with age in mammalian tissues. This appears to be the case, with the important caveat that at present we have very few markers with which to identify senescent cells. In addition to the enlarged

senescent morphology, one marker that is widely used is a neutral (pH 6) β -galactosidase, termed the senescence-associated β -galactosidase (SA-Bgal) (Dimri et al., 1995). The expression of this enzyme correlates strongly, although not exclusively, with the induction of senescence by any of the known senescence-inducing stimuli in a variety of cell types in culture. Because SA-Bgal is easily detected *in situ* by histochemical staining, it has been used to search for cells with senescent characteristics *in vivo*. Indeed, such (SA-Bgal-positive) cells have been found in several tissues from humans and rodents. More important, their frequency has been shown to rise with increasing age in human skin, monkey skin and retina, human prostate, rodent kidney, human liver and human vascular endothelium (Dimri et al., 1995; Mishima et al., 1999; Pendergrass et al., 1999; Choi et al., 2000; Ding et al., 2001; Paradis et al., 2001; Vasile et al., 2001; Melk et al., 2003). Moreover, as discussed below, cells with senescent characteristics have been found at sites of age-related pathology, including atherosclerotic plaques and benign and premalignant lesions of the liver and prostate.

While these studies constitute little more than a ‘smoking gun’, they at least suggest that senescent cells appear to be present at the predicted times and locations.

6.2. Do gatekeeper tumor suppressors promote aging?

Another prediction is that gatekeeper tumor suppressors should promote aging. As noted earlier, genes encoding two tumor suppressor proteins—p53 and pRB—are pivotal for establishing and maintaining cellular senescence. p53, a multifunctional transcriptional regulator (Sherr, 1998; Prives and Hall, 1999; Ryan et al., 2001; Wahl and Carr, 2001; Hofseth et al., 2004), is of particular interest because it is dispensable for mammalian embryogenesis and postnatal development, but crucial for preventing cancer. Indeed, most, if not all, malignant tumors harbor mutations in p53 or one of its critical regulators. p53 is also a quintessential gatekeeper tumor suppressor because it is a crucial regulator of both the apoptotic and senescence responses.

Recently, two groups created mouse models with constitutively hyperactive p53 (Tyner et al., 2002; Maier et al., 2004). p53 acts as a tetramer (Prives and Hall, 1999). In both mouse models, a truncated form of p53 was ubiquitously expressed, and the truncated forms were thought to form mixed tetramers with wild-type p53. Moreover, indirect evidence indicated that the mixed tetramers were hyperactive relative to tetramers composed solely of wild-type p53.

Consistent with p53’s role as a potent tumor suppressor, the mutant mice were strikingly resistant to cancer. Since cancer is a major cause of death in laboratory mice, one might expect the mutant mice to be long-lived. This was not the case, however. Rather, both mutant mouse strains had a modestly shorter life span and displayed multiple signs of premature aging! Moreover, cells from these mice were more prone to undergo apoptosis (Tyner et al., 2002) or

senescence (Maier et al., 2004) when stressed in culture. Thus, hyperactive p53 conferred enhanced protection from cancer, but at the cost of accelerated aging, and the accelerated aging was associated with heightened sensitivity to apoptosis and senescence.

How might hyperactive p53 might cause accelerated aging? At least a partial answer to this question was provided by analysis of the mouse model created by Maier et al. In these animals, p53 appeared to upregulate components of the IGF-1 signaling pathway, which delivers mitogenic, survival and metabolic signals to mammalian cells. Supraphysiological IGF-1 signaling, in turn, stimulated a senescence response in cells from these mice. This finding is significant because components of the IGF-1/insulin pathway are among the evolutionarily conserved genes that have been shown to be important positive regulators of aging in diverse species (Guarente and Kenyon, 2000; Finch and Ruvkun, 2001; Bluhner et al., 2003; Holzenberger et al., 2003; Rincon et al., 2004).

Together, these mouse models indicate that hyperactive tumor suppression by p53 promotes apoptosis and cell senescence and accelerates aging, supporting the idea that gatekeeper tumor suppressor functions are antagonistically pleiotropic (Campisi, 2004).

6.3. Do senescent cells promote age-related pathology?

A third prediction is that senescent cells should promote or accelerate the development of age-related pathology.

Although cancer is often studied independent of age, it is in fact a major age-related disease among mammals, age being the largest single risk factor for its development (Miller, 1991; DePinho, 2000; Balducci and Beghe, 2001; Campisi, 2003a, 2003b). The age-dependence with which cancer develops is sometimes attributed to the fact that cancer requires the accumulation of multiple mutations (Bishop, 1995; Simpson and Camargo, 1998; Gray and Collins, 2000; Knudson, 2000), which takes time. However, there is increasing evidence that mutation accumulation alone cannot fully explain why cancer incidence rises so sharply with age. What else is required for the development of cancer? Several decades of cell biology have established that many cells with oncogenic mutations also require a permissive tissue microenvironment in which to progress into a malignant tumor (DePinho, 2000; Park et al., 2000; Bissell and Radisky, 2001; Liotta and Kohn, 2001; Coussens and Werb, 2002). Of particular importance for the development of epithelial tumors—the major type of age-related cancer that develops in humans—are the interactions between the epithelium and the underlying stroma (Birchmeier and Birchmeier, 1995; DePinho, 2000; Bissell and Radisky, 2001; Chrenek et al., 2001; Liotta and Kohn, 2001; Tlsty and Hein, 2001).

Recent studies show that senescent stromal fibroblasts can stimulate the hyperproliferation of premalignant, but not normal, epithelial cells in culture (Krtolica et al., 2001; Dilley et al., 2003). Moreover, senescent fibroblast can stimulate the tumorigenic conversion of premalignant

epithelial cells into frankly malignant tumors in vivo (Krtolica et al., 2001). The phenotype of senescent fibroblasts, described above, strongly resembles that of “activated stroma” or carcinoma-associated fibroblasts, both which have been shown to strongly stimulate tumor progression in cell culture models and in vivo (Skobe and Fusenig, 1998; Olumi et al., 1999; Shekhar et al., 2001; Martens et al., 2003). Interestingly, irradiated fibroblasts, which were likely senescent albeit not explicitly characterized as such, were also shown to promote epithelial tumor progression in vivo (Barcellos-Hoff and Ravani, 2000).

In many of these cases, the cancer-promoting activity of the senescent or activated stroma was due at least in part to their secretory phenotype (Skobe and Fusenig, 1998; Olumi et al., 1999; Krtolica et al., 2001; Shekhar et al., 2001; Martens et al., 2003; Parrinello et al., submitted for publication). Thus, the age-dependent accumulation of senescent cells, particularly senescent stromal cells, may synergize with the age-dependent accumulation of mutations, resulting in the rise in epithelial cancers. This idea is consistent with the identification of cells with senescent characteristics at sites of hyperplastic and premalignant lesions (Choi et al., 2000; Paradis et al., 2001; Castro et al., 2003). It is ironic indeed that an effective early life tumor suppressor mechanism (cellular senescence) can fuel the development of cancer late in life (Campisi, 2003a, 2003b)!

Much less is known about whether or to what extent the presence of senescent cells contribute to other age-related pathologies. Cells that express SA-Bgal and lack expression of thymosin- β 10, characteristics of senescent endothelial cells in culture have been found at site of atherosclerosis in human aorta (Vasile et al., 2001). Given that senescent cells secrete inflammatory cytokines, and that inflammation is thought to be an important contributory factor in atherogenesis, this result is consistent with the idea that senescent cells can initiate or promote age-related vascular disease. Similar types of evidence have implicated senescent cells in the etiology and/or progression of kidney fibrosis and age-related kidney dysfunction (Ding et al., 2001; Melk et al., 2003), osteoarthritis of the joints (Martin and Buckwalter, 2003) and venous ulcers of the lower extremities (Stanley and Osler, 2001).

Taken together, these findings support the idea that senescent cells can promote age-related pathologies, but the data thus far are of course still correlative and indirect. In some cases, cell culture and limited in vivo data indicate that senescent cells *can* promote pathological phenotypes. In other cases, senescent cells appear to be a ‘smoking gun’ by virtue of their presence.

6.4. Does reversal of the senescent phenotype or elimination of senescent cells prevent or ameliorate age-related pathology?

At present, we know very little about how to reverse the senescent phenotype or how to eliminate senescent cells in

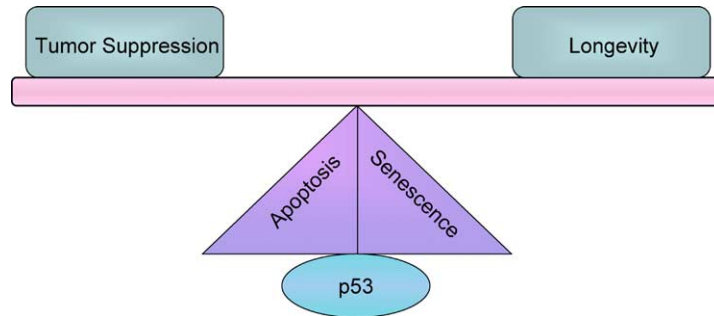


Fig. 1. Tumor suppression and longevity are balanced. In complex organisms such as mammals, we suggest that mechanisms that protect against cancer are balanced against those that ensure longevity. Specifically, the gatekeeper tumor suppressors such as p53, which regulate apoptosis and the senescence of replicative cells, are antagonistically pleiotropic, suppressing cancer in young organisms but promoting aging—including late life cancer—in old organisms.

483 vivo. We do know it is possible to reactivate the growth of
 484 some types of senescent cells by inactivating p53 (Gire and
 485 Wynford-Thomas, 1998; Beausejour et al., 2003). Cells that
 486 express the p16 tumor suppressor protein, which activates
 487 the pRB tumor suppressor, are refractory to reversal by p53
 488 inactivation (Beausejour et al., 2003). We do not yet know
 489 how prevalent the p16 block to senescence reversal is in
 490 vivo. Whatever the case, inactivation of p53 is not a practical
 491 means for reversing the effects of senescent cells in vivo, as
 492 this will surely increase the incidence of cancer.

493 **7. Summary, conclusions and challenges**

494 We hypothesize that mechanisms that protect complex
 495 organisms from cancer (gatekeeper tumor suppressors) do so
 496 in balance against the mechanisms that promote longevity
 497 (Fig. 1). These cancer protection mechanisms engage the
 498 cellular processes of apoptosis and senescence. Both
 499 apoptosis and senescence are crucial for suppressing
 500 malignant tumorigenesis, yet both have the potential to
 501 contribute to aging and age-related pathology. The
 502 senescence response is notable in that the senescent
 503 phenotype may, ironically, also promote cancer at advanced
 504 ages. Critical testing of the hypothesis that the senescence
 505 response is antagonistically pleiotropic faces the challenge
 506 of developing ways to reverse the senescent phenotype
 507 without reversing the growth arrest, or switching the
 508 senescence response to an apoptotic response. If the
 509 hypothesis is correct, however, we face an even larger
 510 challenge—how to develop practical interventions that will
 511 minimize the pro-aging actions of gatekeeper tumor
 512 suppressor genes, while maintaining protection from cancer.
 513 The path to meeting this challenge is not yet clear, but will
 514 likely emerge as we develop a deeper understanding of how
 515 the senescent phenotype is established and maintained.

516 **Acknowledgements**

517 As always, thanks to past and present lab members for
 518 their refreshing ideas and hard work, colleagues for

stimulating discussions and valuable reagents, and the
 National Institute on Aging, Department of Defense and
 University of California Breast Cancer Research Programs,
 Ellison Medical Foundation and Department of Energy for
 research support.

References

Artandi, S.E., DePinho, R.A., 2000. A critical role for telomeres in
 suppressing and facilitating carcinogenesis. *Curr. Opin. Genet. Dev.*
 10, 39–46.
 Balducci, L., Beghe, C., 2001. Cancer and age in the USA. *Crit. Rev. Oncol.*
Hematol. 37, 137–145.
 Barcellos-Hoff, M.H., Ravani, S.A., 2000. Irradiated mammary gland
 stroma promotes the expression of tumorigenic potential by unirradiated
 epithelial cells. *Cancer Res.* 60, 1254–1260.
 Beausejour, C.M., Krtolica, A., Galimi, F., Narita, M., Lowe, S.W., Yaswen,
 P., Campisi, J., 2003. Reversal of human cellular senescence: roles of the
 p53 and p16 pathways. *EMBO J.* 22, 4212–4222.
 Birchmeier, W., Birchmeier, C., 1995. Epithelial-mesenchymal transitions
 in development and tumor progression. *EXS* 74, 1–15.
 Bishop, J.M., 1995. Cancer: The rise of the genetic paradigm. *Genes Dev.* 9,
 1309–1315.
 Bissell, M.J., Radisky, D., 2001. Putting tumours in context. *Nature Rev.*
Cancer 1, 46–54.
 Blasco, M.A., 2003. Mammalian telomeres and telomerase: why they
 matter for cancer and aging. *Eur. J. Cell Biol.* 82, 441–446.
 Blucher, M., Kahn, B.B., Kahn, C.R., 2003. Extended longevity in mice
 lacking the insulin receptor in adipose tissue. *Science* 299, 572–574.
 Bringold, F., Serrano, M., 2000. Tumor suppressors and oncogenes in
 cellular senescence. *Exp. Gerontol.* 35, 317–329.
 Campisi, J., 1996. Replicative senescence: an old lives tale? *Cell* 84, 497–
 500.
 Campisi, J., 1997. Aging and cancer: the double-edged sword of replicative
 senescence. *J. Am. Geriatr. Soc.* 45, 1–6.
 Campisi, J., 2001. Cellular senescence as a tumor-suppressor mechanism.
Trends Cell Biol. 11, 27–31.
 Campisi, J., 2003a. Cancer and ageing: rival demons? *Nature Rev. Cancer*
 3, 339–349.
 Campisi, J., 2003b. Cellular senescence and apoptosis: how
 cellular responses might influence aging phenotypes. *Exp. Geront.*
 38, 5–11.
 Campisi, J., 2004. Fragile fugue: p53 in aging, cancer and IGF signaling.
Nature Med. 10, 231–232.
 Campisi, J., Dimri, G.P., Hara, E., 1996. Control of replicative senescence.
 In: Schneider, E., Rowe, J. (Eds.), *Handbook of the Biology of Aging*.
 Academic Press, New York, pp. 121–149.

- 64 Castro, P., Giri, D., Lamb, D., Ittmann, M., 2003. Cellular senescence in the
65 pathogenesis of benign prostatic hyperplasia. *Prostate* 55, 30–38.
- 66 Chen, Q., Fischer, A., Reagan, J.D., Yan, L.J., Ames, B.N., 1995. Oxidative
67 DNA damage and senescence of human diploid fibroblast cells. *Proc.*
68 *Natl. Acad. Sci. U.S.A.* 92, 4337–4341.
- 69 Choi, J., Shendrik, I., Peacocke, M., Peehl, D., Buttyan, R., Ikeguchi, E.F.,
70 Katz, A.E., Benson, M.C., 2000. Expression of senescence-associated
71 beta-galactosidase in enlarged prostates from men with benign prostatic
72 hyperplasia. *Urology* 56, 160–166.
- 73 Chrenek, M.A., Wong, P., Weaver, V.M., 2001. Tumour-stromal interac-
74 tions. Integrins and cell adhesions as modulators of mammary cell
75 survival and transformation. *Breast Cancer Res.* 3, 224–229.
- 76 Coussens, L.M., Werb, Z., 2002. Inflammation and cancer. *Nature* 420, 860–
77 867.
- 78 Cristofalo, V.J., Pignolo, R.J., 1993. Replicative senescence of human
79 fibroblast-like cells in culture. *Physiol. Rev.* 73, 617–638.
- 80 Dai, C.Y., Enders, G.H., 2000. p16 INK4a can initiate an autonomous
81 senescence program. *Oncogene* 19, 1613–1622.
- 82 DePinho, R.A., 2000. The age of cancer. *Nature* 408, 248–254.
- 83 DiLeonardo, A., Linke, S.P., Clarkin, K., Wahl, G.M., 1994. DNA damage
84 triggers a prolonged p53-dependent G1 arrest and long-term
85 induction of Cip1 in normal human fibroblasts. *Genes Dev.* 8, 2540–
86 2551.
- 87 Dilley, T.K., Bowden, G.T., Chen, Q.M., 2003. Novel mechanisms of
88 sublethal oxidant toxicity: induction of premature senescence in
89 human fibroblasts confers tumor promoter activity. *Exp. Cell Res.*
90 290, 38–48.
- 91 Dimri, G.P., Itahana, K., Acosta, M., Campisi, J., 2000. Regulation of a
92 senescence checkpoint response by the E2F1 transcription factor and
93 p14/ARF tumor suppressor. *Mol. Cell. Biol.* 20, 273–285.
- 94 Dimri, G.P., Lee, X., Basile, G., Acosta, M., Scott, G., Roskelley, C.,
95 Medrano, E.E., Linskens, M., Rubelj, I., Pereira-Smith, O.M., Peacocke,
96 M., Campisi, J., 1995. A novel biomarker identifies senescent human
97 cells in culture and in aging skin in vivo. *Proc. Natl. Acad. Sci. U.S.A.*
98 92, 9363–9367.
- 99 Ding, G., Franki, N., Kapasi, A.A., Reddy, K., Gibbons, N., Singhal, P.C.,
600 2001. Tubular cell senescence and expression of TGF-beta1 and
601 p21(WAF1/CIP1) in tubulointerstitial fibrosis of aging rats. *Exp.*
602 *Mol. Pathol.* 70, 43–53.
- 603 Dolle, M.E., Snyder, W.K., Dunson, D.B., Vijg, J., 2002. Mutational
604 fingerprints of aging. *Nucleic Acids Res.* 30, 545–549.
- 605 Donjacour, A.A., Cunha, G.R., 1991. Stromal regulation of epithelial
606 function. *Cancer Treat. Res.* 53, 335–364.
- 607 Faragher, R.G., 2000. Cell senescence and human aging: where's the link?
608 *Biochem. Soc. Trans.* 28, 221–226.
- 609 Finch, C.E., Ruvkun, G., 2001. The genetics of aging. *Annu. Rev. Genomics*
610 *Hum. Genet.* 2, 435–462.
- 611 Friedberg, E.C., Wagner, R., Radman, M., 2002. Specialized DNA poly-
612 merases, cellular survival, and the genesis of mutations. *Science* 296,
613 1627–1630.
- 614 Gire, V., Wynford-Thomas, D., 1998. Reinitiation of DNA synthesis and cell
615 division in senescent human fibroblasts by microinjection of anti-p53
616 antibodies. *Mol. Cell. Biol.* 18, 1611–1621.
- 617 Gray, J.W., Collins, C., 2000. Genome changes and gene expression in
618 human solid tumors. *Carcinogenesis* 21, 443–452.
- 619 Green, D.R., Evan, G.I., 2002. A matter of life and death. *Cancer Cell* 1, 19–
620 30.
- 621 Guarente, L., Kenyon, C., 2000. Genetic pathways that regulate ageing in
622 model organisms. *Nature* 408, 255–262.
- 623 Hasty, P., Campisi, J., Hoeijmakers, J., van Steeg, H., Vijg, J., 2003. Aging
624 and genome maintenance: lessons from the mouse? *Science* 299, 1355–
625 1359.
- 626 Hayflick, L., 1965. The limited in vitro lifetime of human diploid cell
627 strains. *Exp. Cell Res.* 37, 614–636.
- 628 Hickman, E.S., Moroni, M.C., Helin, K., 2002. The role of p53 and pRB in
629 apoptosis and cancer. *Curr. Opin. Genet. Dev.* 12, 60–66.
- 630 Hofseth, L.J., Hussain, S.P., Harris, C.C., 2004. p53: 25 years after its
631 discovery. *Trends Pharmacol. Sci.* 25, 177–181.
- 632 Holzenberger, M., Dupont, J., Ducos, B., Leneuve, P., Geloën, A., Even,
633 P.C., Cervera, P., Le Bouc, Y., 2003. IGF-1 receptor regulates lifespan
634 and resistance to oxidative stress in mice. *Nature* 421, 182–187.
- 635 Itahana, K., Dimri, G., Campisi, J., 2001. Regulation of cellular senescence
636 by p53. *Eur. J. Biochem.* 268, 2784–2791.
- 637 Itahana, K., Zou, Y., Itahana, Y., Martinez, J.L., Beausejour, C., Jacobs, J.J.,
638 Van Lohuizen, M., Band, V., Campisi, J., Dimri, G.P., 2003. Control of
639 the replicative life span of human fibroblasts by p16 and the polycomb
640 protein Bmi-1. *Mol. Cell. Biol.* 23, 389–401.
- 641 Jacobs, J.J., Kieboom, K., Marino, S., DePinho, R.A., van Lohuizen, M.,
642 1999. The oncogene and Polycomb-group gene bmi-1 regulates cell
643 proliferation and senescence through the ink4a locus. *Nature* 397, 164–
644 168.
- 645 Joaquin, A.M., Gollapudi, S., 2001. Functional decline in aging and disease:
646 a role for apoptosis. *J. Am. Geriatr. Soc.* 49, 1234–1240.
- 647 Kim, S.H., Kaminker, P., Campisi, J., 2002. Telomeres, aging and cancer: in
648 search of a happy ending. *Oncogene* 21, 503–511.
- 649 Kinzler, K.W., Vogelstein, B., 1997. Cancer susceptibility genes: gate-
650 keepers and caretakers. *Nature* 386, 761–763.
- 651 Kirkwood, T.B., Austad, S.N., 2000. Why do we age? *Nature* 408, 233–238.
- 652 Knudson, A.G., 2000. Chasing the cancer demon. *Annu. Rev. Genet.* 34, 1–
653 19.
- 654 Krtolica, A., Campisi, J., 2002. Cancer and aging: a model for the cancer
655 promoting effects of the aging stroma. *Int. J. Biochem. Cell Biol.* 34,
656 1401–1414.
- 657 Krtolica, A., Parrinello, S., Lockett, S., Desprez, P., Campisi, J., 2001.
658 Senescent fibroblasts promote epithelial cell growth and tumorigenesis:
659 a link between cancer and aging. *Proc. Natl. Acad. Sci. U.S.A.* 98,
660 12072–12077.
- 661 Kunkel, T.A., Bebenek, K., 2000. DNA replication fidelity. *Annu. Rev.*
662 *Biochem.* 69, 497–529.
- 663 Levy, M.Z., Allsopp, R.C., Fletcher, A.B., Greider, C.W., Harley, C.B., 1992.
664 Telomere end-replication problem and cell aging. *J. Mol. Biol.* 225,
665 951–960.
- 666 Liotta, L.A., Kohn, E.C., 2001. The microenvironment of the tumour-host
667 interface. *Nature* 411, 375–379.
- 668 Lundberg, A.S., Hahn, W.C., Gupta, P., Weinberg, R.A., 2000. Genes
669 involved in senescence and immortalization. *Curr. Opin. Cell Biol.*
670 12, 705–709.
- 671 Maier, B., Gluba, W., Bernier, B., Turner, T., Mohammad, K., Guise, T.,
672 Sutherland, A., Thorne, M., Scoble, H., 2004. Modulation of mam-
673 malian life span by the short isoform of p53. *Genes Dev.* 18, 306–319.
- 674 Martens, J.W., Sieuwerts, A.M., Vries, J.B., Bosma, P.T., Swiggers, S.J.,
675 Klijn, J.G., Foekens, J.A., 2003. Aging of stromal-derived human breast
676 fibroblasts might contribute to breast cancer progression. *Thromb.*
677 *Haemost.* 89, 393–404.
- 678 Martin, G.M., 1966. Somatic mutagenesis and antimutagenesis in aging
679 research. *Mutat. Res.* 350, 35–41.
- 680 Martin, J.A., Buckwalter, J.A., 2003. The role of chondrocyte senescence in
681 the pathogenesis of osteoarthritis and in limiting cartilage repair. *J. Bone*
682 *Joint Surg. Am.* 85, 106–110.
- 683 McConnell, B.B., Starborg, M., Brookes, S., Peters, G., 1998. Inhibitors of
684 cyclin-dependent kinases induce features of replicative senescence in
685 early passage human diploid fibroblasts. *Curr. Biol.* 8, 351–354.
- 686 Melk, A., Kittikowit, W., Sandhu, I., Halloran, K.M., Grimm, P., Schmidt,
687 B.M., Halloran, P.F., 2003. Cell senescence in rat kidneys in vivo
688 increases with growth and age despite lack of telomere shortening.
689 *Kidney Int.* 63, 2134–2143.
- 690 Miller, R.A., 1991. Gerontology as oncology: research on aging
691 as a key to the understanding of cancer. *Cancer* 68, 2496–2501.
- 692 Mishima, K., Handa, J.T., Aotaki-Keen, A., Lutton, G.A., Morse, L.S.,
693 Hjelmeland, L.M., 1999. Senescence-associated beta-galactosidase his-
694 tochemistry for the primate eye. *Invest. Ophthalmol. Vis. Sci.* 40, 1590–
695 1593.
- 696

- 696 Narita, M., Nunez, S., Heard, E., Narita, M., Lin, A.W., Hearn, S.A.,
697 Spector, D.L., Hannon, G.J., Lowe, S.W., 2003. Rb-mediated hetero-
698 chromatin formation and silencing of E2F target genes during cellular
699 senescence. *Cell* 113, 703–716.
- 700 Ogrzyzko, V.V., Hirai, T.H., Russanova, V.R., Barbie, D.A., Howard, B.H.,
701 1996. Human fibroblast commitment to a senescence-like state in
702 response to histone deacetylase inhibitors is cell cycle dependent.
703 *Mol. Cell. Biol.* 16, 5210–5218.
- 704 Olumi, A.F., Grossfeld, G.D., Hayward, S.W., Carroll, P.R., Tlsty, T.D.,
705 Cunha, G.R., 1999. Carcinoma-associated fibroblasts direct tumor
706 progression of initiated human prostatic epithelium. *Cancer Res.* 59,
707 5002–5011.
- 708 Paradis, V., Youssef, N., Dargere, D., Ba, N., Bonvoust, F., Bedossa, P.,
709 2001. Replicative senescence in normal liver, chronic hepatitis C, and
710 hepatocellular carcinomas. *Hum. Pathol.* 32, 327–332.
- 711 Park, C.C., Bissell, M.J., Barcellos-Hoff, M.H., 2000. The influence of the
712 microenvironment on the malignant phenotype. *Mol. Med. Today* 6,
713 324–329.
- 714 Parrinello, S., Coppe, J.P., Krtolica, A., Campisi, J., Stromal-epithelial
715 interactions in aging and cancer: senescent fibroblasts can alter epithe-
716 lial cell differentiation, submitted for publication.
- 717 Pendergrass, W.R., Lane, M.A., Bodkin, N.L., Hansen, B.C., Ingram, D.K.,
718 Roth, G.S., Yi, L., Bin, H., Wolf, N.S., 1999. Cellular proliferation
719 potential during aging and caloric restriction in rhesus monkeys
720 (Macaca mulatta). *J. Cell. Physiol.* 180, 123–130.
- 721 Prives, C., Hall, P.A., 1999. The p53 pathway. *J. Pathol.* 187, 112–126.
- 722 Reddel, R.R., 2000. The role of senescence and immortalization in carci-
723 nogenesis. *Carcinogenesis* 21, 477–484.
- 724 Reed, J.C., 1999. Mechanisms of apoptosis in avoidance of cancer. *Curr.*
725 *Opin. Oncol.* 11, 68–75.
- 726 Rincon, M., Muzumdar, R., Atzmon, G., Barzilai, N., 2004. The paradox of
727 the insulin/IGF-1 signaling pathway in longevity. *Mech. Ageing Dev.*
728 125, 397–403.
- 729 Robles, S.J., Adami, G.R., 1998. Agents that cause DNA double strand
730 breaks lead to p16INK4a enrichment and the premature senescence of
731 normal fibroblasts. *Oncogene* 16, 1113–1123.
- 732 Rose, M.R., 1991. *The Evolutionary Biology of Aging*. Oxford University
733 Press, Oxford.
- 734 Ryan, K.M., Phillips, A.C., Voudsen, K.H., 2001. Regulation and
735 function of the p53 tumor suppressor protein. *Curr. Opin. Cell Biol.*
736 13, 332–337.
- 737 Sager, R., 1991. Senescence as a mode of tumor suppression. *Environ.*
738 *Health Perspect.* 93, 59–62.
- 739 Serrano, M., Lin, A.W., McCurrach, M.E., Beach, D., Lowe, S.W., 1997.
740 Oncogenic ras provokes premature cell senescence associated with
741 accumulation of p53 and p16INK4a. *Cell* 88, 593–602.
- 742 Shay, J.W., Wright, W.E., 2001. Telomeres and telomerase: implications for
743 cancer and aging. *Radiat. Res.* 155, 188–193.
- 744 Shay, J.W., Wright, W.R., Werbin, H., 1991. Defining the molecular
745 mechanisms of human cell immortalization. *Biochim. Biophys. Acta*
746 1071, 1–7.
- 747 Shekhar, M.P., Werdell, J., Santner, S.J., Pauley, R.J., Tait, L., 2001. Breast
748 stroma plays a dominant regulatory role in breast epithelial growth and
749 differentiation: implications for tumor development and progression. *Cancer Res.* 61, 1320–1326.
- 750 Sherr, C.J., 1998. Tumor surveillance via the ARF-p53 pathway. *Genes Dev.*
751 12, 2891–2984.
- 752 Simpson, A.J., Camargo, A.A., 1998. Evolution and the inevitability of
753 human cancer. *Sem. Cancer Biol.* 8, 439–445.
- 754 Skobe, M., Fusenig, N.E., 1998. Tumorigenic conversion of immortal
755 human keratinocytes through stromal activation. *Proc. Natl. Acad. Sci. U.S.A.* 95, 1050–1055.
- 756 Smith, J.R., Pereira-Smith, O.M., 1996. Replicative senescence: implica-
757 tions for in vivo aging and tumor suppression. *Science* 273, 63–67.
- 758 Stanley, A., Osler, T., 2001. Senescence and the healing rates of venous
759 ulcers. *J. Vasc. Surg.* 33, 1206–1211.
- 760 Sugrue, M.M., Shin, D.Y., Lee, S.W., Aaronson, S.A., 1997. Wild-type p53
761 triggers a rapid senescence program in human tumor
762 cells lacking functional p53. *Proc. Natl. Acad. Sci. U.S.A.* 94, 9648–
763 9653.
- 764 Thompson, L.H., Schild, D., 2002. Recombinational DNA repair and human
765 disease. *Mutat. Res.* 509, 49–78.
- 766 Tlsty, T.D., Hein, P.W., 2001. Know thy neighbor: stromal cells can
767 contribute oncogenic signals. *Curr. Opin. Genet. Dev.* 11, 54–59.
- 768 Tyner, S.D., Venkatachalam, S., Choi, J., Jones, S., Ghebranious, N.,
769 Ingelmann, H., Lu, X., Soron, G., Cooper, B., Brayton, C., Park,
770 S.H., Thompson, T., Karsenty, G., Bradley, A., Donehower, L., 2002.
771 p53 mutant mice that display early aging-associated phenotypes. *Nature*
772 415, 45–53.
- 773 van Brabant, A.J., Stan, R., Ellis, N.A., 2000. DNA helicases, genomic
774 instability, and human genetic disease. *Annu. Rev. Genomics Hum. Genet.* 1, 409–459.
- 775 Vasile, E., Tomita, Y., Brown, L.F., Kocher, O., Dvorak, H.F., 2001.
776 Differential expression of thymosin beta-10 by early passage and
777 senescent vascular endothelium is modulated by VPF/VEGF: evidence
778 for senescent endothelial cells in vivo at sites of atherosclerosis. *FASEB J.* 15, 458–466.
- 779 Vijg, J., Dolle, M.E., 2002. Large genome rearrangements as a primary
780 cause of aging. *Mech. Ageing Dev.* 123, 907–915.
- 781 Wahl, G.M., Carr, A.M., 2001. The evolution of diverse biological responses
782 to DNA damage: insights from yeast and p53. *Nature Cell Biol.* 3, 277–
783 286.
- 784 Walker, D.W., McColl, G., Jenkins, N.L., Harris, J., Lithgow, G.J., 2000.
785 Evolution of lifespan in *C. elegans*. *Nature* 405, 296–297.
- 786 Weinstein, B.S., Ciszek, D., 2002. The reserve capacity hypothesis: evolu-
787 tionary origins and modern implications between tumor suppression and
788 tissue repair. *Exp. Geront.* 37, 615–627.
- 789 Williams, G.C., 1957. Pleiotropy, natural selection, and the evolution of
790 senescence. *Evolution* 11, 398–411.
- 791 Wright, W.E., Shay, J.W., 2001. Cellular senescence as a tumor-protection
792 mechanism: the essential role of counting. *Curr. Opin. Genet. Dev.* 11,
793 98–103.
- 794 Zhang, Y., Herman, B., 2002. Ageing and apoptosis. *Mech. Ageing Dev.*
795 123, 245–260.
- 796 Zhu, J., Woods, D., McMahon, M., Bishop, J.M., 1998. Senescence of
797 human fibroblasts induced by oncogenic raf. *Genes Dev.* 12, 2997–3007.
- 798 800
799 801
802