

**UC Davis**

**The Proceedings of the International Plant Nutrition  
Colloquium XVI**

**Title**

Higher plants have the ability to reduce iodate to iodide.

**Permalink**

<https://escholarship.org/uc/item/23r7j0kw>

**Author**

Sekimoto, Hitoshi

**Publication Date**

2009-04-10

Peer reviewed

## Introduction

Although iodine is one of the essential elements of animals, it is not essential, but harmful, to plants. “Akagare” disease, a physiological disorder by iodine toxicity, occurs in lowland rice cultivated under flooded condition (Tensho and Yeh, 1970, Watanabe and Tensho, 1970). On the other hand, iodine is an essential microelement for human health. Iodine deficiency disorders are believed to be one of the commonest preventable human health problems (Welch and Graham, 1999, Grusak, 2002, Ramakrishnan, 2002). Therefore, enrichment of iodine in crops is a way to contribute to increasing the intake of iodine for human health. To avoid iodine toxicity in crops and to make iodine-biofortified crops, it is important to study iodine uptake and metabolism in plant depending on its chemical forms ( $\text{I}^-$ ,  $\text{IO}_3^-$  and  $\text{I}_2$ ).

Generally the forms of iodine found in natural environments are dependent on pH and electrochemical potential (Eh) (Mackowiak *et al.*, 2005).  $\text{I}^-$  should be the most prevalent (Yuita, 1992), while  $\text{IO}_3^-$  exists under more oxidizing conditions. It is known that plants absorb  $\text{I}^-$  more readily than  $\text{IO}_3^-$  so that  $\text{I}^-$  is more phytotoxic than  $\text{IO}_3^-$  (Mackowiak and Grossl, 1999; Umaly and Poel, 1971), and  $\text{IO}_3^-$  is electrochemically reduced to  $\text{I}^-$  before uptake (Böszörményi and Cseh, 1960; Whitehead, 1975). Also  $\text{I}^-$  may oxidize to  $\text{I}_2$ . However, iodine metabolism in higher plants using sensitive analytical techniques to separate  $\text{I}^-$  and  $\text{IO}_3^-$  was not conducted. Using the ion chromatography and inductively coupled plasma-mass spectrometry system (IC-ICP-MS) for the determination of  $\text{I}^-$  and  $\text{IO}_3^-$ , plant uptake and metabolism of inorganic iodine ( $\text{I}^-$  and  $\text{IO}_3^-$ ) and the r behavior in the rhizosphere were studied.

## Materials and methods

### *Plant culture*

Rice seeds (cv. Koshihikari) were surface sterilized with 70 mL L<sup>-1</sup> ethanol, and the seeds were germinated and grown for 16 days with tap water. Seedlings were transferred to 3L culture solution containing 4 mM N as  $\text{NH}_4\text{NO}_3$ , 0.5 mM P, 3 mM K, 0.5 mM Ca, 1 mM Mg, 1.5 mM S and standard levels of micronutrients in deionized water. The plants were grown in a growth chamber with 14/10 h photoperiod at 120  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  at 27/25°C. The media was adjusted to pH 6 and replaced every 3 days.

### *Experiment 1*

After 28 days of growing in culture media,  $\text{I}^-$  or  $\text{IO}_3^-$  as sodium salts was added to the media at 100 or 1000  $\mu\text{g L}^{-1}$ , respectively. Plants were harvested and separated into shoot and roots, and they were homogenized with 5 times-fold of deionised water after 24 h. of iodine treatments. Also xylem sap was collected with cutting at the basal shoot.  $\text{I}^-$ ,  $\text{IO}_3^-$  and total iodine concentration in the extraction of plant tissues, xylem sap and culture media were analyzed after pre-purification with a membrane filter (0.2 $\mu\text{m}$ ).

### *Experiment 2*

Detached roots (500mg FW) sterilized with 700 mL L<sup>-1</sup> ethanol for 2 min and rinsed with sterile deionized water were obtained from 22-d old rice seedlings grown in the culture media. They were immersed in 10 mL of autoclaved Tris-HCl buffer (50mM, pH 8.0) containing  $\text{I}^-$  or  $\text{IO}_3^-$  (20  $\mu\text{g L}^{-1}$  as I). After 1, 6 and 24 h. of immersion in the dark at 20 °C,  $\text{I}^-$  and  $\text{IO}_3^-$  concentration in the buffer solution were analyzed.

### *Iodine analysis*

The ion chromatography-inductively coupled plasma-mass spectrometry system (IC-ICP-MS)

(Yoshida *et al.*, 2007) was used for the determination of  $\Gamma^-$  and  $\text{IO}_3^-$  in solution samples. Detection limit is  $0.1 \mu\text{g L}^{-1}$  (as I). The analysis of total iodine developed by Muramatsu and Yoshida (1998) was applied. Powdered samples were mixed with  $\text{V}_2\text{O}_5$  in a small ceramic boat and placed in a quartz combustion tube. The end of the quartz tube was connected with a ball-joint to a trap containing  $\text{H}_2\text{O}$ , with tetramethyl ammonium hydroxide (TMAH), and  $\text{Na}_2\text{SO}_3$  solution. A wet oxygen flow was passed through the tube during the heating at  $1000^\circ\text{C}$  for 15 min. Sample solutions containing trapped iodine were adjusted to 10 mL by adding  $\text{H}_2\text{O}$ , and iodine content was determined by ICP-MS. Detection limit of total iodine in dry samples is  $1 \mu\text{g kg (DW)}^{-1}$ .

## Results and discussion

The most of iodine in rice plants was the fraction of the others that would be composed of water soluble organic iodine compounds and molecular iodine ( $\text{I}_2$ ).  $\Gamma^-$  was detected 24% of total-I in shoots and 22% in roots by  $\text{IO}_3^-$  treatment, but  $\text{IO}_3^-$  was not detected in  $\Gamma^-$ -treated plants. Furthermore, xylem sap of  $\text{IO}_3^-$ -treated plants did not contain  $\text{IO}_3^-$  and 70% of total-I in xylem sap was  $\Gamma^-$ , and the others. Similarly in the culture media, 11% of supplied  $\text{IO}_3^-$  was reduced to  $\Gamma^-$  for 24 h, but  $\Gamma^-$  was not changed to  $\text{IO}_3^-$ . Ratio of total-I concentration in shoot and roots to  $\Gamma^-$  concentration in the culture media of  $\text{IO}_3^-$  treatment after 24 h (12 in shoot, 42 in roots) was the same level as that of  $\Gamma^-$  treatment (14 in shoot, 51 in roots), but the ratio to total-I concentration in culture media of  $\text{IO}_3^-$  treatment (1.4 in shoot, 4.8 in roots) was quite different from that of  $\Gamma^-$  treatment (12 in shoot, 43 in roots), implying that total-I concentration in  $\text{IO}_3^-$ -treated plants depended on  $\Gamma^-$  concentration changing from  $\text{IO}_3^-$  in the culture media.  $\text{IO}_3^-$  would be reduced to  $\Gamma^-$  by roots or in the rhizosphere readily, and rice plants would absorb the reduced  $\Gamma^-$ . In Experiment 2, pH and Eh of the buffer solution were pH 8.1-7.9 and 140-40 mV, respectively. It was indicated that  $\text{IO}_3^-$  reduction in rice roots could also be performed under alkaline and oxidized conditions.

It is known that phytoplankton can reduce  $\text{IO}_3^-$  to  $\Gamma^-$  (Wang *et al.*, 2002, Chance *et al.*, 2007) as well as nitrate, sulfate or Fe (III) reducing bacteria (Councell *et al.*, 1997, Farrenkopf *et al.*, 1997, Tsunogai and Sase, 1969). In addition to results shown here,  $\text{IO}_3^-$  reduction was confirmed in soybean roots and culture cells derived from them. Not only microorganisms but also higher plants would have the ability of  $\text{IO}_3^-$  reduction. Mechanisms and meanings of  $\text{IO}_3^-$  reduction and absorption of the reduced  $\Gamma^-$ , despite the fact that iodine is not essential for higher plants, remain to be examined.

## References

- Böszörményi Z and Cseh E 1960 *Curr. Sci.* 29, 340–341.  
Chance R, Malin G, Jickells T and Baker AR 2007 *Marine Chem.* 105, 169-180.  
Councell TB, Landa ER and Lovleg PR 1997 *Water Air Soil Pollution.* 100, 99-106.  
Farrenkopf AM, Dollhopf ME, Chadhain SN, Luther GW and Nealson KH 1997 *Marine Chem.* 57, 347-354.  
Grusak MA 2002 *J Am Coll Nutr.* 21, 178S-183S.  
Mackowiak CL and Grossl PR 1999 *Plant Soil.* 212, 135–143.  
Mackowiak CL, Grossl PR and Cook KI 2005 *Plant Soil.* 269, 141-150.  
Muramatsu Y and Yoshida S 1998 *Geomicrobiol. J.* 16, 85-93.

- Ramakrishnan U 2002 *Nutrition Reviews*. 60,S46-S52.
- Tensho K and Yeh KL 1970 *Radioisotopes*. 19, 574–579.
- Tsunogai S and Sase T 1969 *Deep-Sea Res.* 16, 489-496.
- Umaly RC and Poel LW 1971 *Ann. Bot.* 35, 127–131.
- Wang GTF, Piumsomboon AU and Dustan WM 2002 *Mar. Ecol. Prog. Ser.* 237, 27-39.
- Watanabe and Tensho 1970 *Soil Sci. Plant Nutr.* 16, 30-37.
- Welch RM and Graham RD 1999 *Field Crop Res.* 60, 1-10.
- Whitehead DC 1975 *J. Sci. Fd. Agric.* 26, 361–367.
- Yoshida S, Muramatsu Y, Katou S and Sekimoto H 2007 *J. Radioanal. Nucl. Chem.* 273, 211-214.
- Yuita K 1992 *Soil Sci. Plant Nutr.* 38, 281–287