1 Dermal Uptake of Organic Vapors Commonly Found in Indoor Air

- 2
 3 Charles J. Weschler^{1,2*} and William W Nazaroff³
- 4
- 5 ¹Environmental and Occupational Health Sciences Institute, Rutgers University, Piscataway, NJ
- 6 08854, USA
- ⁷²International Centre for Indoor Environment and Energy, Technical University of Denmark,
- 8 DK-2800 Lyngby, Denmark
- ⁹ ³Department of Civil and Environmental Engineering, University of California, Berkeley, CA
- 10 94720-1710 USA
- 11
- 12 Keywords: Dermal absorption, exposure, percutaneous transport, permeability coefficient,
- 13 transdermal permeation
- 14 Short Title: Transdermal uptake of indoor organics
- 15
- 16 *** Corresponding Author:**
- 17 Environmental and Occupational Health Sciences Institute
- 18 Rutgers University
- 19 Piscataway, NJ 08854, USA
- 20 Phone: (732) 445-2073
- 21 Email: weschlch@umdnj.edu
- 22

23 Table of Contents Art



27 Abstract

28 Transdermal uptake directly from air is a potentially important vet largely overlooked pathway 29 for human exposure to organic vapors indoors. We recently reported (Indoor Air 2012, 22, 356) 30 that transfermal uptake directly from air could be comparable to or larger than intake via 31 inhalation for many semivolatile organic compounds (SVOCs). Here, we extend that analysis to 32 approximately eighty organic compounds that (a) occur commonly indoors and (b) are primarily 33 in the gas-phase rather than being associated with particles. For some compounds, the modeled 34 ratio of dermal-to-inhalation uptake is large. In this group are common parabens, lower 35 molecular weight phthalates, o-phenylphenol, Texanol, ethylene glycol, and α -terpineol. For 36 other compounds, estimated dermal uptakes are small compared to inhalation. Examples include 37 aliphatic hydrocarbons, single ring aromatics, terpenes, chlorinated solvents, formaldehyde, and 38 acrolein. Analysis of published experimental data for human subjects for twenty different 39 organic compounds substantiates these model predictions. However, transdermal uptake rates 40 from air have not been measured for the indoor organics that have the largest modeled ratios of 41 dermal-to-inhalation uptake; for such compounds, the estimates reported here require 42 experimental verification. In accounting for total exposure to indoor organic pollutants and in 43 assessing potential health consequences of such exposures, it is important to consider direct 44 transdermal absorption from air.

45 Introduction

46 Direct uptake of selected organic compounds from air through skin has been demonstrated in many studies conducted over the past half-century. The primary emphasis in these studies has 47 48 been on occupational exposures. Experiments include cases in which the whole body of a human 49 subject was exposed and other cases in which only an arm was exposed. Dutkiewicz and Piotrowski [1] reported that, "... a resting, fully relaxed person absorbs through the skin amounts 50 (of aniline) comparable with those absorbed simultaneously through the respiratory tract." 51 Specifically, they estimated that dermal absorption of aniline vapors accounted for 47-64% of a 52 53 resting person's aggregate aniline intake. Piotrowski [2] more fully described experiments in 54 which both naked and dressed men were exposed to nitrobenzene vapors in a chamber while 55 breathing clean air. He concluded that "about half as much vapour was absorbed through the skin 56 as through the lungs" and that "normal working clothes reduced the absorption by only 20 to 57 30%." Subsequently, Piotrowski [3] conducted similar chamber experiments in which the entire 58 bodies of seven men were exposed to phenol vapors; the dermal absorption rate averaged 70% of 59 the inhalation rate. Kežić et al. [4] exposed only the forearm of five volunteers to vapors of either 60 2-methoxyethanol or 2-ethoxyethanol. Based on the urine concentrations of the metabolites of these glycol ethers, the authors estimated that — for whole body exposure — skin uptake would 61 62 be approximately 120% of inhalation uptake for 2-methoxyethanol and 70% of inhalation uptake 63 for 2-ethoxyethanol. Altogether, we have identified twenty studies published in peer-reviewed 64 archival journals that have measured direct uptake of various organic vapors by human skin in 65 either whole body or arm/hand experiments [1-20]. Most, but not all, of these studies have been reviewed by Rehal and Maibach [21] and by Rauma et al. [22]. 66

67 In assessing human exposure to organic pollutants indoors, inhalation and ingestion of dust 68 are routinely included as exposure pathways. The dermal pathway is frequently assumed to be 69 negligible [23, 24]. When considered, the focus has commonly been on dermal uptake following 70 contact transfer to the skin of a pollutant in dust, on particles and from contaminated surfaces 71 [25-32]. Direct transdermal uptake from air is not routinely considered. Yet the studies outlined 72 in the previous paragraph suggest that, for at least some indoor pollutants, direct dermal uptake 73 from air may occur at rates that are comparable to or larger than inhalation intake. We recently 74 published a critical review of the state of knowledge concerning indoor exposures to semivolatile 75 organic compounds (SVOCs) via dermal pathways [33]. That assessment included predictive 76 equations, based on idealized mass-transport considerations, to estimate the steady rate of 77 transdermal uptake of an SVOC from the gas-phase. We concluded that air-to-skin transdermal 78 uptake was potentially comparable to or larger than inhalation uptake for many SVOCs found 79 indoors. The present paper extends that assessment to volatile organic compounds and includes 80 almost eighty organic compounds that are (a) common in indoor environments and (b) found in 81 air primarily in the gas-phase (rather than being associated with airborne particulate matter). A 82 specific aim of this study is to identify those indoor, gas-phase organic pollutants for which 83 dermal absorption via transport directly from air is potentially significant in relation to the more 84 commonly assessed inhalation exposure. A second objective is to make quantitative comparisons 85 between modeled and measured results for those twenty compounds for which the rate of 86 transdermal permeation directly from the air has been measured in human experiments. The 87 paper's third objective is to examine the physical and chemical attributes that most influence the 88 tendency of an airborne organic pollutant to be transported through air adjacent to the skin, across the stratum corneum and viable epidermis, and ultimately to the blood. Overall, the goals 89

90 of this assessment are to raise awareness of dermal uptake of organic vapors as a route of
91 environmental exposure, to focus research attention on dermal absorption directly from air, and
92 to facilitate inclusion of this pathway in future assessments of total exposure to organic
93 environmental pollutants encountered indoors.

94 Methods

95 **Transdermal permeability coefficient.** The indoor air transdermal permeability coefficient, $k_{p,g}$, is a mass-transfer coefficient that describes the rate of transport of an organic 96 97 compound from bulk air through the boundary layer adjacent to the skin and then from air at the surface of the skin through the epidermis to the dermal capillaries. In the present paper, k_{p_g} is 98 99 estimated for numerous gas-phase organic compounds using a procedure that we outlined 100 previously [33]. Wilschut and ten Berge have reported an analogous approach [34]. 101 The procedure begins with a deterministic model proposed by Mitragotri [35] to calculate 102 the compound's permeability coefficient through the stratum corneum when the vehicle in 103 contact with the skin is water ($k_{p cw}$). We then use a relationship developed by Bunge et al. [36] 104 to estimate B, the ratio of $k_{p_{ew}}$ to the viable epidermis permeability coefficient ($k_{p_{ew}}$) for the 105 compound in question (see equation S2). The parameter B is used to estimate the compound's 106 permeability coefficient through the stratum corneum/viable epidermis composite when the vehicle in contact with skin is water $(k_{p w})$. The permeability coefficient through the stratum 107 108 corneum/viable epidermis composite when the vehicle in contact with the skin is air $(k_{p,b})$ is calculated using Henry's constant (H, expressed in units of (mol/liter) atm⁻¹; note that this 109 110 convention is the inverse of that commonly used in the dermal literature):

111
$$k_{p b} = k_{p w} \times (HRT)$$
(1)

where *R* is the gas constant (0.0821 atm liter mole⁻¹ K⁻¹) and *T* is the skin temperature (305 K = 32 °C). Finally, k_{p_g} is calculated using a resistor-in-series model:

114
$$1/k_{p_g} = 1/v_d + 1/k_{p_b}$$
 (2)

Here, v_d is the mass-transfer coefficient that describes the external transport of a compound from the gas-phase in the core of a room through the boundary layer adjacent to the skin. Throughout the work reported in this paper, we assume that $v_d \sim 6 \text{ m h}^{-1}$ [33]. Further details are provided in section S1 of the *Supporting Information*. The key parameters in calculating k_{p_g} are the organic compound's molecular weight (MW), octanol-water partition coefficient (K_{ow}), and Henry's constant (H). Once k_{p_g} has been estimated, the transdermal flux of an organic compound, J, can be evaluated:

$$J = C_{g} \times k_{p_g}$$
(3)

123 Here, C_{g} is the compound's gas-phase concentration.

124 Several assumptions are implicit in this procedure. The Mitragotri model used to calculate $k_{\rm p}$ cw assumes a simplified one-component lipid system to obtain the required bilayer parameters, 125 avoiding the complexities of the actual multicomponent system (comprising ceramides, fatty 126 127 acids, cholesterol and various other species) that constitutes the lipid bilayer in the stratum 128 corneum. In comparing predictions made with his model against experimental data, Mitragotri 129 reported a mean error of 5% [35]. The model assumes that the organic permeant moves in a 130 stationary frame of lipid molecules; this assumption breaks down for compounds with MW > 131 400 g/mol. With the exception of chlordane (MW = 410 g/mol), the indoor pollutants considered 132 in this paper have MW's less than 400 g/mol. The model assumes that clearance is fast and that the concentration of the permeant in the blood is close to zero. A distributed clearance model 133

[37] would be a better approximation, but computationally more complicated to a degree that is
not justified by the expected improvement in predicted results. Finally, the procedure is based on
a fully hydrated stratum corneum, whereas under typical indoor conditions the stratum corneum
is only partially hydrated. The consequences of this assumption are examined in the *Limitations*subsection of the *Discussion*.

139 **Dermal uptake and inhalation intake.** Uptake of gas-phase organics via the dermal 140 pathway, *D*, is estimated as the product of three terms — C_g , k_{p_g} and the total body surface area 141 (BSA):

142
$$D = C_{g} \times k_{p_{g}} \times BSA$$
(4)

Based in part on the findings of Piotrowski [2], we assume that clothing presents negligible resistance to the transport of organic compounds from air to skin. We further assume that the flux through skin achieves steady state. More precisely, we assume that the time-averaged flux is well modeled as the product of the time-averaged airborne concentration multiplied by a masstransfer coefficient derived for steady-flux conditions. Because of loss processes that may interfere with transdermal transport, such as desquamation, the dermal uptake from air estimated herein represents an upper limit.

150 Intake of gas-phase organics via inhalation, *I*, is estimated as the product of C_g and the 151 volumetric breathing rate, Q_b :

$$152 I = C_{\rm g} \times Q_{\rm b} (5)$$

We assume here that 100% of what is inhaled is absorbed. Hence, the estimated inhalation intakeis also an upper bound.

155 The ratio of dermal uptake to inhalation intake for gas-phase organics (D/I) is then 156 estimated as:

157
$$D/I = k_{\rm p \ g} \times {\rm BSA}/Q_{\rm b} \tag{6}$$

158 For the baseline values that we use for a typical adult, i.e. body surface area (BSA ~ 2 m²) and volumetric breathing rate ($Q_b \sim 0.5 \text{ m}^3 \text{ h}^{-1}$ while at rest), the dermal uptake to inhalation 159 intake ratio, D/I, is simply 4 k_{p_g} when k_{p_g} is expressed in units of m/h. (Units for the parameters 160 161 used in this paper can be found in the *Nomenclature* section of the *Supporting Information*.) 162 Because particles diffuse much more slowly than gases, the gas phase is expected to 163 dominate over the particle phase for dermal absorption of airborne organics. Conversely, for 164 inhalation exposure, the volumetric breathing rate is a limiting process for intake: both particle-165 and gas-phase organics are introduced into the respiratory tract at rates proportional to their 166 respective airborne concentrations. Equation (5) only addresses inhalation intake of gas-phase 167 organics. Hence, the D/I ratios estimated by equation (6) apply only to the gaseous portion of an 168 airborne organic compound. This distinction is unimportant for the indoor pollutants considered 169 in this paper, since they are present primarily in the gas-phase (> 97%; see Table S1). However, 170 the reader is cautioned that equation (6) is inappropriate for organic compounds that have a 171 meaningful fraction of their airborne concentration in the particle phase.

172 Fraction of indoor organic in the gas phase. The values listed in Table S1 for the fraction 173 of an indoor organic pollutant in the gas-phase, $f_g = C_g/(C_g + C_p)$, were estimated as follows:

174
$$f_{\rm g} = C_{\rm g}/(C_{\rm g} + C_{\rm p}) = 1/(1 + (\text{TSP} \times K_{\rm p}))$$
 (7)

where C_p is the airborne concentration of organic in the particle phase, TSP is the average mass concentration of airborne particles, and K_p is the particle/gas-phase partition coefficient for the 177 organic of interest [38]. We have estimated K_p based on the assumption that partitioning of 178 organics into particles is governed primarily by absorption into the condensed-phase organic 179 matter:

180
$$K_{\rm p} = (f_{\rm om} \times K_{\rm og})/\rho_{\rm part}$$
(8)

Here, f_{om} is the fraction of airborne particulate matter that is organic, K_{og} is the octanol/air partition coefficient and ρ_{part} is the airborne particle density [39]. For typical indoor conditions, we assume a temperature of 25 °C, TSP = 20 µg/m³, f_{om} = 0.4 and ρ_{part} = 1 × 10⁶ g/m³ (= 1 g/cm³).

185 **Results**

186 Estimated transdermal uptake of organic vapors. For thirty-three organic compounds 187 with indoor sources, Table 1 lists relevant physical and chemical parameters and calculated 188 overall transdermal permeability coefficients $(k_{p,g})$. Table S1 is an expanded version of Table 1, 189 with data for approximately eighty indoor pollutants and additional columns listing the ratio of 190 stratum corneum permeability to viable epidermis permeability (B), estimated ratios of dermal 191 uptake to inhalation intake (D/I), and predicted fractions in the gas-phase (f_{s}) under typical 192 indoor conditions. Although some of the indoor pollutants in Tables 1 and S1 may be classified 193 as SVOCs, all exist primarily in the gas-phase: gas-phase partitioning is predicted to be greater 194 than 97% in each case for typical indoor conditions. For the first 15 compounds listed in Table 1, 195 $k_{p,g}$ exceeds 2.5 m/h, indicating that direct transdermal absorption of these compounds is 196 anticipated to be an important exposure pathway relative to inhalation intake (the estimated 197 dermal uptake to inhalation intake rate, D/I, exceeds a factor of 10). Noteworthy among these 198 entries are several organic compounds that are frequently found indoors at concentrations larger

199 than 100 ng/m³ [38-43]. These include common parabens, lower molecular weight phthalates, synthetic musks and o-phenylphenol. For the 18 compounds in the lower portion of Table 1, $k_{p g}$ 200 201 is between 0.25 and 2.5 m/h, implying D/I values in the range 1-10. Among the ubiquitous 202 indoor pollutants in this group are nonylphenol, Texanol, α -terpineol, 4-oxopentanal, chlorpyrifos, linalool, and 2-butoxyethanol. Table S1 includes indoor pollutants with k_{p_g} less 203 204 than 0.25 m/h. For compounds (n = 20) with $k_{p g}$ values in the range 0.025-0.25 m/h (D/I ratios 205 of 0.1-1), the dermal pathway is marginally important. This group includes PCB28, PCB52, 206 chlordane, and some aliphatic alcohols (e.g., 1-octen-3-ol, butanol, hexanal and 3-octanol). For compounds (n = 26) in Table S1 with $k_{p,g}$ less than 0.025 m/h, the dermal pathway appears 207 208 unimportant relative to inhalation. This group includes aliphatic hydrocarbons, single ring 209 aromatics, one- or two-carbon chlorinated solvents, formaldehyde, terpenes and isoprene. 210 Empirical evidence supporting estimated transdermal uptakes. What is the basis for

211 confidence that estimates of transdermal uptake parameters reported in Table 1 and Table S1 are 212 approximately correct? For twenty of the listed compounds, published studies [1-20] have measured $k_{p,g}$ or the ratio of dermal uptake to inhalation uptake. Table 2 compares values 213 214 estimated here with published measurement results. (Note: in the case of D/I, when estimated 215 values are compared to measured values, we are assuming that metabolism is equivalent for the 216 dermal and inhalation pathways once the compound enters the blood.) We have calculated the 217 ratio "modeled-to-measured" for each compound in each study. The median value of this ratio is 218 0.7, i.e. within 30% of unity; the interquartile range is 0.3 to 1.2. With the exception of 219 tetrachloroethylene, the modeled values of $k_{p,g}$ are within a factor of five of the measured values 220 in each case, while modeled values of D/I lie within a factor of seven of measured results. For 221 the 14 compounds with modeled dermal to inhalation ratios (D/I) greater than 0.1, the ratios of

222 "modeled-to-measured" results have a median value of 0.96 and range from 0.17 to 6.7. The 223 "modeled-to-measured" outcomes are not strongly biased relative to the expected value of unity: 224 eight of the values are larger than one and six are smaller. We have plotted log (measured) versus log (modeled) results for both $k_{p,g}$ (Figure S1) and D/I (Figure S2). The relationships are 225 approximately linear, with $R^2 = 0.88$ for $k_{p,g}$ (n = 17; MW = 76-166 g/mol), and $R^2 = 0.84$ for D/I226 (n = 27; MW = 72-166 g/mol). Given anticipated subject-to-subject variation in the dermal 227 228 uptake of organic vapors [5, 6] along with substantial uncertainties in the estimated properties of 229 the compounds, these comparisons support a finding that the modeling approach provides 230 reasonable estimates of transdermal uptake rates. An important caveat for interpreting these 231 modeled-to-measured comparisons is that transdermal permeation from the gas-phase has not been experimentally studied for those compounds in Table 1 with the highest modeled k_{pg} 232 233 values. Additional experimental studies are warranted to test the model predictions for 234 compounds predicted to have high transdermal permeability coefficients. 235 Although we are aware of no studies that have directly measured uptake of lower molecular 236 weight phthalate vapors via dermal absorption from air, two studies [44,45] have measured 237 transdermal uptake of diethyl phthalate (DEP) and di(n-butyl) phthalate (DnBP) when these 238 compounds were contained in a cream (each at 2% by mass) that was directly applied to human skin at a surface coverage of 2 mg/cm². Based on metabolites measured in blood [44], the 239 reported maximum measured flux was 2000 μ g m⁻² h⁻¹ for DEP and 52 μ g m⁻² h⁻¹ for DnBP. 240 Based on metabolites measured in urine [45], the maximum measured flux was 1500 μ g m⁻² h⁻¹ 241 for DEP and 450 μ g m⁻² h⁻¹ for DnBP. For air saturated with DEP and DnBP vapors, we 242 calculate that the maximum fluxes for direct dermal absorption are 4600 μ g m⁻² h⁻¹ for DEP and 243 185 µg m⁻² h⁻¹ for DnBP (see Supporting Information (S2) for details). A comparison of these 244

modeled fluxes from air with the measured fluxes for absorption from a 2% cream indicates that our estimated values of k_{p_g} for DEP and DnBP are plausible.

247 **Discussion**

248 Attributes of organics likely to be dermally absorbed rapidly from air. The transdermal 249 permeation coefficient, $k_{p,g}$, for an organic compound depends primarily on its molecular weight, 250 MW, and its stratum corneum/air partition coefficient, $K_{sc g}$ [33]. Where a compound lies in a plot of MW versus log (K_{sc} g) can provide a rapid visual indication of the potential importance of 251 252 direct dermal uptake from air. Figure 1 summarizes such information for all of the compounds 253 in Table S1 with MWs less than 350 g/mol. Different symbols indicate the magnitude of $k_{p,g}$ on a decadal scale. For compounds with k_{p_g} larger than 0.99 m/h (as denoted by purple diamonds), 254 255 the dermal intake may exceed inhalation intake by a factor of four or more. For compounds with k_{p_g} between 0.1 and 0.99 m/h (blue circles), the ratio of dermal to inhalation intake is estimated 256 to be between 0.4 and 4. For compounds with $k_{p,g}$ between 0.01 and 0.099 m/h (green triangles), 257 258 the dermal to inhalation intake ratio is estimated to lie in the range 0.04 and 0.4. For other 259 compounds (orange squares and red diamonds), dermal intake can normally be neglected when 260 estimating indoor exposures. In summary, transdermal uptake directly from air is progressively 261 more significant, relative to inhalation, for organic compounds with larger values of $K_{sc g}$ and among compounds with similar K_{sc_g} values — for those with smaller molecular weights. 262 263 In using the approach presented here to assess the potential importance of the dermal 264 pathway, one must also consider the exposure time necessary for a steady-state transdermal flux 265 model to reasonably approximate reality. The time that an individual spends in a given indoor 266 environment, the frequency of bathing (and its effectiveness in removing skin absorbed organic 267 contaminants), and the time scale for shedding the stratum corneum each pose a constraint on the

268 time available to achieve steady state. In general, for larger values of MW and K_{sc} , longer times 269 are required to reach steady state (see Supporting Information S3 and Table S2). For compounds 270 with MW larger than 225 g/mol and log (K_{sc} g) larger than 7, an interval longer than a day 271 appears necessary to legitimize the use of a steady-flux, two-resistor model to accurately 272 represent transport from the gas-phase through the skin. For intervals shorter than the time 273 needed to reach steady state, a transient model such as that presented in Gong et al. [46] should 274 yield better estimates of the transdermal permeation of organic vapors. The time required to 275 establish steady flux can be an important consideration for accurately modeling the air-mediated 276 dermal uptake for many SVOCs; however, it is not a limitation for most VOCs. 277 Sensitivity to key parameters. The accuracy of the estimates for $k_{p g}$ reported in Tables 1 278 and S1 depends not only on the fidelity of the transdermal permeation model but also on the 279 accuracy of key input parameters, i.e., the octanol-water partition coefficient, K_{ow} , and Henry's 280 constant, H. In addition to limitations in model accuracy [35], determinations of these

thermodynamic parameters may be prone to large errors. In the present study, these parameters

have been calculated using the chemical property estimation software SPARC (v4.6). Using

other software (e.g., EPA's EpiSuite), the calculated values of K_{ow} and H for certain compounds

differ from the SPARC values by an order of magnitude or more [47]. We have assessed the

sensitivity of k_{p_g} to an order-of-magnitude change in either direction in these key parameters.

For the full complement of compounds addressed in this study (Table S1), the results are plotted

in Figures S4 and S5, respectively showing sensitivity to K_{ow} and H. In the case of K_{ow} ,

substituting values that are an order of magnitude smaller or larger than the baseline value

results, on average, in a factor of 0.3 or 3.7 change in $k_{p,g}$. The permeability coefficient is more

sensitive to *H*. Substituting values for *H* that are an order of magnitude smaller or larger than the

291 baseline value results, on average, in a factor of 6.5 or 0.2 change in $k_{p g}$. Nevertheless, it is 292 reassuring that for the organic compounds in Table 2, with the exception of tetrachloroethylene, 293 the ratio of modeled-to-measured values of $k_{\rm p}$ g spans a much narrower range, from 0.2 to 3.7, as 294 compared to the measured $k_{p,g}$ values, which span a factor of 3600. Furthermore, there is 295 reasonable agreement between the estimates in Tables 1 and S1 and estimates for a subset of the 296 same compounds (n = 36) as predicted with ten Berge's *SkinPermMultiScen v1.1* model [48], for 297 which the thermodynamic parameters were calculated using EpiSuite rather than SPARC (see 298 Supporting Information (S4)).

299 Limitations. The basis for the analysis used in this paper is a model proposed by Mitragotri 300 [35] to calculate an organic compound's permeability coefficient through the stratum corneum 301 when the vehicle in contact with skin is water. This model is most applicable to a fully hydrated 302 stratum corneum. However, in the case of dermal absorption from indoor air, we anticipate that 303 the stratum corneum will be only partially hydrated. There are procedures for calculating 304 permeability coefficients when the stratum corneum is partially hydrated (e.g., Table 4 in [49]), 305 but the calculations are more complicated than the relatively simple equation derived in 306 Mitragotri's model. For eighteen of the indoor pollutants considered in this paper, most with relatively large values of $k_{p,g}$, Table S3 compares $k_{p,g}$ values calculated using the procedure in 307 308 the *Methods* section for fully hydrated stratum corneum with values calculated using the 309 procedure outlined in Wang et al. [49] for partially hydrated stratum corneum. On average, the 310 values calculated assuming a partially hydrated stratum corneum are about two thirds those 311 calculated assuming a fully hydrated stratum corneum. This factor of 2/3 should be considered in the context of the more than five-order-of-magnitude range for $k_{p,g}$ revealed in Table S1 – i.e., 312 313 from 0.00001 to 6 m/h. For initial estimates of the relative contribution of dermal uptake of

314 organic pollutants from air compared to uptake via other exposure pathways, the procedure 315 outlined in this paper should be adequate. However, the reader is cautioned that the procedure 316 used in this paper to estimate dermal uptake from air is likely less accurate than estimates 317 obtained by treating the stratum corneum as only partially hydrated. 318 The estimates for dermal uptake presented here do not account for the presence of enzymes 319 in the skin that metabolize certain compounds. For example, di(n-butyl) phthalate is partially 320 hydrolyzed by esterases during penetration through human skin [50]. The reader is cautioned to 321 be mindful of the potential for pollutant metabolism in the skin; inclusion of this process is 322 beyond the scope of the present assessment. 323 Still another factor to consider is ionization of absorbed organics [33]. Compounds that are 324 acidic or basic (e.g., 2,4-D, pentachlorophenol and nicotine) can exist in both neutral and ionized

325 forms in skin-surface films. Only the neutral form is expected to permeate skin rapidly. For 326 relevant species, acidic or basic ionizing reactions can substantially increase the number of 327 molecules that must be transported from the gas-phase to skin-surface films to achieve a steady 328 balance between the neutral species at the skin surface and its gas-phase counterpart. A 329 consequence of this larger capacity of skin-surface films is a correspondingly longer time to 330 reach steady state for such compounds. Skin pH tends to be the range of 5 to 6 [51]; however, the 331 extent to which small organic molecules ionize in skin-surface films is not well known. Given 332 the potential importance of transdermal permeation as an exposure pathway for certain acidic or 333 basic organic gases, the topic of species ionization in skin surface films warrants further study. 334 Implications. The compounds in Figure 1 have been identified in indoor air and settled dust 335 [38-43] and most of them, or their metabolites, have been identified in human blood or urine [52]. When investigators attempt to connect the levels of an indoor pollutant measured in various 336

337 indoor media with the levels of that pollutant (or its metabolites) measured in blood or urine, the 338 focus has been on inhalation and dust ingestion. If the dermal pathway is considered, the 339 pollutant is commonly assumed to have reached the skin surface via contact with dust or 340 contaminated surfaces [25-32], rather than being transported to skin directly via the air. Yet for 341 roughly a third of the compounds in Figure 1 — including parabens, lower molecular weight 342 phthalate esters, terpene alcohols and Texanol — direct dermal uptake from air appears to occur 343 at rates that are comparable to or larger than inhalation intake. A primary intent in reporting this 344 work is to raise awareness, to promote further measurements, and to facilitate inclusion of 345 transdermal uptake directly from air when researchers and practitioners assess an individual's 346 total exposure to organic pollutants in indoor environments.

347 Acknowledgements

We thank John Kissel and Wil ten Berge for useful comments during a workshop on dermal
absorption at the 2012 meeting of the International Society of Exposure Science. WWN's efforts
on this research were funded in part by the Republic of Singapore's National Research
Foundation through a grant to the Berkeley Education Alliance for Research in Singapore
(BEARS) for the Singapore-Berkeley Building Efficiency and Sustainability in the Tropics
(SinBerBEST) Program. BEARS has been established by the University of California, Berkeley
as a center for intellectual excellence in research and education in Singapore.

355 Supporting Information

356 S1 – Calculating transdermal permeability coefficients; S2 – Calculating maximum flux for DEP

- and DnBP vapors; S3 Time scale to achieve steady state; S4 Comparison with *SkinPerm*
- 358 model predictions; S5 Nomenclature (for primary paper and for supporting information); Table

359	S1 – For selected indoor pollutants, MW, K_{sc_g} , K_{ow} , K_{og} , H , B , k_{p_b} , k_{p_g} , D/I and f_g ; Table S2 –
360	For selected indoor pollutants, MW, K_{og} , k_{p_b} and τ_s ; Table S3 – Comparison of k_{p_g} calculated
361	for fully and partially hydrated stratum corneum; Figure S1 – Measured versus modeled values
362	for k_{p_g} ; Figure S2 – Measured versus modeled values for D/I ; Figure S3 – For selected indoor
363	pollutants, log (K_{sc_g}) versus log (K_{og}); Figure S4 – Sensitivity of k_{p_g} to an order of magnitude
364	change in K_{ow} ; Figure S5 – Sensitivity of k_{p_g} to an order of magnitude change in H ; Figure S6 –
365	Comparisons between k_{p_g} estimated using the approach presented in the present paper and that
366	presented by ten Berge (SkinPermMultiScen v1.1).

367 **References**

- Dutkiewicz, T.; Piotrowski, J. Experimental investigations on the quantitative estimation of
 aniline absorption in man. *Pure Appl. Chem.* 1961, *3*, 319-324.
- Piotrowski, J. Further investigations on the evaluation of exposure to nitrobenzene. *Br. J. Ind. Med.* 1967, *24*, 60-65.
- 372 3. Piotrowski, J. K. Evaluation of exposure to phenol: absorption of phenol vapour in the lungs
 373 and through the skin and excretion of phenol in urine. *Br. J. Ind. Med.* 1971, *28*, 172-178.
- 4. Kežić, S.; Mahieu, K.; Monster, A. C.; de Wolff, F. A. Dermal absorption of vaporous and
- 375 liquid 2-methoxyethanol and 2-ethoxyethanol in volunteers. *Occup. Environ. Med.* **1997**, *54*,
- 376 38-43.
- 5. Nomiyama, T.; Omae, K.; Ishizuka, C.; Yamauchi, T.; Kawasumi, Y.; Yamada, K.; Endoh,
- H.; Sakurai, H. Dermal absorption of N,N-dimethylacetamide in human volunteers. *Int. Arch.*
- 379 *Occup. Environ. Health* **2000**, *73*, 121-126.
- 380 6. Nomiyama, T.; Nakashima, H.; Chen, L. L.; Tanaka, S.; Miyauchi, H.; Yamauchi, T.;
- 381 Sakurai, H.; Omae, K. N,N-dimethylformamide: significance of dermal absorption and

- 382 adjustment method for urinary N-methylformamide concentration as a biological exposure
- 383 item. Int. Arch. Occup. Environ. Health 2001, 74, 224-228.
- Johanson, G.; Boman, A. Percutaneous absorption of 2-butoxyethanol vapour in human
 subjects. *Br. J. Ind. Med.* 1991, *48*, 788-792.
- 386 8. Corley, R. A.; Markham, D. A.; Banks, C.; Delorme, P.; Masterman, A.; Houle, J. M.
- 387 Physiologically based pharmacokinetics and the dermal absorption of 2-butoxyethanol vapor
- 388 by humans. *Fundam. Appl. Toxicol.* **1997**, *39*, 120-130.
- 389 9. Maxfield, M. E.; Barnes, J. R.; Azar, A.; Trochimowicz, H. T. Urinary excretion of
- 390 metabolite following experimental human exposures to DMF or to DMAC. J. Occup. Med.
- **1975**, *17*, 506-511.
- 392 10. Mráz, J.; Nohová, H. Percutaneous absorption of N,N-dimethylformamide in humans. *Int.* 393 *Arch. Occup. Environ. Health* 1992, *64*, 79-83.
- 394 11. Flek, J.; Šedivec, V. The absorption, metabolism and excretion of furfural in man. *Int. Arch.*395 *Occup. Environ. Health* 1978, *41*, 159-168.
- 396 12. Kežić, S.; Monster, A. C.; Krüse, J.; Verberk, M. M. Skin absorption of some vaporous
 397 solvents in volunteers. *Int. Arch. Occup. Environ. Health* 2000, *73*, 415-422.
- 398 13. Brooke, I.; Cocker, J.; Delic, J. I.; Payne, M.; Jones, K.; Gregg, N. C.; Dyne, D. Dermal
- 399 uptake of solvents from the vapour phase: an experimental study in humans. *Ann. Occup.*
- 400 *Hyg.* **1998**, *42*, 531-540.
- 401 14. Shih, T.-S.; Wang, P.-Y.; Chen, C.-Y.; Smith, T. J.; Hu, Y.-P. Measurement of percutaneous
- 402 uptake of 2-methoxy ethanol vapor in humans. J. Occup. Environ. Med. 2000, 42, 475-482.

- 403 15. Bader, M.; Wrbitzky, R.; Blaszkewicz, M.; Schäper, M.; van Thriel, C. Human volunteer
- study on the inhalational and dermal absorption of N-methyl-2-pyrrolidone (NMP) from the
 vapour phase. *Arch. Toxicol.* 2008, *82*, 13-20.
- 406 16. Riihimäki, V.; Pfäffli, P. Percutaneous absorption of solvent vapors in man. *Scand. J. Work*407 *Environ. Health* 1978, *4*, 73-85.
- 408 17. Wieczorek, H. Evaluation of low exposure to styrene. II. Dermal absorption of styrene
- 409 vapours in humans under experimental conditions. *Int. Arch. Occup. Environ. Health* 1985,
 410 57, 71-75.
- 411 18. Giardino, N. J.; Gordon, S. M.; Brinkman, M. C.; Callahan, P. J.; Kenny, D. V. Real-time
- 412 breath analysis of vapor phase uptake of 1,1,1-trichloroethane through the forearm:
- 413 implications for daily absorbed dose of volatile organic compounds at work. *Appl. Occup.*
- 414 Environ. Hyg. 1999, 14, 719-727.
- 415 19. Loizou, G. D.; Jones, K.; Akrill, P.; Dyne, D.; Cocker, J. Estimation of the dermal absorption
- 416 of *m*-xylene vapor in humans using breath sampling and physiologically based
- 417 pharmacokinetic analysis. *Toxicol. Sci.* **1999**, *48*, 170-179.
- 418 20. Kežić, S.; Janmaat, A.; Krüse, J.; Monster, A. C.; Verberk, M. M. Percutaneous absorption of
- 419 *m*-xylene vapour in volunteers during pre-steady and steady state. *Toxicol. Lett.* **2004**, *153*,
- 420 273-282.
- 421 21. Rehal, B.; Maibach, H. Percutaneous absorption of vapors in human skin. *Cutan. Ocular*422 *Toxicol.* 2011, *30*, 87-91.
- 423 22. Rauma, M.; Boman, A.; Johanson, G. Predicting the absorption of chemical vapours. Adv.
- 424 Drug Deliv. Rev. **2013**, *65*, 306-314.

425	23. Wilson, N. K.; Chuang, J. C.; Morgan, M. K.; Lordo, R. A.; Sheldon, L. S. An observational
426	study of the potential exposures of preschool children to pentachlorophenol, bisphenol-A,
427	and nonylphenol at home and daycare. Environ. Res. 2007, 103, 9-20.
428	24. Wilson, N. K.; Strauss, W. J.; Iroz-Elardo, N.; Chuang, J. C. Exposures of preschool children
429	to chlorpyrifos, diazinon, pentachlorophenol, and 2,4-dichlorophenoxyacetic acid over 3
430	years from 2003 to 2005: A longitudinal model. J. Expos. Sci. Env. Epid. 2010, 20, 546-558.
431	25. Fenske, R. A.; Black, K. G.; Elkner, K. P.; Lee, C. L.; Methner, M. M.; Soto, R. Potential
432	exposure and health risks of infants following indoor residential pesticide applications. Am.
433	J. Public Health 1990, 80, 689-693.
434	26. Lioy, P. J. Assessing total human exposure to contaminants. A multidisciplinary approach.
435	Environ. Sci. Technol. 1990, 24, 938-945.
436	27. U.S. EPA. 2007. Dermal Exposure Assessment: A Summary of EPA Approaches. U.S.
437	Environmental Protection Agency, Washington, DC, EPA/600/R-07/040F. Available:
438	http://cfpub.epa.gov/ncea/cfm/recordisplay.cfm?deid=183584#Download [accessed 5 Dec
439	2013].
440	28. Gurunathan, S.; Robson, M.; Freeman, N.; Buckley, B.; Roy, A.; Meyer, R.; Bukowski, J.;

- Lioy, P. J., Accumulation of chlorpyrifos on residential surfaces and toys accessible to
 children. *Environ. Health Perspect.* 1998, *106*, 9-16.37.
- 443 29. Wormuth, M.; Scheringer, M.; Vollenweider, M.; Hungerbuhler, K., What are the sources of
- 444 exposure to eight frequently used phthalic acid esters in Europeans? *Risk Analysis* 2006, *26*,
- 445 803-824.

- 446 30. Cohen-Hubal, E. A.; Egeghy, P. P.; Leovic, K. W.; Akland, G. G., Measuring potential
- 447 dermal transfer of a pesticide to children in a child care center. *Environ. Health Perspect.*

2006, *114*, 264-269.

- 449 31. Cohen-Hubal, E. A.; Nishioka, M. G.; Ivancic, W. A.; Morara, M.; Egeghy, P. P., Comparing
- 450 surface residue transfer efficiencies to hands using polar and nonpolar fluorescent tracers.
- 451 *Environ. Sci. Technol.* **2008**, *42*, 934-939.
- 452 32. Guo, Y.; Kannan, K., Comparative assessment of human exposure to phthalate esters from
 453 house dust in China and the United States. *Environ. Sci. Technol.* 2011, *45*, 3788-3794.
- 454 33. Weschler, C. J.; Nazaroff, W. W. SVOC exposure indoors: fresh look at dermal pathways.
- 455 *Indoor Air* **2012**, *22*, 356-377.
- 456 34. Wilschut, A.; ten Berge, W. F. Two mathematical skin permeation models for vapours. In
- 457 Prediction of Percutaneous Penetration, Brain, K. R.; James, V. J.; Walters, K. A., Eds. STS
- 458 Publishing Ltd: Cardiff, UK, 1996; Vol. 4b, pp 182-185.
- 459 35. Mitragotri, S. A theoretical analysis of permeation of small hydrophobic solutes across the
 460 stratum corneum based on scaled particle theory. *J. Pharm. Sci.* 2002, *91*, 744-752.
- 461 36. Bunge, A. L.; Cleek, R. L.; Vecchia, B. E. A new method for estimating dermal absorption
- 462 from chemical exposure. 3. Compared with steady-state methods for prediction and data
- 463 analysis. *Pharm. Res.* **1995**, *12*, 972-982.
- 464 37. Kretsos, K.; Kasting, G. B. A geometrical model of dermal capillary clearance. *Math. Biosci.*465 2007, *208*, 430-453.
- 466 38. Weschler, C. J.; Nazaroff, W. W. SVOC partitioning between the gas phase and settled dust
- 467 indoors. *Atmos. Environ.* **2010**, *44*, 3609-3620.

- 468 39. Weschler, C. J.; Nazaroff, W. W. Semivolatile organic compounds in indoor environments.
 469 *Atmos. Environ.* 2008, *42*, 9018-9040.
- 470 40. Fromme, H.; Lahrz, T.; Piloty, M.; Gebhart, H.; Oddoy, A.; Rüden, H. Occurrence of
- 471 phthalates and musk fragrances in indoor air and dust from apartments and kindergartens in
- 472 Berlin (Germany). *Indoor Air* **2004**, *14*, 188-195.
- 473 41. Logue, J. M.; McKone, T. E.; Sherman, M. H.; Singer, B. C. Hazard assessment of chemical
 474 air contaminants measured in residences. *Indoor Air* 2011, *21*, 92-109.
- 475 42. Liu, N.; Shi, Y.; Xu, L.; Li, W.; Cai, Y. Occupational exposure to synthetic musks in
- barbershops, compared with the common exposure in the dormitories and households.
- 477 *Chemosphere* **2013**, *93*, 1804-1810.
- 478 43. Zhu, J.; Wong, S. L.; Cakmak, S. Nationally representative levels of selected volatile organic
 479 compounds in Canadian residential indoor air: population-based survey. *Environ. Sci.*
- 480 *Technol.* **2013**, *47*, 13276-13283.
- 481 44. Janjua, N. R.; Mortensen, G. K.; Andersson, A.-M.; Kongshoj, B.; Skakkebaek, N. E.; Wulf,
- 482 H. C. Systemic uptake of diethyl phthalate, dibutyl phthalate, and butyl paraben following
- 483 whole-body topical application and reproductive and thyroid hormone levels in humans.
- 484 Environ. Sci. Technol. 2007, 41, 5564-5570.
- 485 45. Janjua, N. R.; Frederiksen, H.; Skakkebaek, N. E.; Wulf, H. C.; Andersson, A.-M. Urinary
- 486 excretion of phthalates and paraben after repeated whole-body topical application in humans.
- 487 Int. J. Androl. 2008, 31, 118-130.
- 488 46. Gong, M.; Zhang, Y.; Weschler, C.J. Predicting dermal absorption of gas-phase chemicals:
- 489 transient model development, evaluation and application. *Indoor Air* **2014**, *24*, in press: doi
- 490 10.1111/ina.12079.

- 491 47. Schossler, P.; Schripp, T.; Salthammer, T.; Bahadir, M. Beyond phthalates: Gas phase
- 492 concentrations and modeled gas/particle distribution of modern plasticizers. *Sci. Total*
- 493 *Environ.* **2011**, *409*, 4031-4038.
- 494 48. Tibaldi, R.; ten Berge, W.; Drolet, D. Dermal absorption of chemicals: estimation by IH
- 495 SkinPerm, J. Occup. Environ. Hyg. 2014, 11, 19-31.
- 496 49. Wang, T. F.; Kasting, G. B.; Nitsche, J. M. A multiphase microscopic diffusion model for
- 497 stratum corneum permeability. II. Estimation of physicochemical parameters, and application
- to a large permeability database. J. Pharm. Sci. 2007, 96, 3024-3051.
- 499 50. Beydon, D.; Payan, J.-P.; Grandclaude, M.-C. Comparison of percutaneous absorption and
- 500 metabolism of di-*n*-butylphthalate in various species. *Toxicol. in Vitro* **2010**, *24*, 71-78.
- 501 51. Fluhr, J. W.; Darlenski, R.; Lachmann, N.; Baudouin, C.; Msika, P.; De Belilovsky, C.;
- 502 Hachem, J.-P. Infant epidermal skin physiology: adaptation after birth. *Br. J. Dermatol.*
- **2012**, *166*, 483-490.
- 504 52. CDC (Centers for Disease Control and Prevention). 2009. National Report on Human
- 505 Exposure to Environmental Chemicals: Fourth Report. Available:
- 506 http://www.cdc.gov/exposurereport [accessed 5 Dec 2013].
- 507

508

510 Figure Caption

- 511 **Figure 1.** Dependence of indoor air transdermal permeability coefficient, k_{p_g} on molecular
- 512 weight (MW) and stratum corneum to gas-phase partitioning coefficient (K_{sc} g) for numerous
- organic compounds commonly found in indoor air. Different symbols denote magnitude of $k_{\rm p~g}$
- 514 on a decade-by-decade scale (see legend). Abbreviations: 1,1,1-TCE 1,1,1-trichloroethane; 2-
- 515 BE 2-butoxyethanol; 2-EE 2-ethoxyethanol; 2,4-D 2,4-dichlorophenoxyacetic acid; 4-OPA
- 516 4-oxopentanal; BHT butylated hydroxy toluene; DCB dichlorobenzene; DEP –
- 517 diethylphthalate; DiBP di(isobutyl)phthalate; DMA dimethylacetamide; DMF -
- 518 dimethylformamide; DMP dimethylphthalate; DnBP di(n-butyl)phthalate; MEK methyl
- 519 ethyl ketone; NMP n-methyl-2-pyrrolidone; PCB28 2,4,4'-trichlorobiphenyl; PCB52 –
- 520 2,2',5,5'-tetrachlorobiphenyl; PCP pentachlorophenol; PGME 1-methoxypropan-2-ol; THF –
- 521 tetrahydrofuran.

Table 1. For selected organics that are found indoors and exist primarily in the gas phase, relevant physical and chemical properties (MW, K_{ow} , H, K_{sc_g}) and overall permeability coefficients (k_{p_g}), with compounds rank ordered according to k_{p_g} .

Compound	MW	$\log(K_{\rm ow})^{\rm a}$	$\log(H)^{a}$	$\log(K_{sc_g})$	k_{p_g}
	g/mol	[—]	(mol/liter)	[—] ^a	m/h
			atm ⁻¹		
diethanolamine	105	-2.5	8.68	8.2	6.0
2,4-D ^b	221	2.9	5.16	8.7	5.8
butyl paraben	194	3.4	4.10	8.0	5.4
propyl paraben	180	2.8	4.22	7.7	5.2
ethyl paraben	166	2.2	4.39	7.4	4.9
di(n-butyl) phthalate	278	4.6	3.61	8.4	4.8
methyl paraben	152	1.5	4.61	7.1	4.7
o-phenylphenol	170	3.5	3.42	7.4	4.6
di(isobutyl) phthalate	278	4.2	3.76	8.3	4.6
nicotine ^b	162	2.0	4.31	7.2	4.4
diethyl phthalate	222	2.6	4.06	7.3	3.4
diazinon	304	4.9	3.10	8.1	3.3
dimethyl phthalate	194	1.5	4.45	6.9	2.9
Galaxolide (HHCB)	258	4.6	2.85	7.6	2.8
Tonalide (AHTN)	258	5.0	2.58	7.7	2.6
monoethanolamine	61	-1.8	5.32	5.4	2.5
nonylphenol	220	6.2	2.00	8.0	2.3
Phantolide	244	4.8	2.35	7.3	1.8
pentachlorophenol ^b	266	4.9	2.30	7.3	1.6
Texanol	216	2.4	3.46	6.7	1.4
ethylene glycol	62	-1.4	4.62	5.0	1.2
hexyl cinnamal	216	5.0	1.86	6.9	1.2
n-methyl-2-pyrrolidone	99	0.063	3.97	5.4	1.2
α-terpineol	154	2.5	2.72	6.0	0.98
phenol	94	1.5	2.62	5.2	0.70
eugenol	164	3.2	2.12	5.9	0.60
4-oxopentanal	100	0.10	3.57	5.0	0.56
chlorpyrifos	351	6.4	1.39	7.5	0.41
linalool	154	3.2	1.85	5.6	0.40
BHT	220	4.7	1.44	6.3	0.38
2-butoxyethanol	118	1.1	2.78	5.0	0.33
dimethylacetamide	87	-0.18	3.37	4.6	0.32
p-tert-bucinal	204	4.0	1.52	5.9	0.26

^a Computed for T = 32 °C. ^b Compound assumed nonionized. Abbreviations: 2,4-D – 2,4dichlorophenoxyacetic acid; BHT – butylated hydroxy toluene

Compound	Modeled	Measured	Modeled/	Modeled	Measured	Modeled/	Ref. ^a
•	$k_{\rm p g}$	$k_{p g}$	Measured	D/I	D/I	Measured	
	m/h	m/h	$k_{p g} []$	[—]	[—]	D / I [—]	
aniline	0.21			0.84	0.9-1.8	0.70 ^b	1
2-butoxyethanol	0.33			1.3	2.4-3.8	0.42 ^b	7
2-butoxyethanol	0.33			1.3	0.18-0.37	4.7 ^b	8
dimethylacetamide	0.32			1.3	0.43	3.0	9
dimethylacetamide	0.32			1.3	0.68 ^c	1.9	5
dimethylformamide	0.081			0.33	0.16-0.64	0.83 ^b	9
dimethylformamide	0.081			0.33	0.20	1.65	10
dimethylformamide	0.081			0.33	0.68 ^d	0.49	6
2-ethoxyethanol	0.19	0.19	1.0	0.74	0.72	1.0	4
furfural	0.14			0.56	0.2-0.3	2.2 ^b	11
hexane	0.000029	0.00013	0.22	0.00012			12
1-methoxy-2-	0.13			0.54	0.044-0.11 ^e	7.0 ^b	13
propanol							
2-methoxyethanol	0.14	0.36	0.39	0.56	1.2	0.47	4
2-methoxyethanol	0.14	0.14-0.18	0.88 ^b	0.56			14
methyl ethyl ketone	0.0075			0.030	0.032-0.040 ^e	0.83 ^b	13
n-methyl-2-	1.2			4.8	0.72	6.7	15
pyrrolidone							
nitrobenzene	0.033			0.13	0.50	0.26	2
phenol	0.70	0.19	3.7	2.8	0.70	4.0	3
styrene	0.0025	0.0037	0.68	0.010	0.019	0.53	16
styrene	0.0025	0.012	0.21	0.010	0.052	0.19	17
tetrachloroethylene	0.00008	0.0017	0.05	0.0003	0.011	0.03	16
tetrachloroethylene	0.00008	0.00054	0.15	0.0003			12
tetrahydrofuran	0.0056			0.022	0.016-0.050 ^e	0.67 ^b	13
toluene	0.0010	0.0019	0.53	0.0038	0.009	0.42	16
toluene	0.0010			0.0038	0.018	0.21	13
toluene	0.0010	0.0014	0.71	0.0038			12
1,1,1-	0.00016	0.0001	1.6	0.00065	0.0008	0.81	16
trichloroethane							
1,1,1-	0.00016			0.00065	0.001	2.6	18
trichloroethane							
1,1,1-	0.00016	0.00021	0.76	0.00065			12
trichloroethane							
trichloroethylene	0.00009	0.00049	0.18	0.00036			12
m-xylene	0.0014	0.0026	0.54	0.0056	0.013-0.014	0.47 ^b	16
xylene ^f	0.0014			0.0056	0.013-0.026 ^e	0.32 ^b	13
m-xylene	0.0014			0.0056	0.018	0.35	19
m-xylene	0.0014	0.0012	1.2	0.0056			12
m-xylene	0.0014	0.00062	2.3	0.0056			20

Table 2. Comparisons between modeled and measured values for either the transdermal permeability coefficient (k_{p_g}) or the ratio of dermal to inhalation intake (D/I).

^a Cited studies exposed whole body or arm of humans to vapors of the listed chemicals. ^b Midpoints of the measured values are used to compute the ratio. ^c Ten subjects; range: 0.15-2.7; see Figure 2 of Nomiyama et al., 2000 [5]. ^d Thirteen subjects; range: 0.28-3.7; see Figure 2 of Nomiyama et al., 2001 [6]. ^e Uptake assessed by monitoring compound or metabolite in blood, breath and urine following exposure. Tabulated ranges are mean values for blood assessment, breath assessment and urine assessment. ^f Unspecified isomer(s).