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Head-to-head intra-individual comparison of biodistribution and tumor uptake of ^{68}Ga -FAPI and ^{18}F -FDG PET/CT in cancer patients

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Abstract

Purpose FAPI ligands (fibroblast activation protein inhibitor), a novel class of radiotracers for PET/CT imaging, demonstrated in previous studies rapid and high tumor uptake. The purpose of this study is the head-to-head intra-individual comparison of ^{68}Ga -FAPI versus standard-of-care ^{18}F -FDG in PET/CT in organ biodistribution and tumor uptake in patients with various cancers.

Material and Methods This international retrospective multicenter analysis included PET/CT data from 71 patients from 6 centers who underwent both ^{68}Ga -FAPI and ^{18}F -FDG PET/CT within a median time interval of 10 days (range 1–89 days). Volumes of interest (VOIs) were manually drawn in normal organs and tumor lesions to quantify tracer uptake by SUVmax and SUVmean. Furthermore, tumor-to-background ratios (TBR) were generated (SUVmax tumor/ SUVmax organ).

Results A total of 71 patients were studied of, which 28 were female and 43 male (median age 60). In 41 of 71 patients, the primary tumor was present. Forty-three of 71 patients exhibited 162 metastatic lesions. ^{68}Ga -FAPI uptake in primary tumors and metastases was comparable to ^{18}F -FDG in most cases. The SUVmax was significantly lower for ^{68}Ga -FAPI than ^{18}F -FDG in background tissues such as the brain, oral mucosa, myocardium, blood pool, liver, pancreas, and colon. Thus, ^{68}Ga -FAPI TBRs were significantly higher than ^{18}F -FDG TBRs in some sites, including liver and bone metastases.

Conclusion Quantitative tumor uptake is comparable between ^{68}Ga -FAPI and ^{18}F -FDG, but lower background uptake in most normal organs results in equal or higher TBRs for ^{68}Ga -FAPI. Thus, ^{68}Ga -FAPI PET/CT may yield improved diagnostic information in various cancers and especially in tumor locations with high physiological ^{18}F -FDG uptake.

Keywords FAPI PET/CT · FDG PET/CT · Cancer-associated fibroblast · Various cancer diseases

Introduction

Altered metabolism is one of the hallmarks of cancer [1]. The metabolic switch to utilize glucose (often equated to the Warburg effect) has been successfully exploited by ^{18}F -FDG PET imaging since the 1980s. ^{18}F -FDG is the most frequently used tracer for oncological PET imaging. However, its known limitations are high uptake in many

normal tissues, low glucose transporter and hexokinase activity in certain tumor types, and a lack of specificity [2, 3].

PET imaging of the fibroblast activation protein (FAP) expression in cancer was recently introduced [4, 5]. The first generation of radiolabeled FAP inhibitors (FAPI) is peptidomimetic quinoline derivatives that bind with high affinity to FAP expressed on cancer-associated fibroblasts (CAFs). CAFs are a key component of the tumor stroma or tumor microenvironment. The tumor stroma consists of a variety of benign cells which interact with the tumor cells to promote growth, invasion, and metastasis. However, these cancer-associated cells, which are thought to promote tumor growth, often express unique receptors not found in the same cells outside of the tumor microenvironment. Therefore, an

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extensive stroma is often associated with metastatic spread and poor prognosis [6, 7].

While many activated CAFs overexpress FAP, ubiquitous resting fibroblasts in healthy tissue have no or low FAP expression. FAP is a type II transmembrane serine protease with post proline dipeptidyl peptidase as well as endopeptidase activity [8]. FAP is overexpressed by stromal fibroblasts in over 90% of epithelial carcinomas. However, FAP expression is not cancer specific: many activated fibroblasts express FAP in wound healing and in nonmalignant diseases such as chronic inflammation, rheumatological diseases, myocardial infarction, lung fibrosis, or liver cirrhosis, among others [8, 9].

The objective of this multicenter retrospective analysis was to perform a head-to-head comparison of ^{68}Ga -FAPI to ^{18}F -FDG in patients with a variety of tumor types to establish generalizable differences between the two agents. Therefore, we intra-individually compared the biodistribution in normal tissue and tumor uptake of ^{68}Ga -FAPI and ^{18}F -FDG.

Methods

Patients

For this international multicenter retrospective analysis, we screened the database of 6 centers (University Hospital Heidelberg (UKHD, GER), University Hospital Würzburg (UKW, GER), University Hospital Essen (UHE, GER), Technical University Hospital Munich (TUM, GER), University of California Los Angeles (UCLA, USA), University of Pretoria (UP, RSA)) for patients with various cancers who underwent both ^{18}F -FDG and ^{68}Ga -FAPI PET/CT scans within a maximum of 3 months apart. Seventy-one patients were finally included (UKHD $n=29$, UKW $n=16$, UHE $n=6$, TUM $n=1$, UCLA $n=14$, UP $n=5$). Data were anonymized, centralized (UKHD), and retrospectively analyzed (UKHD) (Supplement Table 1). This present study enhances and evaluates novel aspects of the UKHD patient cohort partially previously described (Ref: 17).

The ^{18}F -FDG PET/CT scans were performed as per standard of care for oncologic indications. All patients from German sites gave written informed consent to undergo a ^{68}Ga -FAPI PET/CT scan following national regulations, the Declaration of Helsinki, and Good Clinical Practice (GCP). The radiopharmaceutical was produced in accordance with the German Pharmaceuticals Act §13(2b) and the responsible regulatory bodies. The retrospective evaluation of data was approved by the ethics committee of Heidelberg University (permit S016/2018), by the ethics committee from University of Pretoria (881/2019), by the ethics committee of University Hospital Würzburg (permit 2021031005), ethics committee of Technical University Hospital Munich (permit 332/21S).

Patients at UCLA were enrolled into the prospective study NCT04147494 conducted under the Radioactive Drug Research Committee (RDRC) program and approved by the institutional review board (IRB#19-000756) and the Jonsson Comprehensive Cancer Center (JCCC) Internal Scientific Peer Review Committee (ISPRC). All UCLA patients gave written informed consent to participate in the parent prospective study. The UCLA IRB approved this anonymized retrospective analysis (approval #20-001663), and the requirement to obtain informed consent was waived.

PET image acquisition

As this is an international retrospective multicenter study, a variety of PET/CT systems were used for image acquisition and reconstruction (Supplemental Table 2). Overall, whole body images encompassing the patients' head to mid thighs were obtained for both ^{18}F -FDG and ^{68}Ga -FAPI scans. For both tracers, PET/CT datasets acquired approximately 1 h after injection were used for analysis. All PET scans were acquired in 3D mode with an acquisition time of 3–5 min/bed position at all sites. The median time interval between ^{18}F -FDG PET/CT and ^{68}Ga -FAPI PET/CT was 10 days (range 1–89 days). No change of therapy took place in between the scans.

^{18}F -FDG imaging Patients were instructed to fast for at least 6 h before the scan, and blood glucose levels were measured before injection. All patients had serum glucose levels of <150 mg/dl prior to the scan. Median injected activity was 316 MBq (range 233–680 MBq).

^{68}Ga -FAPI imaging Several different ^{68}Ga -labeled FAPI ligands of similar chemical composition were used in this study: FAPI-02, $n=6$; FAPI-04, $n=32$; FAPI-46, $n=32$; and FAPI-74, $n=1$. Radiosynthesis and labeling were performed as described previously [10, 11]. Median injected activity was 185 MBq (range 52–325).

Image analysis

Volumes of interest (VOI) were placed over the normal organs by one UKHD investigator (JS; supervised by FLG&CK) with a diameter of 1 cm for the small organs (thyroid, parotid gland, myocardium, oral mucosa, and spinal cord) to 2 cm for the other organs (brain, muscle, liver, spleen, kidney, fat, aortic lumen, and lung). Circular regions of interest (ROI) were placed on axial slices around lesions with focally increased tracer uptake and were automatically incorporated into a 3-dimensional volume of interest (ESoft; Siemens). A 40% isocontouring approach was used for organs as well as lesions. Normal organs and tissue tracer uptake and biodistribution were quantified by SUVmean and SUVmax. Tumor-to-

background ratios (TBRs) were determined to quantify the image contrast. The TBR was obtained from the geometric mean of the intra-individual quotients of lesion (SUVmax) to background tissue (SUVmax). TBRs were calculated for metastases in lymph nodes (relative to fat tissue), bone (relative to bone spongiosa), liver (relative to liver parenchyma), and lung (relative to lung parenchyma). In addition, TBRs of all tumors were calculated in relation to different tissue types (blood pool, muscle, fat tissue).

Statistical analysis

We used descriptive analyses for demographics and tumor characteristics. For description of SUV, arithmetic mean, standard deviation, and median were used. Comparison between ⁶⁸Ga-FAPI- and ¹⁸F-FDG-SUV in tumor and normal tissue and TBRs was compared with the Wilcoxon signed-rank test. A *p* value of <0.05 was considered statistically significant. All statistical analyses were performed using SPSS Statistics Version 24 (IBM, Armonk, NY, USA), Excel for Mac Version 15.41 (Microsoft, Redmond, Washington, USA) and SigmaPlot 12.5 (Systat Software Inc.) for graphical visualization.

Results

Study population

The clinical characteristics of the study population are summarized in Supplement Table 1. The following cancer types

were included: head and neck cancer (*n* = 16), lung carcinoma (*n* = 9), biliary-pancreatic cancer (*n* = 12), gastrointestinal tract cancer (*n* = 14, including colon carcinoma, rectal carcinoma, anal carcinoma, cecum carcinoma, sigmoid carcinoma, gastro-esophageal cancer, duodenal cancer), and gynecologic cancer (*n* = 12). The group of other cancers (*n* = 8) consists of a neuroendocrine bladder carcinoma, a prostate carcinoma, a B-cell lymphoma, a synovial sarcoma of the lung, an adrenal gland carcinoma, a malignant solitary fibrous cancer, and two cancers of unknown primary (CUP). Forty-one of 71 patients with primary tumors were included. One hundred sixty-four metastatic lesions were found in 43 patients.

Biodistribution in normal organs

Biodistribution of normal organs for ⁶⁸Ga-FAPI and ¹⁸F-FDG is presented in Fig. 1. The SUVmean, SUVmax, and TBRs are provided in Supplemental Table 3a, b. ⁶⁸Ga-FAPI uptake was lower than ¹⁸F-FDG uptake in most normal tissues (11 of 15 organs). Mean SUVmax was significantly lower for ⁶⁸Ga-FAPI than ¹⁸F-FDG in brain parenchyma (⁶⁸Ga-FAPI vs. ¹⁸F-FDG: 0.09 vs. 10.72; *p* < 0.001), oral mucosa (2.04 vs. 3.33; *p* < 0.001), parotid gland (1.71 vs. 2.04; *p* < 0.001), myocardium (1.50 vs. 3.27; *p* < 0.001), blood pool (1.81 vs. 2.34; *p* < 0.001), liver (1.42 vs. 3.10; *p* < 0.001), pancreas (1.82 vs. 1.99; *p* = 0.027), spleen (1.33 vs. 2.60; *p* < 0.001), and kidney cortex (2.20 vs. 2.80; *p* < 0.001). Furthermore, the GI tract showed no relevant ⁶⁸Ga-FAPI uptake as compared to ¹⁸F-FDG (measured in colon transversum: mean SUVmax 1.40 vs. 2.05; *p* < 0.001; SUVmean 0.74 vs. 1.08; *p* < 0.001) (Fig. 2). No significant

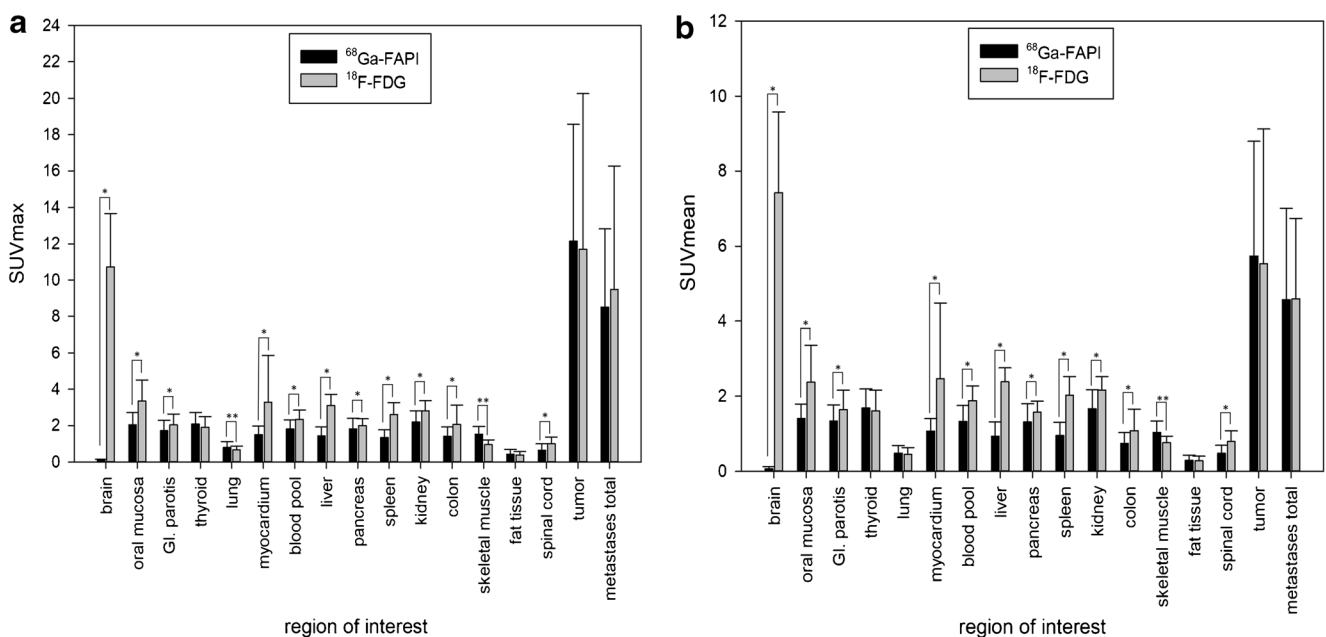
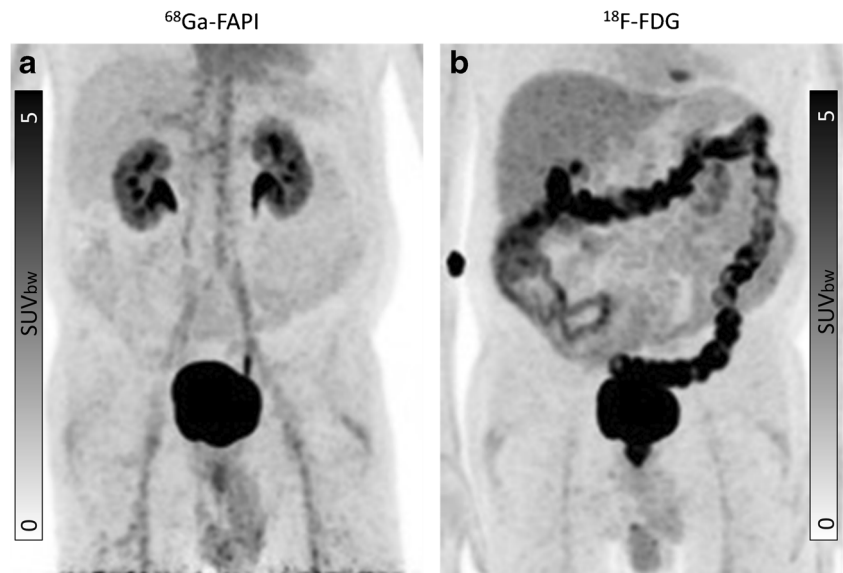


Fig. 1 Biodistribution (SUVmax and SUVmean) of ⁶⁸Ga-FAPI in comparison to ¹⁸F-FDG in normal organs and tumor lesions (mean values and standard deviations; *, ⁶⁸Ga-FAPI sign, higher; **, ¹⁸F-FDG sign, higher)

Fig. 2 Intra-individual comparison of ^{18}F -FDG and ^{68}Ga -FAPI in a patient with oral squamous cell carcinoma presenting with inflammation in the colon as incidental finding in the ^{18}F -FDG PET/CT (**b**), while no acute inflammatory process is associated with a ^{68}Ga -FAPI-positive PET signal (**a**). The direct quantitative comparison presents a high SUVmax of 8.11 in the ^{18}F -FDG PET/CT, while ^{68}Ga -FAPI PET/CT demonstrates low tracer accumulation (SUVmax: 0.38)



difference between ^{68}Ga -FAPI and ^{18}F -FDG was found in normal fat tissue (SUVmax 0.44 vs. 0.39; $p=0.057$). Moreover, no significant difference was found in the SUVmean in lung parenchyma (SUVmean 0.48 vs. 0.46; $p=0.056$) and thyroid tissue (SUVmean 1.68 vs. 1.61; $p=0.306$), whereas the SUVmax of lung parenchyma (SUVmax 0.79 vs. 0.66; $p<0.001$) and thyroid tissue (SUVmax 2.08 vs. 1.90; $p=0.049$) as well as skeletal muscles SUVmean und SUVmax (SUVmax 1.50 vs. 0.95; $p<0.001$) showed significantly higher uptake for ^{68}Ga -FAPI than for ^{18}F -FDG.

Uptake in tumor lesions

No significant difference in mean SUVmax of ^{68}Ga -FAPI and ^{18}F -FDG was present in primary tumors ($n=41$;

^{68}Ga -FAPI vs. ^{18}F -FDG: 12.14 vs. 11.69; $p=0.429$) or metastatic lesions in total (SUVmax 8.49 vs. 9.48; $p=0.814$). The site of metastasis had no significant impact on uptake of ^{18}F -FDG or ^{68}Ga -FAPI. Mean SUVmax of ^{68}Ga -FAPI vs. ^{18}F -FDG was 7.89 vs. 11.17; $p=0.334$ for lymph node metastases, 9.82 vs. 8.84; $p=1.000$ for liver metastases, 7.83 vs. 7.46; $p=0.542$ for bone metastases, 6.68 vs. 11.48; $p=0.641$ for lung metastases and 10.67 vs. 8.17; and $p=0.119$ for other types of metastases (pleural, peritoneal and soft tissue metastases), respectively. However, in individual patients, marked differences between ^{18}F -FDG and ^{68}Ga -FAPI tumor uptake were observed. An example of a patient with tumor lesions with low uptake on ^{18}F -FDG PET but high uptake on ^{68}Ga -FAPI scan is shown in Fig. 3.

Fig. 3 Intra-individual comparison of ^{18}F -FDG and ^{68}Ga -FAPI in two patients with ovarian cancer (**a**) and pancreas cancer (**b**), respectively. Both present with strong ^{68}Ga -FAP uptake in the primary and metastatic lesions while only slight to moderate uptake on ^{18}F -FDG PET/CT (arrow: green (^{68}Ga -FAPI) and red (^{18}F -FDG))

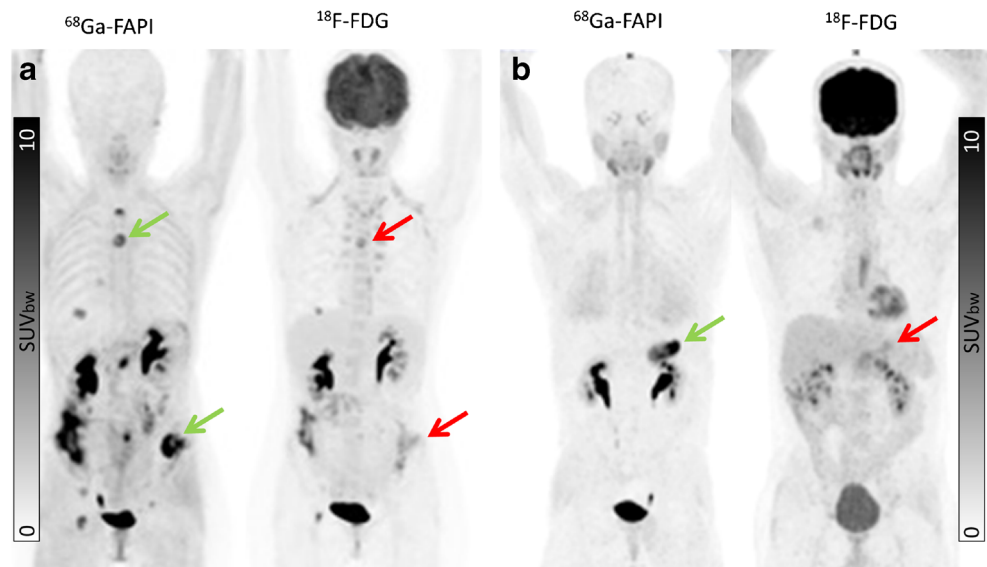


Table 1 TBRs for different metastases (Lymph node, Bone, Liver, Lung)

VOI	Lymph node metastases/fat tissue (n=26)		Bone metastases/bone spongiosa (n=13)		Hepatic metastases/liver parenchyma (n=14)		Lung metastases/lung parenchyma (n=8)	
	⁶⁸ Ga-FAPI	¹⁸ F-FDG	⁶⁸ Ga-FAPI	¹⁸ F-FDG	⁶⁸ Ga-FAPI	¹⁸ F-FDG	⁶⁸ Ga-FAPI	¹⁸ F-FDG
TBR	17.38	24.76	7.16	3.25	5.84	2.62	7.21	11.31
p value	0.132		0.033		0.011		0.313	

Table 2 Primary tumor TBRs

VOI	Tumor/blood pool		Tumor/skeletal muscle		Tumor/fat tissue	
	⁶⁸ Ga-FAPI	¹⁸ F-FDG	⁶⁸ Ga-FAPI	¹⁸ F-FDG	⁶⁸ Ga-FAPI	¹⁸ F-FDG
TBR	6.01	3.57	6.99	9.48	27.69	23.96
p value	0.002		< 0.001		0.331	

Tumor-to-background ratios

TBRs for lymph node metastases with fatty tissue as background (⁶⁸Ga-FAPI vs. ¹⁸F-FDG 17.4 vs. 24.8; $p = 0.132$) were the same as were lung metastases to pulmonary parenchyma as background (7.21 vs. 11.31; $p = 0.313$) (Table 1). In contrast, TBRs of bone metastases to bone spongiosa (⁶⁸Ga-FAPI vs. ¹⁸F-FDG, 7.2 vs. 3.3; $p = 0.033$) and hepatic metastases to liver parenchyma (⁶⁸Ga-FAPI vs. ¹⁸F-FDG, 5.8 vs. 2.6; $p = 0.011$) were substantially higher with ⁶⁸Ga-FAPI than ¹⁸F-FDG (Table 1). Primary tumor TBRs were approximately equal between the two agents (Table 2).

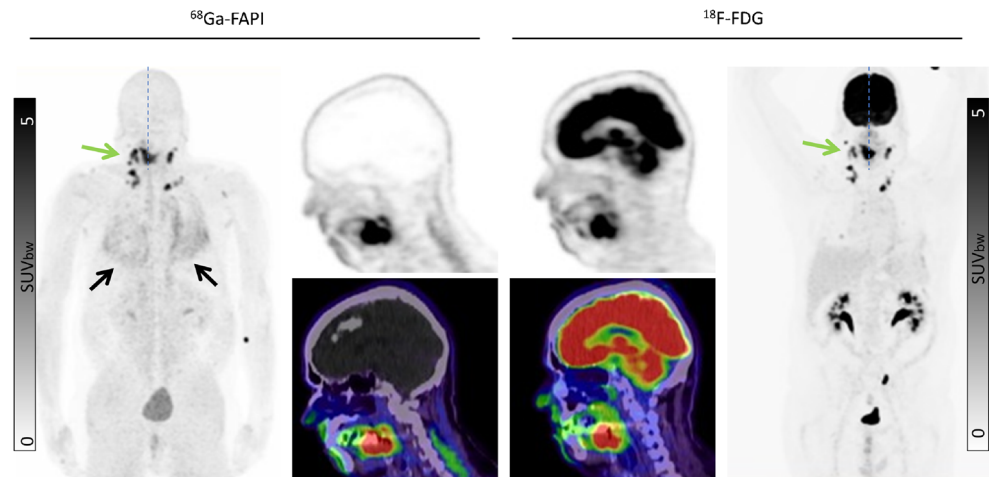
Discussion

This is an international multicenter retrospective analysis for the intra-individual comparison of ¹⁸F-FDG with

newly developed ⁶⁸Ga-labeled FAP inhibitors [4, 5]. Regardless of the various equipment and imaging protocols used, we report high tumor-to-background ratios for ⁶⁸Ga-FAPI, comparable or superior to ¹⁸F-FDG that may result in high diagnostic performance for cancer staging and restaging.

Our findings suggest that ⁶⁸Ga-FAPI may improve tumor delineation in organs with physiologically high ¹⁸F-FDG background activity. This may be relevant in cancers such as pancreatic, ovarian, head-and-neck, hepatocellular and cholangiocarcinoma, colon cancer, as well as brain tumors and lung cancer [12–15]. Due to low ⁶⁸Ga-FAPI uptake in most normal parenchyma, favorable tumor delineation was found in head-and-neck region (Fig. 4), liver metastases (Fig. 5), and abdominal cancer (Fig. 6). Furthermore, in the mediastinal region, ⁶⁸Ga-FAPI uptake in the cardiac muscle is very low compared to ¹⁸F-FDG (Fig. 7). However, we also examined a diffuse large B-cell lymphoma which showed relatively low ⁶⁸Ga-FAPI

Fig. 4 A 68-year-old patient with a histologically confirmed squamous cell carcinoma of the edge of the tongue for pre-radiotherapeutic staging using PET/CT. The quantified uptake in the primary tumor (green arrow) on ⁶⁸Ga-FAPI was SUV_{max} 20.26 compared to the ¹⁸F-FDG uptake with an SUV_{max} 13.35. As a secondary finding, fibrotic, scarred changes of the lung indicative of pulmonary fibrosis were observed (black arrow)



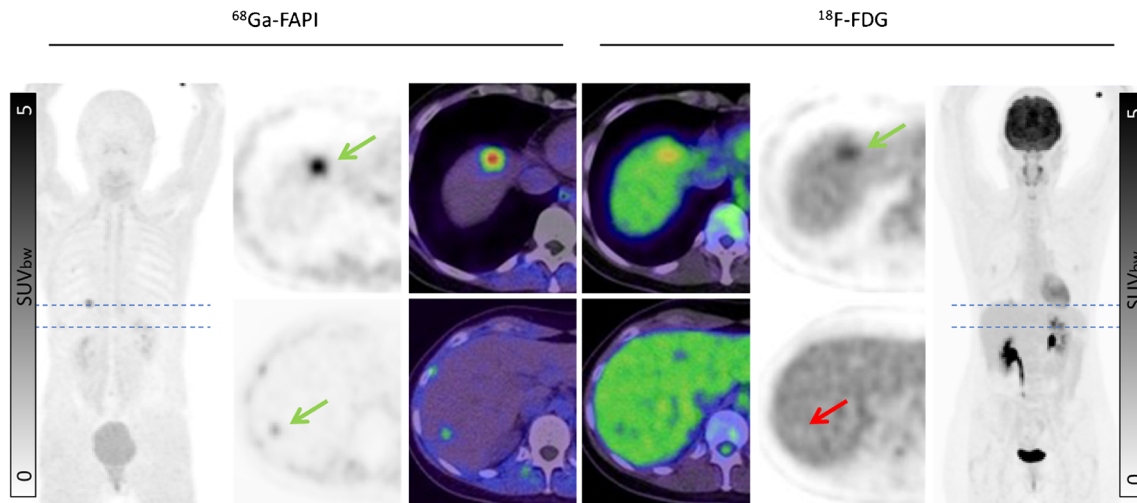


Fig. 5 A 40-year-old female patient with ovarian cancer underwent restaging due to a suspicious subdiaphragmatic hepatic lesion. Tracer uptake in the normal liver parenchyma was markedly different on the two tracers: ^{68}Ga -FAPi SUVmax 0.79 vs. ^{18}F -FDG SUVmax 2.69. In

the liver segment III and VII, a strong ^{68}Ga -FAP uptake was found (green arrow) compared to ^{18}F -FDG. The liver metastatic lesion in segment VII was not detected by ^{18}F -FDG (red arrow)

uptake. FAP-negative tumor phenotypes have also previously been described in the literature, e.g., in differentiated thyroid carcinoma and renal cell cancer [16–18]. Unfortunately, in this work, we can only report the mean values over a relatively heterogenic group of patients. Further research will be needed to explore dedicated kinds of tumor where staging per FAPi PET might provide a clinical advantage in comparison to FDG PET. In addition to tumor entity, it would also be interesting whether FAPi uptake depends on tumor aggressiveness or proliferation rate as it is for ^{18}F -FDG. However, due to small sample sizes within the respective histopathological subtypes, we could not reasonably perform a correlation between

grading and FAPi uptake, yet. This investigation would have to be carried out in a future evaluation.

According to previous reports about their respective biodistribution and pharmacokinetics, the ligands FAPi-02, FAPi-04, and FAPi-46 do not differ in early uptake phase 10 min to 1 h p.i. and were thus considered widely exchangeable with regard to our diagnostic evaluation. In comparison to FAPi-02, the ligands FAPi-04 and FAPi-46 have longer tumor retention time beyond 1 h p.i., which however would only be relevant in the perspective of FAP-targeted radionuclide therapy [18, 19]. FAPi-74 either can be used as ^{68}Ga -FAPi-74 (this work) or alternatively could also be labeled with ^{18}F providing an advantage in large batch production.

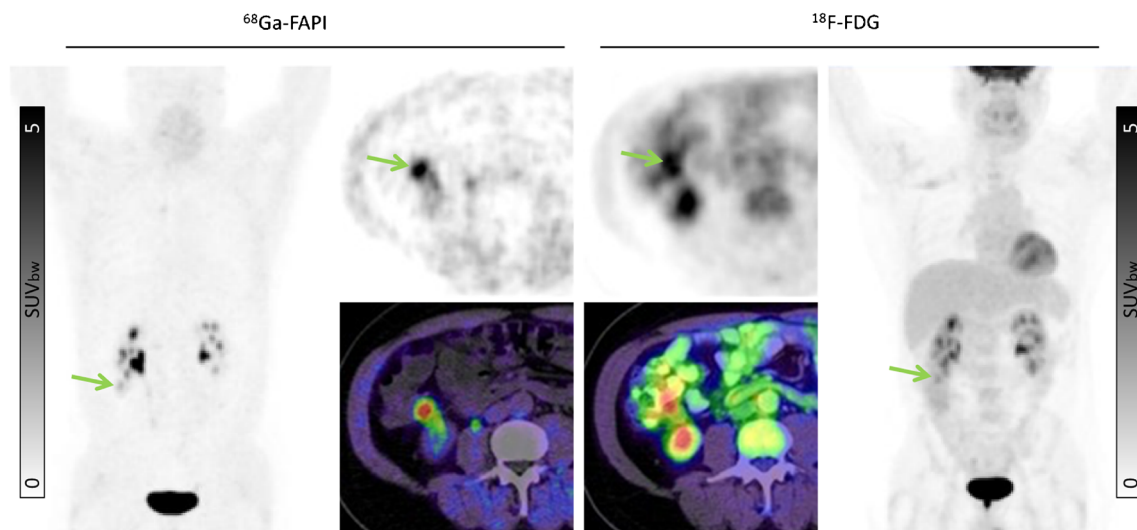


Fig. 6 A 55-year-old male underwent pre-operative staging after being diagnosed with colon cancer in the right ascending colon (green arrow). Both tracers presented uptake in the primary tumor (SUVmax: ^{68}Ga -

FAPi 6.25 vs. ^{18}F -FDG 5.34), but there is considerably more background signal on ^{18}F -FDG

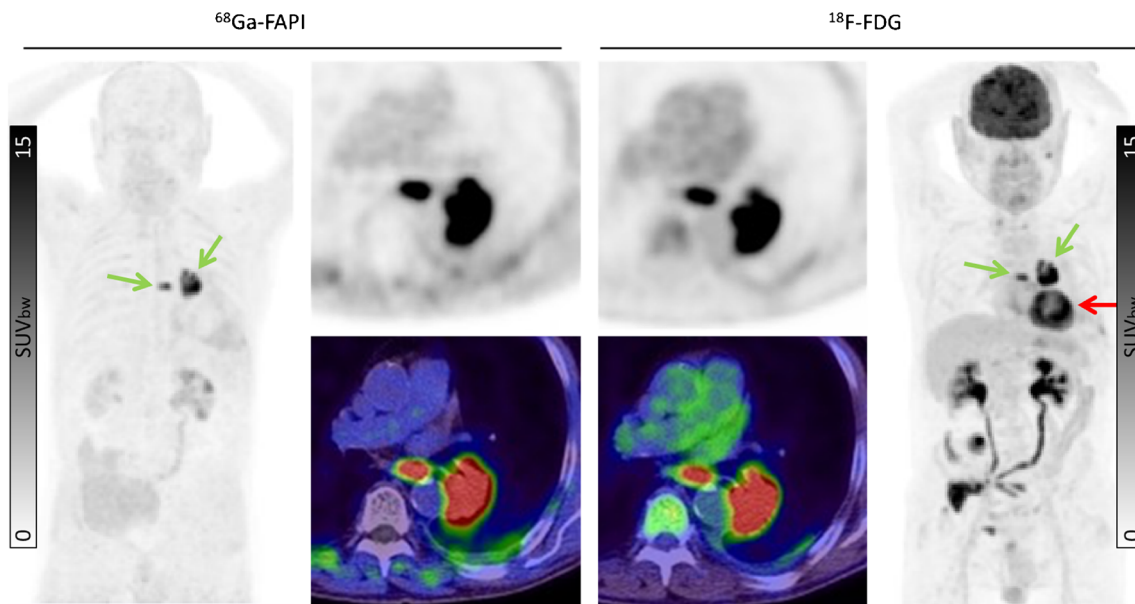


Fig. 7 An 80-year-old male patient with lung cancer (green arrows) was diagnosed by ^{18}F -FDG PET/CT. ^{68}Ga -FAPI PET/CT demonstrated similar tracer uptake (SUVmax: ^{18}F -FDG 15.99 vs. ^{68}Ga -FAPI 17.95).

One advantage of ^{68}Ga -FAPI in this instance is the lack of cardiac muscle uptake which is prominent with ^{18}F -FDG (red arrow)

However, biodistribution at 1 h p.i. was reported comparable to previous ligands [20].

^{68}Ga -FAPI and ^{18}F -FDG reflect two different aspects of tumor behavior. ^{18}F -FDG directly targets tumor cell metabolism. In contrast, ^{68}Ga -FAPI targets FAP on CAFs in the stroma of tumors, which is an indicator of desmoplastic reaction and was reported to be one of the key determinants of tumor immunity [19] and multidrug resistance [21] possibly related to reductions in transtumoral transport of cells and drugs. Thus, in addition to its function as a staging modality, FAPI PET/CT may be helpful in understanding tumor biology related to the tumor microenvironment.

Due to the limited number of studies to date, the tumor entities with low or high ^{68}Ga -FAPI uptake are also not yet sufficiently known.

Furthermore, FAPI could also play a role in the examination of myocardial infarction and IgG4-related diseases [22, 23].

Our retrospective analysis has several limitations: ^{68}Ga -FAPI PET/CT was performed with four different ligands, which however share a common backbone and their early phase biodistribution as well as tumor uptake are comparable [18–20]. Lacking gold standard validation of discrepant lesions, we are not able to evaluate the respective sensitivity, specificity, and accuracy of ^{18}F -FDG and ^{68}Ga -FAPI, which was beyond the scope of the current work but has already been addressed by other researchers [24].

The main limitation is the long (up to 3 month) interval between the examinations, which may cause “interval progression” that might be non-neglectable in aggressive cancers. However,

we had to find a good tradeoff between test vs. re-test reliability and inclusion of a sufficient large patient cohort. For future clinical trials, more stringent inclusion criteria—and alternating order of tracer administration—should be encouraged.

Conclusion

In this international multicenter retrospective analysis, 71 patients with various cancers underwent both ^{18}F -FDG and ^{68}Ga -labeled FAP inhibitors. ^{68}Ga -FAPI showed equal or higher TBR at lower radiation doses than ^{18}F -FDG. These findings suggest that ^{68}Ga -FAPI may demonstrate higher diagnostic performance for cancer staging and restaging in various indications. In addition, the high target to background ratios and the low uptake in normal organs also suggest potential use of FAPI ligands as a potential means of treating tumors with therapeutic radioisotopes.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00259-021-05307-1>.

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Declarations

Conflict of interest UH, TL, CK, and FLG have a patent application for quinolone based FAP-targeting agents for imaging and therapy in nuclear

medicine. UH, TL, CK, and FLG also have shares of a consultancy group for iTheranostics. FLG is also advisor at ABX, Telix, and SOFIE Biosciences. JCa is supported by the Prostate Cancer Foundation (2020 Young Investigator Award 20YOUN05) and the Society of Nuclear Medicine and Molecular imaging (2019 ERF Molecular Imaging Research Grant for Junior Academic Faculty). The other authors declare no conflict of interest regarding this manuscript.

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References

- Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646–74. <https://doi.org/10.1016/j.cell.2011.02.013>.
- Almuhaidab A, Papatthanasiou N, Bomanji J. 18F-FDG PET/CT imaging in oncology. *Ann Saudi Med*. 2011;31(1):3–13. <https://doi.org/10.4103/0256-4947.75771>.
- Agrawal A, Rangarajan V. Appropriateness criteria of FDG PET/CT in oncology. *Indian J Radiol Imaging*. 2015;25(2):88–101. <https://doi.org/10.4103/0971-3026.155823>.
- Lindner T, Loktev A, Altmann A, Giesel F, Kratochwil C, Debus J, et al. Development of quinoline-based theranostic ligands for the targeting of fibroblast activation protein. *J Nucl Med*. 2018;59(9):1415–22. <https://doi.org/10.2967/jnumed.118.210443>.
- Loktev A, Lindner T, Mier W, Debus J, Altmann A, Jäger D, et al. A tumor-imaging method targeting cancer-associated fibroblasts. *J Nucl Med*. 2018;59(9):1423–9. <https://doi.org/10.2967/jnumed.118.210435>.
- Gascard P, Tlsty TD. Carcinoma-associated fibroblasts: orchestrating the composition of malignancy. *Genes Dev*. 2016;30(9):1002–19. <https://doi.org/10.1101/gad.279737.116>.
- Barbazán J, Matic VD. Cancer associated fibroblasts: is the force the path to the dark side? *Curr Opin Cell Biol*. 2019;56:71–9. <https://doi.org/10.1016/j.ceb.2018.09.002>.
- Hamson EJ, Keane FM, Tholen S, Schilling O, Gorrell MD. Understanding fibroblast activation protein (FAP): substrates, activities, expression and targeting for cancer therapy. *Proteomics Clin Appl*. 2014;8(5–6):454–63. <https://doi.org/10.1002/prca.201300095>.
- Schmidkonz C, Rauber S, Atzinger A, Agarwal R, Götz TI, Soare A, et al. Disentangling inflammatory from fibrotic disease activity by fibroblast activation protein imaging. *Ann Rheum Dis*. 2020;79(11):1485–91. <https://doi.org/10.1136/annrheumdis-2020-217408>.
- Lindner T, Loktev A, Giesel F, Kratochwil C, Altmann A, Haberkorn U. Targeting of activated fibroblasts for imaging and therapy. *EJNMMI Radiopharm Chem*. 2019;4(1):16. <https://doi.org/10.1186/s41181-019-0069-0>.
- Loktev A, Lindner T, Burger EM, Altmann A, Giesel F, Kratochwil C, et al. Development of fibroblast activation protein-targeted radiotracers with improved tumor retention. *J Nucl Med*. 2019;60(10):1421–9. <https://doi.org/10.2967/jnumed.118.224469>.
- Giesel FL, Heussel CP, Lindner T, Rohrich M, Rathke H, Kauczor HU, et al. FAPI-PET/CT improves staging in a lung cancer patient with cerebral metastasis. *Eur J Nucl Med Mol Imaging*. 2019;46(8):1754–5. <https://doi.org/10.1007/s00259-019-04346-z>.
- Chen H, Pang Y, Wu J, Zhao L, Hao B, Wu J, et al. Comparison of [(68)Ga]Ga-DOTA-FAPI-04 and [(18)F] FDG PET/CT for the diagnosis of primary and metastatic lesions in patients with various types of cancer. *Eur J Nucl Med Mol Imaging*. 2020;47(8):1820–32. <https://doi.org/10.1007/s00259-020-04769-z>.
- Koerber SA, Staudinger F, Kratochwil C, Adeberg S, Haefner MF, Ungerechts G, et al. The role of FAPI-PET/CT for patients with malignancies of the lower gastrointestinal tract - first clinical experience. *J Nucl Med*. 2020. <https://doi.org/10.2967/jnumed.119.237016>.
- Syed M, Flechsig P, Liermann J, Windisch P, Staudinger F, Akbaba S, et al. Fibroblast activation protein inhibitor (FAPI) PET for diagnostics and advanced targeted radiotherapy in head and neck cancers. *Eur J Nucl Med Mol Imaging*. 2020;47(12):2836–45. <https://doi.org/10.1007/s00259-020-04859-y>.
- Wang G, Jin X, Zhu H, Wang S, Ding J, Zhang Y, et al. 68Ga-NOTA-FAPI-04 PET/CT in a patient with primary gastric diffuse large B cell lymphoma: comparisons with [18F] FDG PET/CT. *Eur J Nucl Med Mol Imaging*. 2020. <https://doi.org/10.1007/s00259-020-04946-0>.
- Kratochwil C, Flechsig P, Lindner T, Abderrahim L, Altmann A, Mier W, et al. (68)Ga-FAPI PET/CT: tracer uptake in 28 different kinds of cancer. *J Nucl Med*. 2019;60(6):801–5. <https://doi.org/10.2967/jnumed.119.227967>.
- Giesel FL, Kratochwil C, Lindner T, Marschalek MM, Loktev A, Lehnert W, et al. (68)Ga-FAPI PET/CT: biodistribution and preliminary dosimetry estimate of 2 DOTA-containing FAP-targeting agents in patients with various cancers. *J Nucl Med*. 2019;60(3):386–92. <https://doi.org/10.2967/jnumed.118.215913>.
- Meyer C, Dahlbom M, Lindner T, Vauclin S, Mona C, Slavik R, et al. Radiation dosimetry and biodistribution of (68)Ga-FAPI-46 PET imaging in cancer patients. *J Nucl Med*. 2019. <https://doi.org/10.2967/jnumed.119.236786>.
- Giesel F, Adeberg S, Syed M, Lindner T, Jimenez LD, Mavriopoulou E, et al. FAPI-74 PET/CT using either (18)F-AIF or cold-kit (68)Ga-labeling: biodistribution, radiation dosimetry and tumor delineation in lung cancer patients. *J Nucl Med*. 2020. <https://doi.org/10.2967/jnumed.120.245084>.
- Erin N, Grahovac J, Brozovic A, Efferth T. Tumor microenvironment and epithelial mesenchymal transition as targets to overcome tumor multidrug resistance. *Drug Resist Updat*. 2020;53:100715. <https://doi.org/10.1016/j.drug.2020.100715>.
- Luo Y, Pan Q, Yang H, Peng L, Zhang W, Li F. Fibroblast activation protein targeted PET/CT with (68)Ga-FAPI for imaging IgG4-related disease: comparison to (18)F-FDG PET/CT. *J Nucl Med*. 2020. <https://doi.org/10.2967/jnumed.120.244723>.
- Tillmanns J, Hoffmann D, Habbaba Y, Schmitto JD, Sedding D, Fraccarollo D, et al. Fibroblast activation protein alpha expression identifies activated fibroblasts after myocardial infarction. *J Mol Cell Cardiol*. 2015;87:194–203. <https://doi.org/10.1016/j.yjmcc.2015.08.016>.
- Chen H, Zhao L, Ruan D, Pang Y, Hao B, Dai Y, et al. Usefulness of [(68)Ga]Ga-DOTA-FAPI-04 PET/CT in patients presenting with inconclusive [(18)F]FDG PET/CT findings. *Eur J Nucl Med Mol Imaging*. 2020. <https://doi.org/10.1007/s00259-020-04940-6>.

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