# **UC Irvine UC Irvine Previously Published Works**

# **Title**

Sex- and genotype-dependent nicotine plus cue-primed reinstatement is enhanced in adolescent Sprague Dawley rats containing the human CHRNA6 3′-UTR polymorphism (rs2304297)

**Permalink** <https://escholarship.org/uc/item/2bn19330>

**Authors**

Carreño, Diana Lotfipour, Shahrdad

**Publication Date**

2023

# **DOI**

10.3389/fpsyt.2022.1064211

Peer reviewed

#### Check for updates

#### OPEN ACCESS

EDITED BY Kristine C-Y McGrath, University of Technology Sydney, Australia

#### REVIEWED BY

Lawrence Toll, Florida Atlantic University, United States Hui Chen, University of Technology Sydney, Australia

\*CORRESPONDENCE Diana Carreño lcarreno@uci.edu

SPECIALTY SECTION This article was submitted to Addictive Disorders, a section of the journal Frontiers in Psychiatry

RECEIVED 07 October 2022 ACCEPTED 21 December 2022 PUBLISHED 10 January 2023

#### CITATION

Carreño D and Lotfipour S (2023) Sex- and genotype-dependent nicotine plus cue-primed reinstatement is enhanced in adolescent Sprague Dawley rats containing the human *CHRNA*6 3 <sup>0</sup>-UTR polymorphism (rs2304297). *Front. Psychiatry* 13:1064211. doi: [10.3389/fpsyt.2022.1064211](https://doi.org/10.3389/fpsyt.2022.1064211)

#### COPYRIGHT

© 2023 Carreño and Lotfipour. This is an open-access article distributed under the terms of the [Creative](http://creativecommons.org/licenses/by/4.0/) [Commons](http://creativecommons.org/licenses/by/4.0/) [Attribution](http://creativecommons.org/licenses/by/4.0/) [License](http://creativecommons.org/licenses/by/4.0/) [\(CC BY\).](http://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Sex- and genotype-dependent nicotine plus cue-primed reinstatement is enhanced in [adolescent Sprague Dawley rats](https://www.frontiersin.org/articles/10.3389/fpsyt.2022.1064211/full) containing the human *CHRNA*6 3 0 -UTR polymorphism (rs2304297)

#### Diana Carreño<sup>1\*</sup> and Shahrdad Lotfipour<sup>1,2,3</sup>

<sup>1</sup>Department of Pharmaceutical Sciences, University of California, Irvine, Irvine, CA, United States, <sup>2</sup> Department of Pathology and Laboratory Medicine, University of California, Irvine, Irvine, CA, United States, <sup>3</sup>Department of Emergency Medicine, University of California, Irvine, Irvine, CA, United States

Rationale: Large-scale human candidate gene studies have indicated that a genetic variant (rs2304297) in the alpha(α)6 nicotinic acetylcholine receptor (nAChR) subunit, encoded by the *CHRNA*6 gene, may play a key role in adolescent nicotine addictive behavior. We hypothesized that the polymorphism selectively enhances nicotine + cue-primed reinstatement, but not nicotine- or cue-reinstatement in α6 *GG* (risk) vs. α6 *CC* (non-risk) allele carriers, without having baseline effects on natural rewards.

Methods: Using CRISPR-Cas9 genomic engineering, we developed a humanized rat line with the human gene variant of the CHRNA6 3'-UTRC123G polymorphism in Sprague-Dawley rats. Genetically modified adolescent male and female rats were food trained under a fixed-ratio (FR)1 schedule of reinforcement and progressively increased to FR5. Animals were implanted with catheters and began nicotine self-administration (15 µg/kg/infusion) at FR5. Upon reaching stable responding, reinforced behavior was extinguished by removal of drug and cues. Reinstatement testing began for cue only, nicotine only, and nicotine + cue in a Latin Square Design. Animals were returned to extinction conditions for 2 days minimum between testing.

Results: For natural food rewards, nicotine self-administration, progressive ratio, and extinction, adolescent male and female (α6 *GG* and α6 *CC*) rats exhibited equivalent behaviors. Male α6 *GG* rats show enhanced nicotine + cue-primed reinstatement when compared with male α6 *CC* rats. This genotype effect on reinstatement was not seen in female rats.

Conclusion: Our findings support the *in vivo* functional role of the human CHRNA6 3'-UTR SNP genetic variant in sex-dependently enhancing nicotine seeking behavior in adolescent rats. Overall, the findings support clinical and preclinical data highlighting a role of α6 nAChRs mediating sex heterogeneity in substance use and related phenotypes.

#### KEYWORDS

alpha6 nicotinic acetylcholine receptor subunits, addiction, drug-seeking behavior, pharmacogenetics, relapse

#### 1. Introduction

In recent years the prevalence of adolescent electronic nicotine use has dramatically increased [\(1,](#page-10-0) [2\)](#page-10-1). Adolescent electronic nicotine use remains a public health concern given that its use can progress into combustible cigarette smoking and conditions the developing brain for addiction to other drugs of abuse [\(3–](#page-10-2)[6\)](#page-10-3). Young people are highly sensitive to nicotine, exhibiting symptoms of dependence soon after smoking initiation and before the start of daily smoking [\(7\)](#page-10-4). Further, electronic cigarette cessation interest has increased among adolescents. In 2019 an estimated 57.5% middle and high school (about 3.5 million) students using tobacco/nicotine made an attempt to quit [\(8\)](#page-10-5). The prevalence of unsuccessful quit attempts among middle and high school students who had either used e-cigarettes or cigarettes, was higher in 2020 than previous years [\(9\)](#page-10-6).

Nicotine is the principal reinforcing constituent in tobacco products responsible for drug seeking and addiction. Nicotine directly activates neuronal nicotinic acetylcholine receptors (nAChR), which are ligand-gated ion channels consisting of five membrane-spanning subunits located on several neurotransmitter systems in the brain, most notably dopaminergic neurons [\(10\)](#page-10-7). In the mammalian brain the predominant and most prevalent nAChRs implicated in the addictive properties of nicotine are those containing α4 and β2 subunits [\(11–](#page-10-8)[13\)](#page-10-9). These subunits can independently partner with  $\alpha$ 6<sup>\*</sup> nAChR subunits to makeup  $\alpha$ 6β2β3 (non-α4) and α6α4β2β3 nAChRs [\(10,](#page-10-7) [13–](#page-10-9)[17\)](#page-10-10). α6 <sup>∗</sup> nAChRs are largely localized in expression on the cell body or terminal regions of dopaminergic neurons and act as critical modulators of dopamine release in reward regions of the brain [\(10,](#page-10-7) [14,](#page-10-11) [17,](#page-10-10) [18\)](#page-10-12). α6 <sup>∗</sup> nAChRs reach highest mRNA expression in the ventral tegmental area (VTA) and substantia nigra (SN) during adolescence [\(19\)](#page-10-13). α6 knockout (KO) mice do not self-administer nicotine even with an extensive range of doses, an effect which can be rescued using lentiviral re-expression of α6 nAChR subunits directly within the VTA [\(20\)](#page-10-14). Further, administration of alpha(α)-conotoxin MII, an α6β2β3 antagonist, into the nucleus accumbens shell attenuate nicotine self-administration in rats [\(21\)](#page-10-15). These studies suggest the  $\alpha$ 6<sup>\*</sup> nAChRs in the VTA

and/or its projections, e.g., nucleus accumbens and prefrontal cortex, are necessary and sufficient to establish nicotine selfadministration. Evidence also demonstrates that a selective α6 nAChRs antagonist, bPiDDB, is able to decrease nicotineinduced reinstatement in nicotine self-administering rats [\(22\)](#page-10-16). Drug paired cues are likely important factors in such behavioral effects, as intra-VTA infusion of an α6 nAChRs antagonist, α-conotoxin MII, blocks the rewarding effects of cues paired with a drug reinforcer [\(23,](#page-10-17) [24\)](#page-10-18). Such studies are important as cue and drug-induced reinstatement is a preclinical model of drug-seeking behavior (see reviews [\(25](#page-11-0)[–27\)](#page-11-1)) and highlight the importance of α6 <sup>∗</sup> nAChRs critical involvement.

Clinically, a large body of literature highlights that a single nucleotide polymorphism (SNP) in the α6 nAChR subunit (encoded by the CHRNA6 gene) is associated with nicotine/tobacco use and related problems [\(28–](#page-11-2)[39\)](#page-11-3). The human CHRNA6 SNP is located in the 3'-untranslated region (UTR), a genomic region known to regulate mRNA stability, localization, and translation [\(40,](#page-11-4) [41\)](#page-11-5). The α6 nAChR subunit SNP has been associated with increased cigarette smoking and drug experimentation during adolescence with  $\alpha$ 6<sup>GG</sup>- more impacted than  $\alpha$ 6<sup>CC</sup>-allele carriers [\(34\)](#page-11-6). Additionally, a clinical study revealed a sex-treatment interaction for rs230497, with a twofold greater abstinence in the bupropion arm in males versus females by end of treatment, although not surpassing Bonferroni corrections [\(33\)](#page-11-7). Sex heterogeneity has been observed as a critical factor to be considered with α6 nAChRs, the CHRNA6 gene and the CHRNA6 3'-UTR SNP in nicotine addiction and other neurological diseases [\(33,](#page-11-7) [42](#page-11-8)-45). The CHRNA6 3'-UTR SNP has been associated with nicotine dependence in males [\(46\)](#page-11-10). Further, other CHRNA6 SNPs in general show greater nicotine dependence associations in males than females [\(43\)](#page-11-11). As such, clinical findings provide evidence for the CHRNA6 3'-UTR SNP contributing to the susceptibility of tobacco/nicotine dependence with the strong likelihood of sex-dependent effects. Our prior pre-clinical data support this hypothesis [\(47\)](#page-11-12).

Mechanisms underlying how the CHRNA6 3'-UTR mediates adolescent substance use are not known. To investigate the human CHRNA6 3'-UTR SNP in vivo, our lab generated a novel, humanized rodent line. Our lab replaced the entire CHRNA6 3'-UTR of the rat line with the human CHRNA6

3 0 -UTR, generating a translational model of α6 nAChRs [\(47\)](#page-11-12). Our recent results using this novel rat line confirm in vivo functionality of the CHRNA6 3'-UTR SNP, with genotype- and sex-dependent effects observed on adolescent nicotine-induced behaviors [\(47\)](#page-11-12). The aim of our current study is to assess sexand genotype-dependent effects on the influence of nicotine selfadministration, progressive ratio, extinction and reinstatement of nicotine-seeking behavior in adolescent males and females containing the humanized CHRNA6 3'-UTR SNP.

### 2. Materials and methods

#### 2.1. Generation of human CHRNA6 3 0 -UTR SNP rodents

Human CHRNA6 3'-UTR SNP knock-in rats were designed and created with CRISPR/Cas9 gene editing techniques by Cyagen Biosciences as described in Cardenas et al. [\(47\)](#page-11-12). Briefly, donor vectors and sgRNA were designed to target the 3'-UTR of the rat CHRNA6 gene (GenBank accession number: NM\_057184.1; Ensembl: ENSRNOG00000012283). Donor vectors contained the human CHRNA6 3'-UTR (559 nucleotides) with either the minor SNP rs2304297 allele, C, or major SNP rs2304297 allele, G, at nucleotide position 123 and replaced the rat 3'-UTR (95 nucleotides).

#### 2.2. Animals

Male and female wild type (WT) Sprague-Dawley rats were purchased from Charles river and bred in house with human CHRNA6 3'-UTRC123G SNP rats. Upon weaning at postnatal day (PN) 21, animals were transferred out of the mother's cage and separated by sex (**[Figure 1](#page-4-0)**). All animals were handled for 3-days prior to experimentation and group-housed (2 per cage) throughout the experiment. All rats were maintained on a 12 h light/dark cycle (lights on at 07:00 am). All experimental procedures were in compliance with NIH guidelines and were approved by the Institutional Animal Care and Use Committee of the University of California, Irvine. Animals were handled for 2 min daily before testing began. No more than one male and female animal per litter per experimental group was used to avoid potential confounds. Rats were minimally food-restricted beginning two days prior to operant conditioning to increase the motivation to learn the self-administration paradigm similar to our prior study [\(48\)](#page-11-13). Group-housed (2 per cage) adolescent rats were fed 15–25 g of food to maintain normal growth during selfadministration testing [\(49\)](#page-11-14). Animal weights based on postnatal days are presented in **[Supplementary Figure 1](#page-10-19)**, with postnatal days across condition highlighted in **[Supplementary Figure](#page-10-19) [2](#page-10-19)**. Attrition rates for animals in our current study are shown in **[Table 1](#page-4-1)**. Similar to our previous wild type Sprague Dawley nicotine reinstatement studies in adolescent rats [\(48\)](#page-11-13) we did not administer vaginal swabs in the present study to assess estrous cycle [\(50–](#page-11-15)[52\)](#page-11-16). The age range of adolescence was defined based on the prior studies of Spear [\(53\)](#page-11-17).

#### 2.3. Apparatus

Animals were tested in plexiglass operant chambers (Med Associates, St Albans, VT, USA), equipped with two levers. The required number of responses at the reinforced (Reinf) lever turned on a cue light over the lever, turned off the house light, and activated an externally mounted syringe pump that infused drug. During the infusion (5.6 s yielding 100 µl of solution) and timeout period (20 s) the cue light remained illuminated, and the house light remained off.

After the timeout period, the house light turned on and signaled the availability of a reinforcer. Responses on the non-reinforced (Non-Reinf) lever were recorded but had no consequences. Procedures modeled previous work [\(54,](#page-11-18) [55\)](#page-11-19).

#### 2.4. Food self-administration

To facilitate learning, male and female adolescent (PN 24) rats were trained twice per day in a 30 min session to lever press for food pellets (45 mg rodent purified diet; Bio-Serv, Frenchtown, NJ, USA) in lever pressing operant testing chambers (Med Associates, St. Albans, VT, USA), based on previous studies [\(54,](#page-11-18) [56\)](#page-11-20) (**[Figure 1](#page-4-0)**). One wall of the chamber contained two levers, a cue light over each, and a house light. The right lever was assigned as the active (Reinf) lever, each response at which was rewarded with delivery of food and presentation of cue light and tone. The left lever was inactive (Non-Reinf) had no consequences but was recorded as a measure of non-specific activity. The animals started at an FR1TO1 (fixed-ratio 1, 1s timeout) schedule of reinforcement, followed by FR1TO10, FR2TO20 and finally FR5TO20, progressing upon earning 35 reinforcers.

#### 2.5. Surgery

Following successful acquisition of food training, rats were anesthetized with Equithesin (0.0035 ml/g body weight) and implanted with indwelling jugular vein catheters [\(57–](#page-11-21)[59\)](#page-11-22) (**[Figure 1](#page-4-0)**). After surgery, rats were given the analgesic carprofen (5 mg/kg, subcutaneous). During the 2–3-day recovery period, catheters were flushed daily with heparinized saline solution (1 ml of 1,000 units/ml heparin into 30 ml of bacteriostatic saline) to maintain patency. Catheter patency was tested for rapid (5–10 s) anesthesia by infusing propofol (5 mg/kg, i.v.) before and after the completion of self-administration experiments. Only animals showing rapid anesthesia were included in analyses.



<span id="page-4-1"></span><span id="page-4-0"></span>TABLE 1 Attrition rates for CHRNA6 3'-UTR SNP rats in the study.



#### 2.6. Nicotine intravenous self-administration and extinction

Animals (PN 34) intravenous self-administered (IVSA) nicotine (0.015 mg/kg/infusion) at an FR5 schedule for 1-h daily session for a minimum of 5 days, or until they reached stable responding (Reinforced responses (Reinf) within 20% of the mean over the last 3 days;  $R > 2 \times$  Non-Reinforced responses;  $R > 5$ ) [\(48\)](#page-11-13) (**[Figure 1](#page-4-0)**). A compound stimulus, light and tone, were paired upon delivery of nicotine infusion. A dose of 0.015 mg/kg/infusion was chosen based on previous adult and adolescent studies [\(54,](#page-11-18) [55\)](#page-11-19). Baseline responding was defined as the average reinforced responses over the last three days of selfadministration. Rats were then allowed to respond to at the dose of 0.015 mg/kg/infusion on Progressive Ratio (PR) schedule (∼PN 39) (**[Figure 1](#page-4-0)**). The PR schedule of reinforcement is a measure of motivation to obtain the drug [\(60\)](#page-11-23). The sequence was determined using the exponential formula (5 exp (0.2  $\times$  infusion

number)-5) such that the required responses per infusion were as followed: 1, 2, 4, 6, 9, 12, 15, 20, 25, 32, 40, 50, 62, 77, 95, 118, 145, 178, 219, 268, 328, 402, 492 [\(60\)](#page-11-23). PR conditions were the same for FR sessions, with the exception that the sessions were 4 h duration. Breakpoint was achieved when >20 min of inactivity on the active lever elapsed. After reaching stable responding and two-day of PR schedule extinction-reinstatement testing began.

During extinction (∼PN 41), animals were placed in the same operant testing chambers the animals were not connected to the infusion tubing, the house light remained on and responses on the levers were counted but had no consequences, as such no cues or reward were delivered (**[Figure 1](#page-4-0)**). Extinction sessions were 1-h per day for a minimum of 5 days, or until responding was reduced to 25% of baseline [\(48\)](#page-11-13).

#### 2.7. Cue and nicotine-induced reinstatement

After meeting extinction criteria, reinstatement testing began (∼PN 47) [\(48\)](#page-11-13) (**[Figure 1](#page-4-0)**). Nicotine-seeking was reinstated using three reinstatement conditions given in a within-subjects counterbalanced design: Cues, nicotine-primed alone, and nicotine-primed paired with cues. Presentation of cues consisted of cue light illumination and sound in the testing chamber. Nicotine-prime injections contained 0.15 mg/kg nicotine and was administered intraperitoneally immediately before the reinstatement test. The nicotine-prime dose was chosen based on previous work [\(26,](#page-11-24) [54\)](#page-11-18). Between reinstatement tests, animals were returned to extinction condition for a minimum of two days, or until extinction criteria were

met. Reinstatement was defined as a significant increase in responding from the last day of extinction.

#### 2.8. Data analysis

Data were analyzed using JMP (SAS Institute) software [\(48\)](#page-11-13). Food acquisition was analyzed by a compound 4 way multivariate ANOVA for lever presses (Reinf and Non-Reinf)  $\times$  sex (male and female)  $\times$  Genotype (α6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup>)  $\times$  FR schedule (FR1TO1, FR1TO10, FR2TO20, and FR5TO20) with repeated measures on lever presses and FR schedule, with Bonferroni corrected t-test post hoc comparisons. Nicotine self-administration data were analyzed by a compound 4-way multivariate ANOVA Reinf/Non-reinf responses  $\times$  day  $\times$  sex (male and female) genotype (α6 $^{GG}$  and α6<sup>CC</sup>)  $\times$  day (day 3–5) with repeated measures on Reinf/Nonreinf responses and day. Reinstatement data were analyzed as normalized reinforced responding. Mean responses for reinstatement condition were analyzed by a 3-way multivariate ANOVA for sex x genotype ( $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup>)  $\times$  reinstatement condition (cue only, nicotine only, and nicotine plus cues), with repeated measure on reinstatement condition. Significant main effects were further analyzed with 1-way ANOVAs and Bonferroni-corrected paired or unpaired t-tests, as appropriate. Food reinstatement data were analyzed as normalized reinforced responding. Mean responses for reinstatement condition were analyzed by a 3-way multivariate ANOVA for sex x genotype ( $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup>)  $\times$  reinstatement condition (cue only, food only, and food plus cues), with a repeated measure on reinstatement condition. Significant main effects were further analyzed with 1-way ANOVAs and Bonferroni-corrected paired or unpaired t-tests, as appropriate.

### 3. Results

# 3.1. The *CHRNA6* 3'-UTR SNP knock-in does not impact food selfadministration in male and female rats

To facilitate acquisition of lever pressing, adolescent male and female  $\alpha$ 6 $^{\rm GG}$  and  $\alpha$ 6 $^{\rm CC}$  rats were evaluated for food reinforcement at Fixed Ratio (FR)1 timeout (TO)1 and then escalated to higher schedules of reinforcement at FR1TO10, FR2TO20, and FR5TO20. A 4-way ANOVA revealed a significance for Reinf/Non-Reinf lever presses  $[F(1,39) = 1169.4560, p < 0.0001]$  and FR schedule  $[F(3,117) = 295.4014, p < 0.0001]$ . A significant interaction was observed between Reinf/Non-Reinf lever presses x FR schedule  $[F(1,117) = 302.9465, p < 0.0001]$ . Data are collapsed by sex since there was no sex or genotype interaction, but graphs (**[Figures 2A, B](#page-6-0)**) are shown separately by sex and genotype for

clarity. Male and female adolescent  $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup> equally learn to press a lever for a natural reward.

#### 3.2. No sex or genotype differences for intravenous self-administration, progressive ratio, and extinction in male and female adolescent *CHRNA6* 3'-UTR SNP knock-in rats

During nicotine intravenous self-administration there was a significant effect of Reinf/Non-reinf responses for days 1– 5  $[(F1,39) = 183.4430, p = 0.0001]$ , Day  $[F(4,36) = 11.7757,$  $p = 0.0001$ , interaction between Reinf/Non-reinf responses and Day  $[F(4,36) = 19.2790, p = 0.0001]$  ([Figures 3A, B](#page-6-1)). No sex or genotype effects were observed for total nicotine intake (**[Figure 3C](#page-6-1)**) or breakpoint values (**[Figure 3D](#page-6-1)**). Following stable nicotine self-administration, nicotine-seeking behavior was extinguished by removal of nicotine and associated cues. All animals significantly reduced their responding on the reinforced lever beginning on Day 1 and continued throughout extinction (**[Figures 4A, B](#page-7-0)**). No sex differences were observed for days to meet extinction criteria as shown in **[Figure 4C](#page-7-0)**, males and females showing equivalent 25% or on less last day of extinction responding with an average of 6 days.

### 3.3. Sex- and genotype-dependent effects on reinstatement of nicotine in the CHRNA6 3'-UTR SNP adolescent rats

Following extinction, animals were triggered to reinstate to nicotine-seeking behavior with cues-, nicotine- and combination of nicotine plus cues (**[Figures 5A, B](#page-7-1)**). For reinstatement overall ANOVA reveal a sex x genotype interaction  $[F(1,38) = 8.0052, p = 0.0074]$ , data was separated by sex and genotype. Main effects of reinstatement stimuli were found  $[F(3,36) = 26.0462, p < 0.0001]$  and a trend for reinstatement stimuli x sex x genotype  $[(F(3,36) = 2.6865,$  $p = 0.061$ . Further one-way ANOVA for nicotine + cue reveal  $[F(1,39) = 10.0598, p = 0.0030]$  and a one-tail t-test reveal male α6 GG show enhanced nicotine + cue primed reinstatement as compared to  $\alpha$ 6<sup>CC</sup> (*p* = 0.01) (**[Figure 5B](#page-7-1)**). In females,  $\alpha$ 6<sup>CC</sup> rats show a trend for enhanced nicotine + cue primed reinstatement as compared with  $\alpha$ 6<sup>GG</sup> rats ( $p = 0.08$ ) (**[Figure 5A](#page-7-1)**).

### 4. Discussion

We have previously demonstrated equivalent adolescent WT Sprague Dawley male and female behavior in the



#### <span id="page-6-0"></span>FIGURE 2

The human *CHRNA*6 3'-UTR SNP knock-in does not impact food self-administration. Female **(A)** and male **(B)**, α6<sup>CC</sup> and α6<sup>GG</sup>, mean daily 30 min responses ± SEM for food self-administration at Fixed Ratio (FR)1 Time out (TO)1, FR2TO10, FR2TO20, FR5TO20 schedules of reinforcement. ∗∗∗*p* = 0.0001 Reinforced (Rein) vs. Non-Reinforced (Non-reinf) responses. *N* = 8–14/group. Circles represent females and squares represents males.



<span id="page-6-1"></span>acquisition of a natural and drug reward and nicotine seeking behavior using a reinstatement model [\(48\)](#page-11-13). In the current study, we investigate the role of humanized CHRNA6 3'-UTR SNP knock-in in male and female adolescent rats. Our results show no sex or genotype effects for food selfadministration, nicotine self-administration, progressive ratio, and extinction. We observe a sex and genotype bidirectional effect during reinstatement testing specifically, α6GG males



<span id="page-7-0"></span>

<span id="page-7-1"></span>exhibit enhanced nicotine + cues reinstatement when compared to α6<sup>CC</sup> males and a trending for α6<sup>CC</sup> when compared to α6 GG females. Our study is the first to use an in vivo translational model of nicotine behavioral phenotype during adolescence. The current study uses an age range for male and female rats between PN 28–42 as the typical period of adolescence [\(53\)](#page-11-17). Ages in the "gray zone," i.e., earlier than PN 28 and later PN 42 have been considered as part of adolescence up to around PN 60 [\(53\)](#page-11-17). Thus, our studies were done during adolescence, a gradual transition period of soft events with sexual maturity as part of a developmental milestone [\(53\)](#page-11-17).

#### 4.1. The CHRNA6 3'-UTR SNP knock-in does not influence a natural reward

The CHRNA6 3'-UTR SNP knock-in does not influence the ability for male and female,  $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup>, adolescents to acquire and maintain food self-administration. Our results are in accord with our recent manuscript using a separate cohort characterizing the CHRNA6 3'-UTR SNP knock-in rats [\(47\)](#page-11-12) and in adolescent wild type Sprague Dawley rats [\(48\)](#page-11-13). These studies show that age matched male and female wild type and CHRNA6 3'-UTR SNP knock-in rats do not exhibit sex effects. Comparing food reinforcement between CHRNA6 3'-UTR SNP and wild type rats show similar behavior at different schedules of reinforcement, with the exception of slightly higher FR5TO20 for the 3'-UTR SNP rats [\(48\)](#page-11-13). Assessing palatable food conditioned place preference (CPP) in α6 KO and WT C57BL/6J(B6) mice, exhibited similar place preference scores for the context associated with palatable food suggesting the α6 inactivation does not result in the incentive value of a natural reward [\(61\)](#page-11-25). Intra-VTA perfusion of α-CntxMII did not yield an effect on food selfadministration in adult male rats on a fixed-ratio schedule of reinforcement [\(15\)](#page-10-20). Additionally, administration of a range of doses for r-bPiDI (2.47-74 µmol/kg; s.c.), a selective small molecule antagonist of α6β2<sup>\*</sup> nAChR, a tertiary amino analog, 1,10-bis(3-methyl-5-6,-dihydropyridin-1(2H)decane (r-bPiDI), derived from N,N-decane-1,10-diyl-bis-3-picolinium diiodide (bPiDI), did not alter the number of food pellets earned in male rats [\(62\)](#page-11-26). Future studies should examine the role of a6<sup>∗</sup> nAChR subunit in food-seeking behavior given the mechanism for food self-administration may differ from food-induced reinstatement.

## 4.2. The CHRNA6 3'-UTR SNP knock-in does not influence nicotine intravenous self-administration, progressive ratio, and extinction

In our current studies, the CHRNA6 3'-UTR SNP knock-in does not influence nicotine self-administration in adolescent (PN 34) males and females  $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup> at a dose of 0.015 mg/kg/injection on a fixed ratio and progressive ratio schedule of reinforcement. Our results are consistent with adolescent male and female Sprague Dawley WT rats (PN 34), which exhibit similar nicotine self-administration in an equivalent paradigm and show lack of sex effect [\(48\)](#page-11-13). Comparing CHRNA6 3-UTR SNP to age matched wild type rats show higher nicotine self-administration and nicotine intake [\(48\)](#page-11-13). This is likely influenced by the higher responses at FR5TO20 for food reinforcement prior to nicotine selfadministration, as progressive ratio values were similar between CHRNA6 3'-UTR SNP and age matched wild type

rats [\(48\)](#page-11-13). Decreased nicotine self-administration in both, fixed ratio and progressive ratio have been observed with microinjections into either the nucleus accumbens shell or the ventral tegmental area with an α6β2 <sup>∗</sup> nAChR antagonist in rats [\(15,](#page-10-20) [21\)](#page-10-15). Intra-VTA α-CntxMII pretreatment significantly reduced responding for nicotine self-administration at a dose of 0.03 mg/kg/infusion in adult male WT Sprague Dawley rats suggesting the role of α6<sup>\*</sup> nAChR in nicotinereinforcing properties [\(15\)](#page-10-20). Further, in adult rats, nicotine self-administration was decreased with a selective antagonist of α6β2 <sup>∗</sup> nAChR, r-bPiDI, at a dose of 0.03 mg/kg/infusion [\(62\)](#page-11-26). Additionally, bPiDI prevented the acquisition of nicotine self-administration in WT and α4-S248F mice, mutant α4 <sup>∗</sup> nAChR that are insensitive to blockade by mecamylamine, a non-selective nicotinic receptor antagonist [\(63\)](#page-11-27). These studies suggest the involvement in  $\alpha 6^*$  nAChR in nicotine reinforcement, particularly at higher nicotine doses that were not assessed in our current studies. Future studies are needed to examine dose-, sex-, or genotypedependent effects, along with  $α6*$  nAChR blockade to suppress nicotine self-administration in the adolescent humanized CHRNA6 3'-UTR SNP rats. Whereas our results suggest that adolescent male and female  $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup> have equivalent nicotine self-administration and reward response at the 0.015 mg/kg/injection, a dose response is warranted. It is possible that nicotine pharmacodynamics and pharmacokinetic properties may be dose-dependently shifted in male and female humanized CHRNA6 3'-UTR SNP rats, which could be assessed in future studies.

Our results illustrate that male and female adolescent containing the CHRNA6 3'-UTR SNP exhibit equivalent extinction learning. These effects are similar to age matched wild type rats [\(48\)](#page-11-13). Both male and females,  $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup>, exhibited extinguished lever-pressing behavior beginning at day 1 and decreasing over time. Self-administration, extinction/reinstatement paradigm in adolescent males (PN 45) and young adult (PN 58) provides an understanding of the molecular mechanisms of extinction learning revealing key structures involved including the mPFC, OFC, Nucleus accumbens (NAc, core and shell) and amygdala [\(64\)](#page-11-28). Further research is needed to understand how dose-dependent nicotine levels alter learning and memory processing during extinction in the humanized male and female CHRNA6 3'-UTR SNP rats.

### 4.3. Sex- and genotype dependent effects with a greater impact in  $\alpha$ 6<sup>GG</sup> males versus  $\alpha$ 6<sup>CC</sup> males

For nicotine-induced reinstatement, male  $\alpha$ 6<sup>GG</sup> and female α6 CC rats illustrated enhanced nicotine combined with cue responding with males being more impacted. Cue only failed

to reinstate responding in all groups. In age matched wild type rats, cue-induced reinstatement was observed in both males and females [\(48\)](#page-11-13). Male  $\alpha$ 6<sup>GG</sup> and female  $\alpha$ 6<sup>CC</sup> rats show that a non-contingent administration of nicotine during extinction of nicotine self-administration reinstates responding. These results are in accord with our recent studies using adolescent male and female wild type Sprague Dawley rats [\(48\)](#page-11-13). There is mounting evidence for significant straindependent differences for nicotine self-administration [\(65,](#page-11-29) [66\)](#page-12-0). Whereas we observe no sex or genotype differences for natural-, drug-reward and extinction between wild type Sprague Dawley and the CHRNA6 3'-UTR SNP rats with a Sprague Dawley genetic background, our reinstatement results suggest otherwise. In particular, male  $\alpha$ 6<sup>GG</sup> rats show the greatest nicotine + cue-induced reinstatement as compared to age matched wild type rats [\(48\)](#page-11-13). The strain difference between the WT Sprague Dawley and the CHRNA6 3'-UTR SNP is the replacement of the rat CHRNA6 3'-UTR with the human CHRNA6 3'-UTR. Polymorphisms in 3'-UTR are known to be associated with neurological disorders and behaviors via involvement of micro(mi)RNAs and RNA binding protein (RBP) by influencing mRNA translation [\(67,](#page-12-1) [68\)](#page-12-2). The human CHRNA6 3'-UTR may bind to miRNA present in humans, but not rodents. The technology to understand the mechanism of the CHRNA6 3'-UTR across species in mediating nicotine-induced behavioral sensitization is limited [\(69\)](#page-12-3).

Assessing α6 <sup>∗</sup> nAChR mRNA and protein expression could shed light into lack of sex or genotype behavioral differences. Sub-chronically treated humanized CHRNA6 3'-UTR SNP rats, following a 4-day nicotine pretreatment paradigm, found no sex-, genotype- or nicotine-dependent alterations in α6 nAChR mRNA expression; however, behavior test showed α6<sup>CC</sup> females and α6<sup>GG</sup> males exhibited nicotine induced locomotor and anxiolytic behavior compared to their salinetreated counterparts [\(47\)](#page-11-12). Further, in adult male and female mice with fluorescently tagged α4 and α6 nAChRs showed no sex differences for CPP scores at 0.5 mg/kg nicotine s.c. however, there was a significant correlation between upregulation of α4α6, but not α6 (non-α4) nAChRs and reward related behavior in males versus females [\(70\)](#page-12-4). Future studies are essential in assessing neuronal activation, α6<sup>\*</sup> nAChR mRNA and protein expression following nicotine + cue reinstatement in adolescent CHRNA6 3'-UTR SNP male and female rats.

# 5. Conclusion

Taken together, our data suggest no genotype and/or sex effects were observed for natural food reinforcement, five-day total drug intake for nicotine self-administration, progressive ratio schedules of reinforcement, extinction behavior, or other parameters of reinstatement (i.e., cueprimed). Our results suggest that adolescent male  $\alpha$ 6<sup>GG</sup> (and potentially female  $\alpha$ 6<sup>CC</sup>) rats may be at risk for addictive effects as assessed by reinstatement paradigm. Future studies will need to evaluate the role of nicotine dose, age, and sex-dependent effects during adulthood. Further assessments of the neurobiological mechanisms involved are also warranted in order to identify what drives the nicotine-induced effects in our CHRNA6 3'-UTR SNP line in a sex- and genotype dependent manner. Such work may assist in translational studies in humans for improved prevention and intervention strategies to curb nicotine-seeking behavior.

#### Data availability statement

The original contributions presented in this study are included in the article/**[Supplementary material](#page-10-19)**, further inquiries can be directed to the corresponding author.

#### Ethics statement

This animal study was reviewed and approved by Institutional Animal Care and Use Committee of the University of California, Irvine.

#### Author contributions

DC collected data and conducted the literature review. Both authors conceived the study design, conducted data analysis, contributed to the manuscript writing, contributed to the critical revision of data analysis, and approved the final version of the manuscript.

#### Funding

This research was supported by the Tobacco and Related Disease Research Program (TRDRP) award (22RT-0103 and T31IP1427) (SL), UCI School of Medicine Startup Funds (SL), and Institute of Clinical Translational Science (ICTS) Pilot Award (TR001414) (SL). DC was supported by the University of California, Irvine, Summer Inclusive Excellence Ambassador Program as well as the Vertex Pharmaceutical Fellowship from the UCI School of Pharmacy and Pharmaceutical Sciences.

#### Acknowledgments

We thank Frances Leslie, James Belluzzi, and Emily Castro with their assistance on this manuscript.

We also thank Michelle Cano, Marisela Martinez, Angelica Cardenas, and Antonella Facundo for their technical assistance and discussion.

### Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

#### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or

#### References

<span id="page-10-0"></span>1. Miech R, Johnston L, O'malley P, Bachman J, Schulenberg J, Patrick M. Monitoring the Future National Survey Results on Drug Use, 1975-2021: Volume 1, Secondary School Students. Ann Arbor, MI: Institute for Social Research (2022).

<span id="page-10-1"></span>2. Wang T, Neff LJ, Park-Lee E, Ren C, Cullen K, King B. E-cigarette use among middle and high school students — United States, 2020. JAMA J Am Med Assoc. (2019) 322:2095–103.

<span id="page-10-2"></span>3. Soneji S, Barrington-Trimis J, Wills T, Leventhal A, Unger J, Gibson L, et al. Association between initial use of E-cigarettes and subsequent cigarette smoking among adolescents and young adults a systematic review and meta-analysis. JAMA Pediatr. (2017) 171:788–97. [doi: 10.1001/jamapediatrics.201](https://doi.org/10.1001/jamapediatrics.2017.1488) 7.1[488](https://doi.org/10.1001/jamapediatrics.2017.1488)

4. Ren M, Lotfipour S. Nicotine gateway effects on adolescent substance use. Western J Emerg Med. (2019) 20:696–709. [doi: 10.5811/westjem.2019.7.41661](https://doi.org/10.5811/westjem.2019.7.41661)

5. Creamer M, Dutra L, Sharapova S, Gentzke A, Delucchi K, Smith R, et al. Effects of e-cigarette use on cigarette smoking among U.S. youth, 2004–2018. Prev Med (Baltim). (2021) 142:106316. [doi: 10.1016/j.ypmed.2020.106316](https://doi.org/10.1016/j.ypmed.2020.106316)

<span id="page-10-3"></span>6. Yuan M, Cross S, Loughlin S, Leslie F. Nicotine and the adolescent brain. J Physiol. (2015) 593:3397–412. [doi: 10.1113/JP270492](https://doi.org/10.1113/JP270492)

<span id="page-10-4"></span>7. DiFranza J, Savageau J, Fletcher K, O'Loughlin J, Pbert L, Ockene J, et al. Symptoms of tobacco dependence after brief intermittent use: the development and assessment of nicotine dependence in youth-2 study. Arch Pediatr Adolesc Med. (2007) 161:704–10. [doi: 10.1001/archpedi.161.7.704](https://doi.org/10.1001/archpedi.161.7.704)

<span id="page-10-5"></span>8. Wang T, Gentzke A, Creamer M. Tobacco product use and associated factors among middle and high school students. MMWR Surveill Summ. (2019) 68:1–22. [doi: 10.15585/mmwr.ss6812a1](https://doi.org/10.15585/mmwr.ss6812a1)

<span id="page-10-6"></span>9. American Medical Association. Letters Failed Attempts to Quit Combustible Cigarettes And E-Cigarettes Among US Adolescents. Chicago, IL: American Medical Association (2022).

<span id="page-10-7"></span>10. le Novère N, Corringer P, Changeux J. The diversity of subunit composition in nAChRs: evolutionary origins, physiologic and pharmacologic consequences. J Neurobiol. (2002) 53:447–56. [doi: 10.1002/neu.10153](https://doi.org/10.1002/neu.10153)

<span id="page-10-8"></span>11. Tapper A, Mckinney S, Nashmi R, Schwarz J, Deshpande P, Labarca C, et al. Nicotine activation of a4<sup>\*</sup> receptors: sufficient for reward, tolerance, and sensitization. Science. (2004) 306:1029–32. [doi: 10.1126/science.1099420](https://doi.org/10.1126/science.1099420)

12. Corrigal W, Franklin K, Coen K, Clarke P. The mesolimbic dopaminergic system is implicated in the reinforcing effects of nicotine. Psychopharmacology.  $(1991)$  107:285-9.

<span id="page-10-9"></span>13. Picciotto M, Zoli M, Rimondini R, Lena C, Marubio L, Pich E, et al. Acetylcholine receptors containing the B2 subunit are involved in the reinforcing properties of nicotine. Nature. (1998) 391:173–7. [doi: 10.1038/34413](https://doi.org/10.1038/34413)

claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

# <span id="page-10-19"></span>Supplementary material

The Supplementary Material for this article can be found online at: [https://www.frontiersin.org/articles/10.3389/](https://www.frontiersin.org/articles/10.3389/fpsyt.2022.1064211/full#supplementary-material) [fpsyt.2022.1064211/full#supplementary-material](https://www.frontiersin.org/articles/10.3389/fpsyt.2022.1064211/full#supplementary-material)

#### SUPPLEMENTARY FIGURE 1

Body weight (g) across postnatal days. Male and female  $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup>. Circles represent females and squares represent males.

#### SUPPLEMENTARY FIGURE 2

Postnatal day across nicotine self-administration, extinction, and reinstatement. No sex differences for the age of initiation of behavior for nicotine self-administration, progressive ratio, extinction, and reinstatement. Mean  $\pm$  SEM postnatal days across study conditions for male and female  $\alpha$ 6<sup>GG</sup> and  $\alpha$ 6<sup>CC</sup>. Circles represent females and squares represents males.

<span id="page-10-11"></span>14. Changeux J, Bertrand D, Corringer P, Dehaene S, Edelstein S, Léna C, et al. Brain nicotinic receptors: structure and regulation, role in learning and reinforcement. Brain Res Rev. (1998) 26:198–216.

<span id="page-10-20"></span>15. Gotti C, Guiducci S, Tedesco V, Corbioli S, Zanetti L, Moretti M, et al. Nicotinic acetylcholine receptors in the mesolimbic pathway: primary role of ventral tegmental area α6β2<sup>\*</sup> receptors in mediating systemic nicotine effects on dopamine release, locomotion, and reinforcement. J Neurosci. (2010) 30:5311–25. [doi: 10.1523/](https://doi.org/10.1523/JNEUROSCI.5095-09.2010) JN[EUROSCI.5095-09.2010](https://doi.org/10.1523/JNEUROSCI.5095-09.2010)

16. Champtiaux N, Gotti C, Cordero-Erausquin M, David D, Przybylski C, Léna C, et al. Subunit composition of functional nicotinic receptors in dopaminergic neurons investigated with knock-out mice. J Neurosci. (2003) 23:7820–9. [doi: 10.](https://doi.org/10.1523/JNEUROSCI.23-21-07820.2003) 152[3/JNEUROSCI.23-21-07820.2003](https://doi.org/10.1523/JNEUROSCI.23-21-07820.2003)

<span id="page-10-10"></span>17. Salminen O, Drapeau J, McIntosh J, Collins A, Marks M, Grady S. Pharmacology of α-conotoxin MII-sensitive subtypes of nicotinic acetylcholine receptors isolated by breeding of null mutant mice. Mol Pharmacol. (2007) 71:1563– 71. [doi: 10.1124/mol.106.031492](https://doi.org/10.1124/mol.106.031492)

<span id="page-10-12"></span>18. Gotti C, Moretti M, Gaimarri A, Zanardi A, Clementi F, Zoli M. Heterogeneity and complexity of native brain nicotinic receptors. Biochem Pharmacol. (2007) 74:1102-11.

<span id="page-10-13"></span>19. Azam L, Chen Y, Leslie F. Developmental regulation of nicotinic acetylcholine receptors within midbrain dopamine neurons. Neuroscience. (2007) 23:1347–60.

<span id="page-10-14"></span>20. Pons S, Fattore L, Cossu G, Tolu S, Porcu E, McIntosh J, et al. Crucial role of α4 and α6 nicotinic acetylcholine receptor subunits from ventral tegmental area in systemic nicotine self-administration. J Neurosci. (2008) 28:12318–27. [doi:](https://doi.org/10.1523/JNEUROSCI.3918-08.2008) 10.[1523/JNEUROSCI.3918-08.2008](https://doi.org/10.1523/JNEUROSCI.3918-08.2008)

<span id="page-10-15"></span>21. Brunzell D, Boschen K, Hendrick E, Beardsley P, Mcintosh J. Alpha; conotoxin MII-sensitive nicotinic acetylcholine receptors in the nucleus accumbens shell regulate progressive ratio responding maintained by nicotine. Neuropsychopharmacology. (2010) 35:665-73. [doi: 10.1038/npp.2009.171](https://doi.org/10.1038/npp.2009.171)

<span id="page-10-16"></span>22. Dwoskin L, Pivavarchyk M, Joyce B, Neugebauer N, Zheng G, Zhang Z, et al. Targeting reward-relevant nicotinic receptors in the discovery of novel pharmacotherapeutic agents to treat tobacco dependence. Nebr Symp Motiv. (2009) 55:31–63. [doi: 10.1007/978-0-387-78748-0\\_4](https://doi.org/10.1007/978-0-387-78748-0_4)

<span id="page-10-17"></span>23. Löf E, Olausson P, DeBejczy A, Stomberg R, McIntosh J, Taylor J, et al. Nicotinic acetylcholine receptors in the ventral tegmental area mediate the dopamine activating and reinforcing properties of ethanol cues. Psychopharmacology. (2007) 195:333–43. [doi: 10.1007/s00213-00](https://doi.org/10.1007/s00213-007-0899-4) 7-0[899-4](https://doi.org/10.1007/s00213-007-0899-4)

<span id="page-10-18"></span>24. Brunzell D. Preclinical evidence that activation of mesolimbic alpha 6 subunit containing nicotinic acetylcholine receptors supports nicotine addiction phenotype. Nicotine Tob Res. (2012) 14:1258–69. [doi: 10.1093/ntr/nts089](https://doi.org/10.1093/ntr/nts089)

<span id="page-11-0"></span>25. Chiamulera C. Cue reactivity in nicotine and tobacco dependence: a "multiple- action" model of nicotine as a primary reinforcement and as an enhancer of the effects of smoking-associated stimuli. Brain Res Rev. (2005) 48:74–97. [doi:](https://doi.org/10.1016/j.brainresrev.2004.08.005) 10.[1016/j.brainresrev.2004.08.005](https://doi.org/10.1016/j.brainresrev.2004.08.005)

<span id="page-11-24"></span>26. Shaham Y, Shalev U, Lu L, de Wit H, Stewart J. The reinstatement model of drug relapse: history, methodology and major findings. Psychopharmacology. (2003)  $168:3 - 20$ 

<span id="page-11-1"></span>27. Epstein D, Preston K, Stewart J, Shaham Y. Toward a model of drug relapse: an assessment of the validity of the reinstatement procedure. Psychopharmacology. (2006) 189:1–16. [doi: 10.1007/s00213-006-0529-6](https://doi.org/10.1007/s00213-006-0529-6)

<span id="page-11-2"></span>28. Pugach O, Cannon D, Weiss R, Hedeker D, Mermelstein R. Classification tree analysis as a method for uncovering relations between CHRNA5A3B4 and CHRNB3A6 in predicting smoking progression in adolescent smokers. Nicotine Tob Res. (2017) 19:410–6. [doi: 10.1093/ntr/ntw197](https://doi.org/10.1093/ntr/ntw197)

29. Cannon D, Mermelstein R, Hedeker D, Coon H, Cook E, McMahon W, et al. Effect of neuronal nicotinic acetylcholine receptor genes (CHRN) on longitudinal cigarettes per day in adolescents and young adults. Nicotine Tob Res. (2014) 16:137– 44. [doi: 10.1093/ntr/ntt125](https://doi.org/10.1093/ntr/ntt125)

30. Fletcher J. Why have tobacco control policies stalled? Using genetic moderation to examine policy impacts. PLoS One. (2012) 7:e50576. [doi: 10.1371/](https://doi.org/10.1371/journal.pone.0050576) jou[rnal.pone.0050576](https://doi.org/10.1371/journal.pone.0050576)

31. Pedneault M, Labbe A, Roy-Gagnon M, Low N, Dugas E, Engert J, et al. The association between CHRN genetic variants and dizziness at first inhalation of cigarette smoke. Addict Behav. (2014) 39:316–20. [doi: 10.1016/j.addbeh.2013.](https://doi.org/10.1016/j.addbeh.2013.\break08.033) \br[eak08.033](https://doi.org/10.1016/j.addbeh.2013.\break08.033)

32. Thorgeirsson T, Gudbjartsson D, Surakka I, Vink J, Amin N, Geller F, et al. Sequence variants at CHRNB3-CHRNA6 and CYP2A6 affect smoking behavior. Nat Genet. (2010) 42:448–53.

<span id="page-11-7"></span>33. Lee W, Bergen A, Swan G, Li D, Liu J, Thomas P, et al. Gender-stratified gene and gene-treatment interactions in smoking cessation. Pharmacogenomics J. (2012) 12:521–32. [doi: 10.1038/tpj.2011.30](https://doi.org/10.1038/tpj.2011.30)

<span id="page-11-6"></span>34. Lotfipour S, Leonard G, Perron M, Pike B, Richer L, Séguin J, et al. Prenatal exposure to maternal cigarette smoking interacts with a polymorphism in the α6 nicotinic acetylcholine receptor gene to influence drug use and striatum volume in adolescence. Mol Psychiatry. (2010) 15:6–8. [doi: 10.1038/mp.2009.63](https://doi.org/10.1038/mp.2009.63)

35. Hoft N, Corley R, McQueen M, Huizinga D, Menard S, Ehringer M. SNPs in CHRNA6 and CHRNB3 are associated with alcohol consumption in a nationally representative sample. Genes Brain Behav. (2009) 8:631–7. [doi: 10.1111/j.1601-183X.](https://doi.org/10.1111/j.1601-183X.2009.00495.x) 200[9.00495.x](https://doi.org/10.1111/j.1601-183X.2009.00495.x)

36. Hoft N, Corley R, McQueen M, Schlaepfer I, Huizinga D, Ehringer M. Genetic association of the CHRNA6 and CHRNB3 genes with tobacco dependence in a nationally representative sample. Neuropsychopharmacology. (2009) 34:698–706. [doi: 10.1038/npp.2008.122](https://doi.org/10.1038/npp.2008.122)

37. Zeiger J, Haberstick B, Schlaepfer I, Collins A, Corley R, Crowley T, et al. The neuronal nicotinic receptor subunit genes (CHRNA6 and CHRNB3) are associated with subjective responses to tobacco. Hum Mol Genet. (2008) 17:724-34. [doi: 10.](https://doi.org/10.1093/hmg/ddm344) 109[3/hmg/ddm344](https://doi.org/10.1093/hmg/ddm344)

38. Ehringer M, McQueen M, Hoft N, Saccone N, Stitzel J, Wang J, et al. Association of CHRN genes with "dizziness" to tobacco. Am J Med Genet B Neuropsychiatr Genet. (2010) 153:600–9.

<span id="page-11-3"></span>39. Rigbi A, Kanyas K, Yakir A, Greenbaum L, Pollak Y, Ben-Asher E, et al. Why do young women smoke? V. Role of direct and interactive effects of nicotinic cholinergic receptor gene variation on neurocognitive function. Genes Brain Behav. (2008) 7:164–72. [doi: 10.1111/j.1601-183X.2007.00329.x](https://doi.org/10.1111/j.1601-183X.2007.00329.x)

<span id="page-11-4"></span>40. Mayya V, Duchaine T. Ciphers and executioners: how 3'-untranslated regions determine the fate of messenger RNAs. Front Genet. (2019) 10:6. [doi: 10.3389/fgene.](https://doi.org/10.3389/fgene.2019.00006) 201[9.00006](https://doi.org/10.3389/fgene.2019.00006)

<span id="page-11-5"></span>41. Mayr C. What are 3 ' UTRs doing? Cold Springs Harb Perpect Biol. (2019) 11:a034728. [doi: 10.1101/cshperspect.a034728](https://doi.org/10.1101/cshperspect.a034728)

<span id="page-11-8"></span>42. Moen J, Lee A. Sex differences in the nicotinic acetylcholine receptor system of rodents: impacts on nicotine and alcohol reward behaviors. Front Neurosci. (2021) 15:745783. [doi: 10.3389/fnins.2021.745783](https://doi.org/10.3389/fnins.2021.745783)

<span id="page-11-11"></span>43. Bureau A, Liu G, Lu Q, Zhang X, Lan T, Wang T, et al. Considering genetic heterogeneity in the association analysis finds genes associated with nicotine dependence. Front Genet. (2019) 10:448. [doi: 10.3389/fgene.2019.00448](https://doi.org/10.3389/fgene.2019.00448)

44. Moen J, DeBaker M, Myjak J, Wickman K, Lee A. Bidirectional sex-dependent regulation of α6 and β3 nicotinic acetylcholine receptors by protein kinase Cε. Addict Biol. (2021) 26:e12954.

<span id="page-11-9"></span>45. Wieskopf J, Mathur J, Limapichat W, Post M, Al-Qazzaz M, Sorge R, et al. The nicotinic a6 subunit gene determines variability in chronic pain sensitivity via cross-inhibition of P2X2/3 receptors. Sci Transl Med. (2015) 7:287ra72. [doi:](https://doi.org/10.1126/scitranslmed.3009986) 10.[1126/scitranslmed.3009986](https://doi.org/10.1126/scitranslmed.3009986)

<span id="page-11-10"></span>46. Wen L, Han H, Liu Q, Su K, Yang Z, Cui W, et al. Significant association of the CHRNB3-CHRNA6 gene cluster with nicotine dependence in the Chinese Han population. Sci Rep. (2017) 7:9745. [doi: 10.1038/s41598-017-09492-8](https://doi.org/10.1038/s41598-017-09492-8)

<span id="page-11-12"></span>47. Cardenas A, Bai Y, Hajy Heydary Y, Li J, Leslie F, Lotfipour S. Sex-and genotype-dependent nicotine-induced behaviors in adolescent rats with a human polymorphism (rs2304297) in the 3-UTR of the CHRNA6 Gene. Int J Mol Sci. (2022) 23:3145. [doi: 10.3390/ijms23063145](https://doi.org/10.3390/ijms23063145)

<span id="page-11-13"></span>48. Carreño, D, Lotfipour S. Male and female sprague dawley rats exhibit equivalent natural reward, nicotine self-administration, extinction, and<br>reinstatement during adolescent-initiated behaviors. Nicotine Tob Res. (2022) ntac234. [doi: 10.1093/ntr/ntac234](https://doi.org/10.1093/ntr/ntac234) [Epub ahead of print].

<span id="page-11-14"></span>49. Mojica C, Belluzzi J, Leslie F. Age-dependent alterations in reward-seeking behavior after brief nicotine exposure. Psychopharmacology. (2014) 231:1763–73. [doi: 10.1007/s00213-013-3266-7](https://doi.org/10.1007/s00213-013-3266-7)

<span id="page-11-15"></span>50. Donny E, Caggiula A, Rowell P, Gharib M, Maldovan V, Booth S, et al. Nicotine self-administration in rats: estrous cycle effects, sex differences and nicotinic receptor binding. Psychopharmacology. (2000) 151:392–405. [doi: 10.1007/](https://doi.org/10.1007/s002130000497) s00[2130000497](https://doi.org/10.1007/s002130000497)

51. Feltenstein M, Ghee S, See R. Nicotine self-administration and reinstatement of nicotine-seeking in male and female rats. Drug Alcohol Depend. (2012) 121:240–6. [doi: 10.1016/j.drugalcdep.2011.09.001](https://doi.org/10.1016/j.drugalcdep.2011.09.001)

<span id="page-11-16"></span>52. Lynch W. Sex and ovarian hormones influence vulnerability and motivation for nicotine during adolescence in rats. Pharmacol Biochem Behav. (2009) 94:43–50. [doi: 10.1016/j.pbb.2009.07.004](https://doi.org/10.1016/j.pbb.2009.07.004)

<span id="page-11-17"></span>53. Spear L. The adolescent brain and age-related behavioral manifestations. Neurosci Biobehav Rev. (2000) 24:417–63. [doi: 10.1016/S0149-7634\(00\)0](https://doi.org/10.1016/S0149-7634(00)00014-2) 001[4-2](https://doi.org/10.1016/S0149-7634(00)00014-2)

<span id="page-11-18"></span>54. Cross S, Reynaga D, Cano M, Belluzzi J, Zaveri N, Leslie F. Differences in mechanisms underlying reinstatement of cigarette smoke extract- and nicotineseeking behavior in rats. Neuropharmacology. (2020) 162:107846. [doi: 10.1016/j.](https://doi.org/10.1016/j.neuropharm.2019.107846) neu[ropharm.2019.107846](https://doi.org/10.1016/j.neuropharm.2019.107846)

<span id="page-11-19"></span>55. Gellner C, Belluzzi J, Leslie F. Self-administration of nicotine and cigarette smoke extract in adolescent and adult rats. Neuropharmacology. (2016) 109:247–53. [doi: 10.1016/j.neuropharm.2016.06.026](https://doi.org/10.1016/j.neuropharm.2016.06.026)

<span id="page-11-20"></span>56. Costello M, Reynaga D, Mojica C, Zaveri N, Belluzzi J, Leslie F. Comparison of the reinforcing properties of nicotine and cigarette smoke extract in rats. Neuropsychopharmacology. (2014) 39:1843–51. [doi: 10.1038/npp.2](https://doi.org/10.1038/npp.2014.31) 014[.31](https://doi.org/10.1038/npp.2014.31)

<span id="page-11-21"></span>57. Cardenas, A, Lotfipour S. Age- and sex-dependent nicotine pretreatment effects on the enhancement of methamphetamine self-administration in spraguedawley rats. Nicotine Tob Res. (2022) 24:1186–92. [doi: 10.1093/ntr/ntab218](https://doi.org/10.1093/ntr/ntab218)

58. Cardenas A, Martinez M, Saenz Mejia A, Lotfipour S. Early adolescent subchronic low-dose nicotine exposure increases subsequent cocaine and fentanyl self-administration in sprague-dawley rats. Behav Pharmacol. (2021) 32:86–91. [doi:](https://doi.org/10.1097/FBP.0000000000000593) 10.[1097/FBP.0000000000000593](https://doi.org/10.1097/FBP.0000000000000593)

<span id="page-11-22"></span>59. Belluzzi J, Wang R, Leslie F. Acetaldehyde enhances acquisition of nicotine self-administration in adolescent rats. Neuropsychopharmacology. (2005) 30:705-12. [doi: 10.1038/sj.npp.1300586](https://doi.org/10.1038/sj.npp.1300586)

<span id="page-11-23"></span>60. Richardson N, Roberts D. Progressive ratio schedules in drug selfadministration studies in rats: a method to evaluate reinforcing efficacy. J Neurosci Methods. (1996) 66:1–11. [doi: 10.1016/0165-0270\(95\)00153-0](https://doi.org/10.1016/0165-0270(95)00153-0)

<span id="page-11-25"></span>61. Sanjakdar S, Maldoon P, Marks M, Brunzell D, Maskos U, Mcintosh J, et al. Differential roles of α6β2<sup>\*</sup> and α4β2<sup>\*</sup> neuronal nicotinic receptors in nicotine- and cocaine-conditioned reward in mice. Neuropsychopharmacology. (2015) 40:350–60. [doi: 10.1038/npp.2014.177](https://doi.org/10.1038/npp.2014.177)

<span id="page-11-26"></span>62. Beckmann J, Meyer A, Pivavarchyk M, Horton D, Zheng G, Smith A, et al. rbPiDI, an α6β2 <sup>∗</sup> nicotinic receptor antagonist, decreases nicotine-evoked dopamine release and nicotine reinforcement. Neurochem Res. (2015) 40:2121–30. [doi: 10.1007/](https://doi.org/10.1007/s11064-015-1680-4) s11[064-015-1680-4](https://doi.org/10.1007/s11064-015-1680-4)

<span id="page-11-27"></span>63. Madsen HB, Koghar HS, Pooters T, Massalas JS, Drago J, Lawrence AJ. Role of α4- and α6-containing nicotinic receptors in the acquisition and maintenance of nicotine self-administration. Addict Biol. (2015) 20:500–12. [doi: 10.1111/adb.12148](https://doi.org/10.1111/adb.12148)

<span id="page-11-28"></span>64. Funk D, Coen K, Tamadon S, Hope B, Shaham Y, Lê A. Role of central amygdala neuronal ensembles in incubation of nicotine craving. J Neurosci. (2016) 36:8612–23. [doi: 10.1523/JNEUROSCI.1505-16.2016](https://doi.org/10.1523/JNEUROSCI.1505-16.2016)

<span id="page-11-29"></span>65. Matta S, Balfour D, Benowitz N, Boyd R, Buccafusco J, Caggiula A, et al. Guidelines on nicotine dose selection for in vivo research. Psychopharmacology. (2007) 190:269–319.

<span id="page-12-0"></span>66. Leyrer-Jackson J, Overby P, Bull A, Marusich J, Gipson C. Strain and sex matters: differences in nicotine self– administration between outbred and recombinase-driver transgenic rat lines HHS public access. Exp Clin Psychopharmacol. (2021) 29:375–84. [doi: 10.1037/pha0000376](https://doi.org/10.1037/pha0000376)

<span id="page-12-1"></span>67. Bae B, Miura P. Emerging roles for 3 UTRs in neurons. Int J Mol Sci. (2020) 21:3413. [doi: 10.3390/ijms21103413](https://doi.org/10.3390/ijms21103413)

<span id="page-12-2"></span>68. Egervari G, Jutras-Aswad D, Landry J, Miller M, Anderson S, Michaelides<br>M, et al. A functional 3′UTR polymorphism (rs2235749) of prodynorphin alters<br>microRNA-365 binding in ventral striatonigral neurons to influence no

and positive reward traits. Neuropsychopharmacology. (2016) 41:2512–20. [doi: 10.](https://doi.org/10.1038/npp.2016.53) 103[8/npp.2016.53](https://doi.org/10.1038/npp.2016.53)

<span id="page-12-3"></span>69. Mayr C. Regulation by 3 -untranslated regions. Annu Rev Genet. (2017) 51:171–94. [doi: 10.1146/annurev-genet-120116-024704](https://doi.org/10.1146/annurev-genet-120116-024704)

<span id="page-12-4"></span>70. Akers AT, Cooper SY, Baumgard ZJ, Casinelli GP, Avelar AJ, Henderson BJ. Disorders of the nervous system upregulation of nAChRs and changes in excitability<br>on VTA dopamine and GABA neurons correlates to changes in nicotine-reward-<br>related behavior. *eNeuro*. (2020) 7:ENEURO.189–120. doi: 10.152 20.[2020](https://doi.org/10.1523/ENEURO.0189-20.2020)