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Title

Syroto Bioprinter

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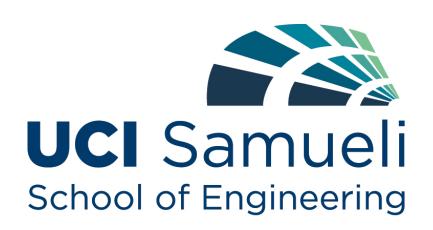
Publication Date

2018-03-15

Supplemental Material

https://escholarship.org/uc/item/2bt1x2j1#supplemental

Peer reviewed



Syroto Bioprinter McKell Davis¹, Ethan Lieberman¹, Sydney Minar¹, Derek Lublin², Alex Schmidt² Mentor: Dr. Elliot Botvinick^{1,3}

Motivation

past fifty years the pharmaceutical industry's Over the productivity in releasing new molecular entities (NME) has remained relatively constant, despite an annual compounding financial investment at approximately 13%¹. Reviews across major pharmaceutical companies attribute an NME's failure to commercialization primarily on lack of efficacy reach (approximately 30%), and safety (approximately 30%)². One approach to validating NMEs is by testing on physiologically relevant in vitro tissues, an intermediate between the current standard of cultured cells and the ideal in vivo organ. Creating a way to streamline research using high sample arrays could accelerate discovery of NME's and repurpose existing ones. Commercialized bioprinters such as Allevi³, 3D-Bioplotter⁴, and Cellink Bio X⁵ feature the pneumatic syringe dispenser but are unable to perform other methods of dispensing such as micro dispensing inkjets, thermoplastic extrusion and mechanical dispensing of whole cell spheroids. Bioprinters with multiprintheads allowing diverse material dispensing are yet to be seen.

Objectives

- Modify the cell dispensing print-head for optimal cell viability and precision.
- Build a novel prototype for fast and reliable printing of biological constructs in a multi-array format (i.e. 96 well array).

References

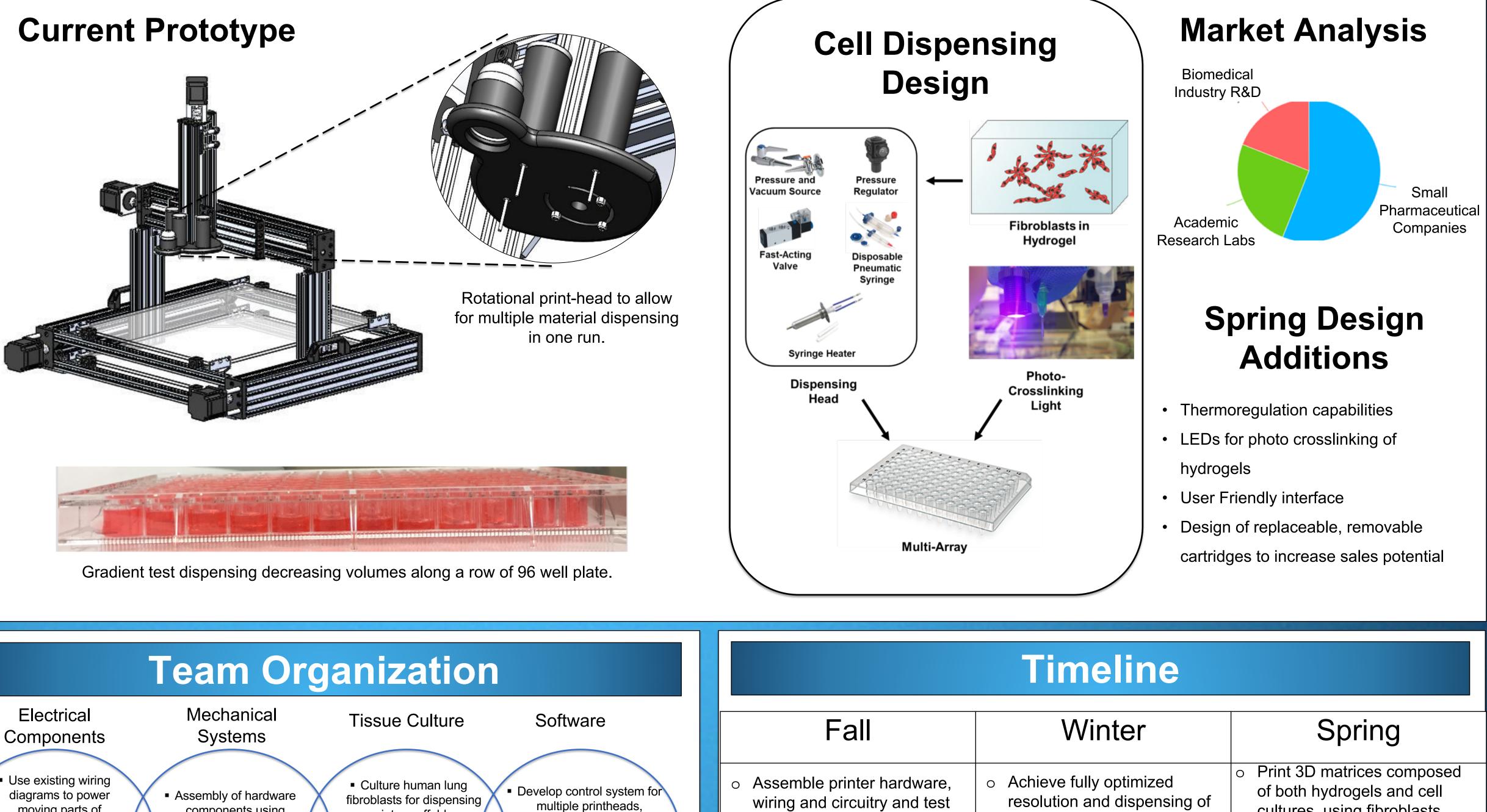
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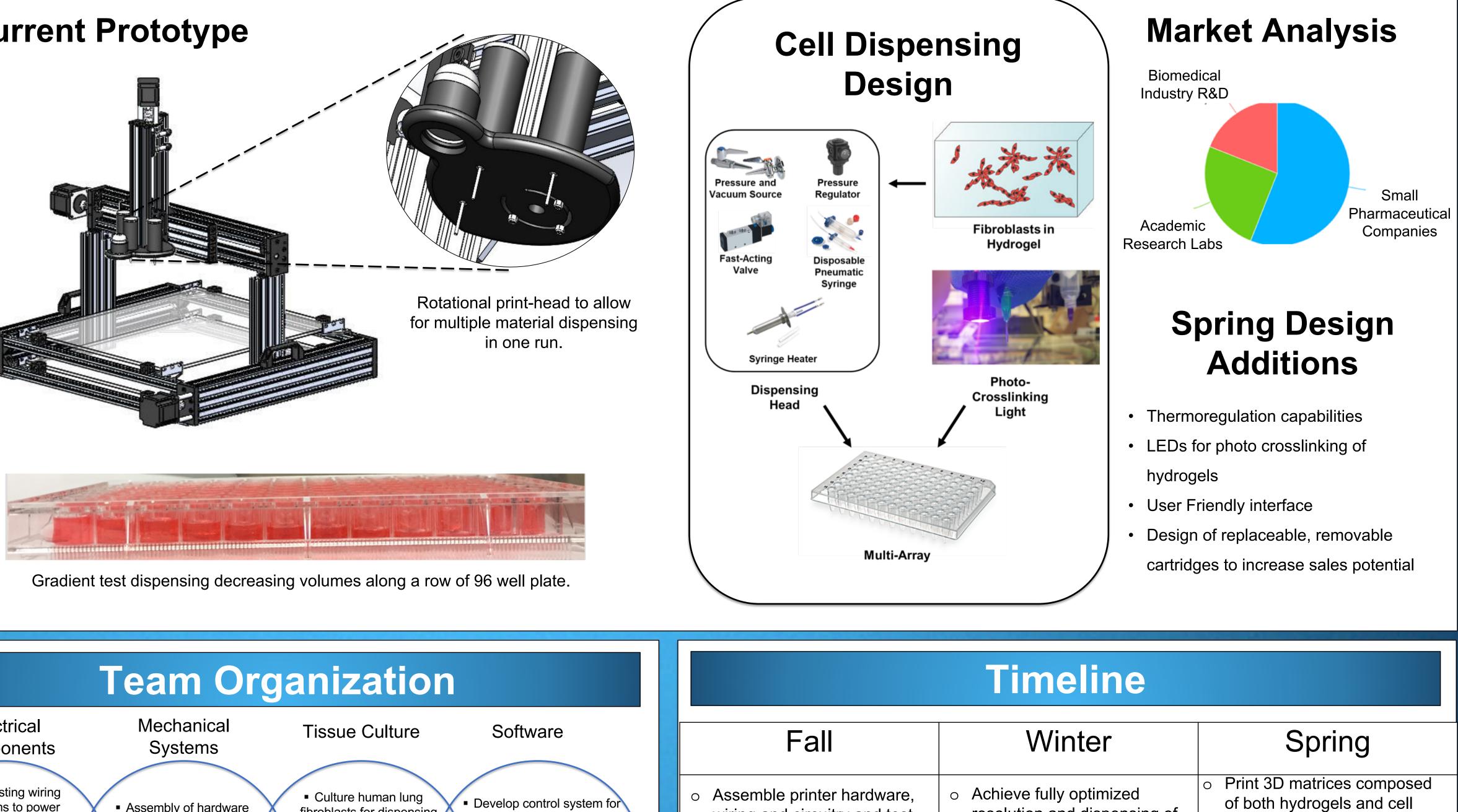
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Acknowledgements



Rachel Hatano, Edwin Shen Graduate Program in Bioengineering and Small Scale Technologies, UC Merced





diagrams to power moving parts of bioprinter

 Create efficient circuit composed of actuators, stepper motors and solenoid valves

Lead: Derek

All team members will contribute to each aspect of the project under direction from respective leads.

Department of Biomedical Engineering 2. Department of Materials Science and Engineering 3. Beckman Laser Institute

Design Process

SketchUp Assembly File Draw additional design features in SolidWorks

components using

- Lead: Alex

Market Research, Lead: Sydney Interviews will be conducted throughout the design process to receive feedback from leading industry representatives to further aid in developing our design.

into scaffold

Fabricate

PEG and alginate

hydrogels for co-culture and

Lead: McKell

dispensing

maximizing resolution of

dispensing

Develop improved user

Optimize current

modules

Lead: Ethan

interface and calibration



Fall	Winter	Spring
 Assemble printer hardware, wiring and circuitry and test with actuated control programming. Use a laser in place of dispensing syringe to determine needs for resolution requirements and adjustment to target into 96 or more well plate. 	 Achieve fully optimized resolution and dispensing of different colored gelatin to test dispensing with a similar material but with a slow polymerization time. 	 Print 3D matrices composed of both hydrogels and cell cultures, using fibroblasts and PEG or alginate, keeping cells alive for at least 3 days in multi-well plates.
	 When testing is sufficiently optimized, we will print target hydrogels in a 96 well tissue culture plate. 	 Assign known drugs to each well and observe distinguishable effects,
		 Finalize project presentation and attend industry night in the spring.