

UC Davis

UC Davis Previously Published Works

Title

Species and population specific gene expression in blood transcriptomes of marine turtles

Permalink

<https://escholarship.org/uc/item/2cb4n5ct>

Journal

BMC Genomics, 22(1)

ISSN

1471-2164

Authors

Banerjee, Shreya M

Stoll, Jamie Adkins

Allen, Camryn D

et al.

Publication Date

2021-12-01

DOI

10.1186/s12864-021-07656-5

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

ARTICLE

Open Access



Species and population specific gene expression in blood transcriptomes of marine turtles

Shreya M. Banerjee¹, Jamie Adkins Stoll¹, Camryn D. Allen^{2,3}, Jennifer M. Lynch⁴, Heather S. Harris³, Lauren Kenyon¹, Richard E. Connon⁵, Eleanor J. Sterling⁶, Eugenia Naro-Maciel⁷, Kathryn McFadden⁸, Margaret M. Lamont⁹, James Benge¹⁰, Nadia B. Fernandez¹, Jeffrey A. Seminoff³, Scott R. Benson^{11,12}, Rebecca L. Lewison¹³, Tomoharu Eguchi³, Tammy M. Summers¹⁴, Jessy R. Hapdei¹⁵, Marc R. Rice¹⁶, Summer Martin², T. Todd Jones², Peter H. Dutton³, George H. Balazs¹⁷ and Lisa M. Komoroske^{1,3*}

Abstract

Background: Transcriptomic data has demonstrated utility to advance the study of physiological diversity and organisms' responses to environmental stressors. However, a lack of genomic resources and challenges associated with collecting high-quality RNA can limit its application for many wild populations. Minimally invasive blood sampling combined with de novo transcriptomic approaches has great potential to alleviate these barriers. Here, we advance these goals for marine turtles by generating high quality de novo blood transcriptome assemblies to characterize functional diversity and compare global transcriptional profiles between tissues, species, and foraging aggregations.

Results: We generated high quality blood transcriptome assemblies for hawksbill (*Eretmochelys imbricata*), loggerhead (*Caretta caretta*), green (*Chelonia mydas*), and leatherback (*Dermochelys coriacea*) turtles. The functional diversity in assembled blood transcriptomes was comparable to those from more traditionally sampled tissues. A total of 31.3% of orthogroups identified were present in all four species, representing a core set of conserved genes expressed in blood and shared across marine turtle species. We observed strong species-specific expression of these genes, as well as distinct transcriptomic profiles between green turtle foraging aggregations that inhabit areas of greater or lesser anthropogenic disturbance.

(Continued on next page)

* Correspondence: ikomoroske@umass.edu

¹Department of Environmental Conservation, University of Massachusetts, Amherst, MA, USA

³Marine Mammal and Turtle Division, Southwest Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, La Jolla, CA, USA

Full list of author information is available at the end of the article



© The Author(s). 2021 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

(Continued from previous page)

Conclusions: Obtaining global gene expression data through non-lethal, minimally invasive sampling can greatly expand the applications of RNA-sequencing in protected long-lived species such as marine turtles. The distinct differences in gene expression signatures between species and foraging aggregations provide insight into the functional genomics underlying the diversity in this ancient vertebrate lineage. The transcriptomic resources generated here can be used in further studies examining the evolutionary ecology and anthropogenic impacts on marine turtles.

Keywords: Comparative transcriptomics, Sea turtle, Minimally invasive sampling, Conservation physiology, RNA-sequencing, Ortholog

Background

Transcriptomics has become a powerful tool to study the underpinnings of ecological and physiological diversity within and between species [1]. In particular, RNA-sequencing can be used to characterize global gene expression and sequence diversity across functional components of the genome. Combined with advances in bioinformatics approaches, high-throughput sequencing has enabled the completion of studies in wild populations with limited genomic resources that were previously not possible. De novo transcriptome assemblies paired with analyses to identify orthologs derived from common ancestral genes have facilitated comparisons of functional diversity and gene expression between closely-related species, especially when reference genomes are not available [2–5]. Additionally, transcriptomics is becoming increasingly employed to complement other methods of assessing physiological responses to environmental conditions, such as hormone assays and blood biochemistry analyses [6–9]. For example, transcriptomics has been used to identify differing physiological responses in urban and rural dwelling great tits (*Parus major* [8]) and for setting baselines and identifying potential cold adaptation mechanisms in dolphins (*Tursiops truncatus* [10]) and beluga whales (*Delphinapterus leucas* [11]).

Although RNA-sequencing techniques have become more feasible in non-model systems, collecting tissues that yield high-quality RNA remains a challenge in many wild populations. This is especially true for protected or long-lived species where non-lethal, minimally-invasive sampling is necessary. Characterizing transcriptomes from blood samples is appealing because blood circulates through the whole body and perfuses most organs and other tissues. Its utility as a liquid biopsy has been developed in human and wildlife medicine [12–14]. While blood does not capture the full array of physiological functions within an organism's tissues, blood transcriptomes have been shown to contain two thirds of orthologous genes present in liver samples (an organ with high functional gene expression diversity frequently used in transcriptomics studies) in six species of reptiles

[15], and contain 61% of protein coding genes in the genome of a species of bat [16]. Additionally, reptile blood samples include both nucleated red and white blood cells, so it is possible to obtain a sufficient amount of RNA from a small volume of blood [15, 17, 18], making blood transcriptomes a valuable tool to understand functional diversity in reptiles and potentially to develop biomarkers for physiological and health assessments.

Marine turtles are reptiles of conservation concern with a growing but limited body of genomic resources [19]. This taxon is globally distributed and has some of the longest known migrations on the planet, so a single individual may experience a wide range of environmental conditions and anthropogenic impacts, which have the potential to be cumulative, within its lifetime [20]. Six out of seven extant species are listed in an elevated threat category (vulnerable, endangered, or critically endangered) on the IUCN Red List and under the U.S. Endangered Species Act [21, 22]. Marine turtles face a myriad of threats, such as fisheries interactions, intentional harvest of eggs and meat for consumption, environmental contaminants, climate change, and disease [23–27]. While there are some characteristics shared by all or multiple species of marine turtle, each species, and sometimes populations within a species, have unique ecological adaptations and life history traits. For example, the trophic ecology varies widely between hawksbill (*Eretmochelys imbricata*; primarily spongivores), loggerhead (*Caretta caretta*; omnivores), green (*Chelonia mydas*; herbivores or omnivores depending on population or life stage), and leatherback (*Dermochelys coriacea*; gelatinivores) turtles [28]. Leatherback turtles also exhibit regional endothermy and other specialized physiological adaptations to inhabit cold water [29, 30]. The evolutionary divergence between Dermochelidae-Cheloniidae (the two extant marine turtle families containing the leatherback and hardshell marine turtle species, respectively) is estimated at 55–100 million years ago [31, 32], but turtles have slower rates of evolution compared to other vertebrates [33] and marine turtles can have high rates of sequence conservation between species [34]. Thus, these unique physiological

and ecological adaptations may be driven largely by key functional differences within a small proportion of their total genomes. Modulating gene expression can also be a mechanism of local adaptation and a source of evolutionary novelty between populations within a species [35, 36]. Gene expression profiles vary between geographically distinct populations and can also change based on environmental conditions such as water temperatures and life stage [9]. Thus, comparative transcriptomics approaches can identify potential drivers of the observed ecological diversity between and within marine turtle species, and offer key insight into how they modulate their physiology in response to natural and anthropogenically driven environmental conditions.

Here, we present the first multi-species comparison of marine turtle transcriptomes. In this study, we assembled *de novo* blood transcriptomes and examined gene expression across four species of marine turtles to characterize and compare the transcriptomic diversity within and across species. We also conducted functional annotation to explore the biological processes represented in genes expressed in blood. To further assess the utility of blood transcriptomes compared to other tissues commonly used for transcriptomic studies, we quantified the proportion of genes shared between blood, brain, lung, and ovary transcriptomes for leatherback turtles. Finally, we used differential gene expression and functional gene enrichment analyses to explore potential drivers of responses to varying environmental conditions within green turtle foraging aggregations. Green turtles have a global distribution comprised of eleven distinct population segments [37] that are genetically differentiated, have different life histories, and face varying levels of anthropogenic disturbance. Here, we include samples from three populations (East Pacific, Central North Pacific, and Central West Pacific), including individuals (East Pacific) that inhabit highly urbanized estuaries. Collectively, these analyses serve to demonstrate the potential of transcriptomics studies using minimally invasive blood sampling to advance our understanding of marine turtle evolutionary ecology and conservation biology.

Results

Transcriptome assessment & annotation

We conducted RNA-sequencing of blood samples from green, hawksbill, leatherback, and loggerhead turtles ($n = 43$), and used these data to assemble four species-specific blood transcriptomes. We also used public data in the NCBI Sequence Read Archive to assemble leatherback tissue-specific transcriptomes. Sequencing yielded 32.7 ± 5 million raw reads per sample (mean \pm standard deviation; Table S1), with an average of $5.5 \pm 2.3\%$ (mean \pm standard deviation) of reads mapping to

hemoglobin. Filtering to collapse transcripts with high sequence similarity and to remove redundant, low quality, or chimeric transcripts reduced the number of transcripts in assemblies by $27.9 \pm 7.6\%$ (mean \pm standard deviation) compared to raw assemblies. Transcriptomes had >75 and 71% mapping rates for conspecific and heterospecific samples, respectively (Table 1). All filtered assemblies had BUSCO completeness scores $>72\%$ (Table 2), and $N50 > 2000$. A total of 844 (0.8%) of all amino acid sequences in the green turtle filtered assembly matched to bacterial, archaeal, or viral sequences, indicating low levels of non-host contamination.

We functionally annotated the green turtle blood transcriptome using Blast2GO to investigate the functions of genes shared or differentially expressed between species or green turtle foraging aggregations [38]. Biological processes represented in the green turtle blood transcriptome are shown in Figure S1 and Table S2. Blast2GO retrieved BLAST hits for 44.4% of transcripts, gene ontology (GO) mappings for 33.9% of transcripts, and 24.7% of transcripts were ultimately annotated with GO terms. These annotated transcripts were associated with 19,583 GO terms across all three GO domains (cellular component, molecular function, and biological process). Of the annotated GO terms in the biological process category, the majority fell within biosynthetic processes ($\sim 15,000$), followed by cellular protein modification processes, signal transduction, cellular nitrogen compound metabolic processes, and stress response (Figure S1). Sequences in the green turtle blood transcriptome were involved with 140 KEGG (Kyoto Encyclopedia of Genes and Genomes) pathways [39]. The most complete KEGG pathways (highest number of pathway enzymes represented in transcriptome) included purine, amino sugar, glycine, glycerophospholipid, and pyrimidine metabolism. We also observed high numbers of sequences mapping to specific enzymes involved in numerous pathways. For example, 979 transcripts were annotated with enzyme code 3.1.3.16-phosphatase, which was involved in the T cell receptor signaling pathway, PD-L1 expression and PD-1 checkpoint pathway in cancer, and Th1 and Th2 cell differentiation (Table S3).

To examine the functions of genes shared between leatherback tissues and blood, we also functionally annotated a combined-tissue leatherback transcriptome. Annotation of the combined leatherback tissue transcriptome yielded BLAST hits for 63% of transcripts, GO mappings for 48.9% of transcripts, and 48.5% of transcripts were ultimately annotated with GO terms (Figure S2 and Table S4). However, we note that the higher annotation percentages here compared to the green turtle blood transcriptome were likely due to an additional filtering step applied in our computational streamlined methods using Transdecoder (i.e., smaller

Table 1 Quality assessment metrics of unfiltered and filtered transcriptome assemblies for multiple tissue types collected from four marine turtle species

	Loggerhead - blood		Hawksbill - blood		Green turtle - blood		Leatherback - blood		Leatherback - brain		Leatherback - lung		Leatherback - ovary	
	raw	filtered	raw	filtered	raw	filtered	raw	filtered	raw	filtered	raw	filtered	raw	filtered
Total trinity transcripts	132,146	77,392	280,711	220,458	489,355	376,736	347,717	276,709	216,942	140,332	243,118	165,611	163,840	119,574
Contig N50	3032	2552	3143	2276	3221	2303	2867	2187	3618	2788	3288	2526	3050	2373
Median contig length	675	707	574	529	606	575	597	553	666	629	632	601	673	593
Mean mapping rates														
<i>Conspecific samples</i>	91.50%	75.36%	95.53%	93.58%	94.88%	93.94%	95.49%	94.95%	92.98%	83.22%	92.52%	82.02%	94.96%	93.89%
<i>Heterospecific samples</i>	82.65%	69.54%	88.56%	85.44%	86.24%	85.99%	83.58%	83.14%	N/A	N/A	N/A	N/A	N/A	N/A
Transrate scores														
<i>Assembly score</i>	0.23	0.35	0.25	0.36	0.29	0.42	0.26	0.37	0.21	0.31	0.21	0.29	0.20	0.29
<i>Optimal assembly score</i>	0.35	0.36	0.36	0.37	0.42	0.43	0.36	0.38	0.33	0.32	0.30	0.30	0.30	0.30

input file containing only 77,387 transcripts identified as containing open reading frames). Annotated transcripts were associated with 23,859 unique GO terms across all three GO domains. Within the biological process category, the most abundant GO terms were related to signal transduction, biosynthetic process, cell differentiation, cellular protein modification, and response to stress. Annotated leatherback transcripts were involved in 149 KEGG pathways ([39], Table S3). The most complete KEGG pathways were also all related to amino acid metabolism (e.g. purine, glycine, pyrimidine, arginine), though these differed slightly in comparison to the green turtle annotation above. We also observed high numbers of sequences mapping to specific enzymes involved in numerous pathways. For example, 680 transcripts were annotated as part of the serine/threonine protein kinase enzyme, which is involved in thermogenesis, relaxin signaling, and numerous viral infection KEGG pathways.

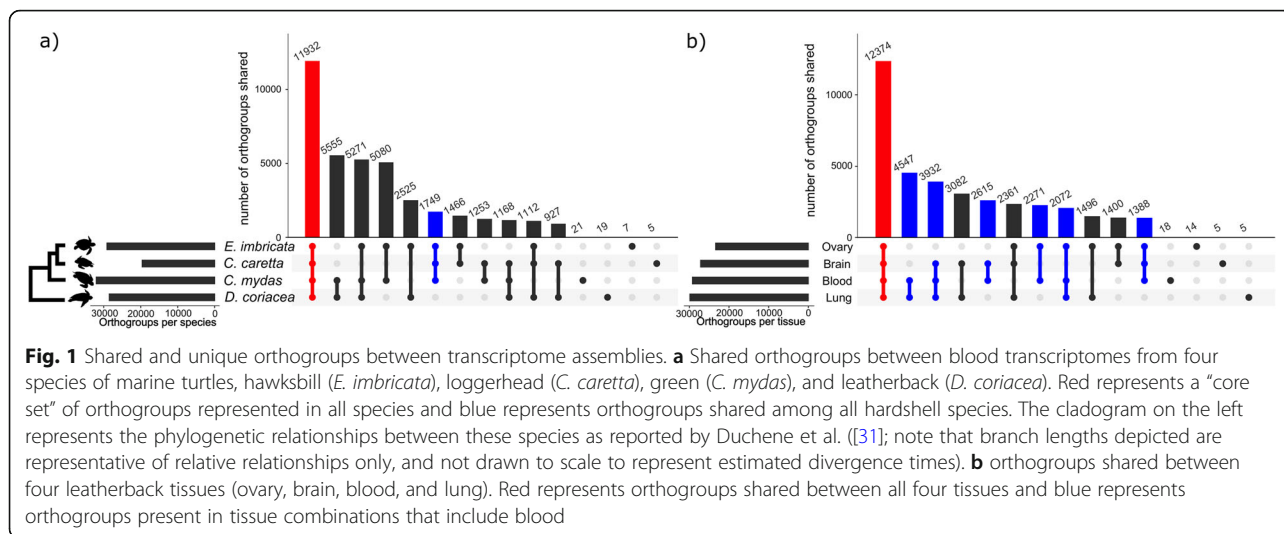
Shared orthology between species and tissues

There was a combined total of 267,039 transcripts in all four species-specific blood transcriptomes, and 64.3% of these transcripts were assigned to orthogroups (Fig. 1a; Table S5) via protein orthology analysis. A total of 11,

932 orthogroups were shared between all four species-specific blood transcriptomes (31.3% of all orthogroups identified). This was the largest shared set of orthogroups, and likely represents a core set of genes expressed in blood across marine turtles. The largest functional groups of genes in this core set based off the green turtle transcriptome annotation were biosynthetic processes ($n = 1447$ genes), cellular protein modification processes ($n = 1348$ genes), and signal transduction ($n = 1269$ genes; Fig. 2a, Table S2). Additionally, this 'marine turtle core gene set' contained 84.4% of the genes in the core set across reptilian blood transcriptomes previously identified by Waits et al. [15]. There were few species-specific orthogroups identified (≤ 60 , Fig. 1a), however, it is important to note that this is distinct from species-specific unique genes expressed because orthogroups are only assigned if more than one transcript (within or between species) is in the set [40]. The relative set size of shared orthogroups was not in complete concordance with phylogenetic distances between species. Specifically, although leatherback turtles have the greatest divergence from the other species ([31], Fig. 1a), the number of orthogroups shared among the three hardshell species was lower than the numbers of orthogroups shared among several other groups containing hardshell species

Table 2 BUSCO completeness percentage scores based on the vertebrata database for unfiltered and filtered transcriptome assemblies for multiple tissue types collected from four marine turtle species

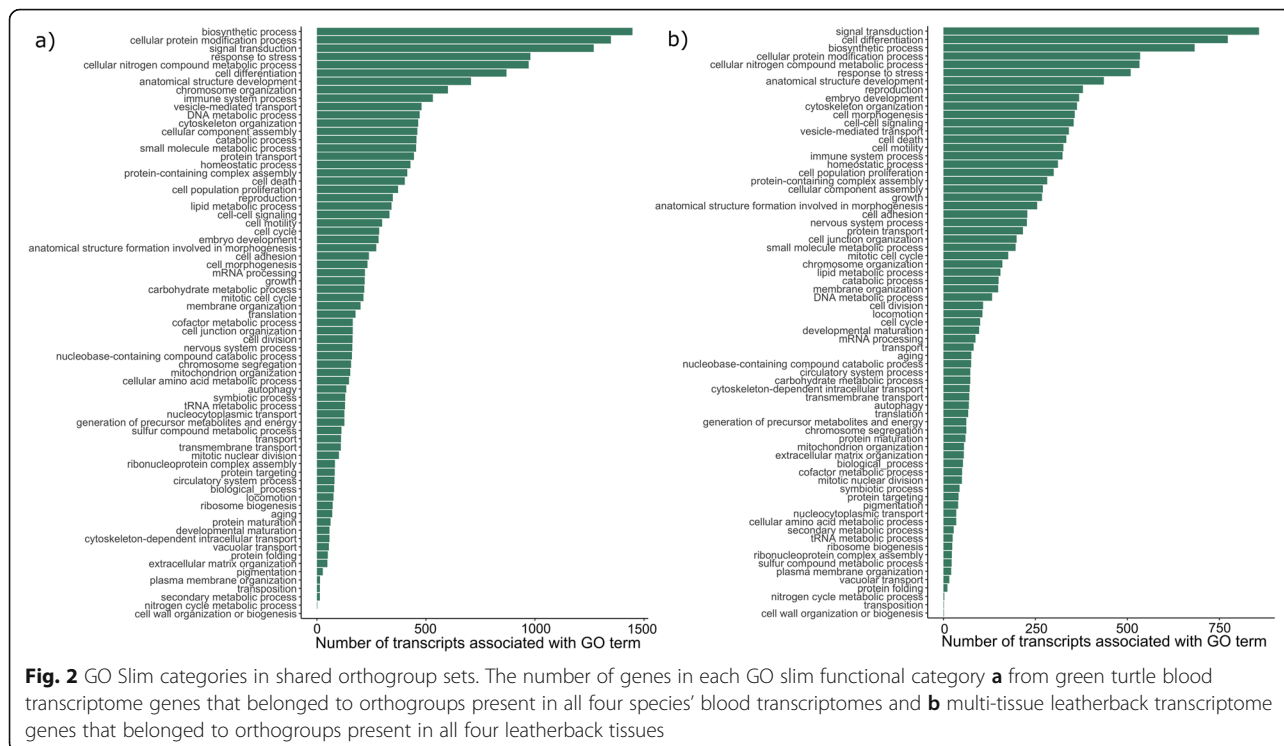
	Loggerhead - blood		Hawksbill - blood		Green turtle - blood		Leatherback turtle - blood		Leatherback - brain		Leatherback - lung		Leatherback - ovary	
	raw	filtered	raw	filtered	raw	filtered	raw	filtered	raw	filtered	raw	filtered	raw	filtered
Total Complete BUSCOs	76.7	72.8	81.1	80.7	83.7	83.7	84.9	85	90.6	86.3	89.5	86.4	88.9	89
Single-copy complete BUSCOs	37.3	50.9	33.9	46.6	31.2	43.4	32.8	45.4	40.9	57.2	39.7	55.5	37.2	57.5
Duplicated Complete BUSCOs	39.4	21.9	47.2	34.1	52.5	40.3	52.1	39.6	49.7	29.1	49.8	30.9	51.7	31.5
Fragmented BUSCOs	6.3	7.1	5.5	5.6	5.4	5.5	4.5	4.2	3.1	4.1	4.1	5	3.9	3.7
Missing BUSCOs	17	20.1	13.4	13.7	10.9	10.8	10.6	10.8	6.3	9.6	6.4	8.6	7.2	7.3



and the leatherback turtle. However, all of the groups in the latter category were missing the loggerhead, for which only a single sample was available.

In a comparison of the leatherback blood transcriptome to those of more traditionally sampled organs, 69.5% of 228,977 total transcripts were assigned to an orthogroup by protein orthology analysis (Fig. 1b and Table S6). This comparison revealed that a large proportion of identified orthogroups were expressed in all four tissues (12,374 orthogroups, 32.9% of total orthogroups identified; Fig. 1b and Table S6). The largest functional

groups of genes in this core set based off the multi-tissue leatherback transcriptome annotation were signal transduction ($n = 858$ genes), biosynthetic processes ($n = 683$ genes), and cell differentiation ($n = 773$ genes; Fig. 2b, Table S4). Secondly, 44.8% of orthogroups were expressed in other combinations of tissues that included blood. Similar to blood transcriptome comparisons across species, there were few tissue-specific orthogroups (42 orthogroups, 0.11% of total orthogroups), which contained 137 transcripts (0.06% of all transcripts present in the four assemblies).



Transcriptional signatures across species

Multi-dimensional scaling (MDS) revealed distinct clustering by species (Fig. 3a), indicating that transcriptional signatures of shared genes vary among species. Exploratory differential expression analysis including only orthogroups shared between the three species with more than one sample available (green turtles, hawksbills, and leatherbacks) further identified that 47.4–57.4% of shared orthogroups were significantly different among the species (Table S7).

Differential gene expression among green turtle foraging aggregations

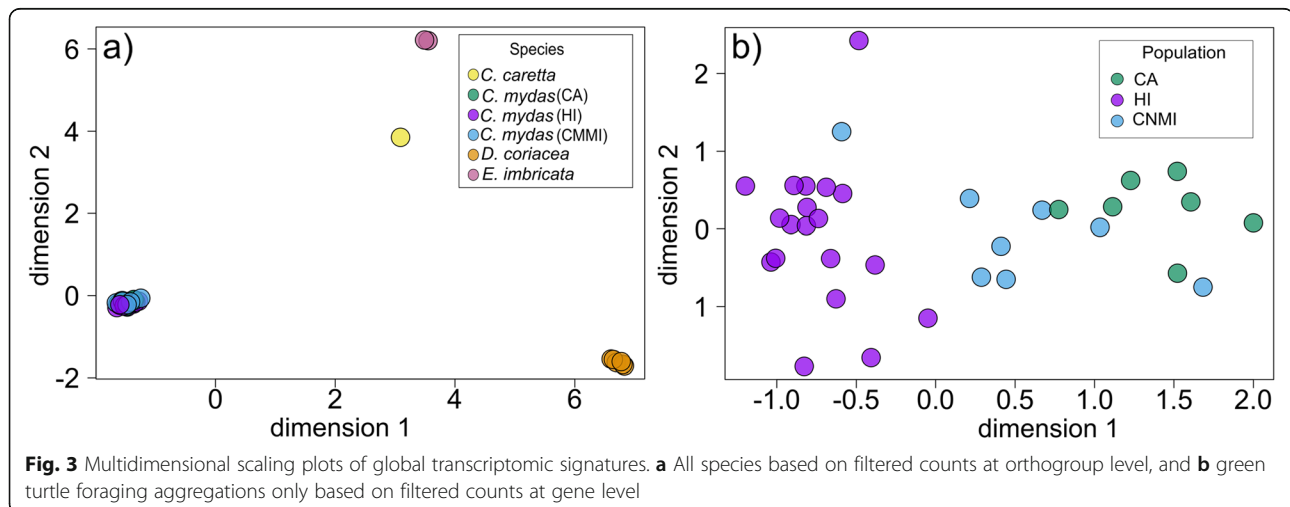
Green turtle gene expression signatures in our MDS analysis clustered by foraging aggregation, but to a lesser degree than among species (Fig. 3b). We found significant differential gene expression between all three pairwise comparisons of green turtle foraging aggregations, with the most differentially expressed genes between Hawai'i and California green turtles (6649 genes, FDR < 0.05), and the least between Hawai'i and Commonwealth of the Northern Mariana Islands (CNMI) green turtles (600 genes, FDR < 0.05) (Fig. 4 and Table S8). Thirty genes were differentially expressed in all three pairwise foraging aggregation comparisons (Table S8). Biological functions of these genes included response to oxidative stress, immune response, DNA repair, and others (see annotations in Table S2). Functional enrichment analyses for each pairwise comparison revealed a total of 16 enriched GO terms at $P < 0.01$ and 78 enriched GO terms at $0.001 < P < 0.05$ (Fig. 5, Table S9). The top three most significantly enriched GO terms represented stem cell population maintenance, organelle organization, and processes using autophagic mechanisms, all in the California and Hawai'i pairwise comparison. The top two enriched GO terms were found in all three pairwise comparisons ($P < 0.05$). Some other enriched ($0.001 <$

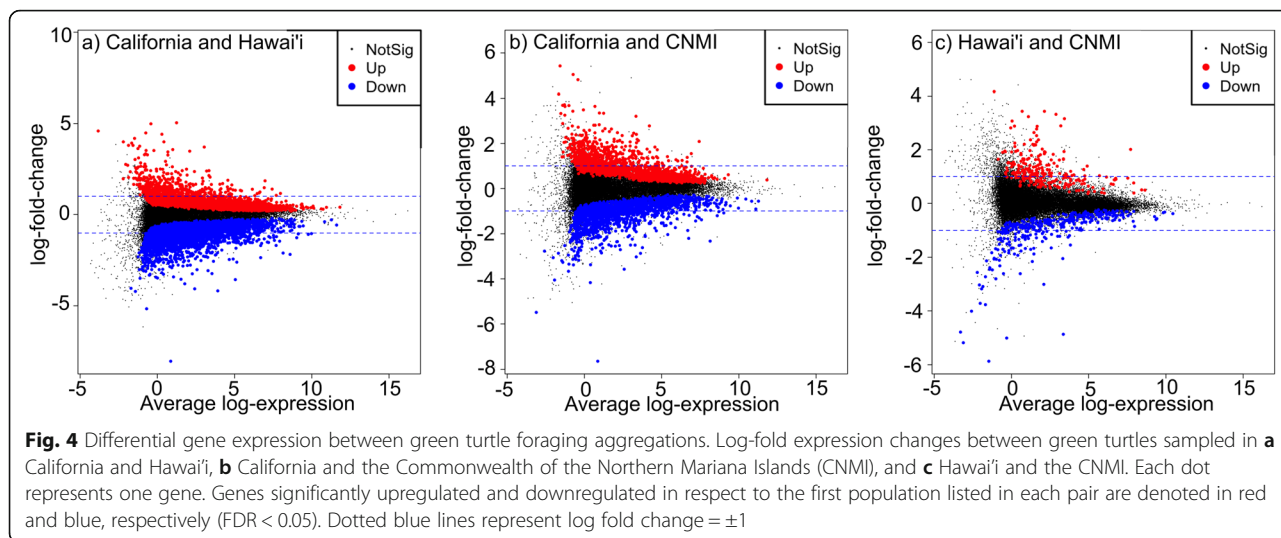
$P < 0.05$) GO terms of potential interest for future biomarker development included cellular response to stress, cell activation involved in immune response, and leukocyte mediated immunity.

Discussion

Global transcriptomics has emerged as a robust approach to understand the mechanistic underpinnings of biodiversity and organisms' responses to environmental stressors [1, 2, 7, 8]. It is also well-suited to complement traditional physiological datasets, such as clinical blood panels and hormone assays. However, until genomic resources and techniques for high quality sample collection are available, its practical utility for isolated and endangered populations will remain limited. Here, we generated high quality de novo transcriptome assemblies for four species of marine turtles and demonstrate that blood is a promising tissue that can be collected using non-lethal and minimally invasive sampling methods for transcriptomic studies. We reported sample collection and sequencing preparation techniques that yield high quality data from marine turtle blood and provide transcriptomes which can be used by other researchers. We characterized gene expression differences at both the species and population levels, which, in future studies, can be paired with complementary data sets to investigate linkages with environmental conditions. We also identified core sets of shared and unique genes among species that may have applications in studies of marine turtle ecological and physiological diversity, as well as the development of potential biomarkers for environmental stress responses, as has been done in other wild species [41–44].

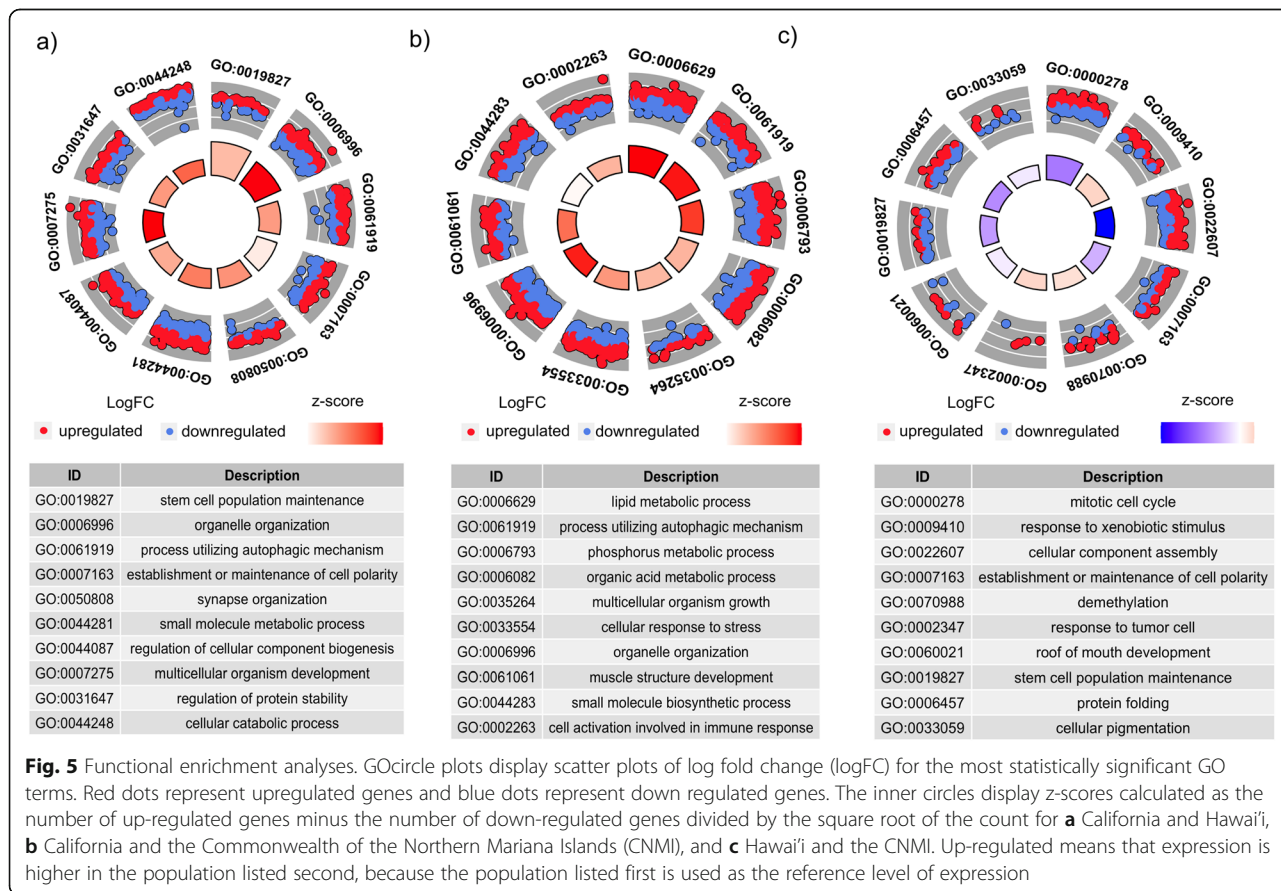
Turtle blood transcriptome assemblies from this study generally had high species-specific mapping rates, BUSCO completeness scores, and transcript diversity. Although at our depth of sequencing, some genes that





were lowly expressed in blood may be omitted, overall, these metrics indicated that our blood transcriptome assemblies were robust and high quality [3, 5, 11, 45–47]. The lower mapping rate and BUSCO completeness score of the loggerhead relative to other species is likely a result of this assembly being constructed from only one individual. Notably, it also was the species missing from

sets with numbers of shared orthogroups that did not align with phylogenetic distance (Fig. 2a), suggesting lower transcript diversity was likely due to shallower sequencing. Although the individual we sequenced had reasonable depth (~28 M reads), these results are in concordance with prior studies' recommendations that using multiple individuals results in more complete de



novo transcriptome assemblies [4] and that at least 50 M input reads are ideal for robust assemblies [5, 45]. These high-quality de novo assemblies add to the growing number of genomic resources for marine turtles and can serve as references for future gene expression or functional gene sequence analyses studies.

Using blood for transcriptomics analyses from minimally invasive non-lethal sampling can substantially expand the species and life stages from which transcriptomic data can be gathered, which is particularly important for protected species like marine turtles. Additionally, because ample high-quality RNA can be extracted from a very small volume of blood samples in reptiles [14], transcriptomic data from blood can be gathered from individuals repeatedly as part of long-term monitoring of their health and used to answer a host of ecological and evolutionary questions. However, we recognize there are limitations in using blood for transcriptomic studies, as tissue-specific expression is common [48, 49]. Nonetheless, our findings that the functional diversity in global gene expression and BUSCO completeness scores in leatherback blood was similar to three organs traditionally used in transcriptomic analyses confirm that blood has high potential in yielding informative gene expression data in marine turtles. Our orthologous protein analysis also demonstrated that the loss of represented genes is relatively modest using blood samples instead of traditionally used tissues, and is similar to what has been demonstrated in other reptile and mammal species [15, 16, 50]. Another possible concern when using blood for RNA-sequencing studies is the high hemoglobin content consuming a large proportion of sequencing effort, making it cost-inefficient without hemoglobin depletion methods [51] that are challenging in non-model species. However, we found low alignment rates to publicly available green turtle hemoglobin sequences in our samples. This was consistent across samples, distinct population segments, and species, and considerably lower than has been previously reported in other reptiles [15]. Combined with the high diversity of functional genes in blood transcriptomes, the rapidly declining costs of high-throughput sequencing, and the risk of introducing bias with more sample processing, our results suggest the most effective approach in marine turtles may be to ‘sequence through’ over-representation of hemoglobin instead of investing in taxon-specific hemoglobin depletion methods. Lastly, contamination of transcriptomes can be a concern when employing de novo assembly methods, particularly when using tissues that can harbor pathogens and parasites such as blood. However, we found very low levels of microbial contamination (0.8%) in our assemblies suggesting that microbial contamination may not be a large issue for marine turtles and can be mitigated by

including a filtering step during bioinformatics analyses if needed [15]. Further, for studies that wish to characterize both host and pathogen gene expression (i.e., dual RNA-Seq [52, 53]), blood may actually offer additional advantages as a tissue of choice in this regard. Interestingly, some of the top BLAST hits for species within bacteria, archaea, and viruses from the green turtle assembly indicated the presence of pathogenic microbial species (e.g. *Acinetobacter baumannii*). Similar analyses of transcriptomic data have identified parasites in lemurs (*Indri indri* and *Propithecus diadema*) and poison frogs (Dendrobatidae) [54, 55]. By detecting pathogen RNA in blood samples, one would be able to confirm that the pathogen was alive at the time of sampling [53]. Thus, in addition to using transcriptomic data to study immune genes and identify signatures of adaptive evolution in host species alone [56], these data can be paired with pathogen screening or cultures of host blood to evaluate bacteremia or septicemia to address a diversity of complex disease ecology and co-evolutionary research questions. Overall, our findings support that blood is an excellent tissue for a minimally invasive and non-lethal liquid biopsy for marine turtle species.

The orthogroups shared between hawksbill, loggerhead, green, and leatherback blood assemblies likely represent a core set of functional genes expressed in the blood of marine turtles, though future studies including the other three extant species of marine turtles will need to confirm this finding. Moreover, the large percentage of genes shared between this core set of marine turtle blood genes and the core set of reptilian blood genes identified in Waits et al. [15] reveals that many physiological pathways are likely conserved at broader taxonomic scales, and may be useful targets for studies developing biomarkers or investigating functional diversity across Reptilia. In particular, the marine turtle core blood gene set included 138 genes from all a priori candidate groups defined by Waits et al. ([15]; following McGaugh et al. [57]) that are of high interest for molecular evolution and functional ecology studies and biomarker development, including the mitochondrial electron transport chain, stress response, oxidative stress, and insulin-signaling pathway genes. Conversely, although we found relatively few species-specific orthogroups, this is likely not an accurate depiction of species-specific expression because Orthofinder only assigns transcripts with at least two orthologous transcripts, within or among species, to orthogroups [40], and our assemblies were constructed with variable sequencing depths among species. It is therefore likely that single copy species-specific expressed genes are underestimated in our analyses. Although it was outside the scope of aims for this study, future analyses can examine the transcripts not assigned to orthogroups to explore

genes that may only be expressed in one species. Finally, within the shared orthologous genes, we observed strong differences in expression levels between species, highlighting the potential role of gene expression regulation in underlying physiological and ecological differences among species. Further studies with larger sample sizes that assess both expression regulation and sequence divergence are needed to confirm these expression differences and understand these mechanisms. While our sample sizes for inter-specific gene expression analyses were small, our data offer exciting preliminary findings that can inform future work.

Within green turtles, we found population-specific clustering and differential expression of many genes among all three foraging locations, California, Hawai'i, and the CNMI. These groups are largely demographically isolated [37], and also inhabit areas that strongly differ in habitat and anthropogenic impacts. As such, the observed location-based differences could be caused by genetic divergence related to neutral evolutionary processes (e.g., drift following reproductive isolation), different physiological responses based on environment, or most likely, a combination of both genetic and environmental influences. Population-specific gene expression has been documented in a diversity of other marine taxa, such as stony corals, teleost fishes, and intertidal copepods [58–60]. While experimental work using traditional approaches such as common garden experiments is challenging in protected species, our data can inform hypotheses about what is driving these differences [1, 61] that can be further assessed with candidate gene profiling linked to complementary datasets. To understand environmental influences, transcriptomic data can also be paired with contaminant analysis to identify correlations between gene expression and environmental pollution [62–64]. Additionally, environmental degradation has been associated with fibropapillomatosis, a tumor-forming disease of marine turtles, within the Hawai'ian Islands and globally [65–68]. Thus, identifying gene expression profiles associated with pollution could provide insight into disease emergence and aid in developing biomarkers of disease. The stronger differential expression between both insular foraging aggregations (i.e., Hawai'i and CNMI) and the California foraging aggregation is suggestive of environmental drivers because although California and Hawai'i populations originate from the same evolutionary clade that is distinct from the CNMI population [69], the California aggregation forages in a much more highly urbanized temperate environment [70] compared to the tropical, less impacted foraging grounds of both island aggregations. Further, the Hawai'i aggregation largely forages

and nests within the greater Hawai'ian islands [71, 72], so there is likely limited contemporary gene flow among the Hawai'ian and CNMI populations. Finally, GO terms representing cellular response to stress and cell activation involved in immune response were significantly enriched in comparisons between California and both island foraging aggregations, and leukocyte-mediated immunity was also enriched between the California and Hawai'i foraging aggregations. Thus, although we cannot draw causative conclusions from our current dataset, it is a plausible hypothesis that differential expression of genes in these functional groups may be in response to differences in exposure to pollution and other stressors in urbanized versus insular locations. Relationships between environmental stressors such as pollutant and pathogen exposure and transcriptomic responses have been documented in a wide variety of taxa, including great tits [8], killifish (*Fundulus heteroclitus* [73]), and wild salmon smolts (*Oncorhynchus nerka* [74]). Additionally, correlations between stress hormones and gene expression have been documented in elephant seals (*Mirounga angustirostris* [75]). Transcriptomic data can also be paired with sex or life stage information to explore the potential for developing gene expression-based biomarkers to determine important demographic information such as sex, which would be a useful addition to the current methods available for sexing immature marine turtles [76, 77]. Thus, future studies pairing gene expression data with contaminant profiles, disease status, environmental data, and health assessment biomarkers, while factoring baseline expression differences between groups demonstrated in this study, can strengthen our understanding of potential relationships between differential gene expression and environmental stress in wild populations.

Conclusions

Minimally invasive blood sampling combined with de novo transcriptomic approaches has strong potential to alleviate key barriers of applying transcriptomic tools in wild, protected populations. Our study provides genomic resources for non-model species of high interest for conservation and demonstrates how global gene expression data from blood can be used to explore evolutionary ecology and anthropogenic impacts on marine turtles and other species where traditional lethal sampling is unwarranted. The distinct differences in gene expression signatures between species and populations yield insight into the functional genomics underlying the diversity in this ancient vertebrate lineage, and the high-quality transcriptomes and expression analyses provide key baseline information to inform a variety of future transcriptomic applications in marine turtles.

Methods

Sample collection and RNA extraction

Blood samples were collected between 2012 and 2016 from hawksbill turtles from off Palmyra Atoll (U.S. Minor Outlying Islands) and the CMNI ($n = 2$), a loggerhead off the southern California coast ($n = 1$), leatherback turtles off the central California coast (USA, $n = 6$), and from three foraging aggregations of green turtles in Southern California (USA, $n = 7$, East Pacific population), Hawai'ian Islands (USA, $n = 19$, Central North Pacific population), and the CNMI ($n = 8$, Central West Pacific population). See Table S1 for specific sampling locations and additional details. Blood was collected from the dorsal cervical sinus using 21-gauge 3.8-cm needles for hardshell turtles [78] and 18-gauge 8.75 cm spinal needles flushed with sodium heparin for leatherback turtles, connected to a vacutainer adapter to directly fill PAXgene™ Blood tubes (Qiagen, Valencia, CA, USA) to stabilize RNA. Blood-filled PAXgene tubes were initially stored at -20°C followed by -80°C within 48 h until analysis according to manufacturer's instructions. Samples ($n = 29$) obtained from the National Institute of Standards and Technology (NIST) Biological and Environmental Monitoring and Archival of Sea Turtle Tissue (BEMAST) cryogenics biorepository project were stored at liquid nitrogen vapor temperature.

Total RNA isolation using Qiagen's PAXgene kits developed for mammalian samples was previously determined to be unable to yield high-quality RNA from samples with nucleated red blood cells, likely due to higher nuclease enzymes and protein content (L. Komoroske, unpublished data). Therefore, we developed a method optimized specifically for total RNA extraction from marine turtle whole blood [18], modified from Chiari and Galtier [17]. RNA quality (RNA integrity number (RIN)) and quantity were determined on a Fragment Analyzer (model number: 5200, Agilent Technologies, Santa Clara, CA) using a standard sensitivity RNA kit (Agilent Technologies, Santa Clara, CA, DNF-471). Total RNA was isolated from blood samples within 1–4 years after collection. Samples with $\text{RIN} > 7.5$ were used for further analysis.

Library preparation and sequencing

We isolated mRNA from Total RNA using the NEBNext Poly(A) mRNA Magnetic Isolation Module (New England Biolabs, Ipswich, MA) followed by library preparation using the NEBNext Ultra Directional RNA Library Prep Kit for Illumina and the NEBNext Multiplex Oligos for Illumina (New England Biolabs, Ipswich, MA) for dual indexing with modifications for half reactions [79]. Individual sample libraries were pooled in equimolar quantities, and the pooled library was sequenced in 150 bp paired-end reads across three lanes of an

Illumina HiSeq 4000 (Illumina, San Diego, CA) by Novogene Corporation (Sacramento, CA). To compare blood to tissues more traditionally used for transcriptomic analyses, we used RNA-Seq data for leatherback ovary, lung, and brain tissue available in the NCBI Sequence Read Archive (SRA accession numbers: SRX8787566, SRX8787565, SRX8787564). See Availability of Data and Materials section for further information and the location of all scripts used for the analyses described below.

Transcriptome assembly, filtering and mapping

Sequences were demultiplexed and concatenated across lanes by sample, followed by trimming for adaptor content with scythe [80] and quality with sickle (minimum Phred quality score of 20 [81]). We assembled de novo transcriptomes for each of the four species to capture species-specific transcripts and avoid mapping biases towards the two species (i.e. green and leatherback) with draft reference genomes available. We tested multiple numbers of samples to use as input for our de novo transcriptomes ($n = 34$, $n = 19$, and $n = 4$) to determine the optimal threshold of individuals for maximizing transcriptome completeness while minimizing computational demands, chimeric sequences, and false-splitting due to sequence divergence between populations [82], and concatenated reads from four Hawai'ian green turtle individuals to generate the green turtle transcriptome. Exploratory mapping of green turtle sequences to the green turtle reference genome [83] also determined that Hawai'ian green turtles expressed the highest sequence diversity, further supporting this sample selection for the de novo transcriptome assembly. We then similarly assembled species-specific blood transcriptomes for leatherbacks ($n = 3$), hawksbills ($n = 2$), and a loggerhead ($n = 1$), as well as tissue specific assemblies for leatherback ovary, brain, and lung ($n = 1$ per tissue). We also included leatherback brain, ovary, and lung in a single assembly in order to annotate the maximum number of functional genes. We used Trinity to assemble each transcriptome with in silico read normalization included to increase computational speed, and we set minimum contig length = 300 bp to minimize fragmented transcripts (v.2.85) [84]. We then filtered the assemblies to remove redundant and low quality or chimeric transcripts using TransRate (v.1.0.3 [85]); to retain only contigs that were most likely to be structurally complete and correct, and then CD-HIT-EST (v.4.8.1) [86] to collapse transcripts with greater than 95% similarity. We then used Salmon (v.1.1.0) [87] to quasi-map reads for each individual to their species-specific assembly and quantify transcript expression, followed by a modification of the gathercounts.py script [88] to convert raw counts from the

salmon output into a count format compatible with edgeR [89].

Transcriptome evaluation & sequencing efficiency

We evaluated assemblies before and after filtering using a combined suite of metrics including the N50 values and median contig lengths reported by the TrinityStats.pl script from the Trinity assembler [84], BUSCO completeness scores [90], TransRate scores [85], and mapping rates of conspecific and heterospecific reads to each assembly using Salmon [87] as described above. We also estimated the percentage of potential contaminant proteins in the green turtle assembly using the Diamond protein sequence aligner [91] against the NCBI nr database, filtered to only include top hits to species within bacteria archaea, and viruses (NCBI taxon ids: 2, 2157, 10,239). We considered sequences to be of non-host origin if blast sequence similarity was greater than 90%. Finally, high hemoglobin expression in blood has previously been shown to hinder effective RNA-sequencing in blood of some species but not others [92, 93]. However, alternative approaches to ‘sequencing-through’ this problem such as hemoglobin depletion requires custom bait design for non-model species, which incurs additional costs and adds another step of sample manipulation that may interject bias into expression profiles [51]. Thus, to assess this issue for marine turtles, we calculated the percentage of reads that aligned to five green turtle (*Chelonia mydas*) hemoglobin sequences available in the NCBI gene database (LOC102939173, LOC102945818, LOC102938944, LOC102946728, LOC102945589, NCBI, Accessed 28 April 2020) using bowtie2 (v. 2.3.4.3 [94]).

Protein orthology between transcriptomes

To enable estimation of the proportion of shared versus unique genes between species, we first determined orthologous transcripts across our species-specific transcriptomes. This approach avoids biases that can arise when mapping multiple species to one reference transcriptome due to sequence divergence between species [4]. We translated and predicted coding regions for each of our assemblies using Transdecoder (v.5.5.0) [95] and then employed Orthofinder with default parameters to group transcripts from our species-specific assemblies into orthogroups, which are defined as sets of genes descended from a single gene in the most recent common ancestor within species groups (v.2.3.3) [40]. Using this method, only transcripts that have at least one orthologous transcript in any transcriptome are assigned to an orthogroup, so transcripts without any orthologs (within or across transcriptomes) are not retained [55]. We quantified the proportion of shared orthogroups between species in R (v3.6.3) [96] and visualized the results

with the package ‘UpSetR’ [97]. After identifying a core set of orthogroups shared across all species in our study, we also compared to those in the core set of reptilian genes expressed in red blood cells previously identified by Waits et al. [15] using the Diamond protein sequence aligner [91] to identify Uniprot gene names matching those in our core set of orthogroups based on our green turtle annotation (see below). Finally, to evaluate the proportion of the exome present in blood transcriptomes, we followed the same procedure to identify and estimate shared and unique orthologs in leatherback turtles between blood and tissues with known high gene expression diversity traditionally employed in transcriptomic studies (brain, lung, and ovary).

Functional annotation

We used Blast2GO (v.5.2.5) [38, 98] to functionally annotate the de novo green turtle transcriptome, and linked annotations to the other species-specific transcriptomes via orthogroups identified as described above. We chose to use this approach instead of annotating each transcriptome separately in order to relate the functional processes of orthogroups across species, and because we were largely focusing on differential expression and functional enrichment analyses between foraging aggregations of green turtles. Additionally, we conducted pilot analyses to confirm our expectation that species-specific transcriptome annotations would have a high concordance of gene identities within each orthogroup because each annotation is based on homologous genes in other vertebrate taxa using the same databases (all pairwise comparisons showed $\geq 95\%$ concordance). In brief, sequences in the final filtered green turtle assembly were compared to protein sequences in the NCBI non-redundant protein database (version 5) filtered to include only hits within vertebrata (NCBI taxonomy ID 7742) using the BLASTX-fast algorithm (e-value = $1.0e^{-3}$, word size = 6, and otherwise default parameter selections [38]). BLAST hits were then mapped to GO terms (GO database accessed 11/2019), followed by annotation of GO terms to sequences (annotation cut-off threshold = 60, E-value = $1e^{-6}$, otherwise default). GO annotations were confirmed and augmented using EggNOG mapping (version 2) to clusters of orthologous groups (version 5.0) [99]. Transcripts were also mapped to enzyme codes using the KEGG pathway analysis module, from which we generated KEGG pathway maps and statistics. Finally, we employed GO-Slim to reduce the specificity of GO terms assigned to sequences to yield a final set of broader functional summary statistics. To explore the identity and biological functions of shared and unique genes between blood and other tissues, we also annotated a combined assembly of all three leatherback tissues (brain, lung, and ovary). This followed the same

procedure with the exception of two adjustments for computational streamlining informed by the results of the green turtle annotation: filtering with Transdecoder (5.5.0) [95] to retain only predicted coding regions prior to annotation, and blasting sequences against the NCBI non-redundant protein database (version 5) filtered to include only hits within tetrapoda (NCBI taxonomy ID 32523).

Gene expression analyses between species

To compare gene expression between species, we condensed transcript counts to orthogroup level counts. Transcripts not assigned to an orthogroup were also excluded. We then filtered orthogroup level counts so that only orthogroups with at least one count per million (cpm) in at least two individuals (out of all individuals included) were retained and then normalized raw orthogroup level counts by library size using edgeR's 'TMM' method [89]. We included all remaining orthogroups in a multi-dimensional scaling plot to visualize how orthogroup expression signatures differed between species. We conducted differential expression analysis to identify genes that may be candidates for driving species-specific signatures, although we recognize that this analysis is exploratory given our limited sample sizes and other challenges of comparing expression between divergent wild non-model species (e.g., establishing control groups is not possible and individuals may be captured in varied circumstances [4]). We excluded loggerheads from differential expression analyses because we only had data from one individual. To reduce issues of unequal sample sizes between groups, we randomly selected three green (one from each population) and three leatherback individuals to include with the two hawksbills. We included only orthogroups present in all three species to reduce biases driven by strong signals from species-specific orthogroups. We then filtered orthogroup level counts so that only orthogroups with at least one count per million (cpm) in at least two individuals (out of all individuals included) were retained, normalized raw counts by library size using edgeR's 'TMM' method, and conducted differential expression analyses using the R packages edgeR and limma [89, 100].

Differential expression & functional enrichment between green turtle foraging aggregations

To compare gene expression between green turtle foraging aggregations we condensed transcript counts to gene level counts, filtered counts to retain only genes with at least one cpm in at least seven individuals (the smallest group size), and normalized raw counts by library size using edgeR's 'TMM' method [89]. We conducted multi-dimensional scaling visualization and

differential expression analyses between the three green turtle foraging aggregations using the R packages edgeR and limma [89, 100]. We then performed functional enrichment analyses of pairwise comparisons between green turtle foraging aggregations with a Kolmogorov-Smirnov test (weight01 algorithm) implemented in the R package TopGO [101]. We report raw p -values rather than reporting p -values adjusted for multiple testing correction, and do not attribute statistical significance to an $\alpha = 0.05$ threshold, because authors of TopGO caution that adjusted p -values may be misleading because p -values for each GO term are not calculated independently of other GO terms [102]. Input GO terms derived from the green turtle transcriptome annotation were filtered to biological process GO terms \leq level 5. The median number of transcripts per GO term was 17, so we included up to 50 transcripts per GO term. Transcript level GO annotations were combined to gene level and redundant terms were removed. The top ten most enriched GO terms for each pairwise analysis were visualized with GOCircle plots using the R package GOploit [103]. GOCircle plots show scatterplots of log fold change values and z -scores (calculated as the number of up-regulated genes minus the number of down-regulated genes divided by the square root of the count) for genes that belong to the top ten most significantly enriched GO terms [103]. It is important to note that the z -score does not give any information about how significant GO terms are as highly significant GO terms can have z -scores close to zero [103].

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12864-021-07656-5>.

Additional file 1: Table S1. Sample metadata. Detailed metadata, raw read count, and the percent of reads that map to green (*Chelonia mydas*) turtle hemoglobin genes for each individual ($n = 43$) included in this study. Individuals without straight carapace length (SCL) denoted were not measured for these parameters.

Additional file 2: Table S2. Annotation and orthogroup information for green turtle blood transcriptome assembly transcripts. Transcripts that did not match to a Gene Ontology (GO) ID are not included.

Additional file 3: Table S3. KEGG pathway analyses results. KEGG pathways for the green turtle transcriptome and the leatherback multi-tissue assembly.

Additional file 4: Table S4. Annotation and orthogroup information for leatherback multi-tissue transcriptome assembly transcripts. Transcripts that did not match to a Gene Ontology (GO) ID are not included.

Additional file 5: Table S5. Species-specific orthogroups. Orthogroups and the transcripts that belong to each orthogroup from each species-specific blood transcriptome.

Additional file 6: Table S6. Tissue-specific orthogroups. Orthogroups and the transcripts that belong to each orthogroup from each individual tissue-type leatherback transcriptome.

Additional file 7: Table S7. Differential orthogroup expression analyses results for comparisons between marine turtle species. Log fold change

and adjusted p -values are listed for the green v. leatherback, green v. hawksbill, and leatherback v. hawksbill turtle comparisons.

Additional file 8: Table S8. Differential gene expression analyses results for comparisons between green turtle foraging aggregations. Log fold change and adjusted p -values are listed for the California and Hawai'i, California and the Commonwealth of the Northern Mariana Islands (CNMI), and Hawai'i and CNMI Islands comparisons.

Additional file 9: Table S9. Functional enrichment analysis results. Functional enrichment analysis results for contrasts between green turtle foraging aggregations.

Additional file 10: Figure S1. Green turtle GO slim plots. Bar plots representing the number of genes in each Gene Ontology (GO) slim functional category from the green turtle blood transcriptome.

Additional file 11: Figure S2. Leatherback GO slim plots. Bar plots representing the number of genes in each Gene Ontology (GO) slim functional category from the multi-tissue (brain, lung, and ovary) leatherback turtle transcriptome.

Acknowledgements

We mourn the loss of our colleague, Kate McFadden, who played an integral role in this work, and whose presence and intellect we sorely miss. We thank Frannie Nilsen, Angela Hansen, and Jessica Jacob for sample collection, Paul Becker, Rebecca Pugh, and Brenda Jensen for sample archival, Rina McPherson, Nann Fangue and Victoria Pease for assistance with laboratory work and logistics, and Blair Bentley for assistance with bioinformatics. We thank the CNMI Department of Lands and Natural Resources Sea Turtle Program for facilitating sample collection. We are also grateful to Moss Landing Marine Laboratories small boat operations and the St. Croix Leatherback Project (Kelly Stewart) at Sandy Point National Wildlife Refuge (Claudia Lombard) for leatherback nesting beach field work assistance and sample collection, as well as John Douglas for leatherback in-water field work assistance and facilitation of sample collection. Many thanks to Erin LaCasella and Gabriela Serra-Valente for their assistance with curation and processing of samples accessioned into the Marine Mammal and Sea Turtle Research (MMSTR) Collection at the Southwest Fisheries Science Center's La Jolla Laboratory in California, USA. Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

Authors' contributions

SMB collected samples, performed laboratory work, conducted data analysis, and wrote the manuscript. Jamie Adkins Stoll conducted data analysis and wrote the manuscript. CDA, JML, EJS, EN, HSH, KM, RLL, and MML contributed to study design, collected samples, and provided editorial feedback on the manuscript. REC provided initial training and support with the optimization of sample processing techniques, and editorial feedback on the manuscript. LK and NBF performed data analyses and JB performed laboratory work. JAS, PHD, SRB, SM, TMS, JRH, MRR, TE, TTJ, and GHB supported fieldwork for sample collection and provided editorial feedback on the manuscript. LMK created the study design, collected samples, performed laboratory work, conducted data analyses and wrote the manuscript. The author(s) read and approved the final manuscript.

Funding

The sample collection was supported by Southwest Fisheries Science Center, Pacific Islands Fisheries Science Center, The Commonwealth of the Northern Mariana Islands Department of Fish and Wildlife, National Institute of Standards and Technology, and US Fish and Wildlife Service. Funding for study design, data analysis, interpretation, and manuscript writing was provided to SMB, JAS, and LMK by the University of Massachusetts Amherst and NSF-IOS award #1904439. CDA was funded by the National Research Council and Ocean Associates, Inc.

Availability of data and materials

All raw RNA-seq reads are archived under NCBI BioProject # PRJNA660024 (<https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA660024>) and SRA accession numbers: SRX8787564 (<https://www.ncbi.nlm.nih.gov/sra/SRX8787564> [accn]), SRX8787565 (<https://www.ncbi.nlm.nih.gov/sra/SRX8787565> [accn]), and SRX8787566 (<https://www.ncbi.nlm.nih.gov/sra/SRX8787566> [accn]).

Annotation information is available in Supplementary Tables S2 and S4. All scripts used for data analysis as well as de novo transcriptomes assembled for this study are available at <https://github.com/komoro/Marine-Turtle-Blood-Transcriptomes>.

Declarations

Ethics approval and consent to participate

No animals were anesthetized or euthanized as part of this study. All sample collection methods were rigorously reviewed and approved under IACUC protocols and NMFS permits (see Table S1 for specific permit numbers for each sample) which are in compliance with US guidelines for work with animals.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Environmental Conservation, University of Massachusetts, Amherst, MA, USA. ²Marine Turtle Biology and Assessment Program, Protected Species Division, Pacific Islands Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Honolulu, HI, USA. ³Marine Mammal and Turtle Division, Southwest Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, La Jolla, CA, USA. ⁴Chemical Sciences Division, National Institute of Standards and Technology, Hawai'i Pacific University, Waimanalo, HI, USA. ⁵Department of Anatomy, Physiology and Cell Biology, University of California, Davis, CA, USA. ⁶Center for Biodiversity and Conservation, American Museum of Natural History, New York, NY, USA. ⁷New York University, New York, NY, USA. ⁸School of Agricultural, Forest, and Environmental Sciences, Clemson University, Clemson, SC, USA. ⁹United States Geological Survey, Wetland and Aquatic Research Center, Gainesville, FL, USA. ¹⁰Section of Molecular Biology, Division of Biological Sciences, University of California, San Diego, La Jolla, CA, USA. ¹¹Marine Mammal and Turtle Division, Southwest Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Moss Landing, CA 95039, USA. ¹²Moss Landing Marine Laboratories, San Jose State University, Moss Landing, CA 95039, USA. ¹³Department of Biology, San Diego State University, San Diego, CA, USA. ¹⁴Rainbow Connection Research, Guam, USA. ¹⁵Jessy's Tag Services, Saipan, Commonwealth of the Northern Mariana Islands, USA. ¹⁶Hawai'i Preparatory Academy, Kamuela, HI, USA. ¹⁷Golden Honu Services of Oceania, Honolulu, HI, USA.

Received: 4 December 2020 Accepted: 23 April 2021

Published online: 13 May 2021

References

- Alvarez M, Schrey AW, Richards CL. Ten years of transcriptomics in wild populations: what have we learned about their ecology and evolution? *Mol Ecol*. 2015;24(4):710–25. <https://doi.org/10.1111/mec.13055>.
- Roberts WR, Roalson EH. Comparative transcriptome analyses of flower development in four species of *Achimenes* (*Gesneriaceae*). *BMC Genomics*. 2017;18(1):240.
- Carruthers M, Yurchenko AA, Augley JJ, Adams CE, Herzyk P, Elmer KR. De novo transcriptome assembly, annotation and comparison of four ecological and evolutionary model salmonid fish species. *BMC Genomics*. 2018;19(1):32.
- Moreno-Santillán DD, Machain-Williams C, Hernández-Montes G, Ortega J. De novo transcriptome assembly and functional annotation in five species of bats. *Sci Rep*. 2019;9(1):6222.
- Page TM, McDougall C, Diaz-Pulido G. De novo transcriptome assembly for four species of crustose coralline algae and analysis of unique orthologous genes. *Sci Rep*. 2019;9(1):12611.
- Bay RA, Palumbi SR. Transcriptome predictors of coral survival and growth in a highly variable environment. *Ecol Evol*. 2017;7(13):4794–803. <https://doi.org/10.1002/ece3.2685>.
- Kumaresan V, Nizam F, Ravichandran G, Viswanathan K, Palanisamy R, Bhatt P, et al. Transcriptome changes of blue-green algae, *Arthrospira* sp. in

- response to sulfate stress. *Algal Res.* 2017;23:96–103. <https://doi.org/10.1016/j.algal.2017.01.012>.
8. Watson H, Videvall E, Andersson MN, Isaksson C. Transcriptome analysis of a wild bird reveals physiological responses to the urban environment. *Sci Rep.* 2017;7(1):1–10.
 9. Trego ML, Whitehead A, Kellar NM, Lauf M, Lewison RL. Tracking transcriptomic responses to endogenous and exogenous variation in cetaceans in the Southern California Bight. *Conserv Physiol.* 2019;7(1):coz018.
 10. Morey JS, Neely MG, Lunardi D, Anderson PE, Schwacke LH, Campbell M, et al. RNA-Seq analysis of seasonal and individual variation in blood transcriptomes of healthy managed bottlenose dolphins. *BMC Genomics.* 2016;17(1):720. <https://doi.org/10.1186/s12864-016-3020-8>.
 11. Morey JS, Burek Huntington KA, Campbell M, Clauss TM, Goertz CE, Hobbs RC, et al. De novo transcriptome assembly and RNA-Seq expression analysis in blood from beluga whales of Bristol Bay. *AK Mar Genomics.* 2017;35:77–92. <https://doi.org/10.1016/j.margen.2017.08.001>.
 12. Liew C-C, Ma J, Tang H-C, Zheng R, Dempsey AA. The peripheral blood transcriptome dynamically reflects system wide biology: a potential diagnostic tool. *J Lab Clin Med.* 2006;147(3):126–32. <https://doi.org/10.1016/j.lab.2005.10.005>.
 13. Di Meo A, Bartlett J, Cheng Y, Pasic MD, Yousef GM. Liquid biopsy: a step forward towards precision medicine in urologic malignancies. *Mol Cancer.* 2017;16(1):80. <https://doi.org/10.1186/s12943-017-0644-5>.
 14. Caza F, de Boissel PGJ, Villemur R, Betoulle S, St-Pierre Y. Liquid biopsies for omics-based analysis in sentinel mussels. *PLoS One.* 2019;14(10):e0223525. <https://doi.org/10.1371/journal.pone.0223525>.
 15. Waits DS, Simpson DY, Sparkman AM, Bronikowski AM, Schwartz TS. The utility of reptile blood transcriptomes in molecular ecology. *Mol Ecol Res.* 2020;20(1):308.
 16. Huang Z, Gallot A, Lao NT, Puechmaile SJ, Foley NM, Jebb D, et al. A nonlethal sampling method to obtain, generate and assemble whole blood transcriptomes from small, wild mammals. *Mol Ecol Res.* 2016;16(1):150–62. <https://doi.org/10.1111/1755-0998.12447>.
 17. Chiari Y, Galtier N. RNA extraction from sauropsids blood: evaluation and improvement of methods. *Amphib-Reptil.* 2011;32(1):136–9. <https://doi.org/10.1163/017353710X543010>.
 18. Townsend K, Ness J, Hoguet J, Stacy NI, Komoroske LK, Lynch JM. Testing the stability of plasma protein and whole blood RNA in archived blood of loggerhead sea turtles, *Caretta caretta*. *Biopreserv Biobank.* 2020;18(5):358.
 19. Komoroske LM, Jensen MP, Stewart KR, Shamblin BM, Dutton PH. Advances in the application of genetics in marine turtle biology and conservation. *Front Mar Sci.* 2017;4. <https://doi.org/10.3389/fmars.2017.00156>.
 20. Hays GC, Hawkes LA. Satellite tracking sea turtles: opportunities and challenges to address key questions. *Front Mar Sci.* 2018;5. <https://doi.org/10.3389/fmars.2018.00432>.
 21. IUCN. The IUCN Red List of Threatened Species. 2020. Available from: <https://www.iucnredlist.org>
 22. U.S. Fish & Wildlife Service. Summary of listed species and recovery plans—US Fish & Wildlife Service species reports. In: U.S. Fish and Wildlife Service, ECOS Environmental Conservation Online System. 2020; Available from: http://ecos.fws.gov/tess_public/pub/Boxscore.do
 23. Alpanidou V, Markantonatou V, Mazaris AD. Thermal heterogeneity along the migration corridors of sea turtles: implications for climate change ecology. *J Exper Mar Biol Ecol.* 2019;520:151223. <https://doi.org/10.1016/j.jembe.2019.151223>.
 24. Chaloupka M, Balazs GH, Work TM. Rise and fall over 26 years of a marine epizootic in Hawaiian green sea turtles. *J Wildl Dis.* 2009;45(4):1138–42. <https://doi.org/10.7589/0090-3558-45.4.1138>.
 25. Clukey KE, Lepczyk CA, Balazs GH, Work TM, Li QX, Bachman MJ, et al. Persistent organic pollutants in fat of three species of Pacific pelagic longline caught sea turtles: accumulation in relation to ingested plastic marine debris. *Sci Tot Environ.* 2018;610–611:402–11.
 26. Jensen MP, Allen CD, Eguchi T, Bell IP, LaCasella EL, Hilton WA, et al. Environmental warming and feminization of one of the largest sea turtle populations in the world. *Curr Biol.* 2018;28(1):154–159.e4.
 27. Work TM, Balazs GH. Pathology and distribution of sea turtles landed as bycatch in the Hawaii-based North Pacific pelagic longline fishery. *J Wildl Dis.* 2010;46(2):422–32. <https://doi.org/10.7589/0090-3558-46.2.422>.
 28. Naro-Maciel E, Le M, FitzSimmons NN, Amato G. Evolutionary relationships of marine turtles: a molecular phylogeny based on nuclear and mitochondrial genes. *Mol Phylogenet Evol.* 2008;49(2):659–62. <https://doi.org/10.1016/j.ympev.2008.08.004>.
 29. Bostrom BL, Jones TT, Hastings M, Jones DR. Behaviour and physiology: the thermal strategy of leatherback turtles. *PLoS One.* 2010;5(11):e13925. <https://doi.org/10.1371/journal.pone.0013925>.
 30. Wyneken J. Anatomy of the leatherback turtle. In: *The Leatherback Turtle: Biol and Conservation*: Johns Hopkins University Press; 2015. p. 32–48.
 31. Duchene S, Frey A, Alfaro-Núñez A, Dutton PH, Thomas P, Gilbert M, et al. Marine turtle mitogenome phylogenetics and evolution. *Mol Phylogenet Evol.* 2012;65(1):241–50. <https://doi.org/10.1016/j.ympev.2012.06.010>.
 32. Pereira AG, Sterli J, Moreira FRR, Schrago CG. Multilocus phylogeny and statistical biogeography clarify the evolutionary history of major lineages of turtles. *Mol Phylogenet Evol.* 2017;113:59–66. <https://doi.org/10.1016/j.ympev.2017.05.008>.
 33. Avise JC, Bowen BW, Lamb T, Meylan AB, Bermingham E. Mitochondrial DNA evolution at a turtle's pace: evidence for low genetic variability and reduced microevolutionary rate in the Testudines. *Mol Biol Evol.* 1992;9(3):457–73. <https://doi.org/10.1093/oxfordjournals.molbev.a040735>.
 34. Komoroske LM, Miller MR, O'Rourke SM, Stewart KR, Jensen MP, Dutton PH. A versatile rapture (RAD-capture) platform for genotyping marine turtles. *Mol Ecol Res.* 2019;19(2):497–511. <https://doi.org/10.1111/1755-0998.12980>.
 35. Martínez-Fernández M, Bernatchez L, Rolán-Alvarez E, Quesada H. Insights into the role of differential gene expression on the ecological adaptation of the snail *Littorina saxatilis*. *BMC Evol Biol.* 2010;10(1):356. <https://doi.org/10.1186/1471-2148-10-356>.
 36. Kenkel CD, Matz MV. Gene expression plasticity as a mechanism of coral adaptation to a variable environment. *Nat Ecol Evol.* 2016;1(1):1–6.
 37. Seminoff JA, Allen CD, Balazs GH, Dutton PH, Eguchi T, Haas HL, et al. Status review of the green turtle (*Chelonia mydas*) under the Endangered Species Act. NOAA Tech Memor NOAA-NMFS-SWFSC-539; 2015. p. 571.
 38. Conesa A, Götz S, García-Gómez JM, Terol J, Talón M, Robles M. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics.* 2005;21(18):3674–6. <https://doi.org/10.1093/bioinformatics/bti610>.
 39. Kanehisa M. Toward understanding the origin and evolution of cellular organisms. *Protein Sci.* 2019;28(11):1947–51. <https://doi.org/10.1002/pro.3715>.
 40. Emms DM, Kelly S. OrthoFinder: phylogenetic orthology inference for comparative genomics. *Genome Biol.* 2019;20(1):1.
 41. Akbarzadeh A, Günther OP, Houde AL, Li S, Ming TJ, Jeffries KM, et al. Developing specific molecular biomarkers for thermal stress in salmonids. *BMC Genomics.* 2018;19(1):749. <https://doi.org/10.1186/s12864-018-5108-9>.
 42. Miller KM, Günther OP, Li S, Kaukinen KH, Ming TJ. Molecular indices of viral disease development in wild migrating salmon. *Conserv Physiol.* 2017;5(1):cox036.
 43. Yadette F, Karlsen OA, Lanzén A, Berg K, Olsvik P, Hogstrand C, et al. Global transcriptome analysis of Atlantic cod (*Gadus morhua*) liver after in vivo methylmercury exposure suggests effects on energy metabolism pathways. *Aquat Toxicol.* 2013;126:314–25. <https://doi.org/10.1016/j.aquatox.2012.09.013>.
 44. Trego ML, Hoh E, Whitehead A, Kellar NM, Lauf M, Datuin DO, et al. Contaminant exposure linked to cellular and endocrine biomarkers in Southern California bottlenose dolphins. *Environ Sci Technol.* 2019;53(7):3811–22. <https://doi.org/10.1021/acs.est.8b06487>.
 45. Haas BJ, Papanicolaou A, Yassour M, Grabherr M, Blood PD, Bowden J, et al. De novo transcript sequence reconstruction from RNA-seq using the trinity platform for reference generation and analysis. *Nat Protoc.* 2013;8(8):1494–512. <https://doi.org/10.1038/nprot.2013.084>.
 46. Freedman AH, Clamp M, Sackton TB. Error, noise and bias in de novo transcriptome assemblies. *Mol Ecol Res.* 2021;21(1):18–29. <https://doi.org/10.1111/1755-0998.13156>.
 47. Bentley BP, Haas BJ, Tedeschi JN, Berry O. Loggerhead Sea turtle embryos (*Caretta caretta*) regulate expression of stress response and developmental genes when exposed to a biologically realistic heat stress. *Mol Ecol.* 2017;26(11):2978–92. <https://doi.org/10.1111/mec.14087>.
 48. Lübke A, Schaffner W. Tissue-specific gene expression. *Trends Neurosci.* 1985;8:100–4. [https://doi.org/10.1016/0166-2236\(85\)90046-3](https://doi.org/10.1016/0166-2236(85)90046-3).
 49. Sonawane AR, Platig J, Fagny M, Chen C-Y, Paulson JN, Lopes-Ramos CM, et al. Understanding tissue-specific gene regulation. *Cell Rep.* 2017;21(4):1077–88. <https://doi.org/10.1016/j.celrep.2017.10.001>.
 50. Telemeco RS, Simpson DY, Tylan C, Langkilde T, Schwartz TS. Contrasting responses of lizards to divergent ecological stressors across biological levels of organization. *Integr Comp Biol.* 2019;59(2):292–305. <https://doi.org/10.1093/icb/icz071>.

51. Byrne A, Supple MA, Volden R, Laidre KL, Shapiro B, Vollmers C. Depletion of hemoglobin transcripts and long-read sequencing improves the transcriptome annotation of the polar bear (*Ursus maritimus*). *Front Genet.* 2019;10. <https://doi.org/10.3389/fgene.2019.00643>.
52. Westermann AJ, Gorski SA, Vogel J. Dual RNA-seq of pathogen and host. *Nat Rev Microbiol.* 2012;10(9):618–30. <https://doi.org/10.1038/nrmicro2852>.
53. Lehman BM, Johnson RC, Adkison M, Burgess OT, Cannon RE, Fangué NA, et al. Disease in central valley salmon: status and lessons from other systems. *San Francisco Estuary Watershed Sci.* 2020;18(3):1–31. <https://doi.org/10.15447/SFEWS.2020V18ISS3ART2>.
54. Santos JC, Tarvin RD, O'Connell LA, Blackburn DC, Coloma LA. Diversity within diversity: parasite species richness in poison frogs assessed by transcriptomics. *Mol Phylogenet Evol.* 2018;125:40–50. <https://doi.org/10.1016/j.ympev.2018.03.015>.
55. Larsen PA, Hayes CE, Williams CV, Junge RE, Razafindramanana J, Mass V, et al. Blood transcriptomes reveal novel parasitic zoonoses circulating in Madagascar's lemurs. *Biol Lett.* 2016;12(1):20150829. <https://doi.org/10.1098/rsbl.2015.0829>.
56. Liu G, Zhang H, Sun G, Zhao C, Shang S, Gao X, et al. Characterization of the peripheral blood transcriptome and adaptive evolution of the MHC I and TLR gene families in the wolf (*Canis lupus*). *BMC Genomics.* 2017;18(1):584. <https://doi.org/10.1186/s12864-017-3983-0>.
57. McGaugh SE, Bronikowski AM, Kuo C-H, Reding DM, Addis EA, Flagel LE, et al. Rapid molecular evolution across amniotes of the IIS/TOR network. *PNAS.* 2015;112(22):7055–60. <https://doi.org/10.1073/pnas.1419659112>.
58. Griffiths JS, Pan T-CF, Kelly MW. Differential responses to ocean acidification between populations of *Balanophyllia elegans* corals from high and low upwelling environments. *Mol Ecol.* 2019;28:2715.
59. Whitehead A, Crawford DL. Variation in tissue-specific gene expression among natural populations. *Genome Biol.* 2005;6(2):R13. <https://doi.org/10.1186/gb-2005-6-2-r13>.
60. Schoville SD, Barreto FS, Moy GW, Wolff A, Burton RS. Investigating the molecular basis of local adaptation to thermal stress: population differences in gene expression across the transcriptome of the copepod *Tigriopus californicus*. *BMC Evol Biol.* 2012;12(1):170. <https://doi.org/10.1186/1471-214-8-12-170>.
61. Garcia TI, Shen Y, Crawford D, Oleksiak MF, Whitehead A, Walter RB. RNA-Seq reveals complex genetic response to Deepwater horizon oil release in *Fundulus grandis*. *BMC Genomics.* 2012;13(1):474. <https://doi.org/10.1186/1471-2164-13-474>.
62. Cortés-Gómez AA, Morcillo P, Guardiola FA, Espinosa C, Esteban MA, Cuesta A, et al. Molecular oxidative stress markers in olive ridley turtles (*Lepidochelys olivacea*) and their relation to metal concentrations in wild populations. *Environ Pollut.* 2018;233:156–67. <https://doi.org/10.1016/j.envpol.2017.10.046>.
63. Cocci P, Mosconi G, Bracchetti L, Nalocca JM, Frapiccini E, Marini M, et al. Investigating the potential impact of polycyclic aromatic hydrocarbons (PAHs) and polychlorinated biphenyls (PCBs) on gene biomarker expression and global DNA methylation in loggerhead sea turtles (*Caretta caretta*) from the Adriatic Sea. *Sci Tot Environ.* 2018;619–620:49–57.
64. Lehnert K, Siebert U, Reißmann K, Bruhn R, McLachlan MS, Müller G, et al. Cytokine expression and lymphocyte proliferative capacity in diseased harbor porpoises (*Phocoena phocoena*) – biomarkers for health assessment in wildlife cetaceans. *Environ Pollut.* 2019;247:783–91. <https://doi.org/10.1016/j.envpol.2019.01.079>.
65. Van Houtan KS, Hargrove SK, Balazs GH. Land use, macroalgae, and a tumor-forming disease in marine turtles. *Thrush S, editor. PLoS One.* 2010;5(9):e12900.
66. Aguirre AA, Lutz PL. Marine turtles as sentinels of ecosystem health: is fibropapillomatosis an indicator? *EcoHealth.* 2004;1(3):275.
67. Work TM. Cancer in sea turtles. *Hawaii Med J.* 2005;64:23–4. <https://core.ac.uk/download/pdf/77122956.pdf>.
68. Hargrove SA, Work TM, Brunson S, Foley AM, Balazs GH. Proceedings of the 2015 international summit on fibropapillomatosis : global status, trends, and population impacts. NOAA Technical Memorandum NMFS-PIFSC-54; 2016.
69. Jensen MP, FitzSimmons NN, Bourjea J, Hamabata T, Reece J, Dutton PH. The evolutionary history and global phylogeography of the green turtle (*Chelonia mydas*). *J Biogeogr.* 2019;46(5):860–70. <https://doi.org/10.1111/jbi.13483>.
70. Komoroske LM, Lewison RL, Seminoff JA, Deheyn DD, Dutton PH. Pollutants and the health of green sea turtles resident to an urbanized estuary in San Diego. *CA Chemosphere.* 2011;84(5):544–52. <https://doi.org/10.1016/j.chemosphere.2011.04.023>.
71. Balazs GH, Parker DM. Ocean pathways and residential foraging locations for satellite tracked green turtles breeding at French Frigate Shoals in the Hawaiian Islands. *Micronesica.* 2017;4:1.
72. Bennett P, Keuper-Bennett U, Balazs GH. Remigration and residency of Hawaiian green turtles in coastal waters of Honokowai, West Maui, Hawaii. *Proceedings of the Twentieth Annual Symposium on Sea Turtle Biology and Conservation US Dept Commerce, NOAA Tech Memo NMFS-SEFSC-477;* 2002. p. 289290.
73. Greytak SR, Champlin D, Callard GV. Isolation and characterization of two cytochrome P450 aromatase forms in killifish (*Fundulus heteroclitus*): differential expression in fish from polluted and unpolluted environments. *Aquat Toxicol.* 2005;71(4):371–89. <https://doi.org/10.1016/j.aquatox.2004.12.007>.
74. Jeffries KM, Hinch SG, Gale MK, Clark TD, Lotto AG, Casselman MT, et al. Immune response genes and pathogen presence predict migration survival in wild salmon smolts. *Mol Ecol.* 2014;23(23):5803–15. <https://doi.org/10.1111/mec.12980>.
75. Pujade Busqueta L, Crocker DE, Champagne CD, McCormley MC, Deyarmin JS, Houser DS, et al. A blubber gene expression index for evaluating stress in marine mammals. *Conserv Physiol.* 2020;8(1):coa082.
76. Allen CD, Robbins MN, Eguchi T, Owens DW, Meylan AB, Meylan PA, et al. First assessment of the sex ratio for an East Pacific green sea turtle foraging aggregation: validation and application of a testosterone ELISA. *PLoS One.* 2015;10(10):e0138861. <https://doi.org/10.1371/journal.pone.0138861>.
77. Tezak B, Sifuentes-Romero I, Milton S, Wyneken J. Identifying sex of neonate turtles with temperature-dependent sex determination via small blood samples. *Sci Rep.* 2020;10(1):5012. <https://doi.org/10.1038/s41598-020-61984-2>.
78. Owens DW, Ruiz GJ. New methods of obtaining blood and cerebrospinal fluid from marine turtles. *Herpetologica.* 1980;36(1):17–20.
79. Komoroske LM. NEB directional kit working RNA-sequencing library preparation protocol. Available from: <https://github.com/MolEcolConsLab/Wet-Lab-Protocols/blob/master/NEBdirectionalkit.working>. SOP.Komoroske.lab_JP_LK_SB.docx.
80. Buffalo V. Scythe – a Bayesian adapter trimmer [internet]. 2014. Available from: <https://github.com/vsbuffalo/scythe>
81. Joshi J, Fass J. Sickle: A sliding-window, adaptive, quality-based trimming tool for FastQ files. Available from: <https://github.com/najoshi/sickle>
82. DeBiasse MB, Kawji Y, Kelly MW. Phenotypic and transcriptomic responses to salinity stress across genetically and geographically divergent *Tigriopus californicus* populations. *Mol Ecol.* 2018;27(7):1621–32. <https://doi.org/10.1111/mec.14547>.
83. Wang Z, Pascual-Anaya J, Zadissa A, Li W, Niimura Y, Huang Z, et al. The draft genomes of soft-shell turtle and green sea turtle yield insights into the development and evolution of the turtle-specific body plan. *Nat Genet.* 2013;45(6):701–6. <https://doi.org/10.1038/ng.2615>.
84. Grabherr MG, Haas BJ, Yassour M, Levin JZ, Thompson DA, Amit I, et al. Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nat Biotechnol.* 2011;29(7):644–52. <https://doi.org/10.1038/nbt.1883>.
85. Smith-Unna R, Boursnell C, Patro R, Hibberd JM, Kelly S. TransRate: reference-free quality assessment of de novo transcriptome assemblies. *Genome Res.* 2016;26(8):1134–44. <https://doi.org/10.1101/gr.196469.115>.
86. Fu L, Niu B, Zhu Z, Wu S, Li W. CD-HIT: accelerated for clustering the next-generation sequencing data. *Bioinformatics.* 2012;28(23):3150–2. <https://doi.org/10.1093/bioinformatics/bts565>.
87. Patro R, Duggal G, Love MI, Irizarry RA, Kingsford C. Salmon provides fast and bias-aware quantification of transcript expression. *Nat Methods.* 2017;14(4):417–9. <https://doi.org/10.1038/nmeth.4197>.
88. Brown CT. gather-counts.py. 2015. Available from: <https://github.com/dib-lab/2017-dibsi-metagenomics/blob/master/gather-counts.py>
89. Robinson MD, McCarthy DJ, Smyth GK. edgeR: a bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics.* 2010;26(1):139–40. <https://doi.org/10.1093/bioinformatics/btp616>.
90. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics.* 2015;31(19):3210–2. <https://doi.org/10.1093/bioinformatics/btv351>.

91. Buchfink B, Xie C, Huson DH. Fast and sensitive protein alignment using DIAMOND. *Nat Methods*. 2015;12(1):59–60. <https://doi.org/10.1038/nmeth.3176>.
92. Demasius W, Weikard R, Hadlich F, Müller KE, Kühn C. Monitoring the immune response to vaccination with an inactivated vaccine associated to bovine neonatal pancytopenia by deep sequencing transcriptome analysis in cattle. *Vet Res*. 2013;44(1):93. <https://doi.org/10.1186/1297-9716-44-93>.
93. Correia CN, McLoughlin KE, Nalpas NC, Magee DA, Browne JA, Rue-Allbrecht K, et al. RNA sequencing (RNA-Seq) reveals extremely low levels of reticulocyte-derived globin gene transcripts in peripheral blood from horses (*Equus caballus*) and cattle (*Bos taurus*). *Front Genet*. 2018;9. <https://doi.org/10.3389/fgene.2018.00278>.
94. Langmead B, Salzberg SL. Fast gapped-read alignment with bowtie 2. *Nat Methods*. 2012;9(4):357–9. <https://doi.org/10.1038/nmeth.1923>.
95. Haas B, Papanicolaou A. Transdecoder (find coding regions within transcripts) [internet]. 2018. Available from: <http://transdecoder.sf.net>
96. R Core Team. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing; 2020.
97. Conway JR, Lex A, Gehlenborg N. UpSetR: an R package for the visualization of intersecting sets and their properties. *Bioinformatics*. 2017;33(18):2938–40. <https://doi.org/10.1093/bioinformatics/btx364>.
98. Götz S, García-Gómez JM, Terol J, Williams TD, Nagaraj SH, Nueda MJ, et al. High-throughput functional annotation and data mining with the Blast2GO suite. *Nucleic Acids Res*. 2008;36(10):3420.
99. Huerta-Cepas J, Szklarczyk D, Heller D, Hernández-Plaza A, Forslund SK, Cook H, et al. eggNOG 5.0: a hierarchical, functionally and phylogenetically annotated orthology resource based on 5090 organisms and 2502 viruses. *Nucleic Acids Res*. 2019;47(D1):D309–14. <https://doi.org/10.1093/nar/gky1085>.
100. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. Limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res*. 2015;43(7):e47. <https://doi.org/10.1093/nar/gkv007>.
101. Alexa A, Rahnenfuhrer J. topGO: enrichment analysis for gene ontology; 2019.
102. Alexa A, Rahnenfuhrer J. Gene set enrichment analysis with topGO. 2020. <https://bioconductor.org/packages/release/bioc/vignettes/topGO/inst/doc/topGO.pdf>
103. Walter W, Sánchez-Cabo F, Ricote M. GOplot: an R package for visually combining expression data with functional analysis. *Bioinformatics*. 2015; 31(17):2912–4. <https://doi.org/10.1093/bioinformatics/btv300>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

