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REVIEW ARTICLE Mechanisms of perinatal arterial ischemic stroke

David Fernández-López¹, Niranjana Natarajan², Stephen Ashwal³ and Zinaida S Vexler¹

The incidence of perinatal stroke is high, similar to that in the elderly, and produces a significant morbidity and severe long-term neurologic and cognitive deficits, including cerebral palsy, epilepsy, neuropsychological impairments, and behavioral disorders. Emerging clinical data and data from experimental models of cerebral ischemia in neonatal rodents have shown that the pathophysiology of perinatal brain damage is multifactorial. These studies have revealed that, far from just being a smaller version of the adult brain, the neonatal brain is unique with a very particular and age-dependent responsiveness to hypoxia-ischemia and focal arterial stroke. In this review, we discuss fundamental clinical aspects of perinatal stroke as well as some of the most recent and relevant findings regarding the susceptibility of specific brain cell populations to injury, the dynamics and the mechanisms of neuronal cell death in injured neonates, the responses of neonatal blood-brain barrier to stroke in relation to systemic and local inflammation, and the long-term effects of stroke on angiogenesis and neurogenesis. Finally, we address translational strategies currently being considered for neonatal stroke as well as treatments that might effectively enhance repair later after injury.

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INTRODUCTION

Perinatal arterial ischemic stroke is common and produces a significant morbidity and severe long-term neurologic and cognitive deficits, including cerebral palsy, epilepsy, neurodevelopmental disabilities, behavioral disorders, and impaired vision and language function.¹⁻⁴ More than half of all children with cerebral palsy are born at term and in many instances the etiology is related to some form of cerebrovascular focal or global insult.⁵ The pathophysiology of perinatal brain damage has proven to be multifactorial.⁶ With the availability and safe use of magnetic resonance imaging (MRI) in newborns, older infants, and children over the past two decades, much has been learned about how the stroke phenotype differs in these age groups compared with adults, as well as the underlying causes and pathophysiology. From an experimental perspective, stroke models in neonatal animals are technically very challenging when one considers the surgical difficulties based on size (e.g., body weight of a postnatal day 7 (P7) rat pup is < 15 g and of a P9 mouse pup is <5 g). Therefore, the first and most commonly used model to mimic ischemic brain injury in term neonates is an hypoxia-ischemia (H-I) model in P7 to P12 rats and mice, which consists of ligation of the common carotid artery followed by systemic hypoxia.⁷ This model more commonly is used to mimic 'global' hypoxic-ischemic encephalopathy (HIE) in term human babies although models of bilateral H-I may be more advantageous for this purpose.⁸ More recently, age-appropriate models of focal arterial stroke involving middle cerebral artery (MCA) occlusion (MCAO) have been developed to mimic pediatric⁹ and neonatal¹⁰⁻¹³ focal arterial stroke. We will briefly review some of the salient clinical aspects of neonatal stroke, describe some of the known differences compared with childhood stroke, and discuss in more detail current experimental knowledge regarding both distinct and common pathophysiologic mechanisms of perinatal stroke as compared with those in childhood and adult stroke. We will then discuss how maturational differences may affect ways to protect the neonatal brain and enhance long-term repair.

BRAIN IMMATURITY AT BIRTH AFFECTS THE PATTERNS OF ISCHEMIC INJURY

The maturational stage of the brain at the time of injury is a key factor in the pattern of brain damage, including regional and cell type-specific susceptibility. In preterm newborns, brain injury associated with hypoxia-ischemia generally coincides with a time window of high susceptibility of oligodendrocyte progenitors (OLPs) to excitotoxicity, oxidative stress, and inflammation,^{14,15} adversely affecting OLP differentiation into mature, myelinating oligodendrocytes. The arrest in OLP differentiation in turn predisposes the brain to defective white-matter tract development, including periventricular white-matter injury and cerebral palsy.^{16–18} Subplate neurons, which exist transiently during human fetal brain development, are involved in the formation of visual thalamocortical projections, and are another cell population selectively vulnerable to H-I in preterm babies that leads to long-term deficits. Several reviews have discussed the unique features of fetal brain neuroinflammation and injury in humans and in corresponding injury models.^{17,19,20}

The patterns of ischemic injury in full-term newborns are different from those in preterm newborns;⁶ injury is no longer diffuse and is manifested focally in gray-matter regions, most commonly in striatum, thalamus, and cortex.



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ETIOLOGY OF PERINATAL STROKE

Perinatal stroke is defined as an insult occurring from 20 weeks gestational age, including *in utero*, to 28 days postnatally.¹ Subcategories within perinatal stroke include neonatal arterial ischemic stroke, neonatal cerebral sinovenous ischemic stroke, and presumed perinatal stroke. Perinatal arterial ischemic stroke, the primary focus of this review, has been defined more specifically by the NIH Workshop on Perinatal Stroke as a condition with acute encephalopathy, seizures, or neurologic deficit presenting in the term or preterm infant before the 29th postnatal day, with brain imaging confirming a parenchymal infarct in the appropriate arterial territory.^{4,21}

Perinatal stroke occurs in 1 in 2,300 to 1 in 5,000 live infant births,¹⁻⁴ and the estimated mortality rate of neonatal stroke is 3.49/100,000 annually.² These likely are underestimations, as to some degree neonatal brain imaging may be less commonly acquired in remote or low resource settings.

The maternal-fetal dyad, present in pregnancy and at delivery, gives rise to specific etiologic considerations with regard to neonatal stroke. Pathologic changes resulting in gestational diabetes, pregnancy-induced hypertension, and preeclampsia have long been linked to neonatal stroke.²² Preeclampsia reduces placental blood flow, resulting in fetal cerebral hypoperfusion and the potential for emboli or global or focal ischemic injury. In case control studies, factors present at delivery including fetal bradycardia, fetal decelerations, prolonged rupture of membranes, or prolonged second stage of labor are associated with neonatal stroke, although it remains unclear whether these represent a causal relationship, or are reflective of intrauterine or ongoing stroke. Additional risk factors associated with difficult delivery such as vacuum delivery and emergency Caesarian section are associated with an increased risk of neonatal stroke. A recent study identified an Apgar score of <7 at 5 minutes, as well as maternal fever >38°C and hypoglycemia as independent risk factors for perinatal stroke.23,24

Increasing evidence suggests that the placenta has a significant role in neonatal neurologic disorders. Placental abruption with resultant decreased perfusion has been associated with neonatal stroke as well as HIE. A study of placental pathology in neonates with perinatal arterial ischemic stroke showed evidence of chronic poor perfusion of the placenta with acute changes at the time of delivery.²⁵ Additionally, placental chorioamnionitis even in the absence of maternal symptomatology is associated with neonatal stroke. Previous studies in which infants and children were diagnosed with cerebral palsy, including hemiplegic cerebral palsy, later in life and in whom the placenta was available for pathologic examination, often showed a high incidence of chronic infarctions, vascular pathology, anomalies of umbilical cord insertion, or the length of the umbilical cord or vessels.^{26,27}

Pregnancy is considered to be a natural prothrombotic state.⁴ Pregnancy is associated with changes in hemostasis due to evolutionary changes to protect the pregnant mother from fatal hemorrhage at the time of delivery. These changes predispose the mother to thromboembolism and place the fetus and the placenta at risk for thromboemboli. During pregnancy, coagulation factors V, VII, VIII, IX, X, XII, and von Willebrand factor and plasma fibrinogen concentrations significantly increase.²⁸ Concomitantly, fibrinolytic activity is decreased in pregnancy, and returns to normal within 1 hour after placental delivery. Tissue plasminogen activator decreases, while PAI-1 activity increases.²⁸ Additionally, consumption of cocaine during pregnancy places the fetus at risk for arterial stroke due to vasoconstriction and vasospasm. Vasospasm increases the risk of *in utero* stroke in the first and third trimester.²⁹

The presence of at least one prothrombotic factor substantially increased the incidence of stroke from 24% in neonates in the control group to 68% of 91 term neonates.³⁰ Increased lipoprotein

(A) is considered as the most important prothrombotic risk factor in newborns. Other common prothrombotic factors associated with neonatal stroke include genetic polymorphisms of MTHFR (C677T), Factor V Leiden (G1691A), and prothrombin (G20210A).³⁰⁻³⁴ Acquired prothrombotic states of anticardiolipin antibodies, activated protein C or protein S deficiency, or antithrombin deficiency also have been linked to neonatal stroke.^{31–34} These factors in combination with other triggering factors such as neonatal septicemia, perinatal asphyxia, or patent foramen ovale may potentiate the risk of stroke. Intracerebral hemorrhage accounts for 10% to 15% of stroke and is a particularly severe form of stroke. Mutations of COL4A1 have been shown to cause intracerebral hemorrhage in mice and humans.^{35,36} COL4A1 encodes the alpha1 chain of type IV collagen (Cola1), a major component of basement membrane. COL4A1 mutant mice and humans with COL4A1 mutations show increased sensitivity of brain vessels to antithrombotic agents and markedly increase the incidence of intracerebral hemorrhage that is associated with birth trauma.

Infants with congenital heart disease are at a higher risk for perinatal stroke although the incidence is not clearly known.³⁷ Additionally, there is an increased incidence of stroke in the setting of cardiac surgery. With the incidence of 1 in 125 newborns having congenital heart disease, cardioembolic events resulting in stroke are another potential etiology. In contrast, one case control study with a relatively small number of newborns with stroke found that none had associated congenital heart disease.⁵ There is currently no consensus regarding the evaluation of neonates suspected of having a stroke. Recent reviews and workshops have suggested various diagnostic approaches.^{4,38,39}

The rate of neonatal stroke recurrence is low (i.e., <3%), and recurrence typically occurs in neonates who have prothrombotic factors or congenital heart disease.⁴⁰ Recurrence rates are higher in neonates with congenital heart disease, presumably due to a combination of factors including the type of congenital heart disease, the peri-operative cardiovascular status of the neonate, and the potential risk of complication from the needed surgical interventions such as cardiac reconstruction or cardiac bypass.

CLINICAL PRESENTATION OF PERINATAL STROKE

Most babies with neonatal stroke present during the first 3 days of life with symptoms including focal seizures, apnea, chewing or bicycling movements, and persistent feeding difficulties. Many newborns will have no lateralizing findings on examination. A variable percentage of neonates will have focal findings such as tone differences or an asymmetric Moro response. Hemiplegia is rare, and evolves over time. This has been attributed to the structural and functional changes in the ipsilateral corticospinal tract in the immature brain at the time of injury. Studies utilizing transcranial magnetic stimulation in infants with hemiplegic cerebral palsy have found that the ipsilateral corticospinal tract hypertrophies, when corticospinal tract injury is present in the contralateral hemisphere.⁴¹

Stroke more frequently affects the left hemisphere and often involves the MCA territory. The International Pediatric Stroke Study has shown that perinatal stroke occurs in the anterior circulation 70% of the time and in 73% of newborns the left hemisphere was affected.⁶ It has been suggested that the origin of the left carotid artery from the aorta allows a more direct vascular route to the brain as a corridor for cardiac emboli.^{42,43} Other considerations take into account the fetal circulatory system or the presence of a patent ductus arteriosus or patent foramen ovale.

Perinatal stroke may be missed initially if findings on examination are subtle or absent, but if new symptoms such as pathologic early handedness, hemiplegia, neurodevelopmental delay, or spasticity appear later in the first year of life, the condition will be described as 'presumed perinatal ischemic stroke'. Newborns who present with seizures secondary to stroke may be



nonencephalopathic and appear normal in the hours before the onset of seizures.

Epilepsy or motor impairment occurs in approximately one-half to two-thirds of injured neonates.^{4,22,23,44–47} Recent reports on the occurrence of recurrent seizures after perinatal arterial ischemic stroke has ranged between one-quarter and two-thirds of patients, with variable responses to therapy,^{47–49} with larger stroke size associated with the likelihood of seizures beyond the acute period.⁴⁹ Later development of infantile spasms is also an important complication of epilepsy due to perinatal stroke, accounting for 5% to 8% of cases.^{50,51}

Motor outcomes in neonates with stroke are largely dependent on the location of injury. The internal capsule as well as the deep gray matter involving the basal ganglia more often is associated with neuromotor disability.^{52,53} Cortical involvement in the MCA distribution with involvement of the basal ganglia and the internal capsule is associated with hemiplegia, while those with basal ganglia and internal capsule without cortical involvement also may have motor coordination difficulties.⁵⁴

Magnetic resonance imaging has played a pivotal role in detecting stroke in term neonates.^{6,55} Diffusion-weighted MRI early after injury has identified the presence of severely injured regions as well as regions with more modest diffusion restriction, 'the ischemic penumbra', with higher likelihood of recovery in the penumbra (Figure 1A). Advanced MRI methods have been developed to quantitatively measure several aspects of brain development. For example, diffusion tensor imaging has become a standard technique to measure the direction and magnitude of water movement, which becomes more restrictive with increasing development of brain microstructure such as neuronal dendritic structure, axonal connections, and myelination.⁵⁷ Diffusion tensor imaging fiber tractography has enabled delineation of abnormalities in the white-matter fiber tracts of injured newborns.58,59 Optical imaging is an emerging method that allows mapping of the structural connectivity of the developing human brain (brain 'connectome').60

ETIOLOGY AND PATTERNS OF PEDIATRIC STROKE DIFFER FROM THOSE OF PERINATAL STROKE

Epidemiologic data from the International Pediatric Stroke Study show that the incidence of pediatric stroke is ~ 1 in 50,000⁶¹, that ~ 50% of strokes in children are hemorrhagic, that the majority of the remaining strokes are arterial ischemic strokes,⁵⁵ that only 24% of children are neurologically normal after a stroke, and that 20% to 30% have recurrent strokes.

In contrast to neonates, children with stroke present with the acute onset of weakness or altered mentation; only one-fourth will present with seizures.⁶²⁻⁶⁴ In comparison, the incidence of seizures as a presenting symptom is $\sim 3\%$ in adults with stroke.⁶⁵ While the recurrence risk in neonatal strokes is low, the risk of recurrence for older children is nearly 20%.⁶⁶ Stroke etiology in children differs from strokes in adults or in neonates, in part owing to a higher incidence of certain arteriopathies.⁶⁷ Vascular damage associated with infection/inflammation is an additional risk factor.⁶⁸ Pediatric stroke may result from arteriopathies due to arterial dissection, moyamoya disease, as well as focal cerebral arteriopathies. Ongoing research continues to explore the role of inflammatory or infectious causes such as varicella or meningitis in pediatric arteriopathies. Moyamoya disease and infarction due to sickle cell disease occur in pediatric but not in neonatal stroke. Additional etiologies, similar to those found in neonates with stroke, include inherited and acquired thrombophilias and various subtypes of congenital heart disease. The Canadian Pediatric Stroke Registry found that 25% of children with stroke had underlying cardiac involvement.⁶

Neurodevelopmental outcomes in pediatric and neonatal stroke are distinguished not only by the differences in motor outcomes,

but also by cognitive outcomes such as perceptual reasoning, working memory, processing speed, and intellectual ability. In a study of 145 children, cognitive outcomes were compared in children with perinatal, early childhood, and later childhood onset arterial stroke.⁶⁹ The perinatal subset had poorer neuro-developmental outcomes compared to children with later stroke onset regardless of lesion location; neonates with subcortical lesions performed significantly worse than those with childhood onset stoke, while conversely, cortical lesions led to greater cognitive impairment in childhood onset stroke compared with perinatal stroke.⁶⁹

In a prospective study comparing MRI and subsequent motor function, hemiparesis was found in 24% of neonatal and 56% of pediatric stroke patients, with comparable distribution of strokes.⁷⁰ The explanation for the different types of motor disability has been suggested to be related to brain/vascular maturational differences at the time of injury. In normal infants without stroke, transcranial magnetic stimulation of the corticospinal tract resulting in contralateral and ipsilateral responses, typically are extinguished by 18 months of age. In infants with neonatal stroke, this ipsilateral response may remain in an attenuated state, suggesting cortical reorganization.^{70,71} As this ipsilateral response extinguishes with normal development, subsequent brain injury after childhood stroke may not allow reorganization in such a robust manner.

ANIMAL MODELS AND THE UNDERLYING MECHANISMS OF PERINATAL ARTERIAL STROKE

Numerous models have been developed in fetal and neonatal nonhuman primate, sheep, lambs, piglets, rabbits, and rodents to elucidate the pathophysiology of brain damage relevant to term and preterm infants.⁷² Although human physiology is more closely related to that of larger animals than to rodents, brain maturation at birth in most of these species corresponds to the developmental stage of the fetal human brain.⁷² This limitation, along with practical issues, has limited the use of larger species for studying mechanisms of neonatal stroke.

Models of permanent MCA ligation¹¹ and transient MCAO (tMCAO) induced in P7 to P10 rats^{10,12,73} accomplished by a transient insertion of a suture filament enabled studies of perinatal arterial stroke. Varying the length of tMCAO has allowed production of injuries of different severity.^{73–75} The presence of recirculation in the tMCAO model^{76,77} also serves to mimic reperfusion, which frequently occurs in arterial stroke in term babies.

Diffusion-weighted MRI has been instrumental in delineating early injury, in experimental neonatal models of focal stroke, including the ischemic core and penumbra.^{13,74,78,79} Figure 1B shows that diffusion-weighted MRI depicts expansion of tissue 'at risk' associated with longer MCAO duration. Other MRI modalities have been proven useful in determining the extent of microcirculatory disturbances during MCAO and after reperfusion (perfusion-sensitive MRI),⁷⁶ disruption of the blood-brain barrier (BBB) (Gd-enhanced T1W imaging),⁷⁶ elucidation of the evolving injury (T2W imaging),^{56,73,80} and changes in brain connectivity after injury during the neonatal period (diffusion tensor imaging).⁸¹

Studies in tMCAO and H–I models in neonatal rodents have shown that although the components of injury that induce cell death and brain injury are similar to those occurring in experimental models of preterm injury—excitotoxic, oxidant, and inflammatory components—the targeted cell populations and regions affected by cerebral hypoxia and ischemia are different. Excitotoxicity, triggered by accumulation of glutamate and other excitotoxic molecules in extracellular spaces after neuronal membrane depolarization, glutamate efflux, and failure of its uptake, is a common initial damaging process caused by focal or global H–I.⁸² The neonatal brain is more excitable and prone to oxidative stress than the adult brain due to higher levels

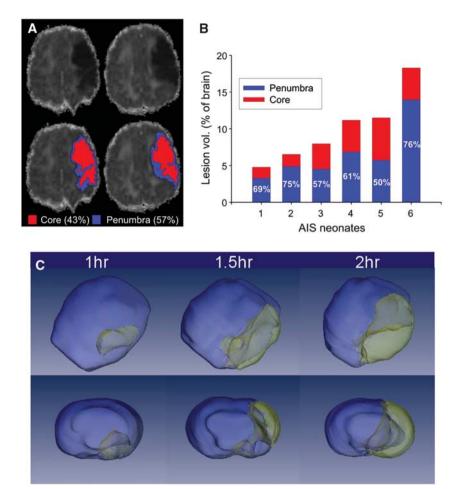


Figure 1. Magnetic resonance imaging (MRI)-based delineation of the ischemic core and penumbra. (**A**, **B**) Diffusion-weighted MRI (DWI) in human term babies 3 to 5 days after arterial ischemic stroke delineates the presence of penumbra (in red). Core and penumbra can be determined using a computational analysis method (Hierarchical Region Splitting) based on apparent diffusion coefficient threshold values as described in Ashwal *et al.*⁷⁹ The axis in (**B**) denotes data from six individual term neonates showing total lesion, core, and penumbral volumes. Note the wide variability in volumes that can occur in neonatal arterial ischemic stroke (AIS). (**C**) An example of injury volumes in a rat pup model of AIS. The 3D volume of 'tissue at risk' corresponds to the increasing duration of transient middle cerebral artery occlusion in the postnatal day 10 (P10) rats. Note the increasing size of the ischemic core (yellow) and penumbra (gray) in the hemisphere ipsilateral to the occlusion (courtesy of Stephen Ashwal and Andy Abenous, Department of Pediatrics, Lomo Linda University). These types of translational studies may provide better data regarding injury severity that could be applied to future clinical trials.

of glutamate receptor expression^{83,84} and the different composition of individual NMDA receptor subunits,⁸⁵ and interaction with downstream signaling cascades.⁸⁶ Several other distinct characteristics of the immature brain, such as high oxygen consumption, higher iron levels, and lower expression of several endogenous antioxidant enzymes, contribute to the higher vulnerability of the immature brain due to the deleterious actions of free radicals.⁸⁷ However, overexpression of antioxidant enzymes does not necessarily protect the neonatal brain against H-I, as was shown for superoxide dismutase overexpressing pups, and is dependent on the balance of free radical scavenging pathways^{88,89} and cell types.⁹⁰ Inflammation, the third major contributor to neonatal brain injury can induce neuronal death by itself as well as by enhancing excitotoxicity and oxidative stress through the release of cytokines, free radicals, and other toxic products or trigger release of excitotoxic molecules, including glutamate.91

MODES OF CELL DEATH IN THE INJURED NEONATAL BRAIN

Several concepts have emerged regarding the complexity of mechanisms underlying neuronal death in the injured immature brain. Neuronal apoptosis is commonly observed and may be

declines with maturation.⁹³ Apoptotic pathways are more readily activated in the immature brain in part due to the high expression of many of the key components in apoptotic pathways that counteract neuronal overproduction during early brain development (programmed neuronal death). Although necrosis has been typically considered as the predominant early neuronal death mechanism in the ischemic core, and apoptosis and autophagy have been related more to delayed cell death in the ischemic penumbra,⁹⁴ apoptosis is an important contributing neuronal death mechanism in the core after neonatal focal stroke.⁷⁴ While pharmacological inhibition of caspase-3 protects the neonatal brain against H-I,⁹² a lack of caspase-3 exacerbates injury via amplification of necrosis and caspase-3-independent injury,95 suggesting that complete abolishment of caspase-3 activity can be injurious rather than beneficial. The modes and timing of cell death may depend on the cell type. For example, death of OLPs via caspase-3-dependent mechanisms is relatively low early after H-I, whereas a more delayed and sustained increase in OLP cell death is largely caspase-3 dependent.⁹⁶

quite severe after global⁹² or focal ischemia-reperfusion⁷⁴ and

Mitochondria have a central role in modes of cell death and the ability to attenuate mitochondrial responses by counteracting oxidative stress via modulation of expression of proteins within the apoptotic pathways has been shown.^{97,98} Both the extrinsic (cell death receptor dependent) and intrinsic (mitochondria dependent) apoptotic pathways are activated in response to neonatal brain injury. Inhibition of apoptotic effectors downstream from death receptors, such as caspases, has resulted only in partial protection.^{99–101} It has been suggested that autophagy may precede apoptosis after neonatal brain ischemia.¹⁰² Importantly, failure to complete apoptosis results in programmed cell necrosis (also known as necroptosis), an intermediate form of cell death that exhibits features of both necrosis and apoptosis.^{94,103} Coexistence of several cell death mechanisms (i.e., necrosis, apoptosis, and autophagy) in the injured neonatal brain leads to the presence of a 'continuum' of hybrid cell death states and is thought to be unique to the newborn.⁹⁴

GENDER AND PERINATAL STROKE

Many central nervous system diseases display sexual dimorphism. Stroke, cerebral palsy, and related developmental disorders are more common in males than in females, ^{104,105} but the reasons for these differences remain uncertain. Sex hormones can provide protection against ischemic injury, but the neonatal brain may not be as influenced by these hormones as the adult brain. Gender predominance in the mechanisms of caspase-dependent and -independent apoptotic death has been shown after neonatal H–I^{106,107} and focal ischemia.¹⁰⁸ While pharmacological inhibition of poly ADP ribose polymerase prevents neuronal death after H–I preferentially in male pups,¹⁰⁶ inhibition of caspase-3 primarily protects female pups,¹⁰⁹ There is also increasing evidence that the response to therapy may be gender dependent, as has been shown for protection of female but not male neonates against H-I by an inducible nitric oxide synthase inhibitor 2-iminobiotin.¹¹⁰ A recent study did not observe gender differences in short-term injury outcome in tMCAO in a P20-P25 mouse model but did show that exogenously administered estrogen preferentially protected female but not males,¹¹¹ pointing out that gender and sex hormones may have independent effects on stroke in adults as well as in children.^{112,113} The existence of intrinsic gender-specific differences in cell death pathways in the neonatal period warrants further study to better understand the need for gender-specific therapies in neonatal and childhood stroke. A possible role of gender in stem cell treatment for neonatal HIE and stroke is being investigated.¹¹⁴

NEUROINFLAMMATION AND INJURY

Clinical and translational studies have shown that inflammatory responses after stroke are different in neonates compared with adults.^{91,115} For example, underdeveloped nonclassical pathways of complement activation are present in term infants as well as in rat pups^{116,117} likely have important age-dependent effects. Genetic deletion of various major mediators of inflammation, such as NADPH oxidase,¹¹⁸ interleukin 1 β (IL-1 β), IL-1 α , or both IL-1 $\alpha\beta^{119}$, is not neuroprotective in the neonatal brain, an effect opposite to that observed in adults. Inhibition of NFkB activation produces opposite effects after H–I, depending on the duration and timing of inhibition.¹²⁰ While data on the injurious role of toll-like receptor activation on the development of the immature brain are emerging,¹²¹ the extent to which long-lasting injury depends on receptor upregulation as opposed to the expression of appropriate ligands during early postnatal brain development has not yet been established.

Microglial activation has been traditionally considered as injurious in the ischemic brain due to their production of inflammatory mediators, free radicals, and other toxic molecules.¹²² This classic view of activated 'cytotoxic' microglia/macrophages, the 'bad guys' in the ischemic brain, is supported by numerous

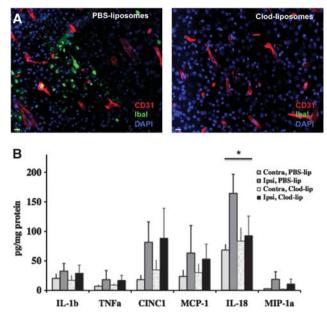


Figure 2. Depletion of microglial cells before a transient 3 hour middle cerebral artery occlusion (MCAO) in postnatal day 7 (P7) rats does not reduce the levels of several cytokines and chemokines elevated by injury induction. (**A**) While Ibal ⁺ microglia are abundant in the penumbra 24 hours after injury in pups treated with phosphate buffered saline (PBS)-liposomes, essentially no Ibal ⁺ microglia are seen in pups after intracerebral injection of liposomeencapsulated clodronate (Clod-liposomes). (**B**) Concentrations of cytokines and chemokines in injured (lpsi) and contralateral (Contra) hemispheres. Note that only the levels of interleukin 18 (IL-18) are significantly reduced by depletion of microglial cells. **P* < 0.05.

studies showing that inhibition of inflammatory pathways, which are typically activated in microglia, results in neuroprotection in adult stroke and neonatal H–I animal models.^{91,122} However, more recent data suggest that microglia may have a dual role after stroke, injurious or beneficial, depending on the heterogeneity of the macrophage population, the origin of these, resident microglia versus infiltrated monocytes, as well as their phenotypes. Microglia also produce mediators that can initially be harmful but later enhance repair through remodeling of the extracellular matrix during the chronic recovery phase after stroke.¹²³ Although it is well established that the number of macrophages is markedly increased 1 to 4 weeks after H-I, little is known about the effects of individual microglial phenotypes and the relative contribution of microglia and invading differentiated monocytes after neonatal brain injury. One study showed that the macrophage population predominantly consists of activated microglia, not invading peripheral monocytes, within the first 24 hours after tMCAO in neonatal rats.¹²⁴ Selective depletion of microglia before induction of tMCAO in neonatal rats revealed that, far from reducing injury, the absence of microglia leads to increased brain levels of inflammatory mediators (Figure 2) and larger infarct size.⁹⁰ This study also showed that when microglia are absent, cytokine and chemokine synthesis and release are increased in other brain cell types. These data suggest that at least a subpopulation of activated microglia acts as a 'buffering' component in regulating the 'pro' and 'anti' neuroinflammatory response induced by neonatal stroke. Removal of apoptotic debris after neonatal stroke is another potential beneficial effect of microglia. Depletion of these cells or genetic deletion of the scavenger receptor CD36, a receptor that participates in the recognition and engulfment of apoptotic cells by microglia and macrophages, worsens injury and leads to accumulation of apoptotic neurons that die by caspase-3-dependent mechanisms.^{13,90} The distinct patterns of superoxide accumulation in microglia/macrophages after acute adult¹²⁵ and neonatal^{13,90} stroke may be central to the age-related differences in stroke mechanisms.

As a rapid responder to alterations in the local microenvironment, microglia 'monitor' the changes in BBB homeostasis and leakage of plasma components into the brain, preserving vascular integrity, as was shown in adult models.^{126,127} Macrophages also promote vasculogenesis during brain development,¹²⁸ but the role of microglia in neurovascular function after neonatal stroke is unknown.

SYSTEMIC INFLAMMATION AND BLOOD-BRAIN BARRIER INTEGRITY

Epidemiologic studies have shown a strong association between fetal infection and inflammation (e.g., chorioamnionitis) with brain damage in the newborn or neurologic impairments in survivors.^{129,130} The sensitizing effect of systemic inflammation on perinatal brain lesions induced by H–I or excitotoxic insults has been shown but the mechanisms underlying these effects are not yet fully understood, particularly as to how BBB function is altered.

In the adult, disruption of the BBB has a key role in injury after stroke due to entrance of unwanted molecules and cells from the peripheral circulation. However, many drugs that cannot access the brain through the intact BBB are able to enter through disrupted BBB and act centrally. Emerging evidence suggests that the early postnatal BBB is not as permeable as once thought. Mechanisms of BBB function in the fetus are different from those in the adult.¹³¹ Tight junctions are present early in embryonic development, restricting entrance of proteins into the brain, while influx and efflux transporters are present during midgestation. At birth, the BBB is functional with no fenestrations in most brain regions.¹³² Blood-brain barrier endothelial proteins undergo major changes in expression from the postnatal period to adulthood.⁷⁷ As pericyte and astrocyte coverage increases with brain maturation, their control of neurovascular integrity in the normal brain increases.¹³³ However, BBB permeability after injury does not decrease linearly with age, as was shown by the finding that BBB disruption is much higher after a local inflammatory challenge (i.e., intrastriatal IL-1 β injection) in P21 rats than in 2-hour-old rat pups.¹³⁴

A recent comparative study provided direct evidence for the different functional BBB response to acute experimental stroke between neonates and adults.⁷⁷ Extravasation of albumin that circulated for 20 hours starting at 2 hours after reperfusion was shown to be increased in the injured adult rat brain (5- to 25-fold), but, in stark contrast, only a 2-fold increase occurred in the injured cortex and caudate in the neonate.⁷⁷ Blood-brain barrier permeability to intravascular tracers of various sizes, from large $(\sim 65 \text{ kDa})$ to relatively small (560 Da), also remained low in the cortex and caudate of neonates after acute injury, while leakage and brain accumulation of tracers are significantly increased in these regions in adults.⁷⁷ The transcriptional profiles in endothelial cells isolated from injured and noninjured adult and neonatal tissue in part explain the observed age differences in BBB susceptibility to stroke by showing that the patterns of upregulated and downregulated endothelial genes are largely nonoverlapping between the two ages (Figure 3), and that gene and protein expression of several tight junction components and basement membrane components (collagen IV and laminin) is better preserved in neonates.⁷⁷ Other studies observed increased BBB permeability during a subchronic injury phase after tMCAO in P7 rats⁷⁷ and P10 rats.⁷⁸ The affected region was larger in spontaneously hypertensive pups⁷⁸ than in normotensive pups of the same age.¹³⁵ As expected, permanent MCAO in P7 rats resulted in rapid BBB disruption and leukocyte extravasation, 136 suggesting that persistent lack of cerebral microcirculation contributes to the BBB collapse. Together, these data suggest

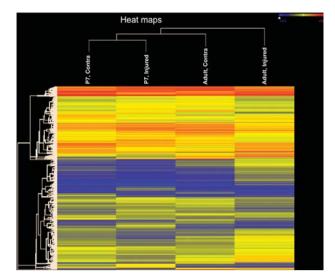


Figure 3. Stroke induces rapid nonoverlapping changes in gene expression in endothelial cells within injured regions in adults and neonates. Heatmaps obtained in endothelial cells isolated from injured adult and neonatal brains 24 hours after transient 3 hours MCAO, as described in Fernandez *et al.*⁷⁷ Heatmap visualization from a total of 31,042 probe sets shows that the expression levels of endothelial genes are markedly changed in injured regions compared with the corresponding contralateral anatomic regions in the same rats in each age group but that the patterns of changes are distinct after adult and neonatal stroke. The expression levels of genes are indexed by color.

that intrinsic developmental differences in basement membrane and extracellular matrix formation may contribute to a better preserved BBB integrity after acute neonatal arterial stroke.

Interaction of endothelial cells with peripheral leukocytes serves as an important contributor to BBB disruption (Figure 4). Both animal and human studies in the adult show that neutrophils infiltrate the brain and contribute to BBB disruption after transient cerebral ischemia (Figure 4B) and that neutropenia is neuroprotective.^{137,138} In contrast to adults, neutrophil infiltration in neonates is negligible after tMCAO⁷⁷ and is very limited¹³⁹ or brief¹⁴⁰ after H–I. Although the exact mechanisms that restrict neutrophil infiltration in the injured neonatal brain are yet to be understood, and it remains unclear whether the higher resistance of the neonatal BBB to stroke is a cause or a consequence of reduced leukocyte infiltration, the different patterns of expression of adhesion molecules and matrix metalloproteinases between neonatal and adult rodents, along with changing chemokine gradients between the brain and the blood, may limit neutrophil extravasation in the neonate.⁷⁷

Astrocytes and pericytes, cells known to participate in the maintenance of vascular impermeability, may contribute differently to neonatal and adult stroke (Figure 4A). In adults, astrocyte swelling, mediated by water channels (aquaporin-4) present in astrocyte end feet, is one of the mechanisms involved in the generation of cytotoxic edema during the early phase after stroke.¹⁴¹ Retraction of astrocyte end feet from the parenchymal basal lamina of vessels and cellular redistribution and/or degradation of aguaporin-4 at later stroke stages are involved in BBB leakage and formation of vasogenic edema.¹⁴² While coverage of vessels with astrocytic end feet begins before birth, it continues to increase during the first postnatal week.¹³³ However, these particular mechanisms have not yet been specifically characterized after neonatal stroke. Pericytes are another important cell determining the adequate formation and maintenance of the BBB.^{133,143} Pericyte coverage continues to increase during postnatal brain maturation and may affect the

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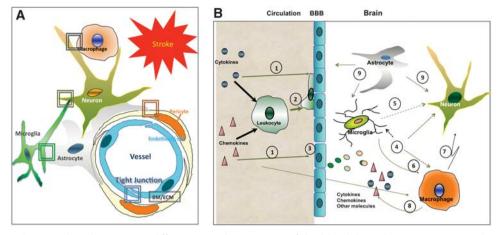


Figure 4. A schematic diagram that shows major differences in the response of the blood-brain barrier to acute stroke between adults and neonates. (A) Under normal conditions the peripheral and local brain compartments are separated by fine-tuned cell-cell interactions at the neurovascular interface. After stroke, direct damage to several cell types, in concert with activation of immune cells and altered cell-cell communications, adversely affect functional integrity of the blood-brain barrier (BBB). Boxes point to known differences in processes in cerebral cortical regions affected by transient middle cerebral artery occlusion (tMCAO) in adult and neonatal rodents. BM/ECM, basal membrane/extracellular matrix. (B) Events at the neurovascular interface after tMCAO in neonatal rats: 1-peripheral cytokines and chemokines activate leukocytes and prime endothelium; 2-extravasation of neutrophils and monocytes is low, contributing to functional preservation of the BBB. Note that in adult rats after tMCAO leukocytes extravasate into ischemic-reperfused brain regions through distorted BBB; 3—BBB leakage is low compared with major BBB leakage and associated entrance of variety of molecules from peripheral circulation entering the injured adult brain; 4-inflammatory mediators that either enter the brain or are produced by local cells activate resident brain cells, including further activation of microglia, macrophages, endothelial cells, and astrocytes; 5-microglial cells signal to neurons; 6-processes shown as 1 to 3 activate microglial cells, prompting morphologic and functional transformation of these cells to brain macrophages. Peripheral monocytes that enter the brain gradually differentiate to become brain macrophages, but activated microglial cells are the predominant population of brain macrophages after acute injury; 7-activated microglia/macrophages directly signal to neurons; 8-microglia/macrophages produce various cytokines and chemokines; and 9-astrocytes directly communicate with neurons and cells of the neurovascular units. 4 to 9-compared with injury in the adult brain, cell-cell communications differ in the newborn in at least several respects, as described in the text.

interactions of these cells with brain vessels in distinct ways after neonatal versus adult stroke that likely differentially affect angiogenesis. Mast cells can be a source of toxicity.¹⁴⁴

As will be discussed later, emerging data on the relative intactness of the BBB after acute neonatal stroke may have broad implications for developing therapies for injured newborns as many classes of drugs may still be precluded from entering the injured brain, at least acutely.

NEURAL REPAIR RESPONSES AFTER NEONATAL STROKE

Compared with the body of knowledge accumulated from adult animal models of stroke, characterization of long-term neural repair responses, including angiogenesis, neurogenesis, oligodendrogenesis, and neural plasticity in experimental models of neonatal stroke is still in its infancy. Transient MCAO gives rise to cell proliferation in the subventricular zone (SVZ) in the adult¹⁴⁵ and neonatal^{146,147} brain. Neuroblasts migrate from the SVZ into the damaged striatum, a region in which neurogenesis does not occur in the intact brain, and begins to express phenotypic region-specific mature neuronal markers,^{148,149} and forms synapses.¹⁵⁰ Gain- and loss-of-function stroke studies in the adult have shown that neuroblast migration occurs in association with remodeling of blood vessels,^{151⁻} via a link between angiogenesis and neurogenesis within the 'neurovascular niche', and that blockage of angiogenesis abolishes neurogenesis after adult stroke. Several recent studies have shown that as opposed to the common belief that the immature brain has greater capacity for endogenous neural repair after injury, repair responses in neonates are more limited and more delayed in time compared with adults.

The low rate of stroke-induced endogenous neural repair observed in neonatal rodents may be related to insufficient induction of angiogenesis after focal ischemia compared with adults. In the adult brain, angiogenesis is a latent process and the

turnover of endothelial cells is very slow, whereas the neonatal brain's vasculature is very dynamic, with the vascular network growing in complexity during the first three postnatal weeks,¹⁵² along with the gradual increase in the brain's increased metabolic activity that results in increasing cerebral blood flow.¹⁵³ Stroke in adult rodents induces a relatively rapid angiogenic response, which starts within 24 hours of injury,^{154,155} while existing data on angiogenesis after neonatal stroke reveal surprisingly delayed induction of angiogenesis after neonatal focal stroke.¹⁵⁶ Vessel density and the number of proliferating endothelial cells are markedly reduced in the ischemic core and even in the ischemic penumbra, and the first signs of increased endothelial proliferation are not observed in the ischemic boundaries until 14 days after tMCAO in P7 rats.¹⁵⁶ Studies utilizing tMCAO in P10 rats also have shown a limited degree of angiogenesis 1 and 2 weeks later^{157,158} despite rapid induction of neuronal and then astrocytic vascular endothelial growth factor (VEGF).¹² Angiogenesis and neurogenesis are enhanced by delayed administration of recombinant human VEGF and disrupted by VEGFR2 inhibition, showing the role of VEGF signaling for the maintenance of angiogenesis and repair in the injured ischemic core.157,158 In this regard, therapies oriented to promote vascular remodeling after neonatal stroke may have a great impact on the long-term progression of brain injury and recovery.

Another factor limiting endogenous repair is the pattern of differentiation of cells that proliferate in the SVZ of the postnatal brain. The formation of GFAP⁺ cells,^{159,160} and not neurons or oligodendrocytes, contributes to astrogliosis and inhibits neural repair in the ischemic boundaries. Brain connectivity depends on white matter, and lack of OLP maturation adversely affects myelination of white-matter tracts^{19,96} and may be the underlying cause for some of the functional deficits in children with neonatal white-matter injury,¹⁹ such as cerebral palsy and long-term cognitive impairment.^{17,161–163} Arrested differentiation and

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delayed cell death of OLPs limit the potentially regenerative effect of increased OLP proliferation observed after H-I.96 While myelination is affected (both ipsilaterally and contralaterally) after neonatal H-I, it is not well understood whether the effect is due to dysregulation of myelination from regulators of myelination, such as Axin2, or altered individual pathways, such as the Wnt pathway, Along with alterations in OLP differentiation, axonal death, malfunction of the subpopulation of astrocytes promoting myelination, and disturbed cell-cell communications can also contribute to myelination defects.¹⁶³ While the effects of growth factors are viewed as purely positive, individual VEGF isoforms, for example, can differentially affect the proliferation and differentiation of SVZ progenitors, since VEGF-A promotes the production of astrocytes from SVZ glial progenitors while VEGF-C stimulates the proliferation of both early and late OLPs, cautioning interpretation of the relative isotype-specific effects on myelination after perinatal brain injury.¹⁶⁴ While these phenomena have not been studied after neonatal stroke, the presence of injury in white-matter structures strongly suggests that these events could be extrapolated to neonatal focal stroke.

TRANSLATIONAL ASPECTS AND TREATMENTS

Therapies available in clinical practice in the perinatal period are very limited and the most commonly used ones such as antiepileptic drugs have side effects that can affect recovery after brain injury. Studies in human neonates have shown that perinatal administration of phenobarbital and phenytoin is associated with reduced brain size and cognitive decline.¹⁶⁵ Consistent with this observation, in rats, a broad range of antiepileptic drugs with various mechanisms of action (GABA_A mimetics or inhibitors of voltage-gated sodium channels) trigger neuronal apoptosis throughout the brain during synaptogenesis^{166–168} but not later in life. Over the past two decades, an array of therapeutic agents has been tested in neonatal ischemic brain injury models to target excitotoxic, oxidative, and inflammatory injury components, but, as in adult stroke, with limited success.

Currently, hypothermia is the only neuroprotective treatment for injury resulting from perinatal global HIE. Recent multicenter clinical trials show that when hypothermia is initiated within hours in neonates with moderate HIE, it can reduce the risk of major neurologic disabilities.^{169,170} A recent analysis of 11 randomized controlled trials revealed that therapeutic hypothermia is beneficial in term newborns with HIE and that cooling reduces mortality without increasing major disability in survivors.¹⁷¹ Selective brain cooling has been shown to potentially induce antiinflammatory effects.¹⁷² The beneficial effects of hypothermia seen in experimental models of H-I, from mice to sheep, may be the result of a wide range of biologic effects, including preservation of energy metabolism and reduction of reactive oxygen species production and neuroinflammation. Hypothermia may also widen the window for repair but the beneficial effects of hypothermia alone seem insufficient, suggesting that outcomes may improve when hypothermia is used in conjunction with other therapies.^{173–177}

Considering that initial observations on a limited endogenous neural repair are not specially promising, as injury can adversely affect postnatal brain development and reprogram the neonatal brain after stroke, more recent studies have begun targeting repair as a way to improve long-term recovery after focal or global H–l injury during the newborn period. Growth factors may prove potent for perinatal stroke and HIE when given alone or as adjunctive therapies. Erythropoietin administration has been shown to enhance neurogenesis and promotes functional recovery after neonatal H–l¹⁷⁸ and MCAO^{179,180} in rodents, with long-lasting effects after multidose erythropoietin treatment. Currently, promising clinical studies are being conducted on the effects of hypothermia and erythropoietin after perinatal HIE¹⁸¹ but much needs to be learned about the optimal conditions for these interventions and alternative means to target the severely injured neonatal brain, especially in the setting of stroke.

As in experimental adult stroke studies, transplantation of stem cells and cell-based therapies, including mesenchymal stem/ progenitor cells, provide promising results in animal models of neonatal brain injury^{182–185} but several aspects of stem-cell therapy including stem-cell origin, number and location of the transplanted cells, and timing of transplantation after injury still need to be refined before considering their use in humans. Another aspect that has yet to be understood is whether engrafted cells themselves enhance the repair by replacement of dead neurons or by stimulating the microenvironment (gene expression of growth factors and inflammatory molecules), which, in turn, permits remodeling and improvement of neurologic function. Some studies have shown that engrafted cells survive for weeks¹⁸² while other studies show that the survival of engrafted cells sharply decline even when the beneficial effects continue.¹⁸⁴

Human umbilical cord blood (HUCB) cells are a rich source of immature stem cells, which have the potential to repair the injured brain. Transplantation of HUCB can induce functional recovery in postischemic neonatal brain in multiple ways, including direct protection, reduced neuroinflammation, production of growth factors, angiogenesis, and neurogenesis, but the exact mechanisms are currently unknown.¹⁸⁶ Some studies suggest that particular subpopulations of HUCB such as endothelial progenitor cells isolated from HUCB may be even more potent and thus preferred. Endothelial progenitor cells, bone marrow-derived precursors are capable of proliferating, circulating, and differentiating into mature endothelial cells. Endothelial progenitor cells have been shown to have a critical role in endothelial repair and vasculogenesis associated with stroke. Their use may have benefits compared with the use of HUCB due to endothelial progenitor cells differentiation into mature endothelial cells and incorporation into vessels or indirectly by secreting factors to stimulate neovascularization.¹⁸⁷ Studies are needed to establish the safety and the temporal and spatial effects before considering clinical trials.

An ongoing preclinical effort to characterize the effects of transplanted cells is supported by MRI to track migration, proliferation, and spatial location and proliferation of neural stem cells implanted after focal stroke in neonatal rats.¹⁸² The ability to monitor cell therapies noninvasively with three-dimensional MRI quantification of ferromagnetically labeled neural stem cells would be important in preclinical and clinical trials using neural stem cells.

CONCLUSIONS AND FUTURE DIRECTIONS

Considerable progress has been made in early stroke diagnosis and MRI-based outcome prognosis in humans and in delineating the complexity of cellular injury, including better understanding of the network of intracellular signaling pathways and the rationale for identification of therapeutic targets and development of effective therapies. Over the past two decades, a major milestone has been the acknowledgment that age at stroke onset is the key factor determining the mechanisms and progression of ischemic brain injury. Recognition of the dynamic nature of changes during brain maturation and the integrative nature of brain injury, which evolves via communication between different cell types and between regions that may reprogram injured immature brain, is another conceptual advance. While much is to be learned about the underlying mechanisms, the knowledge gained is important for advancement to clinical trials. Distinct age-related susceptibility of particular cell populations and mechanisms controlling local inflammation and immune cell infiltration may guide a timedependent selection of therapeutic targets. In view of new knowledge of the relative impermeability of the BBB after acute



neonatal stroke, the distribution and bioavailability of therapies must be carefully considered to ensure that agents that are expected to produce effects locally in the brain do in fact reach the brain.

Short-lived protection of acutely administered therapies, paradoxical responses to antiinflammatory and antioxidant drugs, and the plasticity of the immature brain suggest that one of the critical features to improve outcomes should be a focus on rebuilding the injured neonatal brain by enhancing repair. It is also likely that combinatorial therapies may be superior to single therapies but future research should address the optimal timing for modulating effects of interventions and the role of genetic factors. Among other unanswered questions are why pediatric stroke is recurrent while perinatal stroke is rarely recurrent. Also, the role of prothrombotic pathways after injury is unknown due to lack of neonatal or pediatric thrombotic stroke models.

Elucidating how to enhance the repair process, with a focus on angiogenesis, neurogenesis, and preservation of function will improve the opportunities to enhance recovery and improve outcomes. Emerging evidence also suggests that interventions may be gender specific. Studies are obviously needed to examine the relative risk and benefit of any treatment and to determine whether proposed interventions may adversely affect long-term development.

DISCLOSURE/CONFLICT OF INTEREST

The authors declare no conflict of interest.

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REFERENCES

- 1 Nelson KB. Perinatal ischemic stroke. Stroke 2007; 38: 742-745.
- 2 Lynch JK, Nelson KB. Epidemiology of perinatal stroke. *Curr Opin Pediatr.* 2001; **13**: 499–505.
- 3 Nelson KB, Lynch JK. Stroke in newborn infants. Lancet Neurol 2004; 3: 150-158.
- 4 Raju TN, Nelson KB, Ferriero D, Lynch JK. Ischemic perinatal stroke: summary of a workshop sponsored by the National Institute of Child Health and Human Development and the National Institute of Neurological Disorders and Stroke. *Pediatrics* 2007; **120**: 609–616.
- 5 Lee J, Croen LA, Lindan C, Nash KB, Yoshida CK, Ferriero DM *et al.* Predictors of outcome in perinatal arterial stroke: a population-based study. *Ann Neurol* 2005; 58: 303–308.
- 6 Kirton A, Armstrong-Wells J, Chang T, Deveber G, Rivkin MJ, Hernandez M et al. Symptomatic neonatal arterial ischemic stroke: the International Pediatric Stroke Study. *Pediatrics* 2011; **128**: e1402–e1410.
- 7 Rice JEd, Vannucci RC, Brierley JB. The influence of immaturity on hypoxicischemic brain damage in the rat. *Ann Neurol* 1981; **9**: 131–141.
- 8 Recker R, Adami A, Tone B, Tian HR, Lalas S, Hartman RE *et al.* Rodent neonatal bilateral carotid artery occlusion with hypoxia mimics human hypoxic-ischemic injury. *J Cereb Blood Flow Metab* 2009; **29**: 1305–1316.
- 9 Ashwal S, Cole DJ, Osborne S, Osborne TN, Pearce WJ. A new model of neonatal stroke: reversible middle cerebral artery occlusion in the rat pup. *Pediatr Neurol* 1995; **12**: 191–196.
- 10 Derugin N, Ferriero DM, Vexler ZS. Neonatal reversible focal cerebral ischemia: a new model. *Neurosci Res* 1998; 32: 349–353.
- 11 Renolleau S, Aggoun-Zouaoui D, Ben-Ari Y, Charriaut-Marlangue C. A model of transient unilateral focal ischemia with reperfusion in the P7 neonatal rat: morphological changes indicative of apoptosis. *Stroke* 1998; 29: 1454–1460.
- 12 Mu D, Jiang X, Sheldon RA, Fox CK, Hamrick SE, Vexler ZS *et al.* Regulation of hypoxia-inducible factor 1alpha and induction of vascular endothelial growth factor in a rat neonatal stroke model. *Neurobiol Dis* 2003; **14**: 524–534.
- 13 Woo MS, Wang X, Faustino J, Derugin N, Wendland MF, Zhou P et al. Genetic deletion of CD36 enhances injury after acute neonatal stroke. Ann Neurol 2012; 72: 961–970.

- 14 Back SA, Han BH, Luo NL, Chricton CA, Xanthoudakis S, Tam J et al. Selective vulnerability of late oligodendrocyte progenitors to hypoxia-ischemia. J Neurosci 2002; 22: 455–463.
- 15 Follett PL, Deng W, Dai W, Talos DM, Massillon LJ, Rosenberg PA *et al.* Glutamate receptor-mediated oligodendrocyte toxicity in periventricular leukomalacia: a protective role for topiramate. *J Neurosci* 2004; 24: 4412–4420.
- 16 Back SA, Rivkees SA. Emerging concepts in periventricular white matter injury. Semin Perinatol 2004; 28: 405–414.
- 17 McQuillen PS, Ferriero DM. Selective vulnerability in the developing central nervous system. *Pediatr Neurol* 2004; **30**: 227–235.
- 18 Volpe JJ. Brain injury in premature infants: a complex amalgam of destructive and developmental disturbances. *Lancet Neurol* 2009; 8: 110–124.
- 19 Buser JR, Maire J, Riddle A, Gong X, Nguyen T, Nelson K et al. Arrested preoligodendrocyte maturation contributes to myelination failure in premature infants. Ann Neurol 2012; 71: 93–109.
- 20 Kinney HC, Haynes RL, Xu G, Andiman SE, Folkerth RD, Sleeper LA et al. Neuron deficit in the white matter and subplate in periventricular leukomalacia. Ann Neurol 2012; 71: 397–406.
- 21 Saver JL, Warach S, Janis S, Odenkirchen J, Becker K, Benavente O et al. Standardizing the structure of stroke clinical and epidemiologic research data: the National Institute of Neurological Disorders and Stroke (NINDS) Stroke Common Data Element (CDE) project. Stroke 2012; 43: 967–973.
- 22 Wu YW, Lynch JK, Nelson KB. Perinatal arterial stroke: understanding mechanisms and outcomes. *Semin Neurol* 2005; **25**: 424–434.
- 23 Lee J, Croen LA, Backstrand KH, Yoshida CK, Henning LH, Lindan C *et al*. Maternal and infant characteristics associated with perinatal arterial stroke in the infant. *JAMA* 2005; **293**: 723–729.
- 24 Harteman JC, Groenendaal F, Kwee A, Welsing PM, Benders MJ, de Vries LS. Risk factors for perinatal arterial ischaemic stroke in full-term infants: a case-control study. Arch Dis Child Fetal Neonatal Ed 2012; 97: F411–F416.
- 25 Elbers J, Viero S, MacGregor D, DeVeber G, Moore AM. Placental pathology in neonatal stroke. *Pediatrics* 2011; **127**: e722–e729.
- 26 Kraus FT. Cerebral palsy and thrombi in placental vessels of the fetus: insights from litigation. *Hum Pathol* 1997; **28**: 246–248.
- 27 Kraus FT, Acheen VI. Fetal thrombotic vasculopathy in the placenta: cerebral thrombi and infarcts, coagulopathies, and cerebral palsy. *Hum Pathol* 1999; **30**: 759–769.
- 28 Bremme KA. Haemostatic changes in pregnancy. *Best Pract Res Clin Haematol* 2003; **16**: 153–168.
- 29 Heier LA, Carpanzano CR, Mast J, Brill PW, Winchester P, Deck MD. Maternal cocaine abuse: the spectrum of radiologic abnormalities in the neonatal CNS. *AJNR Am J Neuroradiol* 1991; **12**: 951–956.
- 30 Gunther G, Junker R, Strater R, Schobess R, Kurnik K, Heller C et al. Symptomatic ischemic stroke in full-term neonates: role of acquired and genetic prothrombotic risk factors. Stroke 2000; 31: 2437–2441.
- 31 Silver RK, MacGregor SN, Pasternak JF, Neely SE.. Fetal stroke associated with elevated maternal anticardiolipin antibodies. *Obstet Gynecol* 1992; **80**: 497–499.
- 32 Curry CJ, Bhullar S, Holmes J, Delozier CD, Roeder ER, Hutchison HT. Risk factors for perinatal arterial stroke: a study of 60 mother-child pairs. *Pediatr Neurol* 2007; 37: 99–107.
- 33 Nelson KB. Thrombophilias, perinatal stroke, and cerebral palsy. *Clin Obstet Gynecol* 2006; **49**: 875–884.
- 34 Lynch JK, Han CJ, Nee LE, Nelson KB. Prothrombotic factors in children with stroke or porencephaly. *Pediatrics* 2005; **116**: 447–453.
- 35 Gould DB, Phalan FC, Breedveld GJ, van Mil SE, Smith RS, Schimenti JC *et al.* Mutations in Col4a1 cause perinatal cerebral hemorrhage and porencephaly. *Science* 2005; **308**: 1167–1171.
- 36 Labelle-Dumais C, Dilworth DJ, Harrington EP, de Leau M, Lyons D, Kabaeva Z et al. COL4A1 mutations cause ocular dysgenesis, neuronal localization defects, and myopathy in mice and Walker-Warburg syndrome in humans. *PLoS Genet* 2011; **7**: e1002062.
- 37 McQuillen PS, Miller SP. Congenital heart disease and brain development. Ann NY Acad Sci 2010; **1184**: 68–86.
- 38 Govaert P, Smith L, Dudink J. Diagnostic management of neonatal stroke. *Semin Fetal Neonatal Med* 2009; **14**: 323–328.
- 39 Roach ES, Golomb MR, Adams R, Biller J, Daniels S, Deveber G *et al*. Management of stroke in infants and children: a scientific statement from a Special Writing Group of the American Heart Association Stroke Council and the Council on Cardiovascular Disease in the Young. *Stroke* 2008; **39**: 2644–2691.
- 40 Kurnik K, Kosch A, Strater R, Schobess R, Heller C, Nowak-Gottl U. Recurrent thromboembolism in infants and children suffering from symptomatic neonatal arterial stroke: a prospective follow-up study. *Stroke* 2003; **34**: 2887–2892.
- 41 Eyre JA, Smith M, Dabydeen L, Clowry GJ, Petacchi E, Battini R et al. Is hemiplegic cerebral palsy equivalent to amblyopia of the corticospinal system? Ann Neurol 2007; 62: 493–503.

- 930
- 42 de Vries LS, Groenendaal F, Eken P, van Haastert IC, Rademaker KJ, Meiners LC. Infarcts in the vascular distribution of the middle cerebral artery in preterm and fullterm infants. *Neuropediatrics* 1997; 28: 88–96.
- 43 Govaert P, Matthys E, Zecic A, Roelens F, Oostra A, Vanzieleghem B. Perinatal cortical infarction within middle cerebral artery trunks. Arch Dis Child Fetal Neonatal Ed 2000; 82: F59–F63.
- 44 Golomb MR, MacGregor DL, Domi T, Armstrong DC, McCrindle BW, Mayank S et al. Presumed pre- or perinatal arterial ischemic stroke: risk factors and outcomes. Ann Neurol 2001; 50: 163–168.
- 45 Mercuri E, Rutherford M, Cowan F, Pennock J, Counsell S, Papadimitriou M et al. Early prognostic indicators of outcome in infants with neonatal cerebral infarction: a clinical, electroencephalogram, and magnetic resonance imaging study. *Pediatrics* 1999; **103**: 39–46.
- 46 Wu YW, March WM, Croen LA, Grether JK, Escobar GJ, Newman TB. Perinatal stroke in children with motor impairment: a population-based study. *Pediatrics* 2004; **114**: 612–619.
- 47 Golomb MR, Garg BP, Carvalho KS, Johnson CS, Williams LS. Perinatal stroke and the risk of developing childhood epilepsy. *J Pediatr* 2007; **151**: 409–413413 e401-402.
- 48 Wanigasinghe J, Reid SM, Mackay MT, Reddihough DS, Harvey AS, Freeman JL. Epilepsy in hemiplegic cerebral palsy due to perinatal arterial ischaemic stroke. *Dev Med Child Neurol* 2010; **52**: 1021–1027.
- 49 Wusthoff CJ, Kessler SK, Vossough A, Ichord R, Zelonis S, Halperin A *et al*. Risk of later seizure after perinatal arterial ischemic stroke: a prospective cohort study. *Pediatrics* 2011; **127**: e1550–e1557.
- 50 Golomb MR, Garg BP, Williams LS. Outcomes of children with infantile spasms after perinatal stroke. *Pediatr Neurol* 2006; **34**: 291–295.
- 51 Osborne JP, Lux AL, Edwards SW, Hancock E, Johnson AL, Kennedy CR et al. The underlying etiology of infantile spasms (West syndrome): information from the United Kingdom Infantile Spasms Study (UKISS) on contemporary causes and their classification. *Epilepsia* 2010; **51**: 2168–2174.
- 52 Sreenan C, Bhargava R, Robertson CM. Cerebral infarction in the term newborn: clinical presentation and long-term outcome. J Pediatr 2000; 137: 351–355.
- 53 Yi YY, Lee JS, Jang SI, Song JS, Yang S, Kim SK et al. Clinical outcomes of cerebral infarctions in neonates. *Pediatr Neurol* 2011; **45**: 368–372.
- 54 Mercuri E, Barnett A, Rutherford M, Guzzetta A, Haataja L, Cioni G et al. Neonatal cerebral infarction and neuromotor outcome at school age. *Pediatrics* 2004; 113: 95–100.
- 55 Mackay MT, Wiznitzer M, Benedict SL, Lee KJ, Deveber GA, Ganesan V. Arterial ischemic stroke risk factors: the International Pediatric Stroke Study. Ann Neurol 2011; 69: 130–140.
- 56 Ashwal S, Tone B, Tian HR, Chong S, Obenaus A. Comparison of two neonatal ischemic injury models using magnetic resonance imaging. *Pediatr Res* 2007; 61: 9–14.
- 57 Partridge SC, Mukherjee P, Berman JI, Henry RG, Miller SP, Lu Y et al. Tractography-based quantitation of diffusion tensor imaging parameters in white matter tracts of preterm newborns. J Magn Reson Imaging 2005; 22: 467–474.
- 58 Limperopoulos C. Advanced neuroimaging techniques: their role in the development of future fetal and neonatal neuroprotection. *Semin Perinatol* 2010; 34: 93–101.
- 59 Miller SP, Ferriero DM. From selective vulnerability to connectivity: insights from newborn brain imaging. *Trends Neurosci* 2009; **32**: 496–505.
- 60 Tymofiyeva O, Hess CP, Ziv E, Tian N, Bonifacio SL, McQuillen PS *et al.* Towards the 'baby connectome': mapping the structural connectivity of the newborn brain. *PLoS ONE* 2012; **7**: e31029.
- 61 Fullerton HJ, Wu YW, Zhao S, Johnston SC. Risk of stroke in children: ethnic and gender disparities. *Neurology* 2003; 61: 189–194.
- 62 Abend NS, Beslow LA, Smith SE, Kessler SK, Vossough A, Mason S et al. Seizures as a presenting symptom of acute arterial ischemic stroke in childhood. J Pediatr 2011; 159: 479–483.
- 63 Singh RK, Zecavati N, Singh J, Kaulas H, Nelson KB, Dean NP *et al.* Seizures in acute childhood stroke. *J Pediatr* 2012; **160**: 291–296.
- 64 Chadehumbe MA, Khatri P, Khoury JC, Alwell K, Szaflarski JP, Broderick JP *et al.* Seizures are common in the acute setting of childhood stroke: a populationbased study. *J Child Neurol* 2009; **24**: 9–12.
- 65 Szaflarski JP, Rackley AY, Kleindorfer DO, Khoury J, Woo D, Miller R *et al.* Incidence of seizures in the acute phase of stroke: a population-based study. *Epilepsia* 2008; **49**: 974–981.
- 66 Fullerton HJ, Wu YW, Sidney S, Johnston SC. Risk of recurrent childhood arterial ischemic stroke in a population-based cohort: the importance of cerebrovascular imaging. *Pediatrics* 2007; **119**: 495–501.
- 67 Amlie-Lefond C, Bernard TJ, Sebire G, Friedman NR, Heyer GL, Lerner NB et al. Predictors of cerebral arteriopathy in children with arterial ischemic stroke: results of the International Pediatric Stroke Study. *Circulation* 2009; **119**: 1417–1423.

- 68 Fullerton HJ, Elkind MS, Barkovich AJ, Glaser C, Glidden D, Hills NK *et al.* The vascular effects of infection in Pediatric Stroke (VIPS) Study. *J Child Neurol* 2011; 26: 1101–1110.
- 69 Westmacott R, Askalan R, MacGregor D, Anderson P, Deveber G. Cognitive outcome following unilateral arterial ischaemic stroke in childhood: effects of age at stroke and lesion location. *Dev Med Child Neurol* 2010; **52**: 386–393.
- 70 Boardman JP, Ganesan V, Rutherford MA, Saunders DE, Mercuri E, Cowan F. Magnetic resonance image correlates of hemiparesis after neonatal and childhood middle cerebral artery stroke. *Pediatrics* 2005; **115**: 321–326.
- 71 Maegaki Y, Maeoka Y, Ishii S, Shiota M, Takeuchi A, Yoshino K et al. Mechanisms of central motor reorganization in pediatric hemiplegic patients. *Neuropediatrics* 1997; 28: 168–174.
- 72 Yager JY, Ashwal S. Animal models of perinatal hypoxic-ischemic brain damage. Pediatr Neurol 2009; 40: 156–167.
- 73 Ashwal S, Tone B, Tian HR, Chong S, Obenaus A. Serial magnetic resonance imaging in a rat pup filament stroke model. *Exp Neurol* 2006; **202**: 294–301.
- 74 Manabat C, Han BH, Wendland M, Derugin N, Fox CK, Choi J et al. Reperfusion differentially induces caspase-3 activation in ischemic core and penumbra after stroke in immature brain. Stroke 2003; 34: 207–213.
- 75 Dingman A, Lee SY, Derugin N, Wendland MF, Vexler ZS. Aminoguanidine inhibits caspase-3 and calpain activation without affecting microglial activation following neonatal transient ischemia. J Neurochem 2006; 96: 1467–1479.
- 76 Derugin N, Wendland M, Muramatsu K, Roberts T, Gregory G, Ferriero D *et al.* Evolution of brain injury after transient middle cerebral artery occlusion in neonatal rat. *Stroke* 2000; **31**: 1752–1761.
- 77 Fernandez Lopez D, Faustino J, Daneman R, Zhou L, Lee SY, Derugin N et al. Blood-brain barrier permeability is increased after acute adult stroke but not neonatal stroke. J Neurosci 2012; 32: 9588–9600.
- 78 Wang R, Ashwal S, Tone B, Tian HR, Badaut J, Rasmussen A et al. Albumin reduces blood-brain barrier permeability but does not alter infarct size in a rat model of neonatal stroke. *Pediatr Res* 2007; 62: 261–266.
- 79 Ghosh N, Yuan X, Turenius CI, Tone B, Ambadipudi K, Snyder EY et al. Automated core-penumbra quantification in neonatal ischemic brain injury. J Cereb Blood Flow Metab 2012; 32: 2161–2170.
- 80 Derugin N, Dingman A, Wendland M, Fox C, Vexler ZS. Magnetic resonance imaging as a surrogate measure for histological sub-chronic endpoint in a neonatal rat stroke model. *Brain Res* 2005; **1066**: 49–56.
- 81 Stone BS, Zhang J, Mack DW, Mori S, Martin LJ, Northington FJ. Delayed neural network degeneration after neonatal hypoxia-ischemia. *Ann Neurol* 2008; 64: 535–546.
- 82 McLean C, Ferriero D. Mechanisms of hypoxic-ischemic injury in the term infant. Semin Perinatol 2004; 28: 425–432.
- 83 Jensen FE. The role of glutamate receptor maturation in perinatal seizures and brain injury. *Int J Dev Neurosci* 2002; **20**: 339–347.
- 84 Johnston MV. Excitotoxicity in perinatal brain injury. *Brain Pathol.* 2005; **15**: 234–240.
- 85 Knox R, Zhao C, Miguel-Perez D, Wang S, Yuan J, Ferriero D et al. Enhanced NMDA receptor tyrosine phosphorylation and increased brain injury following neonatal hypoxia-ischemia in mice with neuronal Fyn overexpression. *Neurobiol Dis* 2012; **51**: 113–119.
- 86 Jiang X, Mu D, Sheldon RA, Glidden DV, Ferriero DM. Neonatal hypoxia-ischemia differentially upregulates MAGUKs and associated proteins in PSD-93-deficient mouse brain. *Stroke* 2003; **34**: 2958–2963.
- 87 Lafemina MJ, Sheldon RA, Ferriero DM. Acute hypoxia-ischemia results in hydrogen peroxide accumulation in neonatal but not adult mouse brain. *Pediatr Res* 2006; 59: 680–683.
- 88 Sheldon RA, Aminoff A, Lee CL, Christen S, Ferriero DM. Hypoxic preconditioning reverses protection after neonatal hypoxia-ischemia in glutathione peroxidase transgenic murine brain. *Pediatr Res* 2007; 61: 666–670.
- 89 Sheldon RA, Jiang X, Francisco C, Christen S, Vexler ZS, Tauber MG et al. Manipulation of antioxidant pathways in neonatal murine brain. *Pediatr Res* 2004; 56: 656–662.
- 90 Faustino J, Wang X, Jonhson C, Klibanov A, Derugin N, Wendland M et al. Microglial cells contribute to endogenous brain defenses after acute neonatal focal stroke. J Neurosci 2011; 31: 12992–13001.
- 91 Vexler ZS, Yenari MA. Does inflammation after stroke affect the developing brain differently than adult brain? *Dev Neurosci* 2009; **31**: 378–393.
- 92 Han BH, Xu D, Choi J, Han Y, Xanthoudakis S, Roy S et al. Selective, reversible caspase-3 inhibitor is neuroprotective and reveals distinct pathways of cell death following neonatal hypoxic- ischemic brain injury. J Biol Chem 2002; 277: 30128– 30136.
- 93 Hu BR, Liu CL, Ouyang Y, Blomgren K, Siesjo BK. Involvement of caspase-3 in cell death after hypoxia-ischemia declines during brain maturation. J Cereb Blood Flow Metab 2000; 20: 1294–1300.

- 94 Northington FJ, Chavez-Valdez R, Martin LJ. Neuronal cell death in neonatal hypoxia-ischemia. Ann Neurol 2011; 69: 743–758.
- 95 West T, Atzeva M, Holtzman DM. Caspase-3 deficiency during development increases vulnerability to hypoxic-ischemic injury through caspase-3-independent pathways. *Neurobiol Dis* 2006; 22: 523–537.
- 96 Segovia KN, McClure M, Moravec M, Luo NL, Wan Y, Gong X *et al.* Arrested oligodendrocyte lineage maturation in chronic perinatal white matter injury. *Ann Neurol* 2008; **63**: 520–530.
- 97 Zhu C, Xu F, Fukuda A, Wang X, Fukuda H, Korhonen L et al. X chromosomelinked inhibitor of apoptosis protein reduces oxidative stress after cerebral irradiation or hypoxia-ischemia through up-regulation of mitochondrial antioxidants. Eur J Neurosci 2007; 26: 3402–3410.
- 98 Matsumori Y, Hong SM, Aoyama K, Fan Y, Kayama T, Sheldon RA et al. Hsp70 overexpression sequesters AIF and reduces neonatal hypoxic/ischemic brain injury. J Cereb Blood Flow Metab 2005; 25: 899–910.
- 99 Cheng Y, Deshmukh M, D'Costa A, Demaro JA, Gidday JM, Shah A et al. Caspase inhibitor affords neuroprotection with delayed administration in a rat model of neonatal hypoxic-ischemic brain injury [see comments]. J Clin Invest 1998; 101: 1992–1999.
- 100 Gill R, Soriano M, Blomgren K, Hagberg H, Wybrecht R, Miss MT et al. Role of caspase-3 activation in cerebral ischemia-induced neurodegeneration in adult and neonatal brain. J Cereb Blood Flow Metab 2002; 22: 420–430.
- 101 Joly LM, Mucignat V, Mariani J, Plotkine M, Charriaut-Marlangue C. Caspase inhibition after neonatal ischemia in the rat brain. J Cereb Blood Flow Metab 2004; 24: 124–131.
- 102 Puyal J, Vaslin A, Mottier V, Clarke PG. Postischemic treatment of neonatal cerebral ischemia should target autophagy. Ann Neurol 2009; 66: 378–389.
- 103 Blomgren K, Leist M, Groc L. Pathological apoptosis in the developing brain. *Apoptosis* 2007; **12**: 993–1010.
- 104 Johnston MV, Hagberg H. Sex and the pathogenesis of cerebral palsy. *Dev Med Child Neurol* 2007; **49**: 74–78.
- 105 Nunez J. Sex and steroid hormones in early brain injury. *Rev Endocr Metab Disord* 2012; **13**: 173–186.
- 106 Hagberg H, Wilson MA, Matsushita H, Zhu C, Lange M, Gustavsson M et al. PARP-1 gene disruption in mice preferentially protects males from perinatal brain injury. J Neurochem 2004; 90: 1068–1075.
- Renolleau S, Fau S, Charriaut-Marlangue C. Gender-related differences in apoptotic pathways after neonatal cerebral ischemia. *Neuroscientist* 2008; 14: 46–52.
- 108 Renolleau S, Fau S, Goyenvalle C, Charriaut-Marlangue C. 'Sex, neuroprotection, and neonatal ischemia'. *Dev Med Child Neurol* 2007; **49**: 477.
- 109 Renolleau S, Fau S, Goyenvalle C, Joly LM, Chauvier D, Jacotot E et al. Specific caspase inhibitor Q-VD-OPh prevents neonatal stroke in P7 rat: a role for gender. J Neurochem 2007; 100: 1062–1071.
- 110 Nijboer CH, Groenendaal F, Kavelaars A, Hagberg HH, van Bel F, Heijnen CJ. Gender-specific neuroprotection by 2-iminobiotin after hypoxia-ischemia in the neonatal rat via a nitric oxide independent pathway. J Cereb Blood Flow Metab 2007; 27: 282–292.
- 111 Herson PS, Bombardier CG, Parker SM, Shimizu T, Klawitter J, Quillinan N et al. Experimental pediatric arterial ischemic stroke model reveals sex-specific estrogen signaling. Stroke 2013; 44: 759–763.
- 112 Normann S, de Veber G, Fobker M, Langer C, Kenet G, Bernard TJ et al. Role of endogenous testosterone concentration in pediatric stroke. Ann Neurol 2009; 66: 754–758.
- 113 Vannucci SJ, Hurn PD. Gender differences in pediatric stroke: is elevated testosterone a risk factor for boys? *Ann Neurol* 2009; **66**: 713–714.
- 114 Titomanlio L, Kavelaars A, Dalous J, Mani S, El Ghouzzi V, Heijnen C *et al.* Stem cell therapy for neonatal brain injury: perspectives and challenges. *Ann Neurol* 2011; **70**: 698–712.
- 115 Winerdal M, Winerdal ME, Kinn J, Urmaliya V, Winqvist O, Aden U. Long lasting local and systemic inflammation after cerebral hypoxic ischemia in newborn mice. *PLoS ONE* 2012; 7: e36422.
- 116 Lassiter HA, Walz BM, Wilson JL, Jung E, Calisi CR, Goldsmith ⊔ *et al.* The administration of complement component C9 enhances the survival of neonatal rats with Escherichia coli sepsis. *Pediatr Res* 1997; **42**: 128–136.
- 117 Lassiter HA, Watson SW, Seifring ML, Tanner JE. Complement factor 9 deficiency in serum of human neonates. J Infect Dis 1992; **166**: 53–57.
- 118 Doverhag C, Keller M, Karlsson A, Hedtjarn M, Nilsson U, Kapeller E et al. Pharmacological and genetic inhibition of NADPH oxidase does not reduce brain damage in different models of perinatal brain injury in newborn mice. *Neurobiol Dis* 2008; **31**: 133–144.
- 119 Hedtjarn M, Mallard C, Iwakura Y, Hagberg H. Combined deficiency of IL-1beta18, but not IL-1alphabeta, reduces susceptibility to hypoxia-ischemia in the immature brain. *Dev Neurosci* 2005; **27**: 143–148.
- 120 Nijboer CH, Heijnen CJ, Groenendaal F, May MJ, van Bel F, Kavelaars A. A dual role of the NF-kappaB pathway in neonatal hypoxic-ischemic brain damage. *Stroke* 2008; **39**: 2578–2586.

- 121 Du X, Fleiss B, Li H, D'Angelo B, Sun Y, Zhu C *et al.* Systemic stimulation of TLR2 impairs neonatal mouse brain development. *PLoS ONE* 2011; **6**: e19583.
- 122 Wang Q, Tang XN, Yenari MA. The inflammatory response in stroke. J Neuroimmunol. 2007; **184**: 53–68.
- 123 Zhao BQ, Wang S, Kim HY, Storrie H, Rosen BR, Mooney DJ et al. Role of matrix metalloproteinases in delayed cortical responses after stroke. Nat Med 2006; 12: 441–445.
- 124 Denker S, Ji S, Lee SY, Dingman A, Derugin N, Wendland M et al. Macrophages are comprised of resident brain microglia not infiltrating peripheral monocytes acutely after neonatal stroke. J Neurochem 2007; 100: 893–904.
- 125 Cho S, Park EM, Febbraio M, Anrather J, Park L, Racchumi G *et al.* The class B scavenger receptor CD36 mediates free radical production and tissue injury in cerebral ischemia. *J Neurosci* 2005; **25**: 2504–2512.
- 126 Nimmerjahn A, Kirchhoff F, Helmchen F. Resting microglial cells are highly dynamic surveillants of brain parenchyma *in vivo. Science* 2005; **308**: 1314–1318.
- 127 Davalos D, Ryu JK, Merlini M, Baeten KM, Le Moan N, Petersen MA et al. Fibrinogen-induced perivascular microglial clustering is required for the development of axonal damage in neuroinflammation. Nat Commun 2012; 3: 1227.
- 128 Fantin A, Vieira JM, Gestri G, Denti L, Schwarz Q, Prykhozhij S *et al.* Tissue macrophages act as cellular chaperones for vascular anastomosis downstream of VEGF-mediated endothelial tip cell induction. *Blood* 2010; **116**: 829–840.
- 129 Dammann O, Leviton A. Perinatal brain damage causation. Dev Neurosci 2007; 29: 280–288.
- 130 Bartha AI, Foster-Barber A, Miller SP, Vigneron DB, Glidden DV, Barkovich AJ et al. Neonatal encephalopathy: association of cytokines with MR spectroscopy and outcome. *Pediatr Res* 2004; 56: 960–966.
- 131 Saunders NR, Habgood MD, Dziegielewska KM. Barrier mechanisms in the brain, II. Immature brain. *Clin Exp Pharmacol Physiol* 1999; 26: 85–91.
- 132 Engelhardt B. Development of the blood-brain barrier. *Cell Tissue Res* 2003; **314**: 119–129.
- 133 Daneman R, Zhou L, Kebede AA, Barres BA. Pericytes are required for bloodbrain barrier integrity during embryogenesis. *Nature* 2010; 468: 562–566.
- 134 Anthony DC, Bolton SJ, Fearn S, Perry VH. Age-related effects of interleukin-1 beta on polymorphonuclear neutrophil-dependent increases in blood-brain barrier permeability in rats. *Brain* 1997; **120**(Pt 3): 435–444.
- 135 Dzietko M, Faustino J, Derugin N, Wendland MF, Ferriero DM, Vexler ZS. MRI as a translational tool for the study of neonatal stroke. *J Child Neurol* 2011; 26: 1145–1153.
- 136 Benjelloun N, Renolleau S, Represa A, Ben-Ari Y, Charriaut-Marlangue C. Inflammatory responses in the cerebral cortex after ischemia in the P7 neonatal Rat. Stroke 1999; 30: 1916–1923, discussion 1923-1914.
- 137 Garcia JH, Liu KF, Yoshida Y, Lian J, Chen S, del Zoppo GJ. Influx of leukocytes and platelets in an evolving brain infarct (Wistar rat). Am J Pathol 1994; 144: 188–199.
- 138 Matsuo Y, Kihara T, Ikeda M, Ninomiya M, Onodera H, Kogure K. Role of neutrophils in radical production during ischemia and reperfusion of the rat brain: effect of neutrophil depletion on extracellular ascorbyl radical formation. J Cereb Blood Flow Metab 1995; 15: 941–947.
- 139 Hudome S, Palmer C, Roberts RL, Mauger D, Housman C, Towfighi J. The role of neutrophils in the production of hypoxic-ischemic brain injury in the neonatal rat. *Pediatr Res* 1997; **41**: 607–616.
- 140 Bona E, Andersson AL, Blomgren K, Gilland E, Puka-Sundvall M, Gustafson K et al. Chemokine and inflammatory cell response to hypoxia-ischemia in immature rats. Pediatr Res 1999; 45: 500–509.
- 141 Zador Z, Stiver S, Wang V, Manley GT. Role of aquaporin-4 in cerebral edema and stroke. *Handb Exp Pharmacol* 2009; **190**: 159–170.
- 142 Papadopoulos MC, Manley GT, Krishna S, Verkman AS. Aquaporin-4 facilitates reabsorption of excess fluid in vasogenic brain edema. FASEB J. 2004; 18: 1291–1293.
- 143 Armulik A, Genove G, Mae M, Nisancioglu MH, Wallgard E, Niaudet C *et al.* Pericytes regulate the blood-brain barrier. *Nature* 2010; **468**: 557–561.
- 144 Jin Y, Silverman AJ, Vannucci SJ. Mast cells are early responders after hypoxiaischemia in immature rat brain. *Stroke* 2009; **40**: 3107–3112.
- 145 Carmichael ST. Cellular and molecular mechanisms of neural repair after stroke: making waves. *Ann Neurol* 2006; **59**: 735–742.
- 146 Plane JM, Liu R, Wang TW, Silverstein FS, Parent JM. Neonatal hypoxic-ischemic injury increases forebrain subventricular zone neurogenesis in the mouse. *Neurobiol Dis* 2004; 16: 585–595.
- 147 Yang Z, Covey MV, Bitel CL, Ni L, Jonakait GM, Levison SW. Sustained neocortical neurogenesis after neonatal hypoxic/ischemic injury. *Ann Neurol* 2007; 61: 199–208.
- 148 Arvidsson A, Collin T, Kirik D, Kokaia Z, Lindvall O. Neuronal replacement from endogenous precursors in the adult brain after stroke. *Nat Med* 2002; 8: 963–970.

- 149 Parent JM, Vexler ZS, Gong C, Derugin N, Ferriero DM. Rat forebrain neurogenesis and striatal neuron replacement after focal stroke. Ann Neurol 2002; 52: 802–813.
- 150 Li S, Overman JJ, Katsman D, Kozlov SV, Donnelly CJ, Twiss JL et al. An agerelated sprouting transcriptome provides molecular control of axonal sprouting after stroke. Nat Neurosci 2010; 13: 1496–1504.
- 151 Ohab JJ, Fleming S, Blesch A, Carmichael ST. A neurovascular niche for neurogenesis after stroke. J Neurosci 2006; 26: 13007–13016.
- 152 Ogunshola OO, Stewart WB, Mihalcik V, Solli T, Madri JA, Ment LR. Neuronal VEGF expression correlates with angiogenesis in postnatal developing rat brain. *Brain Res Dev Brain Res* 2000; **119**: 139–153.
- 153 Tuor Ul, Grewal D. Autoregulation of cerebral blood flow: influence of local brain development and postnatal age. *Am J Physiol* 1994; **267**: H2220–H2228.
- 154 Hayashi T, Noshita N, Sugawara T, Chan PH. Temporal profile of angiogenesis and expression of related genes in the brain after ischemia. J Cereb Blood Flow Metab 2003; 23: 166–180.
- 155 Marti HJ, Bernaudin M, Bellail A, Schoch H, Euler M, Petit E *et al.* Hypoxia-induced vascular endothelial growth factor expression precedes neovascularization after cerebral ischemia. *Am J Pathol* 2000; **156**: 965–976.
- 156 Fernandez-Lopez D, Faustino J, Derugin N, Vexler ZS. Acute and chronic vascular responses to experimental focal arterial stroke in the neonate rat. *Transl Stroke Res* 2013; **4**: 179–188.
- 157 Shimotake J, Derugin N, Wendland M, Vexler ZS, Ferriero DM. Vascular endothelial growth factor receptor-2 inhibition promotes cell death and limits endothelial cell proliferation in a neonatal rodent model of stroke. *Stroke* 2010; 41: 343–349.
- 158 Dzietko M, Derugin N, Wendland MF, Vexler ZS, Ferriero DM. Delayed VEGF treatment enhances angiogenesis and recovery after neonatal focal rodent stroke. *Transl Stroke Res* 2013; **4**: 189–200.
- 159 Spadafora R, Gonzalez FF, Derugin N, Wendland M, Ferriero D, McQuillen P. Altered fate of subventricular zone progenitor cells and reduced neurogenesis following neonatal stroke. *Dev Neurosci* 2010; **32**: 101–113.
- 160 Gonzalez FF, Larpthaveesarp A, McQuillen P, Derugin N, Wendland M, Spadafora R et al. Erythropoietin increases neurogenesis and oligodendrogliosis of subventricular zone precursor cells after neonatal stroke. Stroke 2013; 44: 753–758.
- 161 Fancy SP, Chan JR, Baranzini SE, Franklin RJ, Rowitch DH. Myelin regeneration: a recapitulation of development? Annu Rev Neurosci 2011; 34: 21–43.
- 162 Back SA, Gan X, Li Y, Rosenberg PA, Volpe JJ. Maturation-dependent vulnerability of oligodendrocytes to oxidative stress-induced death caused by glutathione depletion. J Neurosci 1998; 18: 6241–6253.
- 163 Fancy SP, Harrington EP, Yuen TJ, Silbereis JC, Zhao C, Baranzini SE et al. Axin2 as regulatory and therapeutic target in newborn brain injury and remyelination. *Nat Neurosci* 2011; 14: 1009–1016.
- 164 Bain JM, Moore L, Ren Z, Simonishvili S, Levison SW. Vascular endothelial growth factors A and C are induced in the SVZ following neonatal hypoxia-ischemia and exert different effects on neonatal glial progenitors. *Transl Stroke Res* 2013; 4: 158–170.
- 165 Dessens AB, Cohen-Kettenis PT, Mellenbergh GJ, Koppe JG, van De Poll NE, Boer K. Association of prenatal phenobarbital and phenytoin exposure with small head size at birth and with learning problems. *Acta Paediatr* 2000; 89: 533–541.
- 166 Bittigau P, Sifringer M, Genz K, Reith E, Pospischil D, Govindarajalu S et al. Antiepileptic drugs and apoptotic neurodegeneration in the developing brain. Proc Natl Acad Sci USA 2002; 99: 15089–15094.
- 167 Olney JW, Wozniak DF, Jevtovic-Todorovic V, Farber NB, Bittigau P, Ikonomidou C. Drug-induced apoptotic neurodegeneration in the developing brain. *Brain Pathol* 2002; **12**: 488–498.

- 168 Olney JW, Young C, Wozniak DF, Ikonomidou C, Jevtovic-Todorovic V. Anesthesia-induced developmental neuroapoptosis. Does it happen in humans? *Anesthesiology* 2004; **101**: 273–275.
- 169 Azzopardi DV, Strohm B, Edwards AD, Dyet L, Halliday HL, Juszczak E et al. Moderate hypothermia to treat perinatal asphyxial encephalopathy. N Engl J Med 2009; 361: 1349–1358.
- 170 Edwards AD, Brocklehurst P, Gunn AJ, Halliday H, Juszczak E, Levene M et al. Neurological outcomes at 18 months of age after moderate hypothermia for perinatal hypoxic ischaemic encephalopathy: synthesis and meta-analysis of trial data. BMJ 2010; 340: c363.
- 171 Jacobs SE, Berg M, Hunt R, Tarnow-Mordi WO, Inder TE, Davis PG. Cooling for newborns with hypoxic ischaemic encephalopathy. *Cochrane Database Syst Rev* 2013; 1: CD003311.
- 172 Perrone S, Szabo M, Bellieni CV, Longini M, Bango M, Kelen D *et al*. Whole body hypothermia and oxidative stress in babies with hypoxic-ischemic brain injury. *Pediatr Neurol* 2010; **43**: 236–240.
- 173 Barks JD, Liu YQ, Shangguan Y, Silverstein FS. Phenobarbital augments hypothermic neuroprotection. *Pediatr Res.* 2010; **67**: 532–537.
- 174 Carlsson Y, Wang X, Schwendimann L, Rousset CI, Jacotot E, Gressens P *et al.* Combined effect of hypothermia and caspase-2 gene deficiency on neonatal hypoxic-ischemic brain injury. *Pediatr Res* 2012; **71**: 566–572.
- 175 Fang AY, Gonzalez FF, Sheldon RA, Ferriero DM. Effects of combination therapy using hypothermia and erythropoietin in a rat model of neonatal hypoxiaischemia. *Pediatr Res* 2013; **73**: 12–17.
- 176 Hobbs CE, Oorschot DE. Neonatal rat hypoxia-ischemia: long-term rescue of striatal neurons and motor skills by combined antioxidant-hypothermia treatment. *Brain Pathol* 2008; **18**: 443–454.
- 177 Liu Y, Barks JD, Xu G, Silverstein FS. Topiramate extends the therapeutic window for hypothermia-mediated neuroprotection after stroke in neonatal rats. *Stroke* 2004; **35**: 1460–1465.
- 178 Iwai M, Stetler RA, Xing J, Hu X, Gao Y, Zhang W et al. Enhanced oligodendrogenesis and recovery of neurological function by erythropoietin after neonatal hypoxic/ischemic brain injury. Stroke 2010; 41: 1032–1037.
- 179 Gonzalez F, McQuillen PS, Mu D, Chang YS, Wendland M, Vexler ZS et al. Erythropoietin enhances long-term neuroprotection and neurogenesis in neonatal stroke. Dev Neurosci 2007; 29: 321–330.
- 180 Gonzalez FF, Abel R, Almli CR, Mu D, Wendland M, Ferriero DM. Erythropoietin sustains cognitive function and brain volume after neonatal stroke. *Dev Neurosci* 2009; **31**: 403–411.
- 181 Wu YW, Bauer LA, Ballard RA, Ferriero DM, Glidden DV, Mayock DE et al. Erythropoietin for neuroprotection in neonatal encephalopathy: safety and pharmacokinetics. *Pediatrics* 2012; **130**: 683–691.
- 182 Ashwal S, Obenaus A, Snyder EY. Neuroimaging as a basis for rational stem cell therapy. *Pediatr Neurol* 2009; **40**: 227–236.
- 183 Daadi MM, Davis AS, Arac A, Li Z, Maag AL, Bhatnagar R et al. Human neural stem cell grafts modify microglial response and enhance axonal sprouting in neonatal hypoxic-ischemic brain injury. Stroke 2010; 41: 516–523.
- 184 van Velthoven CT, Kavelaars A, van Bel F, Heijnen CJ. Mesenchymal stem cell transplantation changes the gene expression profile of the neonatal ischemic brain. Brain Behav Immun 2011; 25: 1342–1348.
- 185 van Velthoven CT, Sheldon RA, Kavelaars A, Derugin N, Vexler ZS, Willemen HL et al. Mesenchymal stem cell transplantation attenuates brain injury after neonatal stroke. Stroke 2013; 44: 1426–1432.
- 186 Pabon MM, Borlongan CV. Advances in the cell-based treatment of neonatal hypoxic-ischemic brain injury. *Future Neurol* 2013; **8**: 193–203.
- 187 George AL, Bangalore-Prakash P, Rajoria S, Suriano R, Shanmugam A, Mittelman A et al. Endothelial progenitor cell biology in disease and tissue regeneration. J Hematol Oncol 2011; 4: 24.

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