UC San Diego UC San Diego Previously Published Works

Title

Ventilation—perfusion heterogeneity measured by the multiple inert gas elimination technique is minimally affected by intermittent breathing of 100% O2

Permalink

https://escholarship.org/uc/item/2g9825f9

Journal Physiological Reports, 8(13)

ISSN 2051-817X

Authors

Elliott, Ann R Puliyakote, Abhilash S Kizhakke Tedjasaputra, Vincent <u>et al.</u>

Publication Date

2020-07-01

DOI

10.14814/phy2.14488

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <u>https://creativecommons.org/licenses/by/4.0/</u>

Peer reviewed

Ventilation–perfusion heterogeneity measured by the multiple inert gas elimination technique is minimally affected by intermittent breathing of 100% O_2

Ann R. Elliott^{1,2} | Abhilash S. Kizhakke Puliyakote^{2,3} | Vincent Tedjasaputra^{1,2} | Beni Pazár^{2,3} | Harrieth Wagner¹ | Rui C. Sá^{1,2} | Jeremy E. Orr¹ | G. Kim Prisk^{1,2,3} | Peter D. Wagner¹ | Susan R. Hopkins^{1,2,3}

¹Department of Medicine, University of California San Diego, La Jolla, CA, USA

²The Pulmonary Imaging Laboratory, University of California San Diego, La Jolla, CA, USA

³Department of Radiology, University of California San Diego, La Jolla, CA, USA

Correspondence

Ann R. Elliott, Department of Medicine 0852 University of California, San Diego 9500 Gilman Dr. La Jolla, CA 92093. Email: aelliott@ucsd.edu

Funding information

National Heart, Lung, and Blood Institute, Grant/Award Number: HL119201, HL119263 and HL129990

Abstract

Proton magnetic resonance (MR) imaging to quantify regional ventilation-perfusion (\dot{V}_A/\dot{Q}) ratios combines specific ventilation imaging (SVI) and separate proton density and perfusion measures into a composite map. Specific ventilation imaging exploits the paramagnetic properties of O2, which alters the local MR signal intensity, in an F₁O₂-dependent manner. Specific ventilation imaging data are acquired during five wash-in/wash-out cycles of breathing 21% O2 alternating with 100% O2 over ~20 min. This technique assumes that alternating F_1O_2 does not affect \dot{V}_A/\dot{Q} heterogeneity, but this is unproven. We tested the hypothesis that alternating F_1O_2 exposure increases \dot{V}_A/\dot{Q} mismatch in nine patients with abnormal pulmonary gas exchange and increased \dot{V}_A/\dot{Q} mismatch using the multiple inert gas elimination technique (MIGET). The following data were acquired (a) breathing air (baseline), (b) breathing alternating air/100% O₂ during an emulated-SVI protocol (eSVI), and (c) 20 min after ambient air breathing (recovery). MIGET heterogeneity indices of shunt, deadspace, ventilation versus \dot{V}_A/\dot{Q} ratio, LogSD \dot{V} , and perfusion versus \dot{V}_A/\dot{Q} ratio, LogSD \dot{Q} were calculated. LogSD \dot{V} was not different between eSVI and baseline (1.04 \pm 0.39 baseline, 1.05 \pm 0.38 eSVI, p = .84); but was reduced compared to baseline during recovery $(0.97 \pm 0.39, p = .04)$. There was no significant difference in LogSDQ across conditions (0.81 \pm 0.30 baseline, 0.79 \pm 0.15 eSVI, 0.79 ± 0.20 recovery; p = .54); Deadspace was not significantly different (p = .54) but shunt showed a borderline increase during eSVI ($1.0\% \pm 1.0$ baseline, $2.6\% \pm 2.9$ eSVI; p = .052) likely from altered hypoxic pulmonary vasoconstriction and/or absorption atelectasis. Intermittent breathing of 100% O2 does not substantially alter \dot{V}_A/\dot{Q} matching and if SVI measurements are made after perfusion measurements, any potential effects will be minimized.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

^{© 2020} The Authors. Physiological Reports published by Wiley Periodicals LLC on behalf of The Physiological Society and the American Physiological Society.

KEYWORDS

hyperoxia, magnetic resonance imaging, pulmonary perfusion distribution, pulmonary ventilation distribution, specific ventilation imaging, ventilation-perfusion ratio

1 INTRODUCTION

Many lung diseases are characterized by an increase in ventilation–perfusion (\dot{V}_A/\dot{Q}) mismatch, which impairs pulmonary gas exchange. Our laboratory has developed a proton magnetic resonance imaging (MRI) technique, to measure regional ventilation-perfusion mismatch, which combines specific ventilation imaging (SVI) with proton density (Holverda et al., 2011; Theilmann et al., 2009) and perfusion (Bolar et al., 2006) images to give regional \dot{V}_A/\dot{Q} ratios (Henderson et al., 2013; Sá et al., 2017).

Specific ventilation imaging noninvasively measures regional specific ventilation (Sá et al., 2010, 2014). During SVI, the subject breathes 20 breaths of 100% O_2 (~2 min, washin) followed by 20 breaths of air (~2 min, washout) and repeats this alternating block five times. Oxygen is paramagnetic; by breathing 100% O₂ and using an appropriate magnetic resonance sequence, inhaled O₂ is used as an MRI contrast agent (Sá et al., 2010, 2014). By analyzing the time course of the change in MR signal intensity on a voxel by voxel basis, a specific ventilation map is constructed (Sá et al., 2010, 2014), and then combined with density and perfusion measures.

Although it is a common tool in pulmonary function testing, (Comroe & Fowler, 1951; Fowler, 1949; Lewis, Evans, & Jalowayski, 1978; Martin, Tsunoda, & Young, 1974; Robinson et al., 2013; Verbanck et al., 1997, 1998), breathing hyperoxic gas could affect the underlying pulmonary physiology and \dot{V}_A/\dot{Q} matching. For example, O₂ breathing can alter regional blood flow and ventilation predominately by either the release of hypoxic pulmonary vasoconstriction and/or absorption atelectasis (Dawson, 1969, 1984; Lee & Read, 1967; Morrell, Nijran, Biggs, & Seed, 1994), but other mechanisms can also modulate these processes. Hyperoxia can either directly or indirectly alter bronchomotor tone (Astin & Penman, 1967; Libby, Briscoe, & King, 1981), which in turn potentially increases \dot{V}_{A}/\dot{Q} heterogeneity (Robinson, Freiberg, Regnis, & Young, 2000; Sassoon, Hassell, & Mahutte, 1987). Additionally, hyperoxia, due to its effects on blood flow and/or ventilatory drive (Aubier et al., 1980; Robinson et al., 2000; Sassoon et al., 1987) may alter local alveolar CO₂ which may in turn affect local lung mechanics and ventilation (Emery, Eveland, Kim, Hildebrandt, & Swenson, 2007; Emery, Eveland, Min, Hildebrandt, & Swenson, 2013; Traystman, Batra, & Menkes, 1976), and thus the underlying \dot{V}_A/\dot{Q} distribution.

The effect of intermittent hyperoxia, as used in proton MRI evaluation of \dot{V}_A/\dot{Q} relationships, on \dot{V}_A/\dot{Q} mismatch is unknown. Previously we showed that in healthy subjects, exposure to either hyperoxia or hypoxia did not affect ventilation heterogeneity (Hopkins, Elliott, Prisk, & Darquenne, 2017), but in healthy subjects any underlying \dot{V}_{A}/\dot{Q} mismatch is expected to be minimal and thus potential changes may be minimized. In the present study, we evaluated the effect of an emulated SVI (eSVI) O2 breathing protocol on \dot{V}_{A}/\dot{Q} heterogeneity in nine subjects with a clinical diagnosis of COPD. This subject population was chosen because they could reasonably be expected to show \dot{V}_A/\dot{Q} mismatch at baseline. We hypothesized that an eSVI protocol would increase \dot{V}_A/\dot{Q} heterogeneity consistent with underlying effects of O₂ either on hypoxic pulmonary vasoconstriction or by altering the ventilation distribution or a combination of both. To test this, we measured \dot{V}_A/\dot{Q} mismatch with the multiple inert gas elimination technique (MIGET) (Hlastala, 1984; Hlastala & Robertson, 1978; Roca & Wagner, 1994; Wagner, Naumann, & Laravuso, 1974), a well-established method for quantitatively assessing \dot{V}_A/\dot{Q} matching.

METHODS 2

2.1 **Subjects**

This study was approved by the University of California, San Diego's Human Subjects' Research Protection Program. Subjects were recruited via advertisements, pulmonary clinics, and existing research databases. Recruited subjects all had a diagnosis of COPD made by their personal physician, and all had a significant history of cigarette smoking (20 ± 12) pack-years). All subjects gave written informed consent prior to participating. Recruited subjects were screened with a medical questionnaire for assessment of exclusion criteria that included any history of significant illness aside from a diagnosis of COPD or a current history of a recent exacerbation. Nine subjects, seven males and two females, between 49 and 70 years old participated. Subjects remained on their daily medication such as long-acting bronchodilators and inhaled steroids for the study. Eight subjects completed the entire protocol; and one individual, subject #7, declined to complete the recovery portion of the study because of time constraints.

2.2 **Overall study design**

On the day of the study, all subjects underwent spirometry using an EasyOne spirometer (ndd Medical Technologies, Zurich, Switzerland). To assess the influences of alternating F_1O_2 concentrations on \dot{V}_A/\dot{Q} heterogeneity, subjects were studied using MIGET during three conditions: (a) steady-state quiet breathing of ambient air, baseline; (b) at the end of eSVI which consisted of alternating breathing air for 2 min followed by 2 min of 100% O_2 for a total of 20 min; and (c) after a 20-min period of air breathing, recovery (Figure 1). During the eSVI section of the protocol, to maximize the chance of seeing potential effects of F_1O_2 on the \dot{V}_A/\dot{Q} distribution, data and samples were acquired during the last two blocks (14–16 and 18–20 min) of 100% O_2 breathing.

2.3 | MIGET protocol

2.3.1 | Experimental setup

The subjects were supine for the study duration duplicating the necessary posture of subjects during SVI MRI data acquisition. The nondominant arm radial artery was catheterized using a 20 gauge, 1.25" catheter, for the collection of arterial blood samples, and for direct arterial pressure monitoring. A peripheral vein from the contralateral arm was catheterized with an 18 gauge, 2" catheter for the infusion of 5% dextrose solution containing the inert gas mixture. Body temperature was measured using an oral thermometer. ECG and cardiac output were monitored with a tetrapolar impedance cardiograph (model BioZ ICG, CardioDynamics), which was previously validated against direct Fick measures of cardiac output (Yung, Fedullo, Kinninger, Johnson, & Channick, 2004).

2.3.2 | Multiple inert gas elimination technique

The multiple inert gas elimination technique has been previously described in detail (Wagner, Laravuso, Uhl, & West, 1974; Wagner, Saltzman, & West, 1974), therefore is only briefly described here. Maintaining sterility, the six usual MIGET inert gases, (sulfur hexafluoride, ethane, cyclopropane, enflurane, ether, and acetone) were dissolved in 5% dextrose. This solution was continuously infused intravenously for the entire study duration at a rate of 2-5 ml/min. The rate was selected based on the subject's minute ventilation, at approximately 1 ml/min per 2 L/min ventilation, and has been shown to provide adequate signal to noise in the measurement of all inert gas levels. The infusion was started 30 min prior to baseline measurements to ensure steady-state conditions for inert gases. From the last 10 min onward, the subject breathed on a mouthpiece into a heated nonrebreathing valve (model 2700, Hans Rudolph Inc) that was connected to a heated (~45°C) low resistance expired gas mixing box. Steady-state conditions were confirmed by ensuring stable inert gas infusion, ventilation, and cardiac output prior to and during measurements.

2.3.3 | Sampling protocol

For each condition (baseline, eSVI, recovery), mixed expired O₂ and CO₂ concentrations were continuously sampled and measured from the mixing box using a mass spectrometer (Perkin Elmer, MGA 1100) and recorded using data acquisition software (LabView, National Instruments), while simultaneously measuring minute ventilation $(\dot{V}_{\rm F})$ using a calibrated gas rotameter (100 Liter, BOC), and mixing box temperature. For each condition, the data were averaged to calculate O2 consumption, carbon dioxide production, and the respiratory exchange ratio (VO₂, VCO₂, RER, respectively). Four arterial blood samples (5-7 ml each), and four time-aligned mixed expired gas samples (20 ml each) were collected for quadruplicate measurement of inert gas levels, while simultaneously measuring cardiac output. Immediately after each inert gas sample, 2 ml of arterial blood gas sample was drawn into a preheparinized 3-ml syringe, debubbled, stored on ice, and analyzed with a GEM 3000 analyzer (Instrumentation Laboratories) to measure P_aO_2 , P_aCO₂, pH_a, hematocrit, hemoglobin, and O₂ saturation. The



FIGURE 1 MIGET protocol sequence for data collection and sampling

alveolar–arterial PO₂ difference (A-aDO₂) was derived from respiratory data and the corresponding average of the four arterial blood gas samples. All measurements were repeated during eSVI where the mixed expired gas samples and associated arterial blood gases were collected in duplicate at the end of the last two 100% O₂ breathing periods (providing a quadruplicate sample set), and in duplicate at the end of a 20-min recovery air breathing period. \dot{VO}_2 and respiratory exchange ratio were not calculated at the end of the eSVI because O₂ consumption cannot be accurately measured during 100% O₂ breathing.

Inert gas concentrations were measured in the mixed expired and arterial blood samples using gas chromatography (5890A; Hewlett-Packard) (Wagner, Naumann, et al., 1974). Mixed venous concentrations of the inert gases were calculated by the Fick principle. Retention-solubility and excretion-solubility curves were generated for the six gases in each sample and transformed into a continuous plot of perfusion against \dot{V}_A/\dot{Q} and ventilation against \dot{V}_A/\dot{Q} using the 50-compartment model (Wagner, Saltzman, et al., 1974). The residual sum of squares (RSS) for the measured 6-gas data and the least squares best fit to the 50 compartment model was calculated for each data set (Hopkins & Wagner, 2017). The degree of mismatch was determined by calculating the second moments, about their mean, of the perfusion and ventilation distributions, on a log \dot{V}_A/\dot{Q} scale excluding shunt and deadspace compartments, which are evaluated separately. The second moments are termed LogSDO representing heterogeneity of the perfusion versus \dot{V}_A/\dot{Q} distribution, and LogSDV representing heterogeneity of the ventilation versus \dot{V}_A/\dot{Q} distribution.

2.4 | Statistical analysis

Statistical analyses were performed using KaleidaGraph (v4.5, Synergy Software). ANOVA for repeated measures was used to evaluate differences between baseline, eSVI, and recovery for all indices derived from MIGET, ventilation, and blood gas data. Where overall significance occurred, post hoc testing was conducted using a two-tailed Student's *t* test to evaluate differences between the three conditions. Significance was accepted at p < .05. All data are expressed as mean \pm standard deviation.

3 | RESULTS

3.1 | Subject demographics

All subjects tolerated the study well. The subjects' demographics, spirometric and blood gas data are listed in Table 1. The extent of pulmonary disease in this population ranged

TABLE 1	Baseline	individual	subject's demog	graphics, spirom	etric data, and b	lood gas data						
Subject	Sex	Age years	Height cm	BMI kg/m ²	FVC % pred.	FEV ₁ % pred.	FEV ₁ /FVC % pred.	Ý _E l/min BTPS	Ý O₂l /min BTPS	P _a O ₂ mmHg	P _a CO ₂ mmHg	AaDO ₂ mmHg
01	Μ	64	178	31.8	42	22	53	9.6	311	73	44	29
02	М	57	187	20.6	50	73	69	9.2	235	LL	42	24
03	М	67	178	32.8	59	52	88	7.1	254	75	45	23
04	М	70	172	28.7	74	61	82	8.1	268	74	34	38
05	М	55	158	18.0	91	75	83	7.0	165	81	39	26
90	ц	49	178	33.5	66	101	100	9.7	280	73	39	35
07	М	53	159	34.8	73	81	108	10.0	292	LT LT	42	29
08	ц	66	154	34.6	92	92	98	9.9	238	83	23	42
60	М	69	178	24.3	78	60	77	6.7	221	64	34	42
Mean $\pm SD$		61 ± 8	171 ± 12	28.8 ± 6.3	73 ± 20	69 ± 23	84 ± 17	8.5 ± 1.4	244 ± 40	76 ± 6	37 ± 7	32±8
Note: $n = 9$; (model) Abbreviations: I	$an \pm SD$).	. alveolar-arte	erial difference fo	r oxvœn: Ϋ́,, mir	uite ventilation: P.	.O. alveolar oxy	oen nartial nressure.	FEV.% nred fo	rced exniratory vol	ume in 1 s nero	ent nredicted [.] FFV./FV	C % nred forced

expiratory volume in 1 s to forced vital capacity ratio percent predicted; FVC % pred, forced vital capacity percent predicted; P_xCO_x, arterial carbon dioxide partial pressure; P_xO_y, arterial oxygen partial pressure;

from very mild to severe based on spirometry with FEV1 percent predicted ranging from 99% to 22%. Three of the subjects (Subjects 6, 7, and 8) with self-reported smoking histories of 37, 30, and 10 pack years, had relatively normal spirometry, despite their clinical diagnosis of COPD. These three subjects had A-aDO₂ differences of 35, 29, and 42 mmHg, respectively, and LogSD \dot{Q} values above the normal LogSD \dot{Q} range of 0.3-0.6 (Figure 2). These gas exchange values are similar to that of GOLD stage 1 COPD patients (Rodríguez-Roisin et al., 2009).

3.2 Metabolic, arterial blood gas data, and A-aDO₂ (Tables 1 and 2)

Cardiac output, \dot{V}_E and $\dot{V}CO_2$ all showed no significant differences across baseline, eSVI, and recovery (p = .69, p = .22, and p = .61, respectively) confirming steady-state conditions. $\dot{V}O_2$ cannot be assessed during hyperoxic breathing and is not reported, but V O_2 was not different between baseline and recovery (p = .28). At baseline all nine subjects had an abnormally low P_aO_2 (mean $P_aO_2 = 75.5 \pm 5.7$ mmHg). P_aO_2 showed the expected increase during eSVI, increasing to a mean $P_aO_2 = 438 \pm 40.5 \text{ mmHg} (p < .0001) \text{ com-}$ pared to baseline and recovery, and P_aO₂ during the recovery period (83.3 \pm 7.1 mmHg) was higher than baseline $(75.5 \pm 5.7 \text{ mmHg})$ (p = .04). Similarly, there was the expected increase in the S_aO_2 from baseline (93.4 \pm 1.9%) to eSVI of 95.7 \pm 1.8% (p < .0001), and the recovery S_aO₂ $(94.0 \pm 1.9\%)$ was also greater (p = .04) than baseline conditions. A-aDO₂ cannot be calculated during O₂ breathing because \dot{VO}_2 and respiratory exchange ratio are unknown. The baseline and recovery A-aDO₂ were elevated in all subjects (mean 32 ± 8 mmHg and 30 ± 12 mmHg, respectively), and were not significantly different from each other (p = .22). P_aCO₂ was not different between baseline $(37.3 \pm 7.0 \text{ mmHg})$ and eSVI $(36.3 \pm 6.8 \text{ mmHg})$, but recovery P_aCO₂ was significantly lower than baseline $(34.6 \pm 6.7 \text{ mmHg}, p = .03)$, suggesting increased alveolar ventilation. There were no significant changes in pH_a, hemoglobin or hematocrit across all three conditions, p = .29, p = .40, and p = .28, respectively.

3.3 **MIGET data**

The residual sum of squares (Table 2) were consistent with good data quality and adequacy of fit to the model for each condition. The RSS values were not different across conditions (p = .33). There were no significant differences between conditions for deadspace ventilation (\dot{V}_{DS}) (p = .54) or



FIGURE 2 Individual LogSDQ, LogSDV and shunt for the nine subjects, at baseline, eSVI, and recovery. $LogSD\dot{Q}$ = heterogeneity of the perfusion versus V_A/Q distribution, $LogSD\dot{V}$ = heterogeneity of the ventilation versus V_A/Q distribution; and \dot{Q} shunt = percent of cardiac output to shunt regions ($\dot{V}_A/\dot{Q} < 0.005$). [#]denotes post hoc statistical difference between baseline and recovery at p < .05 (twotailed t test). Note: normal LogSDQ range of 0.3-0.6; and normal LogSDV range 0.3-0.65 (Hopkins & Wagner, 2017)

the means (i.e., first moments) of the ventilation (p = .38) or perfusion distributions (p = .90).

Baseline Oshunt (Table 2 and Figure 2) showed an small increase (1.0 \pm 1.0% of cardiac output) compared to negligible values seen in normal subjects (Wagner, Laravuso,

EL	LI	CO1	Т	ΕT	AL
		· ·			

TABLE 2 Baseline, eSVI, and recovery supine physiologic data

				ANOVA
	Baseline	eSVI	Recovery	p
Hb (g/dl)	13.7 ± 2.0	13.6 ± 2.0	13.2 ± 0.3	.40
Hct (%)	43.0 ± 7.0	41.9 ± 6.6	41.1 ± 7.1	.28
<i>Q</i> (l/min)	5.4 ± 0.5	5.3 ± 0.6	5.3 ± 0.5	.69
HR (bts/min)	76 ± 11	71 ± 11	73 ± 11	.03
$\dot{V}_{\rm E}$ (l/min)	8.5 ± 1.4	8.8 ± 1.4	9.0 ± 1.2	.22
$\dot{V}O_2$ (ml/min)	244 ± 40	na	224 ± 47	.28
^{VCO} ₂ (ml/min)	206 ± 42	206 ± 44	205 ± 38	.61
RER (ratio)	0.84 ± 0.06	na	$0.92\pm0.09\#$.046
$S_aO_2(\%)$	93.4 ± 1.9	$95.7 \pm 1.8^{*}$	$94.0 \pm 1.9^{\#\$}$	<.0001
A-aDO ₂ (mmHg)	32.4 ± 7.7	na	29.9 ± 11.8	.22
$P_aO_2(mmHg)$	75.5 ± 5.7	$438.3 \pm 40.5^*$	$83.3 \pm 7.1^{\#\$}$.001
P _a CO ₂ (mmHg)	37.3 ± 7.0	36.3 ± 6.8	34.6 ± 6.7	.03
pH _a	7.46 ± 0.05	7.47 ± 0.05	7.47 ± 0.05	.29
RSS	5.26 ± 2.99	5.61 ± 6.22	4.40 ± 4.14	.33
$\dot{V}_{\rm DS}(\%\dot{V})$	0.36 ± 0.04	0.37 ± 0.07	0.37 ± 0.09	.54
\dot{Q} Shunt (% \dot{Q})	1.0 ± 1.0	2.6 ± 2.9	1.8 ± 2.8	.052
Mean <i>V</i>	1.89 ± 0.54	2.03 ± 0.50	1.89 ± 0.36	.38
LogSDV	1.04 ± 0.39	1.05 ± 0.38	$0.97 \pm 0.39^{\#}$.050
Mean Q	0.81 ± 0.30	0.83 ± 0.20	0.86 ± 0.24	.90
LogSDQ	0.81 ± 0.22	0.79 ± 0.15	0.79 ± 0.20	.54

Note: p values for ANOVA testing, when significant *denotes post hoc statistical difference between baseline and emulated-SVI; and [#]denotes post hoc statistical difference between baseline and recovery; and ^{\$}denotes post hoc statistical difference between emulated-SVI and recovery. Post hoc significance was accepted at p < .05 (-tailed *t*-test). Note: normal LogSDQ range of 0.3–0.6; and normal LogSDV range 0.3–0.65 (Hopkins & Wagner, 2017). (n = 9 except recovery n = 8, mean $\pm SD$).

Abbreviations: \dot{Q} shunt, percent of cardiac output to shunt regions ($\dot{V}_A/\dot{Q} < 0.005$); \dot{Q} , cardiac output; \dot{V} CO_2 , carbon dioxide production; \dot{V}_E , minute ventilation; $\dot{V}O_2$, oxygen consumption; A-aDO₂, alveolararterial difference for oxygen; Hb, hemoglobin; Hct, hematocrit; HR, heart rate; LogSDQ, heterogeneity of the perfusion versus \dot{V}_A/\dot{Q} distribution; LogSDV, heterogeneity of the ventilation versus \dot{V}_A/\dot{Q} distribution; mean \dot{Q} , mean of the perfusion distribution; mean \dot{V} , mean of the ventilation distribution; na, values could not be calculated during hyperoxia; PaCO2, arterial carbon dioxide partial pressure; PaO2, arterial oxygen partial pressure; pHa, arterial pH; RER, respiratory exchange ratio; RSS, residual sum of squares; SaO2, arterial oxygen saturation; \dot{V}_{DS} , percent of ventilation to deadspace ($\dot{V}_A/\dot{Q} > 100$).

et al., 1974), and showed a borderline change across conditions (p = .052) increasing during eSVI to $2.6 \pm 2.9\%$.

Baseline LogSD \dot{Q} and LogSD \dot{V} measures of heterogeneity were increased above normal values consistent with the diagnosis of COPD in these subjects (Rodríguez-Roisin et al., 2009; Wagner, Dantzker, Dueck, Clausen, & West, 1977). Figure 2 shows baseline, eSVI and recovery LogSDQ and LogSDV and shunt values for all nine subjects. Mean LogSDQ was not significantly different across the three conditions $(0.81 \pm 0.22$ baseline, 0.79 ± 0.15 eSVI, and 0.79 ± 0.20 recovery, p = .54). Mean LogSDV of 1.05 ± 0.38 during eSVI was not significantly different from baseline $(1.04 \pm 0.39, p = .84)$, but LogSDV in recovery showed a small but statistically significant reduction to 0.97 ± 0.39 (p = .04) compared to baseline, but it was not significantly different than that during eSVI (p = .17).

DISCUSSION 4

The principal findings of this study are that in subjects with varying degrees of underlying \dot{V}_{A}/\dot{Q} mismatch and a clinical diagnosis of COPD, there was no significant change in the MIGET indices that measure global ventilation and perfusion heterogeneity, LogSD \dot{V} , and LogSD \dot{Q} compared to baseline air breathing; the LogSDV, was slightly reduced after 20 min of air breathing. There was also no significant change in deadspace, although there was a borderline increase in shunt during the eSVI protocol. These findings suggest that changes in \dot{V}_A/\dot{Q} mismatch during hyperoxic exposure are small and that SVI is an appropriate method to use in proton MR measures of regional \dot{V}_A/\dot{Q} ratio. However, because SVI may potentially cause small changes in pulmonary blood flow and/or shunt, we suggest the perfusion imaging always be performed before SVI so to minimize any protocol impact on \dot{V}_A/\dot{Q} mapping.

4.1 | Subject population and spirometry

In this study, baseline LogSD \dot{Q} and the LogSD \dot{V} measures of heterogeneity were markedly increased above normal values consistent with the goal of selecting subjects with underlying \dot{V}_A/\dot{Q} mismatch and with their diagnosis of COPD (Rodríguez-Roisin et al., 2009; Wagner et al., 1977). All subjects demonstrated hypoxemia during baseline air breathing, even subjects with very mild air flow obstruction (Table 1). The subjects with normal spirometry had significant elevation in A-aDO₂ and \dot{V}_A/\dot{Q} mismatch (Table 1 and Figure 2), which were similar to those subjects with more severe disease. This is consistent with data from Rodríguez-Roisin et al. (2009) who showed that the extent of \dot{V}_A/\dot{Q} inequality was poorly correlated with disease severity based on spirometry.

4.2 | Potential effects of O_2 on the distribution of ventilation, perfusion, and \dot{V}_A/\dot{Q} mismatch

The distribution of ventilation is affected by the structural heterogeneity of the bronchial and vasculature trees and the feedback systems designed to regulate the local \dot{V}_A/\dot{Q} matching to maximize overall gas exchange. The respiratory gases have some regulatory control of bronchomotor tone and the degree of effect is based on the size of the affected region and local environment of the region, but the effects are small and varied (Swenson, Domino, & Hlastala, 1998). Alveolar hypoxia has a mild bronchoconstrictive effect as evidenced by increases in airway resistance (Saunders et al., 1977; Sterling, 1968), and this effect may be modulated by local CO₂ concentrations; in general, hypocapnia enhances bronchoconstriction (Elshout, van Herwaarden, & Folgering, 1991; Ingram, 1975; Twort, Neild, & Cameron, 1985) while hypercapnia inhibits the increase in collateral airways resistance (Traystman et al., 1976). Hyperoxia exposure in healthy subjects has been shown to have little bronchomotor effect when looking at changes in maximal expiratory flow rates, airway conductances and resistances (Astin & Penman, 1967; Butler, Caro, Alcara, & Dubois, 1960; Libby et al., 1981). Consistent with this, in healthy subjects, we previously showed that neither hypoxic ($F_1O_2 = 0.125$) nor a hyperoxic ($F_1O_2 = 0.9$) (Hopkins et al., 2017) exposure altered ventilation heterogeneity as measured by the multiple breath washout.

All our subjects were hypoxemic during baseline conditions which was relieved during eSVI. In hypoxemic COPD patients, bronchoconstriction can occur (Astin & - The Physiological Physiological Reports-

Penman, 1967; Libby et al., 1981) and is reversed by breathing 30% O_2 ; but it is not known if the bronchoconstriction is the direct effect of low arterial O2 tensions on the bronchial smooth muscle or the reduced lower airway and alveolar O₂ tensions. In the presence of hypoxemia-induced bronchoconstriction, hyperoxic exposure could cause bronchodilation causing an increase in regions of high \dot{V}_A/\dot{Q} ratio or deadspace $(\dot{V}_A/\dot{Q} > 100)$, but this was not seen in the present study. In some COPD patients, hyperoxia has been shown to induce hypercapnia (Abdo & Heunks, 2012; Aubier et al., 1980; Robinson et al., 2000; Sassoon et al., 1987), which is thought to be primarily caused by an increase in \dot{V}_{A}/\dot{Q} mismatch and deadspace. This may be related to the release of hypoxic pulmonary vasoconstriction, secondary bronchodilatory effects, and a decrease in collateral resistance due to the relaxation of local parenchymal tissue associated with elevated P_aCO₂ due to concurrent hypoventilation. The absence of changes in \dot{V}_A/\dot{Q} matching, deadspace, ventilation, or P_aCO_2 during hyperoxia suggests that this is not a major factor in the present study.

One possibility for the borderline increase in shunt is absorption atelectasis due to alveolar de-nitrogenation (Dantzker, Wagner, & West, 1975). However, it is thought that parenchymal interdependence forces of attached neighboring alveoli and collateral ventilation may counteract the loss of volume and preventing alveolar collapse (Briscoe, Cree, Filler, Houssay, & Cournand, 1960; Dantzker et al., 1975). In COPD, it has been shown that the degree of collateral ventilation increases with disease severity (Hogg, Macklem, & Thurlbeck, 1969) and this increase in collateral flow potentially counteracts the development of absorption atelectasis and increase shunt (Barberà et al., 1996; Santos et al., 2000). In some COPD subjects, hyperoxia worsens \dot{V}_A/\dot{Q} matching resulting in an increase in the number of low \dot{V}_A/\dot{Q} units but with little or no increase in shunt (Barberà et al., 1996; Rodriguez-Roisin, 2014; Santos et al., 2000). This is the case for the majority of subjects in the present study, and the reversibility argues against atelectasis as a mechanism. However, the response of subject #8 suggests the possibility of absorption atelectasis in this individual because the shunt did not resolve during recovery, although the changes were small.

Hyperoxia may release hypoxic pulmonary vasoconstriction resulting in a deterioration of \dot{V}_A/\dot{Q} matching by increasing pulmonary blood flow to poorly ventilated or shunt regions. The time course of hypoxic pulmonary vasoconstriction during severe regional hypoxia has been shown to have different profiles based on the type of preparations or protocols, species studied, and global versus regional exposure (Sylvester, Shimoda, Aaronson, & Ward, 2012; Talbot, Balanos, Dorrington, & Robbins, 2005). In general, the hypoxic pulmonary vasoconstrictor response to global hypoxia has an initial rapid onset profile (within seconds) that reaches a maximum by 30 min, followed by a secondary gradual increase over the next 2 hr. The relaxation of the hypoxia induced vasoconstriction is less clear. In humans, 25 min of isocapnic hypoxia exposure causes a rapid rise followed by a plateau in pulmonary vascular resistance, which subsequently after 5 min of normoxia returns to baseline (Talbot et al., 2005). However, with a longer isocapnic hypoxia exposure (105 min), pulmonary vascular resistance rapidly reached a plateau then slowly continued to rise, but took longer than 10 min to return to baseline. This suggests that the relaxation response with the removal of a hypoxic stimulus may follow a different time profile than the activation process. The lack of marked changes in the present study may simply be because the hyperoxic exposure was too short (10 min in total) and intermittent.

There is evidence that structural changes, and thickening of the intimal layer of the pulmonary muscular arteries may affect the ability of the pulmonary vasculature to respond to changes in O₂ and thus hypoxic pulmonary vasoconstriction (Barberà et al., 1994). Work in healthy subjects and in stable mild to severe COPD subjects which used MIGET before and after breathing 100% O_2 for >30 min, (Barbera et al., 1990; Barberà et al., 1994; Wagner et al., 1977; Wagner, Laravuso, et al., 1974) showed no change in either in LogSDQ, and LogSDV and no change in deadspace in healthy and some COPD subjects. However, other COPD subjects showed an increase in LogSDQ, and increased perfusion to regions of low \dot{V}_A/\dot{Q} . This variability between COPD subjects was attributed to the degree of vascular remodeling associated with disease progression (Barberà et al., 1994). Consistent with this, two of our subjects with normal spirometry #6 and #8, (Figure 2), who had the largest increase in shunt with hyperoxia, had a conversion of existing low \dot{V}_{A}/\dot{Q} into shunt during eSVI, which was reversed in recovery. The subjects with more severe airway obstruction (subjects 1-5, 9) showed no change in low \dot{V}_A/\dot{Q} units and no increase in shunt with hyperoxia which agrees with other studies with subjects with advanced airway obstruction (Barberà et al., 1996; Santos et al., 2000). Thus, in this study, the cyclic two-minute intermittent exposures to hyperoxia during SVI are short enough that it is unlikely that complete release of hypoxic pulmonary vasoconstriction occurs during the hyperoxia exposure. This is even less likely in COPD patients if they have lower vascular responsiveness to changes in O2 concentrations. Consequently, this is likely why, on average, we observed minimal changes in the LogSDV and LogSDQ.

4.3 | Post eSVI recovery and ventilation and perfusion heterogeneity

At the end of the recovery period, there was a small but significant reduction in $\text{LogSD}\dot{V}$ compared to baseline; while

LogSD \dot{Q} remained unchanged. The respiratory exchange ratio was significantly increased from 0.84 ± 0.06 during baseline to 0.92 ± 0.09 during recovery (p = .046), associated with a small decrease in PaCO₂. This small increase in ventilation, possibly due to subject discomfort over the study course (~2 hr), may be partly responsible for the increase in P_AO₂ and P_aO₂ in recovery. The resulting \dot{V}_A/\dot{Q} distribution during recovery showed a minor shift toward higher \dot{V}_A/\dot{Q} units compared to baseline. Therefore, the slight improvement in gas exchange after recovery was probably due to a combination of factors, including increased ventilation, small reduction in ventilation–perfusion mismatch and return of hypoxic pulmonary vasoconstriction, none of which were statically significant on their own.

4.4 | Limitations

MIGET is a method that measures pulmonary gas exchange by recovering the global \dot{V}_A/\dot{Q} distribution from excretion and retention patterns of six infused inert gases, thus providing an overall index of \dot{V}_A/\dot{Q} heterogeneity. Because of the global nature of the MIGET results, we cannot identify if there are any regional perfusion or ventilation heterogeneity changes occurring during hyperoxia that were counterbalanced by opposing changes in other regions. We studied a small number of subjects; thus, the generalizability of the findings is limited by how well the population studied is representative population as a whole. When statistical significance is lacking, the study statistical power must be considered, since a small number of subjects may impair the ability to detect meaningful differences. This study used a repeated measures design, thus each subject is their own control, and MIGET indices are highly reliable (test retest correlation ~.9 (Wagner, Hedenstierna, Bylin, & Lagerstrand, 1987)). For the observed differences in LogSDV and LogSDO between baseline and eSVI to be statistically significant (p < .05, twotailed) in a larger sample, post hoc power calculations reveal that at least 2,338 and 228 subjects, respectively, would be required. Thus, any difference between conditions is biologically very small.

As a quality control step, MIGET retention data were matched by a fitting process, the sum of squares of the six gas residuals, and this RSS for all three conditions was calculated. The cumulative chi-square distribution (six degrees of freedom) predicts a mean RSS of 5.3 (50th percentile) and 10.6 (90th percentile) to be indicative of an adequate data fit for the 50 compartment model (Hopkins & Wagner, 2017). Baseline, eSVI and recovery RSS were 5.3 ± 3.0 , 5.6 ± 6.2 , and 4.4 ± 4.1 , respectively, thus demonstrating adequate data quality and supports existence of steady-state conditions. The mean RSS for all three conditions were not statistically different from each other, suggesting that alternating F₁O₂ did

not disturb gas exchange for MIGET gases to any identifiable degree.

5 | CONCLUSIONS

We did not find evidence that alternating inspired O_2 concentration between 21% and 100% for five cycles for 20 min affects \dot{V}_A/\dot{Q} heterogeneity in subjects with lung disease in physiologically significant ways. Results from this study suggest that intermittent hyperoxia exposure that occurs during SVI which is used to calculate of regional \dot{V}_A/\dot{Q} with proton MRI has a minimal effect on \dot{V}_A/\dot{Q} heterogeneity as measured by MIGET. Accordingly, we suggest that the assumption of insignificant effects on \dot{V}_A/\dot{Q} relationships due to SVI protocol is reasonable, supporting its use as part of a methodology to evaluate \dot{V}_A/\dot{Q} inequality noninvasively. Nevertheless, given the findings, collection of perfusion data, which does not require altering F_IO_2 , is recommended before performing SVI.

ACKNOWLEDGMENTS

This study is supported by NIH/NHBLI HL119263, HL129990, and HL119201.

CONFLICT OF INTEREST

The authors listed certify that they have no affiliations with or involvement in any organization or entity with any financial or nonfinancial interest in the subject matter or materials discussed in this manuscript.

AUTHOR CONTRIBUTIONS

ARE, AKP, VT, GKP, PDW, and SRH contributed to study design and conception. ARE, AKP, VT, BP, JEO, HW, GKP, PDW, and SRH carried out data acquisition. All authors performed analysis and interpretation. ARE and SRH drafted the manuscript; all authors revised the manuscript. All authors provided final approval of the manuscript. ARE had full access to the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

ORCID

Ann R. Elliott D https://orcid.org/0000-0002-7959-3376 Jeremy E. Orr D https://orcid.org/0000-0002-4498-5337

REFERENCES

- Abdo, W. F., & Heunks, L. M. (2012). Oxygen-induced hypercapnia in COPD: Myths and facts. *Critical Care*, 16, 323.
- Astin, T. W., & Penman, R. W. (1967). Airway obstruction due to hypoxemia in patients with chronic lung disease. *American Review of Respiratory Disease*, 95, 567–575.
- Aubier, M., Murciano, D., Fournier, M., Milic-Emili, J., Pariente, R., & Derenne, J. P. (1980). Central respiratory drive in acute respiratory

failure of patients with chronic obstructive pulmonary disease. *The American Review of Respiratory Disease*, *122*, 191–199.

- Physiological Physiological Reports

- Barbera, J. A., Ramirez, J., Roca, J., Wagner, P. D., Sanchez-Lloret, J., & Rodriguez-Roisin, R. (1990). Lung structure and gas exchange in mild chronic obstructive pulmonary disease. *American Review of Respiratory Disease*, 141(4_pt_1), 895–901. https://doi. org/10.1164/ajrccm/141.4_Pt_1.895
- Barberà, J. A., Riverola, A., Roca, J., Ramirez, J., Wagner, P. D., Ros, D., ... Rodriguez-Roisin, R. (1994). Pulmonary vascular abnormalities and ventilation-perfusion relationships in mild chronic obstructive pulmonary disease. *American Journal of Respiratory and Critical Care Medicine*, 149(2), 423–429. https://doi.org/10.1164/ ajrccm.149.2.8306040
- Barberà, J. A., Roger, N., Roca, J., Rovira, I., Higenbottam, T. W., & Rodriguez-Roisin, R. (1996). Worsening of pulmonary gas exchange with nitric oxide inhalation in chronic obstructive pulmonary disease. *Lancet*, 347, 436–440. https://doi.org/10.1016/S0140 -6736(96)90011-2
- Bolar, D. S., Levin, D. L., Hopkins, S. R., Frank, L. F., Liu, T. T., Wong, E. C., & Buxton, R. B. (2006). Quantification of regional pulmonary blood flow using ASL-FAIRER. *Magnetic Resonance in Medicine*, 55(6), 1308–1317. https://doi.org/10.1002/mrm.20891
- Briscoe, W. A., Cree, E. M., Filler, J., Houssay, H. E. J., & Cournand, A. (1960). Lung volume, alveolar ventilation and perfusion interrelationships in chronic pulmonary emphysema. *Journal* of Applied Physiology, 15, 785–795. https://doi.org/10.1152/ jappl.1960.15.5.785
- Butler, J., Caro, C. G., Alcara, R., & Dubois, A. B. (1960). Physiological factors affecting airway resistance in normal subjects and in patients with obstructive respiratory disease. *Journal of Clinical Investigation*, 39(4), 584–591. https://doi.org/10.1172/JCI104071
- Comroe, J. H., & Fowler, W. S. (1951). Lung function studies. VI. Detection of uneven alveolar ventilation during a single breath of oxygen. American Journal of Medicine, 10(4), 408–413. https://doi. org/10.1016/0002-9343(51)90285-9
- Dantzker, D., Wagner, P., & West, J. (1975). Instability of lung untis with low VA/Q ratios during O2 breathing. *Journal of Applied Physiology*, 38, 886–895.
- Dawson, A. (1969). Regional pulmonary blood flow in sitting and supine man during and after acute hypoxia. *Journal of Clinical Investigation*, 48(2), 301–310. https://doi.org/10.1172/JCI105986
- Dawson, C. A. (1984). Role of pulmonary vasomotion in physiology of the lung. *Physiological Reviews*, 64(2), 544–616. https://doi. org/10.1152/physrev.1984.64.2.544
- Emery, M. J., Eveland, R. L., Kim, S. S., Hildebrandt, J., & Swenson, E. R. (2007). CO2 relaxes parenchyma in the liquid-filled rat lung. *Journal of Applied Physiology*, 103, 710–716.
- Emery, M. J., Eveland, R. L., Min, J.-H., Hildebrandt, J., & Swenson, E. R. (2013). CO2 relaxation of the rat lung parenchymal strip. *Respiratory Physiology & Neurobiology*, 186(1), 33–39. https://doi. org/10.1016/j.resp.2012.12.014
- Fowler, W. S. (1949). Lung function studies; uneven pulmonary ventilation in normal subjects and in patients with pulmonary disease. *Journal of Applied Physiology*, 2, 283–299.
- Henderson, A. C., Sá, R. C., Theilmann, R. J., Buxton, R. B., Prisk, G. K., & Hopkins, S. R. (2013). The gravitational distribution of ventilation-perfusion ratio is more uniform in prone than supine posture in the normal human lung. *Journal of Applied Physiology*, 115(3), 313–324. https://doi.org/10.1152/japplphysi ol.01531.2012

- Hlastala, M. P. (1984). Multiple inert gas elimination technique. Journal of Applied Physiology, 56(1), 1–7. https://doi.org/10.1152/ jappl.1984.56.1.1
- Hlastala, M. P., & Robertson, H. T. (1978). Inert gas elimination characteristics of the normal and abnormal lung. *Journal of Applied Physiology*, 44(2), 258–266. https://doi.org/10.1152/ jappl.1978.44.2.258
- Hogg, J. C., Macklem, P. T., & Thurlbeck, W. M. (1969). The resistance of collateral channels in excised human lungs. *Journal of Clinical Investigation*, 48(3), 421–431. https://doi.org/10.1172/ JCI105999
- Holverda, S., Theilmann, R. J., Sá, R. C., Arai, T. J., Hall, E. T., Dubowitz, D. J., ... Hopkins, S. R. (2011). Measuring lung water: Ex vivo validation of multi-image gradient echo MRI. *Journal of Magnetic Resonance Imaging*, 34, 220–224.
- Hopkins, S. R., Elliott, A. R., Prisk, G. K., & Darquenne, C. (2017). Ventilation heterogeneity measured by multiple breath inert gas testing is not affected by inspired oxygen concentration in healthy humans. *Journal of Applied Physiology*, *122*(6), 1379–1387. https:// doi.org/10.1152/japplphysiol.01013.2016
- Hopkins, S. R., & Wagner, P. D. (2017). The multiple inert gas elimination technique (MIGET), Methods in Physiology (1st ed., pp. 1–329). US: Springer. https://www.springer.com/gp/book/97814 93974405
- Ingram, R. H. (1975). Effects of airway versus arterial CO2 changes on lung mechanics in dogs. *Journal of Applied Physiology*, 38(4), 603–607. https://doi.org/10.1152/jappl.1975.38.4.603
- Lee, J., & Read, J. (1967). Effect of oxygen breathing on distribution of pulmonary blood flow in chronic obstructive lung disease. *The American Review of Respiratory Disease*, 96, 1173–1180.
- Lewis, S. M., Evans, J. W., & Jalowayski, A. A. (1978). Continuous distributions of specific ventilation recovered from inert gas washout. *Journal of Applied Physiology*, 44(3), 416–423. https://doi. org/10.1152/jappl.1978.44.3.416
- Libby, D. M., Briscoe, W. A., & King, T. K. (1981). Relief of hypoxia-related bronchoconstriction by breathing 30 per cent oxygen. *The American Review of Respiratory Disease*, 123, 171–175.
- Martin, C. J., Tsunoda, S., & Young, A. C. (1974). Lung emptying patterns in diffuse obstructive pulmonary syndromes. *Respiration Physiology*, 21(2), 157–168. https://doi.org/10.1016/0034-5687(74)90091-7
- Morrell, N. W., Nijran, K. S., Biggs, T., & Seed, W. A. (1994). Changes in regional pulmonary blood flow during lobar bronchial occlusion in man. *Clinical Science*, 86(5), 639–644. https://doi.org/10.1042/ cs0860639
- Robinson, P. D., Latzin, P., Verbanck, S., Hall, G. L., Horsley, A., Gappa, M., ... Gustafsson, P. M. (2013). Consensus statement for inert gas washout measurement using multiple- and single- breath tests. *European Respiratory Journal*, 41(3), 507–522. https://doi. org/10.1183/09031936.00069712
- Robinson, T. D., Freiberg, D. B., Regnis, J. A., & Young, I. H. (2000). The role of hypoventilation and ventilation-perfusion redistribution in oxygen-induced hypercapnia during acute exacerbations of chronic obstructive pulmonary disease. *American Journal of Respiratory and Critical Care Medicine*, 161(5), 1524–1529. https://doi.org/10.1164/ajrccm.161.5.9904119
- Roca, J., & Wagner, P. D. (1994). Contribution of multiple inert gas elimination technique to pulmonary medicine. 1. Principles and information content of the multiple inert gas elimination technique. *Thorax*, 49, 815–824. https://doi.org/10.1136/thx.49.8.815

- Rodriguez-Roisin, R. (2014). Nonpulmonary influences on gas exchange. Comprehensive Physiology, 4, 1455–1494.
- Rodríguez-Roisin, R., Drakulovic, M., Rodríguez, D. A., Roca, J., Barberà, J. A., & Wagner, P. D. (2009). Ventilation-perfusion imbalance and chronic obstructive pulmonary disease staging severity. *Journal of Applied Physiology*, *106*(6), 1902–1908. https://doi. org/10.1152/japplphysiol.00085.2009
- Sá, R. C., Asadi, A. K., Theilmann, R. J., Hopkins, S. R., Prisk, G. K., & Darquenne, C. (2014). Validating the distribution of specific ventilation in healthy humans measured using proton MR imaging. *Journal* of Applied Physiology, 116(8), 1048–1056. https://doi.org/10.1152/ japplphysiol.00982.2013
- Sá, R. C., Cronin, M. V., Henderson, A. C., Holverda, S., Theilmann, R. J., Arai, T. J., ... Prisk, G. K. (2010). Vertical distribution of specific ventilation in normal supine humans measured by oxygen-enhanced proton MRI. *Journal of Applied Physiology*, *109*(6), 1950–1959. https://doi.org/10.1152/japplphysiol.00220.2010
- Sá, R. C., Henderson, A. C., Simonson, T., Arai, T. J., Wagner, H., Theilmann, R. J., ... Hopkins, S. R. (2017). Measurement of the distribution of ventilation-perfusion ratios in the human lung with proton MRI: Comparison with the multiple inert-gas elimination technique. *Journal of Applied Physiology*, 123, 136–146.
- Santos, C., Ferrer, M., Roca, J., Torres, A., Hernández, C., & Rodriguez-Roisin, R. (2000). Pulmonary gas exchange response to oxygen breathing in acute lung injury. *American Journal of Respiratory* and Critical Care Medicine, 161(1), 26–31. https://doi.org/10.1164/ ajrccm.161.1.9902084
- Sassoon, C. S., Hassell, K. T., & Mahutte, C. K. (1987). Hyperoxicinduced hypercapnia in stable chronic obstructive pulmonary disease. *The American Review of Respiratory Disease*, 135, 907–911.
- Saunders, N. A., Betts, M. F., Pengelly, L. D., & Rebuck, A. S. (1977). Changes in lung mechanics induced by acute isocapnic hypoxia. *Journal of Applied Physiology*, 42(3), 413–419. https://doi. org/10.1152/jappl.1977.42.3.413
- Sterling, G. M. (1968). The mechanism of bronchoconstriction due to hypocapnia in man. *Clinical Science*, 34, 277–285.
- Swenson, E., Domino, K., & Hlastala, M. P. (1998). Physiological effects of oxygen and carbon dioxide on VA/Q heterogeneity. In M. P. Hlastala & H. T. Robertson (Eds.), *Complexity in structure and function of the lung* (pp. 511–547). Bethesda: Marcel-Dekker.
- Sylvester, J. T., Shimoda, L. A., Aaronson, P. I., & Ward, J. P. (2012). Hypoxic pulmonary vasoconstriction. *Physiological Reviews*, 92(1), 367–520. https://doi.org/10.1152/physrev.00041.2010
- Talbot, N. P., Balanos, G. M., Dorrington, K. L., & Robbins, P. A. (2005). Two temporal components within the human pulmonary vascular response to approximately 2 h of isocapnic hypoxia. *Journal of Applied Physiology*, 98, 1125–1139.
- Theilmann, R. J., Arai, T. J., Samiee, A., Dubowitz, D. J., Hopkins, S. R., Buxton, R. B., & Prisk, G. K. (2009). Quantitative MRI measurement of lung density must account for the change in T(2) (*) with lung inflation. *Journal of Magnetic Resonance Imaging*, 30, 527–534.
- Traystman, R. J., Batra, G. K., & Menkes, H. A. (1976). Local regulation of collateral ventilation by oxygen and carbon dioxide. *Journal* of Applied Physiology, 40(5), 819–823. https://doi.org/10.1152/ jappl.1976.40.5.819
- Twort, C. H., Neild, J. E., & Cameron, I. R. (1985). The effect of verapamil and inspired CO2 on the bronchoconstriction provoked by hyperventilation in normal humans. *Clinical Science*, 69(3), 361– 364. https://doi.org/10.1042/cs0690361

- van den Elshout, F. J., van Herwaarden, C. L., & Folgering, H. T. (1991). Effects of hypercapnia and hypocapnia on respiratory resistance in normal and asthmatic subjects. *Thorax*, 46(1), 28–32. https://doi. org/10.1136/thx.46.1.28
- Verbanck, S., Schuermans, D., Van Muylem, A., Melot, C., Noppen, M., Vincken, W., & Paiva, M. (1998). Conductive and acinar lung-zone contributions to ventilation inhomogeneity in COPD. *American Journal of Respiratory and Critical Care Medicine*, 157(5), 1573– 1577. https://doi.org/10.1164/ajrccm.157.5.9710042
- Verbanck, S., Schuermans, D., Van Muylem, A., Paiva, M., Noppen, M., & Vincken, W. (1997). Ventilation distribution during histamine provocation. *Journal of Applied Physiology*, 83(6), 1907–1916. https://doi.org/10.1152/jappl.1997.83.6.1907
- Wagner, P. D., Dantzker, D. R., Dueck, R., Clausen, J. L., & West, J. B. (1977). Ventilation-perfusion inequality in chronic obstructive pulmonary disease. *Journal of Clinical Investigation*, 59(2), 203–216. https://doi.org/10.1172/JCI108630
- Wagner, P. D., Hedenstierna, G., Bylin, G., & Lagerstrand, L. (1987). Reproducibility of the multiple inert gas elimination technique. *Journal of Applied Physiology*, 62(4), 1740–1746. https://doi. org/10.1152/jappl.1987.62.4.1740
- Wagner, P. D., Laravuso, R. B., Uhl, R. R., & West, J. B. (1974). Continuous distributions of ventilation-perfusion ratios in normal

subjects breathing air and 100 per cent O2. *Journal of Clinical Investigation*, 54, 54-68.

- Wagner, P. D., Naumann, P. F., & Laravuso, R. B. (1974). Simultaneous measurement of eight foreign gases in blood by gas chromatography. *Journal of Applied Physiology*, 36(5), 600–605. https://doi. org/10.1152/jappl.1974.36.5.600
- Wagner, P. D., Saltzman, H. A., & West, J. B. (1974). Measurement of continuous distributions of ventilation-perfusion ratios: Theory. *Journal of Applied Physiology*, 36, 588–599.
- Yung, G. L., Fedullo, P. F., Kinninger, K., Johnson, W., & Channick, R. N. (2004). *Congestive Heart Failure*, 10(s2), 7–10. https://doi. org/10.1111/j.1527-5299.2004.03406.x

How to cite this article: Elliott AR, Kizhakke Puliyakote AS, Tedjasaputra V, et al. Ventilation– perfusion heterogeneity measured by the multiple inert gas elimination technique is minimally affected by intermittent breathing of 100% O₂. *Physiol Rep.* 2020;8:e14488. https://doi.org/10.14814/phy2.14488