UCSF UC San Francisco Previously Published Works

Title

1-BENZYLSPIRO[PIPERIDINE-4,1'-PYRIDO[3,4-b]indole] 'co-potentiators' for minimal function CFTR mutants.

Permalink

<https://escholarship.org/uc/item/2jt1m0z8>

Authors

Son, Jung-Ho Phuan, Puay-Wah Zhu, Jie S [et al.](https://escholarship.org/uc/item/2jt1m0z8#author)

Publication Date

2021

DOI

10.1016/j.ejmech.2020.112888

Peer reviewed

HHS Public Access

Author manuscript Eur J Med Chem. Author manuscript; available in PMC 2022 January 01.

Published in final edited form as:

Eur J Med Chem. 2021 January 01; 209: 112888. doi:10.1016/j.ejmech.2020.112888.

1-BENZYLSPIRO[PIPERIDINE-4,1'-PYRIDO[3,4-B]INDOLE] 'CO-POTENTIATORS' FOR MINIMAL FUNCTION CFTR MUTANTS

Jung-Ho Son†,1, **Puay-Wah Phuan**‡,1, **Jie S. Zhu**†,2, **Elena Lipman**†, **Amy Cheung**†, **Ka Yi Tsui**†, **Dean J. Tantillo**†, **Alan S. Verkman**‡, **Peter M. Haggie**‡,* , **Mark J. Kurth**†,*

†Department of Chemistry, University of California, Davis, CA, 95616, USA

‡Departments of Medicine & Physiology, University of California San Francisco, CA 94143, USA

Abstract

We previously identified a spiro[piperidine-4,1-pyrido[3,4-b]indole] class of co-potentiators that function in synergy with existing CFTR potentiators such as VX-770 or GLGP1837 to restore channel activity of a defined subset of minimal function cystic fibrosis transmembrane conductance regulator (CFTR) mutants. Here, structure-activity studies were conducted to improve their potency over the previously identified compound, **20** (originally termed CP-A01). Targeted synthesis of 37 spiro[piperidine-4,1-pyrido[3,4-b]indoles] was generally accomplished using versatile two or three step reaction protocols with each step having high efficiency. Structureactivity relationship studies established that analog **2i**, with 6'-methoxyindole and 2,4,5 trifluorobenzyl substituents, had the greatest potency for activation of N1303K-CFTR, with EC_{50} ~600 nM representing an ~17-fold improvement over the original compound identified in a small molecule screen.

Graphical Abstract

^{*}Corresponding Authors: Mark J. Kurth, 1 Shields Avenue, University of California, Davis, CA 95616, USA, Phone: (530) 554-2145, Fax: (530) 752-8995, mjkurth@ucdavis.edu, and, Peter M. Haggie, 513 Parnassus Ave, University of California, San Francisco, CA 94143-0521, USA, Phone: (415) 476-8530, Fax: (415) 665-3847, peter.haggie@ucsf.edu. AUTHOR CONTRIBUTIONS:

The manuscript was written by J.-H.S., P.-W.P., J.S.Z., K.Y.T., D.J.T., A.S.V., P.M.H. and M.J.K. Synthesis of substrates was performed by J.-H.S., J.S.Z., A.P.T, and C.K.K. Synthetic work was performed by J.-H.S., J.S.Z., E.L., and A.C. Computational studies were performed by K.Y.T. Biological data collection were performed by P.-W.P. All authors have given approval to the final version of the manuscript. 1The authors contributed equally to the study

²Current Affiliation: Department of Chemistry, Stanford University, 337 Campus Drive, Standford, CA 94305

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Supporting Information: ¹H and ¹³C NMR data (PDF) for all assayed compounds, molecular formula strings (CSV), mol2 files of all energetically relevant conformers, Boltzmann distribution analyses of 2a, 2c, 2e, 4a-4c, and 6a-6b, and the computed H^1 coupling constants (*J*) for **6a** and **6b**. The Supporting Information is available free of charge on the ACS Publications website at DOI: ####.

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Keywords

Cystic Fibrosis; CFTR; Modulator; Potentiator; N1303K-CFTR; c.3700A>G

INTRODUCTION:

Cystic fibrosis transmembrane conductance regulator (CFTR) modulator therapy to restore defective folding and channel gating of CFTR mutants has been a major recent focus in cystic fibrosis (CF) drug development. CFTR modulators include 'potentiators', which restore channel gating, and 'correctors', which rescue misfolded CFTR to improve cell surface presentation [1]. Recently, a 'triple combination' therapy containing two correctors (tezacaftor / VX-661 and elexacaftor / VX-445) and ivacaftor (VX-770) was approved for CF subjects with at least one F508del-CFTR mutation [2–4]. With available potentiators and correctors it is anticipated that up to 90 % of CF subjects may have an efficacious CFTR modulator therapy [5].

There remains an unmet need to develop therapeutics for the remaining \sim 10% of CF subjects [5, 6]. We have described a novel approach that involves two potentiators, with the second potentiator called a 'co-potentiator', that function in synergy to increase chloride conductance for CFTR mutations found mainly in nucleotide binding domain 2 [7–9]. Recent high-throughput screening in cells expressing N1303K-CFTR identified four novel co-potentiator scaffolds, including spiro[piperidine-4,1-pyrido[3,4-b]indoles] and pyrazoloquinolines, some having sub-micromolar potency for activation of N1303K-CFTR when used together with VX-770 [9]. Herein, synthetic chemistry and structure-activity relationship studies were done to advance the spiro[piperidine-4,1-pyrido[3,4-b]indole] class of co-potentiators.

RESULTS:

Chemistry: General synthesis of spiro[piperidine-4,1-pyrido[3,4-b]indoles].

A synthetic scheme to enable investigation of co-potentiator structure-activity relationships (target analogs **1**-**6**) was devised as outlined in Scheme #1 for the efficient synthesis and

modification of the spiro[piperidine-4,1'-pyrido[3,4-b]indole] scaffold (**9**). Adapting technology developed by Mokrosz [10], commercial tryptamines (**8**) were condensed with N -alkylated (11 → 12; R²-X in CH₂Cl₂ + K₂CO₃) [11, 12] piperidin-4-ones (12 where n = 1) to directly deliver the targeted spiro[piperidine-4,1'-pyrido[3,4-b]indole] **9** by a classical Pictet–Spengler reaction [13, 14]. However, when preparing R^2 analogs of 9, it proved more efficient to prepare unalkylated analog **10** [acetic acid-mediated condensation of tryptamine **8** with piperidin-4-one (11; $n = 1$)] and then subsequently perform selective $N¹$ -alkylation of **10** (\mathbb{R}^2 -X in $\text{CH}_2\text{Cl}_2 + \text{K}_2\text{CO}_3$) to give **9**. While the tryptamines employed in this chemistry were commercially available, these starting materials can be readily prepared in two steps by the Büchi/Mak method [15, 16] — 1*H*-indole nitro-olefination followed by exhaustive reduction with LiAlH₄ ($7 \rightarrow 8$).

Figure 1 shows the synthesized analogs of the original hit spiro[piperidine-4,1'-pyrido[3,4^b]indole] **20** that were prepared in six rounds of synthetic study (synthesis rounds **r1** through **r6**). The co-potentiator activities (tested in an N1303K-CFTR expressing FRT cell model) of the synthesized analogs of 20 (10 μ M/V_{MAX} 100%) are summarized at the end of the chemistry description in Table 3, with further description of biological activity provided below (in the *Biology: Characterization of spiro[piperidine-4,1-pyrido[3,4-b]indoles] co*potentiators section).

The synthetic work began by preparing aryl analogs **1a-j** (synthesis round **r1**) to investigate the benzylic aryl moiety by appropriately $N¹$ -alkylating spiro[piperidine-4,1'-pyrido[3,4b]indole] **10**. Of these first ten compounds, nine $(1b-j)$ had a better EC_{50} than **20** and two (**1c** and **1j**) showed improved V_{max} (Table 3). The EC₅₀ and V_{MAX} of the N^1 -3-chloro-2,4difluorobenzyl analog **1j** (4.5 μM/120%) motivated us to next prepare a series of fluorobenzyl-analogs (**2a-i**) to examine the influence of fluoro-substitution patterns on copotentiator activity. These round $r2$ targets (Figure 1) were again prepared by selectively N^1 alkylation of spiro[piperidine-4,1'-pyrido[3,4-b]indole] **10**. Increasing the number of fluorine atoms on the benzyl moiety (e.g., **2ac**/**2f**/**2h** with two fluorines versus **2d**/**2g**/**2i** with three and **2e** with five fluorines) improved the activity of these spirocycles. It was also noted that the positioning of the fluorine atoms on the benzyl moiety was important for maintaining both EC_{50} and V_{MAX} (Table 3: for example, compare 2a versus 2b and 2c).

With results from synthetic rounds **r1** and **r2** in hand, the next objective (synthesis round **r3**, Figure 2) was to more broadly probe the requisite features of the $N¹$ -R³-substituent by preparation of analogs **3a-h**. Notably, deletion of R^3 (i.e., **3a** where $R^3 = H$) gave an inactive compound and lengthening the N^1 —aryl tether $[-N(CH_2)nR^3$ from $n = 1$ (**1a**) to $n = 2$ (**3c**) or $n = 3$ (3d)] greatly reduced activity. Similarly, adding a methyl substituent to the CH₂ tether in **1a** (12.6 μM/V_{MAX} 54%), giving 3b, also reduced activity (3b: 20 μM/V_{MAX} 100%). Replacing the phenyl moiety of **1a** with a furan ring (3e: 16.4 μM/V_{MAX} 67%) reduced activity, while replacing the phenyl moiety of **1a** with a pyridyl ring to electronically mimic the perfluorophenyl moiety of compound **2e** was better tolerated with the nitrogen at position two $(3f: 11.5 \mu M/V_{MAX} 107\%)$, but poorly tolerated with the nitrogen at position three (3g: 17 μM/V_{MAX} 56%).

Having thoroughly investigated the N1-substituent in these spiro[piperidine-4,1'-pyrido[3,4 ^b]indoles], attention was next turned to a probe of the bisheterocyclic spiropiperidine moiety – synthesis round **r4** (Figure 1) targeting analogs **4a-d**. This work began with the preparation of spiroazepane, spiropyrrolidine, and spiropiperidine analogs where the piperidine ring of **2a** was transformed into an azepine (**4a**), pyrrolidine (**4b**), and N-offset piperidine (**4c**) rings. These three analogs of **2a** were prepared by reaction of 2-(5-methoxy-1H-indol-3 yl)ethan-1-amine (**8** where R^1 = methoxy at C5) with the appropriate ketoamine (12), directly giving the targeted **2a** analogs **4a-c**. Interestingly, all three of these analogs were inactive. We next prepared piperidine ring-opened analogs truncated analog **4d**.

Conformational analysis was done to investigate possible mechanisms to explain the differences between active compounds **2a**, **2c** and **2e**, and inactive compounds **4a**-**4c**. We began with conformational searching using Spartan10 (Wavefunction Inc., Irvine, CA). For the more flexible structures **4a** and **4c**, multiple runs of conformational search were performed and the results pooled. The conformational search runs were systematic and used the Merck Molecular Force Field (MMFF). All resulting conformers were subjected to single point energy calculations using PCM(chloroform)-B3LYP/6–31+G(d,p) [17–20]. Geometry optimizations were then performed on all conformers within 4 kcal/mol of the conformer with the lowest electronic energy, with the D3(BJ) dispersion correction [21, 22] included. All quantum chemical calculations were performed using $Gaussian16$ (Gaussian Inc., Wallingford, CT). An implicit chloroform solvent was chosen for these calculations to simulate the polarity inside a general protein binding site [23]. Since **2a**, **2c**, **2e**, **4a**-**4c** contain fluorine atoms, the $6-31+G(d,p)$ basis set might not be sufficient for computing accurate energies; consequently, we also calculated energies for relevant conformers with PCM-(chloroform)-B3LYP-D3(BJ)/6–311+G(2d,2p). B3LYP/6–311+G(2d,2p) has been reported as a standard method for use with high electron affinity atoms, i.e., O and F [24].

Our conformational analysis suggested a trend distinguishing between active **2a**, **2c**, and **2e**, and inactive **4a**-**4c**. Figure 2 shows the optimized geometries of the lowest energy conformers B3LYP/6–31+G(d,p) for each structure. Most of the energetically relevant conformers of the active co-potentiators (i.e., $2a$, $2c$, and $2e$) have their benzene ring (*Ring i*) exposed for interaction, which we define as the "Open" form (Figure 2). Assuming that the CFTR binding site accommodates the Open form better and has residues in the vicinity to interact with the phenyl group of the co-potentiators, the Open conformers would allow for various interactions between the π -system and the CFTR interior. In contrast, most of the energetically relevant conformers for **4a**-**4c** have one face of Ring i interacting with the rest of the molecule, precluding protein interactions with it; we refer to these conformers as "Closed" (Figure 2).

Figure 3 shows the superposition of all optimized conformers within 4 kcal/mol of the lowest energy conformer for **2a**, **2c**, **2e**, and **4a**-**4c**. We aligned the conformers and calculated their RMSD values using an RMSD calculating tool from VMD [25]. In this case, we considered conformers to be unique if they have an RMSD value greater than 0.05 relative to other conformers. We then calculated the Boltzmann weighted averages for the Open and Closed conformers at room temperature. This allows us to determine the population of Open vs. Closed conformers for each structure, assuming all are in equilibrium

[26]. Table 1 summarizes the Boltzmann weighted averages for **2a**, **2c**, **2e**, and **4a**-**4c**. The results of this analysis emphasize the difference in conformational preferences for active and inactive structures

We also considered that these structures contain amines, which can be protonated in biological environments. Therefore, we performed conformational analysis on various protonated states for each structure as well. However, there were no consistent differences in conformational preferences for active and inactive structures. This analysis is complicated by the ability for amines to hydrogen bond with each other within each molecule (see Supporting Information for details), which may or may not occur in biologically relevant environs.

With exploration of the spiropiperidinepyrido aspects of the spiro[piperidine-4,1'-pyrido[3,4-b]indole] scaffold in hand with synthesis rounds **r1-r4**, we next turned to synthesis round **r5** where the indole moiety of scaffold **9** was probed with analogs **5a-e**. Spiro[piperidine-4,1'-pyrido[3,4-b]indoles] **5a-d** were prepared by reacting the appropriate tryptamine **8** with 1-(2,4-difluorobenzyl)piperidin-4-one (12 in Scheme #1 where $n = 1$ and $R^2 = 2.4$ -difluorobenzyl). Whereas **5b** performed in a comparable manner to 20, **5a** (8.2 μ M/ 100%) showed mild improvement, and **5c** (2.5 μM/89%) and **5d** (2.4 μM/100%) were approximately 4–fold more potent. Finally, we prepared indole N-methylated analog **5e** by employing 2-(5-methoxy-1-methyl-1H-indol-3-yl)ethan-1-amine in place of 2-(5methoxy-1H-indol-3-yl)ethan-1-amine in a reaction with 1-(2,4-difluorobenzyl)piperidin-4 one $(8 \rightarrow 9$ in Scheme #1). Compound 5e, however, was inactive In light of the structureactivity insights gained with synthetic rounds **r1-r5**, one final synthesis round (**r6**) was undertaken where targeting constrained spiropiperidinepyrido analog **14** became a high priority in an attempt to address the question of whether there is a conformational requirement for activity vis-à-vis the $N¹$ -(benzyl) moiety. To address this issue, we set out to prepare **14**, a constrained analog of compound **2f**. C-Alkylation of diethyl acetamidomalonate with 4-(bromomethyl)-1,2-difluorobenzene followed by ester hydrolysis/decarboxylation (Scheme #2) set the stage for a subsequent Pictet–Spengler reaction to delivered tetrahydroisoquinoline-3-carboxylic acid **13** [27]. Unfortunately, all attempts at the Pictet-Spengler failed – an apparent consequence of the electron deficient nature of the difluorophenyl ring. In light of this outcome, our constrained analog study was modified to instead target constrained analog **6** (the nor-fluoro analog of **14**). Commercially available 3-amino-4-phenylbutanoic acid (**15**) successfully participated in a Pictet–Spengler reaction to give **16** and subsequent esterification delivered methyl 2-(1,2,3,4 tetrahydroisoquinolin-3-yl)acetate (**17**). Michael addition to methyl acrylate delivered bisester intermediate **18**, which then underwent Dieckmann cyclization/decarboxylation to give ketone **19**. A second Pictet–Spengler reaction [28] of 2-(5-methoxy-1H-indol-3-yl)ethan-1 amine (**8** where R^1 = methoxy at C5) with 19 gave the spiro[piperidine-4,1'-pyrido[3,4b]indole] **6b** as the sole product – i.e., spiro[piperidine-4,1'-pyrido[3,4-b]indole] **6a** was not obtained.

To confirm the product is indeed **6b**, we performed quantum chemical ¹H and ¹³C NMR calculations on both **6a** and **6b** (Figure 4). Both **6a** and **6b** were optimized using

 $SMD(chloroform) - B3LYP-D3(BJ)/(6-31+G(d,p) [21, 22, 29]$. Using the gauge-including atomic orbital (GIAO) [30] method, chemical shift calculations were performed with SMD(chloroform)-mPW1PW91/6–311+G(2d,p) [31] on conformers that are within 4 kcal/mol relative to the lowest energy conformer. The chemical shifts of these energetically relevant conformers were then averaged using Boltzmann distributions. This is a common procedure for computational NMR studies [30, 32, 33]. Linear scaling (using slope= -1.0533 , intercept= 186.524) and (slope= -1.0936 , intercept= 31.802) [\(cheshirenmr.info\)](http://cheshirenmr.info) was applied to computed isotropic shieldings to arrive at the ${}^{13}C$ and ${}^{1}H$ shifts, respectively. Due to the similarity in the calculated ${}^{1}H$ shifts, coupling constants (Hz) were also calculated using the same methods to confirm the identity of some diagnostic ${}^{1}H$ peaks (see SI for detail).

Table 2 shows the calculated chemical shifts for **6a** and **6b** and their deviations from the experimental shifts. Deviations within 6 ppm for ¹³C and 0.3 ppm for ¹H shifts are considered acceptable [30, 32, 33]. Although the 13C chemical shifts for **6a** and **6b** are quite similar, the computed chemical shift at position C13 of **6a** deviates greatly from the experimental value, whereas all ¹³C shifts for **6b** are within the acceptable deviation range. Similarly, the deviations between the calculated 1H shifts for **6b** and the experimental shifts are within the accepted range, but the calculated chemical shifts at positions H15, H16, and H18 for **6a** have deviations greater than 0.3 ppm from the experimental shifts. Both ¹³C and ¹H chemical shifts for **6b** have lower mean absolute deviations (MAD) compared to those for **6a** as well. We also compared the free energies of the lowest energy conformers of **6a** and **6b**; that for **6a** is ~2.3 kcal/mol higher than that for **6b** (Figure 4), consistent with formation of the thermodynamic product during synthesis.

The lowest energy conformer of **6b** resembles more closely the Closed conformers described above, but that of **6a** resembles the Open, i.e., where Ring i is available for interactions (Figure 5). Considering all conformers that are within 4 kcal/mol of the lowest energy conformers, Boltzmann distributions of open and closed forms of **6a** and **6b** were calculated (Figure 5). For **6a**, ~99.3% of conformers are predicted to be Open, while only 23.2% of conformers are predicted to be Open for **6b**. Therefore, we expect **6b** to have low activity at best, if our hypothesis that conformational bias is related to activity is correct. As predicted, **6b** was inactive when tested experimentally.

The structural determinants for activity of compound classes **1**-**6** are summarized in Figure 6. On the indole aryl 6-member ring $(R¹)$, 6'-methoxy $(2i)$ was most potent, whereas substituents such as methoxy at 7' position (**5d**) showed decreased activity. Replacement of 6'-methoxy with bromo (**5b**) led to an inactive compound, although a chloro (**5c**) substitution was tolerated. At the indole nitrogen position (R^2) , hydrogen $(2i)$ was required as methylation (**5e**) led to inactivity. For the benzyl moiety on the terminal piperidine (\mathbb{R}^3) , we found 2,4,5-trifluorobenzyl (**2i**) to be best, di-halo-substituted moieties such as 2,4 dichlorobenzyl (**1d**) and 3-chloro-5-fluorobenzyl (**1e**) showed decreased activity, and methylene-linked heterocycles such as 2-furyl (**3e**), 2-pyridyl (**3f**), or 3-pyridyl (**3g**) were active, but less potent. Likewise, at \mathbb{R}^3 , lack of substitution on the terminal piperidine (3a), or compounds where the linked ring system is extended by longer linkers (**3c** and **3d**) were

also inactive. Finally, in terms of the bis-heterocyclic spiro-fused ring system, a piperidine ring (as in **2a**, **2i** and other compounds) was significantly more potent than an azepine (**4a**), and a pyrrolidine (**4b**), N-offset piperidine (**4c**) or ring-opened piperidine (**4d**) abolished activity.

Additional biological studies were performed for most potent compounds **2i** and **2e**. Copotentiator efficacy was initially determined by short-circuit current measurements in FRT cells expressing N1303K-CFTR in the presence of a transepithelial chloride gradient and with permeabilization of the basolateral cell membrane such that measured current directly reports CFTR channel activity. Representative data in Figure 7A show small increases in CFTR activity following addition of the cAMP agonist forskolin and the potentiator VX-770, followed by concentration-dependent increases in current following addition of the co-potentiators **2i** (*left*) and **2e** (*middle*), with EC₅₀ values 0.6 ± 0.2 and 2.1 ± 0.2 0.3 μM, respectively (*right*). We previously defined potentiators as either class I compounds, including VX-770 and GLPG1837 [34], or class II compounds such as the arylsulphonamidepyrrolopyridine and spiro[piperidine-4,1-pyrido[3,4-b]indole] copotentiators that probably bind at distinct sites on CFTR [9]. Indeed, structural and pharmacological studies have provided evidence that VX-770 and GLPG1837 bind to the same site on CFTR [35, 36]. Thus, as expected from our prior studies [9], N1303KCFTRexpressing FRT cells treated with forskolin and GLPG1837 had similar **2i** co-potentiator EC₅₀ of 0.8 ± 0.2 μ M (Fig. 7B).

Compound **2i** was also tested on a second minimal function CFTR mutation, I1234del-CFTR, which is generated by the c.3700A>G mutation that results in deletion of 6 amino acids from the CFTR polypeptide (p.Ile1234_Arg1239del-CFTR, hereafter termed I1234del-CFTR) due to introduction of a cryptic splice site in the CFTR transcript [37, 38]. As seen in Figure 7C, **2i** further activates I1234del-CFTR following forskolin and VX-770 to increase channel activity with EC_{50} of 0.7 ± 0.2 µM, similar to that found with N1303K-CFTR. As expected, the R347P-CFTR mutant, which is mildly responsive to VX-770 but not responsive to arylsulphonamidepyrrolopyridine co-potentiators [9], was not activated by **2i** indicating that alternative mechanisms (such as increased cAMP signaling) are not responsible for activation of the N1303K- and I1234del-CFTR mutants (Fig. 7D).

The efficacy of **2i** was also tested in 16HBE14o- human bronchial epithelial cell models in which the endogenous CFTR gene was edited to contain the N1303K mutation (16HBEge-N1303K) or the I1234del mutation (16HBEge-I1234del) [38, 39]. As shown in N1303K- (Fig. 8A) and I1234del-CFTR (Fig. 8B) expressing 16HBE14o- cells, addition of forskolin and then VX-770 produced limited channel activity. However, subsequent addition of **2i** produced CFTR_{inh}-172-inhibitable responses of $16.1 \pm 0.4 \mu A/cm^2$ in N1303K- (Fig. 8A) and $6.1 \pm 0.1 \mu A/cm^2$ in I1234del-CFTR (Fig. 8B) expressing cells, approximately 6-fold and 2-fold greater that that produced by VX-770 alone.

CONCLUSIONS:

This study extends our prior work that identified a novel class of CFTR modulators, termed co-potentiators, that act in synergy with existing potentiators such as VX-770 and

GLPG1837 to increase chloride channel function of CFTR mutants [7–9]. The objective of this study was to optimize spiro[piperidine-4,1-pyrido[3,4-b]indoles] co-potentiators previously identified in a high-throughput screen [9]. A straightforward, high yielding synthetic strategy was developed to investigate structure-activity relationships. In total, 37 spiro[piperidine-4,1-pyrido[3,4-b]indoles] were synthesized to investigate the consequences of altering the indole and benzyl moieties. Analogs of the original spiro[piperidine-4,1' pyrido[3,4-b]indole] compound **20** were prepared in six rounds of synthetic studies (synthesis rounds **r1** through **r6**) starting from commercial tryptamines (**8**). These were condensed with N-alkylated piperidin-4-ones (**12**) to directly deliver the targeted spiro[piperidine-4,1'-pyrido[3,4-b]indole] **9** by a classical Pictet–Spengler reaction. When preparing R² analogs of **9**, it proved more efficient to prepare unalkylated analog **10** [acetic acid-mediated condensation of tryptamine **8** with piperidin-4-one (**11**)] and then subsequently perform selective N^1 -alkylation of **10** (\mathbb{R}^2 -X in $\text{CH}_2\text{Cl}_2 + \text{K}_2\text{CO}_3$) to give **9**.

It is estimated that up to 90% of CF subjects will benefit from available CFTR modulators including Trikafta [5], with \sim 10% of CF subjects not benefitted. For co-potentiators, we have demonstrated efficacy for several NBD2 mutants including the missense mutant N1303K, the c.3700A>G splicing mutant, and truncated CFTR protein generated by the premature termination codon (PTC) W1282X [7–9]. In addition, albeit to a lesser degree, channel activity of the VX-770-responsive G551D mutation can be further augmented by co-potentiators [7]. Co-potentiators may be beneficial for some PTCs if sufficient transcript is present, or in combination with future read-through or other therapies [40].

In summary, spiro[piperidine-4,1-pyrido[3,4-b]indoles] represent an evolvable CFTR copotentiator scaffold with nanomolar potency that in synergy with existing potentiators activates certain CF-causing CFTR mutations. The expansion of spiro[piperidine-4,1 pyrido[3,4-b]indole] structure activity relationships may yield insight into the binding site of co-potentiators through computational or structural approaches [35, 36], as well as subsequent rational compound optimization.

MATERIALS AND METHODS:

General Experimental:

All compounds described in this manuscript have 95% purity. The analytical method used to determine purity was 1H NMR (see the accompanying Supporting Information file which provides the 1H- and 13C-NMR for the thirty-seven compounds assayed) and HPLC/HRMS. For HRMS analysis, samples were analyzed by flow-injection analysis into a Thermo Fisher Scientific LTQ Orbitrap (San Jose, CA) operated in the centroided mode. Samples were injected into a mixture of 50% MeOH/H₂O and 0.1% formic acid at a flow of 0.2 mL/min. Source parameters were 5.5kV spray voltage, capillary temperature of 275 °C and sheath gas setting of 20. Spectral data were acquired at a resolution setting of 100,000 FWHM with the lockmass feature, which typically results in a mass accuracy <2ppm.

Cell culture:

Fischer Rat Thyroid (FRT) cells transfected to stably express N1303K-CFTR, I1234del-CFTR and R347P-CFTR were cultured as described [8, 9]. Gene-edited 16HBE14o-cells expressing N1303K-CFTR were provided by CFFT lab and were cultured as described [39]. Gene-edited 16HBE14o- cells expressing I1234del-CFTR were a generous gift of Dr. Christine Bear (The Hospital for Sick Children, Toronto, Canada) and were cultured as described [38].

Short-circuit current measurement.

Short-circuit current measurements were made essentially as described in prior studies [8, 9]. In brief, measurements were made using a DVC-1000 voltage clamp (World Precision Instruments Inc., Sarasota, FL), cells were cultured on Snapwell clear permeable supports (Corning), and experiments were performed using HCO_3^- buffered solutions at 37 °C. For studies using FRT cells the basolateral membrane was permeabilized with 250 μg/ml amphotericin B and experiments were done with a chloride gradient. For 16HBE14o- geneedited cells experiments were done with a chloride gradient using Hepes-buffered solutions.

Statistical analysis.

Data are presented as mean \pm S.E.M. Comparisons between two groups were performed using the unpaired Student's t-test. $P < 0.05$ was considered as statistically significant.

General experimental for the synthesis of 2',3',4',9'-tetrahydrospiro[piperidine-4,1' pyrido[3,4-b]indoles] (8 → **9):**

Tryptamine (**8**, 0.6 mmol) was mixed with glacial acetic acid (3 mL) in a 10 mL vial and stirred with a magnetic stirrer. Corresponding ketone (**12**, 0.5 mmol) was added and the vial was sealed with a plastic cap. The vial was heated at 100 °C for 16 h in an oil bath. After cooling, the solution was diluted with water $(\sim 20 \text{ mL})$ and neutralized by adding 4 M HCl. The product was extracted with dichloromethane, and the organic solution was washed with water, brine and dried over magnesium sulfate. The solvent was removed in vacuo, and the product was purified by using flash column chromatography (2.5 % MeOH/DCM).

General experimental for the synthesis of N-alkylated piperidin-4-one analogs (11 → **12):**

4-Piperidone hydrochloride (**11**, 5 mmol) was mixed with 25 mL of dichloromethane in an Erlenmeyer flask. Small amount of methanol (5 drops) was added and benzyl bromide (2.5 mmol) and potassium carbonate (5 mmol) was added. The mixture was stirred at RT for 16h. Water was added to the reaction mixture and the product was extracted with dichloromethane. The extracted organic solution was washed with water, brine, and dried over magnesium sulfate. The solvent was removed in vacuo, and the product was purified by using flash column chromatography (2.5 % MeOH/DCM).

General experimental for the synthesis of 2',3',4',9'-tetrahydrospiro[piperidine-4,1' pyrido[3,4-b]indoles] (10 → **9):**

Compound **3a** (0.5 mmol) was mixed with 5 mL of dichloromethane in a small vial. Small amount of methanol (1 drop) was added for better solubility, and benzyl bromide (0.5 mmol)

and potassium carbonate (1.5 mmol) was added. The vial was capped, and the mixture was stirred at RT for 16h. Water was added to the reaction mixture and the product was extracted with dichloromethane. The extracted organic solution was washed with water, brine, and dried over magnesium sulfate. The solvent was removed in vacuo, and the product was purified by using flash column chromatography (2.5 % MeOH/DCM).

General experimental for the synthesis of 2',3',4',9'-tetrahydrospiro[piperidine-4,1' pyrido[3,4-b]indoles] (8 → **10):**

5-methoxytryptamine (**8**, 25 mmol) was mixed with glacial acetic acid (20 mL) in a round bottom flask. 4-piperidone hydrochloride (**11**, 25 mmol) was added and the solution was heated at 100 °C for 16 h in an oil bath with stirring. The solution was cooled to RT and diluted with water (100 mL) and neutralized with 4 M NaOH. Tan precipitate formed upon standing within an hour. The precipitate was filtered and washed with water and air dried. Yield $= 54\%$.

General experimental for the synthesis of tryptamines $(7 \rightarrow 8)$:

1-Dimethylamino-2-nitroethylene (3 mmol) was mixed in TFA (4 mL) in a vial. Substituted indole (**7**, 3.6 mmol) was dissolved in dichloromethane (3 mL) separately and added. The mixture was stirred at RT for 2 h and the solution was diluted with dichloromethane. The organic solution was washed with water, brine and dried over magnesium sulfate. Solvent was removed in vacuo and purified by using flash column chromatography (50 % EtOAc/ hexane). LiAlH₄ (12 mmol) was mixed with THF (75 mL) and cooled at −78 °C and stirred. The product from the previous step dissolved in small amount of THF was added dropwise and the mixture was stirred overnight with slowly warming to RT. The reaction mixture was cooled in an ice bath and quenched with water and 4 M NaOH solution. Organic solvent was removed in vacuo and the product was extracted with dichloromethane. The extracted solution was washed with water, brine, and dried over magnesium sulfate. The solvent was removed in vacuo, and the product was directly used.

Synthesis of 2-(1,2,3,4-tetrahydroisoquinolin-3-yl)acetic acid (16):

L-β-homophenylalanine hydrochloride (1 g, 4.6 mmol) was mixed with concentrated HCl (10 mL) in a round bottom flask and formaldehyde (37% aqueous solution, 4 mL, 49 mmol) was added. The mixture was refluxed at 100 °C for 3h with stirring. After cooling, solvent was removed in vacuo to obtain off-white solid in quantitative yield, which was used for the next step without purification.

Synthesis of methyl 2-(1,2,3,4-tetrahydroisoquinolin-3-yl)acetate (17):

16 (1g, 4.3 mmol) was placed in a round bottom flask. Trimethylchlorosilane 1.26 mL was added dropwise with stirring, then methanol (20 mL) was added and stirred for 2 h. Solvent was removed in vacuo and dried in vacuum. The solid product was used for the next step without purification.

17 (0.62g, 3 mmol) was dissolved in dichloromethane (30 mL) and trifluoroacetic acid (3 mL) was added and stirred briefly. Solvent was removed in vacuo and the resulting oil was extracted with ethyl acetate. The organic solution was washed with saturated sodium bicarbonate solution, and the solvent was removed in vacuo. To the resulting oil, methyl acrylate (10 mL) was added and refluxed at 80 °C overnight. Solvent was removed in vacuo, and the product was used without purification for the next step.

Synthesis of 1,3,4,6,11,11a-hexahydro-2H-pyrido[1,2-b]isoquinolin-2-one (19):

Lithium diisopropylamide (LDA, 1M solution in THF/hexane, 10 mL) was cooled at −78 °C under N_2 atmosphere. Crude product of **18** was dissolved in anhydrous THF (8 mL) under $N₂$ atmosphere and added to the LDA solution by syringe. The mixture solution was stirred at −78 °C for 1 h. Concentrated HCl (0.8 mL) was added to the solution and brought to RT. Solvent was removed in vacuo then H_2O (25 mL) and concentrated HCl (25 mL) was added, and the solution was refluxed at 100 °C overnight. After cooling, the solution was basified by adding solid K_2CO_3 portion wise. The product was extracted with diethyl ether and the organic layer was washed with water, brine and dried with magnesium sulfate. Crude product (220 mg) was used for the next step without purification.

Synthesis of $(2R,11aS)$ -6'-methoxy-1,2',3,3',4,4',6,9',11,11a-decahydrospiro[pyrido][1,2^b]isoquinoline-2,1'-pyrido[3,4-b]indole] (6b): Crude 19 (220 mg, 1.1 mmol) was placed in a glass vial and 5-methoxytryptamine (250 mg, 1.3 mmol) was added and glacial acetic acid (3 mL) was added. The vial was capped and heated at 100 °C overnight. After cooling, the solution was diluted with water $(\sim 20 \text{ mL})$ and neutralized by adding 4 M HCl. The product was extracted with dichloromethane, and the organic solution was washed with water, brine and dried over magnesium sulfate. The solvent was removed in vacuo, and the product was purified by using flash column chromatography (2.5 % MeOH/DCM).

1-Benzyl-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4-b]indole] (1a).

Yield = 152 mg (84 %).¹H NMR (400 MHz, CDCl₃) δ 8.30 (d, $J = 35.1$ Hz, 1H), 7.43 (d, $J =$ 7.2 Hz, 2H), $7.39 - 7.27$ (m, 3H), 7.22 (d, $J = 8.7$ Hz, 1H), 6.96 (d, $J = 2.5$ Hz, 1H), 6.82 (dt, $J = 8.9, 1.8$ Hz, 1H), 3.87 (s, 3H), 3.69 (s, 2H), 3.15 (t, $J = 5.6$ Hz, 2H), 2.96 – 2.77 (m, 2H), $2.77 - 2.53$ (m, 4H), 2.25 (td, J = 13.4, 4.4 Hz, 2H), 1.79 (d, J = 13.7 Hz, 2H). ¹³C NMR (101 MHz, CDCl3) δ 153.99, 140.62, 136.72, 130.69, 129.67, 128.46, 127.64, 127.60, 111.62, 111.44, 108.42, 100.50, 63.09, 56.07, 50.52, 48.80, 39.16, 35.86, 23.23. HRMS (ESI) m/z for $C_{23}H_{28}N_3O$ [M+H]. calcd 362.2232, found 362.2229.

1-(2-Fluoro-4-nitrobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (1b).

Yield = 119 mg (56 %).¹H NMR (400 MHz, CDCl₃) δ 7.92 (s, 1H), 7.57 (td, J = 5.2, 2.8 Hz, $2H$), $7.32 - 7.18$ (m, $2H$), 6.95 (d, $J = 2.4$ Hz, 1H), 6.82 (dd, $J = 8.7$, 2.5 Hz, 1H), 3.86 (d, J $= 11.3$ Hz, 5H), 3.14 (t, $J = 5.7$ Hz, 2H), 2.69 (t, $J = 5.6$ Hz, 2H), 2.64 – 2.54 (m, 4H), 2.01 (ddd, $J = 13.8$, 10.3, 6.5 Hz, 2H), 1.79 – 1.71 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ

162.37, 159.88, 154.03, 150.11 (d, $J_{C-F} = 8.4$ Hz), 140.70, 132.64 (d, $J_{C-F} = 7.8$ Hz), 130.61, 130.06 (d, J_{C-F} = 3.7 Hz), 127.72, 119.34 (d, J_{C-F} = 20.7 Hz), 112.21 (d, J_{C-F} = 26.4 Hz), 111.51 (d, J_{C-F} = 7.6 Hz), 108.51, 100.53, 58.88, 56.06, 50.58, 48.83, 39.07, 36.37, 23.15. HRMS (ESI) m/z for $C_{23}H_{26}FN_{4}O_{3}$ [M+H]. calcd 425.1989, found 425.1978.

1-(5-Chloro-2-nitrobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (1c).

Yield = 128 mg (58 %).¹H NMR (400 MHz, CDCl₃) δ 7.90 (s, 1H), 7.83 (d, J = 8.6 Hz, 1H), 7.68 (d, $J = 2.3$ Hz, 1H), 7.39 (dd, $J = 8.6$, 2.3 Hz, 1H), 7.23 (d, $J = 8.7$ Hz, 1H), 6.96 (d, $J =$ 2.4 Hz, 1H), 6.83 (dd, $J = 8.8$, 2.4 Hz, 1H), 3.87 (d, $J = 2.6$ Hz, 5H), 3.16 (t, $J = 5.7$ Hz, 2H), 2.70 (t, $J = 5.7$ Hz, 2H), 2.67 – 2.61 (m, 4H), 2.08 – 1.98 (m, 2H), 1.78 (d, $J = 13.5$ Hz, 2H), 1.29 (s, 1H). 13C NMR (101 MHz, CDCl3) δ 154.05, 147.99, 140.57, 138.85, 136.69, 130.80, 130.62, 127.99, 127.72, 125.99, 111.55, 111.52, 108.55, 100.54, 59.06, 56.07, 50.59, 48.95, 39.08, 36.39, 23.11. HRMS (ESI) m/z for C₂₃H₂₆ClN₄O₃ [M+H]. calcd 441.1693, found 441.1685.

1-(2,4-Dichlorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (1d).

Yield = 156 mg (72 %).¹H NMR (400 MHz, CDCl₃) δ 7.92 (s, 1H), 7.47 (d, J = 8.3 Hz, 1H), 7.41 (d, $J = 2.2$ Hz, 1H), $7.27 - 7.15$ (m, 2H), 6.97 (d, $J = 2.4$ Hz, 1H), 6.83 (dd, $J = 8.7$, 2.4 Hz, 1H), 3.88 (s, 3H), 3.68 (s, 2H), 3.17 (t, $J = 5.6$ Hz, 2H), 2.88 – 2.52 (m, 6H), 2.28 – 2.00 (m, 2H), 1.80 (d, $J = 14.3$ Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 154.05, 140.85, 135.06, 134.79, 133.21, 131.70, 130.57, 129.27, 127.77, 126.95, 111.49, 111.47, 108.59, 100.56, 59.02, 56.08, 50.58, 48.93, 39.12, 36.44, 23.22. HRMS (ESI) m/z for $C_{23}H_{26}Cl_2N_3O$ [M +H]. calcd 430.1453, found 430.1447.

1-(3-Chloro-5-fluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (1e).

Yield = 187 mg (91 %) ¹H NMR (800 MHz, CDCl₃) δ 8.09 (s, 1H), 7.25 – 7.15 (m, 2H), $7.08 - 7.00$ (m, 2H), 6.98 (d, $J = 2.7$ Hz, 1H), 6.84 (dd, $J = 8.7, 2.5$ Hz, 1H), 3.89 (s, 3H), 3.54 (s, 2H), 3.15 (t, $J = 5.7$ Hz, 2H), 2.71 (q, $J = 5.2$ Hz, 4H), 2.61 – 2.46 (m, 2H), 2.10 (td, $J = 13.4$, 4.2 Hz, 2H), 1.83 – 1.69 (m, 2H). ¹³C NMR (201 MHz, CDCl₃) δ 163.35, 162.12, 154.01, 142.79 (d, $J_{\text{C-F}}$ = 7.8 Hz), 140.84, 134.76 (d, $J_{\text{C-F}}$ = 10.4 Hz), 130.65, 127.76, 124.82 (d, $J_{C-F} = 2.9$ Hz), 114.87 (d, $J_{C-F} = 24.9$ Hz), 114.27 (d, $J_{C-F} = 21.4$ Hz), 111.52 (d, $J_{\text{C-F}}$ = 18.3 Hz), 108.56, 100.57, 62.25, 56.07, 50.58, 48.91, 39.09, 36.33, 23.21. HRMS (ESI) m/z for $C_{23}H_{26}CIFN_3O$ [M+H]. calcd 414.1748, found 414.1740.

1-(3-Chloro-2,6-difluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1' pyrido[3,4-b]indole] (1f).

Yield = 183 mg (85 %) ¹H NMR (400 MHz, CDCl₃) δ 8.34 (s, 1H), 7.30 (h, J = 5.4 Hz, 1H), 7.16 (d, $J = 8.8$ Hz, 1H), 6.96 (d, $J = 2.6$ Hz, 1H), 6.87 – 6.72 (m, 2H), 3.88 (s, 3H), 3.76 (s, 2H), 3.13 (t, $J = 5.7$ Hz, 2H), 2.79 (d, $J = 11.4$ Hz, 2H), 2.73 – 2.56 (m, 4H), 2.10 (td, $J =$ 13.3, 4.7 Hz, 2H), 1.77 (d, $J = 12.8$ Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 160.35 (dd, $J_{\text{C-F}} = 248.7, 7.0 \text{ Hz}$), 157.24 (dd, $J_{\text{C-F}} = 250.2, 8.4 \text{ Hz}$), 153.93, 140.88, 130.71, 129.69 (d,

 J_{C-F} = 9.5 Hz), 127.70, 116.55 (dd, J_{C-F} = 19.1, 4.0 Hz), 114.91 (t, J_{C-F} = 20.2 Hz), 111.83 $\left(\text{dd}, J_{\text{C-F}} = 24.6, 4.0 \text{ Hz}\right)$, 111.63, 111.33, 108.38, 100.52, 56.06, 50.44, 49.43, 48.31, 39.10, 36.18, 23.20. HRMS (ESI) m/z for $C_{23}H_{25}CIF_2N_3O$ [M+H]. calcd 432.1654, found 432.1655.

1-(2-Chlorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (1g).

Yield = 126 mg (64 %) ¹H NMR (400 MHz, CDCl₃) δ 8.13 (s, 1H), 7.54 (dd, J = 7.4, 2.3 Hz, 1H), 7.41 (dd, $J = 7.5$, 1.9 Hz, 1H), 7.26 (qd, $J = 7.4$, 6.6, 2.1 Hz, 2H), 7.22 – 7.13 (m, 1H), 6.98 (d, $J = 2.7$ Hz, 1H), 6.84 (dd, $J = 8.8$, 2.6 Hz, 1H), 3.89 (s, 3H), 3.74 (s, 2H), 3.17 $(t, J = 5.7 \text{ Hz}, 2\text{H})$, 2.82 (dt, $J = 12.1$, 3.7 Hz, 2H), 2.74 – 2.58 (m, 4H), 2.14 (td, $J = 13.1$, 4.6 Hz, 2H), 1.80 (dd, $J = 13.9$, 2.4 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 154.00, 140.97, 135.92, 134.57, 131.10, 130.64, 129.58, 128.37, 127.75, 126.67, 111.57, 111.40, 108.45, 100.56, 59.64, 56.10, 50.65, 48.97, 39.16, 36.35, 23.22. HRMS (ESI) m/z for C₂₃H₂₇ClN₃O [M+H]. calcd 396.1843, found 396.1836.

1-(4-Bromo-3-fluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (1h).

Yield = 155 mg (68 %) ¹H NMR (400 MHz, CDCl₃) δ 8.00 (s, 1H), 7.48 (dd, *J* = 8.2, 7.1 Hz, 1H), 7.21 (dd, $J = 9.1$, 2.9 Hz, 2H), 7.07 – 6.93 (m, 2H), 6.85 (dd, $J = 8.7$, 2.5 Hz, 1H), 3.89 (s, 3H), 3.54 (s, 2H), 3.17 (t, $J = 5.7$ Hz, 2H), 2.79 – 2.65 (m, 4H), 2.63 – 2.44 (m, 2H), 2.10 (td, $J = 13.2$, 12.8, 4.4 Hz, 2H), 1.88 – 1.70 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 160.33, 157.87, 154.04, 140.89, 140.84 (d, $J_{C-F} = 6.6$ Hz), 133.17, 130.64, 127.80, 125.78 (d, $J_{C-F} = 3.3$ Hz), 116.90 (d, $J_{C-F} = 22.0$ Hz), 111.51 (d, $J_{C-F} = 4.4$ Hz), 108.62, 107.20 (d, $J_{\text{C-F}}$ = 20.9 Hz), 100.61, 62.20, 56.10, 50.62, 48.92, 39.11, 36.42, 23.23. HRMS (ESI) m/z for $C_{23}H_{26}BrFN_{3}O$ [M+H]. calcd 458.1243, found 458.1239.

1-(4-Bromo-2-fluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (1i).

Yield = 190 mg (83 %) ¹H NMR (400 MHz, CDCl₃) δ 8.59 (s, 1H), 7.24 (t, J = 8.1 Hz, 1H), 7.15 (dd, $J = 7.9$, 2.6 Hz, 3H), 6.98 (d, $J = 2.6$ Hz, 1H), 6.81 (dd, $J = 8.7$, 2.5 Hz, 1H), 3.88 $(s, 3H), 3.60 (s, 2H), 3.14 (t, J = 5.7 Hz, 2H), 2.72 (q, J = 7.2, 5.7 Hz, 4H), 2.59 (td, J = 12.1,$ 2.5 Hz, 2H), 2.10 (td, J = 13.5, 4.6 Hz, 2H), 1.69 (s, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 162.50, 160.00, 153.93, 140.87, 132.96 (d, J_{C-F} = 5.1 Hz), 130.73, 127.72, 127.29 (d, J_{C-F} = 3.7 Hz), 123.77 (d, $J_{C-F} = 15.0$ Hz), 121.33 (d, $J_{C-F} = 9.5$ Hz), 119.03 (d, $J_{C-F} = 25.3$ Hz), 111.47 (d, J_{C-F} = 17.2 Hz), 108.44, 100.56, 56.08, 55.35, 50.50, 48.72, 39.13, 36.03, 23.25. HRMS (ESI) m/z for C23H26BrFN3O [M+H]. calcd 458.1243, found 458.1228.

1-(3-Chloro-2,4-difluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1' pyrido[3,4-b]indole] (1j).

Yield = 124 mg (58 %) ¹H NMR (400 MHz, CDCl₃) δ 8.10 (s, 1H), 7.36 – 7.24 (m, 1H), 7.17 (s, 1H), $7.01 - 6.87$ (m, 2H), 6.83 (dd, $J = 8.7, 2.5$ Hz, 1H), 3.88 (s, 3H), 3.65 (s, 2H), 3.15 (t, $J = 5.7$ Hz, 2H), $2.80 - 2.66$ (m, 4H), 2.60 (td, $J = 11.9$, 2.6 Hz, 2H), 2.10 (td, $J =$ 13.4, 4.6 Hz, 2H), $1.87 - 1.67$ (m, 2H).¹³C NMR (101 MHz, CDCl₃) δ 158.08 (dd, $J_{\text{C-F}}$ =

250.1, 2.9 Hz), 157.44 (d, $J_{C-F} = 251.6$, 2.9 Hz), 154.01, 140.77, 130.62, 129.34 (dd, $J_{C-F} =$ 9.0, 5.7 Hz), 127.74, 121.95 (dd, J_{C-F} = 15.0, 4.0 Hz), 111.50, 111.46, 111.44 (dd, J_{C-F} = 20.6, 4.4 Hz), 109.77 (t, $J_{\text{C-F}} = 21.3$ Hz), 108.58, 100.56, 56.06, 55.41, 50.51, 48.65, 39.10, 36.27, 23.21.HRMS (ESI) m/z for C23H25ClF2N3O [M+H]. calcd 432.1654, found 432.1646.

1-(2,4-Difluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (2a).

Yield = 138 mg (70 %) ¹H NMR (400 MHz, CDCl₃) δ 7.85 (s, 1H), 7.37 (q, J = 7.9 Hz, 1H), 7.18 (d, $J = 8.7$ Hz, 1H), 6.94 (s, 1H), 6.82 (ddt, $J = 14.4$, 8.8, 5.4 Hz, 3H), 3.85 (s, 3H), 3.61 $(s, 2H)$, 3.13 (t, J = 5.7 Hz, 2H), 2.75 (d, J = 11.4 Hz, 2H), 2.67 (t, J = 5.8 Hz, 2H), 2.57 (t, J $= 11.7$ Hz, 2H), 2.07 (td, $J = 13.2$, 4.3 Hz, 2H), 1.79 (s, 2H). ¹³C NMR (101 MHz, CDCl₃) δ , 161.14 – 154.80 (m), 140.93 – 128.43 (m), 120.82 (dd, $J = 14.8$, 3.9 Hz), 111.57 – 108.50 (m), $103.92 - 22.26$ (m). ¹³C NMR (101 MHz, CDCl₃) δ 163.05 (dd, $J_{C-F} = 77.9$, 11.9 Hz), 160.58 (dd, J_{C-F} = 79.0, 12.0 Hz), 154.90, 154.04, 140.83, 132.40 (dd, J_{C-F} = 9.6, 6.2 Hz), 130.53, 128.52, 127.75, 120.82 (dd, $J_{C-F} = 14.8$, 3.9 Hz), 111.04 (dd, $J_{C-F} = 20.9$, 3.7 Hz), 108.60, 103.56 (t, $J_{\text{C-F}} = 25.8 \text{ Hz}$), 100.54, 56.72, 56.05, 55.16, 51.90, 50.54, 48.64, 40.07, 39.10, 37.28, 36.37, 23.22, 22.36. HRMS (ESI) m/z for C23H26F2N3O [M+H]. calcd 398.2044, found 398.2036.

1-(2,5-Difluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (2b).

Yield = 173 mg (87 %) ¹H NMR (400 MHz, CDCl₃) δ 8.12 – 7.83 (m, 1H), 7.19 (d, $J = 8.8$ Hz, 2H), $7.11 - 6.90$ (m, 3H), 6.86 (d, $J = 8.8$ Hz, 1H), 3.90 (s, 3H), 3.64 (s, 2H), 3.16 (d, J $= 5.8$ Hz, 2H), 2.91 – 2.68 (m, 4H), 2.62 (t, $J = 11.8$ Hz, 2H), 2.24 – 1.98 (m, 2H), 1.81 (d, J $= 13.6$ Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 159.22 (dd, $J_{C-F} = 140.6$, 2.3 Hz), 156.81 (dd, $J_{C-F} = 140.5, 2.3$ Hz), 154.04, 140.95, 130.65, 127.81, 127.28 (dd, $J_{C-F} = 17.0, 7.3$ Hz), 117.38 (dd, $J_{C-F} = 24.1$, 4.9 Hz), 116.26 (dd, $J_{C-F} = 25.2$, 8.5 Hz), 114.99 (dd, $J_{C-F} = 24.2$, 8.5 Hz), 111.57, 111.45, 108.59, 100.61, 56.08, 55.32, 50.58, 48.84, 39.11, 36.43, 23.23. HRMS (ESI) m/z for $C_{23}H_{26}F_2N_3O$ [M+H]. calcd 398.2044, found 398.2031.

1-(3,5-Difluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (2c).

Yield = 113 mg (57 %) ¹H NMR (800 MHz, CDCl₃) δ 7.85 (s, 1H), 7.24 (d, J = 8.7 Hz, 1H), $7.00 - 6.92$ (m, 3H), 6.84 (dd, $J = 8.7$, 2.4 Hz, 1H), 6.73 (tt, $J = 8.9$, 2.4 Hz, 1H), 3.88 (s, 3H), 3.58 (s, 2H), 3.16 (t, $J = 5.7$ Hz, 2H), 2.78 – 2.66 (m, 4H), 2.57 (td, $J = 12.0$, 2.4 Hz, 2H), 2.12 (td, $J = 13.3$, 4.3 Hz, 2H), 1.80 (dd, $J = 14.1$, 2.6 Hz, 2H). ¹³C NMR (201 MHz, CDCl₃) δ 163.66 (d, $J_{\text{C-F}}$ = 12.9 Hz), 162.43 (d, $J_{\text{C-F}}$ = 12.5 Hz), 154.08, 140.74, 130.55, 127.74, 111.52, 111.51 (dd, $J_{\text{C-F}} = 20.9$, 4.0 Hz), 111.49, 108.64, 102.48 (t, $J_{\text{C-F}} = 25.5$ Hz), 100.52, 62.34, 56.06, 50.54, 48.88, 39.09, 36.39, 23.21. HRMS (ESI) m/z for C23H26F2N3O [M+H]. calcd 398.2044, found 398.2030.

6'-Methoxy-1-(3,4,5-trifluorobenzyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (2d).

Yield = 159 mg (77 %).¹H NMR (400 MHz, CDCl₃) δ 7.76 (s, 1H), 7.23 (d, J = 8.6 Hz, 1H), 7.04 (t, $J = 7.5$ Hz, 2H), 6.96 (d, $J = 2.4$ Hz, 1H), 6.84 (dt, $J = 8.7$, 1.8 Hz, 1H), 3.87 (s, 3H), 3.52 (s, 2H), 3.16 (t, $J = 5.7$ Hz, 2H), 2.71 (q, $J = 5.3$ Hz, 4H), 2.56 (t, $J = 11.8$ Hz, 2H), 2.10 (td, $J = 13.2$, 4.3 Hz, 2H), 1.81 (d, $J = 13.7$ Hz, 2H). ¹³C NMR (201 MHz, CDCl₃) δ 154.07, 151.14 (ddd, $J_{C-F} = 249.5$, 10.1, 3.7 Hz), 140.68, 139.34 – 138.02 (m), 135.07, 130.58, 127.74, 112.54 (dd, $J_{\text{C-F}} = 17.1$, 3.7 Hz), 111.53, 111.50, 108.64, 100.53, 61.90, 56.05, 50.55, 48.79, 39.08, 36.34, 23.19. HRMS (ESI) m/z for C₂₃H₂₄F₃N₃O [M+H]. calcd 416.1950, found 416.1939.

6'-Methoxy-1-((perfluorophenyl)methyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1' pyrido[3,4-b]indole] (2e).

Yield = 202 mg (90 %).¹H NMR (400 MHz, CDCl₃) δ 7.73 (s, 1H), 7.19 (d, J = 8.7 Hz, 1H), 6.95 (t, $J = 1.8$ Hz, 1H), 6.83 (dt, $J = 8.7$, 1.9 Hz, 1H), 3.87 (d, $J = 1.4$ Hz, 3H), 3.79 (d, $J =$ 2.4 Hz, 2H), 3.13 (t, $J = 5.7$ Hz, 2H), 2.76 (d, $J = 11.1$ Hz, 2H), 2.72 – 2.61 (m, 4H), 2.06 (td, $J = 13.1$, 4.5 Hz, 2H), 1.80 (d, $J = 13.6$ Hz, 2H). ¹³C NMR (201 MHz, CDCl₃) δ 154.03, 145.63 (d, $J_{C-F} = 248.0$ Hz), 140.70, 140.59 (d, $J_{C-F} = 254.2$ Hz), 137.39 (d, $J_{C-F} = 251.6$ Hz), 130.54, 127.73, 111.47, 111.46, 110.75 (t, $J_{C-F} = 18.7$ Hz), 108.66, 100.50, 56.01, 50.26, 48.92, 48.07, 39.04, 36.34, 23.20. HRMS (ESI) m/z for C23H23F5N3O [M+H]. calcd 452.1761, found 452.1753.

1-(3,4-Difluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (2f).

Yield = 88 mg (44 %).¹H NMR (400 MHz, CDCl₃) δ 8.15 – 7.91 (m, 1H), 7.25 (dt, J = 16.3, 7.7 Hz, 2H), $7.17 - 7.03$ (m, 2H), 6.97 (s, 1H), 6.84 (dd, $J = 8.8$, 2.4 Hz, 1H), 3.88 (s, 3H), 3.55 (s, 2H), 3.16 (t, $J = 5.7$ Hz, 2H), 2.72 (q, $J = 8.2$, 6.0 Hz, 4H), 2.55 (t, $J = 11.1$ Hz, 2H), 2.11 (td, $J = 13.5$, 4.8 Hz, 2H), 1.81 (s, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 154.05, δ 151.15 (dd, J_{C-F} = 78.7, 12.7 Hz), 148.69 (dd, J_{C-F} = 78.1, 12.5 Hz), 140.79, 135.47, 130.62, 127.76, 124.91, 117.80 (d, $J_{\text{C-F}} = 17.2 \text{ Hz}$), 116.90 (d, $J_{\text{C-F}} = 17.2 \text{ Hz}$), 111.52, 111.48, 108.59, 100.57, 62.14, 56.07, 50.62, 48.80, 39.10, 36.31, 23.21. HRMS (ESI) m/z for $C_{23}H_{26}F_2N_3O$ [M+H]. calcd 398.2044, found 398.2032.

6'-Methoxy-1-(2,3,4-trifluorobenzyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (2g).

Yield = 200 mg (96 %) ¹H NMR (400 MHz, CDCl₃) δ 8.24 (s, 1H), 7.18 (d, *J* = 8.8 Hz, 1H), $7.15 - 7.03$ (m, 1H), $6.99 - 6.86$ (m, 2H), $6.85 - 6.67$ (m, 1H), 3.87 (d, $J = 2.9$ Hz, 3H), 3.68 -3.59 (m, 2H), 3.13 (t, $J = 5.7$ Hz, 2H), 2.77 – 2.65 (m, 4H), 2.65 – 2.50 (m, 2H), 2.16 – 1.98 (m, 2H), 1.85 – 1.71 (m, 2H). 13C NMR (101 MHz, CDCl3) δ 153.99, 150.40 (ddd, $J_{C-F} = 250.2, 9.9, 2.6 \text{ Hz}$), 150.20 (d, $J_{C-F} = 250.4, 9.5, 3.0 \text{ Hz}$), 140.80, 139.88 (dt, $J_{C-F} =$ 251.8, 15.6 Hz), 130.65, 127.72, 124.92 (ddd, $J_{C-F} = 8.5, 6.3, 3.2$ Hz), 122.42 (dd, $J_{C-F} =$ 12.1, 2.8 Hz), 111.71 (dd, $J_{C-F} = 17.1$, 3.9 Hz), 111.51, 111.42, 108.52, 100.53, 56.05, 55.14, 50.49, 48.58, 39.08, 36.21, 23.20. HRMS (ESI) m/z for C₂₃H₂₄F₃N₃O [M+H]. calcd 416.1950, found 416.1941.

1-(2,3-Difluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (2h).

Yield = 184 mg (93 %) ¹H NMR (400 MHz, CDCl₃) δ 7.85 (s, 1H), 7.20 (d, J = 8.8 Hz, 2H), 7.14 – 7.02 (m, 2H), 6.96 (d, $J = 2.4$ Hz, 1H), 6.83 (dd, $J = 8.8$, 2.4 Hz, 1H), 3.88 (s, 3H), 3.70 (d, $J = 1.7$ Hz, 2H), 3.15 (t, $J = 5.7$ Hz, 2H), 2.85 – 2.73 (m, 2H), 2.70 (t, $J = 5.7$ Hz, 2H), 2.60 (td, $J = 11.9$, 2.7 Hz, 2H), 2.16 – 1.99 (m, 2H), 1.80 (dq, $J = 14.1$, 2.9 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 154.06, 151.26 (dd, J_{C-F} = 119.9, 12.8 Hz), 148.79 (dd, J_{C-F} $= 119.9, 12.8$ Hz), 140.86, 130.54, 127.77, 127.62 (d, $J_{C-F} = 11.4$ Hz), 126.18 (t, $J_{C-F} = 3.5$ Hz), 123.68 (dd, J_{C-F} = 6.6, 4.8 Hz), 116.10, 115.93, 111.47, 108.62, 100.55, 56.06, 55.41, 50.52, 48.74, 39.10, 36.43, 23.21. HRMS (ESI) m/z for C₂₃H₂₆F₂N₃O [M+H]. calcd 398.2044, found 398.2037.

6'-Methoxy-1-(2,4,5-trifluorobenzyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (2i).

Yield = 193 mg (93 %) ¹H NMR (400 MHz, CDCl₃) δ 8.06 (s, 1H), 7.30 (ddd, *J* = 10.7, 8.9, 6.6 Hz, 1H), 7.19 (d, $J = 8.6$ Hz, 1H), 6.98 (d, $J = 2.4$ Hz, 1H), 6.95 – 6.86 (m, 1H), 6.84 $(dd, J = 8.7, 2.5 Hz, 1H), 3.89 (s, 3H), 3.59 (d, J = 1.3 Hz, 2H), 3.16 (t, J = 5.7 Hz, 2H), 2.81$ -2.66 (m, 4H), 2.60 (dd, J = 23.9, 2.7 Hz, 2H), 2.10 (td, J = 13.7, 4.6 Hz, 2H), 1.80 (dd, J = 14.1, 2.7 Hz, 2H).¹³C NMR (101 MHz, CDCl₃) δ 156.20 (ddd, $J_{C-F} = 247.5$, 10.1, 2.2 Hz), 154.03, 149.1 (ddd, $J_{\text{C-F}} = 251.7$, 14.9, 14.0), 146.8 (ddd, $J_{\text{C-F}} = 245.3$, 12.5, 4.0), 140.79, 130.64, 127.76, 121.72 (dt, $J_{C-F} = 16.9$, 4.6 Hz), 118.72 (dd, $J_{C-F} = 18.9$, 5.7 Hz), 111.52, 111.47, 108.60, 105.31 (dd, $J_{\text{C-F}} = 28.6$, 20.5 Hz), 100.57, 56.06, 54.76, 50.52, 48.70, 39.09, 36.34, 23.21. HRMS (ESI) m/z for $C_{23}H_{24}F_3N_3O$ [M+H]. calcd 416.1950, found 416.1940.

6'-Methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4-b]indole] (3a).

 1_H NMR (400 MHz, MeOD) δ 7.18 (d, J = 8.6 Hz, 1H), 6.90 (d, J = 2.4 Hz, 1H), 6.72 (dd, J $= 8.7, 2.5$ Hz, 1H), 3.80 (s, 3H), 3.11 – 2.94 (m, 4H), 2.84 (dt, $J = 12.7, 3.9$ Hz, 2H), 2.66 (t, $J = 5.7$ Hz, 2H), 1.96 (td, $J = 13.5$, 4.7 Hz, 2H), 1.72 (d, $J = 12.8$ Hz, 2H). ¹³C NMR (101) MHz, MeOD) δ 153.53, 140.40, 131.32, 127.35, 110.98, 110.44, 106.88, 99.80, 54.91, 51.11, 40.53, 38.51, 35.15, 21.97. HRMS (ESI) m/z for C₁₆H₂₂N₃O [M+H]. calcd 272.1763, found 272.1755.

6'-Methoxy-1-(1-phenylethyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4-b]indole] (3b).

Yield = 44 mg (36 %)¹H NMR (800 MHz, MeOD) δ 7.45 – 7.42 (m, 2H), 7.39 (t, J = 7.7 Hz, 2H), 7.34 – 7.30 (m, 1H), 7.21 (d, $J = 8.8$ Hz, 1H), 6.92 (d, $J = 2.4$ Hz, 1H), 6.76 (dd, $J =$ 8.7, 2.4 Hz, 1H), 3.81 (s, 3H), 3.21 (td, $J = 6.0$, 2.8 Hz, 2H), 3.18 – 3.15 (m, 1H), 2.82 (s, 1H), 2.80 - 2.76 (m, 2H), 2.65 (s, 1H), 2.56 (s, 1H), 2.32 (ddd, J = 14.4, 12.8, 4.5 Hz, 1H), 2.21 (ddd, $J = 14.3$, 12.9, 4.4 Hz, 1H), 2.05 (s, 1H), 1.99 – 1.94 (m, 1H), 1.85 (dd, $J = 14.5$, 2.9 Hz, 1H), 1.53 (d, $J = 6.8$ Hz, 3H). ¹³C NMR (201 MHz, MeOD) δ 153.78, 137.50, 131.54, 128.28, 127.67, 127.45, 126.96, 116.74, 111.28, 111.16, 106.76, 99.71, 65.04, 54.85, 45.50, 45.38, 38.49, 34.03, 20.81, 18.26. HRMS (ESI) m/z for $C_{24}H_{30}N_{3}O$ [M+H]. calcd 376.2389, found 376.2389.

6'-Methoxy-1-phenethyl-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4-b]indole] (3c).

Yield = 50 mg (40 %)¹H NMR (800 MHz, MeOD) δ 7.32 (t, J = 7.5 Hz, 2H), 7.28 (d, J = 7.4 Hz, 2H), $7.24 - 7.11$ (m, 2H), 6.92 (d, $J = 2.5$ Hz, 1H), 6.76 (dd, $J = 8.7$, 2.5 Hz, 1H), 3.81 (s, 3H), 3.17 (t, $J = 5.8$ Hz, 2H), 3.00 (d, $J = 11.9$ Hz, 2H), 2.92 (dd, $J = 10.7$, 6.0 Hz, 2H), 2.85 (dd, $J = 10.8$, 6.0 Hz, 2H), 2.80 – 2.61 (m, 4H), 2.20 (td, $J = 13.8$, 4.4 Hz, 2H), 1.89 (d, $J = 14.1$ Hz, 2H). ¹³C NMR (201 MHz, MeOD) δ 153.67, 139.20, 138.53, 131.46, 128.34, 128.25, 127.17, 126.05, 111.18, 110.89, 107.14, 99.77, 59.60, 54.89, 51.27, 47.91, 38.56, 33.94, 32.27, 21.47. HRMS (ESI) m/z for C24H30N3O [M+H]. calcd 376.2389, found 376.2387.

6'-Methoxy-1-(3-phenylpropyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4-b]indole] (3d).

Yield = 21 mg (16 %)¹H NMR (400 MHz, MeOD) δ 7.38 – 7.23 (m, 4H), 7.20 (dd, J = 8.3, 5.1 Hz, 2H), $6.97 - 6.86$ (m, 1H), 6.75 (dt, $J = 8.7$, 1.7 Hz, 1H), 3.81 (s, 3H), 3.17 (t, $J = 5.8$) Hz, 2H), 3.01 (d, $J = 12.0$ Hz, 2H), 2.73 (dt, $J = 14.8$, 6.8 Hz, 8H), 2.21 (td, $J = 13.9$, 4.3 Hz, 2H), 2.06 – 1.85 (m, 4H). 13C NMR (101 MHz, MeOD) δ 153.71, 141.28, 138.32, 131.47, 128.11, 128.06, 127.16, 125.72, 111.15, 110.95, 107.23, 99.74, 57.14, 54.87, 51.12, 47.93, 38.56, 33.74, 32.98, 27.41, 21.48. HRMS (ESI) m/z for $C_{25}H_{32}N_3O$ [M+H]. calcd 390.2545, found 390.2572.

1-(Furan-2-ylmethyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (3e).

Yield = 23 mg (18 %)¹H NMR (400 MHz, CDCl₃) δ 8.13 (s, 1H), 7.41 (s, 1H), 7.20 (dd, J = 8.8, 1.3 Hz, 1H), 6.92 (t, $J = 1.8$ Hz, 1H), 6.80 (dt, $J = 8.7$, 1.9 Hz, 1H), $6.37 - 6.30$ (m, 2H), 3.84 (d, $J = 1.4$ Hz, 3H), 3.70 (s, 2H), 3.11 (t, $J = 5.7$ Hz, 2H), 2.84 (d, $J = 11.6$ Hz, 2H), $2.71 - 2.64$ (m, 4H), 2.24 (td, J = 13.5, 4.2 Hz, 2H), 1.80 (d, J = 13.8 Hz, 2H). ¹³C NMR (101 MHz, CDCl3) δ 154.13, 150.38, 142.80, 140.42, 130.74, 127.70, 111.74, 111.62, 110.48, 110.06, 108.63, 100.60, 56.17, 54.84, 50.47, 48.66, 39.29, 35.91, 23.33. HRMS (ESI) m/z for $C_{21}H_{26}N_3O_2$ [M+H]. calcd 352.2025, found 352.2024.

6'-Methoxy-1-(pyridin-2-ylmethyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (3f).

Yield = 118 mg (65 %).¹H NMR (400 MHz, CDCl₃) δ 8.59 (d, J = 4.9 Hz, 1H), 8.34 (s, 1H), 7.63 (t, $J = 7.7$ Hz, 1H), 7.41 (d, $J = 7.8$ Hz, 1H), 7.17 (d, $J = 8.3$ Hz, 2H), 6.93 (s, 1H), 6.78 (dd, $J = 8.7, 2.6$ Hz, 1H), 3.84 (s, 3H), 3.76 (s, 2H), 3.12 (t, $J = 5.7$ Hz, 2H), 2.77 (d, $J = 11.4$ Hz, 2H), $2.71 - 2.58$ (m, 4H), 2.15 (td, $J = 13.4$, 12.8, 4.3 Hz, 2H), 1.75 (d, $J = 13.6$ Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 158.30, 153.99, 149.46, 140.95, 136.54, 130.73, 127.70, 123.53, 122.28, 111.65, 111.37, 108.38, 100.51, 64.61, 56.10, 50.63, 49.14, 39.18, 36.19, 23.26. HRMS (ESI) m/z for C₂₂H₂₇N₄O [M+H]. calcd 363.2185, found 363.2188.

6'-Methoxy-1-(pyridin-3-ylmethyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (3g).

Yield = 61 mg (34 %).¹H NMR (400 MHz, MeOD) δ 8.53 (s, 1H), 8.44 (d, J = 4.9 Hz, 1H), 7.80 (dd, $J = 7.8$, 2.2 Hz, 1H), 7.39 (dd, $J = 7.9$, 5.0 Hz, 1H), 7.15 (d, $J = 8.9$ Hz, 1H), 6.88

(d, $J = 2.3$ Hz, 1H), 6.70 (dt, $J = 8.8$, 1.8 Hz, 1H), 3.78 (s, 3H), 3.58 (s, 2H), 3.02 (t, $J = 5.7$ Hz, 2H), 2.64 (t, $J = 5.6$ Hz, 4H), 2.46 (t, $J = 12.1$ Hz, 2H), 2.06 (tt, $J = 14.1$, 7.0 Hz, 2H), 1.69 (d, $J = 13.7$ Hz, 2H). ¹³C NMR (101 MHz, MeOD) δ 153.55, 149.62, 147.56, 139.80, 137.96, 134.14, 131.35, 127.32, 123.72, 111.09, 110.52, 107.03, 99.77, 59.46, 54.91, 50.90, 48.08, 38.43, 34.75, 21.82. HRMS (ESI) m/z for C₂₂H₂₇N₄O [M+H]. calcd 363.2185, found 363.2180.

1-(4-(tert-Butyl)benzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (3h).

Yield = 111 mg (53 %). ¹H NMR (400 MHz, CDCl₃) δ 8.52 (br, 1H), 7.32 (s, 4H), 7.22 (d, J $= 8.8$ Hz, 1H), 6.97 (d, $J = 2.4$ Hz, 1H), 6.82 (dd, $J = 8.8$, 2.6 Hz, 1H), 3.88 (s, 3H), 3.63 (s, 2H), 3.15 (t, $J = 5.7$ Hz, 2H), 2.82 (d, $J = 11.6$ Hz, 2H), 2.71 (t, $J = 5.7$ Hz, 2H), 2.57 (td, $J =$ 12.1, 2.4 Hz, 2H), 2.20 (td, $J = 13.4$, 4.4 Hz, 2H), 1.79 (s, 2H), 1.34 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 153.97, 150.32, 140.87, 134.19, 130.71, 129.29, 127.67, 125.27, 111.58, 111.39, 108.34, 100.53, 62.95, 56.09, 50.63, 48.87, 39.17, 36.03, 34.51, 31.40, 23.27. HRMS (ESI) m/z for $C_{27}H_{36}N_3O$ [M+H]. calcd 418.2858, found 418.2849.

1-(2,4-Difluorobenzyl)-6'-methoxy-2',3',4',9'-tetrahydrospiro[azepane-4,1'-pyrido[3,4 b]indole] (4a).

Yield = 56 mg (76 %)¹H NMR (400 MHz, MeOD) δ 7.48 – 7.31 (m, 1H), 7.23 (d, J = 8.7 Hz, 1H), $7.09 - 6.92$ (m, 2H), 6.91 (t, $J = 1.8$ Hz, 1H), 6.75 (dt, $J = 8.9$, 1.9 Hz, 1H), 3.79 (s, 3H), 3.67 (s, 2H), 3.15 (dp, $J = 24.7$, 6.3, 5.8 Hz, 2H), 2.75 (h, $J = 6.8$ Hz, 6H), 2.09 (dtd, $J =$ 44.9, 14.8, 7.3 Hz, 3H), 1.86 (dd, $J = 13.8$, 6.4 Hz, 1H), 1.72 (p, $J = 5.9$ Hz, 2H). ¹³C NMR $(101 \text{ MHz}, \text{MeOD})$ δ 163.37 (dd, $J_{\text{C-F}} = 93.9, 12.1 \text{ Hz}$), 160.91 (dd, $J_{\text{C-F}} = 94.0, 12.2 \text{ Hz}$), 153.70, 138.27, 132.81 (dd, $J_{C-F} = 9.7$, 6.1 Hz), 131.21, 127.05, 121.39 (dd, $J_{C-F} = 14.6$, 3.7 Hz), 111.36, 110.90 (dd, $J_{\text{C-F}} = 21.2$, 3.7 Hz), 110.89, 105.10, 103.44 (t, $J_{\text{C-F}} = 26.0 \text{ Hz}$), 99.84, 57.47, 55.62, 54.89, 53.95, 50.44, 38.60, 38.56, 36.75, 24.83, 20.59. HRMS (ESI) m/z for $C_{24}H_{28}F_2N_3O$ [M+H]. calcd 412.2200, found 412.2193.

1'-(2,4-Difluorobenzyl)-6-methoxy-2,3,4,9-tetrahydrospiro[pyrido[3,4-b]indole-1,3' pyrrolidine] (4b).

Yield = 83 mg (43 %) ¹H NMR (800 MHz, CDCl₃) δ 8.87 (s, 1H), 7.33 (td, J = 8.6, 6.6 Hz, 1H), 7.24 (d, $J = 8.7$ Hz, 1H), 6.94 (d, $J = 2.6$ Hz, 1H), 6.89 – 6.84 (m, 2H), 6.83 (dd, $J =$ 8.7, 2.6 Hz, 1H), 3.87 (s, 3H), 3.79 – 3.73 (m, 2H), 3.26 (dt, $J = 13.0$, 4.6 Hz, 1H), 3.18 – $3.07 \text{ (m, 3H)}, 2.78 - 2.68 \text{ (m, 2H)}, 2.64 \text{ (q, } J = 9.0 \text{ Hz}, 1\text{H}), 2.53 \text{ (d, } J = 8.9 \text{ Hz}, 1\text{H}), 2.34 \text{ (g, } J = 9.0 \text{ Hz}, 1\text{ H}), 2.53 \text{ (g, } J = 9.0 \text{ Hz}, 1\text{ Hz}, 1\text{ Hz})$ (dt, $J = 13.3$, 8.5 Hz, 1H), 2.14 (ddd, $J = 13.0$, 9.2, 3.1 Hz, 1H). ¹³C NMR (201 MHz, CDCl₃) δ 162.58 (dd, J_{C-F} = 208.3, 11.7 Hz), 161.34 (dd, J_{C-F} = 208.7, 12.1 Hz), 153.96, 140.73, 132.11–132.03 (m), 130.58, 127.26, 120.86, 111.63, 111.30, 111.21 (dd, J_{C-F} = 21.3, 3.7 Hz), 106.03, 104.09 (t, $J_{C-F} = 25.7$ Hz), 100.49, 65.57, 59.95, 56.05, 52.59, 52.48, 41.80, 39.15, 22.30. HRMS (ESI) m/z for C22H24F2N3O [M+H]. calcd 384.1887, found 384.1894.

Yield = 85 mg (43 %) ¹H NMR (400 MHz, CDCl₃) δ 9.65 (s, 1H), 7.31 – 7.27 (m, 1H), 7.22 (d, $J = 6.7$ Hz, 1H), 6.96 (d, $J = 2.2$ Hz, 1H), 6.90 – 6.76 (m, 3H), 3.88 (s, 3H), 3.60 (q, $J =$ 12.9 Hz, 2H), 3.47 – 3.17 (m, 2H), 3.00 (s, 1H), 2.92 – 2.61 (m, 3H), 2.43 – 2.05 (m, 3H), $1.96 - 1.67$ (m, 2H), 1.58 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 163.37 (dd, $J_{\text{C-F}} = 84.4$, 12.1 Hz), 160.90 (dd, $J_{C-F} = 84.5$, 12.3 Hz), 153.79, 139.99, 132.55 (dd, $J_{C-F} = 9.5$, 6.2 Hz), 130.43, 127.18, 120.51 (dd, $J_{C-F} = 14.3$, 3.7 Hz), 111.74, 111.31, 111.13 (dd, $J_{C-F} = 20.9$, 3.7 Hz), 106.14, 104.29 (t, $J_{\text{C-F}} = 25.9$ Hz), 100.22, 63.38, 56.48, 56.07, 53.34, 52.17, 38.90, 36.31, 22.64, 22.09. HRMS (ESI) m/z for C₂₃H₂₆F₂N₃O [M+H]. calcd 398.2044, found 398.2040.

1-Benzyl-N-(2-(5-methoxy-1H-indol-3-yl)ethyl)piperidin-4-amine (4d).

Yield = 105 mg (53%) ¹H NMR (400 MHz, CDCl₃) δ 8.37 (s, 1H), 7.34 (q, J = 7.8 Hz, 1H), $7.30 - 7.18$ (m, 1H), 7.01 (s, 1H), 6.83 (dt, $J = 32.9$, 9.1 Hz, $3H$), 3.88 (s, $3H$), 3.53 (s, $2H$), 2.98 (dq, $J = 11.1$, 6.8 Hz, 4H), 2.84 (d, $J = 11.2$ Hz, 2H), 2.50 (tt, $J = 10.0$, 4.3 Hz, 1H), 2.07 (t, $J = 11.5$ Hz, 2H), 1.86 (d, $J = 12.4$ Hz, 2H), 1.54 – 1.24 (m, 2H). ¹³C NMR (101) MHz, CDCl₃) δ 162.87 (dd, $J_{C-F} = 77.0$, 12.3 Hz), 160.41 (dd, $J_{C-F} = 78.0$, 11.8 Hz), 153.88, 132.19 (dd, $J_{C-F} = 9.5$, 6.2 Hz), 131.61, 127.85, 122.84, 120.95 (dd, $J_{C-F} = 15.0$, 3.7 Hz), 113.56, 112.16, 111.91, 110.96 (dd, $J_{C-F} = 20.9$, 3.7 Hz), 103.48 (t, $J_{C-F} = 25.7$ Hz), 100.75, 55.97, 54.82, 54.65, 52.15, 46.76, 32.66, 26.02. HRMS (ESI) m/z for $C_{23}H_{28}F_{2}N_{3}O$ [M+H]. calcd 400.2200, found 400.2188.

1-(2,4-Difluorobenzyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4-b]indole] (5a).

Yield = 103 mg (56 %).¹H NMR (400 MHz, CDCl₃) δ 8.94 (s, 1H), 7.51 (d, J = 7.5 Hz, 1H), $7.43 - 7.25$ (m, 2H), 7.13 (hept, $J = 8.4$, 7.5 Hz, 2H), $6.84 - 6.61$ (m, 2H), 3.66 (s, 2H), 3.13 $(t, J = 5.7 \text{ Hz}, 2\text{H})$, $2.86 - 2.69 \text{ (m, 4H)}$, $2.69 - 2.52 \text{ (m, 2H)}$, $2.19 \text{ (dt, } J = 13.0, 7.5 \text{ Hz}, 2\text{H})$, 1.73 (d, $J = 13.7$ Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 163.22 (dd, $J_{C-F} = 91.1$, 11.9 Hz), 160.74 (dd, $J_{\text{C-F}} = 91.6$, 11.9 Hz), 139.72, 135.68, 132.93 (dd, $J_{\text{C-F}} = 9.5$, 5.9 Hz), 127.30, 121.56, 119.77 (dd, J_{C-F} = 15.0, 3.7 Hz), 119.18, 118.12, 111.21 (dd, J_{C-F} = 21.3, 3.7 Hz), 110.99, 108.50, 103.78 (t, $J_{\text{C-F}} = 25.7 \text{ Hz}$), 55.07, 50.44, 48.50, 39.14, 35.77, 23.15. HRMS (ESI) m/z for $C_{22}H_{24}F_2N_3$ [M+H]. calcd 368.1938, found 368.1935.

6'-Bromo-1-(2,4-difluorobenzyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (5b).

Yield = 51 mg (54 %) ¹H NMR (400 MHz, CDCl₃) δ 8.83 (s, 1H), 7.60 (d, J = 2.1 Hz, 1H), 7.36 (td, $J = 8.7$, 6.6 Hz, 1H), 7.19 (dd, $J = 8.6$, 2.0 Hz, 1H), 7.12 (d, $J = 8.5$ Hz, 1H), 6.83 – 6.68 (m, 2H), 3.65 (s, 2H), 3.11 (t, $J = 5.7$ Hz, 2H), 2.77 (dd, $J = 8.4$, 2.9 Hz, 2H), 2.70 – 2.57 (m, 4H), 2.14 (td, $J = 13.3$, 4.6 Hz, 2H), 1.72 (dd, $J = 14.1$, 2.4 Hz, 2H). ¹³C NMR (101) MHz, CDCl₃) δ 163.19 (dd, J_{C-F} = 93.7, 11.9 Hz), 160.72 (dd, J_{C-F} = 94.1, 11.9 Hz), 141.14, 134.23, 132.81 (dd, $J_{C-F} = 9.7, 5.7$ Hz), 129.14, 124.23, 120.79, 119.76 (dd, $J_{C-F} =$ 14.9, 3.9 Hz), 112.38, 112.31, 111.19 (dd, $J_{C-F} = 21.1$, 3.9 Hz), 108.39, 103.80 (t, $J_{C-F} =$ 25.9 Hz), 55.12, 50.39, 48.46, 38.99, 35.76, 22.99. HRMS (ESI) m/z for C₂₂H₂₃BrF₂N₃ [M +H]. calcd 446.1043, found 446.1036.

6'-Chloro-1-(2,4-difluorobenzyl)-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (5c).

Yield = 134 mg (67 %).¹H NMR (400 MHz, CDCl₃) δ 8.86 – 8.49 (m, 1H), 7.43 (d, J = 9.3 Hz, 2H), 7.21 (d, $J = 8.7$ Hz, 1H), 7.08 (d, $J = 8.5$ Hz, 1H), 6.82 (q, $J = 9.0$ Hz, 2H), 3.69 (s, 2H), 3.11 (d, $J = 5.8$ Hz, 2H), 2.81 (d, $J = 11.3$ Hz, 2H), 2.67 (q, $J = 8.5$, 8.0 Hz, 4H), 2.18 $(q, J = 8.0, 5.3 \text{ Hz}, 2\text{H})$, 1.76 (d, $J = 13.6 \text{ Hz}, 2\text{H}$). ¹³C NMR (101 MHz, CDCl₃) δ 163.25 $(\text{dd}, J_{\text{C-F}} = 100.2, 12.4 \text{ Hz}), 160.77 \text{ (dd}, J_{\text{C-F}} = 100.6, 12.3 \text{ Hz}), 141.11, 133.91, 132.90 \text{ (dd)}$ $J_{\text{C-F}}$ = 9.8, 5.8 Hz), 128.44, 124.90, 121.74, 119.48 (d, $J_{\text{C-F}}$ = 15.4 Hz), 117.69, 111.85, 111.29 (d, $J_{\text{C-F}} = 20.7 \text{ Hz}$), 108.51, 103.82 (t, $J_{\text{C-F}} = 25.7 \text{ Hz}$), 54.96, 50.36, 48.43, 39.00, 35.72, 22.99. HRMS (ESI) m/z for C₂₂H₂₃ClF₂N₃ [M+H]. calcd 402.1549, found 402.1539.

1-(2,4-Difluorobenzyl)-7'-methoxy-2',3',4',9'-tetrahydrospiro[piperidine-4,1'-pyrido[3,4 b]indole] (5d).

Yield = 90 mg (45 %).¹H NMR (400 MHz, CDCl₃) δ 8.24 (s, 1H), 7.48 – 7.32 (m, 2H), 6.94 -6.68 (m, 4H), 3.82 (s, 3H), 3.67 (s, 2H), 3.13 (t, $J = 5.7$ Hz, 2H), 2.83 – 2.76 (m, 2H), 2.72 $- 2.54$ (m, 4H), 2.17 (td, J = 13.5, 13.1, 4.4 Hz, 2H), 1.77 (d, J = 13.7 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 163.17 (dd, J_{C-F} = 91.6, 11.9 Hz), 160.70 (dd, J_{C-F} = 92.1, 12.1 Hz), 156.28, 138.34, 136.25, 132.75 (dd, $J_{C-F} = 9.5$, 5.9 Hz), 121.77, 119.96 (dd, $J_{C-F} = 14.8$, 3.8 Hz), 118.65, 111.20 (dd, $J_{\text{C-F}} = 20.8$, 3.8 Hz), 108.93, 108.49, 103.74 (t, $J_{\text{C-F}} = 25.7$ Hz), 94.99, 55.78, 54.99, 50.38, 48.59, 39.15, 35.97, 23.14. HRMS (ESI) m/z for C₂₃H₂₆F₂N₃O [M+H]. calcd 398.2044, found 398.2033.

1-(2,4-Difluorobenzyl)-6'-methoxy-9'-methyl-2',3',4',9'-tetrahydrospiro[piperidine-4,1' pyrido[3,4-b]indole] (5e).

Yield = 47 mg (23 %).¹H NMR (400 MHz, CDCl₃) δ 7.45 (q, J = 8.1 Hz, 1H), 7.20 (d, J = 8.7 Hz, 1H), 6.97 (d, $J = 2.4$ Hz, 1H), 6.87 (dtd, $J = 28.7$, 9.4, 2.5 Hz, 3H), 3.89 (s, 6H), 3.65 $(s, 2H)$, 3.10 (t, $J = 5.7$ Hz, $2H$), 2.72 (tt, $J = 18.0$, 9.9 Hz, $6H$), 2.52 – 2.22 (m, $2H$), 1.72 (d, $J = 13.7$ Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 162.63 (d, $J_{C-F} = 11.9$ Hz), 160.15 (d, J_{C-F} $= 12.1$ Hz), 153.93, 140.67, 132.74, 132.27 (br), 126.69, 120.93 (br), 111.39, 111.02 (d, $J_{\text{C-F}}$ $= 21.6$ Hz), 109.59, 109.17, 103.62 (t, $J_{C-F} = 25.7$ Hz), 100.23, 56.10, 54.96, 51.63, 48.56, 38.81, 34.79, 32.32, 24.10. HRMS (ESI) m/z for C24H28F3N3O [M+H]. calcd 412.2200, found 412.2191.

(+/−)-(2R,11aS)-6'-methoxy-1,2',3,3',4,4',6,9',11,11a-decahydrospiro[pyrido[1,2 b]isoquinoline-2,1'-pyrido[3,4-b]indole] (6b).

¹H NMR (599 MHz, cdcl₃) δ 8.34 (d, *J* = 20.8 Hz, 1H), 7.14 (dq, *J* = 13.2, 6.7, 6.2 Hz, 2H), 7.11 – 6.99 (m, 3H), 6.94 (d, $J = 2.6$ Hz, 1H), 6.78 (dt, $J = 8.7$, 2.3 Hz, 1H), 3.86 (s, 4H), 3.52 (d, $J = 15.3$ Hz, 1H), $3.22 - 3.08$ (m, 2H), $2.94 - 2.88$ (m, 1H), 2.84 (tq, $J = 8.2, 3.1, 2.6$ Hz, 1H), $2.77 - 2.72$ (m, 3H), 2.69 (t, $J = 5.7$ Hz, 2H), $2.29 - 2.19$ (m, 1H), 2.00 (dt, $J =$ 13.9, 3.1 Hz, 1H), 1.92 – 1.82 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 153.98, 140.49, 133.80, 133.42, 130.74, 128.10, 127.67, 126.40, 126.02, 125.81, 111.62, 111.46, 108.47, 100.46, 57.58, 56.05, 52.91, 51.35, 50.40, 43.76, 39.28, 36.37, 35.64, 23.26. HRMS (ESI) m/z for $C_{24}H_{28}N_3O$ [M+H]. calcd 374.2232, found 374.2222.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

ACKNOWLEDGEMENTS:

This work was supported by the Cystic Fibrosis Foundation [HAGGIE19R2 and HAGGIE20XX0]; the National Institutes of Health [DK072517, DK099803, DK075302, EY13574]; and the UC Davis Tara K. Telford CF Fund [fellowship to J.S.Z.]. The authors thank Dr. Christine Bear (Hospital for Sick Children, Toronto, Canada) for provision of the gene edited 16HBE14o- cells expressing I1234del-CFTR, Drs. Martin Mense and Hillary Valley (Cystic Fibrosis Foundation Therapeutics Laboratory) for provision of the gene edited 16HBE14o-cells expressing N1303K-CFTR, and Drs. Andras Rab and Jeong Hong (Emory University) for provision of the FRT cell model expressing R347P-CFTR. The authors also thank Dr. Marc Anderson (San Francisco State University) for helpful discussions.

ABBREVIATIONS USED:

REFERENCES

- 1. Elborn JS (2016) Cystic Fibrosis. Lancet 388:2519–2531. [PubMed: 27140670]
- 2. Keating D, Marigowda G, Burr L, Daines C, Mall MA, McKone EF, Ramsey BW, Rowe SM, Sass LA, Tullis E, McKee CM, Moskowitz SM, Robertson S, Savage J, Simard C, Van Goor F, Waltz D, Xuan F, Young T, Taylor-Cousar and JL VX16-445-001 Study Group (2018) VX-445-Tezacaftor-Ivacaftor in patients with cystic fibrosis and one or two Phe508del alleles. N. Engl. J. Med 379:1612–1620. [PubMed: 30334692]
- 3. Heijerman HGM, McKone EF, Downey DG, Van Braeckel E, Rowe SM, Tullis E, Mall MA, Welter JJ, Ramsey BW, McKee CM, Marigowda G, Moskowitz SM, Waltz D, Sosnay PR, Simard C, Ahluwalia N, Xuan F, Zhang Y, Taylor-Cousar JL, McCoy and KS V17-445-103 Trial Group (2019) Efficacy and safety of the elexacaftor plus tezacaftor plus ivacaftor combination regimen in people with cystic fibrosis homozygous for the F508del mutation: a double-blind, randomized, phase 3 trial. Lancet 394:1940–1948. [PubMed: 31679946]

- 4. Middleton PG, Mall MA, Drevinek P, Lands LC, McKone EF, Polineni D, Ramsey BW, Taylor-Cousar JL, Tullis E, Vermeulen F, Marigowda G, McKee CM, Moskowitz SM, Nair N, Savage J, Simard C, Tian F, Waltz D, Xuan F, Rowe SM, Jain and R V17-445-102 Study Group (2019) Elexacaftor-tezacaftor-ivacaftor for cystic fibrosis with a single Phe508del allele. N. Engl. J. Med 381:1809–1819. [PubMed: 31697873]
- 5. Burgener EB and Moss RB (2018) Cystic fibrosis transmembrane conductance regulator modulators: precision medicine in cystic fibrosis. Curr. Opin. Pediatr 30:372–377. [PubMed: 29538046]
- 6. Kym PR, Wang X, Pizzonero M and Van der Plas SE (2018) Recent progress in the discovery and development of small-molecule modulators of CFTR. Prog. Med. Chem 57:235–276. [PubMed: 29680149]
- 7. Haggie PM, Phuan P-W, Tan JA, Xu H, Avramescu RG, Perdomo D, Zlock L, Nielson DW, Finkbeiner WE, Lukacs GL and Verkman AS (2017) Correctors and potentiators rescue function of the truncated W1282X-cystic fibrosis transmembrane regulator (CFTR) translation product. J. Biol. Chem 292:771–785. [PubMed: 27895116]
- 8. Phuan P-W, Son J-H, Tan JA, Li C, Musante I, Zlock L, Nielson DW, Finkbeiner WE, Kurth MJ, Galietta LJ, Haggie PM and Verkman AS (2018) Combination potentiator ('co-potentiator') therapy for CF caused by CFTR mutants, including N1303K, that are poorly responsive to single potentiators. J. Cyst. Fibros 17:595–606. [PubMed: 29903467]
- 9. Phuan P-W, Tan JA, Rivera AA, Zlock L, Nielson D, Finkbeiner WE, Haggie PM and Verkman AS (2019) Nanomolar-potency 'co-potentiators' for therapy of a defined subset of minimal function CFTR mutations. Sci. Rep 9:17640. [PubMed: 31776420]
- 10. Mokrosz MJ, Bartyzel P and Mokorosz JL (1993) Structure and spectral properties of b-carbolines. 5. Synthesis and stereochemistry of new trihydrodiazabicyclo[3.m.n]alkano[4',5':1,2]-pyrido[3,4 b]-indole ring systems. J. Hetero. Chem 30:1543–1548.
- 11. Bridges TM, Brady AE, Kennedy JP, Daniels RN, Miller NR, Kim K, Breininger ML, Gentry PR, Brogan JT, Jones CK, Conn PJ and Lindsley CW (2008) Synthesis and SAR of analogues of the M1 allosteric agonist TBPB. Part I: Exploration of alternative benzyl and privileged structure moieties. Bioorg. Med. Chem. Lett 18:5439–5442. [PubMed: 18805692]
- 12. Abd Rabo M, Mahmoud M and Pagenkopf BL (2010) Synthesis of 5-azaindoles via a cycloaddition reaction between nitriles and donor-acceptor cyclopropanes. Org. Letts 12:3168– 3171. [PubMed: 20550103]
- 13. Pictet A and Spengler T (1911) Uber die bildung von isochinolin-derivaten durch einwirkung von methylal auf phenyl-athylamin, phenyl-alanin und tyrosin. Berichte der Deutschen Chemischen Gesellschaft 44:2030–2036.
- 14. Cox ED and Cook JM (1995) The Pictet-Spengler condensation: a new direction for an old reaction. Chem. Rev 95:1797–1842.
- 15. Buchi G and Mak C-P (1977) Nitro olefination of indoles and some substituted benzenes with 1 dimethylamino-2-nitroethylene. J. Org. Chem 42:1784–1786.
- 16. Ranjan P, Ojeda GM, Sharma UK and Van der Eycken EV (2019) Metal-free dearomatization: Direct access to spiroindol(en)ines in batch and continuous-flow. Chem. Eur. J 25:2442–2446. [PubMed: 30507048]
- 17. Becke AD (1993) Density-functional thermochemistry. III. The role of exact exchange. J. Phys. Chem 98:5648–5652.
- 18. Lee CJ, Wang W and Parr RG (1988) Development of Colle-Selvatti correlation-energy formula into a function of the electron density. Phys. Rev. B: Condens. Matter Maeter. Phys 37:785–789.
- 19. Miehlich B, Savin A, Stoll H and Preuss H (1989) Results obtained with the correlation energy density functionals of Becke and Lee, Yang and Parr. Chem. Phys. Lett 157:200–206.
- 20. Cammi R, Mennucci B and Tomasi J, In computational chemistry; Reviews of current trends, in In computational chemistry; Reviews of current trends., Leszczynski J, Editor. 2003, World Scientific Publishing Co. Pte. Ltd.: Singapore.
- 21. Grimme S, Antony J, Ehrlich S and Krieg H (2010) A consistent and accurante ab initio parameterization of density functional dispersion correction (DFT-D) fir the 94 elements H-Pu. J. Chem. Phys 132:154104. [PubMed: 20423165]

- 22. Grimme S, Ehrlich S and Goerigk L (2011) Effect of damping function in dispersion corrected density functional theory. J. Comp. Chem 132:1456.
- 23. Lee JK and Tantillo DJ (2003) Computational insights into the mechanism of orotidine monophosphate decarboxylase. Adv. Phys. Org. Chem 38:183–218.
- 24. Jursic BS (1996) Computation of electronic affinities of O and F atoms, and energy profile of F-H2 reaction by densiy functional theory. J. Chem. Phys 104:4151.
- 25. Humphrey W, Dalke A and Schulten K (1996) VMD Visual Molecular Dynamics. J. Mol. Graphics 14:33–38.
- 26. Sharp K and Matschinsky F (2015) Translation of Ludwig Boltzmann's paper "on the relationship between second fundamental theorem of the mechanical theory of heat and probability calculations regarding teh conditions for thermal equilibrium" Sitzungberichte der Kaiserlichen Akademie der Wissenschaften. Mathematisch-Naturwissen Classe. Abt II, LXXVI 1877, pp 373–435 Entropy 17:1971–2009.
- 27. Xu X, Ge R, Li L, Wang J, Lu X, Xue S, Chen X, Li Z and Bian J (2018) Exploring the tetrahydroisoquinoline thiohydantoin scaffold blockade of the androgen receptor as potent antiprostate cancer agents. Eur. J. Med. Chem 143:1325–1344. [PubMed: 29117897]
- 28. Harrison JR, O'Brien P, Porter DW and Smith NM (1999) Studies towards the preparation of spartine-like diamines for asymetric synthesis. J. Chem. Soc 24:3623–3631.
- 29. Marenich AV and Truhlar DG (2009) Universal solvation model based on solute electron density and on a continuum model of solvent defined by the bulk dielectric constant and atomic surface tensions. J. Phys. Chem. B 113:6378–6396. [PubMed: 19366259]
- 30. Grimblat N and Sarotti AM (2016) Computational chemistry to the rescue: modern toolboxes for the assignment of complex molecules by GIAO NMR calculations. Chem. Eur. J 22:12246–12261. [PubMed: 27405775]
- 31. Adamo C and Barone V (1998) Exchange functionals with improved long-range behavior and adiabetic connection methods without adjustable parameters: The mPW and mPW1PW models. J. Chem, Phys 108:664–675.
- 32. Lodewyk MW, Siebert MR and Tantillo DJ (2012) Computational prediction of 1H and 13C chemical shifts: a useful tool for natural products, mechanistic, and synthetic organic chemistry. Chem. Rev 112:1839–1862. [PubMed: 22091891]
- 33. Willoughby PH, Jansma MJ and Hoye TR (2014) A guide to small-molecule structure assignment through computation of (1H and 13C) NMR chemical shifts. Nat. Protoc 9:643–660. [PubMed: 24556787]
- 34. Van der Plas SE, Kelgtermans H, De Munck T, Martina SLX, Dropsit S, Quinton E, De Blieck A, Joannesse C, Tomaskovic L, Jans M, Christophe T, Van der Aar E, Borgonovi M, Nelles L, Gees M, Stouten PFW, Van Der Schueren J, Mammoliti O, Conrath K and Andres MJ (2018) Discovery of N-(3-carbamoyl-5,5,7,7-tetramethyl-5,7-dihydro-4H-thieno[2,3-c]pyran-2-yl)-1H-pyrazole-5 carboxamide (GLPG1837), a novel potentiator which can open class III mutant cystic fibrosis transmembrane conductance regulator (CFTR) channels to a high extent. J. Med. Chem 61:1425– 1435. [PubMed: 29148763]
- 35. Yeh HI, Qui L, Sohma Y, Conrath K, Zou X and Hwang TC (2019) Identifying the molecular target sites for CFTR potentiators GLPG1837 and VX-770. J. Gen. Physiol 151:912–928. [PubMed: 31164398]
- 36. Liu F, Zhang Z, Levit A, Levring J, Touhara KK, Shoichet BK and Chen J (2019) Structural identification of a hotspot on CFTR for potentiation. Science 364:1184–1188. [PubMed: 31221859]
- 37. Molinski SV, Gonska T, Huan LJ, Baskin B, Janahi IA, Ray PN and Bear CE (2014) Genetic, cell biological, and clinical interrogation of the CFTR mutation c.3700 A>G (p.Ile1234V) informs of strategies for future medical intervention. Genetics Med. 16:625–632. [PubMed: 24556927]
- 38. Molinski SV, Ahmadi S, Ip W, Ouyang H, Villella A, Miller JP, Lee P-S, Kulleperuma K, Du K, Di Paola M, Eckford PDW, Laselva O, Huan LJ, Wellhauser L, Li E, Ray PN, Pomes R, Moraes T, Gonska T, Ratjen F and Bear CE (2017) Orkambi and amplifier co-therapy improves function from a rare CFTR mutation in gene-edited cells and patient tissue. EMBO Mol. Med 9:1224–1243. [PubMed: 28667089]

- 39. Valley HC, Bukis KM, Bell A, Cheng Y, Wong E, Jordan NJ, Allaire NE, Sivachenko A, Liang F, Bihler H, Thomas PJ, Mahlou J and Mense M (2018) Isogenic cell models of cystic-fibrosiscausing variants in natively expressing pulmonary epithelial cells. J. Cyst. Fibros 18:476–483. [PubMed: 30563749]
- 40. Sharma J, Keeling KM and Rowe SM (2020) Pharmacological approaches fror targeting cystic fibrosis nonsense mutations. Eur. J. Med. Chem 299:epub.

HIGHLIGHTS:

- **•** Approximately 10% of cystic fibrosis subject remain without CFTR modulator therapy
- **•** Co-potentiators represent a novel class of CFTR modulators
- **•** Co-potentiators act in synergy with potentiators to activate rare CFTR mutants
- **•** Spiro[piperidine-4,1-pyrido[3,4-b]indole] co-potentiators were optimized by structure-activity studies
- Spiro[piperidine-4,1-pyrido[3,4-b]indoles] with EC₅₀ down to 600 nM, an approximately 17-fold improvement, were identified

Figure 1. Structural variants 1–6 of the spiro[piperidine-4,1'-pyrido[3,4- ^b]indole] scaffold.

Figure 2.

Optimized geometries of the lowest energy conformers of **2a**, **2c**, **2e** and **4a**–**4c**.Distances (\AA) are measured from the centroid of *Ring i*, to those of *Ring ii* and *Ring iii*. The lowest energy conformers for 4a-c preferred a close from conformation, where one face of Ring i is blocked from potential interactions with the protein interior.

Son et al. Page 28

 $2a$

 $2c$

2e

Figure 4.

Structures of **6a** and **6b**. Based on quantum calculations at SMD(chloroform)-B3LYP-D3(BJ)/6–31+G(d,p). **6b** is 2.33 kcal/mol more thermodynamically favorable than **6a**.

Son et al. Page 30

Figure 5.

Lowest energy conformers optimized at SMD(chloroform)6–31+G(d,p) with D3(BJ) for **6a** and 6b respectively (top); the distance between centroids of *Rings i* and *ii* and *Rings i* and *iii* were measured. Superimposed images of energetically relevant conformers for **6a** and **6b** (bottom); **6a** has more open form conformers in equilibrium.

Figure 6.

Structural determinants of spiro[piperidine-4,1'-pyrido[3,4-b]indole] scaffold. See text for explanation.

Figure 7.

Short-circuit current measurement of mutant CFTR activation by spiro[piperidine-4,1 pyrido[3,4-b]indoles]. A. (left) Short-circuit current data in FRT cells expressing N1303K-CFTR in response to 20 μM forskolin (fsk), 5 μM VX-770, indicated concentration of **2i** and **2e**, and 10 μM CFTR_{inh}-172. (right) Summary of concentration-dependence data (n=3, mean ± S.E.M.). B. Measurements done as in A, but with 20 μM GLPG1837 instead of VX-770. ^C. Short-circuit current in FRT cells expressing I1234del-CFTR in response to 20 μM forskolin, 5 μM VX-770, indicated concentration of **2i**, and 10 μM CFTRinh-172. D. Shortcircuit current in FRT cells expressing R347P-CFTR in response to 20 μM forskolin, 5 μM VX-770, 20 μM **2i** and 10 μM CFTRinh-172. In all studies, cells were corrected with 3 μM VX-661 for 18–24 hours prior to measurement. All traces are representative of three replicates.

Figure 8.

Activity of **2i** in human airway epithelial cell cultures. A. Short-circuit current in geneedited 16HBE14o- cells expressing N1303K-CFTR. B. Short-circuit current data in geneedited 16HBE14o- cells expressing I1234del-CFTR. C. Summary of changes in short-circuit current (Isc; mean \pm S.E.M., n = 3, *P < 0.05). Concentrations: 20 μM forskolin, 5 μM VX-770, 10 μM **2i**, and 10 μM CFTRinh-172. 16HBE14o- cell models expressing I1234del-CFTR were corrected with 18 μM VX-661 and 3 μM VX-445 for 18–24 hours prior to measurement.

Scheme #1.

Synthetic route to spiro[piperidine-4,1'-pyrido[3,4-b]indole] analogs 1–6.

Scheme #2. Synthesis of constrained analog **6** .

Table 1.

Predicted Boltzmann distributions for Open vs. Closed forms for 2a, 2c, 2e, and 4a–4c at (PCM)chloroform-B3LYP-D3(BJ)//6–31+G(d,p) (left) and (PCM)chloroform-B3LYP-D3(BJ)//6–331+G(2d,2p) (right).

Table 2.

Calculated 13C and 1H chemical shifts for **6a** and **6b**, and their deviation from experiments.

* Abbreviation: MAD, mean absolute deviation; and Dev., deviation. Deviations of less than 6 ppm for ¹³C and less than 0.3 ppm for ¹H shifts are considered acceptable [30, 32, 33]. Chemical shifts exceeding the accepted deviations are bolded above. Coupling constants (J) were also calculated to confirm the identity of complex proton peaks (see SI for details).

Table 3.

Bioassay results.

Biology: Characterization of spiro[piperidine-4,1-pyrido[3,4-b]indoles] co-potentiators