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Infammation in Chemotherapy‑Induced Cardiotoxicity

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Abstract

Purpose of Review In this review we describe the role of infammation in chemotherapy-induced cardiotoxicity with a particular focus on anthracycline-induced cardiomyopathy (AIC). First, we discuss infammation associated with anthracyclines at a cellular level. Next, we discuss the clinical implications of these infammatory mechanisms for early detection and cardioprotective strategies in patients undergoing anthracycline treatment.

Recent Findings Key infammatory pathways identifed in AIC include cytokine release, upregulation of the innate immune system via toll-like receptors, and activation of the infammasome. Emerging evidence suggests a role for infammatory biomarkers in detecting subclinical AIC. Advanced imaging techniques, such as cardiac PET with novel tracers targeting infammation, may enhance early detection. Both traditional cardioprotective strategies and novel anti-infammatory therapies show potential in preventing and treating AIC.

Summary Understanding the infammatory mechanisms involved in AIC provides new opportunities for early detection and targeted cardioprotective strategies in patients undergoing anthracycline treatment and informs our understanding of other forms of chemotherapy-induced cardiotoxicity.

Keywords Cardio-oncology · Anthracyclines · Cancer treatment related cardiac dysfunction · Infammation · Doxorubicin · Cardioprotection

Introduction

Advances in oncologic care have rapidly reduced mortality in cancer patients. With improved survival there is increased focus on cardiovascular side efects of cancer treatments, collectively termed cancer therapy related cardiac dysfunction (CTRCD). Understanding the underlying mechanisms of CTRCD is critical for developing improved cardioprotective protocols and novel, safer treatments in the future. Additionally, as the mechanisms of cardiotoxicity are better understood, they provide important insights into myocardial physiology and particularly myocardial responses to stressors.

Infammation is a complex set of biological responses to harmful stimuli, acting as an adaptive mechanism that mobilizes the immune system to counter these threats. However, prolonged or chronic infammation can become maladaptive, increasing the risk of disease. Infammation underlies many forms of CTRCD and, in particular, plays a crucial role in the development of cardiomyopathy from anthracyclines, known as anthracycline-induced cardiomyopathy (AIC). In this article, we describe the role of infammation in AIC, which is the most well-characterized cause of CTRCD. This is summarized in Fig. [1](#page-2-0)**.** First, we examine the biologic mechanisms of anthracycline-induced infammation at a cellular level. Next, we discuss the clinical evidence for AICinduced infammation, focusing on laboratory biomarkers and cardiac imaging. Finally, we discuss the role of medications in targeting infammation associated with AIC. Understanding the role of infammation in AIC provides insight into all forms of CTRCD and has the potential to improve the long-term cardiovascular health of patients with cancer.

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Fig. 1 Anthracycline-induced Infammation: Mechanisms, Diagnostics, Cardioprotection. Abbreviations: TOP2B: Topoisomerase II beta; ROS: Reactive Oxygen Species; CRP: C-reactive Protein; TNF-α: Tumor Necrosis Factor-alpha; IL-6: Interleukin-6; MMP-2: Matrix Metalloproteinase-2; MMP-9: Matrix Metalloproteinase-9; MPO: Myeloperoxidase; CASP-1: Caspase-1; GDF-15: Growth Diferentiation Factor 15; PIGF: placental growth factor; GLS: Global Longitudinal Strain; cMRI: Cardiac Magnetic Resonance Imaging; PET: Positron Emission Tomography; ACEi: Angiotensin-Converting Enzyme Inhibitor; ARB: Angiotensin Receptor Blocker; NLRP3: Nucleotide-binding Oligomerization Domain-like Receptor Family Pyrin Domain Containing 3

Biological Mechanisms of Infammation in Anthracycline‑Induced Cardiomyopathy (AIC)

Anthracyclines are potent cytotoxic antibiotics integral to many anti-cancer regimens. However, they are associated with a dose-dependent risk of cardiomyopathy in a signifcant subset of patients. For instance, a recent cohort study reported a 9% incidence of cardiotoxicity with a median follow-up of 5.2 years [[1\]](#page-10-0). The risk is dose-dependent, with a 26% incidence of cardiotoxicity when the dose exceeds 550 mg/m² [[2\]](#page-10-1); currently any dose above 240 mg/m² is considered high-risk for development of AIC [[3\]](#page-10-2). Early studies of AIC primarily focused on oxidative stress; however, the pathophysiology of AIC involves multiple interconnected cellular mechanisms, with infammation playing a crucial role. The detailed mechanisms are outlined below and depicted in Fig. [2.](#page-3-0) A thorough understanding of these pathways may reveal strategies for the early detection and treatment of AIC.

Direct Cellular Cardiotoxic Efects of Anthracyclines

Doxorubicin, the most widely used anthracycline, accumulates at a cellular level primarily in the nuclei and mitochondria [[4](#page-10-3)]. The primary nuclear target of doxorubicin is topoisomerase-IIβ (TOP2B) [[5](#page-10-4)], an enzyme that prevents nuclear DNA breaks. When doxorubicin binds to TOP2B, it inhibits the enzyme's ability to bind to DNA and prevent ligation. Additionally, doxorubicin intercalates with DNA strands, leading to DNA breaks [[6](#page-10-5)]. The resultant DNA damage activates pro-apoptotic signaling pathways, notably those involving p53, resulting in cell apoptosis [[5,](#page-10-4) [7\]](#page-10-6).

In the mitochondria, doxorubicin binds to complex I (NADH dehydrogenase) of the electron transport chain [[8,](#page-10-7) [9](#page-10-8)], and to mitochondrial free iron creating highly reactive iron-anthracycline complexes [\[10\]](#page-10-9). Additionally, doxoru-bicin intercalates with mitochondrial DNA (mtDNA) [\[11](#page-10-10)]. These interactions stimulate excessive redox cycling and disrupt mitochondrial function, leading to the uncoupling of the electron transport chain and the formation of reactive

Fig. 2 Mechanisms of Anthracycline-Induced Infammation. Anthracyclines exert their toxic effects primarily by inhibiting topoisomerase II beta (TOP2B) and intercalating with DNA, leading to double-strand DNA (dsDNA) breaks and the activation of the tumor suppressor protein p53, which promotes immunogenic apoptosis. Anthracyclines also disrupt mitochondrial function by binding to complex I (NADH dehydrogenase) and mitochondrial DNA (mtDNA), leading to excessive reactive oxygen species (ROS) production, lipid peroxidation, and mitochondrial damage. The generation of ROS triggers the activation of the NLRP3 infammasome and the release of infammatory cytokines. These cytokines, in turn, recruit leukocytes to the myocardium and further exacerbate infammation. Additionally, anthracyclines induce the release of damageassociated molecular patterns (DAMPs), which activate Toll-like

oxygen species (ROS) [[12](#page-10-11)]. ROS cause oxidative stress, leading to damage to the mitochondria through processes such as lipid peroxidation of the mitochondrial membrane [\[10\]](#page-10-9) [[13](#page-10-12)]. Severe oxidative damage and ATP depletion can result in cardiomyocyte death.

Activation of the Immune System by Anthracyclines

The actions of doxorubicin in the nucleus and mitochondria are inextricably linked with cardiac infammation via activation of the immune system. Severe oxidative damage and ATP depletion, as described above, result in cell death characterized by cell swelling, membrane rupture, and receptors (TLRs) and lead to the nuclear translocation of NF-κB, driving the transcription of pro-infammatory cytokines. The inhibition of cyclooxygenase-2 (COX-2) and lipoxygenase (LOX) by anthracyclines further diminishes anti-infammatory defenses, amplifying the infammatory response. Abbreviations: TOP2B: Topoisomerase II beta; ROS: Reactive Oxygen Species; mtDNA: Mitochondrial DNA; NLRP3: Nucleotide-binding Oligomerization Domain-like Receptor Family Pyrin Domain Containing 3; IL-1β: Interleukin-1 beta; IL-6: Interleukin-6; IL-18: Interleukin-18; TNF-α: Tumor Necrosis Factor-alpha; DAMPs: Damage-Associated Molecular Patterns; TLRs: Toll-like Receptors; NF-κB: Nuclear Factor kappa-light-chain-enhancer of activated B cells; dsDNA: Double-Strand DNA; p53: Tumor Suppressor Protein p53; COX-2: Cyclooxygenase-2; LOX: Lipoxygenase

infammation [[14](#page-10-13)]. While traditional forms of apoptosis do not induce an infammatory response; doxorubicin induces immunogenic forms of cell death, including pyroptosis (discussed below) and necrosis, which trigger an infammatory response from the immune system [[14](#page-10-13)] [[15](#page-10-14)]. ROS and oxidative stress also trigger an infammatory response through the production of infammatory cytokines [[16](#page-10-15)]. These cytokines attract immune cells, such as macrophages and neutrophils, to the myocardium [[17](#page-10-16)]. These immune cells release additional infammatory mediators, ROS, and proteolytic enzymes, exacerbating myocardial injury.

Doxorubicin-induced cell death leads to the production of key infammatory mediators, including damage-associated

molecular patterns (DAMPs) and endotoxins. DAMPs are signals of host cellular distress [[18\]](#page-10-17), released by both doxorubicin-killed tumor cells [\[19](#page-10-18)] as well as macrophages activated by doxorubicin-induced infammation [[20](#page-10-19)] [[21](#page-10-20)]. Endotoxins are toxins released by gram-negative bacteria, and have been shown to enter the circulation due to doxorubicin-induced increases in intestinal permeability [[22\]](#page-10-21). DAMPs and endotoxins upregulate and activate Toll-like receptors (TLRs) [\[23](#page-10-22)], which are key mediators of the innate immune system's infammatory response. Activation of the TLR intracellular signaling leads to nuclear localization of pro-infammatory transcription factor NF-kB [[24](#page-10-23)], which drives the transcription of genes encoding pro-infammatory cytokines, namely tumor necrosis factor-alpha (TNF-α), interleukin-1 beta (IL-1β), interleukin-6 (IL-6), and interleu-kin-18 (IL-18) [\[25](#page-10-24)]. TNF- α further amplifies this response by stimulating NF-kB in a feed-forward mechanism [[26\]](#page-10-25). In animal models, both TLR2 knockout and antagonism have been shown to attenuate doxorubicin-induced infammation and subsequent cardiac injury [\[20](#page-10-19), [23](#page-10-22)].

Another key mechanism of inflammation in AIC is increased activation of the NOD-like receptor protein 3 (NLRP3) inflammasome in cardiomyocytes by doxorubicin [[27\]](#page-11-0). The NLRP3 inflammasome is a large tripartite protein consisting of an intracellular sensor (NLRP3), an adaptor, and an effector (caspase 1) [[28\]](#page-11-1). The inflammasome functions through two critical steps, priming and activation. Priming leads to upregulation of the expression of component proteins, which is typically triggered by DAMP and pathogen-associated molecular patterns (PAMP) binding to TLRs. Next, activation triggers oligomerization into the NLRP3 inflammasome to activate caspase 1, which in turn cleaves pro-IL-1 β and pro-IL-18 leading to inflammatory cytokine release [[29](#page-11-2)]. The inflammasome is activated by a wide range of stimuli, particularly products of cellular stress, including release of ROS. Doxorubicin has been shown in preclinical models to increase activation of the NLRP3 inflammasome [\[27\]](#page-11-0). In addition to direct release of cytokines, the NLRP3 signaling pathway leads to an inflammatory programmed cell death called pyroptosis, leading to release of inflammatory factors and cell contents [[30](#page-11-3)]. Pyroptosis is an important mechanism for the anticancer properties of anthracyclines but contributes to the off-target side effects. Through this form of cell death, release of DAMPs and cytokines cause an inflammatory cycle that leads to further myocardial damage and cardiotoxicity. In addition, activation of the inflammasome inhibits cardiomyocyte autophagy, a non-inflammatory, protective form of cell death [[31](#page-11-4)].

In addition to triggering the immune system as described above, anthracyclines can inhibit the natural anti-inflammatory mechanisms within the myocardium.

Anthracyclines inhibit cyclooxygenase and lipoxygenase enzymes, reducing the availability of anti-inflammatory mediators like prostacyclins in the heart. This reduction of anti-inflammatory protection mechanisms exacerbates inflammation and contributes to cardiac damage associated with anthracyclines [\[32\]](#page-11-5).

Myocardial Efects of Infammation

While myocyte death from the above-described mechanisms lead to a significant change in myocardial function, inflammatory cytokines also lead to myocyte dysfunction in the surviving myocytes. Inflammatory cytokines, including TNF- α , IFN- γ , IL-6, IL-8 and IL-1 β cause changes in cytoskeleton structure and mitochondrial function, which can significantly affect the contractile properties without inducing cell death, which was previously thought to be the driver of cardiac dysfunction [\[19,](#page-10-18) [33\]](#page-11-6). This is similar to the well described mechanism of sepsis-induced cardiomyopathy and cardiotoxicity seen in cytokine release syndrome with chimeric antigen receptor (CAR) T-cell therapy [[34](#page-11-7)].

Novel Methods of Detection for Cardiotoxicity

The emerging understanding of the role of infammation in CTRCD offers promising new approaches to the early detection and treatment of cardiotoxicity. The current mainstay of detection of cardiotoxicity is serial echocardiography with global longitudinal strain (GLS) [[3](#page-10-2)]. In current practice, there is also a role for laboratory biomarkers, particularly troponin and B-type natriuretic peptide (BNP), for additional monitoring in patients at increased risk for cardiotoxicity. Notably, high sensitivity troponin T has been found to be increased in patients who develop CTRCD from anthracyclines and trastuzumab [\[35\]](#page-11-8). As our understanding of the infammatory underpinnings of AIC grows, there is potential for more advanced laboratory biomarkers to detect AIC in its subclinical state. Similarly, with advancements in imaging techniques, there is new opportunity to develop earlier markers of cardiotoxicity to ensure prompt detection and treatment. However, it is important to note that when detecting infammation in AIC, imaging and cardiac biomarkers are not specifc to AIC and can represent infammation from other sources, including tumor itself. Increased understanding of the role of infammation in AIC at a cellular level, beyond what is described above, may open possibilities for additional clinical diagnostic methods.

Infammatory Biomarkers

Biomarkers are an important potential avenue for the early diagnosis of CTRCD as they may provide a cost efective and reproducible method of early detection. However, the utility of infammatory biomarkers in detection of cardiotoxicity is complex and signifcantly confounded by the overall infammatory milieu of the underlying malignancy. Preclinical and clinical studies have demonstrated that, even prior to the initiation of anthracycline, plasma levels of infammatory markers, such as cytokines and metalloproteinases, are elevated in the presence of tumor. In a mouse model of colorectal cancer, elevated IL-6 and IL-8 correlated with myocardial dysfunction even prior to doxorubicin administration [\[33](#page-11-6)]. Similarly, a clinical study of patients with breast cancer demonstrated that infammatory markers, including myeloperoxidase (MPO), IL-6, and matrix metalloproteases (MMP-2 and MMP-9), were signifcantly elevated compared to normal controls prior to anthracycline administration [[36](#page-11-9)].

The literature presents mixed fndings regarding serum biomarker changes after the initiation of anthracycline therapy. While some biomarkers increase post-anthracycline chemotherapy and potentially signal cardiotoxicity, others decrease. In the previously mentioned study, levels of TOP2β, MPO, MMP-2, MMP-9 and IL-6 increased from baseline in patients receiving anthracyclines at 3 and 6-months post-treatment. Interestingly, these levels correlated with high sensitivity troponin T and I, but not with changes in left ventricular ejection fraction (LVEF), though the study may have been underpowered to detect such a change [[36\]](#page-11-9). In another prospective study of 46 women with breast cancer requiring anthracyclines or trastuzumab, high sensitivity C-reactive protein (hsCRP) increased over a treatment duration of 4 months [[37](#page-11-10)]. In a multicenter prospective study of 78 patients with breast cancer undergoing doxorubicin and trastuzumab therapy, troponin I (TnI), C-Reactive protein (CRP), growth diferentiation factor 15 (GDF-15), MPO, placental growth factor (PIGF) and soluble Fms-like tyrosine kinase-1 (sFlt— 1) levels increased with anthracycline therapy. However, only elevated levels of TnI and MPO were associated with an increased risk of cardiotoxicity [[38](#page-11-11)]. Conversely, in a prospective study of patients undergoing doxorubicin-containing chemotherapeutic regimens $(n=41)$, noncardiac biomarkers CASP-1 and MPO decreased after administration of chemotherapy, particularly in the high-dose anthracycline subgroup, but were not associated with change in

Study	Study Type	Population	Biomarkers	Change Post- Anthracycline Treatment	Association with Car- diotoxicity
Dessi et al. 2013 [40]	Randomized con- trolled trial	Cancer patients on epi- IL-6, TNF- α , serum rubicin randomized to treatment with telmisartan $(n=25)$ versus placebo $(n=24)$	ROS	\uparrow — placebo \leftrightarrow —telmisartan	Correlation between strain rate on echo- cardiography and serum IL-6 and ROS observed
Grover et al. 2013 [37] Prospective cohort	trial	Breast cancer patients on anthracyclines and/or trastuzumab $(n=46)$	hsCRP		No direct cardiotoxicity link reported
Ky et al. 2014 [38]	Prospective cohort trial	Breast cancer patients on anthracyclines and trastuzumab $(n=78)$	TnI, CRP, GDF-15, MPO, PIGF, sFlt-1	↑	TnI and MPO were associated with increased cardiotoxic- ity risk
Lakani et al. 2021 [36]	Prospective cohort trial	Breast cancer patients $(n=17)$, matched healthy controls $(n=17)$	ТОР2В, МРО, ММР- 2, MMP-9, IL-6,	\uparrow	Biomarkers correlated with high sensitivity troponin T and I; no direct association with LVEF change was observed
Dean et al. 2023 [39]	Prospective cohort trial	Cancer patients $(n=41)$ undergoing anthracycline-based treatment	CASP-1, MPO		Decrease in biomarkers was not associated with change in LVEF

Table 1 Summary of Clinical Studies Investigating Infammatory Biomarkers in Patients Undergoing Anthracycline-Based Therapy

TOP2β Topoisomerase II beta, *MPO* Myeloperoxidase, *MMP-2* Matrix Metalloproteinase-2, *MMP-9* Matrix Metalloproteinase-9, *IL-6* Interleukin-6, *hsCRP* High-sensitivity C-reactive protein, *TnI* Troponin I, *GDF-15* Growth Diferentiation Factor 15, *PIGF* Placental Growth Factor, *sFlt-1* Soluble Fms-like Tyrosine Kinase-1, *IL-6* Interleukin-6, *TNF-α* Tumor Necrosis Factor-alpha, *ROS* Reactive Oxygen Species, *CASP-1* Caspase-1, *LVEF* Left Ventricular Ejection Fraction

LVEF [[39\]](#page-11-13). A small number of clinical trials have looked at infammatory markers, particularly IL-6 and ROS, and found that pharmacologic cardioprotection correlated with decreased levels of infammatory biomarkers compared to placebo groups [[40](#page-11-12)]. It is theorized that, as previously seen in preclinical studies, there is increased infammation in the setting of malignancy that is reduced with chemotherapeutic treatment. Therefore, these serum markers of infammation are inextricably linked to both oncologic response and cardiotoxicity. See Table [1](#page-5-0) for a detailed summary of studies evaluating the use of infammatory markers for detection of AIC. Further large-scale studies of multiple infammatory biomarkers are needed to identify potential clinical biomarkers more specifc to AIC.

Targeted Imaging of Infammation

Traditional imaging of AIC has been primary focused on changes to LVEF. More recently, changes in GLS have been found to be important markers of subclinical cardiac dysfunction and have entered the guideline defnition of AIC [\[3\]](#page-10-2). However, more specifc imaging of infammation may provide an additional avenue for targeted early detection. In cardiac MRI, an important marker of infammation can be myocardial edema. Myocardial edema on CMR has been found after anthracycline therapy in about a third of patients at one month and nearly a half of patients at four months. However, this has not been correlated to change in LVEF [[37](#page-11-10)]. Cardiac positron emission tomography (PET) is a promising tool due to the ability to use various radioactive tracers to target specifc mechanisms of cardiotoxicity. Indeed, cardiac PET is a well-established tool for diagnosis of infammatory cardiomyopathies, particularly cardiac sarcoid [\[41](#page-11-14)]. In this modality, 18-fluorodeoxyglucose $(^{18}F-$ FDG) PET detects regions of increased glucose metabolism which can signal infammatory activity. In AIC, the impaired mitochondrial phosphorylation and oxidation drives myocytes towards increased glucose metabolism. Increased ¹⁸F-FDG uptake after anthracycline therapy has been demonstrated in preclinical and retrospective clinical studies. However, prospective studies have not been performed and correlation with cardiotoxicity specifcally has not been established. More targeted PET and SPECT tracers are being developed to image specifc mechanisms of AIC, including apoptosis (99mTechnetium-annexin-V) and myocardial necrosis $\binom{111}{1}$ n-antimyosin)[[42\]](#page-11-15).

Advances in Prevention of AIC: Role of Infammation

Given the signifcant morbidity and mortality related to AIC, it is important to identify potential pharmacologic treatments for both cardioprotection and treatment of AIC. Understanding the role of infammation in this process opens the possibility of novel treatment targets. In some cases, traditional cardioprotective medications have additional pathways that specifcally target the mechanisms of AIC, including infammatory pathways; however, clinical trial data has been mixed.

Role of Infammation in Conventional Cardioprotective Therapies for AIC

Dexrazoxane

Dexrazoxane, a water-soluble analog of the iron chelating agent ethylenediaminetetraacetic acid (EDTA), was the frst FDA-approved treatment for prevention of AIC. Cardioprotective efects of dexrazoxane were initially thought to be via iron chelating efects of the metabolize ADR-925; however, studies of isolated ADR-925 do not reduce cardiotoxicity and inhibition of conversion from dexrazoxane to ADR-925 do not attenuate the cardioprotective effects of dexrazoxane $[43-45]$ $[43-45]$ $[43-45]$. Further studies showed that dexrazoxane depletes TOP2B as a primary mechanism of cardioprotection, which as previously described plays an important role in the infammatory cascade [\[44,](#page-11-18) [46](#page-11-19)]. Attenuation of TOP2B prevents DNA damage, p53 activation and eventually cell apoptosis. Specifc anti-infammatory pathways have not been identifed for cardiomyocytes, however, in a Parkinson's Disease mouse model, dexrazoxane was found to suppress local and systemic infammation, as measured by levels of TNF- α and IL-1 β [\[47\]](#page-11-20).

Cardioprotective efects of dexrazoxane during anthracycline therapy have been demonstrated in multiple clinical trials, particularly in children with hematologic malignancies and adults with breast cancer [\[48–](#page-11-21)[52](#page-11-22)]. Based on the strength of this data, the FDA approved the use of dexrazoxane specifically in patients with advanced or metastatic breast cancer after a cumulative dose greater than 300 mg/m² of doxorubicin equivalent. Unfortunately, early studies suggested an increased risk of secondary malignancy and concern for reduced antitumor efficacy [[53\]](#page-11-23), which has not been reproduced in future studies [\[54,](#page-11-24) [55\]](#page-11-25). Importantly, a large meta-analyses of breast cancer patients undergoing anthracycline-based chemotherapy found that administration of dexrazoxane lead to lower rates of heart failure without a detrimental efect on cancer

outcomes [[56\]](#page-11-26). These studies did not specifcally evaluate for anti-infammatory efects.

Angiotensin Converting Enzyme Inhibitors (ACEi) and Angiotensin Receptor Blockers (ARBs)

Preclinical studies of angiotensin converting enzyme inhibitors (ACEi) and angiotensin receptor blockers (ARB) have been promising in providing cardioprotection against and treatment of AIC, and in particular its associated infammation. Doxorubicin increases plasma angiotensin II and local myocardial ACE, which cause direct cardiotoxicity [[57](#page-11-27)]. In rodent models, ACEi and ARBs decreased ROS production, decreased apoptosis, reduced the risk of heart failure, and improved mortality [[57](#page-11-27), [58](#page-11-28)]. However, larger clinical trials of ACEi and ARBs have produced mixed results. In a placebo-controlled study of telmisartan, treatment with telmisartan led to decreased levels of IL-6 and ROS, and mitigated changes in myocardial strain at high doses of anthracyclines (>300 mg/ m²) [[40,](#page-11-12) [59](#page-11-29)]. The PRADA (Prevention of Cardiac Dysfunction During Adjuvant Breast Cancer Therapy) trial was a 2 × 2 factorial, randomized placebo-controlled trial of monotherapy and combined candesartan and metoprolol succinate during epirubicin therapy in breast cancer patients. Early results showed candesartan prevented a modest reduction in LVEF compared to other groups [[60](#page-11-30)], but this did not persist at two year follow up [[61\]](#page-11-31).

Beta Blockers

Beta blockers are a mainstay of treatment for heart failure primarily due to their role in reducing sympathetic activation and neurohormonal upregulation; however, certain beta blockers also have anti-infammatory and antioxidant efect that increase cardioprotective efects in AIC. In particular, carvedilol and nebivolol have antioxidant properties that reduce ROS [\[62](#page-11-32)]. Bisoprolol and carvedilol reduce infammation and ROS in other etiologies of heart failure with reduced LVEF [[63](#page-12-0)]. A similar anti-inflammatory and antioxidant efect has not been seen with metoprolol. Multiple clinical trials have evaluated cardioprotective efects of various beta blockers, showing a modest beneft to beta blocker therapy, particularly with carvedilol, nebivolol and bisoprolol. In the PRADA study, metoprolol had no efficacy in the prevention of AIC in patients with early breast cancer [\[61](#page-11-31)]. In the CECCY (Carvedilol Effect in Preventing Chemotherapy-Induced Cardiotoxicity) trial, the use of carvedilol in women with HER2 negative breast cancer did not lead to a signifcant diference in the clinical incidence of AIC at six months compared to placebo, but there was a decrease in subclinical markers of cardiac dysfunction including abnormal troponin values and diastolic dysfunction [[64\]](#page-12-1). The OVERCOME trial demonstrated that, in patients with hematologic cancers undergoing high dose anthracycline chemotherapy with possible stem cell transplant, combined enalapril and carvedilol prevented a reduction in LVEF compared to placebo, [[65](#page-12-2)] which was also seen with combined bisoprolol and lisinopril [\[66\]](#page-12-3). In a breast cancer population, nebivolol prevented a change in left ventricular end-systolic and end-diastolic diameters, compared to placebo [\[67](#page-12-4)]. These results indicate that beta blocker therapy, particularly with carvedilol, nebivolol and bisoprolol, do not prevent clinical incidence of anthracycline-induced cardiotoxicity (change in LVEF>10% from baseline), but can prevent subclinical changes. Limitations to these studies include small sample sizes, short follow up, low frequency of the outcome, and variations in the patient population and anthracycline exposure.

Statins

Statins have pleiotropic effects beyond traditional lipid lowering mechanisms, including anti-infammatory and antioxidant properties. In a mouse model of AIC, pretreatment with fuvastatin preserved LV function, attenuated oxidative stress, increased expression of antioxidant enzymes, reduced cardiac infammation via TNF-α expression, and decreased apoptosis compared to controls [[68](#page-12-5)]. Statins also inhibit small Ras homologous (Rho) GTPases, which play a critical downstream role in infammation as part of the NADPH oxidase complex and stimulation of a pro-infammatory process [[69\]](#page-12-6). These studies suggest that the cardioprotective efects of statin therapy are mediated by anti-infammatory, antioxidant, and anti-apoptotic mechanisms [[68,](#page-12-5) [70](#page-12-7)]. This proposed mechanistic efect was frst confrmed with early retrospective studies that suggested that statins prevented reduction in LVEF [[71,](#page-12-8) [72](#page-12-9)]. More recent randomized, placebo-controlled trials have had mixed results. In a study of high risk patients undergoing anthracycline chemotherapy for hematologic malignancies, high-dose atorvastatin prevented a reduction on LVEF compared to placebo [\[73\]](#page-12-10). Notably, an elevation in CRP after initiation of chemotherapy was observed in the control group, but not in the statin group. The STOP-CA (Statins to Prevent the Cardiotoxicity of Anthracyclines) trial was the largest double-blind randomized placebo-controlled trial of atorvastatin for prevention of cardiotoxicity in patients with lymphoma. This study demonstrated a signifcant reduction of the primary endpoint of reduction of LVEF of $>10\%$ from prior to a final value of $< 55\%$ over twelve months [[74\]](#page-12-11). However, the PREVENT (Preventing Anthracycline Cardiotoxicity With Statins) trial of patients with breast cancer (85%) and lymphoma (15%) receiving anthracyclines pretreated with atorvastatin 40 mg daily showed no signifcant diference in the 24-month change in LVEF [\[75](#page-12-12)]. Multiple serum markers of infammation were reported including CRP, IL-6, and TNF- α , which decreased in both statin and placebo groups at 6- and 24-months post treatment [\[75](#page-12-12)]. In the STOP-CA trial, the participants received higher doses of anthracyclines (cumulative median anthracycline dose 300 mg/m² versus 240 mg/m²), were older and was limited to patients with lymphoma, representing a higher risk cohort of patients, indicating a likely beneft in the most high-risk patients.

Targeted Anti‑Infammatory Therapies for Prevention of AIC

Understanding the multiple mechanisms behind AIC are essential to develop new therapies for prevention and treatment of AIC. As seen above, traditional cardioprotective therapies have had mixed or limited responses in clinical trials, so anti-infammatory therapies may prove to be a promising strategy for prevention of AIC. However, these therapies have only been investigated in preclinical models of disease.

Steroids

Only preclinical studies have evaluated the cardioprotective efects of steroid therapy. Dexamethasone reduced the ratio of abnormal cardiomyocytes compared to doxorubicin alone in a mouse model of AIC [[76\]](#page-12-13). Interestingly, the single nucleotide polymorphism rs28714259 has been associated with an increased risk of AIC and, in a human induced pluripotent stem cells (hiPSC)-derived cardiomyocyte cell line, CRISPR-Cas9-mediated deletion of this locus identifed glucocorticoid receptor signaling as a key mediator of cardiotoxicity. Pretreatment with dexamethasone in the knock out cell line improved cell viability and contractility, which was not seen in control cells [\[77](#page-12-14)].

Colchicine

Colchicine is a well-established anti-infammatory medication that has proven efficacy in inflammatory cardiovascular disease including pericarditis, post-operative atrial fbrillation and atherosclerosis [[78](#page-12-15)[–80\]](#page-12-16). Colchicine irreversibly binds to tubulin to block microtubule polymerization. This disrupts multiple cellular processes, including neutrophil adhesion, TNF- α synthesis and activation of the NLRP3 infammasome [[80](#page-12-16)]. In preclinical study of in vivo and in vitro doxorubicin induced cardiac dysfunction, low dose colchicine (0.1 mg/kg daily) improved cardiac function compared to placebo [\[81\]](#page-12-17).

NLRP3 Infammasome Inhibitors

Activation of the NLRP3 infammasome plays an important role in the infammatory mechanism of AIC and inhibition of the NLRP3 is an increasingly recognized target for anti-infammatory therapies. Overexpression of sirtuin 3 (SIRT3) inhibits the NLRP3 infammasome activation via autophagy, reducing doxorubicin-induced cardiotoxicity [[82](#page-12-18)]. In both a rat model and in vitro cellular model, dihydromyrecetin (DHM), a favonoid compound, attenuates doxorubicin induced cardiotoxicity by reducing NLRP3 infammasome-mediated infammation [[83\]](#page-12-19). Similarly, calycosin, the active component in *Astragalus,* reduces cardiotoxicity through the Sirt1-NLRP3 pathway [[84\]](#page-12-20). Fraxetin, a coumarin from Cortex Fraxini, mitigated both oxidative stress and infammation through decreased activation of the NLRP3 in a dose-dependent manner. Resveratrol has also been shown to reduce cardiotoxicity via suppression of the NLRP3 infammasome [\[27\]](#page-11-0). Currently, multiple targeted CRID3-based therapies that inhibit the NLRP3 infammasome are in clinical development in Phase I through Phase III trials in a wide spectrum of infammatory diseases, including nonalcoholic fatty liver disease, ulcerative colitis, COVID-19-associated pneumonia, and neurodegenerative diseases. None have yet been trialed in prevention of AIC but may be promising therapies in the future.

Cytokine Receptor Blockers

In addition to NLRP3 infammasome targeted therapies, there are more downstream targets anti-infammatory therapies that target cytokines, like anakinra (IL-1R antagonist), canakinumab (IL-1β neutralizing antibody) and rilonacept (soluble receptor that binds IL-1β and IL-1 α). None have been studied in this population but are currently being evaluated in the cardiovascular sphere, notably in coronary artery disease [[85](#page-12-21)] and heart failure [[86](#page-12-22)].

Application to Other Forms of Cancer Therapy Related Cardiac Dysfunction

The scope of this review has focused on the mechanism of AIC because it is the most well recognized and well-studied form of CTRCD and the mechanisms of AIC can serve as a model to understand other forms of CTRCD. For example, pyroptosis, a key driver of the cycle of infammation as a result of cell death, has been identifed in other anticancer agents beyond anthracyclines because it is often a key driver of the anticancer properties; however, the off-target effects in the myocardium contribute to cardiotoxicity [[30\]](#page-11-3). Similarly, other chemotherapy agents have been shown to activate the NLRP3 infammasome in drug-induced toxicity of other organs. For example, cisplatin increases Nox4 with downstream activation of NLRP3 infammasome and leads to drug-induced nephrotoxicity [[87\]](#page-12-23). Bleomycin induced pulmonary toxicity occurs via HIF-1 α induced NLRP3 inflammasome activation. Additionally, both bortezomib- and paclitaxel-induced neurotoxicity are partially mediated by activation of NLRP3 infammasome [\[87](#page-12-23)]. By understanding mechanisms of toxicity that are common between multiple chemotherapeutic agents, hopefully more universal cardioprotective strategies can be devised.

In addition to cardiotoxicity from chemotherapy, the rise of immunotherapy has highlighted the important role of myocardial infammation in the development of CTRCD. While this review has focused on the role of the innate immune system, immune checkpoint inhibitors primarily cause cardiotoxicity via alterations in the adaptive immune system, particularly T-cells. Importantly, the rare, but potentially fatal condition immune checkpoint inhibitor myocarditis occurs as a result of the loss of normal checks on infammation leading to a robust infammatory infltration within the myocardium and resulting cardiac dysfunction [[88](#page-12-24)].

The above discussions of the role of cytokines in both triggering an intracellular infammatory cascade and alterations in cardiac contractility have important implications in cardiotoxicity from new T-cell mediated immunotherapies, including CAR T-cell therapy. With CAR T-cell therapy, there is a well-documented side efect of cytokine release syndrome that is associated with cardiotoxicity [[89](#page-12-25)]. Treatment with IL-6 targeted therapies can mitigate this cardiotoxicity.

Conclusions

CTRCD occurs because of multiple complex and overlapping mechanisms that lead to cardiotoxicity. In the case of AIC, infammation plays an important role and is part of a cycle whereby initial cardiac damage triggers an infammatory response which in turn causes further cardiotoxicity. AIC-induced infammation provides an important target for diagnosis and treatment of AIC, though the exact role deserves further study. An improved understanding of this mechanism may lead to novel methods of toxicity detection and prevention to improve the care of all patients with cancer. Additional research and expanded clinical studies are needed to further elucidate the role of infammation in chemotherapy induced cardiotoxicity.

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• Krysko DV, K.A., Krysko O, Heyndrickx L, Woznicki J, Bogaert P, Cauwels A, Takahashi N, Magez S, Bachert C, and Vandenabeele P., *TLR-2 and TLR-9 are sensors of apoptosis in a mouse model of doxorubicininduced acute infammation.* Cell Death Difer, 2011. **18**: p. 1316–1325

 Findings demonstrate that doxorubicin induces an immunogenic form of apoptosis that upregulates the innate immune system via toll-like receptors (TLRs)

• Lakhani, H.V., et al., *Detecting early onset of anthracyclines-induced cardiotoxicity using a novel panel of biomarkers in West-Virginian population with breast cancer.* Scientifc Reports, 2021. **11**(1).

 Findings demonstrate that a novel panel of biomarkers, including infammatory biomarkers, correlated with myocardial damage from AIC, as measured by troponin T and troponin I.

• Jong, J., J.R. Pinney, and R.R.S. Packard, *Anthracycline-induced cardiotoxicity: From pathobiology to identifcation of molecular targets for nuclear imaging.* Frontiers in Cardiovascular Medicine, 2022. **9**.

 This review provides an overview of nuclear imaging techniques for detecting anthracyclineinduced cardiomyopathy (AIC) and evaluates the evidence supporting the use of novel tracers for this purpose, including those capable of identifying infammation associated with AIC.

• Sun, Z., et al., *Dihydromyricetin alleviates doxorubicin-induced cardiotoxicity by inhibiting NLRP3 infammasome through activation of SIRT1.* Biochemical Pharmacology, 2020. **175**: p. 113,888

Findings demonstrate that dihydromyricetin can alleviate AIC via inhibiation of the NOD-like receptor protein 3 (NLRP3) infammasome.

Author Contribution A.F.S and E.H. wrote the main manuscript text. E.H and A.F.S prepared Figs. [1](#page-2-0) and [2.](#page-3-0) E. H. prepared Table 1. A.F.S., E.H. and E.H.Y designed the concept for the paper. All authors reviewed the manuscript.

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Compliance with Ethical Standards

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