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Draft Genome Sequence of *Gordonia* sp. Strain UCD-TK1 (Phylum *Actinobacteria*)

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Here, we present the draft genome of *Gordonia* sp. strain UCD-TK1. The assembly contains 5,470,576 bp in 98 contigs. This strain was isolated from a disinfected ambulatory surgery center.

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Members of the genus *Gordonia* are Gram-positive, aerobic bacilli that are commonly isolated from soil and water (1). Previously classified in the genus *Rhodococcus*, they are frequently misidentified as such following biochemical testing (2). Some *Gordonia* species are opportunistic pathogens that have been implicated in nosocomial infections, particularly in immunocompromised patients or those with medical devices, such as catheters (1, 3).

Gordonia sp. strain UCD-TK1 was isolated from a patient chair in the recovery room of an ambulatory surgery center in Redding, CA, USA, as part of an ongoing undergraduate research project to provide microbial reference genomes from the built environment. The chair had been cleaned with CaviCide, an Environmental Protection Agency–approved disinfectant, prior to swabbing. A sterile cotton-tipped applicator (Puritan) was used to swab the surface of the chair and then plate the sample on lysogeny broth agar. The plate was incubated at 37°C for 5 days. Individual colonies were streaked for isolation and, once isolated, were used to make an overnight culture that was incubated at 37°C. DNA was extracted from the overnight culture following the protocol of a Promega Wizard Genomic DNA purification kit. The 16S rRNA gene was amplified using PCR with 27F and 1391R primers. DNA was then purified and used for Sanger sequencing in which DNA is replicated in the presence of dideoxynucleotides generating varying lengths of DNA sequences. Sequences are then ordered by size and base to reconstruct the original DNA sequence. The resulting consensus sequence was analyzed using BLAST (4). Top hits were aligned using the Ribosomal Database Project (5). The alignment was then used to infer a maximum-likelihood phylogenetic tree, using Fast Tree (6), which was visualized in Dendroscope (7). The organism was found in a clade containing *Gordonia terrae* and *Gordonia lacunae*, along with other unnamed species of *Gordonia*.

For whole-genome sequencing, a paired-end library was prepared using a Nextera XT library preparation kit (Illumina). We selected 600- to 900-bp fragments using a Pippin Prep (Sage Science). A portion of an Illumina MiSeq sequencing run generated 653,024 paired reads with a read length of 300 bp. After quality trimming and error correction were completed by the A5-misec

assembly pipeline (8, 9), 577,554 quality reads remained in 95 scaffolds, with 22× coverage and a GC content of 67.8%. Genome completeness was estimated using the PhyloSift software (10), which searches for a list of 37 highly conserved, single-copy marker genes (11), all of which were found in this assembly in a single copy.

Annotation was performed using RAST (12). *Gordonia* sp. strain UCD-TK1 contains 5,032 coding sequences, and 64 non-coding RNAs. A partial-length 16S sequence (857 bp) was obtained from RAST and analyzed using BLAST. As expected, top hits (100% identity), included *G. terrae* and *G. lacunae*, along with unnamed *Gordonia* species. A phylogenetic tree was generated, as described above, and again was unable to resolve the taxonomy between the two strains. Two *G. terrae* whole-genome sequences have been published, but none for *G. lacunae*. Therefore, we were unable to assign a species name to this organism without further biochemical/chemotaxonomic characterization.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [LZMP000000000](https://www.ncbi.nlm.nih.gov/nuccore/LZMP000000000). The version described in this paper is the first version, LZMP01000000.

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REFERENCES

1. Ramanan P, Deziel PJ, Wengenack NL. 2013. *Gordonia* bacteremia. *J Clin Microbiol* 51:3443–3447. <http://dx.doi.org/10.1128/JCM.01449-13>.
2. Renvoise A, Harle J, Raoult D, Roux V. 2009. *Gordonia sputi* bacteremia. *Emerg Infect Dis* 15:1535–1537. <http://dx.doi.org/10.3201/eid1509.080903>.
3. Blaschke AJ, Bender J, Byington CL, Korgenski K, Daly J, Petti CA, Pavia AT, Ampofo K. 2007. *Gordonia* species: emerging pathogens in

- pediatric patients that are identified by 16S ribosomal RNA gene sequencing. *Clin Infect Dis* 45:483–486. <http://dx.doi.org/10.1086/520018>.
4. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [http://dx.doi.org/10.1016/S0022-2836\(05\)80360-2](http://dx.doi.org/10.1016/S0022-2836(05)80360-2).
 5. Cole JR, Wang Q, Fish JA, Chai B, McGarrell DM, Sun Y, Brown CT, Porras-Alfaro A, Kuske CR, Tiedje JM. 2014. Ribosomal Database Project: data and tools for high throughput rRNA analysis. *Nucleic Acids Res* 42:D633–D642. <http://dx.doi.org/10.1093/nar/gkt1244>.
 6. Price MN, Dehal PS, Arkin AP. 2010. FastTree 2—approximately maximum-likelihood trees for large alignments. *PLoS One* 5:e9490. <http://dx.doi.org/10.1371/journal.pone.0009490>.
 7. Huson DH, Scornavacca C. 2012. Dendroscope 3: An interactive tool for rooted phylogenetic trees and networks. *Syst Biol* 61:1061–1067. <http://dx.doi.org/10.1093/sysbio/sys062>.
 8. Tritt A, Eisen JA, Facciotti MT, Darling AE. 2012. An integrated pipeline for de novo assembly of microbial genomes. *PLoS One* 7:e42304. <http://dx.doi.org/10.1371/journal.pone.0042304>.
 9. Coil D, Jospin G, Darling AE. 2015. A5-miseq: an updated pipeline to assemble microbial genomes from Illumina MiSeq data. *Bioinformatics* 31:587–589. <http://dx.doi.org/10.1093/bioinformatics/btu661>.
 10. Darling AE, Jospin G, Lowe E, Matsen FA, Bik HM, Eisen JA. 2014. PhyloSift: phylogenetic analysis of genomes and metagenomes. *PeerJ* 2:e243. <http://dx.doi.org/10.7717/peerj.243>.
 11. Wu D, Jospin G, Eisen JA. 2013. Systematic identification of gene families for use as “markers” for phylogenetic and phylogeny-driven ecological studies of bacteria and archaea and their major subgroups. *PLoS One* 8:e77033. <http://dx.doi.org/10.1371/journal.pone.0077033>.
 12. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.