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## Identification of Genetic Variation Influencing Metformin Response in a Multiancestry Genome-Wide Association Study in the Diabetes Prevention Program (DPP)

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Genome-wide significant loci for metformin response in type 2 diabetes reported elsewhere have not been replicated in the Diabetes Prevention Program (DPP). To assess pharmacogenetic interactions in prediabetes, we conducted a genome-wide association study (GWAS) in the DPP. Cox proportional hazards models tested associations with diabetes incidence in the metformin (MET; n = 876) and placebo (PBO; n = 887) arms. Multiple linear regression assessed association with 1-year change in metformin-related quantitative traits, adjusted for baseline trait, age, sex, and 10 ancestry principal components. We tested for gene-by-treatment interaction. No significant associations emerged for diabetes incidence. We identified four genome-wide significant variants after correcting for correlated traits ( $P < 9 \times 10^{-9}$ ). In the MET arm, rs144322333 near *ENOSF1* (minor allele frequency [MAF]<sub>AFR</sub> = 0.07; MAF<sub>EUR</sub> = 0.002) was associated with an increase in percentage of glycated hemoglobin (per minor allele,  $\beta$  = 0.39 [95% CI 0.28, 0.50]; *P* = 2.8 × 10<sup>-12</sup>). rs145591055 near *OMSR* (MAF = 0.10 in American Indians) was associated with weight loss (kilograms) (per G allele,  $\beta$  = -7.55 [95% CI -9.88, -5.22]; *P* = 3.2 × 10<sup>-10</sup>) in the MET arm. Neither variant was significant in PBO; gene-by-treatment interaction was significant for both variants [*P*(G×T) < 1.0 × 10<sup>-4</sup>]. Replication in individuals with diabetes did not yield significant findings. A GWAS for metformin response in prediabetes revealed novel ethnic-specific associations that require further investigation but may have implications for tailored therapy.

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See accompanying article, p. 1057.

Metformin is the most commonly used agent for initial treatment of type 2 diabetes (T2D), with more than 100 million users worldwide. Over the years, it has remained the foundation of clinical practice guidelines for the management of diabetes globally because it is cheap, safe, effective, well-tolerated, orally administered, and does not promote weight gain (1–4). However, despite its widespread use, metformin does not work well for everyone; in the ADOPT study, which examined the glycemic durability of three oral glucose-lowering agents, each as monotherapy for T2D, 21% of individuals on metformin failed to meet glycemic goals within the first 5 years of treatment (5). Moreover, treatment failure rates are as high as 50% in children and adolescents, as shown by the TODAY study (6).

The reasons for metformin treatment failure remain unclear and could be related to genetic, pharmacologic, pathophysiologic, or environmental factors. Using the genome-wide complex trait analysis method, the heritability of metformin response has been estimated to be between 20 and 34%, a value comparable to other complex phenotypes (7). Indeed, genome-wide association studies (GWAS) have identified common genetic variants associated with metformin response (8-10). In the first GWAS for metformin response, a variant around the gene encoding the ataxia-telangiectasia mutated kinase (ATM) was found to be associated with metformin response in GoDARTS and UKPDS participants (8). Subsequently, in a meta-analysis of >10,000 participants with a harmonized glycemic measure of metformin response, a genome-wide significant association was observed in an intron of the SLC2A2 gene encoding the GLUT2 glucose transporter (9). However, neither of these findings was reproduced in the Diabetes Prevention Program (DPP) (9,11). While the difference in findings could be related to statistical power, one key distinction between the DPP and other studies is that DPP participants have prediabetes and not established T2D. Therefore, it is possible that the influence of genetics on drug response could vary in the prediabetes state compared with the more advanced T2D state.

Thus, we performed a GWAS in the DPP to uncover novel variation associated with metformin response in this distinctive multiethnic cohort that was selected to be at high risk of T2D by virtue of elevated 2-h glucose, fasting glucose, and BMI. Our objective was to identify genetic variants associated with metformin response, as measured by both diabetes incidence and change in quantitative glycemic and metabolic traits. Through this approach, we were able to examine the genetics of metformin response as it relates to both prediabetes and T2D development as well as glycemic response.

#### **RESEARCH DESIGN AND METHODS**

#### Description of Participants and the DPP Study Design

The DPP study design and baseline characteristics of the participants have been described previously (12,13).

Briefly, the DPP was a multicenter, randomized controlled trial designed to test the effects of intensive lifestyle modification and pharmacologic intervention on preventing progression to T2D in high-risk individuals. Enrolled participants had a fasting plasma glucose ranging from 95 to 125 mg/dL (5.3-6.9 mmol/L) and a 2-h plasma glucose level between 140 and 199 mg/dL (7.8-11.0 mmol/L) on a 75-g oral glucose tolerance test (OGTT). A total of 3,819 participants were randomly assigned to intensive lifestyle modification (goal weight loss  $\geq$ 7% and  $\geq$ 150 min/week of physical activity), standard lifestyle recommendations plus metformin (850 mg twice daily), standard lifestyle recommendations plus troglitazone (400 mg daily), or standard lifestyle recommendations plus placebo. The primary end point was diabetes incidence, diagnosed by a fasting glucose of  $\geq$ 126 mg/dL (7.0 mmol/L) or a 2-h glucose of  $\geq$ 200 mg/dL (11.1 mmol/L) after OGTT and confirmed subsequently on a second test within 6 weeks. The primary study demonstrated that over a mean follow-up of 2.8 years, there was a 58% (95% CI 48, 66) reduction of diabetes incidence in the intensive lifestyle intervention group and a 31% (95% CI 17, 43) reduction in the metformin group compared with placebo (14).

Institutional review board approval was obtained by each participating clinical center. All participants included in this analysis provided written informed consent for the main investigation and for subsequent genetic studies.

#### **Genome-Wide Genotyping and Quality Control**

DNA was extracted from peripheral blood leukocytes. Genotyping was performed on 3,227 samples using the Human CoreExome genome-wide array (Illumina, San Diego, CA). We excluded single nucleotide polymorphisms (SNPs) with a call rate <95% or if they failed Hardy-Weinberg equilibrium ( $P < 1.0 \times 10^{-8}$ ) within each ethnic group. Samples with discrepant sex, call rate <95%, inbreeding coefficient <-1, and identity-by-state as measured by pi-hat close to 1 were discarded. As 9,730 SNPs and 3,222 samples were additionally genotyped on the MetaboChip (Illumina), we performed a concordance check, excluding SNPs and samples with a concordance rate <95%. After all quality checks were performed, 3,168 samples remained (Supplementary Fig. 1).

#### Imputation

We performed a two-stage imputation procedure, which consisted of prephasing the genotypes into whole chromosome haplotypes followed by imputation itself. The prephasing was performed using SHAPEIT2 (15), and IMPUTE2 was used for genotype imputation (16). GWIMp-COMPSs can incorporate the contribution of several reference panels (17), and in this work we used 1000 Genomes Phase 3 haplotypes (October 2014) (18). Supplementary Table 1 summarizes the distribution of the 3,168 samples by self-reported race/ ethnicity and the DPP treatment arm that underwent imputation.

#### **Statistical Analyses**

Because the study was conceived as a pharmacogenetic study on the influence of genetics on metformin response, we focused our primary analyses on the standard lifestyle recommendations plus metformin (MET) group and the placebo (PBO) group. Continuous variables are presented as mean  $\pm$  SD and categorical variables as frequency (percentage). Baseline characteristics were compared with ANOVA tests for quantitative variables and  $\chi^2$  tests for qualitative variables.

#### **Diabetes Incidence**

A Cox proportional hazards model tested the association between genetic variants and diabetes incidence under an additive genetic model in the MET and PBO arms. For any genome-wide significant findings, we planned to check and ensure that the proportionality assumptions were met. We evaluated the impact of genetic variation in the MET arm only, and, in a second model, we included a formal interaction test between the MET and PBO arms. Models were adjusted for age at randomization, sex, and 10 ancestry principal components (PCs) for population structure; secondary models were also adjusted for waist circumference or BMI. We addressed common variants (minor allele frequency [MAF] > 1%), both those directly genotyped and imputed. For imputed markers, we excluded variants that had an imputation quality score <0.7, as well as variants that did not pass a filter test  $(2 \times MAF \times [1 - MAF] \times n.events > 75$ , where n.events is the number of events), as described in the GWASTools package (19). We evaluated for genome-wide significance, defined as  $P < 5 \times 10^{-8}$ .

#### Change in Quantitative Traits

We used a multiple linear regression model to test allelic associations, assuming additive effects, with the change in quantitative traits relevant to metformin action at 1 year; traits included fasting glucose, 2-h glucose after 75-g OGTT, fasting insulin, insulin sensitivity index (ISI), hemoglobin  $A_{1c}$  (Hb $A_{1c}$ ), and weight. One-year change in each quantitative outcome was defined as 1-year minus baseline value. Non-normally distributed traits were natural log transformed. To minimize the influence of outliers, winsorization was performed (at percentiles of 0.5 and 99.5 for normally distributed traits and percentiles of 1 and 99 for natural log-transformed traits) (20). Analyses were adjusted for age, sex, first 10 ancestry PCs, and the baseline value of the trait. Similar to the diabetes incidence analyses, we evaluated the impact of genetic variation in the MET arm only for each of the six outcomes, and, in a second model, we tested for a gene-by-treatment (G×T) interaction for the MET and PBO arms. We filtered our results to a study-wide MAF >1% and imputation quality  $\ge 0.7$ . To account for multiple testing and the possible correlation between quantitative traits, we calculated that there were 5.47 effective outcomes, using a previously described method (21), and we set an experiment-wide significance threshold of  $P < 9 \times 10^{-9}$  (5 ×  $10^{-8}$ /5.47). For top findings that emerged, we performed an exploratory analysis that evaluated these findings in the intensive lifestyle treatment arm to understand whether the variants act through a shared pathway between metformin and lifestyle intervention.

#### Replication

For each of the quantitative trait outcomes, we compiled a list of the top 500 genotyped and imputed variants with an MAF >1% and imputation score  $\geq 0.7$  for imputed markers, for a total of  $\sim$ 6,000 variants (500 markers × 6 traits  $\times$  2 models). We explored their relevance in the Metformin Genetics (MetGen) Consortium, which has data from >10,000 participants from >12 observational studies and clinical trials, in which a pharmacogenetic meta-analysis of metformin response was performed (9). We did additional filtering of variants, excluding those with a minor allele count <10 and with imputation score <0.7 from the Pharmacogenomics of Metformin (PMET) study, which represents the largest cohort comprising MetGen participants. We then calculated the number of effective variants after accounting for correlation between genetic variants (21) and determined the replication significance threshold. In addition, we attempted to replicate our top variant in the Million Veteran Program (MVP) and the Diabetes Multiomic Investigation of Drug Response (DIAMOND). Full details of replication cohorts and the statistical models used in the replication analyses are described in the Supplementary Material and Supplementary Table 2.

#### **Data and Resource Availability**

Genetic data generated and analyzed in the current study are available in dbGap (dbGaP Study Accession: phs000681.v2.p1, https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi? study\_id=phs000681.v2.p1). Phenotype data for the DPP are available through the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK) Data Repository. No applicable resources were generated or analyzed during the current study.

#### RESULTS

#### **Baseline Demographics**

Table 1 describes the characteristics of the subset of DPP participants in the MET and PBO arms in whom genome-wide genotyping was available. The mean baseline age was 50.7  $\pm$  10.4 years, 67% were women, and  $\sim\!\!44\%$  self-reported as non-White. Baseline measurements were consistent with a population at risk for developing T2D. There were no significant differences in baseline measurements by treatment arm. The event rate for diabetes incidence is reported for each self-reported race/ethnicity group for both the MET and PBO arms in Supplementary Table 3.

# Association of Genetic Variation With Diabetes Incidence

In the MET arm only (n = 876) (Supplementary Table 4), we performed association analysis of genetic variants with

Baseline characteristic	Total	PBO	MET	P value**
n	1,768	888	880	
Age, years	50.7 ± 10.4	50.6 ± 10.4	50.9 ± 10.3	0.538
Men	578 (32.7)	281 (31.6)	297 (33.8)	0.345
Women	1,190 (67.3)	607 (68.4)	583 (66.3)	
Self-reported race/ethnicity White Black Hispanic/Latino Asian/Pacific Islander American Indian	997 (56.4) 364 (20.6) 290 (16.4) 69 (3.9) 48 (2.7)	490 (55.2) 186 (20.9) 147 (16.6) 38 (4.3) 27 (3.0)	507 (57.6) 178 (20.2) 143 (16.3) 31 (3.5) 21 (2.4)	0.746
Weight (kg)	94.7 ± 19.8	94.9 ± 20.0	94.5 ± 19.6	0.692
HbA <sub>1c</sub> (%)†	5.917 ± 0.501	$5.920 \pm 0.495$	5.915 ± 0.507	0.832
Fasting insulin (pmol/L)†#	167 (111, 236)	167 (111, 229)	167 (111, 236)	0.595***
Fasting glucose (mmol/L)	$5.940 \pm 0.462$	$5.956 \pm 0.466$	$5.923 \pm 0.458$	0.138
ISI†#	0.161 (0.112, 0.235)	0.161 (0.113, 0.236)	0.161 (0.110, 0.234)	0.762***

Table 1-Baseline characteristics of 1,768 participants in the DPP in the MET and PBO arms with genome-wide genotyping

Data are means  $\pm$  SD for continuous variables and *n* (%) for categorical variables unless otherwise indicated. \*\**P* values from *F* test for continuous variables and  $\chi^2$  for categorical variables. \*\*\**P* value from Kruskal-Wallis test. †*n* = 1,766 for total; *n* = 887 for PBO; *n* = 879 for MET. #Median (25th, 75th percentile).

diabetes incidence during the main DPP study, which had a mean follow-up of 3.2 years. While we did not observe any genome-wide significant findings (Supplementary Fig. 2A), we identified a genetic locus near *HIF1AN* that reached suggestive significance. The C effect allele of rs2489017 had a frequency of 0.59 in the entire DPP cohort and was notably more common in non-White populations (White: 0.49, African American: 0.83, Asian/Pacific Islander: 0.62, Hispanic/Latino: 0.62, and American Indian: 0.78). Per copy of the variant, participants had a decreased risk of progressing to diabetes (hazard ratio 0.54 [95% CI 0.43, 0.67];  $P = 9.2 \times 10^{-8}$ ). In the placebo group, the variant did not influence diabetes incidence (P = 0.72), and we noted the presence of an interaction with treatment arm [ $P(G \times T) = 5.9 \times 10^{-5}$ ].

We subsequently combined the MET and PBO arms (n = 1,763) (Supplementary Table 4) and used a Cox proportional hazards model to assess for a gene-by-treatment arm interaction for the outcome of diabetes incidence. We did not identify any genome-wide significant findings or variants that met a suggestive significance threshold of  $P < 1 \times 10^{-6}$  (Supplementary Fig. 2*B*).

# Association of Genetic Variation With 1-Year Change in Quantitative Traits

With respect to the quantitative trait outcomes, we performed association analysis of genetic variants with 1-year change in any of six quantitative traits (fasting glucose, 2-h glucose after OGTT, fasting insulin, ISI, HbA<sub>1c</sub>, and weight) across two separate models, one examining the MET arm only and another testing for a gene-by-treatment arm interaction. Supplementary Table 3 presents the sample size for each model and quantitative trait outcome tested. Table 2 summarizes the 14 independent signals that met genome-wide significance ( $P < 5 \times 10^{-8}$ ) for at least 1 quantitative trait, 4 of which met experiment-wide significance ( $P < 9 \times 10^{-9}$ ): 2 for change in HbA<sub>1c</sub> (Supplementary Fig. 3) and 2 for change in weight (Supplementary Fig. 4). These results remained the same after additionally adjusting the analyses for BMI (data not shown). The effect sizes and *P* values for the top findings, stratified by self-reported race/ethnicity, are also presented in Supplementary Table 5. Because we report the effect allele frequencies stratified by self-reported ethnicity/race, we compared these groupings with the ancestry groupings determined by PC analysis and illustrated that the selfidentified groups tended to cluster tightly on the PC analysis plot (Supplementary Fig. 5).

The top hit, rs144322333, which was associated with change in HbA<sub>1c</sub> in the metformin treatment group, is a deletion polymorphism near *ENOSF1* (Fig. 1) and was mainly found in African American participants with an MAF of 6%, whereas it was present in under 0.2% in White individuals (Table 2). Carriers had a 0.39% increase in 1-year change in HbA<sub>1c</sub> per copy of the deletion, consistent with a worse response to metformin ( $P = 2.9 \times 10^{-12}$ ) (Fig. 2*A*). The proportion of variance in 1-year change in HbA<sub>1c</sub> explained by the variant was 5.2%, compared with 10.8% for baseline HbA<sub>1c</sub>, the strongest predictor of this quantitative outcome in the model.

Since rs144322333 is more common in African American individuals, we performed a subgroup analysis and found that the effect was detectable in this group ( $P = 1.3 \times 10^{-6}$ ) (Fig. 2*B*). In fact, African American individuals without any copies of the deletion experienced a *decrease* in HbA<sub>1c</sub> of 0.03% after a year of metformin, whereas those who carried one or two copies of the deletion experienced *increases* of

Table 2–Fourteen inde	pend	lent genome	-wide significal	nt finc	dings (/	$^{\circ} < 5 \times 10^{-8}$ ) for 1-year	r chang	ge in quantitative tra	its in the DP	ē.					
											Ш	ffect alle	e freque	ncy	
rsID	Chr	Position*	Nearest gene	EA	NEA	Trait	Ν	β (95% CI)	Ρ	AII†	AfrAm‡	AsnPl‡	Hisp‡	AI‡	White‡
Metformin only model															
rs144322333	18	705550	ENOSF1	0 0	TGTT	HbA <sub>1c</sub> , %	818	0.39 (0.28, 0.50)	$2.9 \times 10^{-12}$	0.013	0.065	0.001	0.009	0.0001	0.002
rs145591055	S	38849463	OSMR	പ	∢	Weight, kg	829	-7.55 (-9.87, -5.22)	$3.2 \times 10^{-10}$	0.014	0.001	0.018	0.064	0.103	0.007
rs13401282§	2	207810690	СРО	∢	⊢	ISI, In	808	0.44 (0.29, 0.59)	$1.7 \times 10^{-8}$	0.029	0.119	0.003	0.015	0.006	0.002
rs186681623	13	81546608	LINC00377	с	⊢	Weight, kg	829	-2.66 (-3.58, -1.74)	$2.0 \times 10^{-8}$	0.068	0.121	0.074	0.122	0.161	0.036
rs17083791	2	93863813	KIAA0825	പ	۷	Weight, kg	829	-1.85 (-2.50, -1.20)	$3.3 \times 10^{-8}$	0.149	0.052	0.194	0.210	0.367	0.137
rs9931871	16	19928315	<b>GPRC5B</b>	പ	4	<sup>-</sup> asting glucose, mmol/L	821	0.68 (0.44, 0.93)	$3.5 \times 10^{-8}$	0.011	0.049	<0.0001	0.003	<0.0001	<0.0001
rs549305231	9	19497504	LOC101928519	∢	G	HbA <sub>1c</sub> , %	818	0.35 (0.23, 0.48)	$3.9 \times 10^{-8}$	0.012	0.046	<0.0001	0.005	<0.0001	0.0003
rs73944532	2	104691634	LOC100287010	∢	G	Fasting insulin, In	802	-0.47 (-0.63, -0.30)	$4.1 \times 10^{-8}$	0.020	0.075	0.005	0.007	0.001	0.002
Gene × treatment model															
rs6838493	4	185879789	LINC01093	∢	⊢	HbA <sub>1c</sub> , %	1,621	-0.66 (-0.87, -0.44)	$1.6 \times 10^{-9}$	0.011	0.041	0.004	0.005	0.004	0.002
rs148219263	40	58808522	CDH20	с	⊢	Weight, kg	1,673	10.73 (7.15, 14.30)	$4.8 \times 10^{-9}$	0.012	0.007	0.011	0.011	0.0009	0.017
rs75147163	4	57626769	EXOC5	∢	J	<sup>-</sup> asting glucose, mmol/L	1,633	-0.73 (-0.98, -0.48)	$1.4 \times 10^{-8}$	0.022	0.053	0.018	0.022	0.0002	0.015
rs78075715	4	6613716	MAN2B2	с	⊢	HbA <sub>1c</sub> , %	1,621	-0.51 (-0.68, -0.33)	$1.4 \times 10^{-8}$	0.013	0.063	<0.0001	0.002	<0.0001	0.0004
rs12314996	42	24567467	SOX5	∢	G	HbA <sub>1c</sub> , %	1,621	-0.47 (-0.63, -0.30)	$3.9 \times 10^{-8}$	0.016	0.062	<0.0001	0.007	0.001	0.002
rs143203347	2	107384199	CBLL1	U	G	-asting glucose, mmol/L	1,633	-0.92 (-1.24, -0.59)	$4.1 \times 10^{-8}$	0.015	0.062	0.0002	0.011	<0.0001	<0.0001
AfrAm. African American	AL.	American Inc	dian: AsnPI. Asia	in/Pac	cific Isla	inder: Chr. chromosom	e: FA.	effect allele: Hisp. His	spanic/Latino	SIL in	sulin ser	asitivity ir	dex: In.	natural Ic	od trans-
formed; NEA, noneffect	allele	; rsID, refere	nce SNP ID. *G	RCh37	7 assen	holy. †This is the effect	allele	frequency for the num	her of partic	ipants	in the m	odel (N) t	that was	calculate	d based
on imputation. ‡Effect	allele	frequency	breakdown by	self-re	eportec	I race/ethnicity is for	all 3,1	68 participants with	genome-wid	de gen	otyping,	calculate	ed base	d on im	outation.
§rs13401282 is also ass	ociate	ed with fastir	ng insulin ( $\beta = -$	0.39	[95% C	(1 - 0.53, -0.25]; P = 4.1	$65 \times 1$	0 <sup>-8</sup> ). G×T models col	ntain an expo	osure te	erm for t	he treatm	nent arm	and a G	×T inter-
action term. Results are	filtere	∋d to P(G×T)	$< 5 \times 10^{-8}$ and	I P(SN	IP) < 1	× 10 <sup>-4</sup> . ∥β Estimates a	Ind Cls	are reported for the	interaction te	erm rath	er than 1	the main	effect o	f metform	Ŀ.



**Figure 1** – Regional association plot of rs144322333 for 1-year change in HbA<sub>1c</sub>. The red line indicates the experiment-wide significance threshold of  $P < 9 \times 10^{-9}$ , and the blue line indicates a suggestive significance threshold of  $P < 1 \times 10^{-6}$ .

0.2% and 0.6%. respectively. Finally, since the association was discovered in the MET arm only, we also evaluated its impact in the PBO arm, in which there was no significant effect (P = 0.32). The interaction with treatment arm was notably significant [ $P(G \times T) = 1.4 \times 10^{-6}$ ] (Fig. 2*C*), reflecting the differential impact of this genetic variant.

Another novel experiment-wide significant variant was rs145591055 in chromosome 5 near *OMSR* (Fig. 3), a gene that encodes the oncostatin M receptor (Table 2).

Carriers of the G effect allele had a 7.6-kg greater decline in weight at 1 year following initiation of metformin treatment in adjusted analyses ( $P = 3.2 \times 10^{-10}$ ) (Fig. 4A). The variant has an effect allele frequency of 10% in American Indians, calculated from the entire DPP study (Table 2). While a subgroup analysis in American Indians was limited by sample size (n = 21) (Fig. 4B), we noted that noncarriers experienced a decrease in weight of 0.5 kg at 1 year, compared with a decrease of 8.6 kg in heterozygous carriers.



**Figure 2**—*A*: Box plot illustrating the mean change in HbA<sub>1c</sub> (1-year minus baseline) by rs144322333 genotype in the MET only arm. n = 818. *B*: Stratified analyses by self-reported race/ethnicity. *P* value was calculated for the subgroup of African American individuals. n = 166. *C*: Comparison of the influence of rs144322333 genotype on the mean change in HbA<sub>1c</sub> in all subjects in the MET (n = 818) and PBO (n = 803) arms. The interaction *P* value is reported. For the purposes of generating the box plots, fractional alleles were converted to "hard calls" by rounding to the nearest integer. AfrAm, African American; AI, American Indian; AsnPI, Asian/Pacific Islander; Hisp, Hispanic/Latino.



**Figure 3**—Regional association plot of rs145591055 for 1-year change in weight. The red line indicates the experiment-wide significance threshold of  $P < 9 \times 10^{-9}$ , and the blue line indicates a suggestive significance threshold of  $P < 1 \times 10^{-6}$ .

We also evaluated the influence of this variant in the placebo arm and found that there was no significant effect on weight (per G allele,  $\beta = -0.69$ ; P = 0.50). Again, there was a significant gene-by-treatment interaction [P(G×T) =  $1.5 \times 10^{-5}$ ] (Fig. 4*C*).

In the gene × treatment models, we found that rs6838493, in chromosome 4 near *LINC01093* (Supplementary Fig. 6), had a differential influence on change in HbA<sub>1c</sub> [ $P(G \times T) =$ 

 $1.6 \times 10^{-9}$ ] (Supplementary Fig. 7), in which heterozygous carriers experienced a *rise* in HbA<sub>1c</sub> at 1 year in the PBO arm ( $\beta = 0.5$ ;  $P = 1.3 \times 10^{-8}$ ) in comparison with a *decline* in HbA<sub>1c</sub> in the MET arm ( $\beta = -0.1$ ; P = 0.03). The impact of rs148219263, located in chromosome 18 near *CDH20* (Supplementary Fig. 8), on 1-year change in weight was also different by treatment arm [ $P(G \times T) = 4.8 \times 10^{-9}$ ] (Supplementary Fig. 9). In this study, in comparison with their TT



**Figure 4**—*A*: Box plot illustrating the mean change in weight (1-year minus baseline) by rs145591055 genotype in the MET only arm. n = 829. *B*: Box plot stratified by self-reported race/ethnicity. *C*: Comparison of the influence of rs144322333 genotype on the mean change in weight in all subjects in the MET (n = 829) and PBO (n = 844) arms. The interaction *P* value is reported. For the purposes of generating the box plots, fractional alleles were converted to "hard calls" by rounding to the nearest integer. AfrAm, African American; AI, American Indian; AsnPI, Asian/Pacific Islander; Hisp, Hispanic/Latino.

counterparts, TC carriers experienced weight loss in the placebo arm ( $\beta = -7.0$ ;  $P = 2.4 \times 10^{-8}$ ) but did not achieve weight loss after receiving metformin ( $\beta = 3.9$ ;  $P = 4.0 \times 10^{-3}$ ).

In an exploratory analysis, we evaluated our top findings from Table 2 in the intensive lifestyle arm. Of the nine variants that emerged from the analysis completed in the MET arm only, one variant (rs13401282) appeared to have a similar effect on ISI in the lifestyle arm as in the MET arm (P = 0.004) (Supplementary Table 6); the other eight variants were nonsignificant in the lifestyle arm. For the five variants identified through the gene × treatment model that combined the MET and PBO arms, we tested their relevance in a similar interaction model that combined the PBO and lifestyle arms. We found that four of the five variants demonstrated similar findings between the two gene × treatment models, as illustrated by the magnitude and direction of the  $\beta$  estimates, as well as the *P* values (Supplementary Table 6).

We searched our lead findings in the Genotype-Tissue Expression (GTex) project (https://gtexportal.org) (22) and eQTLGen Consortium (https://eqtlgen.org) (23) to evaluate for expression quantitative trait loci (eQTL) associations. We observed that rs17083791, which was associated with a greater decline in weight at 1 year following metformin, is a cis-eQTL associated with higher expression of KIAA0825, the nearest gene, in fibroblasts (normalized effect size [NES] 0.411;  $P = 2.7 \times 10^{-12}$ ), subcutaneous adipose tissue (NES 0.285;  $P = 7.9 \times 10^{-7}$ ), skeletal muscle (NES 0.241;  $P = 5.0 \times 10^{-5}$ ), and visceral adipose tissue (NES 0.229; P = $1.6 \times 10^{-4}$ ). In blood samples in eQTLGen Consortium, we noted that the variant was also a *cis*-eQTL for MCTP1 (P = $7.3 \times 10^{-189}$ ). We also evaluated the top variants in the pancreatic islet genotype tissue-expression resource (Translational Human Pancreatic Islet Genotype Tissue-Expression Resource [TIGER]; http://tiger.bsc.es), which aggregates >500 human islet genomic data sets (24), but there were no notable findings. Finally, we searched top variants against phenotypes associated with T2D using the Type 2 Diabetes Knowledge Portal (https://t2d.hugeamp.org). We found that the G allele of rs145591055, which was associated with greater 1-year weight change on metformin, was marginally associated with a higher BMI-adjusted waist-to-hip ratio ( $\beta = 0.079$ ; P =0.001).

#### Independent Replication

We sought to replicate our pharmacogenetic findings in additional cohorts in which metformin response has been defined. As there are few cohorts of patients with prediabetes, our first approach was to explore the relevance of the variants in MetGen, in which the outcome was glycemic response, as measured by baseline minus minimum on-treatment HbA<sub>1c</sub> within 18 months after metformin initiation (9). Out of ~6,000 variants, only 3,050 had available genotype information in MetGen, resulting in 2,610 effective variants; we thus set a significance threshold of  $P < 1.9 \times 10^{-5}$  (0.05/2,610). After filtering, results were available in MetGen for only 6 of the 14 genome-wide significant variants from our DPP GWAS, none of which replicated (Table 3).

Because the study cohorts in MetGen were of predominately European ancestry, we sought to replicate our top HbA<sub>1c</sub> finding in African American cohorts. We first examined 2,733 participants with established T2D from MVP who self-reported as non-Hispanic African Americans and had received metformin monotherapy for up to 15 months. The top variant, rs144322333, which had an MAF of 7% and an imputation score of 0.89 in MVP, was not significantly associated with change in HbA<sub>1c</sub> ( $\beta$  = 0.05; *P* = 0.15).

We also sought replication in DIAMOND, an observational population-based cohort with electronic medical record–linked clinical data on individuals with T2D on metformin monotherapy. We identified a subset of 471 individuals who self-reported as African American and were calculated to have a reliable exposure of at least 500 mg metformin daily as monotherapy for treatment of T2D in the 120 days preceding the follow-up HbA<sub>1c</sub> measurement. In DIAMOND, rs144322333 had an MAF of 8% and an imputation score of 0.98. Per copy of the deletion, carriers had a 0.13% reduction in HbA<sub>1c</sub> lowering over 1 year in DIAMOND, indicating a worse metformin response, which was similar in direction to that seen in the DPP but smaller in magnitude and not reaching statistical significance (P = 0.19).

#### DISCUSSION

To date, GWAS have uncovered several genetic loci influencing the glycemic response to metformin, but these were largely performed in European populations with established T2D (8-10). When these variants were assessed in the DPP, the reported associations were not confirmed for the outcomes of diabetes incidence and other relevant physiologic parameters, such as insulin sensitivity, fasting glucose,  $HbA_{1c}$  or oral disposition index (9,11). One possible explanation for this inconsistency is that metformin response is defined differently in a disease cohort, in which the measure of response is the achievement of an HbA<sub>1c</sub>  $\leq$ 7%, compared with a cohort with prediabetes, in which diabetes incidence or quantitative traits are the studied outcomes. Furthermore, genetic variation may influence drug response differently in individuals with prediabetes, who likely have better pancreatic  $\beta$ -cell function.

Thus, we undertook a GWAS in the DPP examining the outcomes of diabetes prevention as well as 1-year change in six quantitative traits known to be affected by metformin therapy. The design of the DPP is unique for studying metformin pharmacogenetics, as it includes both a PBO and MET arm, permitting the use of an interaction test to assess genetic variants that may have a differential response based on drug exposure. In assessing diabetes incidence, we did not identify any genome-wide

Table 3—Attempt at replication of genome-wide significant fin	ndings from the DPP in the MetGen Consortium
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DPP model	Trait	rsID	DPP β*	MetGen (n)	MetGen β*	MetGen P value
MET	Weight, kg	rs186681623	<ul> <li>–2.66 (greater weight loss)</li> </ul>	10,519	-0.05 (decreased HbA <sub>1c</sub> reduction)	0.10
MET	Weight, kg	rs17083791	<ul> <li>–1.85 (greater weight loss)</li> </ul>	12,578	0.004 (greater HbA <sub>1c</sub> reduction)	0.79
MET	Fasting insulin, In	rs73944532	<ul> <li>-0.47 (greater decrease in fasting insulin)</li> </ul>	7,048	0.03 (greater HbA <sub>1c</sub> reduction)	0.84
G×T	Weight, kg	rs148219263	10.73 (reduced weight loss)	10,519	-0.04 (decreased HbA <sub>1c</sub> reduction)	0.39
G×T	Fasting glucose, mmol/L	rs75147163	<ul> <li>-0.73 (greater decrease in fasting glucose)</li> </ul>	10,288	-0.02 (decreased HbA <sub>1c</sub> reduction)	0.70
G×T	HbA <sub>1c</sub> , %	rs12314996	-0.47 (greater decrease in HbA <sub>1c</sub> )	7,048	0.19 (greater HbA <sub>1c</sub> reduction)	0.24

In, natural log transformed; rsID, reference SNP ID. \*In the DPP, the  $\beta$  estimates represent 1-year change in the quantitative trait as calculated by follow-up value minus baseline value. In MetGen, the outcome was change in HbA<sub>1c</sub> (%), defined as baseline value minus follow-up value within 18 months (will have opposite signs of  $\beta$  estimates for change in HbA<sub>1c</sub> compared with that in the DPP). For each analysis and cohort, the impact of the variant on the metformin response outcome is indicated in parentheses.

significant variants. We acknowledge that the relatively modest sample size of the DPP with a limited range of impaired glucose tolerance may have restricted our ability to detect an interaction effect. However, we did observe an association that met a suggestive threshold that could be investigated in the future or meta-analyzed with other cohorts.

After correcting for multiple testing, our quantitative trait analyses yielded several genome-wide significant associations. *ENOSF1*, the closest gene to the top variant rs144322333, encodes the enolase superfamily member 1, a mitochondrial enzyme involved in the catabolism of L-fucose(a sugar found on cellular glycoproteins). Interestingly, *ENOSF1* is next to a kinase, YES1, which has been shown to regulate the activity of organic cation transporters OCT1 (*SLC22A1*) and OCT2 (*SLC22A2*) that play a critical role in metformin disposition and elimination in the kidney and liver, respectively (25,26). Moreover, *LINC01093* near rs6838493 has been shown to be related to several liver phenotypes, including fibrosis (27) and hepatocellular carcinoma (28), and may play a role in modulating metformin's action on the liver.

Many of our genome-wide significant findings were ancestry-specific (rs144322333 near *ENOSF1*, rs145591055 near *OSMR*, and rs6838493 near *LINC01093*). We further illustrate this through subgroup analyses and demonstrate the persistent effect of top variants of interest after stratifying by self-reported ethnicity/race. Based on these stratified analyses, it appeared that the ancestry specificity of our findings was mainly driven by allele frequency, since similar trends were observed across subgroups. We acknowledge that this may have been a consequence of conducting our analyses in a multi-ancestry cohort, which could have biased our results toward variants with concordant effects across subgroups. One limitation of this approach is that variants with opposite effects in different subgroups could be potentially missed. We also estimated that our top genetic finding (rs144322333) explained 5.2% of the variance in 1-year change in HbA<sub>1c</sub>, which was half of that of baseline HbA<sub>1c</sub>, supporting the notion that the contribution of genetics is modest compared with a traditional predictor of drug response.

In the MET arm of the DPP, the average weight loss was 2.1 kg and was relatively stable throughout the study follow-up period (14). In comparison, the association of rs145591055 (near OSMR) variation with 1-year change in weight was 7.6 kg, more than three times greater in magnitude. Individuals who carry this variant, which has an effect allele frequency of 10% in American Indians, appear to derive a significant weight-loss benefit from metformin, which is especially relevant given the high incidence and prevalence of T2D in this population (29,30). Moreover, the G allele was reported in the T2D Knowledge Portal to be marginally associated with a higher BMI-adjusted waistto-hip ratio, also suggesting that this is a group of individuals who would benefit from metformin. However, the underlying mechanism responsible for this weight effect is not clear and requires further investigation. We also evaluated our top findings in the GTex project, the eQTLGen Consortium, and the TIGER portal. Interestingly, we observed that the variant rs17083791 was a cis-eQTL for the nearest gene KIAA0825 in several relevant tissues, suggesting that changes in KIAA0825 gene expression may modulate metforminrelated weight loss. The same variant was also a strong ciseQTL for MCTP1, which codes for a protein involved in calcium ion binding, though the significance of this finding is unknown. While these genomic resources can be useful for functional follow-up of variants, the majority of participants

are of European ancestry, and thus, their utility is limited for variants that are common only in specific populations.

In an exploratory analysis, we evaluated the top findings from the quantitative trait analysis using the intensive lifestyle arm of the DPP. There were five variants for which the association with the quantitative trait was similar in the lifestyle arm as in the MET arm. Prior studies have suggested that the intensive lifestyle intervention of the DPP worked similarly to metformin in improving insulin sensitivity (31). Therefore, it is possible that the mechanism of action of these variants may be shared between metformin and lifestyle. In contrast, the remaining nine variants that met genome-wide significance were nonsignificant when evaluated in the lifestyle arm, illustrating they may be specific to metformin response.

Finally, we took a comprehensive approach to replicate findings that emerged from our GWAS, though this was a major challenge due to the lack of a suitable replication cohort of individuals with prediabetes who received longitudinal metformin exposure. We evaluated approximately the top 500 variants for each quantitative trait analyses in MetGen, the largest meta-analysis examining glycemic response to metformin in T2D (9). Our efforts did not yield replication, but this could be explained by the composition of MetGen as previously discussed. We subsequently identified two African American cohorts (MVP and DIAMOND) for replication of our top variant rs144322333. Though the study populations again largely reflected individuals with T2D, we constructed our models to reflect a similar length of metformin exposure as in the DPP and included similar covariates as in our quantitative trait analyses. Despite imputation on different reference panels in these cohorts, we reassuringly observed similar MAFs for rs14432233; however, neither replication produced a nominally significant result. We acknowledge the possibility that given the low study-wide MAF of our top findings and the small sample size, our findings could represent false-positive results.

Our work underscores the importance of conducting multi-ancestry pharmacogenetic studies to improve upon treatment algorithms across diverse populations in the future. A prior study based on pharmacy fill data reported that African American adults appear to have a better glycemic response to metformin compared with European Americans (32), and the TODAY study showed that African American youth with T2D on metformin had poorer glycemic response to metformin (6). Though these studies were based on self-reported race/ethnicity information, we demonstrated in the DPP that there is clustering of the self-reported ethnicity/race groups along the genetic ancestry PCs. Thus, the identification of variants in our study that are more prevalent in African American populations suggests that there may be a heritable component to these differences in response to metformin.

Strengths of our study include the diverse composition of the DPP, which includes nearly 45% non-White

individuals by self-report and represents a cohort of individuals at a higher risk of developing T2D in the U.S. Participants were well-characterized under standardized clinical trial protocols, and the genetic data were obtained using the highest standards for genotyping and imputation at the time. As described, limitations of our study are the small sample size, which is a frequent problem in pharmacogenomic GWAS (33), and the lack of replication of our findings in patients with diabetes, though this may reflect differences in genetic modulation of metformin response between the prediabetes and diabetes state. However, we were exhaustive in our attempts to identify independent cohorts, including the largest African American cohorts with diabetes known to us. Future directions should include additional follow-up and meta-analyses in additional independent ancestryspecific cohorts as they arise in order to advance work in this area.

In conclusion, we have identified several genome-wide significant associations with metformin response, as measured by change in quantitative traits, in a well-defined multi-ancestry clinical trial cohort. We believe that we have generated a valuable resource that can be used for future genetic investigation and to gain insight into the genetic underpinnings of interindividual differences in metformin response in a population at risk for developing T2D. Finally, we illustrate the peremptory need to generate suitable pharmacogenomic and transcriptomic resources in diverse populations.

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