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2023

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Chemical Investigations into Pyrrole-Containing Secondary Metabolites Isolated From *Curvularia* Sp. and *Bipolaris Maydis*

By

Paulo Andre Machicao Tello

A dissertation submitted in partial satisfaction of the

requirements for the degree of

Doctor of Philosophy

in

Chemistry

in the

Graduate Division

of the

University of California, Berkeley

Committee in charge:

Professor Thomas J. Maimone, Chair Professor Richmond Sarpong Professor Roberto Zoncu

Fall 2023

Chemical Investigations into the Pyrrole-Containing Secondary Metabolites Isolated From *Curvularia* Sp. and *Bipolaris Maydis*

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Pyrrole-containing alkaloids have long captured the attention of the synthetic chemistry community due to their intriguing and reactive chemical architectures; such is the case of curvulamine and associated polypyrrole natural products. Curvulamine, curindolizine, and the related bipolamines share a unique compact pentacyclic skeleton flanked by two electron-rich pyrroles and up to seven stereocenters. Synthetic access to these complex alkaloids has previously been limited to curvulamine and bipolamine I. In this work, we aim to chronicle the results of our synthetic investigations into this family of unique alkaloids. In the first chapter, we provide an overview into their isolation and antibiotic properties, and discuss our initial synthesis of curvulamine which frames our studies in subsequent chapters. The second chapter describes the total synthesis of curindolizine and the substrate scope of a microwave-assisted method to access pyrroloazepinones, bicyclic heterocycles used in the synthesis of curvulamine and curindolizine. Finally, in the last chapter we describe the synthetic strategies used to access bipolamine C, D, G, and I and describe the unexpected reactivity of some pyrrole-containing intermediates along this journey.

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Acknowledgements

Tom: I just want to say thank you for all your support, patience, and guidance throughout these past few years. You are an incredible chemist and a brilliant mind in the field. During our multiple conversations I have learned so much to the point where I can say that your lessons have reshaped the way I approach science. They have guided me into a new path that I am excited to continue to explore, like you told me when I started " you need to understand chemistry". It took me some time to realize what that meant (maybe more than I or you were excepting) but now I can say that I get it. I still have a long journey ahead and like I told you one day "I wont' stop learning, just because the PhD is over". Again, I cannot express with words the gratitude I feel toward the opportunity you provided me with.

The Curvulamine Project Team:

Dr. Karl Haelsig: We didn't spend much time together in the lab and COVID for sure didn't help, but I feel I have gotten to know you better after you graduated. You are an amazing chemist, and I am shocked by your resilience in and outside the lab. The experiences I have had outside the lab with you have also showed me that you are a funny and an understanding person. Thank you for being a mentor and for giving me the opportunity to continue the work you started.

Dr. Jun Xuan: I am extremely happy that I was able to work alongside you. You taught me so much about the curvulamine project and lab techniques that I have used throughout my PhD. I also enjoyed our conversation about sports.

Former Lab Members:

Dr. Bingqi Tong: It is sad that we didn't get to work together in person. You were always super kind to me and very welcoming. You are a great chemist and a very humble individual (Also really good at sports!). Thank you for your help with the dankastatin alkaloids; your recommendations made it much easier to navigate the finnicky chemistry in the project.

Dr. Linus Shen: Working in the lab with you in there during the pandemic was fun. I got to spend quality time with you and get to know the real you (or at least try...). I enjoyed our hikes and honest conversations outside the lab. Certainty you are a complex person, a very thick book in your own words, and getting to understand you wasn't the easiest task but once I did, I felt I gained a good friend.

Dr. Rachel Rosen: Ranch, I cannot express with words how much I valued our time together in the lab. Our multiple conversations, especially the ones late at night made a huge difference in how I felt in the lab. You are just not a kind and caring person; you are also very empathetic and a real friend. I enjoyed our times outside the labs as well (includes going to the city, going out for chinse food at midnight, going dancing, enjoying drinks, so many...) Thank you for taking the time to teach me stuff, distillation, using instruments, techniques, looking over data, etc. Again, I am super happy I got to spend time together and that I was stuck with you in 907 during the pandemic.

Dr. Danny Thach: I learned so much from you. You are one of a kind, the best combination of good looks and intelligence. You always gave me good advice, not just about science, but also on how to be

a good human being. I have never met someone so resilient and determine to accomplish his dreams. You are an example to follow, you are a story of overcoming different challenges to reach your goals. The times I spent with you inside and outside the lab will always be present in my mind when I think about my time in Berkeley. You were really the glue that kept it together!

Dr. Claire Harmange Magnani: Thank you for being a fun hood mate. I am sorry if sometimes my very disorganized space gave you some headaches. I admire your level of organization, your dedication toward science, and your love for your family. I don't know how you managed to be a mom while finishing a natural product in the middle of the pandemic. I value those honest conversations we had and thank you for laughing at my sometimes niche jokes (Monét X Change).

Dr. Vasil Vasilev: You are an incredible human being with a contagious laugh. Even before I joined the lab you were nice to me. I enjoyed both our time in the lab and outside the lab. I really cherish the times we went hiking, eating (or maybe I should say devouring food), and hanging out. You are also incredible funny even when you are not trying to be funny. I enjoyed our time together in the lab and I greatly appreciate all the words of encouragement that you gave me during these past few years.

Dr. Andre Sanchez: Thank you for sharing your best tunes and playlist with the rest of 907. I want to express my gratitude for helping me during the first years of the PhD. You helped me edit documents and provided me with useful advice. Also thank you for helping me with the calculations and for sometimes reminding me that there is more to life than chemistry and lab work.

Danny Huang: You are brilliant and so dedicated to achieving your goals. Since the moment we met during orientation, you have been a good friend. We shared many many moments together and while not all of them have been happy, I feel they have brought us closer together. I think it is funny now you know what I am thinking and what I am going to say even seconds before I do or say something. Also thank you for teaching me about good Chinese food, especially the noodles with meat that I like.

Beth Zhu: Nopa! You are so funny, I honestly believe you don't realize how much. I am so glad we joined the lab at the same time. Our conversations about our struggles during the beginning of the PhD were extremely helpful. Thank you for keeping the lab organize and for helping me with the multiple lab outings.

Yi Xie: You are a very smart person. I am glad that over the years we have been able to have multiple conversations about science, sports, and anime. I have enjoyed the times we were able to get dim sum and hotpot. You are working on a hard project, and I know you will be able to put the pieces together to solve the puzzle.

Lukas: Congratulations on promptly getting a job and moving to the next chapter in your life. Thank you for the beer and for the energy drinks, both were very helpful during these past few years.

Kelvin Li–Debbie Jones: Thank you for all the help editing multiple documents! It has been a pleasure to work alongside you. You are a very good listener and very kind person. I have enjoyed our conversations about niche topics and we both seem to understand well. You are not just a talented chemist, but also amazing at sports!

Vanessa Gonzalez: Muchas gracias por todas las buenas conversaciones que hemos tenido a lo largo de estos ultimos años. Lo siento si a veces no he sido el mejor compañero de hood, tu sabes que he tenido mucha presión para poder lograr resultados y después tratar de encontrar trabajo y poder graduarme. Me da mucha felicidad ver como has crecido durante tu tiempo en el programa, la verdad eres una persona muy talentosa en muchos aspectos: gran cocinera, great listener, y buena maestra (tienes mucha paciencia). Héchale ganas, ya casi acabas!

Kincade Stevenson: Kin__it has been fun working alongside you. One of the things I admire the most about you is your honesty. You say it like it is and I appreciate that about you. Your understanding and knowledge of chemistry is also amazing. You can think through problems with great ease (at least that is how I perceive it). Thank you for sharing conversations about cars and hikes with me, I really enjoyed it.

Tanner Myers: Enjoy your time in Berkeley. Your project is challenging, but you are surrounded by great chemists that will help you achieve your goals.

Shang Ning: Thank you for being flexible when sharing the photoreactor, it really helped a lot! Also make you sure include the recipe for your spicy skewers somewhere in your dissertation or online blog because they are delicious, and I want the recipe. You are super chill and fun to talk to, I have enjoyed our convos.

Katerina Gorou: First of all, thank you for all the help with the calculations, I learned a lot from you. You are kind, funny, smart, talented, and the list goes on. I appreciate all the laughs, especially the ones super late at night. I am sorry if at times I wasn't the most approachable person when you first started, but I am glad we were able to interact more after your first semester. You have a challenging project ahead of you but remember that you are surrounded by smart people and they are willing to always give you a hand.

Julia Friedli: It was a privilege to mentor you while you were at Berkeley. Your dedication to achieving the best results possible was admirable. You are a very talented scientist (not just chemist) and a super kind person. You are also funny and a person that cares deeply about different issues. Again, working alongside you was a gratifying experience and I hope we get to meet again.

A mi familia: Muchas gracias a todos (primos, primas, tios, tias, hermana, y padres). Yo se que no hemos estado cerca estos ultimos años pero siempre tengo en mi mente todos los recuerdos que he tenido con ustedes. La comida hecha con mucho cariño de mis abuelos como el pie de manzana de mama Maruja o el asado de mi abuela Antonia. Todas las propinas que mi tía Marita me daba cuando iba a visitarme (también recuerdo mucho el Verano que pase contigo). Mayra por enseñarme a disfrutar la vida y que no todo es estudio (gracias por llevarme a fiestas y a divertirme). Erika, yo se que no hemos pasado mucho tiempo juntos ahora como adultos, cuando me fuí estabas muy pequeña pero recuerda que siempre estoy libre para cualquier cosa que necesites. Jose Luis ponle ganas a estudiar, tu carrera es dificil pero vas a ver que el tiempo se va a pasar rapido. Erika y Jose Luis busquen siempre la manera de obtener la mejor educacion y oportunidades que puedan, asi van a lograr sus objetivos. Hermana, no hemos vivido juntos por muchos años pero quiero que sepas que me siento muy orgulloso de lo que has logrado. Como siempre te digo, a veces hay que aguantarse y pues ni modo seguir. Tú tienes todo el potencial para lograr lo que tu quieras solo que tienes hacerlo. Mami muchas gracias por

todo tu apoyo desde que era pequeño. Yo se que dejarme ir hace muchos años tal vez fue una de las cosas mas dificiles que tuviste que hacer pero gracias por confiar en mi. Papá Tito, gracias por ser la persona en la familia que decidio tomar la decision de estudiar en Lima y dejar todo atras. Nunca tuve la oportunidad de decirtelo pero yo entiendo lo que tuviste que pasar.

Carlos Gonzalez: Muchas gracias por apoyarme durante mi PhD. Por esperarme despierto esas noches que llegaba tarde (Yo se que fueron muchas), por siempre hacerme ver que hay cosas fuera del trabajo y por mandarme a la escuela con comida. Muchas de las cosas que me dijiste me ayudaron mucho a aguantar toda la presion y el estres de mi trabajo en el laboratorio. No tengo palabras para expresar la gratitud que siento por todo tu apoyo y ayuda (tambien por el apoyo de tu familia).

List of Abbreviations

ABNO	9-azabicyclo[3.3.1]nonane N-oxyl
AIBN	azobisisobutyronitrile
Boc	tert-butyloxycarbonyl
bpy	2,2'-bipyridyl
CAN	ceric ammonion nitrate
CBS	Corey–Bakshi–Shibata
CDI	1,1'-carbonyldiimidazole
CuDPP	Cu ^I diphenylphosphinate
dba	dibenzalacetone
DBU	1,8-diazabicyclo(5.4.0)undue-7-ene
DCA	9,10-dicyanoanthracene
DCC	N,N'-dicyclohexylcarbodiimide
DCE	1,2-dichloroethane
DCM	dichloromethane
DIPEA	N,N-diisopropylethylamine
DFT	density functional theory
DIBAL	diisobutylaluminium hydride
DMac	dimethylacetamide
DMAP	4-dimethylaminopyridine
DMDO	dimethyldioxirane
DMF	dimethylformamide
DMS	dimethyl sulfide
DMSO	dimethyl sulfoxide
dr	diastereomeric ratio
dpm	dipivaloylmethanato.
dt-Bu	di-tert-butyl
dtbpy	4,4'-di-tert-butyl-2,2'-dipyridyl
ee	enantiomeric excess
Et	ethyl
FVP	flash vacuum pyrolysis
НАТ	hydrogen atom transfer
HFIP	hexafluoroisopropanol
HMPA	hexamethylphosphoramide
hν	photo irradiation
KHMDS	potassium bis(trimethylsilyl)amide
LDA	lithium diisopropylamide
LiHMDS	lithium bis(trimethylsilyl)amide
Lg	leaving group
Me	methyl
MeCN	acetonitrile

Mes	mesitylene		
MeO _{hnv}	4,4'-dimethoxy-2,2'-bipyridine		
Ms	methanesulfonyl		
MVK	methyl vinyl ketone		
MWI	microwave irradiation		
m-CPBA	meta-chloroperoxybenzoic acid		
NaHMDS	sodium bis(trimethylsilyl)amide		
NBS	N-bromosuccinimide		
NIS	N-iodosuccinimide		
NMI	N-methylimidazole		
NMR	nuclear magnetic resonance		
NOE	nuclear Overhauser effect		
	oxidation		
OTF	trifluoromethanesulfonate		
Phen	1,10-phenanthroline		
PhMe	toluene		
	diacetoxyiodobenzene		
	2-phenylpyridinato-C ² ,N		
SEM	2-(trimethylsilyl)ethoxymethyl		
SET	single electron transfer		
TBAF	tetra-n-butylammonium fluoride		
TES	triethylsilyl		
ΤΕΔ	trifluoroacetic anhydride		
TED	tri(2-furyl)phosphine		
TRS	tert-butyldimethylsilyl		
TEMP	2,2,6,6-retramethyl-1-piperidinyloxy		
O THE	tetrahydrofuran		
TIC	thin layer chromatography		
t-Bu	tert-butyl		
TIPS	triisopropoylsilyl		
TMS	trimethylsilyl		
OAc	acetate		
Pin	pinacolato		
PPTS	pyridinium p-toluenesulfonate		
PTSA	p-toluenesulfonic acid monohydrate		
1 1 21 1			

Chapter 1

Overview of the Curvulamine-Type Alkaloids

1.1 Curvulamine-Type Alkaloids, Isolation and Biosynthesis

The *Pleosporaceae* is the most diverse family of ascomycete in the *Dothideomycetes* class of fungi.¹ Species belonging to this family are reported to be endophytes, epiphytes or parasites and inhabit a broad range of ecosystems.² Notable *Pleosporaceae* are found in the genera *Bipolaris* and *Curvularia*, which contain species known to reside in tropical and subtropical environments and have symbiotic relations with plants and marine animals.³⁻⁶ The intimate symbiotic connection of these fungi to other species has driven the interest of the scientific community to investigate the biosynthesis and biological activities of the secondary metabolites produced by organisms in these genera.⁷⁻¹⁰

As part of a campaign to identify novel agents with promising biological activities, Tan and co-workers identified two fungi present in the fish *Argyrosomus argentatus*: *Myrothecium* sp. Z16 and *Curvularia* sp. IFB-Z10. The first fungus was isolated from the spermary glad of the fish and the second was isolated from the intestinal tract.^{7,8} The fungal broth obtained from *Myrothecium* sp. Z16 was found to exhibit antimycotic activity against *Candida albicans*, *Trichophyton rubrum*, and *Aspergillus niger*. The broth obtained from *Curvularia* sp. IFB-Z10 was found to have antibiotic activity and shown to inhibit the growth of the following bacterial strains: *Peptostreptococcus* sp., *Veillonella parvula*, *Bacteroides vulgatus*, and *Streptococcus* sp. with minimum inhibitory concentrations (MICs) in the low micromolar range (0.37 μ M).⁸ Notably, Tan and coworkers identified a novel bis-pyrrole alkaloid named curvulamine (1) as the agent responsible for these antimicrobial activities. The structure of **1** was unequivocally assigned by single crystal X-ray crystallography and confirmed a unique [5,7,6,5]-fused tetracyclic bispyrrole skeleton.

Since the initial discovery of 1, Tan and co-workers have identified more than 11 nitrogen-containing secondary metabolites with skeletons akin to curvulamine (1) (Figure 1.1A).¹⁰ In 2016, as part of an effort to further investigate the biological properties of 1, an unexpected and novel nitrogenated alkaloid, namely curindolizine (2), was discovered when scaling up the cultivation of Curvularia sp. IBF-Z10. Single crystal X-ray crystallography was employed to assign the structure of curindolizine (2).⁹ Due to its structural similarities to curvulamine (1), 2 is believed to be a metabolite derived from 1 after a regiospecific Michael addition of the eastern pyrrole of 1 into 3,5-dimethylindolizin-8(5H)-one.⁹ Recent studies by the Tan group on the biosynthesis of these nitrogenated metabolites shed light on the biosynthetic gene cluster (BGC) responsible for making curvulamine (1).7 In this work, the cuaB gene was identified as being responsible for assembling the C₁₀N indolizine unit, a key component in all curvulamine-type alkaloids. This finding triggered a genome mining campaign to search for homologous fungal biosynthetic gene clusters (BGCs) in genomes catalogued in databases from the Joint Genome Institute (JGI) and the National Center for Biotechnology Information (NCBI). From this study, the fungus Bipolaris maydis (ATCC48331) was identified as not only possessing *cuaB*, but also a set of distinctive genes coding for tailoring enzymes predicted to facilitate the synthesis of novel indolizine-type alkaloids. The culture broth of Bipolaris maydis failed to produce any detectable quantity of nitrogenated secondary metabolites under laboratory conditions; but fermentation along with the overexpression

of the putative transcription factor *bipF* afforded 9 new nitrogenated alkaloids with indolizine scaffolds similar to curvulamine (1). These new nitrogenated metabolites were named bipolamines A-I (3-11) and analysis of their structures revealed various oxidation



A. Curvulamine-Type Alkaloids Isolated from Curvularia sp. IFB-Z10 and Bipolaris Maydis

carbon numbering (12)

Figure 1.1 The secondary metabolites isolated from *Curvularia* sp. IFB-Z10 and *Bipolaris maydis*. **A**. The curvulamine alkaloids. **B**. Carbon numbering and skeletal patters of the curvulamine-type alkaloids.

carbon numbering (13)

patters that are attributed to the tailoring enzymes cytochrome P450 monooxygenase, cofactor F₄₂₀-dependent oxygenase, and an α-ketoglutarate-dependent oxygenase.

carbon numbering (14)

The C₁₀N indolizine unit (see **12**) is a common element in all curvulamine-type alkaloids (*vide supra*), and their members can be grouped into three different categories based on the number of these indolizine units present in their carbon skeleton (Figure 1.1B).¹¹ Bipolamine A (**3**) and bipolamine B (**4**) are the simplest congeners with a single methylated tetrahydroindolizine C₁₀N skeleton (**12**). These metabolites are believed to be early biosynthetic building blocks to higher order curvulamine-type alkaloids.¹⁰ The C₂₀N₂ group of congeners, is the most diverse and contains the largest number of family members. The tetracyclic C₂₀N₂ bisindolizine skeleton (**13**) in these curvulamine type alkaloids is functionalized with different levels of oxidation and hydroxylation patterns. The preserved tetracyclic C₂₀N₂ core is believed to be derived from the fusion of a C₁₀N indolizine unit (**12**) to a cleaved indolizine biosynthetic intermediate. The final category of

curvulamine-type alkaloids is the product of enzymatic addition of a $C_{10}N$ unit to the C-8 position of the tetracyclic $C_{20}N_2$ bisindolizine skeleton. To this date, curindolizine (**2**) is the only known higher-order curvulamine alkaloid in this group containing a $C_{30}N_2$ skeleton (**14**).

Two studies by Tan and co-workers has partially elucidated the biosynthesis of the curvulamine alkaloids.^{8,10} In the first study, evidence was obtaining for a polyketide origin of curvulamine (1) via isotopic labeling experiments. Curvularia sp. IFB-Z10 was cultivated in the presence of 1-13C or 2-13C enriched sodium acetate, and 13C NMR analysis of the fungal culture extracts revealed a curvulamine skeleton with ¹³C enrichment in an alternating fashion, indicative of a polyketide origin of this alkaloid. Notably, no labeling was observed at C-9, C-10, C-19 or C-20 in either of these experiments. This indicated the possible involvement of other carbon sources and enzymes in the biogenesis of the skeleton of **1**. Further ¹³C labeling experiments employing [2,3-13C] alanine resulted in ¹³C enrichment at C-9, C-10, C-19, and C-20 in curvulamine (1). In subsequent biochemical experiments, pyridoxal-5'-phosphate(PLP)dependent oxoamine synthase (AOS) was identified as a key enzyme in the formation of the C₁₀N indolizine unit. As stated, the C₁₀N unit is believed to be an early intermediate in the biosynthesis of all curvulamine alkaloids. This hypothesis was confirmed by cultivating Curvularia sp. IFB-Z10 in the presence of a known inhibitor of AOS, which blocked production of 1. More recently, Tan and co-workers sequenced the genome of Curvularia sp. IFB-Z10 and identified the biosynthetic gene cluster and enzymes involved in the synthesis of early precursors en-route to curvulamine (1) and seemingly the rest of the curvulamine alkaloids.¹⁰

It is proposed that the *cuaA* gene encodes the polyketide synthase machinery to condense one acetyl-CoA and three malonyl-CoA into linear β -keto enone **15**. *cuaB* encodes a multipurpose enzyme that first condenses alanine with PLP forming iminium **16**. *cuaB* then triggers a PLP mediated chain release involving a Claisen condensation of **16** with enone **15**. Subsequent decarboxylation, aerobic oxidation, intramolecular condensation and dehydration produces cyclic hemiaminal **17**. Polyene **17** is proposed to be the key synthetic unit in the biogenesis of all the curvulamine alkaloids. Genes *cuaC* and *cuaD* encode the enzymes responsible for converting hemiaminal **17** into indolizine **18** and epoxide **19**. It is postulated the C₂₀N₂ skeleton is generated after a coupling event between **18** and **19** and a series of redox reactions, however the precise mechanism of this biosynthetic step remains elusive. The biosynthetic gene cluster of *Bipolaris maydis* indicates that tailoring enzymes such as cytochrome P450 dependent monooxygenases and oxidoreductases could further functionalize the C₂₀N₂ skeleton giving rise to the diversity of oxidation patterns found in the curvulamine alkaloids.¹⁰

In a separate study, during the investigations of curindolizine (2), Tan and co-workers isolated procuramine (20) from the culture broth of *Curvularia* sp. IFB-Z10.⁹ Interestingly, incubation of 20 and curvulamine (1) in the presence of *Curvularia* sp. IFB-Z10 protein lysate formed curindolizine (2). The individual enzymes involved in this transformation were not identified, but a plausible route to 2 might involve dehydration of 20 to form 3,5-dimethylindolinone (21), which can undergo regioselective Michael addition with 1 followed by a reduction and dehydration.

It is worth noting that while these previously described studies provide information about the plausible biosynthetic pathway of the curvulamine alkaloids, a complete biogenesis remains elusive, especially with respect to how the seven-membered ring is formed.



B. Proposed Biosynthesis of Higher Order C₃₀N₃ Metabolites



Figure 1.2 Summary of biosynthetic studies on the curvulamine alkaloids. **1.2A**. Proposed biosynthesis of **1**. **1.2B**. Proposed biosynthesis of **2**.

1.2 Biological Activity of the Curvulamine-Type Alkaloids

Compounds	Veillonella parvula	Streptococcus sp.	Bacterioides vulgatus	Peptostreptococcus sp.
curvulamine (1)	0.37	0.37	0.37	0.37
bipolamine A (3)	4.20	8.40	8.40	>10
bipolamine D (6)	2.87	>10	5.74	5.74
bipolamine E (7)	0.73	2.92	2.92	5.85
bipolamine F (8)	3.52	3.52	3.52	3.52
bipolamine G (9)	0.32	0.32	0.32	0.32
bipolamine H (10)	0.35	2.94	1.47	2.94
bipolamine I (11)	1.53	>10	>10	>10

Table 1.1 Antibacterial activity (MICs in μ M) of the curvulamine alkaloids.

Natural products have been employed throughout history for the treatment of various ailments and have been a rich source of potential drug candidates.¹²⁻¹⁴ Natural products sourced from fungi, such as penicillin, have played an important role in modern medicine.¹⁵ Particular focus has been paid to fungal symbionts because they produce secondary metabolites that promote the proliferation and survival of their hosts.¹⁶⁻¹⁸ In 2014, Tan and co-workers discovered the fungal ectosymbiont *Curvularia* sp. IFB-Z10 in the gut of the white croaker, a marine organism that feeds on carrion. Evaluation of the fungal extracts against a broad range of bacterial strains revealed potent antibiotic activity against Gram-positive and negative bacteria. Upon further examination, the antibacterial activity observed on the crude extracts was attributed to the denitrogenated alkaloid curvulamine (1).⁸ Isolated 1 was found to exhibit low micromolar minimum inhibitory concentrations (MICs) against *Veillonella parvula*, *Streptococus* sp., *Bacteroides vulgatus*, and *Peptostreptococcus* sp. (Table 1.1). In an antibacterial assay, 1 was shown to be more selective and potent than tinidazole, a prescribed antibiotic and antiparasitic.^{19,20}

Curindolizine (2), a metabolite also isolated from *Curvularia* sp. IFB-Z10 was also tested for antibacterial activity, but found to be inactive against the bacterial strains used in the curvulamine studies.⁹ In a broader screen for biological properties, 2 was evaluated for anti-inflammatory activity using the Griess method, and gratifyingly displayed anti-inflammatory properties in lipopolyssacharide-induced RAW 264.7 macrophages by inhibiting nitric oxide production.²¹ Nitric oxide is a signaling molecule heavily involved in the pathogenesis of inflammation. The antagonistic property of 2 extends to cytokines TNF-a, IL-1 β , and IL-6, which are also involved in inflammation pathways in cells. It is worth noting that despite the structural similarities of curindolizine (2) with the rest of the curvulamine alkaloids, it is the only congener reported to exhibit anti-inflammatory activity.





Figure 1.3 Proposed mechanism of action of the curvulamine alkaloids.

maydis bioactive metabolites (2-11) is still unknown. It is possible that 1 and its congeners

bind to a protein pocket disrupting bacterial proliferation in a non-covalent mode of action, such is the case for the polyketide antibiotic erythromycin which stops the synthesis of bacterial proteins by blocking the nascent peptide exit tunnel (Figure 1.3).^{22,23} Alternatively, one can also envision a covalent mode of action like β -lactams such as the antibiotic penicillin may be operative.^{24,25} This mode of action would require reactive sites in **1** that can interact with protein residues. A cursory analysis of the structure of curvulamine (**1**) shows no electrophilic sites, however under conditions where the C13-C3 ether bridge is protonated and ionizes, several electrophilic sites could be unveiled. This newly formed intermediate (see **21**) could act as a covalent inhibitor in the presence of nucleophilic amino acid residues to form bacterial protein-small molecule complex **23** inhibiting bacterial proliferation (Figure 1.3). This hypothesis may explain the lack of bioactivity of bipolamine I (**11**), the only metabolite with a C-14–C-3 ether bridge, which would be unable to form electrophilic intermediate **21**.

Our total synthesis campaign was in part motivated by a desire to develop the tools necessary to carry a proper investigation on the biological targets of the curvulamine alkaloids and it will be discussed in greater detailed later in this chapter.

1.3 Distinctive Challenges in the Synthesis of Pyrrole-Containing Natural Products

Pyrroles are electron rich heterocycles with reduced aromatic character compared to benzene.²⁶⁻²⁹ A combination of these two features define the pyrrole reactivity. This fivemembered heterocycle can undergo oxidation via single electron processes, protonation in mild acidic conditions, facile attack by electrophilic reagents, and extrusion of benzylic leaving groups (Figure 1.4).²⁶ These processes can produce highly reactive cationic intermediates that readily polymerize or undergo side reactions in solution, greatly limiting the compatibility of many chemical transformations of synthetic targets containing pyrrole fragments. The pyrrole reactivity is increased when electron-donating substituents are present and attenuated with electron-withdrawing groups.



Figure 1.4 Native pyrrole reactivity.

The curvulamine alkaloids contain a unique [5,7,6,5] tetracyclic core with two electronrich pyrrole units that are particularly sensitive to oxidative and acidic reaction conditions (Figure 1.5). To overcome this challenge, historically a combination of deactivating groups or pyrrole surrogates are employed when attempting to synthesize this subgroup of alkaloids.³⁰ In the case of curvulamine (**2**), one of the pyrrole units can be masked as an aromatic 10π heterocycle, an observation which ultimately proved to be critical in the synthesis of this natural product.





1.4 The Total Synthesis of Curvulamine

The intricate structures and promising biological activities of the curvulamine alkaloids has attracted the attention of the scientific community and they have become appealing targets for synthetic organic chemists.³¹⁻³³ Our group has reported pioneering work in this area beginning with the first total synthesis of (–)-curvulamine (1)³⁴, followed by subsequent reports describing the total synthesis of curindolizine (2)¹¹ and four bipolamines.³⁵ In this chapter, we provide an overview of the synthesis of 1 which is intended to provide context for the subsequent chapters that will describe in detail the synthesis of four *Bipolaris maydis* metabolites (6,7,9,11) and curindolizine (2).

A. Central Principle in the Synthesis of Curvulamine



Figure 1.6 Forays into the development of a strategy toward **1**. **1.6A**. Proposed bond disconnections and synthetic fragments. **1.6B**. Proposed retrosynthesis of **1**.

It was envisioned that **1** could be broken down into three smaller building blocks depicted in Figure 1.6. From the proposed bond disconnections between C-3–C4, C-5–C-6, and C-12–C-13, synthetic fragments that contained functional groups harboring potentially desirable reactivity could be identified. Retrosynthetically, curvulamine (**1**) was traced back to tetracycle **24** after various redox manipulations and removal of a two-



Figure 1.7 The total synthesis of (–)-curvulamine (1). 1.7A. Synthesis of pyrroloazepinone 25. 1.7B. Synthesis of cyanohydrin 26. 1.7C. Synthesis of 1.

carbon unit. Tetracycle **24** in turn, could be disconnected to pyrrolo[1,2-a]azepin-7-one **25** and cyanohydrin **26** via a proposed Michael addition and annulation sequence. This convergent step allowed for an initial foray into the reactivity of nitrogen-containing heterocycle **25**, which will be further discussed in the following chapter.

The synthesis began with the preparation of the pyrrole containing fragments **25** and **26**. The pyrrolo[1,2-a]azepin-7-one **25** fragment was elaborated in two steps. First,

pyrrolecarbaldehyde 27 and (E)-4-methoxybut-3-en-2-one (28) were joined via an aldol condensation, followed by microwave irradiation of the resulting enone to illicit cyclization. An expedient three-step sequence was developed to synthesize cyanohydrin 26 beginning with nucleophilic substitution of methyl 2-bromopropanoate (29) with the metal salt of 2-methyl pyrrole. Then DIBAL reduction of the methyl ester gave an intermediate aldehyde which was reacted with TMSCN in the presence of LiClO₄ to give 26, which was immediately used without further purification. These two fragments were coupled by deprotonation of 26 with NaHMDS to form the sodium anion which, in the presence of LiCl, underwent selective addition to pyrrolo[1,2-a]azepin-7-one 25 aided by lithium chloride. This Michael type addition afforded a product with the desired positional selectivity in moderate yields forging the first C-C bond in the planned annulation strategy. Recognizing that an oxidative process was required to realize the second key bond formation in the annulation, the enolate formed after the 1,4 addition was quenched with NIS to give enone **30** in a combined 64% yield. While this reaction favored the undesired configuration at the C-2 stereocenter, this was addressed later in the synthesis. Simple irradiation of 30 with a 390 nm light source in the presence of a polar protic solvent promoted the cyclization event yielding tetracycle 24. Treatment of 24 with excess ethyl vinyl ether lithiate produced **31** with the desired formal acyl addition in 55% isolated yield. As previously stated, the methyl bearing stereocenter C-2 needed to be epimerized and lactol 31 offered such opportunity. Conformational analysis suggested that allylic 1,3strain between the methyl groups at C-1 and C-10 could be minimized with axial positioning of the C-1 methyl group providing the desired configuration. Heating 31 with sodium methoxide in methanol yielded a 2.3:1 thermodynamic mixture of lactols favoring the desired isomer 32 in 85% yield. At this stage, the last hurdle to overcome was the removal of the hydroxyl group at the bridgehead position, which was accomplished in two steps. First, lactols 31 and 32 were deprotonated with KHMDS and subsequent acylation of the resulting anion with CICSOPh in the presence of DMAP, generated a separable mixture of thiocarbonate epimers 33 and 34. Then, Barton-McCombie deoxygenation under mild heating with concomitant enol ether hydrolysis during acidic workup afforded methyl ketone 35. Finally, stereodivergent reduction of the methyl ketone 35 with (R)-2methyl-CBS-oxazaborolidine and BH3•DMS afforded a separable 1:1 mixture of curvulamine (1) and epi-curvulamine with 97% yield and 95% ee respectively. In summary, the first total synthesis of (-)-curvulamine (1) was accomplished in a total of 10 steps from commercially available materials.

1.5 Aims for the Pursuit of the Cuvulamine Alkaloids

Alkaloids have long captured the attention of the synthetic community due to their intriguing chemical architectures and promising biological activities.¹³ Pyrrole-containing alkaloids exhibit a broad range of bioactivities, and their structural complexity present a noteworthy challenge for the practitioners of total synthesis.³⁰ Among the many families of alkaloids, natural products containing electron-rich pyrroles represent an underexplored family for which limited established strategies and methodologies exist. As a result, chemists must often devise novel approaches for the construction of such

A. Pyrroles in Nature





Figure 1.8 Applications of pyrroles to different scientific areas. 1.8A. Pyrroles found in nature. 1.8B. Pyrroles in commercial drugs.1.8 C. Applications of pyrroles in material science.

complicated frameworks, triggering the development of new methods and synthetic strategies to access various types of pyrrole scaffolds. Due to the paramount importance of the pyrrole nucleus in multiple chemical applications (Figure 1.8), new synthetic methods to access these nitrogenated heterocycles is essential.^{26,27,36,37} We aimed to use our pursuit of the total synthesis of the curvulamine alkaloids as a platform to explore novel chemistries and strategies to access electron rich natural products and functionalized pyrroles.

Preliminary studies on the biological properties of these secondary metabolites have revealed antibiotic or anti-inflammatory properties with MICs in the low micromolar range (*vide supra*). Low isolation yields have so far prevented testing against a wider range of pathogens. Our aim is to improve upon our pioneering synthesis of curvulamine to allow for a scalable and unifying strategy to access all the curvulamine alkaloids, with the end goal of performing a comprehensive evaluation of their biological activities. We also plan in due time, to investigate the antibiotic mode of action of curvulamine (1) and bipolamines (2-11). The synthesis of an alkyne-functionalized curvulamine or bipolamine derivative will enable us to perform isotopic tandem orthogonal proteolysis-activity based protein profiling (isoTOP-ABPP).³⁸ We predict that advanced proteomics will aid our goal of

gaining a fundamental understanding of the antimicrobial mechanism of action of these interesting metabolites.

1.6 Conclusion

The aim of this chapter is to provide a fundamental understanding of the curvulamine alkaloids and a framework of knowledge for subsequent chapters. The following chapters will discuss in greater detail our strategies, obstacles and solutions that ultimately culminated in the syntheses of curindolizine (2) and four *Bipolaris maydis* metabolites: bipolamines C,D,G, and I. It is also the goal of this dissertation to document the chemical reactivity and unique findings obtained in our campaign to synthesize these intriguing natural products.

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Chapter 2

The Synthesis of Novel Pyrrolo[1,2a]azepin-7-ones and the Total Synthesis of (+)-curindolizine

2.1 Introduction

The first part of this chapter will elaborate on the background development and the plausible mechanism of a microwave-induced method used to synthesize substituted pyrrolo[1,2-a]azepin-7-ones, an underexplored family of aromatic heterocycles. In addition, a description of the scope of this method will be provided as well as a brief discussion on some of the applications of pyrrolo[1,2-a]azepin-7-ones in the context of drug discovery. The remainder of the chapter will disclose our findings from exploring an initial biomimetic approach toward curindolizine (**2**) and the lessons we learned from this initial approach. Lastly, this chapter will recount the synthetic strategy used to realize the first total synthesis of curindolizine (**2**).

2.2 Forays into a Microwave-Induced Synthesis of New Pyrrolo[1,2-a]azepin-7ones

During our synthetic investigations toward curvulamine $(1)^1$, we were required to synthesize large quantities of pyrrole[1,2-a]azepin-7-one **25**. Initially, a 3-step procedure, inspired from the work of Radley and co-workers, was applied to the synthesis of 5,7-fused bicycle **36** (Figure 2.1A).² First, intramolecular conjugate addition of pyrrole-2-carboxaldehyde (**37**) onto methyl vinyl ketone **38** produced Michael addition product **39** in good yield using K₂CO₃ in DMF. Then, methyl ketone **39** was heated to reflux under basic conditions (NaOEt, EtOH) to provide a separable mixture of aldol condensation products **36** and **40** in a combined 71% yield. Finally, **41** was obtained after oxidation of



Figure 2.1 Investigations into the synthesis of pyrrolo[1,2-a]azepin-7-ones. **2.1A** Pioneering studies by Radley and co-workers. **2.2B** Studies by Flitsch and coworkers. **2.1C** Microwaved-induced synthesis of **25**.

the lithium enolate of 36 with Mukaiyama's reagent (42).^{3,4} This approach was not implemented during the synthesis of **1** because of the lack of selectivity in the aldol condensation step. In 1988, Flitsch reported the synthesis of analogous pyrroloazepinone **43** from vinylogous amide **44** using flash vacuum pyrolysis (FVP).^{5,6} In analogy to the work of Flitsch, a straightforward alternative was developed that used Boc-protected pyrrole-2-carboxaldehyde (27) as the starting material (Figure 2.1C). An intermolecular aldol condensation was performed after addition of the sodium enolate of (E)-4methoxybut-3-en-2-one (28) to 27 which gave enone 45 in 71% yield (NaHMDS, THF). Notably, concomitant Boc deprotection was observed which presumably occurred via a transesterification-type reaction of the initial aldol addition with the Boc-carbamate group followed by an E1cB reaction to generate enone 45. Microwave-induced thermal cyclization of 45 in the presence of base proceed to give pyrrolo[1,2-a]azepin-7-one 25 in 60% yield (DBU, PhMe, 160 °C). While several mechanisms could be proposed for this transformation, based on the identification of (Z)-45 during reaction, we hypothesized that this cyclization process could be occurring via two mechanisms. In one possible scenario, 25 could be formed after a conjugate addition of the pyrrole unit into the vinylogous ester followed by extrusion of methoxide via an E1cB reaction. Another possible mechanism





involves a concerted 10π electrocyclic ring closing reaction with concomitant elimination of methoxide to yield cyclized product **25**.

Substituted pyrrolo[1,2-a]azepin-7-ones have been employed in drug discovery, particularly in oncology and neuroleptic research.^{7,8} Given the limited number of existing methods to synthesize pyrroloazepinones,^{6,9-11} we investigated the application of this microwave-induced cyclization to synthesize novel pyrrolo[1,2-a]azepin-7-ones (Figure 2.3). We initially examined the synthesis of pyrroazepinones bearing different aryl groups (**46-48**). Compounds **46-48** were prepared in three steps. First. Suzuki coupling (Pd(dppf)Cl₂•DCM, K₂CO₃) of iodopyrrole **49** with an aryl substituted boronic acid gave the corresponding coupling products **50-52** in good yields. Second, the coupling products were subjected to a base-mediated intermolecular aldol condensation (NaHMDS,THF) with vinylogous ester **28** yielding enones **53-55**. Third, microwave irradiation (DBU, μ W) of **53-55** gave monosubstituted pyrroloazepinones **46-48** in excellent yields. Encouraged by these initial results, we synthesized disubstituted pyrrole **56** from methyl pyrrole **57** by means of a Vilsmeier-Haack formylation and Boc protection sequence (Oxalyl chloride, DMF *then* Boc₂O, THF). Aldol condensation of the sodium enolate of **28** and **56** produced

enone **58** in 73% yield using NaHMDS in THF. Thermal cyclization of **58** proceeded to give the corresponding pyrroloazepinone **59** in 52% yield (DBU, μ W). We extended the screening investigations of this method to the synthesis of more complex heterocycles such as **60** and **61**. Cyclization precursor **62** was expediently prepared from tetrahydroindole **63** after formylation (*t*-BuLi, DMF), Boc protection, and addition of the enolate of **28** to intermediate **64** (NaHMDS, THF). Indole **65** was made in two steps: first, regioselective formylation of **66** generated aldehyde **67** (POCl₃, DMF), second, an aldol addition of the sodium enolate of **28** to **67** (NaHMDS, THF). Both tetrahydrindole **62** and indole **65** underwent microwave-induced thermal cyclization (DBU, μ W) to give **61** and **60**, respectively albeit with diminished yields. Lastly, we sought to demonstrate that





Figure 2.3 Scope of microwave-assisted cyclization. **2.3A.** Synthesis of aryl substituted pyrroles. **2.3B.** Synthesis of tetrahydroindolizine and indolizine substrates. **2.3C.** Synthesis of simple pyrroles. **2.3D.** Microwave-assisted cyclization products and isolated yields.

pyrroles substituted with aldehydes and ketones were suitable substrates to prepare pyrroloazepinones. Pyrrolecarboxaldehyde **68** was used to make vinylogous ester **69** (NaHMDS, THF), and microwave irradiation of **69** in the presence of DBU yielded pyrroloazepinone **41**. We also observed that methyl ketone **70** could be transformed into intermediate **71**, which underwent smooth cyclization to **72** in 72% yield (DBU, μ W). In summary, the broad scope of this methodology illustrates its applicability to synthesize diverse pyrroloazepinones, scaffolds of potential importance in drug discovery.¹²⁻¹⁴

2.3 Bioinspired Route Toward Curindolizine

After securing a 10-step route to curvulamine (1),¹ we decided to embark on a total synthesis campaign toward curindolizine (2), the most complex *Curvularia* sp. IFB-Z10 secondary metabolite. **2** was serendipitously isolated from a large-scale cultivation broth of the *Curvularia* sp. fungus, and surprisingly was observed to be produced by the fungus preferentially over curvulamine (1). Motivated by this observation, Tan and co-workers investigated the biosynthetic origin of curindolizine (2).^{1,15,16} During these investigations procuramine (72) was isolated from the fungal cultivation broth, and interestingly, when **1** and **72** were added to the cell lysate containing the fungal proteins, curindolizine (2) was subsequently detected. This led to the researchers to concluded that curvulamine (1) was a precursor of curindolizine (2). The Tan group proposed a biosynthetic process where an enzymatic coupling between **1** and **73** (an intermediate derived from **72**) followed by additional enzymatic transformations ultimately yields curindolizine (2) (Figure 2.4).¹⁶ It is worth noting that while enzymes capable of promoting these reactions are suspected to be expressed by the fungus, Tan and co-workers did not perform sequencing experiments to identify the proteins responsible for producing **2**.¹⁷⁻¹⁹





Inspired by the way nature might synthesize curindolizine (2),¹⁶ we aimed to develop a synthetic plan to access electrophilic units analogous to enone **73**, which we would then couple with curvulamine (1). To this end we devised simple chemistry to first synthesize *tert*-butyl ester **74** and then transform **74** into ketone **72** and allylic alcohol **75**. A basecatalyzed aldol reaction between the lithium enolate of *tert*-butyl acetate (LiHMDS, THF) and **76** generated ester **74** in a 6:1 diastereomeric ratio and combined 82% yield. Friedel-Craft acylation of **74** under mild conditions (TMSOTf) generated procuramine (**72**) in good yields. Ester **74** underwent DIBAL-mediated reduction to generate an intermediate aldehyde, which cyclized and dehydrated upon treatment with SiO₂ to yield allylic alcohol **75**. With synthetic routes to **72** and **75**, we proceeded to screen for conditions to couple these units to curvulamine (1) (Figure 2.5B). Acid-catalyzed allylic substitution of **75** with curvulamine (1) in the presence of Brønsted acids PPTS (entry 1) or HCI (entry 2) failed to give curindolizine (**2**); instead, we only observed formation of 3,4-dimethyl-indolizine. Screening different Lewis acids such as $ZrCl_4$ (entry 3), $Sc(OTf)_3$ (entry 4), $Mo(CO)_6$ (entry 5) also failed to give any detectable amount of curindolizine (**2**); under these conditions curvulamine (**1**) and 3,4-dimethyl-indolizine were detected.^{20,21} Inspired by the work of Hartwig and co-workers, we attempted an iridium-mediated allylic substitution of **75** with curvulamine (**1**), but only **1** was recovered.^{22,23} Lastly, we subjected **75** to Tsuji-Trost conditions in an attempt to form a π -allyl complex which could be intercepted by **1**, but we failed to observe any desired product.^{24,25} In the face of these unfortunate results, we moved to explore nonbiomimetic approaches to access the complex alkaloid **2**.



Figure 2.5 Investigations toward a bioinspired synthesis of 2. 2.5A Syntheses of coupling partners. 2.5B. Selected coupling conditions explored.

2.4 Revised Approach Toward the Synthesis of Curindolizine

The rational solution to our problem was to install a handle on the eastern pyrrole and then investigate conditions for C-C bond-forming reactions. With that goal in mind, we explored conditions for a regioselective halogenation of **1** and **35**. Halogenation of **1** with NBS in DCM resulted on an intractable mixture of products and halogenation of methyl ketone **35** proceed to give **76** as the single product (NIS, MeOH).



Figure 2.6 Studies on the halogenation of curvulamine (1) and methyl ketone 35

Computational studies were performed on **35** to further understand the observed regioselectivity during the halogenation reaction. First, a conformational search on

A. Mulliken charge distribution B. Visualization of HOMO-LUMO $\begin{array}{c}
Me \\
0.097 \\
0.110 \\
0.158 \\
0.170 \\
35
\end{array}$ $\begin{array}{c}
Me \\
0.056 \\
0.556 \\
0.556
\end{array}$ B. Visualization of HOMO-LUMO

Figure 2.7 Computational studies on 35. 2.6A Mulliken values of 35. 2.6B The frontier orbitals of 35.

Maestro was performed on **35** to find its low-energy conformers. Then, a density functional theory computational analysis was performed on the lowest-energy conformation to calculate the vibrational frequencies in the gas phase at B3LYP/6-31G(d,p) level of theory. A population analysis in Gaussian provided us with the Mulliken values as well as information on the frontier orbitals (HOMO-LUMO) of **35** (Figure 2.7). The Mulliken partial charges (Figure 2.7A) indicates a higher electron density in the western pyrrole compared to the eastern pyrrole; and a visual representation of the frontier molecular orbitals in **35** (Figure 2.7B), shows they are located on the western pyrrole. Therefore, we hypothesize that the observed regioselectivity of the halogenation reaction might be controlled by electronic effects.^{26,27}

To overcome this regioselectivity challenge, we hypothesized that an intermediate with a deactivated western pyrrole, might undergo halogenation with the desired regioselectivity. Tetracycle **24** was identified as a suitable candidate to explore this hypothesis. The western pyrrole of **24** is conjugated to an electron-withdrawing group while the eastern pyrrole only has electron-donating substituents. Gratifyingly **24** underwent clean regio- and chemoselective iodination to give **77** in 88% yield (NIS,



Figure 2.8 Early investigation of the synthesis of curindolizine(2) from iodide 77



Figure 2.9 Studies to utilize pentacycle 79 to elaborate curindolizine (2).

Acetone). As anticipated, the more electron rich pyrrole preferably reacts with NIS and halogenates the position with the higher electron density on the eastern pyrrole.

Having secured a practical route to iodide **77**, we proceeded to continue our investigations toward the synthesis of curindolizine (**2**). The dihydroindolizine unit at C-8 was envisioned to be derived from **78**, which in turn could be made from iodide **77** in two steps. Addition of ethyl vinyl lithiate gave the corresponding product lactol product, but subsequent lithium-halogen exchange with *t*-BuLi followed by addition of the aldehyde electrophile gave reduction product **31** instead of **78** (entry 1). The observed product **31** was believed to be formed after the protonation of the organolithiate with the lactol proton.



Figure 2.10 Total synthesis of curindolizine (2).

To avoid this, the lactol was deprotonated prior the lithium-halogen exchange (entry 2). Unfortunately, the same reduction product **31** was isolated. Magnesium-halogen exchange using Turbo Grignard at cryogenic temperatures followed by addition of the electrophile gave a mixture of **31** and recovered starting material (entry 3). To explore other organometallic substrates, an organozinc was produced after transmetalation of the organolithiate with ZnBr₂; addition of the organozinc reagent to aldehyde (**76**) resulted in the isolation of lactol **31**. We rationalized that even after the deprotonation of the lactol such as in entry 2, the acidic alpha positions of the resulting ketone could still quench the organolithiate. To circumvent this problem, TMS protected lactol **79** was prepared after a CeCl₃-assited addition of ethyl vinyl ether lithiate to **77** followed by *in situ* silylation. Lithium-halogen exchange of **79** with *t*-BuLi followed by addition of **76** proceed smoothy to give the desired 1,2 addition product. Unfortunately, all attempts to elaborate this intermediate to the desired indolizine product were met with failure.

Next, we explored palladium-catalyzed reactions to utilize pyrrole **79** to complete the synthesis of curindolizine (**2**). Lithium-halogen exchange of **79** followed by ZnCl₂-mediated transmetalation gave the corresponding organozinc reagent, which underwent Pd-catalyzed Negishi coupling with **80** to yield a mixture of diastereomers **81** and **82** in 80% combined yield. The desired diastereomer **82** could be converted to ester **83** after a stereoselective reduction with Sml₂ and silyl ether deprotection. NOESY experiments confirmed the desired relative stereochemistry between this newly formed stereocenter and the vicinal methyl group. Racemic **83** was converted to enanotiopure **84** in four steps: first, base mediated thermodynamic isomerization of the C-2 stereocenter (NaOMe, MeOH); second, thiocarbamate formation (KHMDS, CISOPh, DMAP); third, Barton McCombie deoxygenation (Bu₃SnH, Et₃B, O₂); and fourth, a CBS reduction (BH₃•DMS, (*R*)-CBS catalyst). Based on our previous experiments in the preparation of **75**, DIBAL reduction of ester **84** gave an aldehyde which cyclized and dehydrated in the presence of SiO₂ to yield (+)-curindolizine (**2**) in 70% yield.

2.5 Conclusions

In this chapter, we have documented the result of our investigations on the mechanism and substrate scope of a microwave-induced method to access pyrroloazepinones. We have demonstrated the broad range of substrates that could be used in this transformation with the goal of enabling its implementation in areas such as drug discovery. Lastly, we have chronicled the first total synthesis of curindolizine (**2**), the only trimeric and most structurally complex curvulamine-type alkaloid.

2.6 Distribution of Credit and Acknowledgements

Pioneering studies into the microwave-assisted thermal cyclization were performed by Karl Thomas Haelsig and Jun Xuan. The investigation and synthesis **41-72** were performed by Paulo Andre Machicao Tello with refinement from Professor Thomas J. Maimone. The synthesis of **2** was spearheaded by Jun Xuan. Characterization of all compounds was completed by Paulo Andre Machicao Tello, Karl Thomas Haelsig, and

Jun Xuan. We thank the NIH NIGMS (R01GM136945 to Thomas J. Maimone, and diversity supplement to Paulo Andre Machicao Tello) for financial support. We thank Dr. Hasan Celik and Dr. Jeffrey G. Pelton for nuclear magnetic resonance spectroscopic assistance (NIH grant GM68933). We also thank QB3/Chemistry Mass Spectrometry Facility scientist Dr. Zhongroui Zhou for mass spectrometry assistance.

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2.8 Experimental Procedures and Characterization Data

2.8.1 General Procedures

All reactions were performed in flame- or oven-dried glassware under a positive pressure of nitrogen or argon, unless otherwise noted. Air-and moisture-sensitive liquids were transferred via syringe. When indicated, solvents or reagents were degassed by sparging with argon for 10 min in an ultrasound bath at 25 °C. Volatile solvents were removed under reduced pressure rotary evaporation below 35 °C. Analytical and preparative thin-layer chromatography (TLC) were performed using glass plates precoated with silica gel (0.25-mm, 60-Å pore size, Merck TLC Silicagel 60 F₂₅₄) impregnated with a fluorescent indicator (254 nm). TLC plates were visualized by exposure to ultraviolet light (UV) and then were stained by submersion in an ethanolic anisaldehyde solution or ceric ammonium molybdate solution, followed by brief heating on a hot plate. Flash column chromatography was performed with silica gel purchased from Silicycle (SiliaFlash[®], 60 Å, 230-400 mesh, 40-63 µm). Ethyl vinyl ether and 2bromopropanic acid methyl ester were distilled over calcium hydride prior to use. NaHMDS solutions were purchased from Sigma. All other reagents were used as received from commercial sources, unless stated otherwise. Anhydrous tetrahydrofuran (THF), dichloromethane (DCM), methanol (MeOH), dimethylformamide (DMF), and toluene (PhMe) were obtained by passing these previously degassed solvents through activated alumina columns. Proton nuclear magnetic resonance (¹H NMR) spectra and carbon nuclear magnetic resonance (¹³C NMR) spectra were recorded on Bruker AV-600 spectrometer at 23 °C. Proton chemical shifts are expressed as parts per million (ppm, δ scale) and are referenced to residual solvent (CDCl₃: δ 7.26, C₆D₆: δ 7.16), unless stated otherwise. Carbon chemical shifts are expressed as parts per million (ppm, δ scale) and are referenced to the solvent (CDCl₃: δ 77.16, C₆D₆: δ 128.06), unless stated otherwise. Data is represented as follows: chemical shift, multiplicity (s = singlet, d = doublet, dd = doublet of doublets, ddd, doublet of doublet of doublet, dt = triplet of doublets, t = triplet, q = quartet, m = multiplet, br = broad), coupling constant (J) in Hertz (Hz), and integration. Infrared (IR) spectra were recorded on a Bruker Alpha FT-IR spectrometer as thin films and are reported in frequency of absorption (cm⁻¹). Only selected resonances are reported. High-resolution mass spectra (HRMS) were obtained by the mass spectrometry facility at the University of California, Berkeley using a Finnigan LTQFT mass spectrometer (Thermo Electron Corporation). X-ray diffraction data was collected at the Small Molecule X-ray Crystallography Facility (CheXray) at University of California, Berkeley using a Rigaku XtaLAB P200 equipped with a MicroMax 007HF rotating anode and Pilatus3 R 200K-A hybrid pixel array detector. Data were collected using CuKa radiation ($\lambda = 1.5418$ Å).

2.8.2 Experimental Procedures and Tabulated Characterization Data



Pyrroloazepinone 41: Enone 36 (2.52 g, 17.1 mmol, 1.0 equiv.) was transferred (in benzene) to a 250 mL round bottom flask and was concentrated to dryness in vacuo. The 250 mL round bottom flask was then sealed, evacuated, and back filled with nitrogen (3x). The sealed flask was charged with THF (70 mL), and cooled to -78 °C. LiHMDS (1.0 M in THF, 20.4 mL, 20.4 mmol, 1.2 equiv.) was added dropwise, and the resulting suspension was stirred for 30 min at -78 °C. In a separate flask, Mukaiyama's reagent 42 (4.04 g, 18.8 mmol, 1.1 equiv.) was azeotropically dried with benzene (3x) and then dissolved in THF (10 mL) under an atmosphere of nitrogen. The solution was then transferred via cannula to the reaction mixture, with an additional THF (5 mL) rinse ensuring quantitative transfer. Upon completion of the consumption of 36 as indicated by TLC, the reaction was guenched with saturated ag. NaHCO₃ (50 mL). The solution was extracted with EtOAc (3 x 50 mL). The combined organic layers were washed with brine (50 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo to afford a brown residue. The resulting crude residue was purified by column chromatography (15% EtOAc in hexanes \rightarrow 35% EtOAc in hexanes) to afford 41 (1.53 g, 10.5 mmol, 62%) as a yellow solid. TLC: R_f = 0.5 (40% EtOAc in hexanes); ¹H NMR (600 MHz, C₆D₆) δ 6.39 (d, J = 12.3 Hz, 1H), 6.21 (dd, J = 12.3, 2.3 Hz, 1H), 6.13 (t, J = 2.3 Hz, 1H), 6.10 (dd, J = 12.3 Hz, 1H), 3.9, 1.6 Hz, 1H), 6.02 (d, J = 10.4 Hz, 1H), 5.98 (t, J = 3.3 Hz, 1H), 5.69 (dd, J = 10.4, 2.4 Hz, 1H); ¹³C NMR (151 MHz, C_6D_6) δ 186.4, 132.7, 128.7, 127.6, 126.3(2C), 118.1. 115.8. 111.9: HRMS (m/z): (ESI) calcd. For C₉H₈ON [M+H]⁺: 146.0600. found 146.0599.



Aldehyde 27: To a 1 L round bottom flask containing 5-methylpyrrole-2-carboxaldehyde (32.4 g, 296.9 mmol, 1.0 equiv.) was added THF (590 mL). The vigorously stirring solution was cooled to 0 °C, and DMAP (1.81 g, 14.8 mmol, 0.05 equiv.) and Boc₂O (77.7 g, 356 mmol, 1.2 equiv.) were added in a single portion sequentially. The resulting suspension was removed from the cooling bath and warmed to room temp, at which point the septum was removed allowing for evolved gases to escape. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NaHCO₃ (300 mL) and stirred for 30 min. The biphasic reaction mixture was poured into a separatory funnel and extracted with EtOAc (3 x 500 mL). The combined organic layers were washed with brine (100 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The dark brown colored crude product was first filtered through a short silica gel plug (eluting with 50% EtOAc in hexanes), concentrated *in vacuo*, and purified by column chromatography (10% EtOAc in hexanes \rightarrow 40% EtOAc) to afford

aldehyde **27** (56.5 g, 270 mmol, 91% yield) as a yellow oil. **TLC:** $R_f = 0.3$ (40% EtOAc in hexanes); ¹H NMR (600 MHz, C₆D₆) δ 9.33 (s, 1H), 6.86 (d, J = 3.6 Hz, 1H), 5.63 (d, J = 3.4 Hz, 1H), 2.13 (s, 3H), 1.28 (s, 9H); ¹³C NMR (151 MHz, C₆D₆) δ 178.3, 148.5, 138.2, 134.4, 120.7, 111.1, 83.9, 26.5, 14.1(3C); **IR** (thin film) v_{max} (cm⁻¹): 2979, 2932, 1745, 1663, 1485, 1300, 1124, 861, 798, 777; **HRMS (m/z):** (ESI) calcd. for C₁₁H₁₆NO₃ [M+H]⁺: 210.1125, found 210.1126.



Compound 45: To a flame-dried 1L round bottom flask was added (*E*)-4-methoxybut-3en-2-one (28) (8.13 mL, 79.7 mmol, 1.0 equiv.) and THF (478 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 104 mL, 104 mmol, 1.3 equiv.) was transferred via cannula to the reaction mixture. The resulting suspension was then stirred for 30 min at -78 °C. In a separate flask, aldehyde 27 (20.0 g, 95.7 mmol, 1.2 equiv.) was azeotropically dried with benzene (3x) and then dissolved in THF (20 mL) under an atmosphere of nitrogen. The aldehyde solution was then transferred via cannula to the reaction mixture, with an additional THF (5 mL) rinse ensuring guantitative transfer. The resulting reaction mixture was then stirred at -78 °C for 1 hour, and upon consumption of the starting material as indicated by TLC, the reaction was then guenched with saturated aq. NH4CI (200 mL) at -78°C. The mixture was warmed to room temperature and extracted with EtOAc (3 x 400 mL). The combined organic layers were washed with brine (200 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The dark brown colored crude material was purified by column chromatography (10% EtOAc in hexanes \rightarrow 45% EtOAc in hexanes) to afford dienone **45** (10.8 g, 56.5 mmol, 71%) as an orange solid. **TLC:** $R_f = 0.4$ (40% EtOAc in hexanes); ¹**H NMR** (600 MHz, CDCl₃) δ 9.08 (s, 1H), 7.70 (d, J = 12.3 Hz, 1H), 7.48 (d, J = 15.7Hz, 1H), 6.50 (br s, 1H), 6.42 (d, J = 15.7 Hz, 1H), 5.98 (br s, 1H), 5.86 (d, J = 12.4 Hz, 1H), 3.75 (s, 3H), 2.34 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 189.3, 162.6, 135.1, 132.8, 128.1, 118.0, 116.8, 109.8, 104.5, 57.6, 13.4; **IR** (thin film) v_{max} (cm⁻¹): 3255, 1600, 1546, 1274, 1089, 1035, 778, 618; HRMS (m/z): (ESI) calcd. for C11H14NO2 [M+H]+: 192.1019, found 192.1020.



Compound 71: To a flame-dried reaction tube was added (*E*)-4-methoxybut-3-en-2one (**28**). (60 mg, 0.60 mmol, 1.0 equiv.) and THF (3 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 0.78 mL, 0.78 mmol, 1.3 equiv.) was added dropwise to the reaction mixture. The resulting suspension was then stirred for 1 hour. In a separate flask 2-Acetyl-1-tert-butoxy carbonyl pyrrole (**70**) (150 mg, 0.72 mmol, 1.2 equiv.) was dissolved in THF under an atmosphere of nitrogen. The ketone solution was then added dropwise to the reaction mixture at -78 °C. The resulting reaction mixture was warmed up to -40 °C and stirred for 2 hours, and upon consumption of the starting material as indicated by TLC, the reaction was then quenched with *aq.* NH₄Cl (1.5 mL). The mixture was warmed to room temperature and extracted with EtOAc (3 x 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 40% EtOAc in hexanes) to afford dienone **71** (69 mg, 0.36 mmol, 60%). **TLC:** R_f = 0.30 (30% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 7.72 (d, *J* = 12.4 Hz, 1H), 7.43 – 7.25 (m, 1H), 6.52 (ddd, *J* = 3.9, 2.7, 1.4 Hz, 1H), 6.32 (td, *J* = 2.7, 1.4 Hz, 1H), 6.20 (dt, *J* = 3.7, 2.5 Hz, 1H), 6.07 (q, *J* = 1.2 Hz, 1H), 5.66 (d, *J* = 12.4 Hz, 1H), 3.00 (s, 3H), 2.60 (d, *J* = 1.1 Hz, 3H); ¹³**C NMR** (151 MHz, C₆D₆) δ 188.6, 162.3, 143.7, 133.5, 121.3, 117.7, 111.8, 110.7, 108.4, 57.2, 16.6; **IR** (thin film) v_{max} (cm⁻¹): 1656, 1617,1542, 1422, 1308, 1248, 1207, 1131, 1093, 1044, 858, 738, 737; **HRMS (m/z)**: (ESI) calcd. for C₁₁H₁₄NO₂ [M+H]⁺: 192.1019, found 192.1019.



Compound 65: To a flame-dried reaction tube was added (*E*)-4-methoxybut-3-en-2-one (28) (25 mg, 0.25 mmol, 1.0 equiv.) and THF (1.5 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 0.33 mL, 0.33 mmol, 1.3 equiv.) was added dropwise to the reaction mixture. The resulting suspension was then stirred for 30 min. In a separate flask N-Boc-indole-2-carboxaldehyde 67 (76 mg, 0.31 mmol, 1.2 equiv.) was dissolved in THF under an atmosphere of nitrogen. The aldehyde solution was then added dropwise to the reaction mixture at -78 °C. The resulting reaction stirred for 1 hour at -78 °C, and upon consumption of the starting material as indicated by TLC, the reaction was then guenched with aq. NH4Cl (1.5 mL). The mixture was warmed to room temperature and extracted with EtOAc (3 x 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 50% EtOAc in hexanes) to afford dienone 65 (33 mg, 0.15 mmol, 60%) as a yellow solid. TLC: Rf = 0.30 (30% EtOAc in hexanes); ¹H NMR (600 MHz, C₆D₆) δ 7.76 (d, J = 12.4 Hz, 1H), 7.64 (d, J = 15.9 Hz, 1H), 7.54 (dd, J = 8.0, 1.0 Hz, 1H), 7.37 (s, 1H), 7.21 (ddd, J = 8.2, 7.0, 1.2 Hz, 1H), 7.13 - 7.05 (m, 2H), 6.57 (d, J = 2.1 Hz, 1H), 6.33 (d, J = 15.9 Hz, 1H), 5.75 (d, J = 12.4 Hz, 1H), 2.99 (s, 3H); ¹³C NMR (151 MHz, C₆D₆) δ 186.9, 163.2, 138.2, 134.4, 131.3, 129.3, 124.8, 124.6, 121.9, 120.9, 111.5, 108.5, 105.1, 57.4; IR (thin film) vmax (cm⁻¹): 1644, 1605, 1588, 1345, 1307, 1254, 1126, 1068, 752, 738; HRMS (m/z): (ESI) calcd. for C14H14NO2 [M+H]+: 228.1019, found 228.1021.



Compound 50: To a flame-dried round-bottom flask were added *tert*-butyl-2-formyl-5iodo-1H-pyrrole-1-carboxylate 49 (100 mg, 0.31 mmol, 1.0 equiv.), phenylboronic acid (46 mg, 0.38 mmol, 1.2 equiv.), Pd(dppf)Cl₂•DCM (30 mg, 36 µmol, 0.1 equiv.), K₃PO₄ (140 mg, 0.66 mmol, 2.1 equiv.), dioxane (1.5 mL) and H₂O (75 µL). The resulting solution was sparged with argon for 5 min, then the reaction mixture was stirred at 85 °C for 1 hour. Upon consumption of the starting material as indicated by TLC, the reaction was then cooled down to room temperature and diluted with Et_2O (3 mL) and H_2O (3 mL). The biphasic mixture was poured into a separatory funnel and extracted with Et₂O (3 x 10 mL). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 30% EtOAc in hexanes) to afford aldehyde 50 (54 mg, 0.20 mmol, 65%). **TLC:** Rf = 0.60 (30% EtOAc in hexanes); ¹**H NMR** (600 MHz, C_6D_6) δ 9.95 (s, 1H), 7.24 – 7.18 (m, 2H), 7.06 – 7.01 (m, 3H), 6.79 (d, J = 3.8 Hz, 1H), 5.95 (d, J = 3.8 Hz, 1H), 1.15 (s, 9H); ¹³C NMR (151 MHz, Acetone) δ 180.5, 150.0, 141.9, 135.7, 133.1, 129.4(3C), 129.2(2C), 121.4, 113.1, 86.4, 27.4(3C); **IR** (thin film) v_{max} (cm⁻¹): 2982, 2935, 1765, 1743, 1663, 1506, 1414, 1396, 1371, 1300, 1285, 1260, 1140, 1075; HRMS (m/z): (ESI) calcd. for C₁₆H₁₇NO₃Na [M+Na]⁺: 294.1101, found 294.1100.



Compound 53: To a flame-dried reaction tube was added (E)-4-methoxybut-3-en-2-one (**28**) (37 mg, 0.37 mmol, 1.0 equiv.) and THF (1.5 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 0.48 mL, 0.48 mmol, 1.3 equiv.) was added dropwise to the reaction mixture. The resulting suspension was then stirred for 30 min. In a separate flask aldehyde **50** (133 mg, 0.49 mmol, 1.3 equiv.) was dissolved in THF under an atmosphere of nitrogen. The aldehyde solution was then added dropwise to the reaction mixture at -78 °C. The resulting reaction stirred for 1 hour at -78 °C, and upon consumption of the starting material as indicated by TLC, the reaction was then quenched with *aq.* NH₄Cl (3.0 mL). The mixture was warmed to room temperature and extracted with EtOAc (3 x 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo.* The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 30% EtOAc in hexanes) to afford dienone **53** (50 mg, 0.20 mmol, 54%). **TLC:** R_f = 0.30 (40% EtOAc in hexanes); **1H NMR** (600 MHz, C₆D₆) δ 9.32

(s, 1H), 7.82 (d, J = 15.7 Hz, 1H), 7.73 (d, J = 12.3 Hz, 1H), 7.41 (d, J = 7.7 Hz, 2H), 7.07 – 7.00 (m, 2H), 6.63 – 6.57 (m, 2H), 6.53 (dd, J = 3.8, 2.5 Hz, 1H), 5.74 (d, J = 12.4 Hz, 1H), 2.98 (s, 3H); ¹³**C** NMR (151 MHz, C₆D₆) δ 187.9, 162.9, 137.0, 132.4, 132.0, 130.9, 129.1(2C), 127.3, 125.1(2C), 121.1, 115.6, 109.5, 105.2, 57.3; **IR** (thin film) v_{max} (cm⁻¹): 1637, 1600, 1549, 1462, 1291, 1258, 1206, 1142, 1090, 1045, 975, 759; **HRMS (m/z)**: (ESI) calcd. for C₁₆H₁₆NO₂ [M+H]⁺: 254.1176, found 254.1176.



Compound 51: To a flame-dried round-bottom flask were added tert-butyl-2-formyl-5iodo-1*H*-pyrrole-1-carboxylate 49 (120 mg, 0.37 mmol. 1.0 eauiv.). 4fluorophenylboronic acid (60 mg, 0.43 mmol, 1.2 equiv.), Pd(dppf)Cl₂•DCM (30 mg, 36 µmol, 0.1 equiv.), K₃PO₄ (130 mg, 0.94 mmol, 2.5 equiv.), dioxane (1.5 mL) and H₂O (75 µL). The resulting solution was sparged with argon for 5 min, then the reaction mixture was stirred at 85 °C for 1 hour. Upon consumption of the starting material as indicated by TLC, the reaction was then cooled down to room temperature and diluted with Et₂O (3 mL) and H₂O (3 mL). The biphasic mixture was poured into a separatory funnel and extracted with Et₂O (3 x 10 mL). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 30% EtOAc in hexanes) to afford aldehyde **51** (71 mg, 0.25 mmol, 68%). **TLC:** $R_f = 0.65$ (30% EtOAc in hexanes); ¹**H NMR** (600 MHz, C_6D_6) δ 9.92 (s, 1H), 7.02 – 6.96 (m, 2H), 6.78 (d, J = 3.7 Hz, 1H), 6.73 - 6.66 (m, 2H), 5.86 (d, J = 3.7 Hz, 1H), 1.15 (s, 9H); ¹³C NMR (151 MHz, C₆D₆) δ 179.5, 164.0, 162.4, 149.3, 140.0, 135.4, 131.1, 131.0, 120.9, 115.4, 115.2, 112.8, 85.4, 27.1(3C); **IR** (thin film) v_{max} (cm⁻¹): 2982, 1744, 1663, 1606, 1462, 1422, 1397, 1299, 1260, 1220, 1159, 1139, 840, 802; HRMS (m/z): (ESI) calcd. for C16H17NFO3 [M+H]+: 290.1187, found 290.1189.



Compound 54: To a flame-dried reaction tube was added (*E*)-4-methoxybut-3-en-2-one (**28**) (39 mg, 0.39 mmol, 1.0 equiv.) and THF (3.0 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 0.51 mL, 0.51 mmol, 1.3 equiv.) was added dropwise to thereaction mixture. The resulting suspension was then stirred for 1 hour. In a separate flask aldehyde **51** (140 mg, 0.48 mmol, 1.2 equiv.) was dissolved in THF under an atmosphere of nitrogen. The aldehyde solution was then added dropwise to the reaction mixture at -78 °C. The resulting reaction stirred for 1 hour at -78 °C, and upon consumption of the starting material as indicated by TLC, the reaction was then quenched

with *aq.* NH₄Cl (1.5 mL). The mixture was warmed to room temperature and extracted with EtOAc (3 x 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 40% EtOAc in hexanes) to afford dienone **54** (51 mg, 0.19 mmol, 48%). **TLC:** R_f = 0.65 (30% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 9.30 (s, 1H), 7.81 (d, *J* = 15.7 Hz, 1H), 7.68 (d, *J* = 12.4 Hz, 1H), 6.84 – 6.77 (m, 2H), 6.63 (d, *J* = 15.7 Hz, 1H), 6.59 (dd, *J* = 3.8, 2.4 Hz, 1H), 6.39 (dd, *J* = 3.8, 2.5 Hz, 1H), 5.73 (d, *J* = 12.4 Hz, 1H), 2.97 (s, 3H); ¹³**C NMR** (151 MHz, C₆D₆) δ 188.1, 163.3, 162.9, 161.7, 136.1, 132.0, 130.9, 126.8(2C), 121.2, 116.1, 115.9, 115.5, 109.4, 105.0, 57.3; **IR** (thin film) v_{max} (cm⁻¹): 1634, 1618, 1599, 1562, 1516, 1467, 1344, 1233, 1211, 851, 834, 781, 684; **HRMS (m/z)**: (ESI) calcd. for C₁₆H₁₅NFO₂ [M+H]⁺: 272.1081, found 272.1082.



Compound 52: To a flame-dried round-bottom flask were added *tert*-butyl-2-formyl-5iodo-1*H*-pyrrole-1-carboxylate 49 (110)0.34 mmol. 1.0 eauiv.). mq, 4methoxyphenylboronic acid (80 mg, 0.53 mmol, 1.6 equiv.), Pd(dppf)Cl₂•DCM (25 mg, 31 µmol, 0.09 equiv.), K₂CO₃ (120 mg, 0.87 mmol, 2.6 equiv.), dioxane (2.0 mL) and H₂O (0.1 mL). The resulting solution was sparged with argon for 5 min, then the reaction mixture was stirred at 85 °C for 3 hours. Upon consumption of the starting material as indicated by TLC, the reaction was then cooled down to room temperature and diluted with EtOAc (3 mL) and H₂O (3 mL). The biphasic mixture was poured into a separatory funnel and extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 40% EtOAc in hexanes) to afford aldehyde 52 (70 mg, 0.23 mmol, 68%). TLC: Rf = 0.65 (30% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 9.86 (s, 1H), 7.26 – 7.20 (m, 2H), 6.77 (d, J = 3.8 Hz, 1H), 6.72 - 6.67 (m, 2H), 5.99 (d, J = 3.7 Hz, 1H), 3.24 (s, 3H), 1.24 (s, 9H); ³C NMR(151 MHz, C₆D₆) δ 179.0, 160.4, 149.8, 141.5, 135.0(2C), 130.5, 124.9, 121.4(2C), 113.9, 112.1, 85.2, 54.8, 27.2(2C); **IR** (thin film) v_{max} (cm⁻¹): 1762, 1743, 1660, 1611, 1577, 1463, 1370, 1287, 1250, 1178, 1137, 1075, 1032, 835; HRMS (m/z): (ESI) calcd. For C17H19NO4Na[M+Na]+: 324.1206, found 324.1207.



Compound 55: To a flame-dried reaction tube was added (E)-4-methoxybut-3-en-2-one (28) (40 mg, 0.40 mmol, 1.0 equiv.) and THF (3.0 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 0.52 mL, 0.52 mmol, 1.3 equiv.) was added dropwise to the reaction mixture. The resulting suspension was then stirred for 1 hour. In a separate flask aldehyde 51 (144 mg, 0.47 mmol, 1.2 equiv.) was dissolved in THF under an atmosphere of nitrogen. The aldehyde solution was then added dropwise to the reaction mixture at -78 °C. The resulting reaction stirred for 1 hour at -78 °C, and upon consumption of the starting material as indicated by TLC, the reaction was then quenched with ag. NH₄Cl (3.0 mL). The mixture was warmed to room temperature and extracted with EtOAc (3 x 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 40% EtOAc in hexanes) to afford dienone 55 (53 mg, 0.19 mmol, 47%). ¹H NMR (600 MHz, Acetone) δ 10.70 (s, 1H), 7.72 – 7.63 (m, 3H), 7.49 (d, J = 15.8 Hz, 1H), 7.00 – 6.95 (m, 2H), 6.77 (d, J = 15.7 Hz, 1H), 6.66 (dd, J = 3.7, 2.4Hz, 1H), 6.56 (dd, J = 3.8, 2.4 Hz, 1H), 5.90 (d, J = 12.5 Hz, 1H), 3.83 (s, 3H), 3.78 (s, 3H); ¹³C NMR (151 MHz, Acetone) δ 187.8, 162.7, 160.1, 137.7, 131.6, 130.9, 126.7(2C), 125.7, 120.4, 117.4, 115.2(2C), 108.5, 105.9, 58.0, 55.7; IR (thin film) v_{max} (cm⁻¹): 1599, 1569, 1549, 1517, 1468, 1439, 1281, 1252, 1206, 1183, 1088, 1075; HRMS (m/z): (ESI) calcd. for C₁₇H₁₈NO₃ [M+H]⁺: 284.1281, found 284.1281.



Compound 69: To a flame-dried reaction tube was added (*E*)-4-methoxybut-3-en-2-one (28) (60 mg, 0.60 mmol, 1.0 equiv.) and THF (3 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 0.78 mL, 0.78 mmol, 1.3 equiv.) was added dropwise to the reaction mixture. The resulting suspension was then stirred for 30 min. In a separate flask N-Boc-pyrrole-2-carboxaldehyde 68 (140 mg, 0.72 mmol, 1.2 equiv.) was dissolved in THF under an atmosphere of nitrogen. The aldehyde solution was then added dropwise to the reaction mixture at -78 °C. The resulting reaction stirred for 1 hour at -78 °C, and upon consumption of the starting material as indicated by TLC, the reaction was then guenched with aq. NH₄Cl (3.0 mL). The mixture was warmed to room temperature and extracted with EtOAc (3 x 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated in vacuo. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 40% EtOAc in hexanes) to afford dienone 69 (50 mg, 0.28 mmol, 47%). TLC: Rf = 0.35 (30% EtOAc in hexanes); ¹H NMR (600 MHz, C₆D₆) δ 7.78 (d, J = 12.4 Hz, 2H), 7.64 (d, J = 15.8 Hz, 1H), 6.57 - 6.39 (m, 1H), 6.37 – 6.24 (m, 2H), 6.18 (dt, J = 3.6, 2.5 Hz, 1H), 5.73 (d, J = 12.3 Hz, 1H), 2.96 (s, 3H); ¹³C NMR (151 MHz, C₆D₆) δ 187.4, 162.6, 131.6, 129.3, 122.4, 121.0, 114.1, 111.1, 105.0, 57.2; **IR** (thin film) v_{max} (cm⁻¹): 1734, 1641, 1603, 1568, 1541, 1437, 1414, 1290, 1125, 1088, 1035, 977, 740; HRMS (m/z): (ESI) calcd. for C10H12NO2 [M+H]+: 178.0863, found 178.0861.



Compound 64: To a round bottom flask containing 4,5,6,7-tetrahydro-1*H*-indole-2-carbaldehyde (55 mg, 0.37 mmol, 1.0 equiv.) was added THF (2 mL). To the resulting solution were added DMAP (2 mg, 16.4 µmol, 0.04 equiv.) and Boc₂O (105 mg, 0.48 mmol, 1.3 equiv.) in a single portion sequentially. The reaction mixture was stirred at room temperature for 1 hour, and upon consumption of the starting material as indicated by TLC, the mixture was diluted with EtOAc (3 mL) and quenched with aq. NaHCO₃ (5 mL). The biphasic reaction mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed were dried over Na₂SO₄, filtered, and concentrated in vacuo. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 40% EtOAc in hexanes) to afford aldehyde 64 (59 mg, 0.24 mmol, 65%). **TLC:** R_f = 0.50 (10% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 10.21 (s, 1H), 6.85 (s, 1H), 2.64 (tt, J = 6.4, 1.6 Hz, 2H), 2.12 (tt, J = 6.1, 1.7 Hz, 2H), 1.41 (pd, J = 5.9, 3.3 Hz, 2H), 1.38 – 1.32 (m, 2H), 1.28 (s, 9H); ¹³C NMR (151 MHz, C₆D₆) δ180.1, 149.4, 137.8, 134.6, 122.3, 121.6, 84.4, 27.6(3C), 25.4, 23.1, 22.9, 22.7; IR (thin film) v_{max} (cm⁻¹): 2935, 1739, 1648, 1483, 1462, 1416, 1348, 1333, 1296, 1259, 1144, 1125, 1086, 849; HRMS (m/z): (ESI) calcd. for C14H19NO3Na[M+Na]+: 272.1257, found 272.1258.



Compound 62: To a flame-dried reaction tube was added (*E*)-4-methoxybut-3-en-2-one (28) (60 mg, 0.60 mmol, 1.0 equiv.) and THF (3.5 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 0.78 mL, 0.78 mmol, 1.3 equiv.) was added dropwise to the reaction mixture. The resulting suspension was then stirred for 1 hour. In a separate flask aldehyde 64 (200 mg, 0.80 mmol, 1.3 equiv.) was dissolved in THF under an atmosphere of nitrogen. The aldehyde solution was then added dropwise to the reaction mixture at -78 °C. The resulting reaction stirred for 1 hour at -78 °C, and upon consumption of the starting material as indicated by TLC, the reaction was then guenched with ag. NH₄Cl (3.0 mL). The mixture was warmed to room temperature and extracted with EtOAc (3 x 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 50% EtOAc in hexanes) to afford dienone 62 (65 mg, 0.28 mmol, 47%). TLC: R_f = 0.45 (30% EtOAc in hexanes); ¹H NMR (600 MHz, C₆D₆) δ 7.83 (d, J = 12.3 Hz, 1H), 7.69 (d, J = 15.7 Hz, 1H), 7.38 (s, 1H), 6.32 (d, J = 15.7 Hz, 1H),6.27 (d, J = 2.5 Hz, 1H), 5.81 (d, J = 12.3 Hz, 1H), 3.01 (s, 3H), 2.40 (tt, J = 4.8, 2.0 Hz, 2H), 2.16 (q, J = 4.3, 3.2 Hz, 2H), 1.53 (td, J = 3.8, 1.9 Hz, 4H); ¹³C NMR (151 MHz, C_6D_6) δ 187.2, 162.2, 132.8, 131.9, 127.6, 120.5, 119.1, 114.1, 105.2, 57.1, 23.9, 23.4, 23.2, 23.1; **IR**(thin film) v_{max} (cm⁻¹): 2932, 1637, 1602, 1570, 1546, 1411, 1362, 1318, 1285, 1198, 1119, 1082, 975; **HRMS (m/z)**: (ESI) calcd. for C₁₄H₁₈NO₂ [M+H]⁺: 232.1332, found 232.1333.



Compound 56: To a round bottom flask containing 4-methyl-5-phenyl-1*H*-pyrrole-2carbaldehyde (155 mg, 0.84 mmol, 1.0 equiv.) was added THF (5 mL). To the resulting solution were added DMAP (4.9 mg, 40 µmol, 0.05 equiv.) and Boc₂O (253 mg, 1.16 mmol, 1.4 equiv.) in a single portion sequentially. The reaction mixture was stirred at room temperature for 1 hour, and upon consumption of the starting material as indicated by TLC, the mixture was diluted with EtOAc (5 mL) and guenched with ag. NaHCO₃ (5 mL). The biphasic reaction mixture was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 35% EtOAc in hexanes) to afford aldehyde 56 (132 mg, 0.46 mmol, 55%). TLC: R_f = 0.60 (10% EtOAc in hexanes); ¹H NMR (600 MHz, C₆D₆) δ 10.21 (s, 1H), 7.09 – 7.00 (m, 5H), 6.81 (s, 1H), 1.70 (s, 3H), 1.08 (s, 9H); ¹³C NMR (151 MHz, Acetone) δ 180.9, 149.8, 138.2, 134.4, 133.1, 130.5(2C), 129.1(2C), 122.2, 121.9, 85.7, 27.3(3C), 11.3; **IR** (thin film) v_{max} (cm⁻¹): 1739, 1663, 1605, 1508, 1465, 1443, 1415, 1371, 1337, 1300, 1154, 1089, 703; HRMS (m/z): (ESI) calcd. for C17H19NO3Na [M+Na]+: 308.1257, found 308.1257.



Compound 58: To a flame-dried reaction tube was added (*E*)-4-methoxybut-3-en-2-one (**28**) (26 mg, 0.26 mmol, 1.0 equiv.) and THF (1.5 mL). The resulting solution was cooled to -78 °C and NaHMDS (1.0 M in THF, 0.33 mL, 0.33 mmol, 1.3 equiv.) was added dropwise to the reaction mixture. The resulting suspension was then stirred for 1 hour. In a separate flask aldehyde **56** (82 mg, 0.29 mmol, 1.1 equiv.) was dissolved in THF under an atmosphere of nitrogen. The aldehyde solution was then added dropwise to the reaction mixture at -78 °C. The resulting reaction stirred for 1 hour at -78 °C, and upon consumption of the starting material as indicated by TLC, the reaction was then quenched with *aq.* NH₄Cl (1.5 mL). The mixture was warmed to room temperature and extracted with EtOAc (3 x 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude material was purified by column chromatography

(0% EtOAc in hexanes \rightarrow 50% EtOAc in hexanes) to afford dienone **58** (50 mg, 0.19 mmol, 73%). **TLC:** R_f = 0.35 (30% EtOAc in hexanes); ¹H NMR (600 MHz, Acetone) δ 7.67 (d, *J* = 12.5 Hz, 1H), 7.62 – 7.58 (m, 2H), 7.51 – 7.42 (m, 3H), 7.31 (t, *J* = 7.4 Hz, 1H), 6.81 (d, *J* = 15.7 Hz, 1H), 6.56 (s, 1H), 5.90 (d, *J* = 12.5 Hz, 1H), 3.78 (s, 3H), 2.26 (s, 3H); ¹³C NMR (151 MHz, Acetone) δ 187.8, 162.8, 133.9, 133.7, 131.6, 129.5(2C), 127.6(3C), 120.7, 119.6, 118.7, 118.6, 106.0, 58.0, 12.9; IR (thin film) v_{max} (cm⁻¹): 1636, 1597, 1550, 1458, 1437, 1410, 1350, 1331, 1291, 1267, 1082, 979, 768; HRMS (m/z): (ESI) calcd. for C₁₇H₁₈NO₂[M+H]⁺: 268.1332, found 268, 1332.

Standard Procedure for the microwave-assisted synthesis of substituted pyrroloazepinones

To a 5 mL Biotage microwave vial was added dienone (50 mg, 1.0 equiv.), PhMe (2 mL), and DBU (2.0 equiv.). The resulting solution was sealed, placed in a Biotage microwave reactor, and heated at 160 °C for 2-4 hours. Upon cooling, the reaction mixture filtered through a cotton plug (eluting with EtOAc), concentrated in vacuo, and then purified by column chromatography chromatography (0% EtOAc in hexanes \rightarrow 60% EtOAc in hexanes) to the corresponding pyrroloazepinone.



Compound 25: The standard procedure was followed with dienone **45** (2.0 g, 10.5 mmol, 1.0 equiv.) and DBU (3.1 mL, 21.0 mmol, 2.0 equiv.) to afford compound **25** (1.0 g, 6.2 mmol, 60% yield) as a yellow solid. **TLC**: $R_f = 0.4$ (50% EtOAc in hexanes); ¹H **NMR** (600 MHz, CDCl₃) δ 7.28 (d, J = 10.6 Hz, 1H), 7.09 (d, J = 12.2 Hz, 1H), 6.61 (d, J = 3.7 Hz, 1H), 6.27 (d, J = 3.3 Hz, 1H), 6.12 (dd, J = 12.2, 2.4 Hz, 1H), 5.90 (dd, J = 10.6, 2.4 Hz, 1H), 2.40 (s, 3H); ¹³**C NMR** (151 MHz, CDCl₃) δ 187.6, 134.0, 132.7, 130.1, 130.0, 123.9, 117.8, 114.9, 112.7, 13.1; **IR** (thin film) v_{max} (cm⁻¹): 3108, 3031, 2980, 2923, 1640, 1610, 1495, 1396, 1347, 1261, 1139, 1027, 844, 763, 660; **HRMS** (**m/z**): (ESI) calcd. for C₁₀H₁₀ON [M+H]⁺: 160.0757, found160.0756.

(Observed with decreased reaction times)

TLC: $R_f = 0.7$ (40% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 13.64 (bs, 1H), 7.79 (d, J = 12.3 Hz, 1H), 6.52 – 6.50 (m, 1H), 6.48 (d, J = 12.0 Hz, 1H), 6.02 (t, J = 3.1 Hz, 1H), 5.62 (d, J = 12.0 Hz, 1H), 5.55 (d, J = 12.3 Hz, 1H), 2.96 (s, 3H), 1.95 (s, 3H); ¹³**C NMR** (151 MHz, C₆D₆) δ 188.8, 162.6, 134.6, 133.7, 130.6, 121.0, 114.8, 111.0, 107.3, 57.0, 13.4; **IR** (thin film) v_{max} (cm⁻¹): 3260, 1651, 1584, 1529, 1497, 1435, 1406, 1352, 1309, 1245, 1205, 1170, 1079, 1029, 831; **HRMS (m/z):** (ESI) calcd for C₁₁H₁₄O₂N [M+H]⁺: 192.1019, found 192.1018.



Compound 72: The standard procedure was followed with dienone **71** (50 mg, 0.26 mmol, 1.0 equiv.) and DBU (77.5 μ L, 0.52 mmol, 2.0 equiv.) to afford compound **72** (30 mg, 0.18 mmol, 72%) as a yellow solid. **TLC:** R_f = 0.30 (50% EtOAc in hexanes); ¹H **NMR** (600 MHz, CDCl₃) δ 7.41 (d, *J* = 10.3 Hz, 1H), 7.18 (dd, *J* = 2.9, 1.7 Hz, 1H), 6.78 (dd, *J* = 4.1, 1.6 Hz, 1H), 6.53 (dd, *J* = 3.9, 2.9 Hz, 1H), 6.31 (dd, *J* = 2.5, 1.2 Hz, 1H), 5.92 (dd, *J* = 10.3, 2.4 Hz, 1H), 2.37 (d, *J* = 1.1 Hz, 3H); ¹³C **NMR** (151 MHz, CDCl₃) δ 187.0, 138.7, 134.3, 133.6, 128.1, 127.0, 116.5, 114.9, 112.4, 24.3; **IR** (thin film) v_{max} (cm⁻¹): 3105, 3055, 3015, 2983, 1643, 1593, 1576, 1530, 1471, 1374, 896, 764. **HRMS (m/z)**: (ESI) calcd. for C₁₀H₁₀NO [M+H]⁺ m/z: 160.0757, found 160.0756.



Compound 60: The standard procedure was followed with dienone **65** (50 mg, 0.22 mmol, 1.0 equiv.) and DBU (65.7 μ L, 0.44 mmol, 2.0 equiv.) to afford compound **60** (18.5 mg, 94.7 μ mol, 43%) as an orange solid. **TLC:** R_f = 0.55 (50% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 7.40 – 7.35 (m, 1H), 7.12 – 7.04 (m, 2H), 6.76 (dd, *J* = 9.3, 4.9 Hz, 2H), 6.43 (d, *J* = 12.4 Hz, 1H), 6.30 – 6.24 (m, 2H), 5.73 (dd, *J* = 10.5, 2.3 Hz, 1H); ¹³**C NMR** (151 MHz, Acetone) δ 187.8, 138.3, 137.5, 131.6, 130.5, 129.3, 128.6, 126.3, 124.4, 122.7, 113.5, 112.5, 111.7; **IR** (thin film) v_{max} (cm⁻¹): 1634, 1595, 1523, 1471, 1426, 1409, 1391, 1357, 1315, 878, 850, 781; **HRMS** (m/z): (ESI) calcd. for C₁₃H₁₀NO [M+H]⁺ m/z: 196.0757, found 196.0758.



Compound 46: The standard procedure was followed with dienone **53** (50 mg, 0.20 mmol, 1.0 equiv.) and DBU (58.9 μ L, 0.40 mmol, 2.0 equiv.) to afford compound **46** (23 mg, 126 μ mol, 63%) as a yellow solid. **TLC**: R_f = 0.50 (45% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 7.04 (dd, *J* = 5.2, 2.0 Hz, 3H), 6.95 (dt, *J* = 6.7, 2.2 Hz, 2H), 6.92 (d, *J* = 10.7 Hz, 1H), 6.49 (d, *J* = 12.2 Hz, 1H), 6.27 (dd, *J* = 12.3, 2.3 Hz, 1H), 6.19 (d, *J* = 3.8 Hz, 1H), 6.14 (d, *J* = 3.8 Hz, 1H), 5.67 (dd, *J* = 10.7, 2.3 Hz, 1H); ¹³**C NMR** (151 MHz, C₆D₆) δ 186.8, 138.7, 133.8, 131.3, 130.0,129.8(2C), 129.4, 129.1(2C), 128.8, 126.0, 117.8, 116.0, 113.1; **IR** (thin film) v_{max} (cm⁻¹): 1637,1613, 1595, 1446, 1435, 1411, 1396, 1346, 1322, 877, 849, 758, 702; **HRMS (m/z)**: (ESI) calcd. for C₁₅H₁₂NO [M+H]⁺ m/z: 222.0913, found 222.0916.



Compound 47: The standard procedure was followed with dienone **54** (50 mg, 0.18 mmol, 1.0 equiv.) and DBU (55 μ L, 0.36 mmol, 2.0 equiv.) to afford compound **47** (26 mg, 108 μ mol, 59%) as a yellow oil which slowly solidified. **TLC:** R_f = 0.50 (50% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 6.74 (d, *J* = 10.7 Hz, 1H), 6.66 (d, *J* = 6.9 Hz, 4H), 6.49 (d, *J* = 12.3 Hz, 1H), 6.28 (dd, *J* = 12.3, 2.4 Hz, 1H), 6.17 (d, *J* = 3.8 Hz, 1H), 6.04 (d, *J* = 3.8 Hz, 1H), 5.72 (dd, *J* = 10.7, 2.3 Hz, 1H); ¹³**C NMR** (151 MHz, C₆D₆) δ 186.7, 164.0, 162.4, 137.5, 133.7, 131.7, 131.6, 129.7, 129.3, 126.1, 117.7, 116.2, 116.1, 116.0, 113.1; **IR** (thin film) v_{max} (cm⁻¹): 1646, 1635, 1615, 1507, 1478, 1417, 1271, 1227, 1159, 1099, 840, 814, 770; **HRMS (m/z)**: (ESI) calcd. for C₁₅H₁₁NFO [M+H]⁺ m/z: 240.0819, found 240.0820.



Compound 48: The standard procedure was followed with dienone **55** (50 mg, 0.18 mmol, 1.0 equiv.) and DBU (52.7 μ L, 0.36 mmol, 2.0 equiv.) to afford compound **48** (26 mg, 103 μ mol, 57%) as an orange oil which slowly solidified. **TLC**: R_f = 0.35 (50% EtOAc in hexanes); ¹H **NMR** (600 MHz, C₆D₆) δ 7.00 (d, *J* = 10.7 Hz, 1H), 6.92 – 6.87 (m, 2H), 6.69 – 6.63 (m, 2H), 6.53 (d, *J* = 12.3 Hz, 1H), 6.29 (dd, *J* = 12.3, 2.3 Hz, 1H), 6.23 (d, *J* = 3.7 Hz, 1H), 6.16 (d, *J* = 3.8 Hz, 1H), 5.75 (dd, *J* = 10.7, 2.4 Hz, 1H), 3.26 (s, 3H); ¹³C **NMR** (151 MHz, C₆D₆) δ 186.8,160.5, 138.8, 133.5, 131.2(2C), 130.1, 129.5, 125.7, 123.4, 117.8, 115.8, 114.6(2C), 112.8, 54.9. **IR** (thin film) v_{max} (cm⁻¹): 1645, 1634, 1608, 1478, 1436, 1406, 1397, 1343, 1287, 1250, 1178, 1087, 877; **HRMS (m/z)**: (ESI) calcd. for C₁₆H₁₄NO₂ [M+H]⁺ m/z: 252.1019, found 252.1021.



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Compound 41: The standard procedure was followed with dienone **69** (50 mg, 0.28 mmol, 1.0 equiv.) and DBU (83.3 μ L, 0.56 mmol, 2.0 equiv.) to afford compound **41** (26 mg, 0.18 mmol, 64%) as a yellow solid. **TLC:** R_f = 0.35 (50% EtOAc in hexanes); ¹H **NMR** (600 MHz, CDCl₃) δ 7.38 (dd, J = 10.4, 0.7 Hz, 1H), 7.21 – 7.16 (m, 2H), 6.73 (dd, J = 3.4, 1.4 Hz, 1H), 6.52 (dd, J = 3.8, 2.9 Hz, 1H), 6.24 (dd, J = 12.4, 2.4 Hz, 1H), 5.94 (dd, J = 10.4, 2.4 Hz, 1H); ¹³C **NMR** (151 MHz, C₆D₆) δ 186.5, 132.7, 132.6, 128.8, 126.3, 126.2, 118.2, 115.8, 111.9; **IR** (thin film) v_{max} (cm⁻¹): 3111, 3091, 1640, 1611, 1589, 1527, 1478, 1432, 1382, 1370, 1291, 868, 846, 749; **HRMS (m/z)**: (ESI) calcd. for C₉H₈NO [M+H]⁺ m/z: 146.0600, found 146.0600.



Compound 61: The standard procedure was followed with dienone **62** (50 mg, 0.22 mmol, 1.0 equiv.) and DBU (65.7 μ L, 0.44 mmol, 2.0 equiv.) to afford compound **61** (17.5 mg, 88 μ mol, 40%) as a yellow solid. **TLC:** R_f = 0.30 (50% EtOAc in hexanes); ¹H **NMR** (600 MHz, C₆D₆) δ 6.52 (d, *J* = 12.2 Hz, 1H), 6.34 (dd, *J* = 12.2, 2.4 Hz, 1H), 6.28 (d, *J* = 10.5 Hz, 1H), 5.97 (s, 1H), 5.88 (dd, *J* = 10.5, 2.4 Hz, 1H), 2.24 – 2.19 (m, 2H), 1.73 – 1.68 (m, 2H), 1.35 (p, *J* = 3.0 Hz, 4H); ¹³C **NMR** (151 MHz, C₆D₆) δ 186.7, 132.6, 132.0, 128.8, 128.7, 125.1, 122.4, 117.1, 114.9, 23.0, 22.9, 22.6, 21.8; **IR** (thin film) v_{max} (cm⁻¹): 2933, 2853, 1636, 1615, 1582, 1497, 1441, 1406, 1356, 1300, 850; **HRMS** (**m/z**): (ESI) calcd. for C₁₃H₁₄NO [M+H]⁺ m/z: 200.1070, found 200.1072.



Compound 59: The standard procedure was followed with dienone **58** (50 mg, 0.19 mmol, 1.0 equiv.) and DBU (56.9 μ L, 0.38 mmol, 2.0 equiv.) to afford compound **58** (23 mg, 97.8 μ mol, 52%) as a yellow oil. **TLC:** R_f = 0.38 (50% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 7.09 – 7.01 (m, 3H), 6.87 – 6.83 (m, 2H), 6.82 (d, *J* = 10.7 Hz, 1H), 6.54 (d, *J* = 12.2 Hz, 1H), 6.33 (dd, *J* = 12.2, 2.4 Hz, 1H), 6.07 (s, 1H), 5.67 (dd, *J* = 10.7, 2.4 Hz, 1H), 1.80 (s, 3H); ¹³**C NMR** (151 MHz, C₆D₆) δ 186.7, 135.2, 132.5, 130.9(2C), 130.5, 130.3, 129.1(2C), 128.9, 128.8, 126.1, 121.6, 119.4, 115.3, 11.5; **IR** (thin film) v_{max} (cm⁻¹): 1636, 1616, 1589, 1480, 1433, 1422, 1302, 1193, 880, 763, 703; **HRMS (m/z)**: (ESI) calcd. for C₁₆H₁₄NO [M+H]⁺ m/z: 236.1070, found 236.1072.



Methyl 2-(2-methyl-1*H***-pyrrol-1-yl)propanoate**: A dry 100 mL round bottom flask was charged NaH (984 mg (60% dispersion in mineral oil), 24.6 mmol, 2.0 equiv.) and evacuated and backfilled with nitrogen three times. Hexanes (10.0 mL) was added and the suspension was swirled. Upon settling of the suspension, the hexane was carefully removed via syringe under nitrogen. The flask containing the rinsed NaH was then charged with DMF (20.0 mL) and cooled to 0 °C, wherein 2-methyl pyrrole (1.06 mL, 12.3 mmol, 1.0 equiv.) was added, and the resulting mixture was stirred for 30 min at 0 °C. Methyl 2-bromopropionate (2.7 mL, 24.6 mmol, 2.0 equiv.) was added dropwise, and upon completion of the addition, the mixture was warmed to room temperature. Upon completion of the reaction, as indicated by TLC, the reaction was cooled to 0 °C and

quenched with saturated *aq.* NH₄Cl (100 mL). The mixture was extracted with Et₂O (3 x 100 mL). The combined organic layers were washed with brine (200 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The brown colored crude material was purified by column chromatography (0% EtOAc in hexanes \rightarrow 15% EtOAc in hexanes) to afford methyl 2-(2-methyl-1*H*-pyrrol-1-yl)propanoate (1.64 g, 9.8 mmol, 80%) as a yellow oil. **TLC:** R_f = 0.4 (10% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 6.68 (dd, J = 3.0, 1.8 Hz, 1H), 6.28 (t, J = 3.2 Hz, 1H), 6.03 (m, 1H), 4.34 (q, J = 7.2 Hz, 1H), 3.16 (s, 3H), 2.01 (s, 3H), 1.31 (d, J = 7.2 Hz, 3H); ¹³C NMR (151 MHz, C₆D₆) δ 171.4, 128.4, 117.4, 108.4, 107.9, 53.6, 51.8, 17.9, 12.1; **IR** (thin film) v_{max} (cm⁻¹): 2989, 2950, 1742, 1420, 1296, 1204, 1086, 773, 700; **HRMS** (m/z): (ESI) calcd. for C₉H₁₄O₂N [M+H]⁺: 168.1019, found 168.1013.



Pyrrole Aldehyde 76: To a 500 mL round bottom flask was added PhMe (250 mL) and methyl 2-(2-methyl-1H-pyrrol-1-yl)propanoate (7.0 g, 41.9 mmol, 1.0 equiv.). The mixture was cooled to -78 °C, and DIBAL (1.0 M in hexanes, 46.1 mL, 46.1 mmol, 1.1 equiv.) was added dropwise over 15 min. Upon completion of the reaction as indicated by TLC, the reaction mixture was guenched with saturated ag. Rochelle's salt (50 mL) and warmed to room temperature by removing the vessel from the cooling bath. The resulting biphasic suspension was stirred until the cloudiness dissipated (~2 hours). The mixture was then poured into a separatory funnel and extracted with Et₂O (3 x 300 mL), the combined organics were washed with brine (100 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The resulting crude residue was purified by column chromatography (10% Et₂O in hexanes \rightarrow 50% Et₂O in hexanes) to afford aldehyde **76** (5.47 g, 40.1 mmol, 95%) as a colorless oil. **TLC:** R_f = 0.4 (40% Et₂O in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 9.03 (d, J = 0.7 Hz, 1H), 6.28 (dd, J = 11.3, 5.2 Hz, 2H), 6.04 (m, 1H), 3.77(q, J = 7.2 Hz, 1H), 1.81 (s, 3H), 1.02 (d, J = 7.2, 3H); ¹³**C** NMR (151) MHz, C₆D₆) δ 197.9, 128.0, 117.4, 109.0, 108.6, 59.8, 14.9, 12.0; IR (thin film) v_{max} (cm⁻¹): 2982, 2936, 2826, 1734,1448, 1231, 703; HRMS (m/z): (ESI) calcd. for C₈H₁₂ON [M+H]⁺: 138.0913, found 138.0913.



Alcohol 74: To a 100 mL round bottom flask containing t-BuOAc (450 mg, 3.88 mmol, 1.0 equiv.) was added THF (30 mL). The mixture was cooled to -78 °C before LiHMDS (1.0 M in THF, 4.3 mL, 4.3 mmol, 1.1 equiv.) was added dropwise. The resulting mixture was stirred for 30 min before aldehyde 76 (591 mg, 4.3 mmol, 1.1 equiv.) in THF (5 mL) was added. Upon consumption of the starting material as indicated by TLC, the reaction mixture was guenched with saturated ag. NH₄Cl solution (20 mL) and extracted with EtOAc (3 x 30 mL). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes \rightarrow 20% EtOAc in hexanes) to afford an inseparable mixture of alcohol epimers 74 (809 mg, 3.18 mmol, 82% yield, 6:1 dr) as a yellow oil. **TLC**: $R_f = 0.4$ (50% EtOAc in hexanes); ¹H NMR (600 MHz, C₆D₆) δ 6.83 (dd, J = 3.0, 1.8 Hz, 0.15H), 6.45 (dd, J = 2.9, 1.8 Hz, 0.85H), 6.30 (t, J = 3.1Hz, 0.15H), 6.26 (t, J = 3.2 Hz, 0.85H), 6.03 (ddd, J = 3.6, 1.9,1.0 Hz, 0.15H), 5.99 (ddd, J = 3.7, 1.8, 0.9 Hz, 0.85H), 3.96 (ddd, J = 9.0, 7.7, 3.2 Hz, 0.85H), 3.91 (ddd, J= 8.8, 5.0, 3.7 Hz, 0.15H), 3.82 (p, J = 7.0 Hz, 0.85H), 3.74 - 3.68 (m, 0.15H), 3.31(s, 0.85H), 2.96 (s, 0.15H), 2.13 – 2.01 (m, 2H), 2.00 (s, 0.45H), 1.98 (s, 2.55H), 1.30 (d, J = 6.8 Hz, 2.55H), 1.28 (s, 0.45H), 1.24 (s, 2.55H), 1.15 (d, J = 7.0 Hz, 0.45H); ¹³C NMR (151 MHz, C₆D₆) δ 172.5, 172.1, 127.8(2C), 117.6, 116.4, 108.5, 108.1, 107.3, 107.1, 81.0, 80.8, 72.5, 71.7, 54.9, 54.4, 39.2, 39.0, 28.0(3C), 27.9(3C), 17.9, 17.6, 12.5, 12.4; **IR** (thin film) v_{max} (cm⁻¹): 3454, 2978, 2934, 1723, 1486, 1417, 1368, 1287, 1155, 1080, 842, 768, 704; HRMS (m/z): (ESI) calcd. for C14H24NO3 [M+H]+: 254.1751, found 254.1751.



Alcohol 75: To a 20 mL round bottom flask containing alcohol 74 (90 mg, 0.36 mmol, 1.0 equiv.) was added PhMe (5 mL). The resulting mixture was cooled to -78 °C before DIBAL (1.0 M in hexanes, 1.78 mL, 1.78 mmol, 5.0 equiv.) was added dropwise. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated *aq*. Rochelle's salt (5 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting crude product was dissolved in DCM (5 mL) followed by the addition of SiO₂ (100 mg). Upon consumption of the starting material as indicated by TLC, the reaction mixture was filtered and concentrated *in vacuo*. The resulting crude product was dissolved in DCM (5 mL) followed by the addition of SiO₂ (100 mg). Upon consumption of the starting material as indicated by TLC, the reaction mixture was filtered and concentrated *in vacuo*. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes) \rightarrow 30% EtOAc in hexanes) to afford alcohol **75** (25 mg, 0.16 mmol, 50% yield) as a yellow oil. **TLC**: R_f = 0.4 (50% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 6.33

(d, J = 9.6 Hz, 1H), 6.15 (d, J = 3.4 Hz, 1H), 6.00 (d, J = 3.3 Hz, 1H), 5.42 (dd, J = 9.6, 5.9 Hz, 1H), 4.04 (q, J = 6.9 Hz, 1H), 3.73 (t, J = 6.6 Hz, 1H), 1.97 (s, 3H), 1.22 (d, J = 8.3 Hz, 1H), 0.72 (d, J = 6.9 Hz, 3H); ¹³**C** NMR (151 MHz, C₆D₆) δ 130.4, 126.1, 122.7, 116.0, 108.2, 108.1, 68.4, 54.6, 18.6, 11.2; **IR** (thin film) v_{max} (cm⁻¹): 3358, 2923, 2852, 1658, 1631, 1464, 1421, 1060, 764; **HRMS** (m/z): (ESI) calcd. for C₁₀H₁₄NO [M+H]⁺: 164.1070, found 164.1071.



3,5-dimethyl-indolizine: To a 10 mL reaction tube containing (–)-curvulamine (1) (2.0 mg, 6.2 μ mol, 1.0 equiv.) and alcohol 17 (2.0 mg, 12.4 μ mol, 2.0 equiv.) was added DCM (1 mL). PPTS (1.6 mg, 0.62 μ mol, 0.1 equiv.) was added dropwise. The resulting mixture was warmed to 25°C. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated *aq.* NaHCO₃ (2 mL) and extracted with DCM (3 x 3 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting crude product was purified by preparative TLC (10% EtOAc in hexanes) to afford 3,5-dimethyl-indolizine (0.6 mg, 3.9 μ mol, 63% yield) as a yellow oil. **TLC**: R_f = 0.7 (10% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 7.13 (d, *J* = 8.9 Hz, 1H), 6.49 (d, *J* = 3.8 Hz, 1H), 6.46 (d, *J* = 3.8 Hz, 1H), 6.30 (dd, *J* = 8.9, 6.4 Hz, 1H), 5.81 (dt, *J* = 6.5, 1.2 Hz, 1H), 2.41 (s, 3H), 2.19 (s, 3H); ¹³C NMR (151 MHz, C₆D₆) δ 135.6, 134.6, 122.0, 118.4, 115.8, 115.8, 111.4, 99.3,1.1, 16.8; **IR** (thin film) v_{max} (cm⁻¹): 2961, 2922, 1588, 1538, 1456, 1290, 1154, 750; **HRMS** (m/z): (ESI) calcd. for C₁₀H₁₂N [M+H]⁺: 146.0964, found 146.0964.



Procuramine 72: To a 20 mL reaction tube containing alcohol **124** (80 mg, 0.32 mmol, 1.0 equiv.) was added DCM (4 mL). The resulting mixture was cooled to 0 °C and TMSOTf (64 μ L, 0.35 mmol, 1.1 equiv.) was added dropwise. The resulting mixture was warmed to 25°C. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated *aq*. NaHCO₃ (5 mL) and extracted with DCM (3 x 5 mL). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes \rightarrow 80% EtOAc in hexanes) to afford procuramine **72** (28 mg, 0.16 mmol, 50% yield) as a yellow oil. **TLC**: R_f = 0.1 (50% EtOAc in hexanes); ¹H **NMR** (600 MHz, CDCl₃) δ 7.01 (d, *J* = 4.0 Hz, 1H), 6.08 (d, *J* = 4.0 Hz, 1H), 4.40 (qd, *J* = 6.8, 1.9 Hz, 1H), 4.35 (s, 1H), 2.87 (dd, *J* = 18.0, 3.2 Hz, 1H), 2.79 (s, 1H), 2.64 (dd,

J = 18.0, 2.7 Hz, 1H), 2.32 (s, 3H), 1.36 (d, J = 6.9 Hz, 3H); ¹³**C** NMR (151 MHz, CDCl₃) δ 183.7, 135.8, 128.3, 115.1, 111.2, 70.5, 53.6, 39.4, 19.3, 12.0; **IR** (thin film) v_{max} (cm⁻¹): 3452, 2923, 1739, 1626, 1492, 1462, 1347, 1256, 1177, 1033, 776, 642; **HRMS** (m/z): (ESI) calcd. for C₁₀H₁₄NO₂ [M+H]⁺: 180.1019, found 180.1018.



Enone 73: To a 10 mL reaction tube containing ketone **72** (10.0 mg, 0.056 mmol, 1.0 equiv.) was added DCM (30 mL). The resulting mixture was cooled to 0 °C, Et₃N (23.0 μ L, 0.17 mmol, 3.0 equiv.) and MsCl (5.1 μ L, 0.067 mmol, 1.2 equiv.) were added dropwise sequentially. The resulting mixture was warmed to room temperature. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated *aq*. NaHCO₃ (5 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting crude product was purified by preparative TLC (50% EtOAc in hexanes) to afford enone **73** (3.9 mg, 0.024 mmol, 43% yield) as a yellow oil. **TLC**: R_f = 0.4 (50% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 7.33 (d, *J* = 3.9 Hz, 1H), 6.19 (dd, *J* = 10.3, 1.6 Hz, 1H), 5.94 (dd, *J* = 3.9, 0.8 Hz, 1H), 5.80 (dd, *J* = 10.4, 4.0 Hz, 1H), 3.67 (tdd, *J* = 6.7, 5.4, 3.4 Hz, 1H), 1.70 (s, 3H), 0.64 (d, *J* = 6.8 Hz, 3H); ¹³**C NMR** (151 MHz, C₆D₆) δ 175.3, 143.1, 133.3, 129.7, 127.7, 112.9, 110.8, 49.3, 21.7, 12.0; **IR** (thin film) v_{max} (cm⁻¹): 2950, 1733, 1685, 1462, 1342, 1033, 766, 651; **HRMS** (m/z): (ESI) calcd. for C₁₀H₁₂NO [M+H]⁺: 162.0913, found 162.0913.



Iodide 77: To a 100 mL round bottom flask containing enone **24** (1.10 g, 2.80 mmol, 1.0 equiv.) was added acetone (30 mL). The resulting mixture was cooled to 0 °C and a solution of NIS (0.63 g, 29.4 mmol, 1.05 equiv.) in acetone (10 mL) was added dropwise. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated *aq*. NH₄Cl solution (30 mL) and extracted with EtOAc (3 x 40 mL). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes \rightarrow 10% EtOAc in hexanes) to afford iodide **77** (1.28 g, 2.46 mmol, 88% yield) as a yellow oil. **TLC**: R_f = 0.6 (20% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 6.28 (dd, *J* = 12.1, 0.8 Hz, 1H), 6.16 (d, *J* = 3.7 Hz, 1H), 6.04 (s, 1H), 5.82 (dd, *J* = 3.8, 0.9 Hz, 1H), 5.69 (d, *J* = 12.1 Hz,

1H), 4.50 (q, J = 6.8 Hz, 1H), 4.40 (d, J = 3.6 Hz, 1H), 3.32 (dd, J =, 1.8 Hz, 1H), 1.94 (s, 3H), 1.89 (s, 3H), 1.07 (d, J = 6.8 Hz, 3H), 0.03 (s, 9H); ¹³**C** NMR(151 MHz, C₆D₆) δ 193.6, 137.9, 132.4, 131.0, 128.6, 128.5, 120.6, 120.4, 120.1, 114.2, 110.9, 73.0, 64.7, 61.6, 60.9, 47.5, 18.4, 13.8, 13.2, 0.8(3C); **IR** (thin film) v_{max} (cm⁻¹): 2954, 2921, 1654, 1620, 1483, 1422, 1255, 1161, 1068, 851, 765; **HRMS** (m/z): (ESI) calcd. for C₂₂H₂₇IN₃O₂Si [M+H]⁺: 520.0912, found 520.0920.



lodide 79: To a 20 mL reaction tube containing ethyl vinyl ether (0.20 mL, 2.1 mmol, 5.5 equiv.) was added THF (2 mL). The mixture was cooled to -78 °C and t-BuLi (1.6 M in pentane, 1.2 mL, 2.0 mmol, 5.0 equiv.) was added dropwise. Upon completion of the addition, the mixture was warmed to 0 °C and stirred for 30 min followed by the addition of CeCl₃ (0.49 g, 2.0 mmol, 5.0 equiv). The mixture was stirred for 1 h at 25 °C. To a 20 mL reaction tube containing iodide 77 (200 mg, 0.39 mmol, 1.0 equiv.) was added THF (5 mL). The mixture was -78 °C and a solution of fresh cerium reagent was added dropwise. The mixture was stirred for 1 h before the addition of TMSCI (0.25 mL, 2.0 mmol, 5.0 equiv.). The resulting mixture was stirred for 30 min before it was guenched with saturated ag. NaHCO₃ (5 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes \rightarrow 10% EtOAc in hexanes) to afford iodide **79** (171 mg, 0.30 mmol, 79% yield) as a yellow oil. **TLC**: R_f = 0.7 (10% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 6.21 (dd, J = 11.5, 0.7 Hz, 1H), 6.09 (d, J = 3.5 Hz, 1H), 6.01 (s, 1H), 5.90 (d, J = 3.5 Hz, 1H), 5.59 (d, J = 11.4 Hz, 1H), 4.56 (d, J = 1.5 Hz, 1H), 4.24 (s, 1H), 4.00 (q, J = 6.3 Hz, 1H), 3.81 (s, 1H), 3.78 (d, J = 1.4 Hz, 1H), 3.36 - 3.30 (m, 1H), 3.23 (ddt, J = 9.0, 7.6, 6.6 Hz, 1H), 2.03 (s, 3H), 1.88 (s, 3H), 1.48 (d, J = 6.3 Hz, 3H), 1.04 (t, J = 7.0 Hz, 3H), 0.10 (s, 9H); ¹³C NMR (151 MHz, C₆D₆) δ 164.2, 135.2, 132.3, 130.8, 128.6, 124.3, 123.9, 114.8, 112.1, 111.4, 108.6, 86.7, 80.8, 65.0, 63.6, 63.1, 62.7, 46.1, 14.6(2C), 14.5, 13.1, 1.7(3C); **IR** (thin film) v_{max} (cm⁻¹): 3269, 2977, 1640, 1491, 1414, 1311, 1250, 1079, 908, 844, 764; HRMS (m/z): (ESI) calcd. for C₂₅H₃₄IN₂O₃Si [M+H]⁺: 565.1378, found 565.1377.



Ester: To a 100 mL round bottom flask containing t-BuOAc (1.00 g, 8.62 mmol, 1.0 equiv.) was added THF (30 mL). The resulting mixture was cooled to -78 °C before LiHMDS (1.0 M in THF, 9.5 mL, 9.48 mmol, 1.1 equiv.) was added dropwise. The resulting mixture was stirred for 30 min before ester SI-11 (1.58 g, 9.48 mmol, 1.0 equiv.) in THF (10 mL) was added. The mixture was warmed to 25 °C. Upon consumption of the starting material as indicated by TLC, the reaction mixture was guenched with saturated aq. NH₄Cl solution (20 mL) and extracted with EtOAc (3 x 30 mL). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes \rightarrow 20% EtOAc in hexanes) to afford ester (1.95 g, 7.76 mmol, 90% yield) as a yellow oil. **TLC**: $R_f = 0.6$ (50% EtOAc in hexanes); ¹H NMR (600 MHz, C_6D_6) δ 12.95 (s, 0.24H), 6.57 (dd, J = 3.0, 1.8 Hz, 0.24H), 6.28 (dd, J = 3.0, 1.8 Hz, 0.76H), 6.24 (t, J = 3.2 Hz, 0.24H), 6.20 (t, J = 3.2 Hz, 0.76H), 6.00 – 5.98 (m, 0.24H), 5.97 (ddd, J =3.5, 1.8, 0.9 Hz, 0.76H), 4.58 (d, J = 0.9 Hz, 0.24H), 4.41 – 4.34 (m,0.24H), 4.30 (q, J = 7.1 Hz, 0.76H), 2.89 (d, J = 16.2 Hz, 0.76H), 2.84 (d, J = 16.2 Hz, 0.76H), 1.90 (s, 0.72H), 1.89 (d, J = 0.8 Hz, 2.28H), 1.35 (d, J = 7.2 Hz, 0.72H), 1.30 (s, 6.84H), 1.25 (s, 16H), 1.22 (d, J = 7.1 Hz, 2.28H); ¹³**C** NMR (151 MHz, C₆D₆) δ 201.2, 178.4, 173.6, 166.4,128.6, 128.3, 117.6, 117.0, 109.3, 108.8, 108.3, 108.0, 89.3, 81.4, 81.3, 59.9, 53.6, 45.8, 28.1(3C), 27.9(3C), 17.9, 16.2, 11.9(2C); **IR** (thin film) v_{max} (cm⁻¹): 2980. 2935, 1742, 1719,1649, 1325, 1294, 1247, 1153, 1054, 814, 703; HRMS (m/z): (ESI) calcd. for C₁₄H₂₁NNaO₃ [M+Na]⁺: 274.1414, found 274.1414.



Triflate 80: To a 50 mL round bottom flask containing ester (1.00 g, 3.98 mmol, 1.0 equiv.) was added DCM (20 mL). The resulting mixture was cooled to -78 °C, then Et₃N (1.6 mL, 11.94 mmol, 3.0 equiv.) and Tf₂O (0.74 mL, 4.38 mmol, 1.1 equiv.) was added dropwise. The resulting mixture was warmed to 0 °C. Upon consumption of the starting material as indicated by TLC, the). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered reaction mixture was guenched with saturated aq. NaHCO₃ (20 mL) and extracted with DCM (3 x 30 mL and concentrated in vacuo. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes \rightarrow 10% EtOAc in hexanes) to afford triflate **80** (1.34 g, 3.51 mmol, 88%) yield) as a yellow oil. **TLC**: R_f = 0.5 (10% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 6.30 (dd, J = 3.0, 1.7 Hz, 1H), 6.18 (t, J = 3.2 Hz, 1H), 5.97 – 5.92 (m,1H), 5.01 (d, J = 1.5 Hz, 1H), 4.52 (qd, J = 7.1, 1.5 Hz, 1H), 1.92 (s, 3H), 1.25 (s, 9H), 1.10 (d, J = 7.1 Hz, 3H); ¹³C NMR (151 MHz, C₆D₆) δ 161.6, 157.7, 128.9, 128.5, 116.1, 113.5, 109.3,109.0, 82.6, 52.8, 27.7(3C), 17.1, 11.4; **IR** (thin film) v_{max} (cm⁻¹): 2982, 2936, 1725, 1673, 1431, 1297, 1209, 1153, 1020, 915, 792, 700, 595; HRMS (m/z): (ESI) calcd. for C₁₅H₂₁F₃NO₅S [M+H]⁺: 384.1087, found 384.1092.



Negishi coupling of 79 & (±)-80: To a 20 mL reaction tube containing iodide **79** (130 mg, 0.23 mmol, 1.0 equiv.) was added THF (5 mL) at -78 °C. *t*-BuLi (1.6 M in pentane, 0.36 mL, 0.58 mmol, 2.5 equiv) was added and the resulting mixture was stirred for 30 min. Then a solution of ZnCl₂ (94 mg, 0.69 mmol, 3.0 equiv.) in THF (1 mL) was added dropwise. The resulting mixture was stirred for 1 h at -78 °C. Then a solution of triflate (±)-**80** (176 mg, 0.46 mmol, 2.0 equiv.) and Pd(PPh₃)₄ (8.0 mg, 0.0069 mmol, 3 mol%.) in THF (3 mL) was added dropwise at -78 °C. The resulting was warmed to 25 °C and stirred for 6 h. Upon completion of the reaction as indicated by TLC, the reaction mixture was quenched with saturated *aq*. NH₄Cl solution (5 mL) and extracted with Et₂O (3 x 10 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes \rightarrow 10% EtOAc in hexanes) to afford **81** (61 mg, 0.092 mmol, 40% yield), **82** (64 mg, 0.092 mmol, 40% yield) and **110** (10 mg, 0.023 mmol, 10% yield) as yellow oils.



TLC: $R_f = 0.4$ (10% EtOAc in hexanes); ¹**H NMR** (600 MHz, C_6D_6) δ 6.71 (dd, J = 3.0, 1.8 Hz, 1H), 6.34 (t, J = 3.1 Hz, 1H), 6.27 (d, J = 11.5 Hz, 1H), 6.15 – 6.13 (m, 1H), 6.13 (d, J = 3.7 Hz,1H), 5.93 (d, J = 3.4 Hz, 1H), 5.72 (s, 1H), 5.70 (d, J = 11.5 Hz, 1H), 5.46 (d, J = 1.1 Hz, 1H), 4.68 (q, J = 6.9 Hz, 1H), 4.61 (d, J = 1.9 Hz, 1H), 4.39 (s, 1H), 4.13 (q, J = 6.3 Hz, 1H), 3.96 (s, 1H), 3.90 (d, J = 1.9 Hz, 1H), 3.48 (dq, J = 9.4, 7.0 Hz, 1H), 3.40 (dq, J = 9.4, 7.1 Hz, 1H), 2.14 (s, 3H), 2.06 (s, 3H), 1.89 (s, 3H), 1.61 (d, J = 6.3 Hz, 3H), 1.30 (d, J = 7.1 Hz, 3H), 1.25 (s, 9H), 1.13 (t, J = 7.0 Hz, 3H), 0.13 (s, 9H); ¹³**C NMR** (151 MHz, C_6D_6) δ 165.9, 164.2, 152.2, 135.2, 130.9, 129.9, 128.5, 126.0, 124.4, 124.1, 118.7, 118.5, 117.1, 114.8, 111.5, 108.6, 108.2, 107.6, 104.5, 86.9, 81.1, 78.9, 65.3, 63.1, 61.9, 57.6, 46.2, 28.2(3C), 19.4, 14.7(2C), 13.0(2C), 12.4, 1.7(3C); **IR** (thin film) v_{max} (cm⁻¹): 2977, 2929, 1718, 1699, 1640, 1419, 1307, 1253, 1164, 1143,1080, 910, 845, 764, 698; **HRMS** (m/z): (ESI) calcd. for $C_{39}H_{53}N_3NaO_5Si$ [M+Na]⁺: 694.3647,found 694.3643.



TLC: $R_f = 0.3 (10\% EtOAc in hexanes); {}^{1}H NMR (600 MHz, C_6D_6) \delta 6.48 (dd, <math>J = 2.9, 1.8$ Hz, 1H), 6.30 – 6.23 (m, 2H), 6.13 (d, J = 3.6 Hz, 1H), 6.09 – 6.07 (m, 1H), 5.96 – 5.94 (m, 1H), 5.72 – 5.67 (m, 2H), 5.62 (s, 1H), 4.62 (d, J = 1.8 Hz, 1H), 4.56 (q, J = 7.1 Hz, 1H), 4.39 (s, 1H), 4.05 (q, J = 6.3 Hz, 1H), 3.96 (s, 1H), 3.86 (d, J = 1.8 Hz, 1H), 3.41 (dq, J = 9.6, 7.0 Hz, 1H), 3.31 (dq, J = 9.5, 7.0 Hz, 1H), 2.10 (s, 3H), 1.90 (s, 3H), 1.81 (s, 3H), 1.53 (d, J = 6.3 Hz, 3H), 1.32 (d, J = 7.1 Hz, 3H), 1.20 (s, 9H), 1.11 (t, J = 7.0 Hz, 3H), 0.15 (s, 9H); ${}^{13}C$ NMR (151 MHz, C_6D_6) \delta 165.6, 164.5, 152.6, 135.2, 130.8, 129.4, 128.5, 125.9, 124.3, 124.0, 120.2, 117.4, 116.8, 114.7, 111.6, 108.5, 107.8, 107.1, 104.8, 86.9, 80.6, 79.0, 65.2, 63.0, 61.7, 58.4, 46.0, 28.1(3C), 19.2, 14.8, 14.6, 13.0, 12.4, 11.8, 1.7(3C); IR (thin film) v_{max} (cm⁻¹): 2977, 2926, 1699, 1641, 1419, 1307, 1253, 1163, 1081, 910, 845, 763, 698; HRMS (m/z): (ESI) calcd. for C₃₉H₅₃N₃NaO₅Si [M+Na]⁺: 694.3647, found 694.3640.



TLC: $R_f = 0.6$ (10% EtOAc in hexanes); ¹**H NMR** (600 MHz, C_6D_6) δ 6.26 (d, J = 11.5 Hz, 1H), 6.11 (d, J = 3.6 Hz, 1H), 5.98 (dd, J = 3.3, 1.0 Hz, 1H), 5.90 (dd, J = 3.6, 0.9 Hz, 1H), 5.86 (d, J = 3.3 Hz, 1H), 5.68 (d, J = 11.5 Hz, 1H), 4.64 (d, J = 1.7 Hz, 1H), 4.35 (s, 1H), 4.12 (q, J = 6.3 Hz, 1H), 4.00 (s, 1H), 3.84 (d, J = 1.7 Hz, 1H), 3.39 (dq, J = 9.3, 7.0 Hz, 1H), 3.29 (dq, J = 9.3, 7.0 Hz, 1H), 2.07 (s, 3H), 1.82 (s, 3H), 1.61 (d, J = 6.3 Hz, 3H), 1.09 (t, J = 7.0 Hz, 3H), 0.14 (s, 9H); ¹³**C** NMR (151 MHz, C_6D_6) δ 164.7, 135.3, 130.9, 130.4, 127.1, 124.3, 124.2, 114.6, 111.7, 108.4, 108.4, 104.7, 86.9, 80.6, 65.5, 63.0, 61.7, 46.4, 14.8, 14.6, 14.2, 13.0, 1.8(3C); IR (thin film) v_{max} (cm⁻¹): 2977, 2958, 1641, 1416, 1309, 1251, 1080, 909, 845, 762; HRMS (m/z): (ESI) calcd. for $C_{25}H_{35}N_2O_3Si$ [M+H]⁺: 439.2411, found 439.2411.



Lactol 83: Ester 82 (100 mg, 0.15 mmol, 1.0 equiv.) was dissolved in THF (9 mL) and MeOH (1 mL) at 0 °C, then the solution was degassed by sparging with argon for 5 min. Sml₂ (0.1 M in THF, 3.0 mL, 0.30 mmol, 2.0 equiv) was added and the resulting mixture was stirred for 30 min. The mixture was warmed to 25 °C and then TBAF (1.0 M in THF, 0.3 mL, 0.30 mmol, 2.0 equiv.) was added. Upon completion of the reaction as indicated by TLC, the reaction mixture was guenched with saturated aq. NH₄Cl solution (5 mL) and extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting crude product was purified by column chromatography (0% EtOAc in hexanes \rightarrow 10% EtOAc in hexanes) to afford lactol 83 (71.6 mg, 0.12 mmol, 80% yield) as a brown foam. **TLC**: $R_f = 0.5$ (20% EtOAc in hexanes); ¹H NMR (600 MHz, C₆D₆) δ 6.40 (dd, J = 2.9, 1.8 Hz, 1H), 6.29 (t, J = 3.2 Hz, 1H), 6.27 (d, J = 11.6 Hz, 1H), 6.14 (d, J = 3.6 Hz, 1H), 6.08 (ddd, J = 3.6, 1.8, 0.9 Hz, 1H), 5.92 (dd, J = 3.6, 0.9 Hz, 1H), 5.77 (d, J = 11.5 Hz, 1H), 5.65 (s, 1H), 4.66 (d, J = 1.9 Hz, 1H), 4.31 (d, J = 0.9 Hz, 1H), 4.18 – 4.12 (m, 1H), 4.10 (q, J = 6.3 Hz, 1H), 3.83 (s, 1H), 3.80 (d, J = 1.9 Hz, 1H), 3.43 – 3.35 (m, 2H), 3.29 (dq, J = 9.4, 7.0 Hz, 1H), 2.67 (s, 1H), 2.47 (dd, J = 14.9, 9.2 Hz, 1H), 2.40 (dd, J = 14.9)14.9, 6.3 Hz, 1H), 2.24 (s, 2H), 1.81 (s, 3H), 1.80 (s, 3H), 1.61 (d, J = 6.2 Hz, 3H), 1.21 (d, J = 6.6 Hz, 3H), 1.20 (s, 9H), 1.12 (t, J = 7.0 Hz, 3H); ¹³**C** NMR (151 MHz, C₆D₆) δ 171.6, 164.4, 134.9, 130.3, 129.1, 127.5, 125.9, 125.7, 122.9, 118.5, 116.8, 114.5, 109.4, 109.2, 107.5, 107.0, 103.5, 85.9, 80.7, 79.5, 64.6, 63.0, 59.5, 55.2, 45.6, 40.6, 39.5, 28.0(3C), 17.9, 15.2, 14.6, 12.9, 12.5, 11.0; **IR** (thin film) v_{max} (cm⁻¹): 2977, 2924, 2361, 1726, 1643, 1424, 1300, 1144, 1081, 951, 767, 702; HRMS (m/z): (ESI) calcd. for C₃₆H₄₇N₃NaO₅ [M+Na]⁺: 624.3408, found 624.3408.



Epimerization of 106: In a nitrogen-filled glovebox, a reaction tube containing lactol **83** (730 mg, 1.21 mmol, 1.0 equiv.) was charged with NaOMe (328 mg, 6.07 mmol, 5.0 equiv.). The reaction tube was sealed, removed from the glovebox, and placed under an atmosphere of nitrogen. Anhydrous MeOH (30 mL) was then added, the N₂ balloon was removed, and the sealed reaction vessel was heated at 90 °C for 1 h. The reaction mixture was then cooled to room temperature and quenched with saturated *aq*. NH₄Cl solution (30 mL). The solution was extracted with EtOAc (3 x 40 mL) and the combined organic layers were washed with brine (40 mL), dried over Na₂SO₄, filtered and concentrated *in vacuo*. The resulting crude residue was purified by column chromatography (0% EtOAc in hexanes \rightarrow 10% EtOAc in hexanes) to afford lactol epimers **106** (416 mg, 0.69 mmol, 57%, 80% BRSM) and **83** (211 mg, 0.35 mmol, 29%) as brown foams. **TLC**: R_f = 0.4 (20% EtOAc in hexanes); ¹**H NMR** (600 MHz, C₆D₆) δ 6.65 (t, *J* = 2.2 Hz, 1H), 6.37 (t, *J* = 3.1 Hz, 1H), 6.28 (d, *J* = 11.5 Hz, 1H), 6.16 (d, *J*

= 3.6 Hz, 1H), 6.10 (dd, J = 3.4, 1.7 Hz, 1H), 5.94 (d, J = 3.5 Hz, 1H), 5.78 (d, J = 11.5 Hz, 1H), 5.68 (s, 1H), 4.57 (s, 1H), 4.45 (d, J = 1.7 Hz, 1H), 4.16 (q, J = 6.4 Hz, 1H), 3.99 (dq, J = 9.1, 6.8 Hz, 1H), 3.85 (s, 1H), 3.58 (d, J = 1.8 Hz, 1H), 3.42 – 3.34 (m, 2H), 3.29 (dq, J = 9.3, 7.1 Hz, 1H), 2.68 (s, 1H), 2.37 (dd, J = 14.8, 10.6 Hz, 1H), 2.26 (dd, J = 14.8, 4.8 Hz, 1H), 2.18 (s, 3H), 1.93 (s, 3H), 1.88 (s, 3H), 1.30 (d, J = 6.5 Hz, 3H), 1.20 (d, J = 6.7 Hz, 3H), 1.18 (s, 9H), 1.12 (t, J = 7.0 Hz, 3H); ¹³**C** NMR (151 MHz, C₆D₆) δ 171.6, 163.9, 135.0, 130.6, 128.6, 127.6, 125.1, 125.0, 122.8, 120.2, 116.2, 114.6, 110.3, 109.1, 108.4, 107.0, 103.0, 87.1, 79.4, 79.1, 62.9, 60.5, 60.3, 56.0, 46.4, 41.6, 39.7, 28.0(3C), 19.8, 19.3, 14.7, 13.3, 12.5, 10.2; **IR** (thin film) v_{max} (cm⁻¹): 2973, 2925, 2853, 2363, 2336, 1727, 1489, 1283, 1192, 1162, 1027, 767; **HRMS** (m/z): (ESI) calcd. for C₃₆H₄₇N₃NaO₅ [M+Na]⁺: 624.3408, found 624.3412.



Thiocarbonate 107: Lactol 106 (500 mg, 0.83 mmol, 1.0 equiv.) was azeotropically dried with benzene (3x) and then dissolved in THF (50 mL) under an inert atmosphere. The solution was cooled to -78 °C and KHMDS (1.0 M in THF, 1.0 mL, 0.99 mmol, 1.2 equiv.) was added dropwise down the side of the reaction vessel. The reaction was stirred for 30 min and added a solution of DMAP (195 mg, 1.6 mmol, 2.0 equiv.) in THF (5 mL) and followed by O-phenyl chlorothionoformate (75 μ L, 1.6 mmol, 2.0 equiv.). The mixture was stirred for 1 h at -78 °C and then guenched by the addition of saturated aq. NaHCO₃ (10 mL). The solution was extracted with EtOAc (3 x 30 mL) and the combined organics were washed with brine (20 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. The resulting crude residue was purified by column chromatography (0% EtOAc in hexanes \rightarrow 10% EtOAc in hexanes) to afford thiocarbonate 107 (509 mg, 0.69 mmol, 83%), as a yellow oil. TLC: Rf = 0.5 (20% EtOAc in hexanes); ¹H NMR (600 MHz, C_6D_6) δ 6.92 – 6.85 (m, 2H), 6.82 – 6.77 (m, 1H), 6.68 (dd, J = 2.9, 1.8 Hz, 1H), 6.64 – 6.60 (m, 2H), 6.37 (t, J = 3.1 Hz, 1H), 6.32 (d, J = 11.6 Hz, 1H), 6.19 (d, J = 3.6 Hz, 1H), 6.14 - 6.07 (m, 1H), 6.00 - 5.94 (m, 2H),5.72 (t, J = 5.7 Hz, 2H), 4.95 (d, J = 1.0 Hz, 1H), 4.69 (d, J = 2.0 Hz, 1H), 4.00 (dq, J = 9.8, 6.7 Hz, 1H), 3.94 (s, 1H), 3.59 (d, J = 2.0 Hz, 1H), 3.43 (td, J = 10.2, 4.6 Hz, 1H), 3.34 (dq, J = 9.4, 7.0 Hz, 1H), 3.28 (dq, J = 9.3, 7.1 Hz, 1H), 2.40 (dd, J = 14.9, 10.5 Hz, 1H), 2.29 (dd, J = 14.9, 4.7 Hz, 1H), 2.19 (s, 3H), 2.02 (s, 3H), 2.00 (s, 3H), 1.47 (d, J = 6.5 Hz, 3H), 1.21 (d, J = 6.9 Hz, 3H), 1.20 (s, 9H), 1.12 (t, J = 7.0 Hz, 3H); ¹³C NMR (151 MHz, C₆D₆) δ 190.3, 171.6, 163.2, 153.6, 135.3, 130.5,129.6, 128.4, 127.6, 126.5, 125.4, 124.3, 122.6, 122.2, 120.7, 116.1, 115.6, 115.3, 109.1, 108.5, 107.1, 103.2, 91.1, 80.2, 79.2, 63.0, 61.7, 57.6, 56.0, 45.4, 41.5, 39.7, 28.0(3C), 19.9, 18.7, 14.7, 13.6, 12.5, 10.4; **IR** (thin film) v_{max} (cm⁻¹): 2977, 2928, 2361, 1726, 1489, 1277,1197, 1161, 1077, 1003, 768; HRMS (m/z): (ESI) calcd. for 1300. C₄₃H₅₁N₃NaO₆S [M+Na]⁺: 760.3391, found 760.3392.



Ketone 108: Thiocarbonate 107 (70 mg, 0.095 mmol, 1.0 equiv.) was dissolved in THF (5 mL) and the solution degassed by sparging with argon for 5 min at 45 °C. Tributyltin hydride (51 μ L, 0.19 mmol, 2.0 equiv.) and BEt₃ (1.0 M in hexanes, 95 μ L, 0.095 mmol, 1.0 equiv.) were then added to the mixture. A syringe containing 3 mL of air was placed into the reaction solution, and air was bubbled into the mixture at a rate of approximately 1mL/hr. Upon completion of the reaction as indicated by TLC, the mixture was cooled to 0 °C and 1M ag. HCI (1 mL) was added dropwise. The resulting solution was vigorously stirred for 30 min, and was guenched with saturated aq. NaHCO₃ (5 mL). The solution was extracted with EtOAc (3 x 10 mL) and the combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. The resulting crude residue was purified by preparative TLC (10% Et₂O in DCM) to afford ketone **108** (21.7 mg, 0.039 mmol, 41%) as a white solid. **TLC**: $R_f = 0.6$ (10% Et₂O in DCM); ¹**H** NMR (600 MHz, C₆D₆) δ 6.57 (dd, J = 3.0, 1.8 Hz, 1H), 6.31 (t, J = 3.1 Hz, 1H), 6.28 (d, J = 11.5 Hz, 1H), 6.17 (d, J = 11.5 HzJ = 3.6 Hz, 1H), 6.09 - 6.05 (m, 1H), 5.95 (dd, J = 3.7, 0.9 Hz, 1H), 5.63 (d, J = 11.4Hz, 1H), 5.61 (s, 1H), 4.48 (s, 1H), 4.16 – 4.13 (m, 1H), 4.11 (q, J = 7.2 Hz, 1H), 3.75 (s, 1H), 3.72 (qd, J = 6.7, 2.2 Hz, 1H), 3.44 – 3.36 (m, 1H), 2.35 – 2.25 (m, 2H), 2.19 (s, 3H), 1.75 (s, 3H), 1.74 (s, 3H), 1.70 (s, 3H), 1.25 (d, J = 6.8 Hz, 3H), 1.17 (s, 9H), 0.78 (d, J = 6.6 Hz, 3H); ¹³C NMR (151 MHz, C₆D₆) δ 209.3, 171.5, 132.8, 130.6, 127.6, 127.3, 125.7, 124.2, 121.4, 120.1, 116.6, 114.7, 109.1, 107.8, 107.2, 104.8, 94.4, 90.3, 79.2, 60.1, 56.7, 55.0, 46.0, 41.1, 40.0, 28.0(3C), 26.8, 19.3, 18.3, 13.4, 12.5, 9.6; IR (thin film) v_{max} (cm⁻¹): 3481, 2925, 2854, 2364, 1725, 1644, 1449, 1391, 1282, 1165, 1067, 770, 750,702; **HRMS** (m/z): (ESI) calcd. for C₃₄H₄₃N₃NaO₄ [M+Na]⁺: 580.3146, found 580.3140.



CBS reduction of 108: In a N₂ filled glovebox, a reaction tube was charged with (*R*)-(+)-2-methyl-CBS-oxazaborolidine (2.0 mg, 9.0 μ mol, 1.0 equiv.). The reaction tube was sealed and brought out of the glovebox under inert atmosphere. DCM (0.2 mL) was added followed by BH₃•DMS (1.1 μ L, 18.0 μ mol, 2.0 equiv.) and the mixture stirred for 15 min. Methyl ketone **108** (5.0 mg, 9.0 μ mol, 1.0 equiv.) was dissolved in DCM (0.2 mL) and added dropwise to the reaction mixture. Additional DCM (0.1 mL) was used to render the transfer quantitative. Upon completion of the reaction as indicated by TLC, the reaction mixture was quenched with saturated *aq*. NH₄Cl solution (1 mL) and the mixture stirred for 5 min. The solution was extracted with DCM (3 x 3 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting crude residue was purified by preparative TLC (10% Et₂O in DCM) to afford (–)-**109** (2.2 mg, 3.8 μ mol, 42%) and (+)-**84** (2.1 mg, 3.8 μ mol, 42%) both as white solids.

D



TLC: $R_f = 0.7$ (10% Et₂O in DCM); ¹**H** NMR (600 MHz, C_6D_6) δ 6.54 (t, J = 2.3 Hz, 1H), 6.48 (d, J = 11.7 Hz, 1H), 6.30 (t, J = 3.2 Hz, 1H), 6.23 (d, J = 3.3 Hz, 1H), 6.08 (s, 1H), 5.99 (d, J = 3.4 Hz, 1H), 5.92 (d, J = 11.7 Hz, 1H), 5.47 (s, 1H), 4.54 (s, 1H), 4.17 (p, J = 6.9 Hz, 1H), 3.99 (d, J = 1.4 Hz, 1H), 3.70 (s, 1H), 3.63 – 3.56 (m, 1H), 3.46 (q, J = 7.5 Hz, 1H), 2.86 (d, J = 6.8 Hz, 1H), 2.44 – 2.35 (m, 2H), 2.21 (s, 3H), 1.78 (s, 3H), 1.71 (s, 3H), 1.44 (d, J = 6.5 Hz, 3H), 1.25 (d, J = 6.8 Hz, 3H), 1.22 (s, 9H), 0.74 (d, J = 6.6 Hz, 3H); ¹³C NMR (151 MHz, C_6D_6) δ 171.5, 132.2, 131.2, 129.1, 128.0, 126.2, 124.3, 120.8, 119.1, 116.7, 113.7, 108.9, 107.5, 107.1, 103.3, 89.8, 88.8, 79.5, 70.7, 60.6, 57.3, 54.7, 45.3, 40.5, 39.4, 28.0(3C), 19.4, 17.8, 17.7, 13.5, 12.5, 9.6; IR (thin film) v_{max} (cm⁻¹): 3592, 3004, 1709, 1420, 1358, 1220, 1092, 902, 529; HRMS (m/z): (ESI) calcd. for $C_{34}H_{45}N_3NaO_4$ [M+Na]⁺: 582.3302, found 582.3295; [α]²⁵ = -15°(c = 0.01, MeOH).



TLC: $R_f = 0.4$ (10% Et₂O in DCM); ¹**H NMR** (600 MHz, C₆D₆) δ 6.53 (dd, J = 3.0, 1.8 Hz, 1H), 6.43 (d, J = 11.8 Hz, 1H), 6.34 (t, J = 3.1 Hz, 1H), 6.21 (d, J = 3.6 Hz, 1H), 6.09 (d, J = 2.8 Hz, 1H), 6.04 (d, J = 11.7 Hz, 1H), 5.98 (dd, J = 3.5, 0.9 Hz, 1H), 5.52 (s, 1H), 4.52 (s, 1H), 4.14 (p, J = 7.0 Hz, 1H), 4.02 – 3.99 (m, 1H), 3.61 (qd, J = 6.6, 2.1 Hz, 1H), 3.48 – 3.42 (m, 1H), 3.32 (s, 1H), 2.91 (q, J = 6.6 Hz, 1H), 2.44 – 2.33 (m, 2H), 2.24 (s, 3H), 2.13 (s, 1H), 1.79 (s, 3H), 1.77 (s, 3H), 1.21 (s, 9H), 1.21 (d, J = 6.6

Hz, 3H), 1.19 (d, J = 7.0 Hz, 3H), 0.78 (d, J = 6.6 Hz, 3H); ¹³**C** NMR (151 MHz, C₆D₆) δ 171.5, 132.3, 131.1, 128.5, 127.7, 125.9, 124.6, 119.8, 118.9, 116.4, 113.9, 108.9, 107.9, 107.2, 104.2, 91.1, 88.9, 79.4, 72.3, 61.2, 57.6, 55.0, 43.9, 40.8, 39.8, 28.0(3C), 19.3, 18.0, 17.3, 13.5, 12.5, 9.7; **IR** (thin film) v_{max} (cm⁻¹): 3360, 2922, 2852, 1727, 1646, 1465, 1425, 1280, 1159, 1080, 950, 786, 701; **HRMS** (m/z): (ESI) calcd. for C₃₄H₄₅N₃NaO₄ [M+Na]⁺: 582.3302, found 582.3296; [a]²⁵ = +12° (c = 0.01, MeOH).



(+)-Curindolizine (2): To a 10 mL reaction tube containing alcohol 84 (1.7 mg, $3.0 \,\mu$ mol, 1.0 equiv.) was added DCM (0.5 mL). The resulting solution was cooled to -78 °C and DIBAL (1.0 M in hexanes, 15.0 μ L, 15.0 μ mol, 5.0 equiv) was added dropwise. Upon completion of the reaction as indicated by TLC, the reaction mixture was quenched with saturated aq. Rochelle's salt (1 mL). The solution was extracted with EtOAc (3 x 5 mL) and the combined organics were washed with brine (5 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. The resulting crude residue was purified by preparative TLC (10% Et₂O in DCM) to afford (+)-curindolizine (2) (1.0 mg, 2.1 μ mol, 70%) as a white solid. **TLC**: $R_f = 0.8$ (10% Et₂O in DCM); ¹H NMR (600 MHz, CDCl₃) δ 6.51 (d, J = 9.6 Hz, 1H), 6.34 (d, J = 11.7 Hz, 1H), 6.03 (d, J = 3.5 Hz, 1H), 5.95 (d, J = 3.4 Hz, 1H), 5.87 (d, J = 3.5 Hz, 1H), 5.83 (d, J = 3.3 Hz, 1H), 5.69 (dd, J = 9.6, 6.3Hz, 1H), 5.59 (d, J = 11.7 Hz, 1H), 5.25 (s, 1H), 4.86 (s, 1H), 4.40 (dd, J = 2.1, 1.1 Hz, 1H), 4.11 (qd, J = 6.7, 2.3 Hz, 1H), 4.02 (q, J = 6.7 Hz, 1H), 3.69 (s, 1H), 3.39 (d, J = 6.3 Hz, 1H), 2.26 (s, 3H), 2.20 (s, 3H), 2.11 (s, 2H), 1.44 (d, J = 6.7 Hz, 3H), 1.33 (d, J = 6.7 Hz, 3H), 1.12 (d, J = 6.4 Hz, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 132.2, 130.4, 128.9, 128.2, 127.4, 124.0, 122.7, 122.5, 120.0, 119.7, 119.3, 113.2, 108.4, 106.5, 105.4, 103.6, 89.7, 88.9, 69.8, 60.0, 57.3, 54.2, 44.7, 38.3, 20.6, 19.6, 17.7, 13.8, 11.2, 10.3; **IR** (thin film) v_{max} (cm⁻¹): 3359, 2921, 2852, 1658, 1633, 1465, 720; **HRMS** (m/z): (ESI) calcd. for C₃₀H₃₅N₃NaO₂ [M+Na]⁺: 492.2621, found 492.2620; $[\alpha]^{25} = +314^{\circ}$ (c = 0.01, MeOH).

Procuramine ¹H spectra comparison:



procuramine

	¹ H NMR (δ)	¹ H NMR (δ)
Position	Natural Sample	Synthetic Sample
	(500 MHz, CDCl ₃) ⁹	(600 MHz, CDCl ₃)
1	1.37 (d, 7.0)	1.36 (d, 6.9)
2	4.40(qd, 7.0, 2.0)	4.40(qd, 6.8, 1.9)
3	4.36 (dt, 3.2, 2.0)	4.35 (br s)
4	2.89 (dd, 18.0, 3.2)	2.87 (dd, 18.0, 3.2)
	2.67 (dd, 18.0, 2.0)	2.64 (dd, 18.0, 2.7)
5		
6		
7	7.01 (br d, 4.0)	7.01 (d, 4.0)
8	6.08 (br d, 4.0)	6.08 (d, 4.0)
9		
10	2.32 (br s)	2.32 (s)

Procuramine ¹³C spectra comparison:



procuramine

	13 C NMR (δ)	^{13}C NMR (δ)
Position	Natural Sample	Synthetic Sample
	(125 MHz, CDCl ₃) ⁹	(151 MHz, CDCl ₃)
1	19.3	19.3
2	53.7	53.6
3	70.5	70.5
4	39.5	39.4
5	183.5	183.7
6	128.3	128.3
7	115.1	115.1
8	111.2	111.2
9	135.7*	135.8
10	12.0	12.0

*Revised shift by ¹³C NMR of the isolation paper

(+)-Curindolizine ¹H spectra comparison:



	1 H NMR (δ)	¹ H NMR (δ)
Position	Natural Sample	Synthetic Sample
	(400 MHz, CDCl ₃) ⁹	(600 MHz, CDCl ₃)
1	1.44 (d, 6.8)	1.44 (d, 6.7)
2	4.11(qd, 6.8, 2.0)	4.11(qd, 6.7, 2.3)
3	4.41 (br s)	4.40 (dd, 2.1, 1.1)
4	4.87 (br s)	4.86 (s)
5	3.70 (br s)	3.69 (s)
6		
7	5.25 (br s)	5.25 (s)
8		
9		
10	2.26 (br s)	2.26 (s)
11	1.13 (d, 6.4)	1.12 (d, 6.4)
12	2.19 (q, 6.4)	2.20 (s)
13		
14	5.60 (d, 11.6)	5.59 (d, 11.7)
15	6.35 (d, 11.6)	6.34 (d, 11.7)
16		
17	6.06 (d, 3.2)	6.03 (d, 3.5)
18	5.88 (d, 3.2)	5.87 (d, 3.5)
19		
20	2.21(br s)	2.20(s)
21	1.33 (d, 6.4)	1.33 (d, 6.7)
22	4.02 (q, 6.8)	4.02 (q, 6.7)
23	3.40 (br d, 6.0)	3.39 (d, 6.3)
24	5.69 (dd, 9.6, 6.4)	5.69 (dd, 9.6, 6.3)
25	6.51 (d, 9.6)	6.51 (d, 9.6)
26		
27	5.96 (d, 3.2)	5.95 (d, 3.4)
28	5.83 (d, 3.2)	5.83 (d, 3.3)
29		
30	2.12 (br s)	2.11 (s)

(+)-Curindolizine ¹³C spectra comparison:



	13 C NMR (δ)	¹³ C NMR (δ)
Position	Natural Sample	Synthetic Sample
	(101 MHz, CDCl ₃) ⁹	(151 MHz, CDCl ₃)
1	19.6	19.6
2	57.3	57.3
3	88.9	88.9
4	59.9	60.0
5	44.6	44.7
6	128.2	128.2
7	103.5	103.6
8	127.4	127.4
9	122.7	122.7
10	10.2	10.3
11	17.6	17.7
12	69.8	69.8
13	89.7	89.7
14	119.7	119.7
15	124.0	124.0
16	130.4	130.4
17	113.2	113.2
18	108.4	108.4
19	132.2	132.2
20	13.7	13.8
21	20.6	20.6
22	54.2	54.2
23	38.2	38.3
24	119.3	119.3
25	119.9	120.0
26	122.4	122.5
27	105.4	105.4
28	106.5	106.5
29	128.2	128.2
30	11.2	11.2





































































































































































































































Chapter 3

The Syntheses of Bipolamines D, E, G, and

3.1 Introduction

This chapter chronicles our chemical investigations of bispyrrole metabolites derived from *Bipolaris maydis*. A total of 9 pyrrole-containing secondary metabolites were identified in 2020 from this source.¹ Apart from simpler bipolamine A (**3**) and bipolamine (**4**), the other congeners are differentially oxidized variants of curvulamine (**1**) (Figure 3.1).^{2,3} In the case of bipolamine C (**5**), the vinyl pyrrole has undergone oxidation at C-18 and C-19 to yield a hydroxypyrrolone structure. Bipolamines D (**6**) and E (**7**) are C-14 epimers wherein a hydroxyl group at C-14 replaces the Δ_{14-15} olefin, and in bipolamine G (**9**) further oxidation of the alkene to a monomethylatated trans diol has occurred. Bipolamine H (**10**) and bipolamine F (**8**) contain oxidation at C-5, in the form of a hydroxyl group in **10** and a THF ring in the case of **8**. Lastly, bipolamine I (**11**) is structurally distinct with its congeners in that is the only isolated metabolite with an ether bridge between C-14 and C-3. Driven to further explore the chemical reactivity of these pyrrole-containing natural products and investigate the superior reported bioactivity of certain bipolamines relative to curvulamine (**1**) (*see* Chapter 1), we aimed to develop a unifying strategy toward these novel bispyrrole alkaloids from a common intermediate.^{4,5}



Figure 3.1 Curvulamine-type alkaloids isolated from Curvularia sp. and Bipolaris maydis.

3.2 Pierce's Total Synthesis of Bipolamine I

In 2022, the Pierce group at North Carolina State University reported the synthesis of bipolamine I (**11**) from mono-ethyl malonate and 2-methylpyrrole.⁶ Their 15-step synthesis of **11** featured a two-component ruthenium-catalyzed transfer hydrogenative coupling that yielded most of the carbon skeleton of **11** (Figure 3.2).⁷ One of the components, alkyne **85** was made in a few steps from mono-ethyl malonate. First, activation of levulinic acid by CDI, followed by Claisen reaction of the malonte and the levulinic-CDI complex with concomitant decarboxylation proceed to give ester **86** (Mg(OEt)₂, CDI). A subsequent Paal-Knorr pyrrole synthesis using propargyl amine then gave **87** in 84% yield (AcOH, PhMe, Δ). DIBAL-mediated reduction of the ethyl ester **87**

to the corresponding aldehyde followed by 1,2 addition of vinyl Grignard to the aldehyde intermediate gave a secondary alcohol that was immediately protected with TBSCI to afford **85** (Figure 3.2A). The other pyrrole-containing component **88** was made from methyl pyrrole in two steps: intermolecular substitution reaction between 2-methyl pyrrole and **29** (NaH, DMF) followed by LiAlH₄ reduction to alcohol **88**. Base mediated isomerization of alkyne **85** (*t*-BuOK, *t*-BuOH) and subsequent ruthenium-catalyzed transfer hydrogenative coupling with alcohol **88** gave bispyrrole **89** in a 3.7:1 mixture of diastereomers with a combined 68% yield (HCIRu(CO)(PPh₃)₃, dippf, 1M dioxane). This intermediate was transformed into **90** after a ring closing metathesis event (Grubbs II, PhMe) and TBAF deprotection sequence. In an unexpected event, exposure of **90** to





Figure 3.2 Pierce's total synthesis of bipolamine I. 3.2A Synthesis of pyrrolecontaining coupling fragments. 3.2B Completion of the synthesis of 11

MnO₂ not only resulted in oxidation of the allylic alcohol to an enone that reacted with the neighboring pyrrole, but also resulted in the formation of the ether bridge between C-14 and C-3 ultimately yielding **91**. An aldol reaction between the kinetic enolate of **91** (LiHMDS, THF) and acetaldehyde gave a secondary alcohol which was protected (TBSCI, imidazole, DMAP) to generate ketone **92**. Exposure of **92** to reductive conditions gave hemiacetal **93** in 95% yield (SmI₂, THF). Finally, **93** was converted to a thiocarbonate (KHMDS, CICOSPh), which underwent smooth radical deoxygenation (BEt₃, n-Bu₃SnH) and TBAF mediated deprotection completing the synthesis of bipolamine I (**11**).



3.3 Unifying Strategy Toward the Synthesis of the Bipolamine Alkaloids

Figure 3.3 Proposed strategy to access all Bipolaris maydis metabolites.

During our synthetic planning, we aimed to develop an approach that targets all the bipolamines from a common intermediate (Figure 3.3). While this strategy might sound routine within a family of natural products,⁸ we anticipated significant challenges due to the notorious sensitivity of these pyrrole-containing compounds to acidic and oxidative conditions.⁹ As discussed in previous chapters, these properties of electron-rich pyrrole units, such as the ones found in the bipolamine alkaloids, greatly limits the chemical reactions that can be used with these natural products, especially in an oxidative context.^{10,11}

In our strategy, we anticipated that bipolamines D (6) and E (7) could be obtained from curvulamine (1) after hydroboration and oxidation.¹² Previously prepared methyl ketone 35 could be elaborated into bipolamine G (9) in two steps: an electrophilic epoxidation (*m*-CPBA) followed by an epoxide ring opening reaction with methanol.^{13,14} Recognizing that bipolamine C (5) has an oxidized pyrrole unit, we envisioned a series of oxidation reactions could furnish this congener. Starting from 5, m-CPBA mediated oxidation of the western pyrrole followed by hydrolysis of the newly formed epoxide ring, could produce allylic alcohol intermediate 94. A chemoselective oxidation of 94 with MnO2 and subsequent base-mediated elimination of the hydroxyl group at C-14 could produce bipolamine C (5).¹⁵ To elaborate bipolamine E (7), we proposed a regioselective ahydroxylation of 24 to make 95 followed by an analogous sequence of reactions as the one used to make 1.¹⁶ We anticipate that bipolamine F (8) could be obtained from bipolamine E (7) after an acid-catalyzed cyclization. Lastly, we designed a strategy to synthesize bipolamine I (11) from methyl ketone 35 in 3 steps: samarium diiodide mediated reduction of 35, then a base-catalyzed intramolecular conjugate addition, and finally a stereodivergent reduction of ketone 96.17 Our proposal employs methyl ketone (35) as a key common synthetic intermediate thus we first investigated synthetic routes to produce large quantities of 35.18,19

3.4 Scalable Synthetic Plan to Access Intermediate 35

During our campaigns toward curvulamine (1) and curindolizine (2),¹⁹ we observed inconsistent results in the Barton-McCombie deoxygenation step. We noticed that the yield of the radical deoxygenation oscillated between 40% to 10% depending on the scale. Presumably, the substrate or the reaction conditions are very sensitive to the amount of oxygen used during the radical initiation step (O₂, Et₃B), which is hard to control accurately.¹⁵ While one could have assumed that this accuracy problem could have been less chronic in larger scales, the issue still persisted. These results prompt us to look for a more robust and scalable strategy to access methyl ketone **35** from pentacycle **31**.

Beginning with pentacycle **31**, thermodynamic isomerization of the C-2 stereocenter proceeded to give an inseparable mixture of diastereomers **31** and **32** (NaOMe, MeOH). This diasteoromeric mixture was acetylated (Ac₂O, Et₃N, 4-DMAP) to yield **97** and the corresponding C-2 epimer. These diastereomers could be easily separated and heating 2-*epi*-**97** in sodium methoxide afforded a mixture containing **31** and **32**. Global reduction (LiAlH₄, THF) of pure **97** gave diol **98** in 85% yield. Additionally, we found that this transformation could be carried out in gram scale without any decrease in yield. Mesylation (MsCl, DMAP, Et₃N) of the more accessible secondary alcohol gave **99**, and intramolecular displacement of the activated alcohol by the tertiary alcohol in **99** (DBU, PhMe, Δ) followed by hydrolysis of the ethyl vinyl ether during the acidic work-up afforded methyl ketone **35**. Even though, this new synthetic route to **35** is one step longer compared to the previous Barton-McCombie deoxygenation sequence, we were able to more than double the overall yield and more easily obtain large quantities of **35** in a short amount of time. In addition, as we will discuss later, some of the lessons learned during this optimization campaign proved critical in the synthesis of bipolamine I (**11**).



Figure 3.3 Scalable route toward intermediate 35.

3.5 The Total Syntheses of Bipolamines D and E

Having secured a scalable procedure to access methyl ketone **35** we then set our sights on the synthesis of bipolamine D (**6**) and bipolamine E (**7**). Stereodivergent reduction of **35** under previously discussed conditions ((*R*)-CBS, BH₃·THF) proceed to give **1** and 12-*epi*-**1** in a combined 90% yield.¹⁸ Brown hydroboration-oxidation of **1** (BH₃•DMS *then* NaOH, H₂O₂) generated **5** and **6** in almost equimolar quantities. To rationalize the regioselectivity of this reaction, we analyzed the Muliken charge distribution of **35** (Chapter 2, section 1.4), but these data did not provide evidence of an electronic basis for the selectivity.^{20,21} Perturbations of hydroboration regioselectivity and stereoselectivity by oxygen-containing substituents are well reported in the literature, however, and a mechanism that involves the hydroxyl group at C-12 directing the reaction could be operating, which would explain the observed regioselectivity.^{22,23}



Figure 3.4 Syntheses of bipolamines D and E.

3.6 The Total Synthesis of Bipolamine G

During our synthetic planning to access bipolamine G (9) (Figure 3.3), we proposed to transform **35** into **9** via an electrophilic epoxidation of the Δ_{14-15} alkene followed by opening of the oxirane ring with MeOH.¹⁵ When we subjected **35** to *m*-CPBA at –35° C, we observed formation of methyl ether compound **100**, likely via a mechanism that involves oxidation of the eastern pyrrole (**101**). This formal benzylic oxidation product was

unexpected because during previous studies on 35 and its derivatives, we only observed reactivity at the western pyrrole unit.^{18,19} For instance, during our synthetic studies of curindolizine (2), treatment of 35 with NIS in methanol at low temperatures gave 76 as the only product. While trying to further investigate the unexpected oxidation of 35, we quickly recognized that 76 could be elaborated into bipolamine G (9) via solvolysis of the iodide atom at C-14. Methyl ether 76 was treated with silver (I) salts in a nitromethanewater mixture, which we hoped would generate a carbocation that could be intercepted by water.²⁴ Under these conditions however, instead of forming bipolamine G (9), we observed exclusive formation of aldehyde 102, presumably formed via a semi pinacoltype rearrangement aided by the neighboring pyrrole. We then evaluated a one-electron approach to this transformation, but subjecting 76 to radical conditions (Bu₃SnH, O₂, AIBN) only afforded the reduction product. In a revised approach, dihydroxylation of 35 (OsO₄, NMO) was found to be productive, albeit with poor conversion due to decomposition upon prolongation of reaction time. Diol 103 proceeded to give substitution product 104 in 85% yield (MeOH, HCI), possibly via the initial formation of an azafulvenium ion, which is then trapped by methanol.²⁵ DIBAL reduction of **104** yielded nearly identical quantities of bipolamine G (9) and its C-12 epimer. Notably, during the synthesis of curvulamine (1), when compound 35 is treated to the same reduction conditions (DIBAL, DCM), primarily the C-12 epimer of the corresponding product is obtained.18



Figure 3.5 Synthesis of bipolamine G. 3.5A. Initial strategy toward 9 3.5B Second generation route toward 9 3.5C Successful synthesis of 9

3.7 The Total Synthesis of Bipolamine I

After completing the synthesis of **9**, we directed our efforts toward bipolamine I (**11**), a topologically distinct molecule relative to its congeners. We rationalized that **11** could be elaborated from enone **105** via an intramolecular Michael addition and ketone reduction sequence. Enone intermediate **105** could be prepared from methyl ketone **35** via a reductive ring cleavage (Figure 3.6).¹⁷



Figure 3.6 Bipolamine I retrosynthetic analysis.

Exposing **35** to reductive conditions (Sml₂, THF) proceeded to smoothly cleave the C-O bond, but upon work-up, the extended samarium enolate yielded a 10:1 mixture of alkene **106** and hemiacetal **107** instead of the desired enone **105**. Different conditions were explored to isomerize the double bond into conjugation with the carbonyl group to obtain **105**. Ketone **106** was heated with *t*-BuOK in THF and at temperatures below 60



B. E(RB3LYP) Obtained from Frenquency Calcultations at the 3LYP, 6-31(d,p) Level of Theory



Figure 3.7 Bipolamine I synthetic investigations. **3.7A** First generation strategy toward bipolamine I. **3.7B** DFT calculations of isomerization intermediates.

°C, only unreacted starting material was recovered (entry 1) while decomposition products were observed at elevated temperatures (entry 2). Heating **106** in the presence of DBU at refluxing temperatures resulted in isomers **106** and **107** (3:1 ratio). This result indicated that deprotonation is possible, but isomerization is unfavorable. Attempts to perform a metal-catalyzed isomerization with RhCl₃•6H₂O or PdCl₂(MeCN)₂ resulted in mixture of decomposition products (entry 4 and entry 5).²⁶⁻²⁸ DFT calculations of the ground state energies of the optimized structures of **105** and **106** at the B3LYP/6-31 G (d,p) level of theory, informed us that alkene **106** is 5.98 kcal/mol lower in energy compared to enone **105**, supporting our results at attempting to carry out a thermodynamic isomerization.



Figure 3.8 Synthesis of bipolamine I.

Recognizing the smooth hydroboration of the Δ_{14-15} olefin during the synthesis of **5** and **6**, a revised strategy was devised (Figure 3.8). Starting from **98**, regioselective mesylation (MsCl, Et₃N, DMAP) with concomitant acidic hydrolysis of the enol ether afforded ketone **108**. Subsequent Sml₂ mediated single-electron reduction of **108** furnished the corresponding ketone product which was equilibrated to a single diastereomer (**109**) after addition of DBU. Methyl ketone **109** was transformed into bipolamine I (**11**) in three steps. First, **109** was reduced (NaBH₄, MeOH) to a separable mixture of C-12 epimers in a 1:1 ratio. Second, regioselective hydroboration oxidation (BH₃•THF *then* NaBO₃) of the Δ_{14-15} alkene yielded the desired isomer in good yields. Finally, DBU mediated intramolecular SN₂ at elevated temperatures proceeded to give bipolamine I (**11**).

3.8 Future Directions

Through our investigations, we have established efficient synthetic routes to several curvulamine type alkaloids. We are currently still investigating the synthesis of the syntheses of bipolamines C, E and F (Figure 3.3). So far, during our studies on the synthesis of bipolamine C (5) and E (7) we haven't observed oxidation of curvulamine (1) at C-5 or C-18/19 π -bond and selective oxidation at these positions is required for the synthesis of the remaining bipolamine congeners. Recently, we discovery conditions to selectively oxidize 24 at C-5, and we are investigating conditions to transform this intermediate into bipolamines E and F. We are concurrently investigating different strategies to synthesize bipolamine C from methyl ketone 35.



Figure 3.9 Possible future synthetic strategies to access bipolamines C, H, and F.

3.9 Conclusion

This chapter has chronicled the chemistry of the electron-rich pyrrole natural products observed while investigating the synthesis of several curvulamine derivatives. Several novel bipolamine alkaloids have been synthesized, including **9** the most potent antibiotic in the family. This work lays the foundation for future mechanism of action studies of these molecules.

3.10 Distribution of Credit

Second-generation synthesis of methyl ketone **35** was performed by Jun Xuan and Paulo Andre Machicao Tello. The investigation and synthesis **5** and **6** were performed by Paulo Andre Machicao Tello with refinement from Professor Thomas J. Maimone. The synthesis of **9** and **11** were performed by Jun Xuan. Preliminary studies on the synthesis of **7** and **8** were performed by Paulo Andre Machicao Tello. Characterization of all compounds was completed by Paulo Andre Machicao Tello and Jun Xuan. We thank the NIH NIGMS (R01GM136945 to Thomas J. Maimone, and diversity supplement to Paulo Andre Machicao Tello) for financial support. We thank Dr. Hasan Celik and Dr. Jeffrey G. Pelton for nuclear magnetic resonance spectroscopic assistance (NIH grant GM68933). We also thank QB3/Chemistry Mass Spectrometry Facility scientist Dr. Zhongroui Zhou for mass spectrometry assistance.

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3.12 Experimental Procedures and Characterization Data

3.12.1 General Procedures

All reactions were performed in flame- or oven-dried glassware under a positive pressure of nitrogen or argon, unless otherwise noted. Air-and moisture-sensitive liquids were transferred via syringe. When indicated, solvents or reagents were degassed by sparging with argon for 10 min in an ultrasound bath at 25 °C. Volatile solvents were removed under reduced pressure rotary evaporation below 35 °C. Analytical and preparative thin-layer chromatography (TLC) were performed using glass plates precoated with silica gel (0.25-mm, 60-Å pore size, Merck TLC Silicagel 60 F₂₅₄) impregnated with a fluorescent indicator (254 nm). TLC plates were visualized by exposure to ultraviolet light (UV) and then were stained by submersion in an ethanolic anisaldehyde solution or ceric ammonium molybdate solution, followed by brief heating on a hot plate. Flash column chromatography was performed with silica gel purchased from Silicycle (SiliaFlash[®], 60 Å, 230-400 mesh, 40-63 µm). Ethyl vinyl ether and 2bromopropanic acid methyl ester were distilled over calcium hydride prior to use. NaHMDS solutions were purchased from Sigma. All other reagents were used as received from commercial sources, unless stated otherwise. Anhydrous tetrahydrofuran (THF), dichloromethane (DCM), methanol (MeOH), dimethylformamide (DMF), and toluene (PhMe) were obtained by passing these previously degassed solvents through activated alumina columns. Proton nuclear magnetic resonance (¹H NMR) spectra and carbon nuclear magnetic resonance (¹³C NMR) spectra were recorded on Bruker AV-600 spectrometer at 23 °C. Proton chemical shifts are expressed as parts per million (ppm, δ scale) and are referenced to residual solvent (CDCl₃: δ 7.26, C₆D₆: δ 7.16), unless stated otherwise. Carbon chemical shifts are expressed as parts per million (ppm, δ scale) and are referenced to the solvent (CDCl₃: δ 77.16, C₆D₆: δ 128.06), unless stated otherwise. Data is represented as follows: chemical shift, multiplicity (s = singlet, d = doublet, dd = doublet of doublets, ddd, doublet of doublet of doublet, dt = triplet of doublets, t = triplet, q = quartet, m = multiplet, br = broad), coupling constant (J) in Hertz (Hz), and integration. Infrared (IR) spectra were recorded on a Bruker Alpha FT- IR spectrometer as thin films and are reported in frequency of absorption (cm⁻¹). Only selected resonances are reported. High-resolution mass spectra (HRMS) were obtained by the mass spectrometry facility at the University of California, Berkeley using a Finnigan LTQFT mass spectrometer (Thermo Electron Corporation). X-ray diffraction data was collected at the Small Molecule X-ray Crystallography Facility (CheXray) at University of California, Berkeley using a Rigaku XtaLAB P200 equipped with a MicroMax 007HF rotating anode and Pilatus3 R 200K-A hybrid pixel array detector. Data were collected using CuKa radiation ($\lambda = 1.5418$ Å).



3.12.2 Experimental Procedures and Tabulated Characterization Data

Tetracyclic diol 13: To a 100 mL round bottom flask containing pentacyclic bispyrrole **31** and **32** (1.5 g, 4.10 mmol, 1.0 equiv.) was added CH₂Cl₂ (30 mL). To the stirring solution was added Ac₂O (577 μ L, 6.15 mmol, 1.5 equiv.), DMAP (500 mg, 4.10 mmol, 1.0 equiv.) and Et₃N (1.71 ml, 12.3 mmol, 3.0 equiv.). Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated *aq*. NaHCO₃ (50 mL) and extracted with CH₂Cl₂ (3 x 50 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, and concentrated *in vacuo*. The resulting crude residue was purified by column chromatography (5% EtOAc in hexanes) to afford **97** (1.12 g, 2.73 mmol) and 2-*epi*-**97** (477 mg,1.17 mmol).

To a reaction tube containing 2-epi-12 (477 mg, 1.17 mmol, 1.0 equiv.) was added NaOMe (317 mg, 5.85 mmol, 5.0 equiv.). The reaction tube was sealed and anhydrous MeOH (20 mL) was then added, the sealed reaction vessel was heated at 90 °C for 4 hours. The reaction mixture was then cooled to room temperature guenched with saturated. aq. NH₄Cl solution (50 mL). The solution was extracted with EtOAc (3 x 30 mL) and the combined organic layers were washed with brine (50 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The resulting crude residue was purified by column chromatography (0% EtOAc in hexanes \rightarrow 20% EtOAc in hexanes) to afford an inseparable mixture of pentacyclic bispyrrole 11 and 2-epi-11 (2.3:1 ratio) (367 mg, 0.99 mmol, 85%) as a brown foam. To a 100 mL round bottom flask containing 97 (1.12 g, 2.73 mmol, 1.0 equiv.) was added THF (30 mL). To the vigorously stirring solution was added LiAlH₄ (1.0 M in THF, 8.19 mL, 8.19 mmol, 3.0 equiv.) dropwise. Upon consumption of the starting material as indicated by TLC, the reaction mixture was cooled to 0 °C and guenched with saturated ag. Rochelle salt (30 mL) and extracted with EtOAc (3 x 50 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, and concentrated *in vacuo*. The resulting crude residue was purified by column chromatography (10% EtOAc in hexanes) to afford tetracyclic diol **98** (854 mg, 2.32 mmol, 85%) as a yellow oil.

TLC: $R_f = 0.5$ (30% EtOAc in hexanes).¹**H NMR** (600 MHz, Benzene-*d*₆) δ 6.34 (d, *J* = 12.5 Hz, 1H), 6.15 (d, *J* = 3.5 Hz, 1H), 6.03 –5.99 (m, 2H), 5.93 (dd, *J* = 3.5, 0.9 Hz,

1H), 5.51 (d, J = 12.4 Hz, 1H), 5.44 (ddd, J = 11.4, 6.1, 3.5 Hz, 1H), 4.85 (dd, J = 11.4, 3.8 Hz, 1H), 4.69 (d, J = 3.7 Hz, 1H), 4.45 (d, J = 2.4 Hz, 1H), 4.36 (p, J = 6.4 Hz, 1H), 3.95 (d, J = 2.4 Hz, 1H), 3.48 – 3.37 (m, 2H), 2.00 (s, 3H), 1.97 (s, 3H), 1.70 (s, 1H), 1.64 (d, J = 2.5 Hz, 1H), 1.21 (d, J = 6.5 Hz, 3H), 0.92 (t, J = 7.0 Hz, 3H) ¹³**C NMR** (151 MHz, C₆D₆) δ 167.1, 134.8, 129.4, 128.4, 126.2, 124.9, 120.5, 112.9, 108.4, 108.2, 106.8, 81.4, 79.9, 64.9, 63.8, 55.8, 54.0, 45.4, 15.4, 14.3, 13.6, 12.3. **IR** (thin film) v_{max} (cm⁻¹): 3451, 2977, 2924, 1710, 1616, 1415, 1239, 1016, 871, 773. 748. **HRMS** (m/z): (ESI) calcd. for C₂₂H₂₉N₂O₃ [M+H]⁺ m/z: 369.2173, found 369.2174.



Mesylate 99: To a 100 mL round bottom flask containing tetracyclic diol **98** (500 mg, 1.36 mmol, 1.0 equiv.) was added CH₂Cl₂ (30 mL). The resulting mixture was cooled to 0 °C, Et₃N (567 μ L,4.08 mmol, 3.0 equiv.), MsCl (116 μ L, 1.5 mmol, 1.1 equiv.) and DMAP (166 mg, 1.36 mmol,1.0 equiv.) were added dropwise sequentially. The resulting mixture was warmed to room temperature. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NaHCO₃ (30 mL) and extracted with EtOAc (3 x 30mL). The combined organic layers were washed with brine (20 mL), dried over Na2SO4, and concentrated in vacuo. The resulting crude product was purified by column chromatography (10% EtOAc in hexanes) to afford mesylate **99** (443 mg, 0.95 mmol, 70%) as a white solid.

TLC: $R_f = 0.6$ (30% EtOAc in hexanes). ¹**H NMR** (600 MHz, Chloroform-*d*) δ 6.30 (d J = 12.5 Hz, 1H), 6.17 (dd, J = 11.8, 6.3 Hz, 1H), 6.09 (d, J = 3.6 Hz, 1H), 5.94 (d, J = 3.5 Hz, 1H), 5.89 (d, J = 3.4 Hz, 1H), 5.70 (d, J = 3.4 Hz, 1H), 5.26 (d, J = 12.4 Hz, 1H), 5.04 (dd, J = 11.8, 3.7 Hz, 1H), 4.82 (p, J = 6.5 Hz, 1H), 4.44 (d, J = 3.7 Hz, 1H), 4.19 (d, J = 2.8 Hz, 1H), 4.01 (d, J = 2.8 Hz, 1H), 3.84 – 3.70 (m, 2H), 2.44 (s, 3H), 2.29 (s, 3H), 2.28 (s, 3H), 1.51 (s, 1H), 1.50 (d, J = 6.9 Hz, 3H), 1.30 (t, J = 7.0 Hz, 3H). ¹³**C** NMR (151 MHz, CDCl₃) δ 165.9, 134.8, 129.1, 128.8, 125.1, 124.4, 120.0, 113.0, 108.5, 108.2, 106.7, 81.4, 79.7, 74.5, 63.8, 53.4, 53.0, 44.2, 36.6, 16.6, 14.5, 13.7, 12.6. IR (thin film) v_{max} (cm⁻¹): 3441, 2924, 2854, 1665, 1607, 1362, 1299, 1179, 885, 758, 406. HRMS (m/z): (ESI) calcd. for C₂₃H₃₁N₂O₅S [M+H]⁺ m/z: 447.1948, found 447.1947.



Ketone 35: To a reaction tube containing mesylate **99** (300 mg, 0.67 mmol, 1.0 equiv.) was added PhMe (20 mL) and DBU (100 μ L, 0.67 mmol, 1.0 equiv.). The resulting mixture was heated up to 120 °C. Upon consumption of the starting material as indicated by TLC, the reaction mixture was cooled to 25 °C followed by the addition of aq. 1 M HCl (5 mL).

Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NH₄Cl (10 mL) and extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude residue was purified by column chromatography (20% EtOAc in hexanes) to afford ketone **35** (186 mg, 0.57 mmol, 86%) as a white solid. **TLC**: $R_f = 0.5$ (40% EtOAc in hexanes). ¹**H NMR** (600 MHz, Benzene-*d*₆) δ 6.28 (d *J* = 11.5 Hz, 1H), 6.15 (d, *J* = 3.6 Hz, 1H), 5.94 (dd, *J* = 3.5, 0.9 Hz, 1H), 5.87 (dd, *J* = 3.4, 1.0 Hz, 1H), 5.77 (d, *J* = 3.4 Hz, 1H), 5.74 (d, *J* = 11.5 Hz, 1H), 4.46 (br s, 1H), 4.14 (br s, 1H), 3.72 (br s, 1H), 3.69 (dq, *J* = 6.7, 2.1 Hz, 1H), 1.83 (d, *J* = 0.9 Hz, 3H), 1.73 (s, 3H), 1.53 (s, 3H), 0.73 (d, *J* = 6.7 Hz, 3H). ¹³**C NMR** (150.9 MHz, C₆D₆) δ 209.7, 132.8, 130.6, 128.6, 127.4, 124.0. 121.6, 114.5, 109.0. 108.9, 106.3, 94.0, 90.5, 60.0, 56.7, 46.1, 25.2, 19.1, 13.4, 12.3. **IR** (thin film) v_{max} (cm⁻¹): 2972, 2932, 1741, 1682, 1640, 1456, 1378, 1214, 1071, 786, 734. **HRMS** (m/z): (ESI) calcd. for C₂₀H₂₃O₂N₂ [M+H]⁺: 323.1754, found 323.1756.



Curvulamine (1): In a N₂ filled glovebox, a reaction tube was charged with (*R*)-(+)-2methyl-CBS-oxazaborolidine (86 mg, 0.31 mmol, 1.0 equiv.). The reaction tube was sealed and brought out of the glovebox under inert atmosphere. CH_2Cl_2 (2.0 mL) was added followed by BH₃•DMS (60 µL, 0.62 mmol, 2.0 equiv.) and the mixture stirred for 15 minutes. Ketone **35** (100 mg, 0.31 mmol, 1.0 equiv.) was dissolved in CH_2Cl_2 (2.0 mL) and added dropwise to the reaction mixture. Additional CH_2Cl_2 (1.5 mL) was used to render the transfer quantitative. Upon completion of the reaction as indicated by TLC, saturated aq. NH₄Cl solution (5 mL) was added, and the mixture stirred for 5 minutes. The biphasic mixture was poured into a separatory funnel and the organic layer was extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The resulting crude residue was purified by preparative TLC (50% Et2O in hexanes) to afford curvulamine **1** (46 mg, 0.14 mmol, 45%) and 12-*epi*-**1** (44 mg, 0.14 mmol, 45%) both as white solids.

curvulamine (1):

TLC: $R_f = 0.4$ (40% EtOAc in hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 6.45 (d, J = 11.6 Hz, 1H), 6.13 (d, J = 3.6 Hz, 1H), 5.94 (br d, J = 3.6 Hz, 1H), 5.93 (d, J = 3.4 Hz, 1H), 5.92 (d, J = 3.4 Hz, 1H), 5.72 (d, J = 11.6 Hz, 1H), 4.94 (br, s, 1H), 4.50 (br, s, 1H), 4.21 (qd, J = 6.7, 1.9 Hz, 1H), 3.95 (br, s, 1H), 2.67 (br s, 1H), 2.31 (br s, 3H), 2.29 (br s, 3H), 1.51 (d, J = 6.7 Hz, 3H), 1.24 (d, J = 6.4 Hz, 3H). ¹³**C NMR** (151 MHz, CDCl₃) δ 132.4,129.8, 128.5, 124.2, 119.7, 113.4, 108.5 103.5, 89.4, 89.0, 70.2, 60.3, 57.4, 44.9, 19.5, 17.3, 13.8, 12.8. **IR** (thin film) v_{max} (cm⁻¹): 3466, 2972, 2925, 3852, 1643, 1425, 1393, 1322,

1301, 1044, 1011, 763. **HRMS** (m/z): (ESI) calcd. for C₂₀H₂₄O₂N₂Na [M+Na]⁺: 347.1730, found 347.1731.

12-epi-**1** :

TLC: R_f = 0.2 (40% EtOAc in hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 6.41(d, *J* =11.7 Hz, 1H), 6.10 (d, *J* = 3.6 Hz, 1H), 5.91 (dd, *J* = 3.6, 0.9 Hz, 1H), 5.86 (dd, *J* = 3.4, 1.0 Hz, 1H), 5.79 (d, *J* = 3.4 Hz, 1H), 5.75 (d, *J* = 11.7 Hz 1H), 4.90 (br, s, 1H), 4.56 (br, s, 1H), 4.23 (qd, *J* = 6.7, 2.1 Hz, 1H), 3.71 (br, s, 1H), 2.71 (q, *J* = 6.3 Hz, 1H), 2.27 (br s, 3H), 2.26 (br s, 3H), 2.05 (d, *J* = 2.2 Hz, 3H), 1.48 (d, *J* = 6.7 Hz, 3H), 1.10 (d, *J* = 6.3 Hz, 3H). 13C NMR (151 MHz, CDCl₃) δ 132.6; 130.3, 128.4, 128.0, 124.1, 118.9, 113.5, 108.5, 107.2, 105.0, 90.6, 89.4, 89.1, 72.0, 61.0, 57.6, 43.7, 19.4, 16.9, 13.8, 12.8. **IR** (thin film) v_{max} (cm⁻¹): 3468, 2925, 1644, 1415, 1299, 1077, 776, 670. HRMS (m/z): (ESI) calcd. for C₂₀H₂₄O₂N₂Na [M+Na]⁺: 347.1730, found 347.1731.



Bipolamine D (6) and bipolamine E (7): To a reaction tube containing curvulamine **(1)** (5.0 mg, 0.015 mmol, 1.0 equiv.) and THF (0.8 mL) was added BH₃•DMS (10.0 μ L, 0.020 mmol, 1.3 equiv.) at 25 °C. The resulting mixture was stirred at 25 °C for 3 hours and then cooled down to 0 °C. At this point, aq. 1 M NaOH (23.1 μ L, 0.023 mmol, 1.5 equiv.) and 50% H₂O₂ (4.4 μ L, 0.077 mmol, 5 equiv.) were added to the reaction mixture. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NaHCO₃ (3 mL) and extracted with CH₂Cl₂ (3 x 5 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude residue was purified by preparative TLC (50% EtOAc in hexanes) to afford bipolamine D **(6)** (2.1 mg, 0.0061 mmol, 40%) as a white solid and bipolamine E **(7)** (1.8 mg, 0.0053 mmol, 34%) as a white solid.

Bipolamine D:

TLC: $R_f = 0.35$ (50% EtOAc in hexanes). ¹H NMR (600 MHz, Acetone-*d*₆) δ 5.82 (d *J* = 3.3 Hz, 1H), 5.75 (br d, *J* = 3.3 Hz, 1H), 5.70 (br d, *J* = 3.3 Hz, 1H), 5.67 (br d, *J* = 3.3 Hz, 1H), 4.84 (s, 1H), 4.38 (s, 1H), 4.36 (s, 1H) , 4.32 (qd, *J* = 6.6, 2.3 Hz, 1H), 4.04 (d, *J* = 4.3 Hz, 1H), 3.91 (s, 1H), 3.73 (d, *J* = 1.9 Hz, 1H), 3.09 (d, *J* = 15.6 Hz, 1H), 2.93 (qd, *J* = 6.4, 4.5 Hz, 1H), 2.87 (dd, *J* = 15.6, 5.0 Hz, 1H), 2.24 (s, 3H), 2.21 (s, 3H), 1.48 (d, *J* = 6.6 Hz, 3H), 1.12 (d, *J* = 6.4 Hz, 3H). ¹³C NMR (151 MHz, Acetone) δ 131.5, 128.8, 128.6, 128.3, 108.8, 107.8, 107.0, 105.5, 91.8, 85.6, 72.3, 68.3, 59.6, 59.0, 43.7, 31.4, 19.2, 18.8, 13.5, 12.8. IR (thin film) v_{max} (cm⁻¹): 3520, 2925, 2855, 2150, 1465, 1298, 1022, 755, 619, 429. HRMS (m/z): (ESI) calcd. for C₂₀H₂₇N₂O₃ [M+H]⁺ m/z: 343.2016, found 343.2016.

Bipolamine E:

TLC: $R_f = 0.32$ (50% EtOAc in hexanes). ¹H NMR (600 MHz, Acetone-*d₆*) δ 5.92 (d, *J* = 3.4 Hz, 1H), 5.78 (d, *J* = 3.4 Hz, 1H), 5.74 (d, *J* = 3.3 Hz, 1H), 5.67 (d, *J* = 3.3 Hz, 1H), 4.91 (s, 1H), 4.35 – 4.32 (m, 1H), 4.30 (br s, 1H), 4.30 – 4.29 (m, 1H), 4.22 (br s, 1H), 3.07 (d, *J* = 15.7 Hz, 1H), 3.03 (d, *J* = 4.2 Hz, 1H), 2.95 (d, *J* = 9.8 Hz, 1H), 2.82 (d, *J* = 4.0 Hz, 1H), 2.53 (q, *J* = 6.5 Hz, 1H), 2.26 (s, 3H), 2.26 (s, 3H), 1.55 (d, *J* = 6.7 Hz, 3H), 1.08 (d, *J* = 6.5 Hz, 3H). ¹³C NMR (151 MHz Acetone) δ 131.4, 128.7, 128.6, 128.2, 108.7, 107.7, 107.0, 105.5, 91.7, 85.5, 72.2, 68.2, 59.5, 59.0, 43.6, 31.3, 19.1, 18.7, 13.4, 12.8. (IR thin film) v_{max} (cm⁻¹): 3575, 2925, 2854, 2106, 1445, 1296, 1048, 802, 760, 621, 429. HRMS (m/z): (ESI) calcd. for C₂₀H₂₇N₂O₃ [M+H]⁺ m/z: 343.2016, found 343.2015.



Formal benzylic oxidation product 100: To a reaction tube containing ketone **35** (5.0 mg, 0.016 mmol, 1.0 equiv.) was added CH₂Cl₂/MeOH (1:1, 1 mL). The resulting mixture was cooled to –35 °C followed by the addition of *m*-CPBA (purified1, 2.8 mg, 0.016 mmol, 1.0 equiv.). Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. Na₂SO₃ (2 mL) and extracted with EtOAc (3 x 3 mL). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude residue was purified by column chromatography (30% EtOAc in hexanes) to afford ketone formal benzylic oxidation product **100** (3.0 mg, 0.0088 mmol, 55%) as a yellow oil.

TLC: $R_f = 0.3$ (40% EtOAc in hexanes). ¹**H NMR** (600 MHz, Benzene-*d*₆) δ 6.28 (d *J* = 11.5 Hz, 1H), 6.15 (d, *J* = 3.6 Hz, 1H), 6.03 (d, *J* = 3.5 Hz, 1H), 5.92 (dd, *J* = 3.6, 0.9 Hz, 1H), 5.77 (d, *J* = 3.5 Hz, 1H), 5.71 (d, *J* = 11.5 Hz, 1H), 4.48 (s, 1H), 4.23 (qd, *J* = 6.7, 2.0 Hz, 1H), 4.18 (d, *J* = 12.8 Hz, 1H), 4.16 (dd, *J* = 2.0, 1.1 Hz, 1H), 3.95 (d, *J* = 12.8 Hz, 1H), 3.72 (s, 1H), 3.01 (s, 3H), 1.69 (s, 3H), 1.62 (s, 3H), 0.99 (d, *J* = 6.7 Hz, 3H). ¹³**C NMR** (151 MHz, C₆D₆) δ 209.9, 132.9, 130.7, 130.5, 128.3, 124.1, 121.5, 114.5, 112.5, 109.0, 106.2, 94.2, 90.6, 66.4, 59.9, 57.6, 56.7, 46.1, 25.6, 19.2, 13.4. **IR** (thin film) v_{max} (cm⁻¹): 3495, 3286, 2953, 2870, 1702, 1471, 1169, 901, 726. **HRMS** (m/z): (ESI) calcd. for C₂₁H₂₅N₂O₃ [M+H]⁺ m/z: 353.1860, found 353.1861.



Iodohydrin 76: To a reaction tube containing ketone **35** (10.0 mg, 0.032 mmol, 1.0 equiv.) was added MeOH (2 mL). The resulting mixture was cooled to 0 °C followed by the slow addition of NIS (7.2 mg, 0.032 mmol, 1.0 equiv.) in THF (0.5 mL). Upon consumption of

the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NaHCO₃ (3 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude residue was purified by column chromatography (10% EtOAc in hexanes) to afford iodohydrin **76** (12.5 mg, 0.026 mmol, 81%) as a yellow oil.

TLC: $R_f = 0.6$ (40% EtOAc in hexanes). ¹**H NMR** (700 MHz, Benzene- d_6) δ 6.18 (d, J = 3.4 Hz, 1H), 6.00 (d, J = 3.3 Hz, 1H), 5.80 (t, J = 3.3 Hz, 3H), 5.24 (s, 1H), 4.66 (d, J = 2.3 Hz, 1H), 4.44 (s, 1H), 3.76 (s, 1H), 3.65 (q, J = 6.9 Hz, 1H), 2.93 (s, 3H), 1.78 (s, 3H), 1.73 (s, 3H), 1.58 (s, 3H), 0.67 (d, J = 6.7 Hz, 3H). ¹³**C NMR** (151 MHz, C₆D₆) δ 209.5*, 131.2, 127.4, 127.1, 125.5, 114.4, 109.3, 108.2, 107.1, 96.0, 88.2, 83.8, 59.9, 56.1, 55.4, 44.5, 38.2, 24.9, 18.8, 13.4, 12.2. *see HSQC **IR** (thin film) v_{max} (cm⁻¹): 3444, 2922, 2851, 1713, 1414, 1351, 1301, 1219, 1040, 765, 597. HRMS (m/z): (ESI) calcd. for C₂₁H₂₅N₂O₃INa [M+Na]+ m/z: 503.0802, found 503.0803.



Aldehyde 102: To a reaction tube containing iodohydrin 76 (10.0 mg, 0.021 mmol, 1.0 equiv.) was added MeNO₂ (2 mL). The resulting mixture was cooled to 0 °C followed by the addition of H₂O (3.8μ L, 0.21 mmol, 10 equiv.) and AgTFA (7.0 mg, 0.032 mmol, 1.5 equiv.). Silver iodide began to precipitate immediately. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NaHCO₃ (3 mL) and extracted with EtOAc ($3 \times 5 \text{ mL}$). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude residue was purified by column chromatography (30% EtOAc in hexanes) to afford aldehyde **102** (5.5 mg, 0.016 mmol, 77%) as a yellow solid.

TLC: $R_f = 0.3$ (40% EtOAc in hexanes). ¹**H NMR** (600 MHz, Benzene-*d₆*) δ 9.52 (s, 1H), 6.04 – 6.01 (m, 2H), 5.87 (dd, *J* = 3.4, 1.0 Hz, 1H), 5.80 (d, *J* = 3.5 Hz, 1H), 4.50 (s, 1H), 4.04 (d, *J* = 1.4 Hz, 1H), 3.64 (qd, *J* = 6.7, 2.6 Hz, 1H), 3.51 (d, *J* = 2.7 Hz, 1H), 2.87 (s, 1H), 1.87 (s, 3H), 1.81 (d, *J* = 0.9 Hz, 3H), 1.72 (s, 3H), 0.59 (d, *J* = 6.7 Hz, 3H). ¹³**C NMR** (151 MHz, C₆D₆) δ 208.9, 197.1, 127.4, 126.0, 125.1, 120.7, 108.6, 108.0, 106.6, 106.5, 92.4, 86.2, 56.4, 54.6, 53.4, 45.7, 26.3, 18.3, 11.9, 11.5. **IR** (thin film) v_{max} (cm⁻¹): 3479, 2924, 2159, 1725, 1635, 1222, 773, 558, 512. **HRMS** (m/z): (ESI) calcd. for C₂₀H₂₃N₂O₃ [M+H]⁺ m/z: 339.1703, found 339.1708.



Diol 103: To a reaction tube containing ketone **35** (30.0 mg, 0.093 mmol, 1.0 equiv.) and H₂O/acetone (1/4, 5 mL) was added OsO₄ (2.5 wt% in *t*-BuOH, 95.2 mg, 0.0093 mmol, 10 mol%) and NMO (50 wt% in H2O, 22.0 mg, 0.093 mmol, 1.0 equiv.). The resulting mixture was stirred at 25 °C for 4 hours and quenched with saturated aq. Na₂SO₃ (3 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄ and concentrated in vacuo. The resulting crude product was purified by column chromatography (100% EtOAc in hexanes) to afford ketone **35** (16.0 mg, 0.050 mmol) as a white solid and diol **103** (10.1 mg, 0.028 mmol, 30% yield, 65% brsm) as a yellow oil.

TLC: $R_f = 0.2 (100\% EtOAc in hexanes). ¹H NMR (600 MHz, Benzene-$ *d₆* $) <math>\delta$ 6.30 (d, *J* = 3.5 Hz, 1H), 5.93 (dd, *J* = 3.4, 0.9 Hz, 1H), 5.84 (dd, *J* = 3.4, 1.0 Hz, 1H), 5.67 (d, *J* = 3.5 Hz, 1H), 4.84 (dd, *J* = 8.8, 3.9 Hz, 1H), 4.34 (s, 1H), 4.26 (dt, *J* = 8.7, 3.4 Hz, 1H), 3.75 (q, *J* = 5.8, 5.2 Hz, 1H), 3.67 (s, 1H), 3.46 (d, *J* = 9.7 Hz, 1H), 3.26 (d, *J* = 9.2 Hz, 1H), 2.81 (s, 1H), 1.82 (s, 3H), 1.79 (s, 3H), 1.65 (s, 3H), 0.72 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (151 MHz, C₆D₆) δ 205.3*, 130.5, 129.7, 127.6, 127.4, 113.1, 108.6, 107.9, 106.6, 101.1, 85.8, 73.4, 70.2, 58.2, 56.2, 46.5, 26.7, 18.6, 13.2, 12.3. *see HSQC IR (thin film) v_{max} (cm⁻¹): 3667, 3146, 2889, 2474, 1728, 1437, 1275, 1047, 950, 920, 868. HRMS (m/z): (ESI) calcd. for C₂₀H₂₄N₂O₄Na [M+Na]⁺ m/z: 379.1628, found 379.1626.



Ketone 104: To a reaction tube containing diol **103** (10.0 mg, 0.028 mmol, 1.0 equiv.) was added MeOH (1.5 mL). The resulting mixture was cooled to 0 °C followed by the addition of aq. 1 M HCI (56.0 μ L, 0.056 mmol, 2.0 equiv.). The resulting mixture was stirred for 10 minutes and quenched with saturated aq. NaHCO₃ (3 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude product was purified by preparative TLC (50% EtOAc in hexanes) to afford ketone **104** (8.9 mg, 0.024 mmol, 85% yield) as a yellow oil.

TLC: $R_f = 0.4$ (50% EtOAc in hexanes).¹**H NMR** (600 MHz, Chloroform-*d*) δ 6.16 (d, J = 3.4 Hz, 1H), 5.94 (dd, J = 3.4, 1.0 Hz, 1H), 5.82 (dd, J = 3.5, 1.0 Hz, 1H), 5.71 (d, J = 3.4 Hz, 1H), 4.88 (s, 1H), 4.51 (s, 0H), 4.49 (td, J = 6.7, 2.1 Hz, 1H), 4.41 (dd, J = 2.2, 1.0 Hz, 1H), 4.37 (dd, J = 8.5, 4.8 Hz, 1H), 4.28 (d, J = 4.7 Hz, 1H), 3.34 (s, 3H), 2.87 (d, J = 8.5 Hz, 1H), 2.27 (d, J = 0.9 Hz, 3H), 2.23 (d, J = 0.9 Hz, 3H), 1.69 (s, 3H), 1.56 (s, 3H), 1.55 (d, J = 6.8 Hz, 3H).¹³**C NMR** (151 MHz, CDCl₃) δ 209.0, 131.2, 128.1, 127.0, 126.5, 112.6, 108.5, 107.9, 106.5, 96.6, 87.6, 80.4, 74.5, 59.4, 57.4, 56.2, 43.9, 26.3, 19.4, 13.6, 12.7. **IR** (thin film) v_{max} (cm⁻¹): 3536, 3253, 2953, 2616, 2396, 1721, 1234, 945, 713. **HRMS** (m/z): (ESI) calcd. for $C_{20}H_{24}N_2O_4Na$ [M+Na]⁺ m/z: 393.1785, found 393.1783.



Bipolamine G (9): To a reaction tube containing ketone **104** (8.0 mg, 0.022 mmol, 1.0 equiv.) was added CH₂Cl₂ (1.5 mL). The resulting mixture was cooled to -78 °C followed by the addition of DIBAL (1.0 M in hexanes, 44.0 µL 0.044 mmol, 2.0 equiv.). Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. Rochelle salt (3 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude residue was purified by preparative TLC (50% EtOAc in hexanes) to afford bipolamine G **(9)** (3.0 mg, 0.0080 mmol, 36%) and 12-*epi*-**9** (3.1 mg, 0.0084 mmol, 38%) both as white solids.

Bipolamine G (9):

TLC: R_f = 0.4 (50% EtOAc in hexanes).¹**H NMR** (600 MHz, Acetone- d_6) δ 6.04 (d, J = 3.4 Hz, 1H), 5.91 (d, J = 3.3 Hz, 1H), 5.84 (dd, J = 3.3, 1.0 Hz, 1H), 5.78 (dd, J = 3.4, 1.0 Hz, 1H), 4.89 (d, J = 1.0 Hz, 1H), 4.86 (s, 1H), 4.36 (dd, J = 10.0, 3.6 Hz, 1H), 4.32 (qd, J = 6.7, 1.9 Hz, 1H), 4.29 (br s, 1H), 4.13 (d, J = 3.6 Hz, 1H), 3.16 (s, 3H), 3.02 (d, J = 10.0 Hz, 1H), 2.92 (s, 1H), 2.47 (td, J = 6.6, 4.4 Hz, 1H), 2.31 (d, J = 0.9 Hz, 3H), 2.26 (d, J = 0.9 Hz, 3H), 1.56 (d, J = 6.7 Hz, 3H), 1.07 (d, J = 6.6 Hz, 3H). ¹³**C NMR** (151 MHz Acetone) δ 131.5, 130.9, 128.4, 127.4, 114.1, 108.2, 108.1, 105.2, 91.5, 87.5, 81.4, 70.4, 69.2, 60.8, 57.6, 55.8, 42.6, 19.5, 18.7, 13.7, 12.8. **IR** (thin film) v_{max} (cm⁻¹): 3480, 3108, 3071, 3041, 2507, 2105, 1431, 1148. **HRMS** (m/z): (ESI) calcd. for C₂₁H₂₈N₂O₄Na [M+Na]⁺ m/z: 395.1941, found 395.1939.

12-*epi*-9:

TLC: $R_f = 0.2$ (50% EtOAc in hexanes). ¹**H NMR** (600 MHz, Acetone-*d*₆) δ 6.04 (d, *J* = 3.3 Hz, 1H), 5.84 (dd, *J* = 3.4, 1.0 Hz, 1H), 5.75 (dd, *J* = 3.4, 1.0 Hz, 1H), 5.72 (d, *J* = 3.4 Hz, 1H), 4.84 (d, *J* = 1.0 Hz, 1H), 4.55 (s, 1H), 4.36 (qd, *J* = 6.7, 2.0 Hz, 1H), 4.31 (t, *J* = 1.5 Hz, 1H), 4.28 (d, *J* = 3.7 Hz, 1H), 4.15 (d, *J* = 3.7 Hz, 1H), 3.45 (s, 1H), 3.18 (s, 3H), 2.71 (q, *J* = 6.3 Hz, 1H), 2.30 (d, *J* = 0.9 Hz, 3H), 2.25 (d, *J* = 0.8 Hz, 3H), 1.54 (d, *J* = 6.7 Hz, 3H), 1.05 (d, *J* = 6.3 Hz, 3H). ¹³**C NMR** (151 MHz Acetone) δ 131.5, 129.7, 128.2, 127.4, 114.0, 108.0, 107.8, 106.0, 92.1, 87.3, 80.7, 73.9, 71.0, 61.3, 57.6, 56.0, 42.3, 19.4, 18.9, 13.7, 12.8. **IR** (thin film) v_{max} (cm⁻¹): 3285, 3192, 2706, 2415, 1487, 1328, 1191, 1021, 811. **HRMS** (m/z): (ESI) calcd. for C₂₁H₂₈N₂O₄Na [M+Na]⁺ m/z: 395.1941, found 395.1937.



Ketone 106 and hemiacetal 107: To a reaction tube containing ketone **35** (20.0 mg, 0.062 mmol, 1.0 equiv.) was added THF (3 mL). The resulting mixture was cooled to 0 $^{\circ}$ C, Sml₂ (0.1 M in THF, 0.930 mL, 0.093 mmol, 1.5 equiv.) was added dropwise. The resulting mixture was warmed to room temperature. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NH₄Cl (5 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude product was purified by preparative TLC (20%

EtOAc in hexanes) to afford ketone **106** (17.2 mg, 0.053 mmol, 86%) and hemiacetal **107** (2.0 mg, 0.006 mmol, 9%) as light yellow oils.

Ketone **106**:

TLC: $R_f = 0.5$ (40% EtOAc in hexanes). ¹**H NMR** (600 MHz, Benzene-*d₆*) δ 6.19 (dd J = 11.7, 1.8 Hz, 1H), 6.11 (d, J = 3.5 Hz, 1H), 6.01 (td, J = 2.9, 2.5, 1.6 Hz, 2H), 5.97 (d, J = 3.4 Hz, 1H), 5.14 (dd, J = 11.7, 5.1 Hz, 1H), 4.68 (dd, J = 6.6, 2.3 Hz, 1H), 4.65 (t, J = 7.2 Hz, 1H), 3.99 (ddd, J = 7.4, 5.1, 1.7 Hz, 1H), 3.95 (qd, J = 6.9, 1.8 Hz, 1H), 3.47 (dt, J = 4.3, 2.0 Hz, 1H), 1.95 (s, 3H), 1.89 (d, J = 4.6 Hz, 1H), 1.83 (s, 3H), 1.73 (s, 3H), 0.78 (d, J = 6.9 Hz, 3H). ¹³**C** NMR (151 MHz, C₆D₆) δ 206.8, 131.7, 131.5, 130.4, 127.6, 122.6, 119.5, 112.6, 109.5, 108.2, 105.2, 75.7, 60.2, 55.0, 54.6, 37.2, 29.5, 19.9, 13.3, 12.0. IR (thin film) v_{max} (cm⁻¹): 3345, 2853, 2409, 1725, 1377, 1301, 1221, 1042, 940, 758. HRMS (m/z): (ESI) calcd. for C₂₀H₂₅N₂O₂ [M+H]⁺ m/z: 325.1911, found 325.1910.

Hemiacetal 107:

TLC: $R_f = 0.4$ (40% EtOAc in hexanes). ¹**H NMR** (600 MHz, Benzene-*d₆*) δ 6.33 (d, *J* = 11.7 Hz, 1H), 6.23 (d, *J* = 3.6 Hz, 1H), 6.07 (dd, *J* = 3.6, 1.0 Hz, 1H), 5.98 (dd, *J* = 3.5, 1.0 Hz, 1H), 5.82 (d, *J* = 3.4 Hz, 1H), 5.26 (dd, *J* = 11.7, 8.4 Hz, 1H), 4.40 – 4.32 (m, 2H), 3.88 (d, *J* = 1.5 Hz, 1H), 3.47 (q, *J* = 2.4 Hz, 1H), 2.53 (dd, *J* = 8.4, 2.3 Hz, 1H), 2.03 (s, 1H), 1.93 (d, *J* = 0.9 Hz, 3H), 1.83 (s, 3H), 1.48 (s, 3H), 0.85 (d, *J* = 6.9 Hz, 3H). ¹³**C NMR** (151 MHz, C₆D₆) 132.8, 132.1, 131.3, 127.3, 122.4, 120.1, 112.7, 109.1, 108.9, 104.2, 98.2, 77.0, 55.8, 51.1, 49.2, 32.9, 29.6, 20.5, 13.9, 12.4. **IR** (thin film) v_{max} (cm⁻¹): 3455, 2925, 2162, 2143, 1429, 1377, 1042, 940, 780, 668. **HRMS** (m/z): (ESI) calcd. for C₂₀H₂₅N₂O₂ [M+H]⁺ m/z: 325.1911, found 325.1912.



Ketone 108: To a reaction tube containing diol **98** (30.0 mg, 0.081 mmol, 1.0 equiv.) was added CH₂Cl₂ (5 mL). The resulting mixture was cooled to 0 °C, Et₃N (33.7 μ L, 0.24 mmol, 3.0 equiv.), MsCl (6.9 μ L, 0.089 mmol, 1.1 equiv.) and DMAP (9.9 mg, 0.081 mmol, 1.0 equiv.) were added dropwise sequentially. The resulting mixture was warmed to room temperature. Upon consumption of the starting material as indicated by TLC, the reaction mixture was cooled to 0 °C followed by the addition of aq. 1 M HCl (2 mL). Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NaHCO₃ (3 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude product was purified by preparative TLC (10% EtOAc in hexanes) to afford ketone **108** (23.5 mg, 0.056 mmol, 69% yield) as a white solid.

TLC: R_f = 0.4 (30% EtOAc in hexanes). ¹**H NMR** (600 MHz, Benzene-*d₆*) δ 6.36 (dd, *J* = 11.8, 6.3 Hz, 1H), 6.02 (d, *J* = 12.2 Hz, 1H), 5.93 (d, *J* = 3.6 Hz, 1H), 5.88 (d, *J* = 3.3 Hz, 1H), 5.80 (d, *J* = 3.6 Hz, 1H), 5.54 (d, *J* = 3.5 Hz, 1H), 4.86 (dd, *J* = 11.9, 3.8 Hz, 1H), 4.82 (p, J = 6.5 Hz, 1H), 4.77 (d, J = 12.1 Hz, 1H), 4.03 (d, J = 3.8 Hz, 1H), 3.70 (s, 1H), 2.00 (s, 3H), 1.96 (s, 3H), 1.80 (s, 3H), 1.80 (s, 3H), 1.23 (d, *J* = 6.6 Hz, 3H). ¹³**C NMR** (151 MHz, C₆D₆) δ 206.0, 135.8, 129.1, 128.7, 124.4, 122.7, 120.0, 114.5, 109.1, 108.5, 105.5, 84.5, 75.0, 53.9, 53.6, 44.0, 36.2, 23.6, 16.7, 13.3, 12.2. **IR** (thin film) v_{max} (cm⁻¹): 3430, 2926, 2855, 1715, 1360, 1179, 1003, 972, 635, 465. **HRMS** (m/z): (ESI) calcd. for C₂₁H₂₇N₂O₅S [M+H]⁺ m/z: 419.1635, found 419.1635.



Ketone 109: To a reaction tube containing ketone **108** (20.0 mg, 0.048 mmol, 1.0 equiv.) was added THF (4 mL). The resulting mixture was cooled to 0 °C, Sml₂ (0.1 M in THF, 0.72 mL, 0.072 mmol, 1.5 equiv.) were added dropwise. The resulting mixture was warmed to room temperature. Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with air followed by the addition of DBU (36.0 μ L, 0.24 mmol, 5.0 equiv.). Upon consumption of the starting material as indicated by TLC, the reaction mixture was quenched with saturated aq. NH₄Cl (3 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude product was purified by preparative TLC (20% EtOAc in hexanes) to afford ketone **109** (17.6 mg, 0.044 mmol, 91% yield) as a white solid.

TLC: $R_f = 0.5$ (30% EtOAc in hexanes). ¹**H NMR** (600 MHz, Benzene-*d*₆) δ 6.02 (d, *J* = 3.5, 1.0 Hz, 1H), 6.01 (d, *J* = 10.7 Hz, 1H), 5.85 (dd, *J* = 3.5, 1.2 Hz, 1H), 5.74 (d, *J* = 3.5 Hz, 1H), 5.66 (dd, *J* = 3.5, 0.9 Hz, 1H), 5.53 (dd, *J* = 10.4, 8.7 Hz, 1H), 5.14 (dd, *J* = 11.3, 5.2 Hz, 1H), 5.06 (dd, *J* = 9.9, 3.3 Hz, 1H), 4.72 (dd, *J* = 11.3, 9.8 Hz, 1H), 4.60 (qd, *J* = 11.3, 5.2 Hz, 1H), 5.06 (dd, *J* = 9.9, 3.3 Hz, 1H), 4.72 (dd, *J* = 11.3, 9.8 Hz, 1H), 4.60 (qd, *J* = 11.3, 5.2 Hz, 1H), 5.06 (dd, *J* = 9.9, 3.3 Hz, 1H), 4.72 (dd, *J* = 11.3, 9.8 Hz, 1H), 4.60 (qd, *J* = 11.3, 5.2 Hz, 1H), 5.06 (dd, *J* = 9.9, 3.3 Hz, 1H), 4.72 (dd, *J* = 11.3, 9.8 Hz, 1H), 4.60 (qd, *J* = 11.3), 5.2 Hz, 1H), 5.2

6.6, 5.1 Hz, 1H), 3.08 (dd, J = 8.7, 3.3 Hz, 1H), 2.02 (s, 3H), 1.91 (s, 3H), 1.83 (s, 3H), 1.63 (s, 3H), 1.10 (d, J = 6.7 Hz, 3H). ¹³**C NMR** (151 MHz, C₆D₆) δ 205.4, 134.7, 129.7, 126.7, 126.2, 125.0, 124.9, 111.3, 108.4, 108.0, 104.6, 80.2, 57.3, 51.4, 51.0, 42.4, 37.0, 27.7, 15.3, 12.9, 11.2. **IR** (thin film) v_{max} (cm⁻¹): 3381, 2924, 2853, 1707, 1358, 1290, 1176, 969, 872, 772. **HRMS** (m/z): (ESI) calcd. for C₂₁H₂₆N₂O₄SNa [M+Na]⁺ m/z: 425.1506, found 425.1509.



Alcohol 110 and 111: To a reaction tube containing ketone **109** (10.0 mg, 0.025 mmol, 1.0 equiv.) and MeOH (2 mL) was added NaBH₄ (1.0 mg, 0.025 mmol, 1.0 equiv.). The resulting mixture was stirred at 25 °C. Upon consumption of the starting material as indicated by TLC, the reaction mixture was cooled to 0 °C and quenched with saturated aq. NH₄Cl (3 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude residue was purified by preparative TLC (50% EtOAc in hexanes) to afford alcohol **110** (4.8 mg, 0.012 mmol, 47%) and alcohol **111** (4.7 mg, 0.012 mmol, 47%) as white solids.

Alcohol 110 (desired):

TLC: $R_f = 0.5 (50\% \text{ EtOAc in hexanes}).^1H$ **NMR**(600 MHz, Benzene-*d* $₆) <math>\delta$ 6.18 (d, *J* = 11.9 Hz, 1H), 5.98 – 5.92 (m, 2H), 5.87 (d, *J* = 3.5 Hz, 1H), 5.78 (d, *J* = 3.4 Hz, 1H), 5.60 (dd, *J* = 11.5, 5.9 Hz, 1H), 5.14 (dd, *J* = 12.0, 6.0 Hz, 1H), 4.72 – 4.64 (m, 2H), 3.90 (t, *J* = 5.9 Hz, 1H), 3.86 (s, 1H), 2.29 (q, *J* = 5.9 Hz, 1H), 2.03 (s, 3H), 1.84 (s, 3H), 1.82 (s, 3H), 1.17 (d, *J* = 6.5 Hz, 3H), 1.05 (s, 1H) 0.94 (d, *J* = 6.4 Hz, 3H). ¹³**C NMR** (151 MHz, C₆D₆) δ 133.4, 131.1, 130.1, 128.0, 123.8, 121.6, 111.6, 109.1, 108.3, 105.6, 76.1, 68.7, 54.0, 52.8, 52.5, 41.9, 37.3, 21.8, 16.1, 12.7, 11.8. **IR** (thin film) v_{max} (cm⁻¹): 3435, 2926, 2857, 1660, 1557, 1359, 1177, 972, 959, 744, 483. **HRMS** (m/z): (ESI) calcd. for C₂₁H₂₉N₂O₄S [M+H]⁺ m/z: 405.1843, found 405.1844.

Alcohol 111:

TLC: $R_f = 0.3$ (50% EtOAc in hexanes). ¹**H NMR** (600 MHz, Benzene-*d₆*) δ 6.14 (d, *J* = 11.3 Hz, 1H), 6.05 (dd, *J* = 3.4, 1.0 Hz, 1H), 6.02 (dd, *J* = 3.4, 0.9 Hz, 1H), 5.81 (d, *J* = 3.5 Hz, 1H), 5.75 (dd, *J* = 3.4, 0.9 Hz, 1H), 5.60 (dd, *J* = 11.5, 5.6 Hz, 1H), 5.55 (dd, *J* = 11.3, 7.6 Hz, 1H), 4.72 – 4.63 (m, 2H), 4.33 (dd, *J* = 8.1, 3.9 Hz, 1H), 3.43 (dt, *J* = 12.1, 6.1 Hz, 1H), 2.47 (td, *J* = 8.0, 3.9 Hz, 1H), 2.01 (s, 3H), 1.95 (s, 3H), 1.84 (s, 3H), 1.17 (d, *J* = 6.5 Hz, 3H), 0.86 (d, *J* = 6.1 Hz, 3H), 0.61 (s, 1H). ¹³**C NMR** (151 MHz, C₆D₆) δ 133.4, 131.1, 129.8, 126.5, 126.3, 123.3, 110.7, 108.9, 107.9, 105.6, 77.8, 68.4, 55.1, 51.9, 51.8, 41.4, 37.3, 22.1, 15.8, 12.8, 11.5. **IR** (thin film) v_{max} (cm⁻¹): 3687, 2929, 2178, 1602, 1357,

1177, 962, 611, 407. **HRMS** (m/z): (ESI) calcd. for $C_{21}H_{29}N_2O_4S$ [M+H]⁺ m/z: 405.1843, found 405.1844.



Diol 112 To a reaction tube containing alcohol **110** (10.0 mg, 0.025 mmol, 1.0 equiv.) and THF (1.5 mL) was added BH₃·THF (6.4 mg, 0.075 mmol, 3.0 equiv.). The resulting mixture was stirred at 50 °C. Upon consumption of the starting material as indicated by TLC, the reaction mixture was cooled to 0 °C followed by the addition of NaBO₃·4H₂O (38.5 mg, 0.25 mmol, 10 equiv.). The reaction mixture was quenched with saturated aq. NH₄Cl (5 mL) and extracted with EtOAc (3 x 10 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, and concentrated in vacuo. The resulting crude residue was purified by preparative TLC (100% EtOAc in hexanes) to afford diol **112** (4.7 mg, 0.011 mmol, 45%) as a white solid.

TLC: $R_f = 0.3 (100\% EtOAc in hexanes). ¹H NMR (600 MHz, Chloroform-$ *d* $) <math>\delta$ 6.17 (dd, *J* = 3.6, 1.4 Hz, 1H), 5.93 (dd, *J* = 3.5, 1.0 Hz, 1H), 5.86 (d, *J* = 3.4 Hz, 1H), 5.78 (d, *J* = 3.1 Hz, 1H), 4.95 (dd, *J* = 10.5, 4.9 Hz, 1H), 4.69 (t, *J* = 11.0 Hz, 1H), 4.60 (qd, *J* = 6.8, 4.8 Hz, 1H), 4.10 (s, 1H), 4.02 (dd, *J* = 11.5, 2.8 Hz, 1H), 3.08 (dd, *J* = 14.2, 5.7 Hz, 1H), 2.62 (dd, *J* = 14.2, 11.0 Hz, 1H), 2.56 (dd, *J* = 9.3, 3.8 Hz, 1H), 2.41 – 2.32 (m, 2H), 2.26 (s, 3H), 2.26 (s, 3H), 2.24 (s, 3H), 1.43 (d, *J* = 6.8 Hz, 3H), 1.38 (s, 1H), 1.18 (d, *J* = 6.2 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 129.6, 127.7, 126.8, 126.5, 108.3, 107.7, 107.6, 105.5, 82.8, 72.0, 68.3, 54.9, 50.5, 50.2, 35.7, 34.0, 33.8, 23.1, 14.4, 12.7, 11.7. IR (thin film) v_{max} (cm⁻¹): 3394, 2925, 2853, 1634, 1401, 1360, 1299, 1177, 1080, 970, 770, 649. HRMS (m/z): (ESI) calcd. for C₂₁H₃₁N₂O₅S [M+H]⁺ m/z: 423.1948, found 423.1950.



Bipolamine I (11): To a reaction tube containing diol 112 (2.4 mg, 0.0056 mmol, 1.0 equiv.) and PhMe (1 mL) was added DBU (8.5μ L, 0.056 mmol, 10.0 equiv.). The resulting mixture was heated up to 120 °C. Upon consumption of the starting material as indicated by TLC, the reaction mixture was cooled to 25 °C. The organic layers were concentrated in vacuo. The resulting crude residue was purified by column chromatography (40% EtOAc in hexanes) to afford bipolamine I (11) (1.5 mg, 0.0046 mmol, 82%) as a white solid.

TLC: $R_f = 0.4$ (50% EtOAc in hexanes). ¹**H NMR** (600 MHz, Acetone- d_6) δ 5.76 (dd, J = 3.4, 1.0 Hz, 1H), 5.75 (dd, J = 3.4, 1.0 Hz, 1H), 5.65 (br d, J = 3.1 Hz, 1H), 5.60 (d, J = 3.4 Hz, 1H), 4.56 (br d, J = 3.6 Hz, 1H), 4.40 – 4.33 (m, 3H), 3.52 (dd, J = 17.3, 4.8 Hz, 1H), 3.26 (dq, J = 11.1, 5.5 Hz, 1H), 3.13 (br d, J = 17.3 Hz, 1H), 2.97 (br t, J = 1.8 Hz, 1H), 2.22 (br d, J = 0.9 Hz, 3H), 2.19 (s, 3H), 2.16 (br dd, J = 11.0, 7.1 Hz, 1H), 1.42 (d, J = 6.6 Hz, 3H), 1.21 (d, J = 6.0 Hz, 3H). ¹³**C NMR** (151 MHz, Acetone) δ 134.9, 128.8, 126.3 (2C), 108.1, 108.0, 105.2, 102.4, 76.9, 71.2, 68.2, 56.5, 55.7, 49.6, 36.4, 31.2, 22.7, 19.1, 12.3, 12.2. **IR** (thin film) v_{max} (cm⁻¹): 3558, 2922, 2164, 1726, 1588, 1446, 1304, 1067, 562. **HRMS** (m/z): (ESI) calcd. for C₂₀H₂₇N₂O₂ [M+H]⁺ m/z: 327.2067, found 327.206.

Natural Product Spectral Comparisons

Bipolamine D (6) ¹H spectra comparison:



	¹ H NMR δ)	¹ H NMR δ)
Position	Natural Sample	Synthetic Sample
	(500 MHz, CDCl ₃) ²	(600 MHz, CDCl ₃)
1	1.49 (d, 6.3)	1.48 (d, 6.6)
2	4.32 (qd, 6.3, 2.1)	4.32 (qd, 6.6, 2.3)
3	4.35 (br s)	4.36 (s)
4	4.85 (br s)	4.84 (s)
5	3.92 (br s)	3.91 (s)
6		
7	5.83 (d, 3.0)	5.82 (d, 3.3)
8	5.76 (br d, 3.0)	5.75 (br d, 3.3)
9		
10	2.25 (br s)	2.24 (s)
11	1.13 (d, 6.6)	1.12 (d, 6.4)
12	2.93 (q, 6.6)	2.93 (qd, 6.4, 4.5)
13		
14	4.38 (br s)	4.38 (s)
15	3.11 (br d, 15.6),	3.09 (d, 15.6),
	2.88 (dd, 15.6, 5.1)	2.87 (dd, 15.6, 5.0)
16		
17	5.71 (s)	5.70 (br d, 3.3)
18	5.69 (br s)	5.67 (br d, 3.3)
19		
20	2.22 (br s)	2.21 (s)

Bipolamine D (6) ¹³C spectra comparison:



	¹³ C NMR δ)	¹³ C NMR δ)
Position	Natural Sample	Synthetic Sample
	(125 MHz, CDCl ₃) ²	(151 MHz, CDCl ₃)
1	19.2	19.2
2	59.0	59.0
3	85.6	85.6
4	59.6	59.6
5	43.7	43.7
6	131.5	131.5
7	105.5	105.5
8	107.8	107.8
9	128.2	128.3
10	12.8	12.8
11	18.7	18.8
12	72.3	72.3
13	91.8	91.8
14	68.3	68.3
15	31.4	31.4
16	128.6	128.6
17	108.8	108.8
18	107.0	107.0
19	128.8	128.8
20	13.5	13.5

Bipolamine E (7) ¹H spectra comparison:



i		
	¹ H NMR δ)	¹ H NMR δ)
Position	Natural Sample	Synthetic Sample
	$(500 \text{ MHz}, \text{CDCl}_3)^2$	(600 MHz, CDCl ₃)
1	1.54 (d, 6.5)	1.55 (d, 6.7)
2	4.32 (qd, 6.5, 1.6)	4.32 (m)
3	4.26 (br s)	4.29 (m)
4	4.89 (br s)	4.91 (s)
5	4.21 (br s)	4.22 (br s)
6		
7	5.92 (d, 3.0)	5.92 (d, 3.4)
8	5.78 (br d, 3.0)	5.78 (d, 3.4)
9		
10	2.25 (br s)	2.26 (s)
11	1.08 (d, 6.5)	1.08 (d, 6.5)
12	2.53 (q, 6.5)	2.53 (q, 6.5)
13		
14	4.29 (br s)	4.30 (br s)
15	3.06	3.07
	2.81 (dd, 15.5, 4.0)	2.82 (dd, 15.7, 4.0)
16		
17	5.68 (br d, 2.8)	5.67 (d, 3.3)
18	5.75 (br d, 2.8)	5.74 (d, 3.3)
19		
20	2.26 (br s)	2.26 (s)

Bipolamine E (7) ¹³C spectra comparison:



	¹³ C NMR δ)	¹³ C NMR δ)
Position	Natural Sample	Synthetic Sample
	(125 MHz, CDCl ₃) ²	(151 MHz, CDCl ₃)
1	19.4	19.3
2	57.5	57.5
3	87.4	87.4
4	61.0	61.0
5	44.1	44.0
6	130.5	130.5
7	105.3	105.2
8	108.0	108.0
9	128.3	128.3
10	12.7	12.7
11	18.5	18.5
12	70.2	70.3
13	92.0	91.9
14	67.3	67.3
15	34.3	34.3
16	128.9	128.8
17	108.7	108.7
18	107.8	107.8
19	128.8	128.7
20	13.5	13.5

Bipolamine G (9) ¹H spectra comparison:



	¹ H NMR δ)	¹ H NMR δ)
Position	Natural Sample	Synthetic Sample
	$(500 \text{ MHz}, \text{Acetone-} d_6)^2$	$(600 \text{ MHz}, \text{Acetone-} d_6)$
1	1.55 (d, 7.0)	1.56 (d, 6.7)
2	4.31 (qd, 7.0, 2.0)	4.32 (qd, 6.7, 1.9)
3	4.26 (br s)	4.29 (br s)
4	4.88 (br s)	4.89 (d, 1.0)
5	4.85 (br s)	4.86 (d, 1.0)
6		
7	5.92 (br d, 3.0)	5.91 (d, 3.3)
8	5.79 (br d, 3.0)	5.78 (dd, 3.4, 1.0)
9		
10	2.26 (br s)	2.26 (d, 0.9)
11	1.08 (d, 7.0)	1.07 (d, 6.6)
12	2.47 (q, 7.0)	2.47 (td, 6.6, 4.4)
13		
14	4.36 (d, 3.5)	4.36 (dd, 10.0, 3.6)
15	4.15 (d, 3.5)	4.13 (d, 3.6)
16		
17	6.07 (br d, 3.0)	6.05 (d, 3.4)
18	5.86 (br d, 3.0)	5.84 (d, 3.3)
19		
20	2.31 (br s)	2.31 (d, 0.9)
15-OMe	3.17 (s)	3.16 (s)

Bipolamine G (9) ¹³C spectra comparison:



	¹³ C NMR δ)	^{13}C NMR δ)
Position	Natural Sample	Synthetic Sample
	$(125 \text{ MHz}, \text{Acetone-}d_6)^2$	$(151 \text{ MHz}, \text{Acetone-}d_6)$
1	19.5	19.4
2	57.5	57.5
3	87.4	87.4
4	60.7	60.7
5	42.6	42.5
6	130.7	130.8
7	105.2	105.1
8	108.1	108.1
9	128.3	128.3
10	12.8	12.8
11	18.6	18.6
12	70.2	70.3
13	91.4	91.4
14	69.1	69.1
15	81.3	81.3
16	127.3	127.3
17	114.1	114.1
18	108.0	108.0
19	131.5	131.4
20	13.6	13.6
15-OMe	55.8	55.8

Bipolamine I (11) ¹H spectra comparison:



bipolamine | (10)

	¹ H NMR δ)	¹ H NMR δ)
Position	Natural Sample	Synthetic Sample
	$(500 \text{ MHz}, \text{Acetone-} d_6)^2$	$(600 \text{ MHz}, \text{Acetone-}d_6)$
1	1.42 (d, 6.6)	1.42 (d, 6.6)
2	4.37 (overlapped)	4.37 (overlapped)
3	4.36 (overlapped)	4.36 (overlapped)
4	4.56 (br d, 3.6)	4.56 (br d, 3.6)
5	2.97 (br t, 1.8)	2.97 (br t, 1.8)
6		
7	5.61 (d, 3.4)	5.60 (d, 3.4)
8	5.77 (dd, 3.4, 0.8)	5.76 (dd, 3.4, 1.0)
9		
10	2.23 (br d, 0.8)	2.22 (br d, 0.9)
11	1.22 (d, 6.0)	1.21 (d, 6.0)
12	3.26 (dq, 11.5, 6.0)	3.26 (dq, 11.1, 5.5)
13	2.16 (br dd, 11.5, 8.0)	2.16 (br dd, 11.0, 7.1)
14	4.39 (dd, 8.0, 4.8)	4.38 (overlapped)
15	3.52 (dd, 17.0, 4.8)	3.52 (dd, 17.3, 4.8)
	3.14 (br d, 17.0)	3.13 (br d, 17.3)
16		
17	5.65 (br d, 3.3)	5.65 (br d, 3.1)
18	no data	5.75 (dd, 3.4, 1.0)
19		
20	no data	2.19 (s)
Bipolamine I (11) ¹³C spectra comparison:



bipolamine I (11)

	¹³ C NMR δ)	¹³ C NMR δ)
Position	Natural Sample	Synthetic Sample
	$(125 \text{ MHz}, \text{Acetone-}d_6)^2$	(151 MHz, Acetone- d_6)
1	19.1	19.1
2	56.5	56.5
3	76.9	76.9
4	49.6	49.6
5	36.4	36.4
6	134.9	134.9
7	102.4	102.4
8	108.1	108.1
9	126.4	126.3
10	12.1	12.2
11	22.6	22.7
12	68.1	68.2
13	55.7	55.7
14	71.2	71.2
15	31.2	31.2
16	128.8	128.8
17	105.2	105.2
18	108.0 from ¹³ C NMR	108.0
19	126.3 from ¹³ C NMR	126.3
20	12.3 from 13 C NMR	12.3






























































































































