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Journal

Microbiology Resource Announcements, 6(3)

ISSN

2576-098X

Authors

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Publication Date

2018-01-18

DOI

10.1128/genomea.01387-17

Peer reviewed







Draft Genome Sequences of Two *Vibrio parahaemolyticus* Strains Associated with Gastroenteritis after Raw Seafood Ingestion in Colorado

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ABSTRACT *Vibrio parahaemolyticus* is a Gram-negative pathogen associated with gastrointestinal and wound infections after exposure to raw seafood or contaminated waters. We report here the whole-genome sequences of two stool isolates (CDC-AM50933 and CDC-AM43539) from patients in Colorado presenting with gastroenteritis after ingesting raw seafood.

Vibrio parahaemolyticus is a halophilic Gram-negative organism found in marine and estuarine environments worldwide (1). The pathogen causes the following three types of infections in patients with exposure to raw or undercooked seafood or contaminated seawaters: gastroenteritis, wound infections, and septicemia (2). Major V. parahaemolyticus virulence factors include thermostable direct hemolysin (TDH), type III secretion systems (TTSS1 and TTSS2), and type VI secretion systems (T6SS1 and T6SS2) (3–5). According to the Cholera and Other Vibrio Illness Surveillance (COVIS) system established by the CDC, the incidence of V. parahaemolyticus per 100,000 population increased from 0.01 to 0.13 in the United States from 1996 to 2010 (6). The increasing incidence of V. parahaemolyticus may be related to climate change, as warming seawaters provide the optimal growth environment for the pathogen (7).

In this paper, we describe the complete genome sequence of two clinical *V. parahaemolyticus* strains isolated from stool cultures of patients presenting with gastroenteritis after eating raw seafood in Colorado, CDC-AM50933 and CDC-AM43539. Both isolates were originally reported to the COVIS as *Vibrio vulnificus*; however, the strains were identified as *V. parahaemolyticus* by 16S rRNA sequencing. The isolates were further confirmed as *V. parahaemolyticus* on CHROMagar Vibrio (CHROMagar Microbiology, France) and TCBS (thiosulfate-citrate-bile-sucrose) agar (BD, USA).

Strain CDC-AM50933 was isolated from a stool culture of a 59-year-old healthy male who presented with *V. parahaemolyticus* gastroenteritis in Colorado. He had a history of consuming raw clams, mussels, oysters, and shrimp and cooked crab, lobster, and crawfish. He presented as an outpatient with fevers, nausea, vomiting, abdominal pain, and more than 10 episodes of diarrhea. The antibiotic treatment received was not reported to the COVIS. The symptoms lasted for 6 days with no other adverse reactions. Strain CDC-AM43539 was isolated from a stool culture of a 69-year-old male with a history of gastric bypass surgery who presented with *V. parahaemolyticus* gastroenteritis in Colorado. He had a recent history of ingesting crab, shrimp, and raw oysters. The patient presented with abdominal cramps and more than 10 episodes of diarrhea.

Received 26 November 2017 **Accepted** 1 December 2017 **Published** 18 January 2018

Citation Trinh SA, Leyn SA, Rodionov ID, Godzik A, Satchell KJF. 2018. Draft genome sequences of two *Vibrio parahaemolyticus* strains associated with gastroenteritis after raw seafood ingestion in Colorado. Genome Announc 6:e01387-17. https://doi.org/10.1128/qenomeA.01387-17.

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During his 26-day hospitalization, he was treated with a course of doxycycline and metronidazole.

Genomic DNA was extracted using the Maxwell 16-cell DNA purification kit, and DNA libraries were prepared using the Illumina Nextera XT DNA library preparation kit according to the manufacturer's protocol. An average coverage of 91× was achieved using HiSeq 4000 paired-end 150-bp sequencing at the Institute for Genome Sciences at the University of Maryland School of Medicine. Assembly was performed using SPAdes version 3.10.0 (8), and annotation was added by the NCBI Prokaryotic Genome Annotation Pipeline as implemented at the PATRIC Bioinformatics Database and Analysis Resource Center (9). Both strains CDC-AM50933 and CDC-AM43539 had genes encoding virulence factors TDH, TTSS1, and TT6SS2.

Accession number(s). The whole-genome sequences reported here have been deposited at DDBJ/ENA/GenBank under the accession numbers NPOM00000000 (CDC-AM50933) and NPOL000000000 (CDC-AM43539).

ACKNOWLEDGMENTS

We thank Cheryl Tarr for providing strains, Anna Newton and Erin Burdette for clinical information, Katherine Murphy for assistance with genome library preparation, and Kasey Cervantes for microbiological identification of strains.

This work was supported by a Ruth L. Kirschstein NRSA in Translational Research in Infectious Diseases T32A1095207 (to S.A.T.) and NIH grants R01Al092825 and R01Al098369 (to K.J.F.S.). The work by I.D.R. was conducted as part of an internship at Sanford Burnham Prebys Medical Discovery Institute.

REFERENCES

- Letchumanan V, Chan KG, Lee LH. 2014. Vibrio parahaemolyticus: a review on the pathogenesis, prevalence, and advance molecular identification techniques. Front Microbiol 5:705. https://doi.org/10.3389/fmicb.2014 .00705.
- Daniels NA, MacKinnon L, Bishop R, Altekruse S, Ray B, Hammond RM, Thompson S, Wilson S, Bean NH, Griffin PM, Slutsker L. 2000. Vibrio parahaemolyticus infections in the United States, 1973–1998. J Infect Dis 181:1661–1666. https://doi.org/10.1086/315459.
- Raghunath P. 2014. Roles of thermostable direct hemolysin (TDH) and TDH-related hemolysin (TRH) in *Vibrio parahaemolyticus*. Front Microbiol 5:805. https://doi.org/10.3389/fmicb.2014.00805.
- 4. Park KS, Ono T, Rokuda M, Jang MH, Okada K, Iida T, Honda T. 2004. Functional characterization of two type III secretion systems of *Vibrio parahaemolyticus*. Infect Immun 72:6659–6665. https://doi.org/10.1128/IAI.72.11.6659-6665.2004.
- Salomon D, Gonzalez H, Updegraff BL, Orth K. 2013. Vibrio parahaemolyticus type VI secretion system 1 is activated in marine conditions to target bacteria, and is differentially regulated from system 2. PLoS One 8:e61086. https://doi.org/10.1371/journal.pone.0061086.
- 6. Newton A, Kendall M, Vugia DJ, Henao OL, Mahon BE. 2012. Increasing

- rates of vibriosis in the United States, 1996–2010: review of surveillance data from 2 systems. Clin Infect Dis 54(suppl 5):S391–S395. https://doi.org/10.1093/cid/cis243.
- Vezzulli L, Brettar I, Pezzati E, Reid PC, Colwell RR, Höfle MG, Pruzzo C. 2012. Long-term effects of ocean warming on the prokaryotic community: evidence from the vibrios. ISME J 6:21–30. https://doi.org/10.1038/ismej .2011.89.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Wattam AR, Davis JJ, Assaf R, Boisvert S, Brettin T, Bun C, Conrad N, Dietrich EM, Disz T, Gabbard JL, Gerdes S, Henry CS, Kenyon RW, Machi D, Mao C, Nordberg EK, Olsen GJ, Murphy-Olson DE, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Vonstein V, Warren A, Xia F, Yoo H, Stevens RL. 2017. Improvements to PATRIC, the all-bacterial Bioinformatics Database and Analysis Resource Center. Nucleic Acids Res 45: D535–D542. https://doi.org/10.1093/nar/gkw1017.

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